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87 Abstract

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89	Over the last decade, there have been substantial advances in our understanding about how
90	viral infections regulate asthma (Table 1). Important lessons have been learned from birth
91	cohort studies examining viral infections and subsequent asthma, understanding the
92	relationships between host genetics and viral infections, the contributions of respiratory viral
93	infections to patterns of immune development, the impact of environmental exposure on severity
94	of viral infections, and how the viral genome influences host immune responses to viral
95	infections. Further, there has been major progress in our knowledge about how bacteria
96	regulate host immune responses in asthma pathogenesis. In this article, we also examine the
97	dynamics of respiratory tract bacterial colonization during viral upper respiratory tract infection,
98	in addition to the relationship of the gut and respiratory microbiomes with respiratory viral
99	infections. Finally, we focus on potential interventions that could decrease virus-induced
100	wheezing and asthma. There are emerging therapeutic options to decrease severity of
101	wheezing exacerbations caused by respiratory viral infections. Primary prevention is a major
102	goal and a strategy toward this end is considered.
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104	Key words: virus, asthma, genetics, immune, microbiome
105	
106	Abbreviations:
107	Respiratory syncytial virus (RSV)
108	Rhinovirus (RV)
109	Airway responsiveness (AR)
110	Fusion (F)
111	Type 2 (Th2)
112	Cadherin-related family member 3 (CDHR3)
113	Genome wide association study (GWAS)
114	Environmental tobacco smoke (ETS)
115	Upper respiratory tract infection (URI)
116	Short chain fatty acids (SCFA)
117	Forced expiratory volume in 0.5 s (FEV _{0.5})
118	Wheezing lower respiratory tract illness (WLRI)
119	Inhaled corticosteroids (ICS)
120	Long acting beta agonists (LABA)
121	Lower respiratory tract illness (LRTI)
122	Plasmacvtoid dendritic cells (pDC)

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123 124 Introduction

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126 Over the last decade, there have been substantial advances in our understanding about how 127 viral infections regulate asthma (Table 1). Important lessons have been learned from birth 128 cohort studies examining viral infections and subsequent asthma, understanding the 129 relationships between host genetics and viral infections, the contributions of respiratory viral 130 infections to patterns of immune development, the impact of environmental exposure on severity 131 of viral infections, and how the viral genome influences host immune responses to viral 132 infections. Further, there has been major progress in our knowledge about how bacteria 133 regulate host immune responses in asthma pathogenesis. In this article, we also examine the 134 dynamics of respiratory tract bacterial colonization during viral upper respiratory tract infection, 135 in addition to the relationship of the gut and respiratory microbiomes with respiratory viral 136 infections. Finally, we focus on potential interventions that could decrease virus-induced 137 wheezing and asthma. There are emerging therapeutic options to decrease severity of 138 wheezing exacerbations caused by respiratory viral infections. Primary prevention is a major 139 goal and a strategy toward this end is considered.

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141 The viral genome and how it influences host immune responses to viral infections

142 Respiratory syncytial virus (RSV) and rhinovirus (RV) are important causes of wheezing 143 in early life and wheezing illness with these viruses have been associated with increased asthma risk later in childhood. At age 6, there is an increased risk of asthma if children had 144 wheezing illness with RSV (odds ratio 2.6), RV (odds ratio 9.8), or both RSV and RV (odds ratio 145 146 10.0) in the first 3 years of life. RSV is a negative-sense, single stranded RNA virus that is a 147 member of the Paramyxoviridae family and is the leading cause of hospitalization each year in the United States in children under 1 year of age.¹ There are three species of RV in the 148 149 enterovirus genus, and all are positive-sense, single-stranded RNA viruses that have protein 150 capsids. RV are the most frequently detected viruses in wheezing children over the age of 1 151 year, and from children and adults with acute exacerbations of asthma.

152 The clinical manifestations of a viral infection in the respiratory tract result from a complex interplay of the host, environment, and virus. To make comparisons between different 153 immune responses elicited by diverse viruses, host and the environmental conditions must be 154 155 held constant in order to prevent the introduction of confounding factors. This requires artificial 156 conditions, such as the use of human cell lines for in vitro infection studies, the infection of 157 genetically identical animals, such as mice, housed in the same environment, and the use of a 158 standard viral inoculum. Determining the effect of specific genes within a virus requires that all 159 other viral genes are identical. Such studies have begun but are still relatively new.

Experiments in models of RSV genomes have provided important insights into how the viral genome influences host immune responses to infection. Three RSV strains commonly used in pathogenesis studies are A2, line 19, and Long. RSV A2 infection in BALB/c mice resulted in a predominant IFN-γ immune response, no production of the Th2 cytokine IL-13 in the lung, an absence of airway mucus, and no airway responsiveness (AR) to methacholine.² Infection with RSV Long similarly did not result in host IL-13 production in the lung nor was there airway mucus.³ However, line 19 infection in genetically identical mice in the same environment

167 caused the host immune response to produce IL-13, decreased IFN-y compared to A2 infection, airway mucus, and heightened AR.² Sequencing of the A2, line 19, RSV Long strains revealed 168 six amino acid differences between line 19 and the A2 and Long strains, of which 5 amino acid 169 differences were in the fusion (F) protein.³ To determine the contribution of the F gene of each 170 171 virus to disease pathogenesis, a reverse genetics approach was undertaken by creating chimeric viruses whereby an A2 virus was manipulated to replace the A2 F gene with either the 172 F gene of either line 19 or Long. Infection with the chimeric virus containing the line 19 F gene 173 caused decreased host IFN-α lung levels, higher viral load in the lungs, greater lung IL-13 174 175 protein, augmented airway mucus, and increased AR, compared to the chimeric viruses containing either A2 or Long F proteins.³ Therefore, this reverse genetic approach provided the 176 opportunity to not only discover which genes in RSV line 19 strain were responsible for the lung 177 178 IL-13, airway mucus, and AR, but also the identification of the specific amino acids that caused 179 airway remodeling. These techniques not only provide the knowledge of unique components of the viral genome that contribute to specific pathogenic features but may also assist in vaccine 180 181 and therapeutic strategies aimed at the proteins responsible for specific disease characteristics. 182 Future perspectives: To date, there have not been studies that reveal a relationship between 183 RSV genotypes and the presence of wheezing in hospitalized children with bronchiolitis or 184 bronchopneumonia; however, this may be a function of the lack of application of technology to 185 sequence strains because of cost. Studies relating viral genetics to severity of illness in mice 186 have demonstrated the intricate interactions between viral genome, viral proteins, host cell 187 function and metabolism and immune response. Developing a greater understanding of this 188 chain of events could highlight several therapeutic opportunities, including identification of high-189 priority pathogens, inhibition of viral or host proteins that are critical for replication, and 190 strategies to inhibit virus-induced skewing of immune responses that favors viral replication over 191 host defense.

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193 Host genetics and viral respiratory infections

194 A number of studies have begun to shed light on the relationships among host genetics, 195 viral infections and acute and long-term respiratory outcomes. Candidate gene approaches 196 have been utilized to identify associations between genetic polymorphisms and viral respiratory 197 illness outcomes. Polymorphisms in several antiviral and innate immune genes have been 198 linked to susceptibility to respiratory viruses, infection severity, and virus-induced asthma 199 exacerbations, and have been replicated across multiple cohorts (Table 2). These genes include STAT4, JAK2, MX1, VDR, DDX58, and EIF2AK2.⁴ Additionally, whole exome 200 201 sequencing has been utilized to identify rare variants in innate immune responses linked to 202 severe respiratory viral infections. Autosomal recessive IRF7 deficiency has been observed in one patient in association with severe influenza infection and acts through impairment of 203 204 interferon amplification.⁵ Dominant negative loss-of-function variants in *IFIH1*, critical to viral 205 RNA sensing, have been shown to be a risk for intensive care unit hospitalization due to viral infections in previously healthy children.⁶ 206

207 A number of polymorphisms have been specifically associated with increased severity of 208 illnesses associated with hospitalization from RSV infection. A candidate gene approach 209 identified SNPs in the innate immune genes VDR, IFNA5, and NOS2 as risk factors for RSV bronchiolitis.⁷ In order to further elucidate associations of host genetics with RSV illness severity 210 211 and asthma risk, a recent review examined overlap amongst genes associated with both 212 outcomes. This approach identified a number of genes involved with both innate immunity and 213 type 2 (Th2) inflammatory responses (ADAM33, IL4R, CD14, TNF, IL13 and IL1RL1) that are highly relevant to these outcomes.⁸ 214

The most replicated association between host genetics and asthma risk is the 17q21 locus. In two birth cohort studies, variants in this locus, including ORMDL3 and GSDMB, were also associated with increased risk of wheezing with RV infections in early life.⁹ Interestingly, these variants were only associated with increased risk of subsequent asthma in children who

developed RV wheezing in the first 3 years of life. In contrast, early life RSV wheezing was not
linked to 17q21 variants in these cohorts. In addition, farm exposures¹⁰ and pets¹¹ in the home
lessen the risk of asthma for children with high-risk 17q21 genotypes. In each case, the genetic
risk associated with 17q21 was buffered by protective environmental exposures.

223 RV virulence varies by species; RV-A and RV-C are more likely to cause illnesses, wheezing and lower respiratory tract infection compared to RV-B,¹² which has a slower rate of 224 replication and induces muted cytokine and chemokine responses.¹³ Whether there are 225 226 individual types within RV species that are more virulent is unknown, and difficult to study given the genetic diversity of these viruses and high mutation rate. A functional polymorphism in 227 228 cadherin-related family member 3 (CDHR3) has been associated by genome wide association study (GWAS) with early childhood asthma and severe wheezing episodes.¹⁴ Interestingly, 229 230 CDHR3 is a receptor that enables binding and replication of RV-C, suggesting that this link between CDHR3 and asthma risk may be mediated by RV-C infections.¹⁵ In support of this 231 hypothesis, children with the risk polymorphism in CDHR3 were recently found to have greater 232 risk of RV-C illnesses, but not illnesses associated with other viruses.¹⁶ 233 234 Future perspectives: These genetic associations among respiratory virus susceptibility, infection severity and subsequent asthma risk may prove to be important to risk stratify populations, and 235 236 potentially provide new therapeutic targets for reducing illness severity and subsequent risk. 237 Further, unbiased approaches have been employed recently to identify pathways of gene 238 expression in the upper airway that differentiate a viral cold that resolves from one that leads to 239 an asthma exacerbation.¹ Efforts are ongoing to understand how these gene expression 240 patterns are regulated in hopes of identifying new personalized therapeutic strategies to prevent asthma exacerbation. The integration of multiple "omics" approaches holds promise to provide 241

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244 Environmental factors affecting the inception and severity of asthma exacerbations

the ability to unravel these complex relationships.

245 The exposome, defined as the measure of all exposures that influence the health of an individual, is an important determinant of asthma risk during the lifespan of an individual.¹⁷ Early 246 247 exposures can set in motion pathways that will ultimately define illnesses and symptom 248 exacerbations, which is especially true when considering the ontogeny of asthma. From birth through school years, children are frequently exposed to a variety of respiratory pathogens, 249 allergens, microbes and airway irritants. The pathogenic or beneficial effects of these 250 251 exposures and their interactions remain the focus of research to develop new interventions and 252 preventive therapies.

253 Most of the initial research devoted to the ontogeny of asthma focused on RSV infections which are frequently detected by culture and tests for RSV antigen in nasal washes from 254 255 wheezing infants during the mid-winter months. Studies in the past reported that flares of 256 wheezing caused by RSV leading to hospitalization during infancy increased the risk for developing asthma and allergy.^{18, 19} However, recent studies indicate that the more severe 257 episodes (i.e., those requiring hospitalization) of infantile wheezing caused by RSV increase the 258 259 risk for subsequent wheeze in infants and toddlers, but it is less certain whether RSV-induced wheeze influences the development of atopy or asthma as children grow older.²⁰ 260

In contrast, flares of wheezing caused by RV are more strongly linked to persistent wheezing and the development of asthma, especially in children who are sensitized to allergens at an early age.²¹⁻²³ In keeping with this, the dominant risk factors for asthma attacks that require hospital care among children after 3 years of age is the combination of allergic airway inflammation and RV infection.²⁴⁻²⁶ As a result, several host factors should be considered in efforts to treat asthma exacerbations more effectively and to reduce the risk for asthma development, For example:

There may be phenotypes of asthmatic children who would benefit from development of
 vaccines to RV or RSV. For example, genetic variations at the 17q21 locus and a coding

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SNP in *CDHR3* (the receptor for RV-C genotypes) increase the risk for wheezing with
 RV during childhood.^{9, 14}

272 2) There is current interest in whether the administration of a biologic such as omalizumab
273 (anti-IgE antibody) during early childhood will have a disease modifying effect after this
274 intervention is discontinued (i.e., the Preventing Asthma in High Risk Kids (PARK) trial;
275 NCT02570984).

3) There is evidence that the asthmatic airway, especially epithelial cells and innate 276 lymphoid cells, has a Type 2 bias with enhanced production of TSLP, IL-13, and IL-25 in 277 278 parallel with decreased type I and III IFN responses that are needed for effective antiviral killing and clearance.²⁷⁻³⁴ This bias may increase the susceptibility of allergic 279 asthmatics to RV infections. Once infected, however, in vivo studies have shown that 280 281 during RV infections viral loads and clearance are similar among children and young adults with asthma compared to non-allergic individuals without asthma.³¹ At present, 282 mechanisms to explain these differences are poorly understood. A better understanding 283 is likely to come from research focused on the cascade of early, innate cellular and 284 285 molecular events that follow RV infection of epithelial cells.

286 Future perspectives: Airway inflammation caused by recurrent infections (predominantly with 287 RV) in the allergic host will continue to be the focus of research designed to develop new 288 therapies to help children and young adults with asthma. Whether treatments targeting allergic 289 inflammation will be sufficient to reduce the frequency and severity of exacerbations (e.g., using 290 new monoclonal antibody-based biologics), or whether additional therapeutics will be needed to 291 decrease the frequency of RV infections, or enhance viral killing, remains to be determined. 292 Looking to the future, the evaluation of other interventions such as the administration of 293 antibiotics to treat secondary bacterial pathogens, or azithromycin to reduce wheezing following 294 virus infection also deserve further study, along with investigations to determine whether the 295 administration of vitamin D, probiotics, and dietary modifications (e.g., fish oil) will have benefits.

296 In contrast, the adverse effects of airway irritants such as environmental tobacco smoke (ETS) 297 and air pollution (e.g., diesel fuel) on the severity and persistence of RV-induced asthma remain 298 poorly understood.

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Effects of respiratory viral infections on patterns of immune development

301 Acute wheezing illnesses with respiratory tract viruses in infancy and early childhood 302 represent an important risk factor for childhood recurrent wheezing and later asthma 303 development. This link is particularly well-established with RV and RSV, suggesting that these 304 viruses may have a causative role, and significant research is directed towards understanding 305 how these viruses can alter immune development to contribute to asthma pathogenesis. That 306 said, causation remains unproven and asthma prevention strategies targeting viral illnesses do 307 not currently exist.

308 RV-associated wheezing, in particular, is associated with a higher asthma risk than other viruses; this has been consistently demonstrated across multiple studies.^{21, 23, 35-38} Many of 309 310 these studies have linked RV-induced wheezing with other asthma risk factors, in particular 311 markers of atopy including allergic sensitization, increased eosinophils, and atopic eczema, 312 suggesting possible additive or synergistic effects in increasing asthma risk. Experimental 313 models have demonstrated alterations in type-2 immune responses to RV that may account for 314 this risk (Figure 1). Mouse models have demonstrated that neonatal RV (RV1B) infection results 315 in persistent airway hyperresponsiveness, mucous cell metaplasia, and IL-13 production that 316 does not occur in adult mice. Furthermore, knockout of IL4R prevents this response, consistent with an IL13-dependent process.³⁹ Subsequent work demonstrated that RV infection leads to 317 expression of epithelial derived cytokines IL-25, IL-33, and TSLP and an increase in ILC2s as 318 319 an important source of airway IL-13; blocking these pathways with anti-IL-25 attenuates neonatal RV-induced AHR and mucous cell metaplasia.^{40, 41} While there is no equivalent human 320

321 evidence regarding the immune effects of RV in early life, these same pathways are known to play a key role in the response to RV leading to exacerbations in established asthma.²⁹ 322 A key guestion however, is whether underlying Th2 inflammation or RV associated 323 324 wheezing comes first. A prospective birth cohort analysis has shown that allergic sensitization 325 generally precedes RV wheezing but not the other way around, suggesting allergic sensitization may lead to more severe RV illnesses and the development of asthma.⁴² Supporting this 326 observation, in vitro studies have shown that Th2 inflammation can inhibit type I and III 327 interferon antiviral responses to RV infections,^{43, 44} which may increase susceptibility to more 328 329 severe RV infections. However in contrast, several human studies have demonstrated 330 increased IFN signatures in asthmatic children with virus infections, as well as in severe asthma in adults;⁴⁵⁻⁴⁷ these might represent different disease states, as a recent report found that early 331 332 life exacerbation-prone asthma was correlated with low IFN signatures, while the highest IFN signatures were associated with later-onset asthma.48 333

Allergy is a major risk factor for the progression from wheezing illnesses to asthma, and 334 this has been a very consistent finding across multiple cohorts.^{21, 49, 50} Allergic sensitization 335 precedes wheezing illnesses in most young children,⁴² and allergic inflammation can impair 336 antiviral responses *in vitro*⁵¹ and *in vivo*.⁵² This suggests that allergic airway inflammation can 337 increase susceptibility to and severity of viral respiratory illnesses. Allergic sensitization in early 338 339 childhood may also modify the relationship between microbial colonization and respiratory 340 outcomes. In preschool children whose airways were colonized with pathogen-dominated 341 microbiomes, sensitized children were at increased risk for chronic asthma while non-sensitized children were likely to have transient wheeze that resolved by age 4 years.⁵³ 342

It is well established that hospitalization for RSV bronchiolitis in the first year of life is
associated with later development of asthma.^{38, 54-56} RSV induces a broad innate immune
response in infants including systemic interferon, neutrophil, and inflammatory pathways, and
distinct RSV strains and concomitant airway bacteria can influence the severity of infection.^{57, 58}

347 The risk for more severe RSV-illnesses has also been linked to polymorphisms in several immune regulatory genes,⁵⁹ many of which also can influence asthma risk. However, whether 348 RSV is causal remains a subject of debate with two large cohort studies showing different 349 conclusions,^{59,60} one suggesting causation and the other an underlying genetic predisposition. 350 351 Notably, two prevention studies using palivizumab (a monoclonal antibody directed against RSV) in high-risk infants found that prevention of more severe RSV-illnesses decreased the risk 352 of childhood recurrent wheezing but not asthma development.^{61, 62} Ultimately RSV infection 353 354 appears to have the greatest impact on asthma risk during a critical window of lung 355 development for infants born during the fall (in the Northern hemisphere) who are at ~4 months 356 of age during the peak of the winter RSV season. It has been well-established that RSV 357 infection can induce pathologic Th2 immune responses, especially within the context of formalin-inactivated RSV vaccination.⁶³⁻⁶⁵ More recently, studies in mice have demonstrated the 358 359 ability for RSV-related Pneumonia Virus of Mice as well as human RSV infection to break tolerance to allergens in neonatal mice.⁶⁶ Furthermore, it is now appreciated that RSV triggers 360 361 release of epithelial-derived cytokines that promote Th2 responses and can induce ILC2 responses following infection.^{67, 68} These same epithelial cytokines have also been implicated in 362 363 RV infection, perhaps suggesting a shared innate Th2-skewing mechanism during viral infection.^{27, 40} 364

<u>Future perspectives:</u> Fully understanding the patterns of immune development that lead to
 asthma inception, and how such patterns are affected by exogeneous exposures including viral
 infections, will direct asthma prevention research. Ongoing studies are focused on altering Th2 skewing in early life including through blocking IgE and through altering microbial exposures. If
 effective, decreasing Th2-inflammation in early life may function in part through enhancing
 antiviral responses.^{51, 69} However, antiviral specific therapies including RV and RSV vaccines,
 may also prove to be critical in asthma prevention.

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373 Dynamics of respiratory tract bacterial colonization during viral upper respiratory tract 374 infection

Detection of viruses in the upper airway during peak viral seasons can be as high as 90% in prospective studies.⁷⁰ However, rates of illness are significantly lower, leading researchers to question why some patients are more susceptible to increased morbidity when they have a viral upper respiratory tract infection (URI). One factor that has been shown to increase upper and lower airway symptoms during viral infections are bacteria.⁷¹

These bacteria collectively constitute the microbiota. The upper airway microbiota develops over the first year of life with alterations in the natural development associated with increased risk for URIs during the first few years of life.^{72, 73} The most abundant bacteria within the upper airway of infants and children are *Staphylococcus*, *Streptococcus*, *Moraxella*, *Haemophilus*, *Dolosigranulum*, and *Corynebacterium*.^{71, 72, 74-78}

385 In several infant cross-sectional and cohort studies, the presence of Streptococcus, Moraxella, or Haemophilus during upper respiratory infection increases the likelihood that the 386 infant will have lower airway symptoms.^{72, 76} Studies examining airway bacteria during RSV 387 bronchiolitis have reported links between an increased abundance of Streptococcus⁹ and 388 Haemophilus.⁷⁸ In contrast, RV-bronchiolitis is associated with an increased abundance of 389 390 Moraxella and Haemophilus.⁹ While these studies suggest that a bacteria-virus interaction 391 occurs during infancy, only a few studies have examined the association between virus and 392 bacteria in school-age children. One such study revealed that children with Streptococcus or 393 Moraxella present in their airway are more likely to have cold and asthma symptoms during a naturally occurring RV infection.⁷¹ Collectively, these studies demonstrate that an association 394 395 exists between specific bacteria and illness severity.

While *Streptococcus*, *Moraxella* and *Haemophilus* are often associated with an increase in viral-associated symptoms, a higher abundance of *Corynebacterium*, *Staphylococcus* and *Dolosigranulum* is often present in the airway in the absence of viral detection and clinical

symptoms.^{75, 77, 78} In addition, when the latter three bacteria are enriched in the upper airway,
infants are less likely to have an acute respiratory illness,⁷² and school-age children are less
likely to have a symptomatic illness during RV infection.⁷⁵ Furthermore, high abundance of *Lactobacillus* in the upper airway during RSV illness is associated with a decreased risk of
childhood wheeze,⁷⁷ suggesting that bacteria present in the airway during viral illnesses may
contribute to both illness severity and long term sequela.

405 Future perspectives: Because most studies examining airway bacteria during viral infection 406 have been cross-sectional, observational studies, it remains unclear how airway microbes affect 407 the epithelium, and whether these interactions contribute to the causation of wheezing illnesses, 408 asthma development in young children, and exacerbations of established asthma. Greater 409 insight is needed into metabolic, immunologic and toxic effects of bacteria on epithelial cells that 410 could contribute to acute illnesses and asthma risk. While many studies have examined changes in bacteria that occur during viral infection, few have examined how the airway 411 microbiome influences susceptibility vs. resilience to viral infection. Some bacteria could 412 promote a "pro-inflammatory" environment thereby making the airway susceptible to viral 413 414 infection. The presence of *H. influenzae* in the infant airway prior to viral infection is associated with increased expression of local inflammatory cytokines suggesting a link between bacteria 415 and airway inflammation.⁷⁹ In contrast, mice receiving intranasal administration of *Lactobacillus* 416 *rhamnosus* prior to viral infection have enhanced antiviral immune responses.⁸⁰ suggesting that 417 418 some bacteria protect the airway and reduce the risk of symptomatic viral infection. Greater 419 understanding of these relationships may lead to new preventive approaches to acute viral-420 bacterial illnesses and perhaps the development of childhood asthma.

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422 The influence of the gut microbiome on viral infections of the respiratory tract

423 The gut microbiome represents the most abundant and diverse microbial environment in 424 the human body, comprised of approximately 40 trillion bacteria.⁸¹ These bacteria have

425 coevolved with humans over millennia to contribute to a symbiosis in which humans consume prebiotic fiber which is metabolized by resident microbes in the gut to create short chain fatty 426 acids (SCFA) which in turn regulate immune responses.^{82, 83} Alterations in this relationship are 427 428 occurring in modern times due to practices such as the frequent use of antibiotics, and the 429 consumption of a high-sugar, low fiber diet. As a consequence, a state of microbial dysbiosis, or an ecological imbalance, may result, which leads to the loss of metabolic capabilities and 430 predisposes infants to both the development of atopic diseases as well as an increased 431 susceptibility to viral infections.⁸⁴ 432

Although epidemiologic evidence strongly supports a role of the gut microbiome in the 433 development of asthma, the mechanisms remain unclear.^{85, 86} The most popular theory to 434 explain these observations is that colonization with certain gut bacteria have a direct anti-435 436 inflammatory effect on the respiratory tract decreasing the likelihood of airway hyperreactivity.⁸⁷ 437 However, there is evidence that certain species of microbiota in the gastrointestinal tract prime the respiratory immune system to effectively fight viral pathogens. Immunologic factors in early 438 life such as low blood cell interferon responses⁸⁸⁻⁹¹ and attenuated cytokine production⁹² have 439 440 been associated with increased risk for wheezing in infancy. Furthermore, patterns of metabolites (which can regulate immune responses⁹³) at birth are associated with the risk for 441 wheezing illnesses.⁹⁴ The idea that delayed immune maturation might contribute to wheezing is 442 supported by studies showing that early life exposures to dogs,^{95, 96} farm life,^{10, 97} and increased 443 microbes and allergens⁹⁸ are inversely related to the risk of wheezing illnesses. Furthermore, 444 exposure to these microbes and allergens during the prenatal period or infancy may be 445 immunostimulatory.^{99, 100} A loss, therefore, of these resident microbes may then lead to a 446 predisposition to viral infections and in turn, the development of asthma. 447

448 Several studies have proposed mechanisms for the influence of the gut microbiota on 449 both local and distant immune functions. SCFA have been shown to have a local effect on 450 immune responses through their influence on mucosal barrier function, and a loss of SCFA-

producing bacteria has been implicated in the development of food allergy.¹⁰¹ Recent advances 451 have also shown that this symbiosis also influences vital immune responses in other systemic 452 453 tissues. For example, in the absence of SCFA, mucosal barrier function can break down and allow for translocation of gut pathobionts, bacteria that are symbiotic under normal conditions 454 455 but pathogenic when removed from their normal environment, which, in turn, can drive autoimmunity.¹⁰² Similarly, in a murine model intact commensal bacteria in the gut were 456 457 required for adaptive immune responses to respiratory influenza virus infection. Specifically, when mice were treated with antibiotics, they had reduced virus-specific antibody titers, CD4+ 458 459 T-cell responses, and cytokine secretion which consequently resulted in elevated viral titers post 460 infection. This impairment, however, was rescued by local or distal injection of Toll-like receptor ligands.¹⁰³ Further, exposure to house dust from homes with dogs enriched the cecal 461 462 microbiome in a murine model with L. johnsonii, which protected them against infection with **RSV**.¹⁰⁴ 463

Future Perspectives: Although the pathways remain incomplete, evidence continues to mount 464 465 that the gut microbiome can influence the maturation of the immune system in viral defense and therefore the development of asthma.¹⁰⁵ Future therapies look to a role of probiotics for the 466 467 prevention and treatment of allergic disorders, with recent evidence that atopy risk may be 468 associated with a dysbiosis of the gut microbiome. Studies have shown that in asthma, MMP9 469 (members of a family of enzymes that cleave extracellular matrix proteins) levels were 470 significantly increased and treatment with the probiotic, L. rhamnosus GG (LCC), decreased MMP9 expression in lung tissue and inhibited inflammatory cell infiltration, as well as reducing 471 exhaled nitric oxide among 4- to 7-year olds in pediatric asthma.¹⁰⁶ 472 In early childhood, total fecal IgE levels appear to be specifically correlated with house dust 473

474 mite-specific IgE levels, indicating that fecal IgE levels represent markers of allergic response to
475 aeroallergens. A significant correlation of fecal IgE levels with *Dorea spp.* and *Clostridium spp.*

476 related to allergic rhinitis and asthma, respectively, suggest that modulation of particular subsets

of gut microbial dysbiosis could contribute to the susceptibility to allergic airway diseases.¹⁰⁷ 477 Future work is required for identification of specific species and functional studies to understand 478 479 the strength and mechanism of these associations. In the future, it is critical to understand more 480 precisely the microbiota composition. Optimized biomarker studies of the microbial taxa and the 481 metabolites involved in asthma-associated dysbiosis could help identify infants at risk of asthma 482 before symptoms. This would also provide a scientific rationale for future therapeutic strategies 483 aimed at restoring an altered infant gut microbiome. Future studies need to revolve around 484 state-of-the-art methods for the evaluation of the microflora to better define indications, the 485 probiotic strains and the type of prebiotic to be used. 486 Potential for primary prevention: clinical trials aiming to prevent the development of the 487 488 episodic wheeze phenotype The inception of childhood asthma is tightly related to early life events such as 489 respiratory infections and the development of aeroallergens sensitization. Other co-factors (e.g., 490 491 vitamin D) may modulate asthma inception pathways. Previous and on-going clinical trials, 492 geared for asthma prevention, have targeted these pathways and co-factors. Early life respiratory infections are significant determinants of childhood asthma¹⁰⁸. In 493 young toddlers, prevention of severe RSV bronchiolitis may reduce the risk of episodic 494 wheeze/asthma development ^{109, 110}. In preterm infants (33-35 wks), palivizumab treatment 495 during the RSV season resulted in a 73% reduction in the number of wheezing days during the 496 first year of life, and outside of the RSV season¹⁰⁹. A follow up study from the same cohort, 497 498 revealed that at the age of 6 years the intervention resulted in a 41% relative risk reduction in parent-reported asthma, but the forced expiratory volume in 0.5 s ($FEV_{0.5}$) percentage predicted 499 500 values, which was an additional primary outcome, were similar between the palivizumab and placebo treated infants¹¹⁰. 501

502 Since early life respiratory infections cannot be completely prevented, attenuation of the 503 immune/inflammatory processes during these infections may be another pathway for asthma 504 prevention. This concept is illustrated by the results of a proof-of concept clinical trial in 40 505 infants hospitalized with RSV bronchiolitis. In this trial, azithromycin treatment for 2 weeks, 506 during acute RSV bronchiolitis, reduced the likelihood of developing recurrent wheeze during 507 the subsequent year¹¹¹. Azithromycin effects were attributed to anti-inflammatory properties 508 and/or its effects on the airway microbiome¹¹². A larger confirmatory trial is ongoing (APW-RSV 509 II; NCT02911935; Table 3).

510 Based on observational studies that linked maternal vitamin D deficiency to childhood asthma, two clinical trials (VDAART¹¹³, COPSAC2010¹¹⁴) investigated whether maternal vitamin 511 D supplementation (2400 IU/day¹¹³, 4000IU/day¹¹⁴) during pregnancy would prevent 512 513 asthma/recurrent wheeze in their children. A recent meta-analysis that combined these two 514 trials revealed that this intervention resulted in a 25% significant reduction in asthma/recurrent wheeze risk during the first 3 years of life ¹¹⁵. The effect was most profound among women with 515 516 sufficient serum vitamin D levels at randomization highlighting the importance of normal preconception vitamin D levels¹¹⁵. It was suggested that vitamin D beneficial effects may be related 517 518 to enhancement of in-utero lung growth and development and promotion of antimicrobial effects, thereby reducing early life respiratory infections, and/or providing immune modulation effects ¹¹⁶. 519 520 Omega-3 fatty acids were suggested to have anti-inflammatory effects, potentially due to 521 decreased production of arachidonic acid metabolites. In a recent clinical trial, high dose 522 Omega-3 fatty acids supplementation (2.4 g daily) to pregnant women, beginning at 24 week of

523 gestation, resulted in a 30% relative risk reduction of persistent wheeze or asthma at age 3 524 years¹¹⁷. These positive effects were driven by subgroups of children born to mothers with a 525 variant of the gene encoding fatty acid desaturase, predisposing to low ability to produce 526 omega-3 fatty acids, and by infants born to mothers with low omega-3 fatty acids baseline blood 527 levels. These sub-group analyses suggest the plausibility of a precision-medicine approach of

this potential future intervention. Nevertheless, it is important to assure that high dose omega-3
fatty acids does not possess any safety issues, before omega-3 fatty acids may be utilized for
asthma prevention.

531 The ongoing ORBEX clinical trial (NCT02148796) is attempting to modulate the infant immune system by treating high-risk preschool children with Broncho-Vaxom® for 2 years to 532 533 prevent/delay the development of wheezing lower respiratory tract illness (WLRI) during a third observation year. Broncho-Vaxom® contains bacterial lysates and was previously shown to 534 reduce the rate of respiratory infections¹¹⁸. Hence, it is postulated that prevention of early life 535 536 WLRI will prevent the development of the recurrent wheeze phenotype. Finally, the ongoing 537 PARK clinical trial (NCT02570984) is targeting the association between allergic sensitization 538 and asthma inception. PARK investigates whether treatment of high-risk preschool children with 539 Omalizumab for 2-years would prevent asthma development, and whether the treatment would 540 decrease asthma severity among infants who will develop asthma, during an additional 2-year 541 observation period. 542 Future perspectives: This is an exciting time for all involved in childhood asthma prevention:

recent clinical trials have shown the feasibility of asthma prevention, and multiple clinical trials are ongoing toward this goal. In addition to targeting type 2 immune responses, new interventions are needed to inhibit viral replication, either with specific inhibitors or strategies to boost the development of global antiviral responses in the airways. Finally, studies in farming environments strongly suggest that environmental exposure can lower the risk of viral respiratory illness in addition to reducing allergy.^{10, 97} Identifying relevant mechanisms is likely to lead to new preventive approaches to virus-induced wheeze and asthma.

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551 New therapeutic options to decrease severity of asthma exacerbations caused by

552 respiratory viral infection

553 Recent studies have focused on short-term increases in standard asthma therapy, 554 vitamin D supplementation, azithromycin and anti-IgE therapy. However, mixed efficacy results 555 limit the widespread application of many of these therapies in clinical practice. 556 Maintenance inhaled corticosteroids (ICS) are effective in reducing the risk of asthma 557 exacerbations and, when combined with inhaled long acting beta agonists (LABA), this 558 decreases the risk further. However, exacerbations continue to occur. Attempts to increase 559 dosing of inhaled steroid with early signs of loss of asthma control with viral infection, termed 560 the "yellow zone", to decrease exacerbation risk, have yielded mixed results. GINA guidelines 561 suggest increasing ICS at onset of symptoms as part of a self-management plan 562 (http://www.ginasthma.org). A Cochrane database review (including five studies in adults and 563 three studies in children) concluded that current evidence does not support increasing ICS in 564 mild to moderate asthma patients as part of a self-management plan to treat exacerbations.¹¹⁹ A clinical trial examined this question further in 254 children aged 5-11 with history of 565 mild to moderate persistent asthma with at least one previous exacerbation in the past year.¹²⁰ 566 567 Children were treated for 48 weeks with low dose inhaled steroid and assigned to either 568 continue this or quintuple the dose for 7 days at onset of loss of asthma control. There was no 569 significant difference in the rate of severe exacerbations in the groups. The total corticosteroid 570 exposure in the high dose group was 16% higher (including both inhaled corticosteroid use and 571 prednisone) and there was an effect on linear growth velocity between the high dose and low 572 dose group (-0.23 cm/year), suggesting potential risk without identifiable benefit of the therapy. 573 Vitamin D levels have been inversely associated with asthma severity, including hospitalization for severe infections.¹²¹ A large study aimed at optimizing low Vitamin D levels 574 through supplementation did not reduce rates of colds or treatment failures in adults with 575 asthma.^{122, 123} In contrast, a meta-analysis of seven randomized trials demonstrated a significant 576 reduction in asthma exacerbations, with the effect seen only in patients with low vitamin D at 577

baseline.¹²⁴ There are ongoing studies in children with asthma examining the possible role of
vitamin D supplementation in preventing asthma exacerbations (Table 3).

580 Current guidelines do not recommend the use of antibiotic treatment for episodes of 581 asthma-like symptoms in children, yet they are commonly used. A randomized, double-blind, placebo-controlled trial conducted in the US, evaluated the role of early administration of 582 583 azithromycin in prevention of progression to severe lower respiratory tract illness (LRTI) 584 symptoms.¹²⁵ Preschool children, age 12-71 months, with history of recurrent severe wheezing 585 in the setting of LRTI, were randomized to azithromycin 12 mg/kg for 5 days (307 patients) or 586 placebo (300 patients). The medications were to be started as soon as the children developed 587 signs or symptoms that typically preceded the development of a severe LRTI. The primary 588 outcome measure was the number of respiratory tract infections not progressing to a severe 589 LRTI. The azithromycin group experienced a lower risk of progression to a severe LRTI than the 590 placebo group.

In the COPSAC₂₀₁₀ (Copenhagen Prospective Studies on Asthma in Childhood 2010 591 592 cohort), children (age 1-3) with recurrent asthma-like symptoms within this cohort were enrolled in a study to assess the duration of episodes when treated with azithromycin.¹²⁶ With each 593 594 episode of 3 days of consecutive symptoms (wheeze, cough, dyspnea), children were 595 randomized to receive 10 mg/kg azithromycin or placebo for 3 days. Seventy-two children from 596 the recurrent asthma-like symptoms group had 158 episodes. The azithromycin treatment 597 shortened the days of symptoms, 3.4 days compared with 7.7 days after placebo, 598 corresponding to a calculated reduction in episode length of 63.3%. More improvement was 599 seen when the treatment was started earlier in the episode; however, treatment did not 600 significantly affect the time to next episode of troublesome lung symptoms in children. 601 With these episodes, a hypopharyngeal aspirate was collected and cultured for common 602 bacterial pathogens and a nasopharyngeal aspirate was collected for viral PCR. Overall, the

603 presence of any cultured pathogenic bacteria did not significantly alter the treatment effect

compared to episodes without bacteria present; however, azithromycin was more effective in
those whose culture grew *H. influenzae*. The treatment effect in these studies is promising;
however, resistance to these antibiotics and eliminating commensal microbes along with
pathogens are concerns with repeated treatment.

Birth cohort studies have shown allergic sensitization to be a risk factor for RV-induced wheeze.⁴² Additionally, in one prospective cohort study, the severity of RV-triggered asthma exacerbation increased as the degree of allergen sensitization increased, with serum IgE levels (total IgE and allergen specific IgE) increasing from baseline during the exacerbation.¹²⁷ Persistence of asthma by age 13 was most strongly associated with wheezing illness with RV and aeroallergen sensitization in early life³⁷ suggesting a role for both viral infection and allergic sensitization in the development of asthma.

615 A possible mechanism for impaired response to viral infections in allergic asthmatics is a decreased secretion of IFN in response to viral infection. Purified plasmacytoid dendritic cells 616 (pDC) from patients with allergic asthma were shown to secrete less IFN-α in response to 617 exposure with influenza A virus.⁵¹ Increased FcεRIα expression and serum IgE levels were 618 619 inversely associated with IFN- a secretion. The increased susceptibility to viral wheeze in atopic 620 patients and impaired antiviral response in these patients suggests a role for possible 621 therapeutic intervention to decrease allergic inflammation with the goal of decreasing asthma 622 exacerbations in response to viral infection.

Omalizumab, a humanized monoclonal antibody that selectively binds to IgE, has
recently been studied as an add-on therapy to prevent fall asthma exacerbations in atopic
asthmatics in the Preventative Omalizumab or Step-Up Therapy for Fall Exacerbations
(PROSE) study.¹²⁸ The PROSE study included 478 children, age 6-17, with respiratory allergy
and asthma, randomized to either inhaled corticosteroid boost, add on omalizumab or placebo.
All patients had guidelines-based care in addition to the add-on treatment (ICS boost,
omalizumab or placebo). Treatment was begun 4-6 weeks before the participant's school start

630 day and ended 90 days after school start date. Omalizumab treatment significantly decreased 631 the odds of having at least 1 exacerbation, whereas boosting ICS did not reduce risk. 632 Omalizumab increased IFN- α responses to RV *ex vivo*. Within the omalizumab group, greater 633 restoration of IFN- α responses were associated with fewer exacerbations. In this trial, 634 omalizumab was associated with a decreased frequency of RV illnesses, decreased duration of RV infection as well as decreased frequency of overall respiratory illness, and reduced peak RV 635 shedding.⁵² Omalizumab reduced expression of FccRIa on the surface of pDC and this 636 637 reduction was associated with lower exacerbation rates and correlated with enhanced IFN-α production, suggestion a possible mechanism for the interaction between allergic sensitization 638 and virus-induced asthma exacerbations.⁶⁹ However, the connection between the pDC type I 639 IFN production and asthma exacerbation will benefit from further study. 640 641 In an observational study following children with asthma presenting with an acute

asthma exacerbation triggered by RV, the use of omalizumab for at least 4 weeks prior to presentation was associated with reduced severity of exacerbation compared with patients primarily treated with ICS.¹²⁹ This suggests a benefit in not only frequency and duration of asthma exacerbation, but also severity of exacerbation.

646 Another possible mechanism for the interaction between allergic sensitization and virusinduced asthma exacerbations is the presence of anti-viral IgE in response to infection. In RSV 647 648 infection in infants, RSV specific IgE was detected in nasopharyngeal secretions, with significantly higher titers in subjects with wheezing.^{130, 131} Correlation of the peak titers with 649 650 degree of hypoxia was also noted. Following known exposure to a specific laboratory strain, RV-specific IgE could be detected in human sera.¹³² While the IgE response to RV and RSV 651 652 are associated with infection, the role of IgE in the host response to these infections is not fully 653 understood. Given the decreased exacerbations with use of omalizumab, further investigation 654 into the role of anti-viral IgE is indicated.

<u>Future perspectives</u>: Given the morbidity of RSV and RV infections in patients with asthma, a
consistent and effective treatment approach is highly desirable. While studies have found
possible benefits to treatment with azithromycin and omalizumab, the widespread use of these
treatment approaches is not currently justified. Further characterization of risk in this patient
population and additional work to delineate the mechanisms by which these drugs are effective
may lead to selection of patients most appropriate for these therapies.

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663 Conclusion

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There have been important advances in our knowledge of the relationship between viruses and 665 666 asthma over the last decade. Advances in scientific methods have provided innovative 667 opportunities to examine host, environment, and viral interactions that either protect against or increase vulnerability to asthma development and exacerbations. The exploration of the 668 669 contribution of the respiratory and gut microbiome to virally-induced asthma is in its infancy and 670 we suspect that over the next 5 years there will be major advances in this area. Finally, primary 671 prevention is a major goal to diminish the morbidity of virally-mediated wheezing, asthma and 672 exacerbations. Until primary prevention becomes a reality, clinical trials examining the impact of 673 established medications, as well as novel therapies, will be critical to diminish the impact of viral 674 infections on wheezing and asthma.

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676 Bibliography

- 677
 678 1. Stier MT, Peebles RS, Jr. Host and Viral Determinants of Respiratory Syncytial Virus679 induced Airway Mucus. Ann Am Thorac Soc 2018; 15:S205-S9.
- Lukacs NW, Moore ML, Rudd BD, Berlin AA, Collins RD, Olson SJ, et al. Differential
 immune responses and pulmonary pathophysiology are induced by two different strains
 of respiratory syncytial virus. Am. J Pathol 2006; 169:977-86.
- Moore ML, Chi MH, Luongo C, Lukacs NW, Polosukhin VV, Huckabee MM, et al. A
 chimeric A2 strain of respiratory syncytial virus (RSV) with the fusion protein of RSV
 strain line 19 exhibits enhanced viral load, mucus, and airway dysfunction. J Virol 2009;
 83:4185-94.
- Loisel DA, Du G, Ahluwalia TS, Tisler CJ, Evans MD, Myers RA, et al. Genetic associations
 with viral respiratory illnesses and asthma control in children. Clin Exp Allergy 2016;
 46:112-24.
- 690 5. Ciancanelli MJ, Huang SX, Luthra P, Garner H, Itan Y, Volpi S, et al. Infectious disease.
 691 Life-threatening influenza and impaired interferon amplification in human IRF7
 692 deficiency. Science 2015; 348:448-53.
- 6. Asgari S, Schlapbach LJ, Anchisi S, Hammer C, Bartha I, Junier T, et al. Severe viral
 694 respiratory infections in children with IFIH1 loss-of-function mutations. Proc Natl Acad
 695 Sci U S A 2017; 114:8342-7.
- 696 7. Janssen R, Bont L, Siezen CL, Hodemaekers HM, Ermers MJ, Doornbos G, et al. Genetic
 697 susceptibility to respiratory syncytial virus bronchiolitis is predominantly associated with
 698 innate immune genes. J Infect Dis 2007; 196:826-34.
- 699 8. Larkin EK, Hartert TV. Genes associated with RSV lower respiratory tract infection and
 700 asthma: the application of genetic epidemiological methods to understand causality.
 701 Future Virol 2015; 10:883-97.
- 702 9. Caliskan M, Bochkov YA, Kreiner-Moller E, Bonnelykke K, Stein MM, Du G, et al.
 703 Rhinovirus wheezing illness and genetic risk of childhood onset asthma. N Engl J Med
 704 2013; 368:1398-407.
- 10. Loss GJ, Depner M, Hose AJ, Genuneit J, Karvonen AM, Hyvarinen A, et al. The Early
 Development of Wheeze. Environmental Determinants and Genetic Susceptibility at
 17q21. Am J Respir Crit Care Med 2016; 193:889-97.
- Stokholm J, Chawes BL, Vissing N, Bonnelykke K, Bisgaard H. Cat exposure in early life
 decreases asthma risk from the 17q21 high-risk variant. J Allergy Clin Immunol 2018;
 141:1598-606.
- 12. Lee WM, Lemanske RF, Jr., Evans MD, Vang F, Pappas T, Gangnon R, et al. Human
 rhinovirus species and season of infection determine illness severity. Am J Respir Crit
 Care Med 2012; 186:886-91.
- Nakagome K, Bochkov YA, Ashraf S, Brockman-Schneider RA, Evans MD, Pasic TR, et al.
 Effects of rhinovirus species on viral replication and cytokine production. J Allergy Clin
 Immunol 2014; 134:332-41.
- 717 14. Bonnelykke K, Sleiman P, Nielsen K, Kreiner-Moller E, Mercader JM, Belgrave D, et al. A
 718 genome-wide association study identifies CDHR3 as a susceptibility locus for early
 719 childhood asthma with severe exacerbations. Nat Genet 2014; 46:51-5.

720 15. Bochkov YA, Watters K, Ashraf S, Griggs TF, Devries MK, Jackson DJ, et al. Cadherin-721 related family member 3, a childhood asthma susceptibility gene product, mediates 722 rhinovirus C binding and replication. Proc Natl Acad Sci U S A 2015; 112:5485-90. 723 Bonnelykke K, Coleman AT, Evans MD, Thorsen J, Waage J, Vissing NH, et al. Cadherin-16. 724 related Family Member 3 Genetics and Rhinovirus C Respiratory Illnesses. Am J Respir 725 Crit Care Med 2018; 197:589-94. 726 17. Vrijheid M. The exposome: a new paradigm to study the impact of environment on 727 health. Thorax 2014; 69:876-8. 728 18. Sigurs N, Bjarnason R, Sigurbergsson F, Kjellman B. Respiratory syncytial virus 729 bronchiolitis in infancy is an important risk factor for asthma and allergy at age 7. Am J 730 Respir Crit Care Med 2000; 161:1501-7. 731 19. Welliver RC. RSV and chronic asthma. Lancet 1995; 346:789-90. 732 20. Carroll KN, Wu P, Gebretsadik T, Griffin MR, Dupont WD, Mitchel EF, et al. The severity-733 dependent relationship of infant bronchiolitis on the risk and morbidity of early 734 childhood asthma. J Allergy Clin Immunol 2009; 123:1055-61. 735 21. Jackson DJ, Gangnon RE, Evans MD, Roberg KA, Anderson EL, Pappas TE, et al. Wheezing 736 rhinovirus illnesses in early life predict asthma development in high-risk children. Am J 737 Respir Crit Care Med 2008; 178:667-72. 738 22. Lemanske RF, Jr., Jackson DJ, Gangnon RE, Evans MD, Li Z, Shult PA, et al. Rhinovirus 739 illnesses during infancy predict subsequent childhood wheezing. J Allergy Clin Immunol 740 2005; 116:571-7. 741 23. Lukkarinen M, Koistinen A, Turunen R, Lehtinen P, Vuorinen T, Jartti T. Rhinovirus-742 induced first wheezing episode predicts atopic but not nonatopic asthma at school age. J 743 Allergy Clin Immunol 2017; 140:988-95. 744 24. Heymann PW, Carper HT, Murphy DD, Platts-Mills TA, Patrie J, McLaughlin AP, et al. 745 Viral infections in relation to age, atopy, and season of admission among children 746 hospitalized for wheezing. J Allergy Clin Immunol 2004; 114:239-47. 747 25. Rakes GP, Arruda E, Ingram JM, Hoover GE, Zambrano JC, Hayden FG, et al. Rhinovirus 748 and respiratory syncytial virus in wheezing children requiring emergency care. IgE and 749 eosinophil analyses. Am J Respir Crit Care Med 1999; 159:785-90. 750 26. Soto-Quiros M, Avila L, Platts-Mills TA, Hunt JF, Erdman DD, Carper H, et al. High titers of 751 IgE antibody to dust mite allergen and risk for wheezing among asthmatic children 752 infected with rhinovirus. J Allergy Clin Immunol 2012; 129:1499-505 e5. 753 27. Beale J, Jayaraman A, Jackson DJ, Macintyre JDR, Edwards MR, Walton RP, et al. 754 Rhinovirus-induced IL-25 in asthma exacerbation drives type 2 immunity and allergic 755 pulmonary inflammation. Sci Transl Med 2014; 6:256ra134. 756 28. Contoli M, Message SD, Laza-Stanca V, Edwards MR, Wark PA, Bartlett NW, et al. Role of 757 deficient type III interferon-lambda production in asthma exacerbations. Nat Med 2006; 758 12:1023-6. 759 29. Jackson DJ, Makrinioti H, Rana BM, Shamji BW, Trujillo-Torralbo MB, Footitt J, et al. IL-760 33-Dependent Type 2 Inflammation during Rhinovirus-induced Asthma Exacerbations In 761 Vivo. Am J Respir Crit Care Med 2014; 190:1373-82.

762 30. Kennedy JL, Koziol-White CJ, Jeffus S, Rettiganti MR, Fisher P, Kurten M, et al. Effects of 763 rhinovirus 39 infection on airway hyperresponsiveness to carbachol in human airways 764 precision cut lung slices. J Allergy Clin Immunol 2018; 141:1887-90 e1. 765 31. Kennedy JL, Shaker M, McMeen V, Gern J, Carper H, Murphy D, et al. Comparison of viral load in individuals with and without asthma during infections with rhinovirus. Am J 766 767 Respir Crit Care Med 2014; 189:532-9. 768 32. Khaitov MR, Laza-Stanca V, Edwards MR, Walton RP, Rohde G, Contoli M, et al. 769 Respiratory virus induction of alpha-, beta- and lambda-interferons in bronchial 770 epithelial cells and peripheral blood mononuclear cells. Allergy 2009; 64:375-86. 771 Laza-Stanca V, Message SD, Edwards MR, Parker HL, Zdrenghea MT, Kebadze T, et al. 33. 772 The role of IL-15 deficiency in the pathogenesis of virus-induced asthma exacerbations. 773 PLoS Pathog 2011; 7:e1002114. 774 34. Message SD, Laza-Stanca V, Mallia P, Parker HL, Zhu J, Kebadze T, et al. Rhinovirus-775 induced lower respiratory illness is increased in asthma and related to virus load and 776 Th1/2 cytokine and IL-10 production. Proc Natl Acad Sci U S A 2008; 105:13562-7. 777 Liu L, Pan Y, Zhu Y, Song Y, Su X, Yang L, et al. Association between rhinovirus wheezing 35. 778 illness and the development of childhood asthma: a meta-analysis. BMJ Open 2017; 779 7:e013034. 780 36. Midulla F, Nicolai A, Ferrara M, Gentile F, Pierangeli A, Bonci E, et al. Recurrent 781 wheezing 36 months after bronchiolitis is associated with rhinovirus infections and 782 blood eosinophilia. Acta Paediatr 2014; 103:1094-9. 783 37. Rubner FJ, Jackson DJ, Evans MD, Gangnon RE, Tisler CJ, Pappas TE, et al. Early life 784 rhinovirus wheezing, allergic sensitization, and asthma risk at adolescence. J Allergy Clin 785 Immunol 2017; 139:501-7. 786 38. Ruotsalainen M, Hyvarinen MK, Piippo-Savolainen E, Korppi M. Adolescent asthma after 787 rhinovirus and respiratory syncytial virus bronchiolitis. Pediatr Pulmonol 2013; 48:633-9. 788 39. Schneider D, Hong JY, Popova AP, Bowman ER, Linn MJ, McLean AM, et al. Neonatal 789 rhinovirus infection induces mucous metaplasia and airways hyperresponsiveness. J 790 Immunol 2012; 188:2894-904. 791 40. Hong JY, Bentley JK, Chung Y, Lei J, Steenrod JM, Chen Q, et al. Neonatal rhinovirus 792 induces mucous metaplasia and airways hyperresponsiveness through IL-25 and type 2 793 innate lymphoid cells. J Allergy Clin Immunol 2014; 134:429-39 e8. 794 41. Han M, Rajput C, Hong JY, Lei J, Hinde JL, Wu Q, et al. The Innate Cytokines IL-25, IL-33, 795 and TSLP Cooperate in the Induction of Type 2 Innate Lymphoid Cell Expansion and 796 Mucous Metaplasia in Rhinovirus-Infected Immature Mice. J Immunol 2017; 199:1308-797 18. 798 42. Jackson DJ, Evans MD, Gangnon RE, Tisler CJ, Pappas TE, Lee WM, et al. Evidence for a 799 causal relationship between allergic sensitization and rhinovirus wheezing in early life. 800 Am J Respir Crit Care Med 2012; 185:281-5. 801 43. Contoli M, Ito K, Padovani A, Poletti D, Marku B, Edwards MR, et al. Th2 cytokines impair 802 innate immune responses to rhinovirus in respiratory epithelial cells. Allergy 2015; 803 70:910-20. 804 44. Bochkov YA, Grindle K, Vang F, Evans MD, Gern JE. Improved molecular typing assay for 805 rhinovirus species A, B, and C. J Clin Microbiol 2014; 52:2461-71.

Miller EK, Hernandez JZ, Wimmenauer V, Shepherd BE, Hijano D, Libster R, et al. A

Mechanistic Role for Type III IFN-lambda1 in Asthma Exacerbations Mediated by Human

806

807

45.

808 Rhinoviruses. Am J Respir Crit Care Med 2012; 185:508-16. 809 46. Altman MC, Reeves SR, Parker AR, Whalen E, Misura KM, Barrow KA, et al. Interferon 810 response to respiratory syncytial virus by bronchial epithelium from children with 811 asthma is inversely correlated with pulmonary function. J Allergy Clin Immunol 2017. 812 47. Raundhal M, Morse C, Khare A, Oriss TB, Milosevic J, Trudeau J, et al. High IFN-gamma 813 and low SLPI mark severe asthma in mice and humans. J Clin Invest 2015; 125:3037-50. 814 48. Custovic A, Belgrave D, Lin L, Bakhsoliani E, Telcian AG, Solari R, et al. Cytokine 815 Responses to Rhinovirus and Development of Asthma, Allergic Sensitization, and 816 Respiratory Infections during Childhood. Am J Respir Crit Care Med 2018; 197:1265-74. 817 49. Illi S, von Mutius E, Lau S, Niggemann B, Gruber C, Wahn U. Perennial allergen 818 sensitisation early in life and chronic asthma in children: a birth cohort study. Lancet 819 2006; 368:763-70. 820 50. Kusel MM, de Klerk NH, Kebadze T, Vohma V, Holt PG, Johnston SL, et al. Early-life 821 respiratory viral infections, atopic sensitization, and risk of subsequent development of 822 persistent asthma. J Allergy Clin Immunol 2007; 119:1105-10. 823 51. Gill MA, Bajwa G, George TA, Dong CC, Dougherty II, Jiang N, et al. Counterregulation 824 between the FcepsilonRI pathway and antiviral responses in human plasmacytoid 825 dendritic cells. J Immunol 2010; 184:5999-6006. 826 52. Esquivel A, Busse WW, Calatroni A, Togias AG, Grindle KG, Bochkov YA, et al. Effects of 827 Omalizumab on Rhinovirus Infections, Illnesses, and Exacerbations of Asthma. Am J 828 Respir Crit Care Med 2017; 196:985-92. 829 Inouye M, Teo SM, Tang H, Mok D, Judd L, Watts S, et al. Dynamics of airway microbiota 53. 830 identify a critical window for interplay of pathogenic bacteria and allergic sensitization in 831 childhood respiratory disease. Cell Host & Microbe 2018. 832 54. Stein RT, Sherrill D, Morgan WJ, Holberg CJ, Halonen M, Taussig LM, et al. Respiratory 833 syncytial virus in early life and risk of wheeze and allergy by age 13 years. The Lancet 834 1999; 354:541-5. 835 55. Henderson J, Hilliard TN, Sherriff A, Stalker D, Shammari NA, Thomas HM. 836 Hospitalization for RSV bronchiolitis before 12 months of age and subsequent asthma, 837 atopy and wheeze: A longitudinal birth cohort study. Pediatric Allergy and Immunology 838 2005; 16:386-92. 839 56. Castro-Rodriguez JA, Forno E, Rodriguez-Martinez CE, Celedon JC. Risk and Protective 840 Factors for Childhood Asthma: What Is the Evidence? J Allergy Clin Immunol Pract 2016; 841 4:1111-22. 842 57. Rodriguez-Fernandez R, Tapia LI, Yang CF, Torres JP, Chavez-Bueno S, Garcia C, et al. 843 Respiratory Syncytial Virus Genotypes, Host Immune Profiles, and Disease Severity in 844 Young Children Hospitalized With Bronchiolitis. J Infect Dis 2017; 217:24-34. 845 de Steenhuijsen Piters WA, Heinonen S, Hasrat R, Bunsow E, Smith B, Suarez-Arrabal 58. 846 MC, et al. Nasopharyngeal Microbiota, Host Transcriptome, and Disease Severity in 847 Children with Respiratory Syncytial Virus Infection. Am J Respir Crit Care Med 2016; 848 194:1104-15.

- 849 59. Wu P, Dupont WD, Griffin MR, Carroll KN, Mitchel EF, Gebretsadik T, et al. Evidence of a
 850 causal role of winter virus infection during infancy in early childhood asthma. Am J
 851 Respir Crit Care Med 2008; 178:1123-9.
- 852 60. Thomsen SF, van der Sluis S, Stensballe LG, Posthuma D, Skytthe A, Kyvik KO, et al.
 853 Exploring the association between severe respiratory syncytial virus infection and
 854 asthma: a registry-based twin study. Am J Respir Crit Care Med 2009; 179:1091-7.
- 61. Carroll KN, Gebretsadik T, Escobar GJ, Wu P, Li SX, Walsh EM, et al. Respiratory syncytial
 virus immunoprophylaxis in high-risk infants and development of childhood asthma.
 Journal of Allergy and Clinical Immunology 2017; 139:66-71.e3.
- Mochizuki H, Kusuda S, Okada K, Yoshihara S, Furuya H, Simoes EAF, et al. Palivizumab
 Prophylaxis in Preterm Infants and Subsequent Recurrent Wheezing. Six-Year Follow-up
 Study. Am J Respir Crit Care Med 2017; 196:29-38.
- 63. Connors M, Giese NA, Kulkarni AB, Firestone CY, Morse HC, 3rd, Murphy BR. Enhanced
 pulmonary histopathology induced by respiratory syncytial virus (RSV) challenge of
 formalin-inactivated RSV-immunized BALB/c mice is abrogated by depletion of
 interleukin-4 (IL-4) and IL-10. J Virol 1994; 68:5321-5.
- 865 64. Waris ME, Tsou C, Erdman DD, Zaki SR, Anderson LJ. Respiratory synctial virus infection
 866 in BALB/c mice previously immunized with formalin-inactivated virus induces enhanced
 867 pulmonary inflammatory response with a predominant Th2-like cytokine pattern. J Virol
 868 1996; 70:2852-60.
- 869 65. Johnson TR, Johnson JE, Roberts SR, Wertz GW, Parker RA, Graham BS. Priming with
 870 secreted glycoprotein G of respiratory syncytial virus (RSV) augments interleukin-5
 871 production and tissue eosinophilia after RSV challenge. J Virol 1998; 72:2871-80.
- Krishnamoorthy N, Khare A, Oriss TB, Raundhal M, Morse C, Yarlagadda M, et al. Early
 infection with respiratory syncytial virus impairs regulatory T cell function and increases
 susceptibility to allergic asthma. Nat Med 2012; 18:1525-30.
- 875 67. Stier MT, Bloodworth MH, Toki S, Newcomb DC, Goleniewska K, Boyd KL, et al.
 876 Respiratory syncytial virus infection activates IL-13-producing group 2 innate lymphoid
 877 cells through thymic stromal lymphopoietin. J Allergy Clin Immunol 2016; 138:814-24
 878 e11.
- Siegle JS, Hansbro N, Herbert C, Rosenberg HF, Domachowske JB, Asquith KL, et al. Earlylife viral infection and allergen exposure interact to induce an asthmatic phenotype in
 mice. Respir Res 2010; 11:14.
- 69. Gill MA, Liu AH, Calatroni A, Krouse RZ, Shao B, Schiltz A, et al. Enhanced plasmacytoid
 dendritic cell antiviral responses after omalizumab. J Allergy Clin Immunol 2018;
 141:1735-43 e9.
- Olenec JP, Kim WK, Lee WM, Vang F, Pappas TE, Salazar LE, et al. Weekly monitoring of
 children with asthma for infections and illness during common cold seasons. J Allergy
 Clin Immunol 2010; 125:1001-6.
- Kloepfer KM, Lee WM, Pappas TE, Kang TJ, Vrtis RF, Evans MD, et al. Detection of
 pathogenic bacteria during rhinovirus infection is associated with increased respiratory
 symptoms and asthma exacerbations. J Allergy Clin Immunol 2014; 133:1301-7, 7 e1-3.

- Teo SM, Mok D, Pham K, Kusel M, Serralha M, Troy N, et al. The infant nasopharyngeal
 microbiome impacts severity of lower respiratory infection and risk of asthma
 development. Cell Host Microbe 2015; 17:704-15.
- 894 73. Hyde ER, Petrosino JF, Piedra PA, Camargo CA, Jr., Espinola JA, Mansbach JM.
 895 Nasopharyngeal Proteobacteria are associated with viral etiology and acute wheezing in 896 children with severe bronchiolitis. J Allergy Clin Immunol 2013.
- 897 74. Bosch A, de Steenhuijsen Piters WAA, van Houten MA, Chu M, Biesbroek G, Kool J, et al.
 898 Maturation of the Infant Respiratory Microbiota, Environmental Drivers, and Health
 899 Consequences. A Prospective Cohort Study. Am J Respir Crit Care Med 2017; 196:1582900 90.
- 901 75. Kloepfer KM, Sarsani VK, Poroyko V, Lee WM, Pappas TE, Kang T, et al. Community902 acquired rhinovirus infection is associated with changes in the airway microbiome. J
 903 Allergy Clin Immunol 2017; 140:312-5 e8.
- 904 76. Hasegawa K, Mansbach JM, Ajami NJ, Espinola JA, Henke DM, Petrosino JF, et al.
 905 Association of nasopharyngeal microbiota profiles with bronchiolitis severity in infants
 906 hospitalised for bronchiolitis. Eur Respir J 2016; 48:1329-39.
- 907 77. Rosas-Salazar C, Shilts MH, Tovchigrechko A, Schobel S, Chappell JD, Larkin EK, et al.
 908 Nasopharyngeal Lactobacillus is associated with a reduced risk of childhood wheezing
 909 illnesses following acute respiratory syncytial virus infection in infancy. J Allergy Clin
 910 Immunol 2018.
- 78. Rosas-Salazar C, Shilts MH, Tovchigrechko A, Schobel S, Chappell JD, Larkin EK, et al.
 Differences in the Nasopharyngeal Microbiome During Acute Respiratory Tract Infection
 With Human Rhinovirus and Respiratory Syncytial Virus in Infancy. J Infect Dis 2016;
 214:1924-8.
- 915 79. Folsgaard NV, Schjorring S, Chawes BL, Rasmussen MA, Krogfelt KA, Brix S, et al.
 916 Pathogenic bacteria colonizing the airways in asymptomatic neonates stimulates topical
 917 inflammatory mediator release. Am J Respir Crit Care Med 2013; 187:589-95.
- 80. Tomosada Y, Chiba E, Zelaya H, Takahashi T, Tsukida K, Kitazawa H, et al. Nasally
 administered Lactobacillus rhamnosus strains differentially modulate respiratory
 antiviral immune responses and induce protection against respiratory syncytial virus
 infection. BMC Immunol 2013; 14:40.
- 81. Sender R, Fuchs S, Milo R. Are We Really Vastly Outnumbered? Revisiting the Ratio of
 Bacterial to Host Cells in Humans. Cell 2016; 164:337-40.
- 82. Moeller AH, Caro-Quintero A, Mjungu D, Georgiev AV, Lonsdorf EV, Muller MN, et al.
 925 Cospeciation of gut microbiota with hominids. Science 2016; 353:380-2.
- 92683.Cummings JH, Macfarlane GT. The control and consequences of bacterial fermentation927in the human colon. J Appl Bacteriol 1991; 70:443-59.
- 84. Laforest-Lapointe I, Arrieta MC. Patterns of Early-Life Gut Microbial Colonization during
 Human Immune Development: An Ecological Perspective. Front Immunol 2017; 8:788.
- 93085.Fujimura KE, Lynch SV. Microbiota in allergy and asthma and the emerging relationship931with the gut microbiome. Cell Host Microbe 2015; 17:592-602.
- 86. Sokolowska M, Frei R, Lunjani N, Akdis CA, O'Mahony L. Microbiome and asthma.
 Asthma Res Pract 2018; 4:1.

- 87. McAleer JP, Kolls JK. Contributions of the intestinal microbiome in lung immunity. Eur J
 935 Immunol 2018; 48:39-49.
- 88. Copenhaver CC, Gern JE, Li Z, Shult PA, Rosenthal LA, Mikus LD, et al. Cytokine response
 patterns, exposure to viruses, and respiratory infections in the first year of life.
 Am.J.Respir.Crit Care Med. 2004; 170:175-80.
- Sumino K, Tucker J, Shahab M, Jaffee KF, Visness CM, Gern JE, et al. Antiviral IFN-gamma
 responses of monocytes at birth predict respiratory tract illness in the first year of life.
 J.Allergy Clin.Immunol. 2012; 129:1267-73.
- 942 90. Gern JE, Brooks GD, Meyer P, Chang A, Shen K, Evans MD, et al. Bidirectional
 943 interactions between viral respiratory illnesses and cytokine responses in the first year
 944 of life. J Allergy Clin Immunol 2006; 117:72-8.
- 945 91. Ly NP, Rifas-Shiman SL, Litonjua AA, Tzianabos AO, Schaub B, Ruiz-Perez B, et al. Cord
 946 blood cytokines and acute lower respiratory illnesses in the first year of life. Pediatrics
 947 2007; 119:e171-8.
- 948 92. Macaubas C, de Klerk NH, Holt BJ, Wee C, Kendall G, Firth M, et al. Association between
 949 antenatal cytokine production and the development of atopy and asthma at age 6 years.
 950 Lancet 2003; 362:1192-7.
- 93. Pelgrom LR, Everts B. Metabolic control of type 2 immunity. Eur J Immunol 2017;
 952 47:1266-75.
- 953 94. Donovan BM, Ryckman KK, Breheny PJ, Gebretsadik T, Turi KN, Larkin EK, et al.
 954 Association of newborn screening metabolites with risk of wheezing in childhood.
 955 Pediatr Res 2018.
- 95. Bufford JD, Reardon CL, Li Z, Roberg KA, Dasilva D, Eggleston PA, et al. Effects of dog
 957 ownership in early childhood on immune development and atopic diseases. Clin Exp
 958 Allergy 2008; 38:1635-43.
- 95996.Ownby DR, Johnson CC, Peterson EL. Exposure to dogs and cats in the first year of life960and risk of allergic sensitization at 6 to 7 years of age. JAMA 2002; 288:963-72.
- 97. Ludka-Gaulke T, Ghera P, Waring SC, Keifer M, Seroogy C, Gern JE, et al. Farm exposure
 962 in early childhood is associated with a lower risk of severe respiratory illnesses. J Allergy
 963 Clin Immunol 2018; 141:454-6 e4.
- 964 98. Lynch SV, Wood RA, Boushey H, Bacharier LB, Bloomberg GR, Kattan M, et al. Effects of
 965 early-life exposure to allergens and bacteria on recurrent wheeze and atopy in urban
 966 children. J Allergy Clin Immunol 2014; 134:593-601 e12.
- 967 99. Gern JE, Calatroni A, Jaffee KF, Lynn H, Dresen A, Cruikshank WW, et al. Patterns of
 968 immune development in urban preschoolers with recurrent wheeze and/or atopy. J
 969 Allergy Clin Immunol 2017.
- 970 100. Schaub B, Liu J, Hoppler S, Schleich I, Huehn J, Olek S, et al. Maternal farm exposure
 971 modulates neonatal immune mechanisms through regulatory T cells. J Allergy Clin
 972 Immunol 2009; 123:774-82 e5.
- 973 101. Stefka AT, Feehley T, Tripathi P, Qiu J, McCoy K, Mazmanian SK, et al. Commensal
 974 bacteria protect against food allergen sensitization. Proc Natl Acad Sci U S A 2014;
 975 111:13145-50.

- 976 102. Manfredo Vieira S, Hiltensperger M, Kumar V, Zegarra-Ruiz D, Dehner C, Khan N, et al.
 977 Translocation of a gut pathobiont drives autoimmunity in mice and humans. Science
 978 2018; 359:1156-61.
- 979 103. Ichinohe T, Pang IK, Kumamoto Y, Peaper DR, Ho JH, Murray TS, et al. Microbiota
 980 regulates immune defense against respiratory tract influenza A virus infection. Proc Natl
 981 Acad Sci U S A 2011; 108:5354-9.
- 982 104. Fujimura KE, Demoor T, Rauch M, Faruqi AA, Jang S, Johnson CC, et al. House dust
 983 exposure mediates gut microbiome Lactobacillus enrichment and airway immune
 984 defense against allergens and virus infection. Proc Natl Acad Sci U S A 2014; 111:805-10.
- 985105.Ciaccio CE. Modulating the microbiome: The future of allergy therapeutics? Ann Allergy986Asthma Immunol 2019; 122:233-5.
- 987 106. Wang HT, Anvari S, Anagnostou K. The Role of Probiotics in Preventing Allergic Disease.
 988 Children (Basel) 2019; 6.
- 989 107. Chiu CY, Chan YL, Tsai MH, Wang CJ, Chiang MH, Chiu CC. Gut microbial dysbiosis is
 990 associated with allergen-specific IgE responses in young children with airway allergies.
 991 World Allergy Organ J 2019; 12:100021.
- 992 108. Beigelman A, Bacharier LB. Early-life respiratory infections and asthma development:
 993 role in disease pathogenesis and potential targets for disease prevention. Curr Opin
 994 Allergy Clin Immunol 2016; 16:172-8.
- 995 109. Blanken MO, Rovers MM, Molenaar JM, Winkler-Seinstra PL, Meijer A, Kimpen JL, et al.
 996 Respiratory syncytial virus and recurrent wheeze in healthy preterm infants. N Engl J
 997 Med 2013; 368:1791-9.
- 998 110. Scheltema NM, Nibbelke EE, Pouw J, Blanken MO, Rovers MM, Naaktgeboren CA, et al.
 999 Respiratory syncytial virus prevention and asthma in healthy preterm infants: a
 1000 randomised controlled trial. Lancet Respir Med 2018; 6:257-64.
- 1001 111. Beigelman A, Isaacson-Schmid M, Sajol G, Baty J, Rodriguez OM, Leege E, et al.
 1002 Randomized trial to evaluate azithromycin's effects on serum and upper airway IL-8
 1003 levels and recurrent wheezing in infants with respiratory syncytial virus bronchiolitis. J
 1004 Allergy Clin Immunol 2015; 135:1171-8 e1.
- 1005 112. Zhou Y, Bacharier LB, Isaacson-Schmid M, Baty J, Schechtman KB, Sajol G, et al.
 1006 Azithromycin therapy during respiratory syncytial virus bronchiolitis: Upper airway
 1007 microbiome alterations and subsequent recurrent wheeze. J Allergy Clin Immunol 2016;
 108 138:1215-9 e5.
- 1009 113. Chawes BL, Bonnelykke K, Stokholm J, Vissing NH, Bjarnadottir E, Schoos AM, et al.
 1010 Effect of Vitamin D3 Supplementation During Pregnancy on Risk of Persistent Wheeze in
 1011 the Offspring: A Randomized Clinical Trial. JAMA 2016; 315:353-61.
- 1012 114. Litonjua AA, Carey VJ, Laranjo N, Harshfield BJ, McElrath TF, O'Connor GT, et al. Effect of
 1013 Prenatal Supplementation With Vitamin D on Asthma or Recurrent Wheezing in
 1014 Offspring by Age 3 Years: The VDAART Randomized Clinical Trial. JAMA 2016; 315:3621015 70.
- 1016 115. Wolsk HM, Chawes BL, Litonjua AA, Hollis BW, Waage J, Stokholm J, et al. Prenatal
 1017 vitamin D supplementation reduces risk of asthma/recurrent wheeze in early childhood:
 1018 A combined analysis of two randomized controlled trials. PLoS One 2017; 12:e0186657.

1010	446	
1019	116.	Mirzakhani H, Al-Garawi A, Weiss ST, Litonjua AA. Vitamin D and the development of
1020	117	allergic disease: how important is it? Clinical and Experimental Allergy 2015; 45:114-25.
1021	117.	Bisgaard H, Stokholm J, Chawes BL, Vissing NH, Bjarnadottir E, Schoos AM, et al. Fish Oil-
1022		Derived Fatty Acids in Pregnancy and Wheeze and Asthma in Offspring. N Engl J Med
1023	110	2016; 375:2530-9.
1024	118.	Del-Rio-Navarro BE, Espinosa Rosales F, Flenady V, Sienra-Monge JJ. Immunostimulants
1025		for preventing respiratory tract infection in children. Cochrane Database Syst Rev
1026	440	2006:CD004974.
1027	119.	Kew KM, Quinn M, Quon BS, Ducharme FM. Increased versus stable doses of inhaled
1028		corticosteroids for exacerbations of chronic asthma in adults and children. Cochrane
1029	420	Database Syst Rev 2016:CD007524.
1030	120.	Jackson DJ, Bacharier LB, Mauger DT, Boehmer S, Beigelman A, Chmiel JF, et al.
1031		Quintupling Inhaled Glucocorticoids to Prevent Childhood Asthma Exacerbations. N Engl
1032		J Med 2018; 378:891-901.
1033	121.	Brehm JM, Celedon JC, Soto-Quiros ME, Avila L, Hunninghake GM, Forno E, et al. Serum
1034		vitamin D levels and markers of severity of childhood asthma in costa rica. Am J Respir
1035		Crit Care Med 2009; 179:765-71.
1036	122.	Denlinger LC, King TS, Cardet JC, Craig T, Holguin F, Jackson DJ, et al. Vitamin D
1037		Supplementation and the Risk of Colds in Patients with Asthma. Am J Respir Crit Care
1038		Med 2016; 193:634-41.
1039	123.	Castro M, King TS, Kunselman SJ, Cabana MD, Denlinger L, Holguin F, et al. Effect of
1040		vitamin D3 on asthma treatment failures in adults with symptomatic asthma and lower
1041		vitamin D levels: the VIDA randomized clinical trial. JAMA 2014; 311:2083-91.
1042	124.	Jolliffe DA, Greenberg L, Hooper RL, Griffiths CJ, Camargo CA, Jr., Kerley CP, et al.
1043		Vitamin D supplementation to prevent asthma exacerbations: a systematic review and
1044		meta-analysis of individual participant data. Lancet Respir Med 2017; 5:881-90.
1045	125.	Bacharier LB, Guilbert TW, Mauger DT, Boehmer S, Beigelman A, Fitzpatrick AM, et al.
1046		Early Administration of Azithromycin and Prevention of Severe Lower Respiratory Tract
1047		Illnesses in Preschool Children With a History of Such Illnesses: A Randomized Clinical
1048		Trial. JAMA 2015; 314:2034-44.
1049	126.	Stokholm J, Chawes BL, Vissing NH, Bjarnadottir E, Pedersen TM, Vinding RK, et al.
1050		Azithromycin for episodes with asthma-like symptoms in young children aged 1-3 years:
1051		a randomised, double-blind, placebo-controlled trial. Lancet Respir Med 2016; 4:19-26.
1052	127.	Kantor DB, Stenquist N, McDonald MC, Schultz BJ, Hauptman M, Smallwood CD, et al.
1053		Rhinovirus and serum IgE are associated with acute asthma exacerbation severity in
1054		children. J Allergy Clin Immunol 2016; 138:1467-71 e9.
1055	128.	Teach SJ, Gill MA, Togias A, Sorkness CA, Arbes SJ, Jr., Calatroni A, et al. Preseasonal
1056		treatment with either omalizumab or an inhaled corticosteroid boost to prevent fall
1057		asthma exacerbations. J Allergy Clin Immunol 2015; 136:1476-85.
1058	129.	Kantor DB, McDonald MC, Stenquist N, Schultz BJ, Smallwood CD, Nelson KA, et al.
1059		Omalizumab Is Associated with Reduced Acute Severity of Rhinovirus-triggered Asthma
1060		Exacerbation. Am J Respir Crit Care Med 2016; 194:1552-5.
1061	130.	Welliver RC, Kaul TN, Ogra PL. The appearance of cell-bound IgE in respiratory-tract
1062		epithelium after respiratory-syncytial-virus infection. N Engl J Med 1980; 303:1198-202.

1063	131.	Welliver RC, Wong DT, Sun M, Middleton E, Jr., Vaughan RS, Ogra PL. The development
1064		of respiratory syncytial virus-specific IgE and the release of histamine in nasopharyngeal
1065		secretions after infection. N Engl J Med 1981; 305:841-6.
1066	132.	Tam JS, Jackson WT, Hunter D, Proud D, Grayson MH. Rhinovirus specific IgE can be
1067		detected in human sera. J Allergy Clin Immunol 2013; 132:1241-3.
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1070 1071 Table 1. Review of most salient points 1072 1073 1. Polymorphisms in several antiviral and innate immune genes have been linked to 1074 susceptibility to respiratory viruses, infection severity, and virus-induced asthma exacerbations, and have been replicated across multiple cohorts; these genes include STAT4, JAK2, MX1, 1075 1076 VDR, DDX58, and EIF2AK2. 1077 1078 2. Rhinovirus virulence varies by species; RV-A and RV-C are more likely to cause illnesses, wheezing and lower respiratory tract infection compared to RV-B. 1079 1080 1081 3. RV infection leads to expression of epithelial derived cytokines IL-25, IL-33, and TSLP and 1082 an increase in ILC2s as an important source of airway IL-13; blocking these pathways with anti-1083 IL-25 attenuates neonatal RV-induced AHR and mucous cell metaplasia in mice. 1084 1085 4. Two prevention studies using palivizumab (a monoclonal antibody directed against RSV) in 1086 high-risk infants found that prevention of more severe RSV-illnesses decreased the risk of 1087 childhood recurrent wheezing but not asthma development. 1088 1089 5. In several infant cross-sectional and cohort studies, the presence of Streptococcus, 1090 Moraxella, or Haemophilus during upper respiratory infection increases the likelihood that the infant will have lower airway symptoms; studies examining airway bacteria during RSV 1091 1092 bronchiolitis have reported links between an increased abundance of Streptococcus and 1093 Haemophilus, while in contrast, RV-bronchiolitis is associated with an increased abundance of 1094 Moraxella and Haemophilus. 1095 1096 6. The presence of *H. influenzae* in the infant airway prior to viral infection is associated with 1097 increased expression of local inflammatory cytokines suggesting that a link exists between 1098 bacteria and airway inflammation; in contrast, mice receiving intranasal administration of Lactobacillus rhamnosus prior to viral infection have an enhanced antiviral immune response⁸⁰, 1099 1100 suggesting that some bacteria may protect the airway and help prevent viral infection. 1101 1102 7. The gut microbiome also regulate pulmonary anti-viral immunity; in a murine model intact 1103 commensal bacteria in the gut were required for adaptive immune responses to respiratory 1104 influenza virus infection. 1105 1106 8. Unique components of the viral genome contribute to respiratory illness and knowledge of 1107 these factors may also assist in vaccine and therapeutic strategies aimed at the proteins responsible for specific disease characteristics. 1108 1109 1110 9. Omalizumab, a humanized monoclonal antibody that selectively binds to IgE, decreased fall asthma exacerbations in atopic asthmatics and increased IFN-a responses to RV ex vivo in the 1111 Preventative Omalizumab or Step-Up Therapy for Fall Exacerbations (PROSE) study, whereas 1112 1113 boosting ICS did not reduce risk. 1114 1115 10. A recent meta-analysis that combined two clinical trials (VDAART, COPSAC2010) investigated whether maternal vitamin D supplementation (2400 IU/day, 4000IU/day) during 1116 pregnancy revealed that this intervention resulted in a 25% significant reduction in 1117 1118 asthma/recurrent wheeze risk during the first 3 years of life. 1119

1120	Table 3. Future/ongoing interventional studies examining treatments for viral triggered
1121	asthma

			Primary Outcome	Estimated Completion	
Study Title	Study Population	Intervention	Measurement	Date	
		Baseline and 3.5			
Miteraia Dita da		month high dose	Niversite and a f		
Vitamin D In the Prevention of Viral-	children age 1-<6 with	vitamin D 100,000	Number of courses of	December	
Induced Asthma in	recurrent cold triggered asthma attacks, expected	IU and daily Vitamin D dose 400 IU	rescue oral	2022	
preschoolers	enrollment 865 subjects	OR placebo	steroids (OCS)	2022	
			over 7 months	enrolling	
			Time to		
			occurrence of		
Azithromycin to Prevent	children 1-18 months of	Azithromycin	a 3rd episode		
Wheezing Following	age, hospitalized due to	(10 mg/kg x 7 days	of post-RSV	December	
Severe RSV	RSV bronchiolitis,	followed by 5 mg/kg	wheezing,	2021	
Bronchiolitis II	expected enrollment 200	x 7days)	observation over 48	notwot	
	subjects	OR placebo	months	not yet enrolling	

Figure 1. Immune responses to virus in the allergic asthmatic host. In the healthy host, anti-viral
IFN responses control and clear respiratory viral infections. In allergic asthmatics, the release of
the type 2-skewing cytokines TSLP, IL-25, and IL-33 promote the induction of Th2 cytokines
and the suppression of IFN responses, in addition to promoting airway hyperreactivity (AHR)
and increased mucus and IgE production. Furthermore, IgE has the capacity to suppress IFN-α
production by Plasmacytoid DCs (pDCs).

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Table 1. Review of most salient points

1. Polymorphisms in several antiviral and innate immune genes have been linked to susceptibility to respiratory viruses, infection severity, and virus-induced asthma exacerbations, and have been replicated across multiple cohorts; these genes include *STAT4*, *JAK2*, *MX1*, *VDR*, *DDX58*, and *EIF2AK2*.

2. Rhinovirus virulence varies by species; RV-A and RV-C are more likely to cause illnesses, wheezing and lower respiratory tract infection compared to RV-B.

3. RV infection leads to expression of epithelial derived cytokines IL-25, IL-33, and TSLP and an increase in ILC2s as an important source of airway IL-13; blocking these pathways with anti-IL-25 attenuates neonatal RV-induced AHR and mucous cell metaplasia in mice.

4. Two prevention studies using palivizumab (a monoclonal antibody directed against RSV) in high-risk infants found that prevention of more severe RSV-illnesses decreased the risk of childhood recurrent wheezing but not asthma development.

5. In several infant cross-sectional and cohort studies, the presence of *Streptococcus*, *Moraxella*, or *Haemophilus* during upper respiratory infection increases the likelihood that the infant will have lower airway symptoms; studies examining airway bacteria during RSV bronchiolitis have reported links between an increased abundance of *Streptococcus* and *Haemophilus*, while in contrast, RV-bronchiolitis is associated with an increased abundance of *Moraxella* and *Haemophilus*.

6. The presence of *H. influenzae* in the infant airway prior to viral infection is associated with increased expression of local inflammatory cytokines suggesting that a link exists between bacteria and airway inflammation; in contrast, mice receiving intranasal administration of *Lactobacillus rhamnosus* prior to viral infection have an enhanced antiviral immune response⁸⁰, suggesting that some bacteria may protect the airway and help prevent viral infection.

7. The gut microbiome also regulate pulmonary anti-viral immunity; in a murine model intact commensal bacteria in the gut were required for adaptive immune responses to respiratory influenza virus infection.

8. Unique components of the viral genome contribute to respiratory illness and knowledge of these factors may also assist in vaccine and therapeutic strategies aimed at the proteins responsible for specific disease characteristics.

9. Omalizumab, a humanized monoclonal antibody that selectively binds to IgE, decreased fall asthma exacerbations in atopic asthmatics and increased IFN- α responses to RV *ex vivo* in the Preventative Omalizumab or Step-Up Therapy for Fall Exacerbations (PROSE) study, whereas boosting ICS did not reduce risk.

10. A recent meta-analysis that combined two clinical trials (VDAART, COPSAC2010) investigated whether maternal vitamin D supplementation (2400 IU/day, 4000IU/day) during pregnancy revealed that this intervention resulted in a 25% significant reduction in asthma/recurrent wheeze risk during the first 3 years of life.

Table 2. Polymorphisms in several antiviral and innate immune genes have been linked to susceptibility to respiratory viruses, infection severity, and virus-induced asthma exacerbations, and have been replicated across multiple cohorts.

Gene	Function
STAT4	Transcription factor required for IL-12 signaling in the development of Th1 cells from naïve CD4 T cells
JAK2	A non-receptor tyrosine kinase critical for signaling of the GM-CSF, gp130, and single chain receptor families.
MX1	Responsible for the antiviral state against influenza infection
VDR	Vitamin D receptor
DDX58	Involved in antiviral signaling in response to viruses containing a dsDNA genome
EIF2AK2	Innate antiviral immune response to viral infection that can trigger apoptosis via FADD-mediated caspase 8
IRF7	Critical role in the innate immune response against DNA and RNA viruses
IFIH1	Provides instructions for making the MDA5 protein that has a critical role in innate antiviral immunity
IFNA5	One of the type I IFN-α isoforms that has antiviral activities
VDR	Vitamin D receptor
DDX58	Involved in antiviral signaling in response to viruses containing a dsDNA genome
EIF2AK2	Innate antiviral immune response to viral infection that can trigger apoptosis via FADD-mediated caspase 8
IRF7	Critical role in the innate immune response against DNA and RNA viruses
IFIH1	Provides instructions for making the MDA5 protein that has a critical role in innate antiviral immunity

	ble 3. Future/ongoing interventional studies examining treatments for viral triggered thma	
as	unna	

			Primary	Estimated
			Outcome	Completion
Study Title	Study Population	Intervention	Measurement	Date
		Baseline and 3.5		
		month high dose		
Vitamin D In the	children age 1-<6 with	vitamin D 100,000	Number of	
Prevention of Viral-	recurrent cold triggered	IU and daily Vitamin	courses of	December
Induced Asthma in	asthma attacks, expected	D dose 400 IU	rescue oral	2022
preschoolers	enrollment 865 subjects	OR placebo	steroids (OCS)	
			over 7 months	enrolling
			Time to	
			occurrence of	
Azithromycin to Prevent	children 1-18 months of	Azithromycin	a 3rd episode	
Wheezing Following	age, hospitalized due to	(10 mg/kg x 7 days	of post-RSV	December
Severe RSV	RSV bronchiolitis,	followed by 5 mg/kg	wheezing,	2021
Bronchiolitis II	expected enrollment 200	x 7days)	observation	
	subjects	OR placebo	over 48	not yet
		*	months	enrolling

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