

Loyola University Chicago

Biology: Faculty Publications and Other Works

Faculty Publications and Other Works by Department

12-2019

Neuromodulation with Electromagnetic Stimulation for Seizure Suppression: From Electrode to Magnetic Coil

Hui Ye Loyola University Chicago, hye1@luc.edu

Stephanie Kaszuba Rosalind Franklin University of Medicine and Science

Follow this and additional works at: https://ecommons.luc.edu/biology_facpubs

Part of the Biology Commons

Author Manuscript This is a pre-publication author manuscript of the final, published article.

Recommended Citation

Ye, Hui and Kaszuba, Stephanie. Neuromodulation with Electromagnetic Stimulation for Seizure Suppression: From Electrode to Magnetic Coil. IBRO Reports, 7, : 26-33, 2019. Retrieved from Loyola eCommons, Biology: Faculty Publications and Other Works, http://dx.doi.org/10.1016/j.ibror.2019.06.001

This Article is brought to you for free and open access by the Faculty Publications and Other Works by Department at Loyola eCommons. It has been accepted for inclusion in Biology: Faculty Publications and Other Works by an authorized administrator of Loyola eCommons. For more information, please contact ecommons@luc.edu.



This work is licensed under a Creative Commons Attribution-Noncommercial-No Derivative Works 3.0 License. © Elsevier, 2019.

Neuromodulation with electromagnetic stimulation for seizure suppression: From electrode to magnetic coil

Hui Ye¹, Stephanie Kaszuba²

¹Department of Biology, Loyola University Chicago, Chicago, 1032 W. Sheridan Rd., IL 60660. Hye1@luc.edu

² Chicago Medical School, Rosalind Franklin University of Medicine and Science, 3333 Green Bay Rd., North Chicago, IL 60064. stephanie.kaszuba@my.rosalindfranklin.edu

Correspondence: Hui Ye, Ph.D. Department of Biology Loyola University Chicago 032 W. Sheridan Rd., Chicago, IL 60660 Tel (773) 508-2720 hye1@luc.edu

Running title: Seizure suppression with magnetic coil

Abstract

Non-invasive brain tissue stimulation with a magnetic coil provides several irreplaceable advantages over that with an implanted electrode, in altering neural activities under pathological situations. We reviewed clinical cases that utilized time-varying magnetic fields for the treatment of epilepsy, and the safety issues related to this practice. Animal models have been developed to foster understanding of the cellular/molecular mechanisms underlying magnetic control of epileptic activity. These mechanisms include (but are not limited to) (1) direct membrane polarization by the magnetic field, (2) depolarization blockade by the deactivation of ion channels, (3) alteration in synaptic transmission, and (4) interruption of ephaptic interaction and cellular synchronization. Clinical translation of stimulation protocols, and evaluation of the long-term safety. Cellular and molecular studies focusing on the mechanisms of magnetic stimulation are of great value in facilitating this translation.

Keywords: epilepsy; magnetic stimulation; animal models; cellular mechanisms

Abbreviations

- 4-AP: 4-aminopyridine
- CD₅₀: Convulsant dose
- DBS: Deep Brain Stimulation
- EcoG: Electrocorticography
- EEG: Electroencephalography
- ELF-MF: Extremely low frequency magnetic fields
- GABA: Gamma-Aminobutyric acid
- HFS: High Frequency Stimulation
- KA: Kainic acid
- LD₅₀: Lethal dose
- LTD: Long-term depression
- LTP: Long-term potential
- MEG: Magnetoencephalography
- MRI: Magnetic resonance imaging
- NMDAR: N-methyl-D-aspartate receptor
- PTZ: Pentylenetetrazol
- REM: Rapid eye movement
- tDCS: Transcranial direct-current stimulation
- TES: Transcranial electrical stimulation
- TLE: temporal lobe epilepsy
- TMS: Transcranial magnetic stimulation
- rTMS: Repetitive transcranial magnetic stimulation
- SMF: Static magnetic field

Epilepsy is one of the most widespread and devastating neurological disorders, with a lifetime prevalence of 7.60 per 1,000 people (Fiest et al., 2017). The disease is characterized by abnormal neural activity in the brain, which ultimately leads to spontaneous recurrent seizures. In the U.S. alone, the annual epilepsy-related medical expenses are close to \$15.5 billion (NIH), with yearly epilepsy-specific healthcare costs ranging up to \$19,749 per patient (Begley and Durgin, 2015). Despite significant advances made in new pharmacological development (Kaur et al., 2016), traditional anti-epileptic drugs demonstrate limited specificity in targeting particular groups of cells and epileptic neural circuitry. One-third of patients continue to experience pharmacologically intractable seizures (Kwan and Brodie, 2000, Laxer et al., 2014) and may have to consider a variety of surgical options, such as resectional surgery (e.g. temporal lobectomy, cortical excision, lesionectomy) and disconnection surgery (e.g. corpus callosotomy and functional hemispherotomy). These surgeries are irreversible, and are often associated with many neurological deficits such as memory, speech, motor, and visual impairments (Josephson et al., 2013).

1. Electric stimulation for seizure control

Electric activation of neurons has been reported for more than two centuries, dating back to the discovery by Luigi Galvani in 1780 (Galvani, 1791), who accidently found that muscle from a deceased frog would twitch upon touch with a charged metal scalpel. The event sparked the appreciation of electricity in relation to animation — or life. At present, electric stimulation of neurons in the central and peripheral nervous systems has been successful in controlling neural network activity (Selimbeyoglu and Parvizi, 2010), regulating synaptic transmission (Nowak and Bullier, 1998), alleviating memory loss (Esmaeilpour et al., 2017) and blocking pain (Coderre et al., 1993, Rodrigo et al., 2017). Electric currents have also been clinically used to modulate or suppress seizure activity, as a reversible and adjustable alternative to surgical removal of epileptic foci. One successful therapy is vagus nerve stimulation (Morris and Mueller, 1999, Yuan and Silberstein, 2016, Oliveira et al., 2017).

Deep bran stimulation (DBS)

Electric currents delivered via deeply implanted electrodes, or deep bran stimulation (DBS), have been effective in alleviating seizures in humans. In this practice, many discrete brain structures have been target areas, including the locus coeruleus (Faber and Vladyka, 1983), centromedian nucleus (Velasco et al., 2000), anterior nucleus of the thalamus (Kerrigan et al., 2004, Salanova et al., 2015), cerebellum (Cooper, 1973, Cooper et al., 1973), and other predetermined epileptic foci (Morrell and Group, 2011, Sun and Morrell, 2014). It is generally

believed that the effects of an electric field on neuronal tissue are caused by the establishment of a transmembrane potential (Esselle and Stuchly, 1994, Schnabel and Struijk, 2001, McIntyre et al., 2004, Ye et al., 2007, Lu et al., 2008). In addition, other field-induced mechanisms, such as K⁺ release (Shin and Carlen, 2008, Sutton et al., 2013), depolarization blockage (Gluckman et al., 1996, Lian et al., 2003), inactivation of voltage-gated currents (Shin et al., 2007), altered synaptic transmission (Chiken and Nambu, 2014), or altered field coupling among neurons (Ghai et al., 2000), are believed to be involved in the modification of neural network activity. A more detailed discussion is also presented in section 5.

Despite the significant improvement and clinical promise of electrical stimulation for seizure control with directly implanted electrodes, its implementation comes with some technical and biological limitations, mainly invasiveness and poor bio-compatibility.

DBS cannot avoid the risks of major surgery including hemorrhage (1-2%) and infection (3-5%) (Doshi, 2011, Bjerknes et al., 2014). Other complications include lead migration (1.60%) and electrode fracture (1.46%) (Jitkritsadakul et al., 2017). The complication rate is dependent on the experience of the surgical team.

A primary concern in DBS device design is biocompatibility of the implanted electrodes (Polikov et al., 2005). Therapeutic effects can be largely altered by inflammatory and immune responses due to the direct contact between the tissue and stimulating electrode (Kim et al., 2004, Liu et al., 2017). The formation of glial scarring around individual electrodes (Polikov et al., 2005, Grill et al., 2009) can block electric currents produced by the electrode, which causes a change (or even loss) of the resultant neural response. Electrodes implanted into the primary visual cortex of macaque monkeys lost effectiveness within a few months, even though each electrode had reliably elicited a visual percept (phosphene) shortly after implantation (Davis et al., 2012). There is some recent exploration into new electrode designs and implantation techniques to minimize tissue response and promote long-term stability of the implants (Liu et al., 2017, Luan et al., 2017).

When DBS patients are examined by Magnetic Resonance Imaging (MRI), excess heat can be produced at the stimulating electrode tip due to the interaction between the MRIgenerated radio-frequency waves and the conductive leads (Angelone et al., 2004), which can result in neurological damage (Rezai et al., 2004). In examination of possible alternatives to the customary platinum-iridium electrodes, carbon nanotube yarns have demonstrated improved biocompatibility and decreased MRI distortion (Jiang et al., 2013, Guo et al., 2015).

Transcranial direct current stimulation (tDCS)

tDCS, a noninvasive method that modulates cortical excitability, was developed in the last 20 years for the treatment of epilepsy. tDCS is applied using two electrodes (anode and cathode) positioned on the cranium to deliver a weak electric current and alter cortical excitability. Cortical excitability decreases under cathodal stimulation (Nitsche and Paulus, 2000), a principle used for suppressing epileptiform discharges and seizures in basic and clinical studies of epilepsy (Fregni et al., 2006c, Varga et al., 2011, Yook et al., 2011, Auvichayapat et al., 2013, San-Juan et al., 2017, San-Juan et al., 2018). tDCS is generally considered safe in clinical practice (Matsumoto and Ugawa, 2017). tDCS experimental protocols have resulted in only minor side effects, including mild headache and itching at the site of electrode placement (Fregni et al., 2005) (Fregni et al., 2006a), but no obvious adverse effects such as cognitive impairment (Fregni et al., 2006a).

New methods have been developed in recent years to further improve the efficiency of neural control by tDCS. Voroslakos et al. developed an "intersectional short pulse" method to increase the intensity of the electric current injected into the brain, and to keep the sensation on the scalp surface relatively low (Voroslakos et al., 2018). Grossman et al. used temporally interfering electric fields to stimulate neurons throughout a region where interference between the multiple fields establishes an electric field envelope (Grossman et al., 2017). The researchers demonstrated that temporally interfering stimulation facilitated the targeting of deep neurons in living mice without stimulating overlying cortical cells. Using a rodent model of generalized epilepsy, Berenyi et al. developed a closed-loop system to provide on-demand stimulation, which avoided detrimental side effects of continuous stimulation (Berenyi et al., 2012). These works have significantly improved the temporal and spatial resolution of tDCS.

2. Magnetic stimulation as an alternative method in neural modulation

While electric currents delivered by tissue-contacting electrodes have provided a physical mechanism to modulate neuronal activity, electric currents can also be generated via magneto-electric induction with magnetic coils (Maccabee et al., 1991, Maccabee et al., 1993, Ye et al., 2010, Ye et al., 2011, Ye and Steiger, 2015). Magnetic stimulation on excitable biological tissues was first reported in the early 20th century by Jacques d'Arsonval (1896) and Silvanus P. Thompson (1910) with their pioneer work on human visual sensations. Effects of a time-varying magnetic field are generally believed to be caused by its induced electric field and the establishment of a transmembrane potential (Ye et al., 2007) (Pashut et al., 2014). In comparison to electrical stimulation, magnetic stimulation offers advantages in biocompatibility and consistency.

While electrodes require direct contact with biological tissue, magnetic stimulation may stimulate neural tissue without requiring surgery for coil implantation. The magnetic field can penetrate biological tissue without much attenuation, thereby maintaining its intensity and stimulation consistency.

More importantly, magnetic stimulation can prevent the direct contact between an electrode and neural tissue, eliminating numerous problems that arise at the brain-electrode interface. For example, issues including charge transfer, electrode surface modification, and corrosion are reduced through coil-based stimulation (Polikov et al., 2005, Cogan, 2008, Koivuniemi et al., 2011). The coils are capable of stimulating specific nuclei with decreased disruption of surrounding regions. By avoiding direct contact with brain tissue, these coils greatly enhance biocompatibility and MRI compatibility (Golestanirad et al., 2018, Zaeimbashi et al., 2018).

Coils may still be implanted if focal stimulation is required. The coils are insulated with a soft biocompatible material, which attenuates the cortical tissue response to implantation (Saxena et al., 2013, Canales et al., 2015, Lee et al., 2016) while increasing stimulation intensity to the target tissue. Regulating the spatial orientation of miniature-sized implantable coils allows the induced electric fields to be specifically designed to activate particular groups of neurons while simultaneously avoiding others (Bonmassar et al., 2012, Lee and Fried, 2014). In the cortex, this could include the ability to activate vertically oriented pyramidal neurons without activating horizontally oriented passing axons (Lee et al., 2016). Encapsulation of the miniature coils could prevent many adverse effects and the diminishing of coil performance over time, as occurs with electrodes (Lee et al., 2016).

3. Epilepsy treatment with magnetic field and its safety

Reports on epilepsy treatment with magnetic field emerged in the 1990s (Anninos et al., 1991, Anninos et al., 1999). It was reported that magnetic field caused attenuation in seizure frequency and alteration in the circadian occurrence of seizure (Sandyk and Anninos, 1992b). The authors proposed that magnetic fields alter the functions of the pineal gland, which is a magnetosensitive organ that "transduces" environmental information of the light-dark cycle and earth's magnetic field into an endocrine message. This message, mediated via circadian release of melatonin, attenuates seizure activity. In a case report, external magnetic field was applied to seizure foci (Sandyk and Anninos, 1992a). This was done by emitting back the same intensity and frequency of magnetic field defined by the magnetoencephalography (MEG) from the patient. The method was successful in mitigating seizure activity in over 150 patients with

various forms of epilepsy. Similarly, it was found that external magnetic stimulation applied in the frontal, occipital, and temporal lobes resulted in rapid attenuation of the MEG activity of epileptic patients (Anninos et al., 2003).

At present, repetitive transcranial magnetic stimulation (rTMS) is amongst emerging options for seizure treatment. Overall, case reports demonstrate reduction of seizure frequency and/or epileptic discharges after rTMS applications. Menkes and Gruenthal investigated the impact of slow-frequency rTMS in a patient with medically refractory partial seizures due to focal cortical dysplasia (Menkes and Gruenthal, 2000). rTMS led to a 70% decrease in the frequency of seizures and a 77% reduction in the occurrence of interictal spikes. In another study, rTMS was delivered to the vertex with a round coil, at an intensity 5% below motor threshold (Brasil-Neto et al., 2004). This led to a 22.8% decrease in the mean daily number of seizures in patients. Kinoshita et al. evaluated the effects of rTMS on seizure frequency in adults with medically intractable extratemporal lobe epilepsy (Kinoshita et al., 2005). After low-frequency rTMS for one week, the frequency of all seizure types, complex partial seizures, and simple partial seizures was reduced by 19.1, 35.9, and 7.4%, respectively. Liu et al. reported that transcranial magnetic stimulation (TMS) decreased seizure frequency for refractory focal status epilepticus in the intensive care unit (ICU) (Liu et al., 2013). In a randomized double-blinded study, rTMS significantly decreased the number of seizures in the active compared with the sham rTMS group for patients with refractory epilepsy and malformations of cortical development (Fregni et al., 2006b). Likewise, low-frequency high intensity rTMS (90% resting motor threshold) delivered into the epileptogenic zone had a significant antiepileptic effect on patients with refractory partial seizures (Sun et al., 2012). A recent meta-analysis revealed a 30% average rate of 50% seizure reduction when low-frequency rTMS was used in the treatment of drug-resistant epilepsy (Cooper et al., 2018). This is consistent with previous analysis that identified a 34% reduction in seizure frequency after rTMS treatment (Hsu et al., 2011).

In general, rTMS is a safe practice with some side effects. Liu et al. reported that rTMS was safe and did not interfere with the functioning of ICU equipment (Liu et al., 2013). In a systematic review, Pereira et al. summarized 46 studies with epileptic subjects undergoing rTMS. Among these subjects, 18.3% reported adverse events. The majority of the adverse effects were mild, with headache or dizziness being most common. The authors calculated a 2.9% per subject seizure risk (Pereira et al., 2016). It appears that in about 2% of the studies, rTMS could induce seizures, especially when rTMS was implemented in relatively high frequency (Rossi et al., 2009). A recent systematic review of available data indicates that risk

7

from TMS/theta-burst stimulation is small in children and is similar to that in adults (Allen et al., 2017).

4. Basic research that investigates magnetic control of epilepsy

Although clinical and experimental results show that magnetic stimulation, including rTMS, is effective and relatively safe in the treatment of epilepsy, the characteristics of the magnetic field are empirical in these studies. Most of the explorations were case reports and randomized trails, which showed high dependency on individual parameters. Animal models were developed to provide better control of these parameters in basic lab research, and to provide in-depth study of the neurological mechanisms underlying magnetic control of seizure.

Interactions between inhibitory and excitatory elements shape neural network activity (Ziburkus et al., 2006). Alteration of this balance is believed to be the underlying mechanism of epileptogenesis (Epsztein et al., 2006, Derchansky et al., 2008, Lasztoczi et al., 2009, Huberfeld et al., 2011). Therefore, animal models of epilepsy were developed mainly by disrupting this balance. Current animal model work focuses on investigating the possibility of magnetic intervention in restoring the excitation/inhibition balance. A few studies have started to delineate the neurological mechanisms of its action.

Magnetic fields may suppress seizure by altering the inhibitory network. Sung et al. found that magnetic field exposure decreases an animals' convulsion susceptibility to bicuculline (an antagonist of GABA_A receptors) (Sung et al., 2003). Mice were exposed to either a placebo, or 20 G of extremely low frequency magnetic fields (ELF-MF) for 24 hours. Bicuculline was administered intraperitoneally at various doses and the seizure onset time and duration were measured. In addition, lethal dose (LD_{50}) and convulsant dose (CD_{50}) of the clonic and tonic convulsions were measured. The mice subjected to ELF-MFs showed moderately higher CD_{50} , LD₅₀, and induction time on the bicuculline-induced seizure. In another investigation, Kistsen et al. studied the anticonvulsive effects of different modes of impulse magnetic field on the picrotoxin (a non-competitive channel blocker of GABA_A receptors) seizure model, with the animals exposed to a big ring coil (Kistsen et al., 2016). In the study, picrotoxin was injected at a dose of 2.5 mg/kg subcutaneously after either rTMS or a placebo. Exposure to rTMS caused a decrease in the number of seizures and a reduction in the convulsive readiness of the brain, which was quantified by the latent period of myoclonuses during the picrotoxin test. These works suggest that magnetic stimulation may alter the convulsion susceptibility through a GABAergic mechanism.

Magnetic fields may suppress seizure by altering the excitatory network. Sung et al. studied the effects of magnetic field exposure on the convulsant and lethal doses of NMDA-

8

induced seizures in animals (Sung et al., 2003). Subjection to magnetic field was followed by a significant increase in glutamate and decrease in GABA levels in this seizure model. Yet, this elevation in glutamate concentration did not precipitate an increase in convulsion response. Pentylenetetrazol (PTZ) increases calcium and sodium influx to the cell, both of which depolarize the neuron. After intraperitoneal administration (60 mg/kg) of PTZ, mice were exposed to a 50 Hz, 2 G (0.2 mT) magnetic field in glass cages for 1 hour. This magnetic field did not demonstrate a significant effect on the average number of PTZ-induced seizures, seizure latency, total seizure duration, or mortality (Keskil et al., 2001). In another work, magnetic stimulation was applied to a rat kainate (a glutamate receptor agonist) status epilepticus model. Bursts of high-frequency rTMS, together with a low dose of lorazepam, suppressed seizures (Gersner et al., 2016a). Therefore, the minimal effect of magnetic field on the excitatory network could be promoted in conjunction with pharmacologic approaches.

Magnetic fields may suppress seizure by altering cellular properties. Repeated stimulation of the amygdala can prompt afterdischarges and motor seizure (Chen et al., 2016). Potschka et al. found that chronic exposure of rats to a 50-Hz, 100-μT magnetic field exerted weak inhibitory effects on some seizure parameters in amygdala kindled rats (Potschka et al., 1998). In another study, application of rTMS during amygdala kindling prevented seizures. A cellular mechanistic study revealed that rTMS administration inhibited kindling-induced changes in the electrophysiological properties of hippocampal CA1 pyramidal neurons (Shojaei et al., 2014).

Magnetic fields may suppress seizure by altering synaptic activity. Varro et al. studied the effects of extremely low frequency electromagnetic field (ELF-EMF) on living rats. Animals were then sacrificed and the brain slices were examined (WhB or "Whole body group"). The authors compared this to a group to rats who were not exposed to magnetic field while they were living, but rather their brain slices were exposed to the ELF-EMF ("slice group"). Interestingly, the development of seizure-like activity was promoted in the WhB group. In contrast, seizure activity was inhibited in the slice group (Varro et al., 2009). The authors concluded that ELF-EMF exposure exerts significant effects on synaptic activity, which depended on the specific spatial parameters and constancy of EMF.

These animal model investigations provide a link between bench studies and clinical practice. Outcomes from such systemic level works beg for in-depth analysis of cellular and molecular studies on the biological effects of magnetic fields.

5. Possible cellular mechanisms underlying magnetic field stimulation

Electric fields that are induced by the magnetic stimulation have been found to control many aspects of neuronal behavior, which set the foundation for the magnetic control of seizure. However, in comparison to the large body of literature studying the cellular basis of seizure control with direct electric stimulation, studies with magnetic field as an inhibitory stimulus are rare. There are several possible molecular/cellular mechanisms by which magnetic fields could affect network activity, and potentially suppress seizure, namely (A) direct alteration of neuronal excitability, (B) alteration of ion channel functions, (C) alteration of synaptic transmission, (D) interruption of ephaptic effects, and so on.

(A) Direct alteration of neuronal excitability by induced membrane polarization

Evidence supports the direct membrane polarization of individual neurons by magneticallyinduced currents in neuronal tissue. In vivo and in vitro studies from TMS revealed that low frequency stimulation decreases neural activity, while high frequency stimulation excites neural circuitry (Dayan et al., 2013, Parkin et al., 2015). Application of rTMS conserved normal neuronal firing of CA1 pyramidal neurons induced by kindling and prevented hyperexcitability in these cells (Shojaei et al., 2014, Moradi Chameh et al., 2015). Micro-magnetic stimulation, using a small coil, has been successful in the local activation of neurons *in vitro* (Bonmassar et al., 2012) and *in vivo* (Park et al., 2013), offering potential advantages over electric field and TMS due to enhanced spatial selectivity in neural control. Lee et al. described a new micro-coil design to activate cortical neurons and drive behavioral responses (Lee et al., 2016).

Computational work on the effects of magnetic stimulation on neurons provides valuable, quantitative insight and supporting evidence for the direct polarization of the cell membrane by the magnetic field. Using analytical methods, we calculated the induced membrane potential for a spherical cell (Ye et al., 2007) and internal organelles (Ye et al., 2010). Other works focused on axonal responses to the fields (Schnabel and Struijk, 2001) (Esselle and Stuchly, 1994, Ye et al., 2011), or axons located at the center of a nerve bundle (Nagarajan and Durand, 1995). Numerical simulation allows for the insertion of ion channels into modeled cells, which provide a close-to real morphological representation of the cell. Using this method, it was found that magnetic stimulation depolarized the soma of central nervous system neurons, followed by initiation of an action potential in the initial segment of the axon (Pashut et al., 2014). Goodwin and Butson modeled cortical neurons subject to external TMS, and found that the sites of neural activation are coil orientation dependent (Goodwin and Butson, 2015). This finding is consistent with previous animal studies, which indicated that the effects of magnetic field on synaptic activity are influenced by spatial parameters (Varro et al., 2009).

(B) Control of neuronal activity and network excitability via alteration of ion channels

Change in neuronal excitability is associated with the modulation of ionic channels under magnetic stimulation. Specifically, modifications in the electrophysiological properties of Na⁺- channels, A-type K⁺ channels, and Ca²⁺ channels are associated with altered neural excitability in rTMS (Tan et al., 2013). Computational simulation suggests that magnetic stimulation could adjust sodium channel currents and field excitatory postsynaptic potentials in rat hippocampal CA1 neurons (Zheng et al., 2017).

Modification in ionic homogeneity could be an interesting strategy for seizure blockage through the electromagnetic field. In support of this notion, it was found that high frequency (140 Hz) electric stimulation of hippocampal slices induced an increase in the extracellular potassium concentration and blocked neuronal depolarization (Lian et al., 2003). Bikson et al. (2001) found that during high frequency stimulation (HFS), the increase in extracellular potassium concentration ($[K^+]_e$) coincided with suppressed epileptiform activity. Likewise, HFS or elevated K⁺ depresses neuronal activity in the rat entopeduncular nucleus (Shin et al., 2007). It is hypothesized that direct magnetic stimulation can suppress seizures via an increase in $[K^+]_e$, leading to an inactivation of voltage-dependent ion channels and depolarization blockade.

(C) Control of neural activity via alteration in synaptic transmission

Magnetic field could alter synaptic transmission via its induced electric field, including long- term and short-term modulation. Magnetic field generated by a circular coil (50Hz, 100 mT) increased long term potential (LTP) induction in the hippocampal area of rats (Komaki et al., 2014). In addition, 100 Hz pulsed sinusoidal magnetic field can also modulate LTP in the hippocampus (Dong et al., 2018), likely due to a NMDAR-dependent mechanism (Tokay et al., 2009).

Research on the effects of magnetic field on short term synaptic plasticity is rare. However, one can make speculations from works of electric field stimulation. Short-term depression of synaptic transmission was observed during high frequency electric stimulation in the globus pallidus in rats (Rav-Acha et al., 2005) and in primates (Erez et al., 2009). Depression of synaptic transmission by HFS could be due to the fact that HFS-induced release of inhibitory GABA molecules is more prominent than the excitatory neurotransmitter (Feuerstein et al., 2011). Alternatively, it could be due to axonal and/or synaptic failure, which suppress the synaptic transfer of firing rate oscillations, synchrony, and rate-coded information during high frequency DBS (Rosenbaum et al., 2014). DBS-induced short term depression is a major therapeutic mechanism of DBS for Parkinson's disease. It was found that stimulation can decouple pre- and post-synaptic spiking patterns and suppress pallidal β oscillations in Parkinson's patients (Rosenbaum et al., 2014).

(D) Interruption of ephaptic coupling

Neurons affect each other via local electric fields, a phenomenon called ephaptic coupling. The functional significance of ephaptic coupling was largely ignored until the 1960s, when an inhibitory function of mauthner cells in goldfish was discovered (Furukawa and Furshpan, 1963). Via ephaptic interaction, these cells control 40-80 interneurons (Faber and Korn, 1983). Ephaptic interactions play a critical role in non-synaptic epileptogenesis (Haas and Jefferys, 1984, Richardson and O'Reilly, 1995). Communication among cells via ephaptic coupling could facilitate synchronized activity, epileptic-like neuronal bursting (Dudek et al., 1998), and neuron-glia communication (Amzica and Steriade, 2000). As such, interruption of ephaptic interaction between neurons, using external electric stimulation, has been proposed as a key neuronal mechanism for seizure suppression (Ghai et al., 2000). Similarly, by interrupting ephaptic coupling mechanisms in epileptogenesis, magnetic field could also de-synchronize neuronal firing and suppress seizure via its induced electric field.

(E) Other mechanisms

The impact of magnetic field is not only limited to neurons and their activation/deactivation. Magnetic fields can also enhance adult neural stem and progenitor cell proliferation (Cullen and Young, 2016). In addition, magnetic fields may effect microglia, which can modulate normal neuronal activity. Low intensity, high frequency rTMS following ischemic injury or demyelination activates microglia (Fang et al., 2010, Raus et al., 2013). On the other hand, high intensity, high frequency rTMS applied to injured spinal cord decreased microglial activation (Kim et al., 2013). In another study, high intensity, low frequency rTMS did not significantly change microglial number in the motor cortex or hippocampus of healthy rats (Liebetanz et al., 2003). The various results and limited number of studies assessing the effects of TMS on microglia suggest the need for further research in this area.

6. Translational Considerations

Several practical strategies could potentially improve the transition into more clinically relevant contexts. It is premature to completely replace current pharmacological treatments with magnetic stimulation in seizure treatment. Rather, a combination of the two could be expected to yield more efficient seizure suppression. For example, bursts of high-frequency rTMS, together with lorazepam, suppresses seizures in a rat kainate status epilepticus model (Gersner et al., 2016a), with the combined methods more effective than rTMS alone. In another example,

static magnetic field (SMF) is more effective in decreasing audiogenic seizure severity when administrated with the anticonvulsant phenytoin (McLean et al., 2003) (McLean et al., 2008). The combined methods can potentially reduce the required dose of the anticonvulsant medicine (and, therefore, the likelihood of medication-induced side effects). Future research should address issues that could potentially improve the outcomes of the complementary therapies.

As a replacement for electric stimulation with invasive electrodes, magnetic stimulation must also overcome some of the limitations that occur with electric stimulation. For example, specific stimulation of certain cell types is not easily addressed. Nevertheless, delicate design of the coil can provide a desired electric field distribution that significantly improves specificity of the target. It is possible to construct customized coils to fit the needs of specific requirements and animal models (Tang et al., 2016). While conventional rTMS stimulators activate only superficial cortical areas, it is possible to reach deep epileptic foci, such as in temporal lobe epilepsy (TLE), by using specially designed H-coils (Gersner et al., 2016b). Furthermore, minicoils can be covered in biocompatible material and positioned inside the animals, closer to the distinct target (Lee et al., 2016).

The ultimate goal of epilepsy therapies is to control seizure while minimizing side effects. A closed-loop system that can automatically detect seizure activity, optimize stimulus input, and apply current to the coil would be an ideal system for precise, low cost, and efficient seizure control with magnetic field. Successful examples of closed-loop control systems for seizure control have emerged from transcranial electrical stimulation (TES) studies (Berenyi et al., 2012, Kozak and Berenyi, 2017). The closed-loop approach also generated promising results in optogenetic inhibition of epilepsy (Krook-Magnuson et al., 2013), in which EEG has been used to predict and trigger optogenetic inhibition of excitatory principal cells, or to activate a subpopulation of GABAergic cells. For rTMS control of seizure, EEG has been used to guide rTMS in a rat kainic acid (KA) epilepsy model. In this study, the idea of closed-loop control emerged but had not been fully developed. EEG was continuously viewed by an operator who identified each seizure onset (Rotenberg et al., 2008).

The above-mentioned, successful closed-loop examples in TES and optogenetic studies could shine some lights on closed-loop seizure control with magnetic fields. This control diagram could be constructed to form a therapeutic loop. First, seizure signal must be reliably measured through electrophysiological recording, such as electrocorticography (EcoG), electroencephalography (EEG), and single unit recording. Other physiological parameters can also be measured to improved seizure prediction, such as blood flow, blood oxygenation, metabolism (Schwartz, 2007, Zhao et al., 2011, Patel et al., 2013), and heart rate (Lockman et

al., 2011). Features of these recordings will then be extracted with computational tools. Previous work done in these areas could be readily adapted into an innovative closed-loop system for seizure control with magnetic field. The above mentioned novel coil design will ultimately provide more specific stimulation, which will be seamlessly integrated into a system that contains hardware and software design for accurate temporal prediction and stimulation.

Concluding remarks

Despite the promising clinical potential for magnetic treatment of epilepsy, significant progress is still necessary. This includes the advancement of magnetic design, optimization of stimulation protocols, and evaluation of the long-term safety of these approaches so that the technique can become more translatable for clinical use in humans. Basic laboratory research that focuses on the mechanisms of magnetic field stimulation at the molecular, cellular, and network levels are of great value in facilitating this translation. We are optimistic that these challenges are not insurmountable, and that magnetic stimulation can become a reliable, practical method for epilepsy treatment with the continued close collaboration of clinicians, neuroscientists, engineers, and regulators.

Acknowledgements

The authors thank Dr. Vincent C.F. Chen for engaging in valuable discussion pertaining to magnetic stimulation and neuromodulation.

Funding: This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Conflict of Interest: None

References

- Allen CH, Kluger BM, Buard I (2017) Safety of Transcranial Magnetic Stimulation in Children: A Systematic Review of the Literature. Pediatr Neurol 68:3-17.
- Amzica F, Steriade M (2000) Neuronal and glial membrane potentials during sleep and paroxysmal oscillations in the neocortex. The Journal of neuroscience : the official journal of the Society for Neuroscience 20:6648-6665.
- Angelone LM, Potthast A, Segonne F, Iwaki S, Belliveau JW, Bonmassar G (2004) Metallic electrodes and leads in simultaneous EEG-MRI: specific absorption rate (SAR) simulation studies. Bioelectromagnetics 25:285-295.
- Anninos P, Kotini A, Adamopoulos A, Tsagas N (2003) Magnelic stimulation can modulate seizures in epileptic patients. Brain Topogr 16:57-64.

- Anninos PA, Tsagas N, Jacobson JI, Kotini A (1999) The biological effects of magnetic stimulation in epileptic patients. Panminerva medica 41:207-215.
- Anninos PA, Tsagas N, Sandyk R, Derpapas K (1991) Magnetic stimulation in the treatment of partial seizures. Int J Neurosci 60:141-171.
- Auvichayapat N, Rotenberg A, Gersner R, Ngodklang S, Tiamkao S, Tassaneeyakul W, Auvichayapat P (2013) Transcranial direct current stimulation for treatment of refractory childhood focal epilepsy. Brain stimulation 6:696-700.
- Begley CE, Durgin TL (2015) The direct cost of epilepsy in the United States: A systematic review of estimates. Epilepsia 56:1376-1387.
- Berenyi A, Belluscio M, Mao D, Buzsaki G (2012) Closed-loop control of epilepsy by transcranial electrical stimulation. Science 337:735-737.
- Bjerknes S, Skogseid IM, Saehle T, Dietrichs E, Toft M (2014) Surgical site infections after deep brain stimulation surgery: frequency, characteristics and management in a 10-year period. PLoS One 9:e105288.
- Bonmassar G, Lee SW, Freeman DK, Polasek M, Fried SI, Gale JT (2012) Microscopic magnetic stimulation of neural tissue. Nature communications 3:921.
- Brasil-Neto JP, de Araujo DP, Teixeira WA, Araujo VP, Boechat-Barros R (2004) Experimental therapy of epilepsy with transcranial magnetic stimulation: lack of additional benefit with prolonged treatment. Arq Neuropsiquiatr 62:21-25.
- Canales A, Jia X, Froriep UP, Koppes RA, Tringides CM, Selvidge J, Lu C, Hou C, Wei L, Fink Y, Anikeeva P (2015) Multifunctional fibers for simultaneous optical, electrical and chemical interrogation of neural circuits in vivo. Nat Biotechnol 33:277-284.
- Chen SD, Wang YL, Liang SF, Shaw FZ (2016) Rapid Amygdala Kindling Causes Motor Seizure and Comorbidity of Anxiety- and Depression-Like Behaviors in Rats. Front Behav Neurosci 10:129.
- Chiken S, Nambu A (2014) Disrupting neuronal transmission: mechanism of DBS? Front Syst Neurosci 8:33.
- Coderre TJ, Katz J, Vaccarino AL, Melzack R (1993) Contribution of central neuroplasticity to pathological pain: review of clinical and experimental evidence. Pain 52:259-285.
- Cogan SF (2008) Neural stimulation and recording electrodes. Annual review of biomedical engineering 10:275-309.
- Cooper IS (1973) Effect of stimulation of posterior cerebellum on neurological disease. Lancet 1:1321.
- Cooper IS, Crighel E, Amin I (1973) Clinical and physiological effects of stimulation of the paleocerebellum in humans. Journal of the American Geriatrics Society 21:40-43.
- Cooper YA, Pianka ST, Alotaibi NM, Babayan D, Salavati B, Weil AG, Ibrahim GM, Wang AC, Fallah A (2018) Repetitive transcranial magnetic stimulation for the treatment of drug-resistant epilepsy: A systematic review and individual participant data meta-analysis of real-world evidence. Epilepsia Open 3:55-65.
- Cullen CL, Young KM (2016) How Does Transcranial Magnetic Stimulation Influence Glial Cells in the Central Nervous System? Front Neural Circuits 10:26.
- Davis TS, Parker RA, House PA, Bagley E, Wendelken S, Normann RA, Greger B (2012) Spatial and temporal characteristics of V1 microstimulation during chronic implantation of a microelectrode array in a behaving macaque. Journal of neural engineering 9:065003.
- Dayan E, Censor N, Buch ER, Sandrini M, Cohen LG (2013) Noninvasive brain stimulation: from physiology to network dynamics and back. Nat Neurosci 16:838-844.
- Derchansky M, Jahromi SS, Mamani M, Shin DS, Sik A, Carlen PL (2008) Transition to seizures in the isolated immature mouse hippocampus: a switch from dominant phasic inhibition to dominant phasic excitation. The Journal of physiology 586:477-494.

- Dong L, Zheng Y, Li ZY, Li G, Lin L (2018) Modulating effects of on-line low frequency electromagnetic fields on hippocampal long-term potentiation in young male Sprague-Dawley rat. J Neurosci Res.
- Doshi PK (2011) Long-term surgical and hardware-related complications of deep brain stimulation. Stereotact Funct Neurosurg 89:89-95.
- Dudek FE, Yasumura T, Rash JE (1998) 'Non-synaptic' mechanisms in seizures and epileptogenesis. Cell biology international 22:793-805.
- Epsztein J, Milh M, Bihi RI, Jorquera I, Ben-Ari Y, Represa A, Crepel V (2006) Ongoing epileptiform activity in the post-ischemic hippocampus is associated with a permanent shift of the excitatoryinhibitory synaptic balance in CA3 pyramidal neurons. The Journal of neuroscience : the official journal of the Society for Neuroscience 26:7082-7092.
- Erez Y, Czitron H, McCairn K, Belelovsky K, Bar-Gad I (2009) Short-term depression of synaptic transmission during stimulation in the globus pallidus of 1-methyl-4-phenyl-1,2,3,6tetrahydropyridine-treated primates. The Journal of neuroscience : the official journal of the Society for Neuroscience 29:7797-7802.
- Esmaeilpour K, Sheibani V, Shabani M, Mirnajafi-Zadeh J (2017) Effect of low frequency electrical stimulation on seizure-induced short- and long-term impairments in learning and memory in rats. Physiol Behav 168:112-121.
- Esselle KP, Stuchly MA (1994) Quasi-static electric field in a cylindrical volume conductor induced by external coils. IEEE transactions on bio-medical engineering 41:151-158.
- Faber DS, Korn H (1983) Field effects trigger post-anodal rebound excitation in vertebrate CNS. Nature 305:802-804.
- Faber J, Vladyka V (1983) Antiepileptic effect of electric stimulation of the locus coeruleus in man. Activitas nervosa superior 25:304-308.
- Fang ZY, Li Z, Xiong L, Huang J, Huang XL (2010) Magnetic stimulation influences injury-induced migration of white matter astrocytes. Electromagn Biol Med 29:113-121.
- Feuerstein TJ, Kammerer M, Lucking CH, Moser A (2011) Selective GABA release as a mechanistic basis of high-frequency stimulation used for the treatment of neuropsychiatric diseases. Naunyn Schmiedebergs Arch Pharmacol 384:1-20.
- Fiest KM, Sauro KM, Wiebe S, Patten SB, Kwon CS, Dykeman J, Pringsheim T, Lorenzetti DL, Jette N (2017) Prevalence and incidence of epilepsy: A systematic review and meta-analysis of international studies. Neurology 88:296-303.
- Fregni F, Boggio PS, Lima MC, Ferreira MJ, Wagner T, Rigonatti SP, Castro AW, Souza DR, Riberto M, Freedman SD, Nitsche MA, Pascual-Leone A (2006a) A sham-controlled, phase II trial of transcranial direct current stimulation for the treatment of central pain in traumatic spinal cord injury. Pain 122:197-209.
- Fregni F, Boggio PS, Mansur CG, Wagner T, Ferreira MJ, Lima MC, Rigonatti SP, Marcolin MA, Freedman SD, Nitsche MA, Pascual-Leone A (2005) Transcranial direct current stimulation of the unaffected hemisphere in stroke patients. Neuroreport 16:1551-1555.
- Fregni F, Otachi PT, Do Valle A, Boggio PS, Thut G, Rigonatti SP, Pascual-Leone A, Valente KD (2006b) A randomized clinical trial of repetitive transcranial magnetic stimulation in patients with refractory epilepsy. Ann Neurol 60:447-455.
- Fregni F, Thome-Souza S, Nitsche MA, Freedman SD, Valente KD, Pascual-Leone A (2006c) A controlled clinical trial of cathodal DC polarization in patients with refractory epilepsy. Epilepsia 47:335-342.
- Furukawa T, Furshpan EJ (1963) Two inhibitory mechanisms in the Mauthner neurons of goldfish. Journal of neurophysiology 26:140-176.
- Galvani A (1791) De viribus electricitatis in motu musculari commentaries. Opuscula 363-418.

- Gersner R, Dhamne SC, Zangen A, Pascual-Leone A, Rotenberg A (2016a) Bursts of high-frequency repetitive transcranial magnetic stimulation (rTMS), together with lorazepam, suppress seizures in a rat kainate status epilepticus model. Epilepsy & behavior : E&B 62:136-139.
- Gersner R, Oberman L, Sanchez MJ, Chiriboga N, Kaye HL, Pascual-Leone A, Libenson M, Roth Y, Zangen A, Rotenberg A (2016b) H-coil repetitive transcranial magnetic stimulation for treatment of temporal lobe epilepsy: A case report. Epilepsy Behav Case Rep 5:52-56.
- Ghai RS, Bikson M, Durand DM (2000) Effects of applied electric fields on low-calcium epileptiform activity in the CA1 region of rat hippocampal slices. Journal of neurophysiology 84:274-280.
- Gluckman BJ, Neel EJ, Netoff TI, Ditto WL, Spano ML, Schiff SJ (1996) Electric field suppression of epileptiform activity in hippocampal slices. J Neurophysiol 76:4202-4205.
- Golestanirad L, Gale JT, Manzoor NF, Park HJ, Glait L, Haer F, Kaltenbach JA, Bonmassar G (2018) Solenoidal Micromagnetic Stimulation Enables Activation of Axons With Specific Orientation. Front Physiol 9:724.
- Goodwin BD, Butson CR (2015) Subject-Specific Multiscale Modeling to Investigate Effects of Transcranial Magnetic Stimulation. Neuromodulation 18:694-704.
- Grill WM, Norman SE, Bellamkonda RV (2009) Implanted neural interfaces: biochallenges and engineered solutions. Annual review of biomedical engineering 11:1-24.
- Grossman N, Bono D, Dedic N, Kodandaramaiah SB, Rudenko A, Suk HJ, Cassara AM, Neufeld E, Kuster N, Tsai LH, Pascual-Leone A, Boyden ES (2017) Noninvasive Deep Brain Stimulation via Temporally Interfering Electric Fields. Cell 169:1029-1041 e1016.
- Guo Y, Duan W, Ma C, Jiang C, Xie Y, Hao H, Wang R, Li L (2015) Biocompatibility and magnetic resonance imaging characteristics of carbon nanotube yarn neural electrodes in a rat model. Biomed Eng Online 14:118.
- Haas HL, Jefferys JG (1984) Low-calcium field burst discharges of CA1 pyramidal neurones in rat hippocampal slices. The Journal of physiology 354:185-201.
- Hsu WY, Cheng CH, Lin MW, Shih YH, Liao KK, Lin YY (2011) Antiepileptic effects of low frequency repetitive transcranial magnetic stimulation: A meta-analysis. Epilepsy Res 96:231-240.
- Huberfeld G, Menendez de la Prida L, Pallud J, Cohen I, Le Van Quyen M, Adam C, Clemenceau S, Baulac M, Miles R (2011) Glutamatergic pre-ictal discharges emerge at the transition to seizure in human epilepsy. Nature neuroscience 14:627-634.
- Jiang CQ, Hao HW, Li LM (2013) Artifact properties of carbon nanotube yarn electrode in magnetic resonance imaging. J Neural Eng 10:026013.
- Jitkritsadakul O, Bhidayasiri R, Kalia SK, Hodaie M, Lozano AM, Fasano A (2017) Systematic review of hardware-related complications of Deep Brain Stimulation: Do new indications pose an increased risk? Brain Stimul 10:967-976.
- Josephson CB, Dykeman J, Fiest KM, Liu X, Sadler RM, Jette N, Wiebe S (2013) Systematic review and meta-analysis of standard vs selective temporal lobe epilepsy surgery. Neurology 80:1669-1676.
- Kaur H, Kumar B, Medhi B (2016) Antiepileptic drugs in development pipeline: A recent update. eNeurologicalSci 4:42-51.
- Kerrigan JF, Litt B, Fisher RS, Cranstoun S, French JA, Blum DE, Dichter M, Shetter A, Baltuch G, Jaggi J,
 Krone S, Brodie M, Rise M, Graves N (2004) Electrical stimulation of the anterior nucleus of the
 thalamus for the treatment of intractable epilepsy. Epilepsia 45:346-354.
- Keskil IS, Keskil ZA, Canseven AG, Seyhan N (2001) No effect of 50 Hz magnetic field observed in a pilot study on pentylenetetrazol-induced seizures and mortality in mice. Epilepsy research 44:27-32.
- Kim JY, Choi GS, Cho YW, Cho H, Hwang SJ, Ahn SH (2013) Attenuation of spinal cord injury-induced astroglial and microglial activation by repetitive transcranial magnetic stimulation in rats. J Korean Med Sci 28:295-299.

- Kim YT, Hitchcock RW, Bridge MJ, Tresco PA (2004) Chronic response of adult rat brain tissue to implants anchored to the skull. Biomaterials 25:2229-2237.
- Kinoshita M, Ikeda A, Begum T, Yamamoto J, Hitomi T, Shibasaki H (2005) Low-frequency repetitive transcranial magnetic stimulation for seizure suppression in patients with extratemporal lobe epilepsy-a pilot study. Seizure 14:387-392.
- Kistsen V, Evstigneev V, Dubovik B, Ulashchik V (2016) The Effects of Repetitive Transcranial Magnetic Stimulation on Picrotoxin-Induced Convulsions in Mice. Adv Clin Exp Med 25:317-325.
- Koivuniemi A, Wilks SJ, Woolley AJ, Otto KJ (2011) Multimodal, longitudinal assessment of intracortical microstimulation. Progress in brain research 194:131-144.
- Komaki A, Khalili A, Salehi I, Shahidi S, Sarihi A (2014) Effects of exposure to an extremely low frequency electromagnetic field on hippocampal long-term potentiation in rat. Brain Res 1564:1-8.
- Kozak G, Berenyi A (2017) Sustained efficacy of closed loop electrical stimulation for long-term treatment of absence epilepsy in rats. Scientific reports 7:6300.
- Krook-Magnuson E, Armstrong C, Oijala M, Soltesz I (2013) On-demand optogenetic control of spontaneous seizures in temporal lobe epilepsy. Nature communications 4:1376.
- Kwan P, Brodie MJ (2000) Early identification of refractory epilepsy. N Engl J Med 342:314-319.
- Lasztoczi B, Nyitrai G, Heja L, Kardos J (2009) Synchronization of GABAergic inputs to CA3 pyramidal cells precedes seizure-like event onset in juvenile rat hippocampal slices. J Neurophysiol 102:2538-2553.
- Laxer KD, Trinka E, Hirsch LJ, Cendes F, Langfitt J, Delanty N, Resnick T, Benbadis SR (2014) The consequences of refractory epilepsy and its treatment. Epilepsy & behavior : E&B 37:59-70.
- Lee SW, Fallegger F, Casse BD, Fried SI (2016) Implantable microcoils for intracortical magnetic stimulation. Sci Adv 2:e1600889.
- Lee SW, Fried SI (2014) The response of L5 pyramidal neurons of the PFC to magnetic stimulation from a micro-coil. Conference proceedings : Annual International Conference of the IEEE Engineering in Medicine and Biology Society IEEE Engineering in Medicine and Biology Society Annual Conference 2014:6125-6128.
- Lian J, Bikson M, Sciortino C, Stacey WC, Durand DM (2003) Local suppression of epileptiform activity by electrical stimulation in rat hippocampus in vitro. The Journal of physiology 547:427-434.
- Liebetanz D, Fauser S, Michaelis T, Czeh B, Watanabe T, Paulus W, Frahm J, Fuchs E (2003) Safety aspects of chronic low-frequency transcranial magnetic stimulation based on localized proton magnetic resonance spectroscopy and histology of the rat brain. J Psychiatr Res 37:277-286.
- Liu A, Pang T, Herman S, Pascual-Leone A, Rotenberg A (2013) Transcranial magnetic stimulation for refractory focal status epilepticus in the intensive care unit. Seizure 22:893-896.
- Liu B, Kim E, Meggo A, Gandhi S, Luo H, Kallakuri S, Xu Y, Zhang J (2017) Enhanced biocompatibility of neural probes by integrating microstructures and delivering anti-inflammatory agents via microfluidic channels. J Neural Eng 14:026008.
- Lockman J, Fisher RS, Olson DM (2011) Detection of seizure-like movements using a wrist accelerometer. Epilepsy & behavior : E&B 20:638-641.
- Lu H, Chestek CA, Shaw KM, Chiel HJ (2008) Selective extracellular stimulation of individual neurons in ganglia. J Neural Eng 5:287-309.
- Luan L, Wei X, Zhao Z, Siegel JJ, Potnis O, Tuppen CA, Lin S, Kazmi S, Fowler RA, Holloway S, Dunn AK, Chitwood RA, Xie C (2017) Ultraflexible nanoelectronic probes form reliable, glial scar-free neural integration. Sci Adv 3:e1601966.
- Maccabee PJ, Amassian VE, Cracco RQ, Eberle LP, Rudell AP (1991) Mechanisms of peripheral nervous system stimulation using the magnetic coil. Electroencephalogr Clin Neurophysiol Suppl 43:344-361.

- Maccabee PJ, Amassian VE, Eberle LP, Cracco RQ (1993) Magnetic coil stimulation of straight and bent amphibian and mammalian peripheral nerve in vitro: locus of excitation. The Journal of physiology 460:201-219.
- Matsumoto H, Ugawa Y (2017) Adverse events of tDCS and tACS: A review. Clinical neurophysiology practice 2:19-25.
- McIntyre CC, Grill WM, Sherman DL, Thakor NV (2004) Cellular effects of deep brain stimulation: modelbased analysis of activation and inhibition. J Neurophysiol 91:1457-1469.
- McLean MJ, Engstrom S, Holcomb RR, Sanchez D (2003) A static magnetic field modulates severity of audiogenic seizures and anticonvulsant effects of phenytoin in DBA/2 mice. Epilepsy research 55:105-116.
- McLean MJ, Engstrom S, Qinkun Z, Spankovich C, Polley DB (2008) Effects of a static magnetic field on audiogenic seizures in black Swiss mice. Epilepsy research 80:119-131.
- Menkes DL, Gruenthal M (2000) Slow-frequency repetitive transcranial magnetic stimulation in a patient with focal cortical dysplasia. Epilepsia 41:240-242.
- Moradi Chameh H, Janahmadi M, Semnanian S, Shojaei A, Mirnajafi-Zadeh J (2015) Effect of low frequency repetitive transcranial magnetic stimulation on kindling-induced changes in electrophysiological properties of rat CA1 pyramidal neurons. Brain research 1606:34-43.
- Morrell MJ, Group RNSSiES (2011) Responsive cortical stimulation for the treatment of medically intractable partial epilepsy. Neurology 77:1295-1304.
- Morris GL, 3rd, Mueller WM (1999) Long-term treatment with vagus nerve stimulation in patients with refractory epilepsy. The Vagus Nerve Stimulation Study Group E01-E05. Neurology 53:1731-1735.
- Nagarajan SS, Durand DM (1995) Analysis of magnetic stimulation of a concentric axon in a nerve bundle. IEEE Trans Biomed Eng 42:926-933.
- Nitsche MA, Paulus W (2000) Excitability changes induced in the human motor cortex by weak transcranial direct current stimulation. The Journal of physiology 527 Pt 3:633-639.
- Nowak LG, Bullier J (1998) Axons, but not cell bodies, are activated by electrical stimulation in cortical gray matter. I. Evidence from chronaxie measurements. Exp Brain Res 118:477-488.
- Oliveira T, Francisco AN, Demartini ZJ, Stebel SL (2017) The role of vagus nerve stimulation in refractory epilepsy. Arq Neuropsiquiatr 75:657-666.
- Park HJ, Bonmassar G, Kaltenbach JA, Machado AG, Manzoor NF, Gale JT (2013) Activation of the central nervous system induced by micro-magnetic stimulation. Nature communications 4:2463.
- Parkin BL, Ekhtiari H, Walsh VF (2015) Non-invasive Human Brain Stimulation in Cognitive Neuroscience: A Primer. Neuron 87:932-945.
- Pashut T, Magidov D, Ben-Porat H, Wolfus S, Friedman A, Perel E, Lavidor M, Bar-Gad I, Yeshurun Y, Korngreen A (2014) Patch-clamp recordings of rat neurons from acute brain slices of the somatosensory cortex during magnetic stimulation. Front Cell Neurosci 8:145.
- Patel KS, Zhao M, Ma H, Schwartz TH (2013) Imaging preictal hemodynamic changes in neocortical epilepsy. Neurosurgical focus 34:E10.
- Pereira LS, Muller VT, da Mota Gomes M, Rotenberg A, Fregni F (2016) Safety of repetitive transcranial magnetic stimulation in patients with epilepsy: A systematic review. Epilepsy & behavior : E&B 57:167-176.
- Polikov VS, Tresco PA, Reichert WM (2005) Response of brain tissue to chronically implanted neural electrodes. Journal of neuroscience methods 148:1-18.
- Potschka H, Thun-Battersby S, Loscher W (1998) Effect of low-intensity 50-Hz magnetic fields on kindling acquisition and fully kindled seizures in rats. Brain Res 809:269-276.

- Raus S, Selakovic V, Manojlovic-Stojanoski M, Radenovic L, Prolic Z, Janac B (2013) Response of hippocampal neurons and glial cells to alternating magnetic field in gerbils submitted to global cerebral ischemia. Neurotox Res 23:79-91.
- Rav-Acha M, Sagiv N, Segev I, Bergman H, Yarom Y (2005) Dynamic and spatial features of the inhibitory pallidal GABAergic synapses. Neuroscience 135:791-802.
- Rezai AR, Phillips M, Baker KB, Sharan AD, Nyenhuis J, Tkach J, Henderson J, Shellock FG (2004) Neurostimulation system used for deep brain stimulation (DBS): MR safety issues and implications of failing to follow safety recommendations. Investigative radiology 39:300-303.
- Richardson TL, O'Reilly CN (1995) Epileptiform activity in the dentate gyrus during low-calcium perfusion and exposure to transient electric fields. J Neurophysiol 74:388-399.
- Rodrigo D, Acin P, Bermejo P (2017) Occipital Nerve Stimulation for Refractory Chronic Migraine: Results of a Long-Term Prospective Study. Pain Physician 20:E151-E159.
- Rosenbaum R, Zimnik A, Zheng F, Turner RS, Alzheimer C, Doiron B, Rubin JE (2014) Axonal and synaptic failure suppress the transfer of firing rate oscillations, synchrony and information during high frequency deep brain stimulation. Neurobiol Dis 62:86-99.
- Rossi S, Hallett M, Rossini PM, Pascual-Leone A, Safety of TMSCG (2009) Safety, ethical considerations, and application guidelines for the use of transcranial magnetic stimulation in clinical practice and research. Clinical neurophysiology : official journal of the International Federation of Clinical Neurophysiology 120:2008-2039.
- Rotenberg A, Muller P, Birnbaum D, Harrington M, Riviello JJ, Pascual-Leone A, Jensen FE (2008) Seizure suppression by EEG-guided repetitive transcranial magnetic stimulation in the rat. Clinical neurophysiology : official journal of the International Federation of Clinical Neurophysiology 119:2697-2702.
- Salanova V, Witt T, Worth R, Henry TR, Gross RE, Nazzaro JM, Labar D, Sperling MR, Sharan A, Sandok E, Handforth A, Stern JM, Chung S, Henderson JM, French J, Baltuch G, Rosenfeld WE, Garcia P, Barbaro NM, Fountain NB, Elias WJ, Goodman RR, Pollard JR, Troster AI, Irwin CP, Lambrecht K, Graves N, Fisher R, Group SS (2015) Long-term efficacy and safety of thalamic stimulation for drug-resistant partial epilepsy. Neurology 84:1017-1025.
- San-Juan D, Espinoza Lopez DA, Vazquez Gregorio R, Trenado C, Fernandez-Gonzalez Aragon M, Morales-Quezada L, Hernandez Ruiz A, Hernandez-Gonzalez F, Alcaraz-Guzman A, Anschel DJ, Fregni F (2017) Transcranial Direct Current Stimulation in Mesial Temporal Lobe Epilepsy and Hippocampal Sclerosis. Brain stimulation 10:28-35.
- San-Juan D, Sarmiento CI, Gonzalez KM, Orenday Barraza JM (2018) Successful Treatment of a Drug-Resistant Epilepsy by Long-term Transcranial Direct Current Stimulation: A Case Report. Frontiers in neurology 9:65.
- Sandyk R, Anninos PA (1992a) Attenuation of epilepsy with application of external magnetic fields: a case report. Int J Neurosci 66:75-85.
- Sandyk R, Anninos PA (1992b) Magnetic fields alter the circadian periodicity of seizures. Int J Neurosci 63:265-274.
- Saxena T, Karumbaiah L, Gaupp EA, Patkar R, Patil K, Betancur M, Stanley GB, Bellamkonda RV (2013) The impact of chronic blood-brain barrier breach on intracortical electrode function. Biomaterials 34:4703-4713.
- Schnabel V, Struijk JJ (2001) Evaluation of the cable model for electrical stimulation of unmyelinated nerve fibers. IEEE Trans Biomed Eng 48:1027-1033.
- Schwartz TH (2007) Neurovascular coupling and epilepsy: hemodynamic markers for localizing and predicting seizure onset. Epilepsy currents 7:91-94.
- Selimbeyoglu A, Parvizi J (2010) Electrical stimulation of the human brain: perceptual and behavioral phenomena reported in the old and new literature. Frontiers in human neuroscience 4:46.

- Shin DS, Carlen PL (2008) Enhanced Ih depresses rat entopeduncular nucleus neuronal activity from high-frequency stimulation or raised Ke+. J Neurophysiol 99:2203-2219.
- Shin DS, Samoilova M, Cotic M, Zhang L, Brotchie JM, Carlen PL (2007) High frequency stimulation or elevated K+ depresses neuronal activity in the rat entopeduncular nucleus. Neuroscience 149:68-86.
- Shojaei A, Semnanian S, Janahmadi M, Moradi-Chameh H, Firoozabadi SM, Mirnajafi-Zadeh J (2014) Repeated transcranial magnetic stimulation prevents kindling-induced changes in electrophysiological properties of rat hippocampal CA1 pyramidal neurons. Neuroscience 280:181-192.
- Sun FT, Morrell MJ (2014) The RNS System: responsive cortical stimulation for the treatment of refractory partial epilepsy. Expert Rev Med Devices 11:563-572.
- Sun W, Mao W, Meng X, Wang D, Qiao L, Tao W, Li L, Jia X, Han C, Fu M, Tong X, Wu X, Wang Y (2012) Low-frequency repetitive transcranial magnetic stimulation for the treatment of refractory partial epilepsy: a controlled clinical study. Epilepsia 53:1782-1789.
- Sung JH, Jeong JH, Kim JS, Choi TS, Park JH, Kang HY, Kim YS, Kim DS, Sohn UD (2003) The influences of extremely low frequency magnetic fields on drug-induced convulsion in mouse. Arch Pharm Res 26:487-492.
- Sutton AC, Yu W, Calos ME, Mueller LE, Berk M, Shim J, Molho ES, Brotchie JM, Carlen PL, Shin DS (2013) Elevated potassium provides an ionic mechanism for deep brain stimulation in the hemiparkinsonian rat. Eur J Neurosci 37:231-241.
- Tan T, Xie J, Tong Z, Liu T, Chen X, Tian X (2013) Repetitive transcranial magnetic stimulation increases excitability of hippocampal CA1 pyramidal neurons. Brain Res 1520:23-35.
- Tang AD, Lowe AS, Garrett AR, Woodward R, Bennett W, Canty AJ, Garry MI, Hinder MR, Summers JJ, Gersner R, Rotenberg A, Thickbroom G, Walton J, Rodger J (2016) Construction and Evaluation of Rodent-Specific rTMS Coils. Front Neural Circuits 10:47.
- Tokay T, Holl N, Kirschstein T, Zschorlich V, Kohling R (2009) High-frequency magnetic stimulation induces long-term potentiation in rat hippocampal slices. Neurosci Lett 461:150-154.
- Varga ET, Terney D, Atkins MD, Nikanorova M, Jeppesen DS, Uldall P, Hjalgrim H, Beniczky S (2011) Transcranial direct current stimulation in refractory continuous spikes and waves during slow sleep: a controlled study. Epilepsy research 97:142-145.
- Varro P, Szemerszky R, Bardos G, Vilagi I (2009) Changes in synaptic efficacy and seizure susceptibility in rat brain slices following extremely low-frequency electromagnetic field exposure. Bioelectromagnetics 30:631-640.
- Velasco M, Velasco F, Velasco AL, Jimenez F, Brito F, Marquez I (2000) Acute and chronic electrical stimulation of the centromedian thalamic nucleus: modulation of reticulo-cortical systems and predictor factors for generalized seizure control. Archives of medical research 31:304-315.
- Voroslakos M, Takeuchi Y, Brinyiczki K, Zombori T, Oliva A, Fernandez-Ruiz A, Kozak G, Kincses ZT, Ivanyi B, Buzsaki G, Berenyi A (2018) Direct effects of transcranial electric stimulation on brain circuits in rats and humans. Nature communications 9:483.
- Ye H, Cotic M, Carlen PL (2007) Transmembrane potential induced in a spherical cell model under lowfrequency magnetic stimulation. Journal of neural engineering 4:283-293.
- Ye H, Cotic M, Fehlings MG, Carlen PL (2011) Transmembrane potential generated by a magnetically induced transverse electric field in a cylindrical axonal model. Medical & biological engineering & computing 49:107-119.
- Ye H, Cotic M, Kang EE, Fehlings MG, Carlen PL (2010) Transmembrane potential induced on the internal organelle by a time-varying magnetic field: a model study. J Neuroeng Rehabil 7:12.

- Ye H, Steiger A (2015) Neuron matters: electric activation of neuronal tissue is dependent on the interaction between the neuron and the electric field. Journal of neuroengineering and rehabilitation 12:65.
- Yook SW, Park SH, Seo JH, Kim SJ, Ko MH (2011) Suppression of seizure by cathodal transcranial direct current stimulation in an epileptic patient a case report. Annals of rehabilitation medicine 35:579-582.
- Yuan H, Silberstein SD (2016) Vagus Nerve and Vagus Nerve Stimulation, a Comprehensive Review: Part III. Headache 56:479-490.
- Zaeimbashi M, Wang Z, Lee SW, Cash S, Fried S, Sun N (2018) Micro-solenoid inductors with magnetic core for neural stimulation. Conf Proc IEEE Eng Med Biol Soc 2018:2230-2233.
- Zhao M, Nguyen J, Ma H, Nishimura N, Schaffer CB, Schwartz TH (2011) Preictal and ictal neurovascular and metabolic coupling surrounding a seizure focus. The Journal of neuroscience : the official journal of the Society for Neuroscience 31:13292-13300.
- Zheng Y, Ma W, Dong L, Dou JR, Gao Y, Xue J (2017) Influence of the on-line ELF-EMF stimulation on the electrophysiological properties of the rat hippocampal CA1 neurons in vitro. Rev Sci Instrum 88:105106.
- Ziburkus J, Cressman JR, Barreto E, Schiff SJ (2006) Interneuron and pyramidal cell interplay during in vitro seizure-like events. Journal of neurophysiology 95:3948-3954.