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Influence of Soil Management on the Red Grapevine (*Vitis vinifera* L.) Mencía Must Amino Acid Composition and Wine Volatile and Sensory Profiles in a Humid Region

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Abstract: Adopting cover crops for vineyard soil management can provide several benefits, including soil protection, reductions in vine vigor, and enhancements in berry composition. However, the effects of this practice on wine aroma have seldom been addressed. This study aimed to determine the influence of different cover crops and soil tillage on the must and wine amino acid composition and wine volatile compounds of the red cultivar, ‘Mencía’ (*Vitis vinifera* L.), grown in Northwest Spain. Treatments consisted of soil tillage (ST), native vegetation (NV), English ryegrass (ER), and subterranean clover (SC). Cover crops did not alter the macro-constituents of musts; however, musts from NV and SC tended to lower concentrations of amino acids. Some color attributes of wines were influenced by cover crops in the vineyard. Methanol and *trans*-linalool oxide (pyran) concentrations in wines were significantly affected by soil management. Professional tasters encountered differences in visual, aroma, and palate descriptors of wines depending on the treatment imposed in the vineyard. These alterations in sensory properties seemed to obey to slight modifications of wine chemical characteristics due to vineyard soil management. According to these results, cover crops might be useful for modulating wine aroma in humid climates.

Keywords: nitrogen fraction; red wine; sensory profile; soil management; tillage; volatile compounds

1. Introduction

Aroma is one of the main attributes of wines and results from the complex balance of the concentrations of a high number of volatile compounds with distinct characteristics and intensities [1]. These volatiles accumulate in grapes during ripening and they are affected by temperature and water availability that, among other factors, can be affected by vineyard management practices [1,2]. Soil management in vineyards has multiple goals that encompass improving weed management, soil conservation, nutrient and water management, enhanced biodiversity for pest control, and reduce the availability of soil resources to control vine vigor [2–4]. Since these aspects are important to vine growth, soil management has relevant implications for wine quality [5–7].

One of the techniques for managing vineyard soils is the use of cover crops, although their application has been limited due to the concern of excessive water and nutrient competition between

these crops and the vines [8,9]. However, a great number of environmental and agronomic benefits can be expected from cover crops in these agroecosystems, including soil protection against erosion [10], improvements in soil properties [11], reductions in vine vigor [12], etc. Despite these advantages, vineyards are usually managed through tillage in the inter-row and herbicides in the vine row [13].

Soil management can contribute to alter the balance between grapevine (*Vitis vinifera* L.) vegetative growth and yield; thereby, modifying temperature and solar irradiation in the cluster zone [14]. Moreover, the use of cover crops as a means of soil management can reduce water availability for grapevines [9]. Furthermore, wine aroma depends on must quality, which is influenced by nitrogen composition that affects fermentation kinetics and the production of ethanol, glycerol, and both aroma and spoilage compounds [15]. Among the nitrogen compounds present in grapes, amino acids are related to wine aroma compounds [16]. Vineyard soil management can alter the concentrations of these amino acids in grapes and musts [17], although some studies reported no effects on the yeast assimilable nitrogen composition [18].

Few studies have addressed the effects that soil management through cover crops may exert on wine aroma and they reached contrasting results [19,20]. For instance, establishing cover crops in a Cabernet Sauvignon vineyard increased the concentrations of wine volatile compounds when compared to soil tillage [19]. In contrast, Negroamaro wines coming from the soil tillage treatment had greater concentrations of aroma compounds than those coming from a cover crop treatment [20]. These discrepancies can be explained by the different climate and soil conditions and the different varieties considered in both studies. Therefore, further research on other grapevine varieties and vine growing regions is needed.

In Galicia (North-West Spain), favorable temperatures and soil water availability during springtime enable a fast canopy establishment in vineyards, which may lead to unbalanced vines with high vegetative growth. In these situations, the employment of cover crops as a soil management system can reduce this excessive vegetative growth [21]. In this region, the main cultivar grown for red wine production is Mencía and the use of cover crops for floor management in these vineyards could help to achieve a better balance between vegetative growth and yield, as well as enhance the color potential of this variety. Although the chemical and phenolic profiles of Mencía wines have been previously characterized [22–25], the effect of soil management systems on Mencía wine aroma has not been previously determined.

In this context, the aims of the current study were to assess the effects of establishing permanent cover crops for vineyard soil management on the amino acid composition of musts and wines, and the aromatic and sensory profiles of wines from the red grapevine cultivar, Mencía, grown under an Atlantic climate during three consecutive years.

2. Materials and Methods

2.1. Description of the Study Site

The current study was conducted in a 0.1 ha rain-fed vineyard (*Vitis vinifera* L. cv. Mencía) located in Leiro (42°21.6' N, 8°7.02' W, elevation 115 m), Ourense, North-West Spain, within the Ribeiro Designation of Origin. The vineyard was planted in 2007, with vines grafted onto 196-17C rootstock and trained to a vertical trellis on a single cordon system (10–12 buds per vine). Rows were East-West oriented; vines were spaced 1.25 m between plants and 2.3 m between rows. Soil at the site is sandy-loamy, acidic, and with a high organic matter content. Soil depth is about 1 m and available water capacity is, approximately, 100 mm m⁻¹. Climate is temperate, humid with cool nights [26], with an average annual rainfall of 900 mm, of which about 70% falls during the dormant period. Figure 1 displays the dynamics of the mean air temperature and monthly rainfall for the study site over the three years considered.

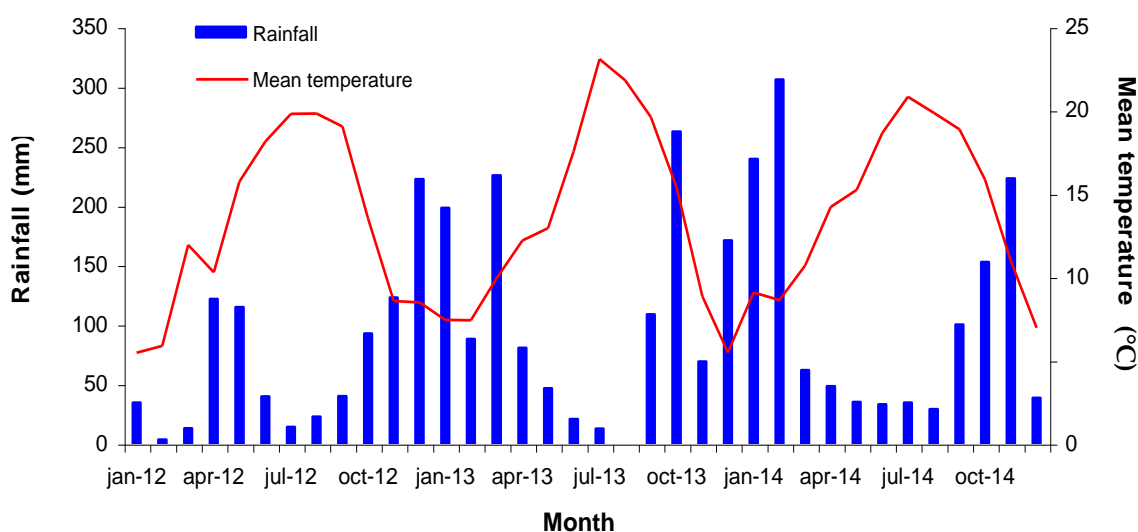


Figure 1. Monthly rainfall and mean temperature at the experimental vineyard over the three years studied (2012–2014).

2.2. Experimental Design

The experiment was conducted over three consecutive years: 2012, 2013, and 2014. Four treatments were established in a randomized block design with three replications. Each replicate consisted of three rows, with seven vines per row. The five vines in the center of the middle row were used for sampling. The treatments consisted of four different soil management systems:

1. Soil tillage (ST);
2. native vegetation (NV);
3. English ryegrass (*Lolium perenne* L.) sown at 40 kg ha⁻¹ (ER); and
4. subterranean clover (*Trifolium subterraneum* L.) sown at 30 kg ha⁻¹ (SC).

The three cover crop treatments were mowed three times per year. The ST treatment was kept with no vegetation through cultivation. With the exception of soil management, agricultural practices (such as pest and disease control, canopy management, etc.) were the same for all treatments. Further information and data on physiological, vegetative growth, and yield variables can be found in a previous report [21].

2.3. Sampling and Winemaking

The different treatments were harvested manually on the same day. Winemaking was performed separately on samples of about 35 kg per treatment. Due to limitations on grape production and fermentation tanks, and to fulfill the requirements of a wider project, only one vinification per treatment was carried out each year, by mixing the grapes from the three replications from each treatment.

Grapes were destemmed mechanically and put into stainless steel tanks. A replicated sample from each treatment was collected. During grape processing, 50 mg L⁻¹ of SO₂ were added to the mass, which was fermented at room temperature (22–24 °C). Excellence XR (Lamothe-Abiet, Bordeaux, France) yeast was added following the producer's instructions. Daily, wine lots were punched down until alcoholic fermentation ended (8 days). Then, lots were pressed, racked into new tanks, and kept at room temperature for two days. Then, wines were stabilized at 4 °C in a chamber for one month, approximately. After this period, wines were filtered, bottled, and stored.

2.4. Basic Parameters of Musts and Wines

The general parameters of Mencía musts (total soluble solids, pH, total acidity, tartaric and malic acid concentrations) and wines (alcohol content, pH, total acidity, volatile acidity, dry extract, and

tartaric and malic acid concentrations) were determined by Fourier transform infrared spectrometry (FTIR) using a WineScan FT120 analyzer (FOSS Electric, Barcelona, Spain) calibrated according to the official methods [27].

Wine color attributes, including color intensity, color hue, total polyphenol index (TPI), total anthocyanins, and total tannins contents, were determined using the methodology described by Zamora [28] using an ultraviolet-visible Helios Zeta spectrophotometer (Thermo Fisher Scientific Ltd., Waltham, MA, USA). Absorbances at 420 nm, 520 nm, and 620 nm were determined and color intensity was computed by summing these values, whereas color hue was calculated as the ration between absorbances at 420 nm over that at 520 nm and expressed in percentage.

2.5. Analytical Methods

2.5.1. Chemical Reagents

A Milli-Q equipment (Millipore, Bedford, MA, USA) was used for obtaining ultra-pure water. Amino acid solutions were prepared using standards from Acros Organics (Geel, Belgium). Acetonitrile and methanol were from Scharlau (Sentmenat, Spain), whereas ammonium chloride was from Merck (Darmstadt, Germany).

Dichloromethane, *n*-pentane, and anhydrous sodium sulphate (Scharlau, Sentmenat, Spain) were used for the extraction of volatile compounds. The internal standards (Merck, Madrid, Spain) were 4-methyl-2-pentanol for major volatile compounds; 4-decanol for terpenes and C6 alcohols; and 1-heptanol for volatile fatty acids, ethyl esters, and acetates of higher alcohols. All the standards were prepared in 50% hydro-alcoholic solutions.

2.5.2. Quantification of Amino Acids in Musts and Wines

A method based on a derivatization reaction in a basic methanolic medium and high performance liquid chromatography (HPLC) allowed us to determine the concentrations of amino acids present in musts and wines [29]. An Agilent 1100 series equipment (Agilent Technologies, Palo Alto, CA, USA) was used for performing the HPLC determinations. Chromatographic separation of amino acids was made in a Zorbax Eclipse AAA column (C18), with 5 μm particle size (150 mm \times 4.6 mm, Agilent Technologies, Palo Alto, CA, USA) thermostated at 22 °C. Extraction method, reagents, and elution conditions are described elsewhere [30]. Determinations were performed in triplicate.

2.5.3. Determination of Volatile Compounds

Concentrations of wine volatiles were determined by gas chromatography (GC) as described elsewhere [30,31], thus methods are outlined briefly.

Major volatile compounds were quantified by direct injection using a 7890A gas chromatograph (Agilent Technologies, Palo Alto, CA, USA) after adding 50 μL of internal standard (5 g L⁻¹ of 4-methyl-2-pentanol in 50% ethanol) to 50 mL of wine [30,31].

Terpenes, C6 alcohols, volatile fatty acids, ethyl esters of fatty acids, and acetates of higher alcohols were extracted by modifying an established protocol [32] consisting of the addition of internal standards to the wine sample, passing this mixture through a cartridge (Isolute ENV + SPE, Biotage, Uppsala, Sweden), cleaning the sample, adding *n*-pentane, and injecting the extract into a chromatograph following previously published conditions [30,31].

The National Institute of Standards and Technology (NIST) Mass Spectral library allowed us to identify volatiles by comparing their mass spectra and retention times with those of pure standard compounds. Calibration curves for each single compound were built for quantifying volatiles as a function of the internal standards. These curves showed high regression coefficients, indicating excellent linearity. All determinations were performed in triplicate.

2.5.4. Odor Activity Values

The contribution of a given volatile compound to the wine aroma was assessed by the calculation of its odor activity value (OAV). This indicator is the ratio between the concentration of an individual compound and its perception threshold [33,34]. Theoretically, this value should be greater than one; however, due to synergic effects among different substances, those compounds with values greater than 0.2 can be active aromas [35]. Nevertheless, we used those compounds with an OAV > 0.5 for data analysis.

2.6. Sensory Evaluation

A panel of nine judges composed of oenologists and technicians with experience in tasting Mencía wines participated in the sensory evaluation of the wines from the current experiment. This panel consisted of five males and four females between 30 and 64 years old. A scorecard including 26 descriptors (6 for color, 10 for aroma, and 10 for palate) was used. The descriptors were chosen for Galician red wines and were scored from 0 (not present) to 9 (most intense) [36]. Furthermore, judges scored the global quality of each wine. The wines were coded and presented arbitrarily to the panel in clear tulip-shaped glasses. The tasting sessions were held in April or May the year after the vintage. Data were processed with Big Sensory Soft 1.02 (Centro Studi Assagiatori, Brescia, Italy).

2.7. Statistical Analysis

As there were no replicates, the different years were considered as ‘temporal replicates’ and the statistical significance between soil management strategies was evaluated using analysis of variance. When needed, means were separated using the Tukey’s test. Principal Component Analysis (PCA) was carried out for separating must samples according to the concentrations of the eight most abundant amino acids. In addition, PCA allowed us to correlate aromatic descriptors from the sensory evaluation with those volatile compounds that appeared in the wines at concentrations at least 0.5 times their corresponding OAV. Statistical tests were performed using R software v3.4.1 [37] (R Core Team, Vienna, Austria).

3. Results and Discussion

3.1. Physiological, Vegetative Growth, and Yield Performance

A previous work reported data on grapevine performance under the four management treatments considered in the current study and for the same years [21]. Therefore, it is interesting to summarize these findings, as they may explain differences in must amino acids and wine volatiles.

Stem water potential at midday was more negative in NV, causing significant reductions in leaf stomatal conductance on certain dates, especially in 2013. This is a common response in field studies comparing grapevines growing with cover crops against a tilled soil in the inter-row [38]. In contrast, vines in SC showed less negative stem water potential values, likely due to a reduced leaf surface that caused a lower transpiration [39].

The competition exerted by cover crops produced significant reductions in leaf surface and pruning and berry weight; whereas soil management did not affect yield and cluster weight [21]. The high water supply by rainfall, which was enough to fulfill the water requirements of both the grapevines and the cover crops, can explain these results. Moreover, these findings were in accordance with those observed in regions with a similar climate to the one studied here [6,8,40].

3.2. General Attributes of Musts and Wines

As pointed out above, the competition between grapevines and cover crops was not severe and this explains the absence of significant differences among treatments for must compositional attributes (Table 1). Despite the fact that cover crops reduced berry size [21], berries from the cover

crop treatments did not accumulate more sugars (Table 1). This is likely caused by a greater leaf surface in vines from ST that compensated the greater berry size in this treatment, as reported for other regions with similar climate conditions [8,40].

Table 1. General parameters of Mencía musts for the different soil management treatments averaged for three seasons (2012–2014). Data are averages \pm standard errors.

Attribute	Soil Tillage	Native Vegetation	English Ryegrass	Subterranean Clover	Statistical Significance
Total soluble solids ($^{\circ}$ Brix)	22.8 \pm 0.7	22.4 \pm 0.5	22.2 \pm 0.7	21.1 \pm 1.1	ns
Total acidity (g L ⁻¹ tartaric acid)	4.1 \pm 0.3	4.0 \pm 0.3	4.5 \pm 0.2	4.8 \pm 0.6	ns
pH	3.85 \pm 0.06	3.79 \pm 0.04	3.74 \pm 0.05	3.65 \pm 0.10	ns
Tartaric acid (g L ⁻¹)	5.0 \pm 0.1	5.0 \pm 0.0	5.1 \pm 0.1	5.2 \pm 0.1	ns
Malic acid (g L ⁻¹)	3.4 \pm 0.2	3.1 \pm 0.2	3.4 \pm 0.1	3.3 \pm 0.3	ns

ns = not significant.

Although not significant, cover crop treatments tended to increase total acidity in Mencía musts, as previously reported for Cabernet Franc [41]. This trend is more evident in the case of the SC treatment, where musts had lower total soluble solids contents and greater total acidities than the rest of the studied treatments. Finally, organic acids contents were very similar for all treatments, as observed for Tempranillo in La Rioja [13]. Therefore, our results are in accordance with previous indications that soil management did not affect grape macro-constituents at maturity [19,42].

Wine composition was determined at the same time as the tasting sessions, five months after bottling, and soil management did not cause significant differences for any of the wine general attributes (Table 2), in accordance with the observations in the musts. However, cover crop treatments significantly altered color intensity, color hue, and the concentration of total tannins (Table 2). Wines from ER showed lower color hue values when compared to those from the other treatments, except for those from SC (Table 2).

Table 2. General parameters of Mencía wines for the different soil management treatments after five months bottled and averaged for three seasons (2012–2014). Data are averages \pm standard errors.

Attribute	Soil Tillage	Native Vegetation	English Ryegrass	Subterranean Clover	Statistical Significance
Alcohol content (% vol.)	13.2 \pm 0.5	12.9 \pm 0.6	12.7 \pm 0.5	12.1 \pm 0.4	ns
Total acidity (g L ⁻¹ tartaric acid)	4.0 \pm 0.7	4.0 \pm 0.7	4.1 \pm 0.4	3.9 \pm 0.2	ns
Volatile acidity (g L ⁻¹ acetic acid)	0.30 \pm 0.07	0.23 \pm 0.04	0.23 \pm 0.04	0.33 \pm 0.06	ns
pH	4.42 \pm 0.14	4.37 \pm 0.10	4.32 \pm 0.07	4.31 \pm 0.05	ns
Tartaric acid (g L ⁻¹)	1.8 \pm 0.4	2.0 \pm 0.4	1.7 \pm 0.4	1.9 \pm 0.2	ns
Malic acid (g L ⁻¹)	1.5 \pm 0.8	1.4 \pm 0.6	1.4 \pm 0.3	1.7 \pm 0.6	ns
Lactic acid (g L ⁻¹)	1.6 \pm 0.4	1.2 \pm 0.3	1.5 \pm 0.1	1.7 \pm 0.3	ns
Dry extract (g L ⁻¹)	32.2 \pm 3.3	29.9 \pm 2.4	28.2 \pm 1.5	28.0 \pm 1.0	ns
Absorbance 420 nm	0.349 \pm 0.032 ab	0.380 \pm 0.070 ab	0.327 \pm 0.012 a	0.431 \pm 0.063 b	*
Absorbance 520 nm	0.388 \pm 0.047 ab	0.394 \pm 0.087 ab	0.387 \pm 0.004 a	0.502 \pm 0.102 b	*
Absorbance 620 nm	0.152 \pm 0.018	0.146 \pm 0.029	0.132 \pm 0.005	0.168 \pm 0.021	ns
Color intensity	9.5 \pm 0.9 a	10.1 \pm 1.6 ab	9.1 \pm 0.7 a	10.8 \pm 1.3 b	*
Color Hue (%)	86.9 \pm 4.6 b	87.1 \pm 9.4 b	77.9 \pm 7.2 a	80.5 \pm 8.0 ab	*
Total Polyphenol Index	54.3 \pm 3.8	53.0 \pm 7.4	53.8 \pm 3.8	54.3 \pm 4.6	ns
Total anthocyanins (mg L ⁻¹)	570 \pm 100	571 \pm 127	606 \pm 112	562 \pm 111	ns
Total tannins (g L ⁻¹)	2.0 \pm 0.1 b	1.8 \pm 0.2 a	2.1 \pm 0.1 b	2.1 \pm 0.1 b	*

Different letters in the row indicate significant differences among treatments. ns = not significant; * = significant at $p < 0.05$.

3.3. Amino Acid Composition of Musts and Wines

Although a previous study in another Galician winegrowing region (Ribeira Sacra) reported total amino acid contents in Mencía musts [43], the current work is the first one to determine the concentrations of individual amino acids in Mencía musts (Table 3). Independently of the soil management treatment, the total amino acid contents were lower in the current study than those

reported for Ribeira Sacra [43], although a high variability was observed in both studies. However, the concentrations of individual compounds (Table 3) were within previously reported ranges [44], except for those of tryptophan, which were slightly greater.

Table 3. Soil management effects on the amino acid concentrations (mean \pm standard error, mg L⁻¹) of musts from Mencía averaged for three seasons (2012–2014).

Compound	Soil Tillage	Native Vegetation	English Ryegrass	Subterranean Clover	Statistical Significance
Aspartic acid	43.5 \pm 9.7	34.8 \pm 4.4	45.3 \pm 9.2	45.8 \pm 10.1	ns
Glutamic acid	81.1 \pm 10.7	59.8 \pm 10.1	80.5 \pm 12.9	62.4 \pm 19.4	ns
Asparagine	6.9 \pm 0.4	4.5 \pm 0.4	8.1 \pm 3.1	5.2 \pm 0.6	ns
Serine	44.9 \pm 5.2	39.5 \pm 4.8	53.2 \pm 12.5	42.0 \pm 7.7	ns
Glutamine	128.1 \pm 9.7	91.7 \pm 12.3	198.8 \pm 98.3	96.1 \pm 20.4	ns
Histidine	33.0 \pm 5.0	27.9 \pm 6.0	43.1 \pm 18.2	28.6 \pm 7.0	ns
Glycine	3.3 \pm 0.6	3.0 \pm 0.4	3.1 \pm 0.4	2.7 \pm 0.4	ns
Threonine	71.2 \pm 13.9	64.9 \pm 15.1	87.8 \pm 32.0	70.7 \pm 19.4	ns
Arginine	339.4 \pm 61.7	258.0 \pm 62.8	361.9 \pm 99.1	291.0 \pm 55.7	ns
Alanine	111.4 \pm 25.5	90.4 \pm 25.8	114.7 \pm 31.9	90.1 \pm 18.4	ns
γ -Aminobutyric acid (GABA)	56.8 \pm 24.1	60.7 \pm 32.8	58.9 \pm 31.0	49.3 \pm 25.5	ns
Proline	12.6 \pm 3.3	11.0 \pm 3.1	13.2 \pm 4.7	7.9 \pm 3.1	ns
Tyrosine	6.9 \pm 2.5	5.7 \pm 1.7	8.9 \pm 3.1	7.3 \pm 2.3	ns
Ammonium ion	174.3 \pm 21.4	162.5 \pm 18.2	182.8 \pm 19.2	183.0 \pm 23.3	ns
Valine	24.0 \pm 1.0	22.2 \pm 2.1	35.5 \pm 15.7	20.8 \pm 4.8	ns
Methionine	4.9 \pm 2.0	4.4 \pm 1.9	8.7 \pm 5.9	3.4 \pm 1.2	ns
Cysteine	2.1 \pm 0.3	2.2 \pm 0.3	2.2 \pm 0.3	2.1 \pm 0.3	ns
Isoleucine	12.5 \pm 0.4	13.4 \pm 2.1	21.2 \pm 10.6	10.2 \pm 1.9	ns
Tryptophan	10.8 \pm 1.8	13.3 \pm 2.3	22.6 \pm 13.0	11.3 \pm 3.6	ns
Leucine	15.9 \pm 1.3	14.7 \pm 1.6	25.3 \pm 13.1	12.9 \pm 3.2	ns
Phenylalanine	9.6 \pm 1.0	7.8 \pm 0.3	15.9 \pm 7.8	7.6 \pm 1.5	ns
Ornithine	7.0 \pm 0.8	4.9 \pm 1.1	13.3 \pm 8.3	6.3 \pm 2.0	ns
Lysine	5.9 \pm 0.7	5.2 \pm 0.8	6.1 \pm 1.5	5.2 \pm 0.9	ns
Sum of amino acids	1031.8 \pm 118.9	839.7 \pm 148.5	1228.3 \pm 401.8	878.8 \pm 162.6	ns

ns = not significant. Sum of amino acids does not account for ammonium ion concentration.

Soil management system did not affect the concentrations of amino acids in Mencía musts (Table 3), due to the high interannual variability observed. In this sense, other authors did not find significant differences in the amino acid concentrations among Tempranillo musts coming from grapevines grown under soil tillage or cover crop treatments during the first year of their study, while the differences among treatments detected in subsequent years did not coincide between years [45]. The concentrations of amino acids in musts depend on many factors, including grapevine variety, maturation stage, weather conditions, and management practices [46], and a combination of these factors might have caused these differences with previous studies on other cultivars and also explain the absence of differences among the treatments considered in the current work.

The NV and SC treatments tended to cause lower (p -value $<$ 0.1) concentrations of amino acids in Mencía musts (Table 3). In contrast, ER tended to increase the concentration of amino acids (p -values $<$ 0.08), such as valine, isoleucine, leucine, and phenylalanine (Table 3). These modifications on the amino acid composition can be explained by the different water status observed on vines from the different treatments [21]. Since these amino acids are precursors of wine volatile compounds [47], these slight alterations might modify the wine aroma.

Independently of soil management, arginine was the most abundant amino acid in Mencía musts (Table 3). Therefore, this cultivar can be considered an arginine accumulator, similarly to other red grapevine varieties, such as Syrah, Merlot [48], Garnacha, and Pinot noir [44]. Moreover, previous studies carried out by our research group proved that other three Galician grapevine varieties—namely Albariño, Godello, and Treixadura—are also arginine accumulators [30,31,49].

In fact, arginine represented about 31.6% of the total free amino acids in Mencía musts, independently of the treatment (Figure 2a). Other amino acids present at relevant concentrations in the musts from this variety were glutamine (12.6%), alanine (10.3%), glutamic acid (7.2%), and threonine (7.5%). These compounds were also abundant in other Spanish grapevine varieties, such as Monastrell and Verdejo [48,50,51]. Soil management caused very slight differences on the percentages of each

free amino acid, although they were greater in the case of minor amino acids (Figure 2b). The high variability among samples prevented the detection of significant differences among treatments. The ER treatment presented the highest variability for most of these compounds (Figure 2b). Therefore, soil management slightly altered the Mencía amino acid profile, and this could allow for discerning among management systems [48].

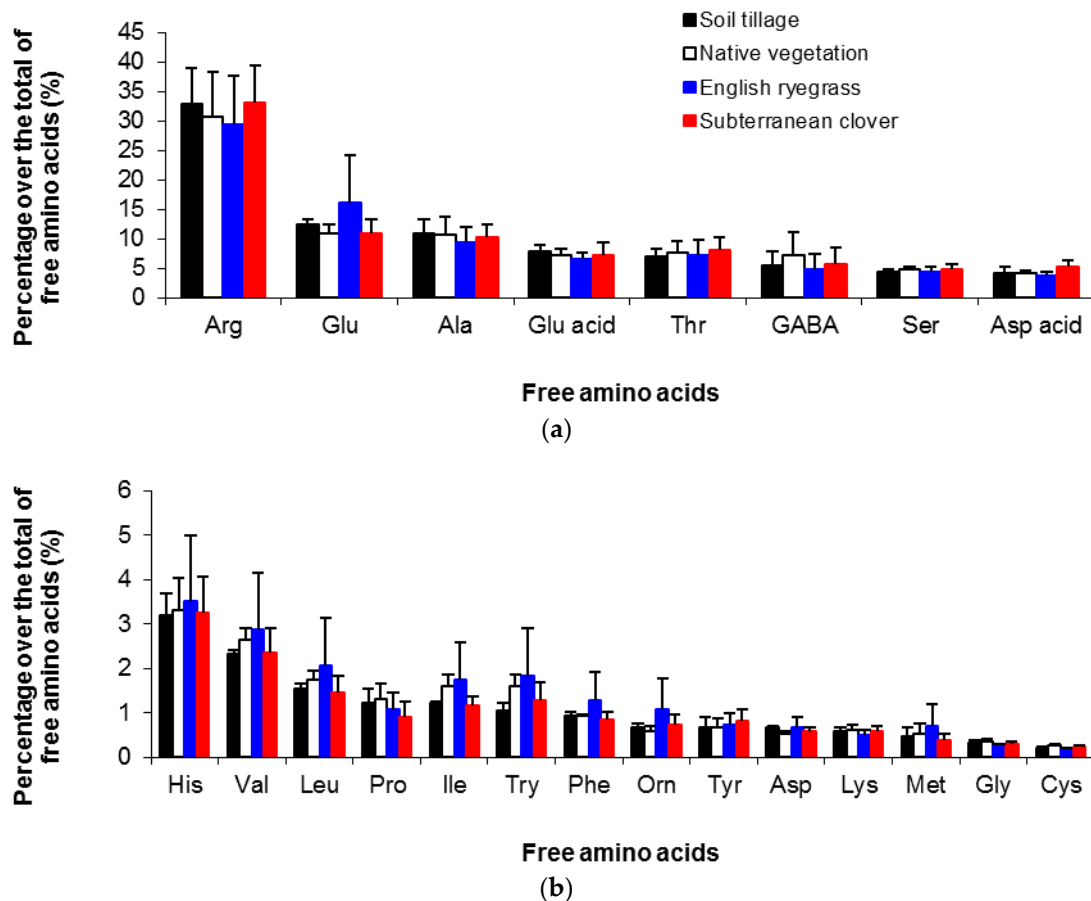


Figure 2. Percentage of each amino acid over the total amino acids in Mencía musts from different soil management systems averaged for three seasons (2012–2014). The upper and lower graphs depict the major (a) and minor (b) amino acids, respectively. Bars indicate standard errors. Arginine (Arg); glutamine (Glu); alanine (Ala); glutamic acid (Glu acid); threonine (Thr); γ -aminobutyric acid (GABA); serine (Ser); aspartic acid (Asp acid); histidine (His); valine (Val); leucine (Leu); proline (Pro); isoleucine (Ile); tryptophan (Try); phenylalanine (Phe); ornithine (Orn); tyrosine (Tyr); asparagine (Asp); lysine (Lys); methionine (Met); glycine (Gly); cysteine (Cys).

When considering the eight most abundant amino acids in Mencía musts, representing around 84% of the total amino acid content, PCA was able to separate samples from each treatment (Figure 3). The two first principal components (PC) explained 93.5% of the total variance in the dataset: PC1 accounted for 73.1% and PC2 for 20.4%. Musts from the ER treatment were located on the positive side of PC1 due to their high concentrations in arginine, threonine, serine, glutamic acid, glutamine, and alanine. Similarly, samples from the ST treatment were located also on the positive side of PC1, although close to the origin. The SC and NV treatments were located on the negative side of PC1; SC was on the positive side of PC2 due to its high concentration of aspartic acid, whereas NV was on the negative side of PC2 due to its high GABA content. These results agree with the fact that the amino acid profile could be a means for discerning vineyard management systems [48].

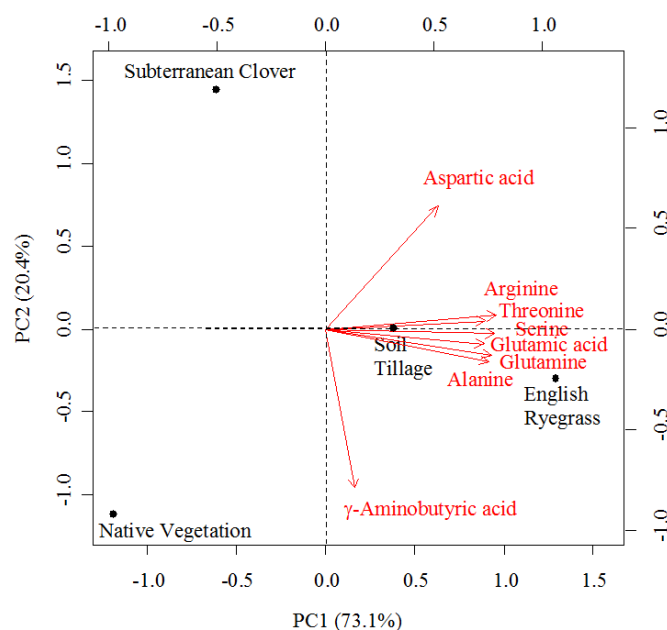


Figure 3. Principal component analysis (PCA) of Mencia musts: Biplot for the first two components (PC) for the eight most abundant amino acids in musts.

Cover crops did not affect the amino acid concentrations of Mencia wines (Table 4) due to the inter-annual variability on these concentrations, as observed for musts. However, wines from the cover crop treatments tended to have lower amino acid concentrations than those from the ST treatment (Table 4); however, these trends were not significant at the 95% level. The total and individual concentrations of amino acids in Mencia wines were greater than those detected in red wines from other varieties [52,53]. This confirmed the significant effect of grapevine variety on the amino acid concentrations observed in wines and could allow detection of the wine origin [48,54].

Table 4. Soil management effects on the amino acid concentrations (mean \pm standard error, mg L⁻¹) of wines from Mencia averaged for three seasons (2012–2014).

Compound	Soil Tillage	Native Vegetation	English Ryegrass	Subterranean Clover	Statistical Significance
Aspartic acid	53.6 \pm 10.7	35.2 \pm 5.1	40.7 \pm 8.0	38.6 \pm 0.7	ns
Glutamic acid	26.9 \pm 9.9	13.8 \pm 4.3	12.9 \pm 5.9	13.9 \pm 6.8	ns
Asparagine	9.0 \pm 4.2	6.8 \pm 3.3	14.3 \pm 7.9	10.3 \pm 5.1	ns
Serine	13.6 \pm 1.7	8.8 \pm 1.9	10.7 \pm 3.1	10.8 \pm 2.6	ns
Glutamine	3.9 \pm 1.3	2.4 \pm 0.7	4.4 \pm 2.3	3.5 \pm 1.3	ns
Histidine	18.9 \pm 3.2	12.5 \pm 1.5	16.5 \pm 5.9	13.3 \pm 2.8	ns
Glycine	26.4 \pm 2.6	16.9 \pm 3.2	22.5 \pm 6.8	20.0 \pm 4.6	ns
Threonine	12.7 \pm 1.5	8.1 \pm 2.1	10.7 \pm 3.5	11.2 \pm 3.3	ns
Arginine	44.3 \pm 18.1	15.7 \pm 3.3	36.3 \pm 21.2	20.0 \pm 11.8	ns
Alanine	43.8 \pm 6.6	25.4 \pm 5.9	41.7 \pm 17.3	35.2 \pm 9.4	ns
γ -Aminobutyric acid (GABA)	31.7 \pm 2.0	19.7 \pm 4.0	34.9 \pm 13.1	27.7 \pm 6.4	ns
Proline	4833.8 \pm 1347.2	4144.6 \pm 1087.3	4377.5 \pm 1673.9	3892.8 \pm 1207.9	ns
Tyrosine	8.1 \pm 3.3	6.2 \pm 2.5	7.6 \pm 3.7	3.3 \pm 2.7	ns
Ammonium ion	25.6 \pm 0.8	20.8 \pm 4.0	22.7 \pm 4.8	25.1 \pm 5.0	ns
Valine	31.2 \pm 6.1	17.8 \pm 3.5	23.8 \pm 9.6	23.4 \pm 5.5	ns
Methionine	5.9 \pm 0.7	4.0 \pm 1.3	5.1 \pm 1.9	4.5 \pm 1.2	ns
Cysteine	0.9 \pm 0.6	0.8 \pm 0.6	1.0 \pm 0.6	0.7 \pm 0.6	ns
Isoleucine	10.4 \pm 1.7	6.3 \pm 1.0	7.5 \pm 2.3	7.8 \pm 1.1	ns
Tryptophan	4.9 \pm 2.1	3.8 \pm 1.7	4.7 \pm 2.2	3.3 \pm 1.4	ns
Leucine	22.8 \pm 5.9	14.0 \pm 4.9	19.1 \pm 7.4	19.1 \pm 5.4	ns
Phenylalanine	13.0 \pm 2.7	8.3 \pm 2.8	10.9 \pm 4.2	11.1 \pm 3.2	ns
Ornithine	25.6 \pm 5.8	22.4 \pm 11.8	28.8 \pm 13.7	34.9 \pm 17.6	ns
Lysine	33.8 \pm 3.5	21.3 \pm 6.1	27.3 \pm 8.9	26.5 \pm 6.0	ns
Sum of amino acids	441.5 \pm 54.1	270.1 \pm 46.5	381.4 \pm 126.3	338.9 \pm 68.7	ns

Sum of amino acids does not account for ammonium ion and proline concentrations. ns = not significant.

3.4. Volatile Composition of Mencía Wines

Forty-three volatile compounds were quantified in Mencía wines (Table 5). According to their chemical characteristics, they were grouped into higher alcohols, other alcohols, acetates of higher alcohols, esters, volatile fatty acids, terpenes, and other compounds.

Table 5. Soil management effects on the concentrations (mean \pm standard error) of volatile compounds in wines from Mencía averaged for three seasons (2012–2014).

Compound	Soil Tillage	Native Vegetation	English Ryegrass	Subterranean Clover	Statistical Significance
Methanol (mg L ⁻¹)	171 \pm 10 c	126 \pm 14 a	136 \pm 12 ab	150 \pm 13 b	**
Ethyl acetate (mg L ⁻¹)	50 \pm 1	53 \pm 8	51 \pm 7	52 \pm 11	ns
Acetaldehyde (mg L ⁻¹)	6 \pm 3	14 \pm 6	8 \pm 2	13 \pm 9	ns
Higher alcohols (mg L ⁻¹)					
1-propanol	39 \pm 6	37 \pm 8	33 \pm 7	36 \pm 9	ns
2-methyl-1-propanol	43 \pm 2	50 \pm 2	43 \pm 2	50 \pm 7	ns
2-methyl-1-butanol	215 \pm 10	235 \pm 6	210 \pm 13	203 \pm 22	ns
3-methyl-1-butanol	57 \pm 4	66 \pm 6	56 \pm 6	54 \pm 6	ns
Σ Higher alcohols	355 \pm 15	389 \pm 7	342 \pm 18	344 \pm 39	ns
Other alcohols (mg L ⁻¹)					
1-hexanol	1.7 \pm 0.6	1.8 \pm 0.6	1.7 \pm 0.6	1.9 \pm 0.6	ns
<i>trans</i> -3-hexen-1-ol	0.11 \pm 0.04	0.11 \pm 0.02	0.09 \pm 0.02	0.10 \pm 0.04	ns
<i>cis</i> -3-hexen-1-ol	0.13 \pm 0.00	0.13 \pm 0.01	0.12 \pm 0.02	0.13 \pm 0.05	ns
Benzyl alcohol	3.91 \pm 0.26	3.89 \pm 0.63	3.79 \pm 0.51	3.53 \pm 0.63	ns
2-phenylethanol	25 \pm 8	34 \pm 6	24 \pm 3	23 \pm 4	ns
Other compounds (mg L ⁻¹)					
Ethyl lactate	31 \pm 5	26 \pm 9	33 \pm 14	29 \pm 10	ns
Acetoine	12 \pm 6	13 \pm 9	10 \pm 3	11 \pm 3	ns
Acetol	38 \pm 4	34 \pm 7	31 \pm 5	37 \pm 8	ns
2,3-butanediol <i>levo</i>	875 \pm 126	806 \pm 67	687 \pm 57	688 \pm 112	ns
2,3-butanediol <i>meso</i>	293 \pm 94	269 \pm 71	224 \pm 67	227 \pm 76	ns
Methionol	1.23 \pm 0.22	1.02 \pm 0.11	1.08 \pm 0.31	1.23 \pm 0.42	ns
γ -butyrolactone	58 \pm 16	50 \pm 19	47 \pm 15	50 \pm 10	ns
Acetates of higher alcohols (mg L ⁻¹)					
Isoamyl acetate	0.6 \pm 0.1	0.6 \pm 0.1	0.5 \pm 0.0	0.6 \pm 0.2	ns
2-phenylethyl acetate	0.25 \pm 0.08	0.26 \pm 0.09	0.22 \pm 0.07	0.23 \pm 0.08	ns
Σ Acetates of higher alcohols	0.85 \pm 0.12	0.86 \pm 0.14	0.72 \pm 0.08	0.78 \pm 0.23	ns
Esters (mg L ⁻¹)					
Ethyl butyrate	0.09 \pm 0.03	0.12 \pm 0.05	0.15 \pm 0.05	0.12 \pm 0.06	ns
Ethyl hexanoate	0.11 \pm 0.04	0.15 \pm 0.09	0.19 \pm 0.07	0.18 \pm 0.08	ns
Ethyl octanoate	0.15 \pm 0.05	0.18 \pm 0.09	0.20 \pm 0.10	0.21 \pm 0.09	ns
Ethyl decanoate	0.03 \pm 0.02	0.05 \pm 0.03	0.26 \pm 0.20	0.03 \pm 0.02	ns
Σ Ethyl esters C6-C10	0.38 \pm 0.13	0.48 \pm 0.26	0.71 \pm 0.36	0.54 \pm 0.25	ns
Volatile fatty acids (mg L ⁻¹)					
Isobutyric acid	1.77 \pm 0.23	2.59 \pm 0.56	1.91 \pm 0.27	2.73 \pm 1.13	ns
Butyric acid	1.41 \pm 0.23	1.60 \pm 0.39	1.52 \pm 0.35	1.41 \pm 0.53	ns
Isovaleric acid	1.04 \pm 0.14	1.40 \pm 0.16	1.09 \pm 0.13	1.37 \pm 0.50	ns
Σ Volatile fatty acids C4-C5	4.22 \pm 0.56	5.60 \pm 1.11	4.52 \pm 0.72	5.51 \pm 1.56	ns
Hexanoic acid	2.06 \pm 0.43	2.46 \pm 0.58	2.34 \pm 0.42	2.15 \pm 0.78	ns
Octanoic acid	1.55 \pm 0.36	1.67 \pm 0.36	1.80 \pm 0.33	1.43 \pm 0.50	ns
Decanoic acid	0.31 \pm 0.06	0.33 \pm 0.07	0.32 \pm 0.10	0.27 \pm 0.10	ns
Σ Volatile fatty acids C6-C10	3.93 \pm 0.84	4.46 \pm 1.01	4.46 \pm 0.82	3.85 \pm 0.67	ns
Free terpenes (μ g L ⁻¹)					
<i>trans</i> -linalool oxide (furan) ^a	5.8 \pm 0.4	5.6 \pm 0.9	6.5 \pm 1.6	4.8 \pm 1.9	ns
<i>cis</i> -linalool oxide (furan) ^a	1.9 \pm 0.8	0.8 \pm 0.3	1.1 \pm 0.4	0.9 \pm 0.3	ns
<i>trans</i> -linalool oxide (pyran) ^a	3.6 \pm 0.1 b	3.3 \pm 0.4 ab	1.9 \pm 0.4 a	2.5 \pm 0.9 ab	*
<i>cis</i> -linalool oxide (pyran) ^a	1.4 \pm 0.3	1.2 \pm 0.2	1.0 \pm 0.3	0.7 \pm 0.4	ns
Linalool (L)	3.8 \pm 1.3	4.5 \pm 1.4	6.4 \pm 2.1	3.9 \pm 1.5	ns
Hotrienol (3,7-dimethyl-1,5,7-octatriene-3-ol) ^a	5.3 \pm 4.1	3.9 \pm 2.9	2.7 \pm 1.5	6.3 \pm 2.7	ns
α -terpineol (α T)	3.9 \pm 0.6	3.0 \pm 0.6	4.0 \pm 0.3	4.7 \pm 1.6	ns
Citronellol (C)	16.6 \pm 0.8	17.8 \pm 0.1	14.9 \pm 3.0	20.0 \pm 7.2	ns
Nerol (N)	17.2 \pm 5.8	10.8 \pm 4.1	19.4 \pm 6.5	17.3 \pm 6.0	ns
Geraniol (G)	30.7 \pm 3.3	34.0 \pm 1.5	36.1 \pm 8.0	32.9 \pm 11.0	ns
Σ Free terpenes (L + α T + C + N + G)	83.8 \pm 13.8	80.9 \pm 6.9	86.8 \pm 11.1	94.0 \pm 31.4	ns
Hodiol I	4.9 \pm 2.0	4.9 \pm 2.5	4.2 \pm 2.1	3.2 \pm 1.3	ns
(<i>trans</i> -3,7-dimethyl-1,5-octadiene-3,7-diol) ^a					
2,7-dimethyloctane-4,5-diol ^a	119.9 \pm 9.7	112.0 \pm 5.0	110.3 \pm 18.5	99.5 \pm 38.2	ns

Different letters in the row indicate significant differences among treatments according to Tukey's test. ns = not significant; * = $p < 0.05$; ** = $p < 0.01$. ^a indicates that the compound is expressed in μ g L⁻¹ of internal standard (4-decanol).

Previous reports characterized the aroma composition of Mencía wines from other regions within Galicia [22,25,43], so it is expected that the concentrations of volatile compounds observed in wines from the current study would differ from those previously published. For instance, the concentrations of 1-hexanol and benzyl alcohol were greater in wines from our study than in those previously reported [22].

Soil management slightly altered the concentrations of volatiles in Mencía wines (Table 5). According to the quantitative data, the total concentration of volatiles in wines from the four treatments ranged from 1616 to 1938 mg L⁻¹. Wine from ER had the lowest amount while that from the ST treatment had the highest. Methanol contents were greater in samples from ST than in those from the cover crop treatments. The other compound that showed significant differences due to soil management was *trans*-linalool oxide (pyran), which appeared at lower levels in wines coming from ER when compared to those from ST (Table 5). The rest of the volatiles did not show significant differences among soil managements, although we observed trends to greater concentrations of esters and volatile fatty acids in wines from the cover crop treatments. Moreover, 2-phenylethanol, which imparts rose nuances [23,55], was present at greater concentrations in wines from NV.

Vineyard-soil management effects on the volatile composition of wines disagree among studies. For instance, significant differences depending on the soil management system for almost all the volatile compounds quantified in Cabernet Sauvignon wines have been found; wines from the soil tillage treatment showed the lowest concentrations of volatiles [19].

In contrast, higher levels of volatiles in Negroamaro wines coming from the cover crop treatments were detected when compared to those coming from soil tillage [20]. These discrepancies are due to the different level of competence between cover crops and grapevines in each study, which greatly depends on soil and climate conditions in the study region. Furthermore, an effect of the grapevine cultivar cannot be discarded. In our case, the lack of differences in grapevine water status and the slight alterations in leaf surface and berry size caused the absence of alterations in wine volatile compound concentrations [21].

In the current study, higher alcohols in wines from all treatments appeared at concentrations greater than the threshold considered to contribute negatively to wine complexity, 300 mg L⁻¹ [56], although they were similar to those previously detected in commercial Mencía wines from Ribeira Sacra and Monterrei [22]. Moreover, higher alcohols constitute more than 95% of the concentration of volatiles in Mencía wines from Valdeorras [23]. Similarly, other alcohols that impart positive nuances, such as 2-phenylethanol, appeared at concentrations within the range reported for commercial wines from this variety [22]. Moreover, the use of commercial yeasts could have homogenized the volatile contents in wines from the different treatments.

Only two acetates of higher alcohols were detected in the wines from the current study: Isoamyl acetate and 2-phenylethyl acetate, which did not differ among treatments (Table 5). They appeared at concentrations similar to those of commercial wines from this cultivar [22].

Ethyl esters constitute one of the most important groups of aroma compounds in wines because they impart fruity notes. Since yeasts produce them during fermentation as secondary products of sugar metabolism [57], soil management is not supposed to alter significantly their concentrations, as occurred in this study (Table 5). However, other studies reported higher concentrations of these compounds in Cabernet Sauvignon wines coming from the cover crop treatments when compared to those coming from soil tillage [19], while the opposite was found in Negroamaro wines [20].

Similarly, soil management did not alter the concentrations of volatile fatty acids in Mencía wines (Table 5). These compounds originate from the metabolism of fatty acids by yeast [57]. These molecules may contribute negatively to wine aroma, providing rancid and cheese notes; however, concentrations between 4 and 10 mg L⁻¹ of C6-C10 volatile fatty acids provide mild and pleasant aromas to wines due to synergistic effects [58]. In this study, all wines had C6-C10 volatile fatty acid contents within this range (Table 5).

Terpenes are relevant contributors to the wine aroma, and correlations between floral sensory attributes and high levels of some of these molecules, such as α -terpineol and linalool, have been documented [1]. In our case, soil management did not affect the concentrations of terpenes in Mencía wines, except for that of *trans*-linalool oxide (pyran) (Table 5). Furthermore, wines from ER and SC tended to have greater concentrations of terpenes, which might be caused by the fact that precursors of these compounds abound in berry skin [23,59] and, in these two treatments, berries were smaller and, likely, they had a lower flesh:skin ratio [21]. However, these compounds appeared in concentrations lower than their perception thresholds and their contribution to wine aroma would be not important. In fact, the contents of terpenes detected in the current study were lower than those previously observed in Mencía wines [22]. Nevertheless, linalool and α -terpineol appeared at concentrations similar to those observed in Mencía wines from Valdeorras [23].

When considering the OAV as an index of the contribution of a given compound to wine aroma [33,34], only 18 compounds had an OAV greater than 0.5 (Table 6). Among these compounds, the ones with the greatest OAV were 1-propanol, ethyl octanoate, isovaleric acid, isoamyl acetate, and acetaldehyde. Soil management did not alter the OAV of the volatile compounds determined in Mencía wines (Table 6) due to the high variability among samples. However, the OAV found in the current study were higher than those reported for the same cultivar [36]. Ethyl esters had a great influence on the aroma of the wines from the current study, and cover crop treatments tended to induce higher OAV for these compounds (Table 6). Ethyl butyrate, hexanoate, and octanoate (notes to strawberry, apple, and pear, respectively) showed high OAV in the wines of Mencía, in accordance with previous reports [23].

Table 6. Soil management effects on the odor activity values (mean \pm standard error) of volatile compounds in wines from Mencía averaged for three seasons (2012–2014).

Compound	Odor Threshold ($\mu\text{g L}^{-1}$)	Descriptor	Soil Tillage	Native Vegetation	English Ryegrass	Subterranean Clover	Statistical Significance
Ethyl acetate	7500	Pineapple	6.6 \pm 0.1	7.0 \pm 1.0	6.8 \pm 1.0	7.0 \pm 1.4	ns
Acetaldehyde	500	Fruity	13.0 \pm 5.5	27.4 \pm 12.8	16.0 \pm 4.2	25.4 \pm 17.6	ns
Higher alcohols							
1-propanol	750	Alcohol	52.2 \pm 7.5	49.6 \pm 11.2	43.9 \pm 9.8	48.2 \pm 11.3	ns
2-methyl-1-propanol	40,000	Alcohol	1.1 \pm 0.1	1.3 \pm 0.1	1.1 \pm 0.0	1.3 \pm 0.2	ns
Other alcohols							
Benzyl alcohol	620	Blackberry	6.3 \pm 0.4	6.3 \pm 1.0	6.1 \pm 0.8	5.7 \pm 1.0	ns
2-phenylethanol	14,000	Rose	1.8 \pm 0.1	2.4 \pm 0.4	1.7 \pm 0.2	1.7 \pm 0.3	ns
Other compounds							
Acetoin	10,000	Butter, almond	1.2 \pm 0.6	1.3 \pm 0.9	1.0 \pm 0.3	1.1 \pm 0.3	ns
Acetates of higher alcohols							
Isoamyl acetate	30	Banana	19.1 \pm 3.9	19.6 \pm 4.2	16.6 \pm 1.7	18.4 \pm 1.6	ns
2-phenylethyl acetate	250	Rose, honey	1.0 \pm 0.0	1.1 \pm 0.0	0.9 \pm 0.0	0.9 \pm 0.0	ns
Esters							
Ethyl butyrate	20	Fruity	4.7 \pm 1.3	5.9 \pm 2.6	7.3 \pm 2.3	6.2 \pm 2.7	ns
Ethyl hexanoate	14	Fruity	7.5 \pm 3.2	10.4 \pm 6.7	13.8 \pm 4.8	12.7 \pm 5.9	ns
Ethyl octanoate	5	Fruity	30.7 \pm 9.8	36.9 \pm 17.9	40.7 \pm 19.1	41.3 \pm 18.5	ns
Volatile fatty acids							
Isobutyric acid	2300	Cheese	0.8 \pm 0.1	1.1 \pm 0.2	0.8 \pm 0.1	1.2 \pm 0.4	ns
Butyric acid	173	Cheese	8.1 \pm 1.3	9.2 \pm 2.3	8.8 \pm 2.0	8.1 \pm 2.0	ns
Isovaleric acid	33	Cheese	31.7 \pm 4.1	42.5 \pm 4.8	32.9 \pm 3.9	41.4 \pm 8.3	ns
Hexanoic acid	3000	Cheese	0.7 \pm 0.1	0.8 \pm 0.2	0.8 \pm 0.1	0.7 \pm 0.1	ns
Octanoic acid	500	Rancid	3.1 \pm 0.7	3.3 \pm 0.7	3.6 \pm 0.7	2.9 \pm 0.4	ns
Free terpenes							
Geraniol	30	Flower	1.0 \pm 0.1	1.1 \pm 0.1	1.2 \pm 0.3	1.1 \pm 0.1	ns

ns = not significant. Only the compounds with odor activity values greater than 0.5 are shown. Odor thresholds have been taken from Guth (1997) and Ferreira et al. (2000) [33,34].

3.5. Sensory Profiles of Mencía Wines

Figure 4 shows the sensory profiles for Mencía wines from the different soil management systems studied, as averaged for the three years considered.

Visually, wines from the ST had lower violet and cherry reflects than wines from the cover crop treatments (Figure 4a). Aromatically (Figure 4b), the wine from SC received the lowest scores for black, red, and ripened fruit. In contrast, wines from ST received the highest marks for these descriptors. Judges perceived floral notes more intensely in wines from the NV and EC treatments. Palate descriptors showed slight differences among treatments (Figure 4c). Wines from ST received higher marks for persistence, but lower for body and green tannins. Finally, wines from NV, ER, and SC received global quality scores 6.5%, 9.7%, and 19.4%, respectively; lower than wines from ST.

These results seem in contradiction with those previously found [6,19], which showed that Cabernet Sauvignon wines coming from the cover crop treatments scored higher than those from the soil tillage control. However, this contradiction can be explained by the different climate conditions on each site, since in our case, grapes from the ST treatment were ripened when harvested in contrast to other studies [6]. Although Xi et al. [19] did not report data from musts in their study, wine alcohol in wines from the tilled soil treatment was significantly lower than that of wines from the cover crop treatments, which could lead to an unbalanced wine.

Furthermore, non-trained consumers tasted the same wines from this study and they ranked the wine from the ST treatment in fourth place [21]. This clearly contrasts with the results from the current sensory analysis, adding information to the debate on the selection of wine tasters [60].

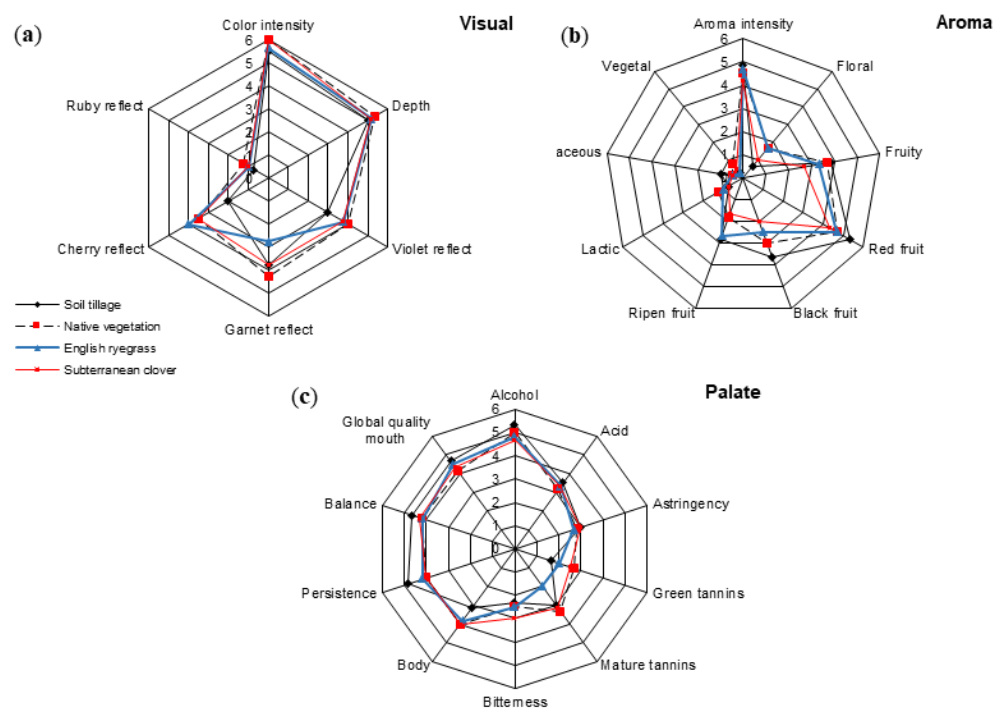


Figure 4. Sensory profile of Mencía wines (average values for 2012–2014) as a function of the soil management system: (a) Visual, (b) aroma, and (c) palate descriptors.

3.6. Relationships among Amino Acids, Volatile Compounds, and Sensory Descriptors

Previous studies proved the close relationship between the concentrations of amino acids in musts and volatiles in wines [16,31]. In the current study, concentrations of several amino acids correlated significantly with those of wine volatiles. For instance, cysteine and aspartic acid concentrations in musts correlated with those of 26 and 22 individual volatile compounds in wines, respectively.

In contrast, asparagine and phenylalanine concentrations in musts related only to that of an individual volatile (data not shown).

When considering those volatiles with an OAV greater than one and the aroma descriptors, a PCA revealed a clear separation of the wines produced under each treatment (Figure 5). The first two principal components accounted for 78.3% of the total variance in our dataset: PC1 explained 46.2% of this variance and PC2 explained 32.1%. Wines from ST appeared on the positive sides of both PC1 and PC2, due to their high scores in red, black, and ripened fruits, as well as for high concentrations in 1-propanol. Wines from NV and ER were located on the positive side of PC1, but on the negative side of PC2. High concentrations in 2-phenylethanol and isobutyric acid, as well as high scores for the floral descriptor, characterized NV wines. Great concentrations of butyric and octanoic acids and geraniol characterized wines from ER. Finally, wines from SC appeared on the negative side of PC1, but on the positive side of PC2 due to their low concentrations on volatile compounds.

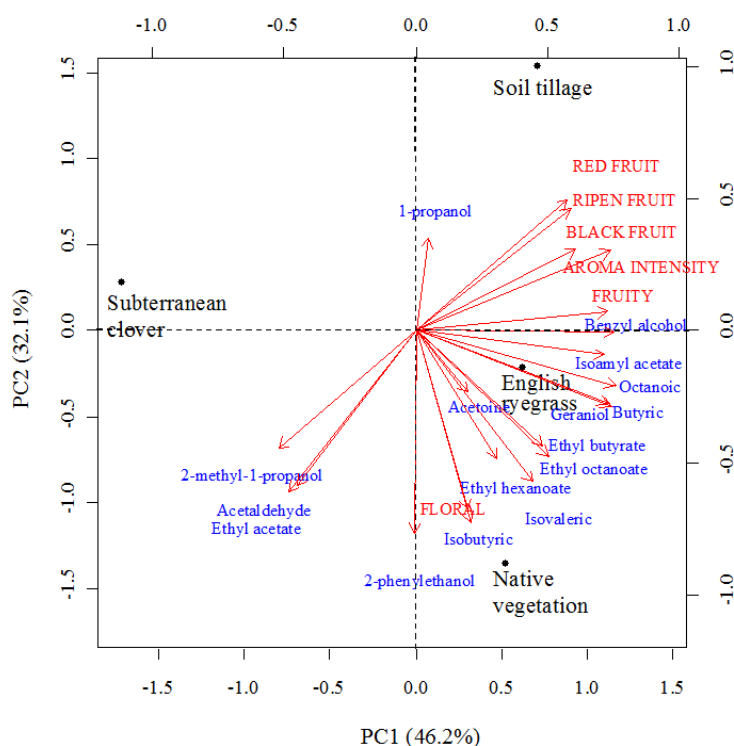


Figure 5. Principal component analysis (PCA) of Mencía wines: Biplot for the first two components (PC) for volatile compounds with odor activity values greater than one (lower-case letters) and the sensory descriptors related to wine aroma (capital letters).

4. Conclusions

The current study provided insights about the effects of establishing cover crops in the vineyard may have on Mencía must and wine composition under the climate conditions of Galicia (NW Spain). Must general attributes were not altered by the use of cover crops; however, the color characteristics of the wines were modified by the soil management system. Moreover, some tendencies were detected, although significant differences among treatments were not established in most cases. Musts from the NV and SC treatments tended to have lower amino acid concentrations. Wines from the cover crop treatments tended to have higher concentrations of ethyl esters, volatile fatty acids, and free terpenes than wines from the ST control. A correlation between volatile compounds with concentrations higher than their perception thresholds and aroma descriptors has been observed. In conclusion, vineyard soil management using cover crops seem to be an appropriate option for modulating must and wine composition in Mencía under humid climate conditions.

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