

# Spawn characteristics in *Adelomelon ferussacii* (Donovan, 1824) (Gastropoda: Volutidae) from southern Patagonia, Argentina

Pablo E. Penchaszadeh

María Eugenia Segade

Museo Argentino de Ciencias Naturales, CONICET  
Av. Angel Gallardo 470  
Buenos Aires, ARGENTINA  
pablop@mail.retina.ar

## ABSTRACT

South American volutids are very homogeneous with regard to their reproductive mode. These gastropods generally spawn egg capsules containing few eggs; the embryos feed on substances contained in the intracapsular fluid and hatch as crawling juveniles. *Adelomelon ferussacii* lives on subtidal mud or sandy bottoms, yet the egg capsules collected in San Julián, Santa Cruz, Argentina, were found on flat smooth subtidal rocks. The egg capsule is globose and hemispherical, flexible, opaque-white, and the attachment base is wide, measuring between 15–30 mm in diameter. One to six eggs were recorded inside each egg capsule. The embryonic development occurred in the interior of the capsule and eight stages are described. Crawling juveniles, with shells measuring between 11.25–14.8 mm, were observed at the last stage before hatching. Also a gregarious spawning event is recorded for the first time in the South American volutes.

*Additional keywords:* Neogastropoda, egg capsules, development, hatching size, gregarious spawning

## INTRODUCTION

South American volutids are relatively homogenous with regard to their reproductive biology. Female volutes produce single, large egg capsules with relatively few eggs that are attached to hard substrates. Embryos develop until metamorphosis and hatch as crawling juveniles. Juveniles usually exceed 10 mm in total shell length, originating from eggs smaller than 300  $\mu$ m that are suspended with extra-vitelline substances such as albumen in the intracapsular liquid (Penchaszadeh and De Mahieu, 1976). Yet, there are exceptions, such as *Voluta virescens* (Lightfoot, 1786), which is reported to spawn egg capsules containing about 200 eggs. Of these, only one or two develop further, ingesting the others as nurse eggs (Bandel, 1976).

The spawnings of three species of the genus *Adelomelon* in the southwestern Atlantic have been described to date. *Adelomelon brasiliiana* (Lamarck, 1811) has the largest known unattached caenogastropod egg capsules,

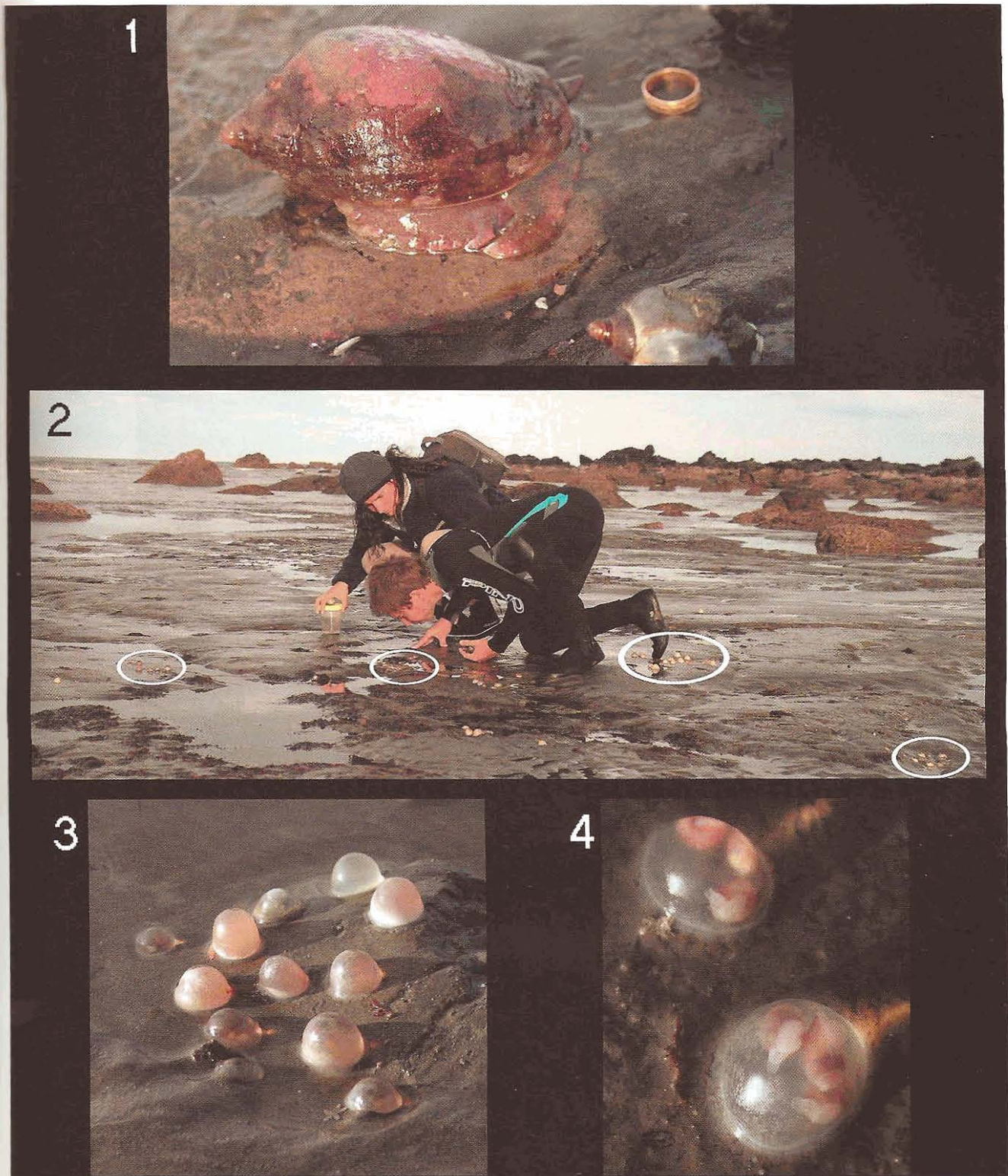
with diameters between 40–80 mm and internal volumes of up to 140 ml (Penchaszadeh and De Mahieu, 1976). *Adelomelon ancilla* (Lightfoot, 1786) have oval and flat egg capsules, which are attached to hard substrates. The minor and major axes of their bases measure between 25–44 mm and 27–46 mm, respectively, and their internal volumes may reach four milliliters (Penchaszadeh et al., 1999). *Adelomelon beckii* (Broderip, 1836) have globose hemispherical egg capsules that are also attached to hard substrates, usually the external surfaces of empty scallop shells. Egg capsules measure approximately 50 mm in basal diameter and have internal volumes between 30–35 ml (Penchaszadeh et al., 1999).

*Adelomelon ferussacii* (Donovan, 1824) are distributed from 42° S (Gulf San Matias) to 52° S (Straits of Magellan) (Carcelles and Williamson, 1951), corresponding to the Magellanic biogeographical province. Little is known about this species, which lives below the low water line on mud or sandy bottoms. This study describes the egg capsules and the first stages of development of representatives of *A. ferussacii*.

## MATERIALS AND METHODS

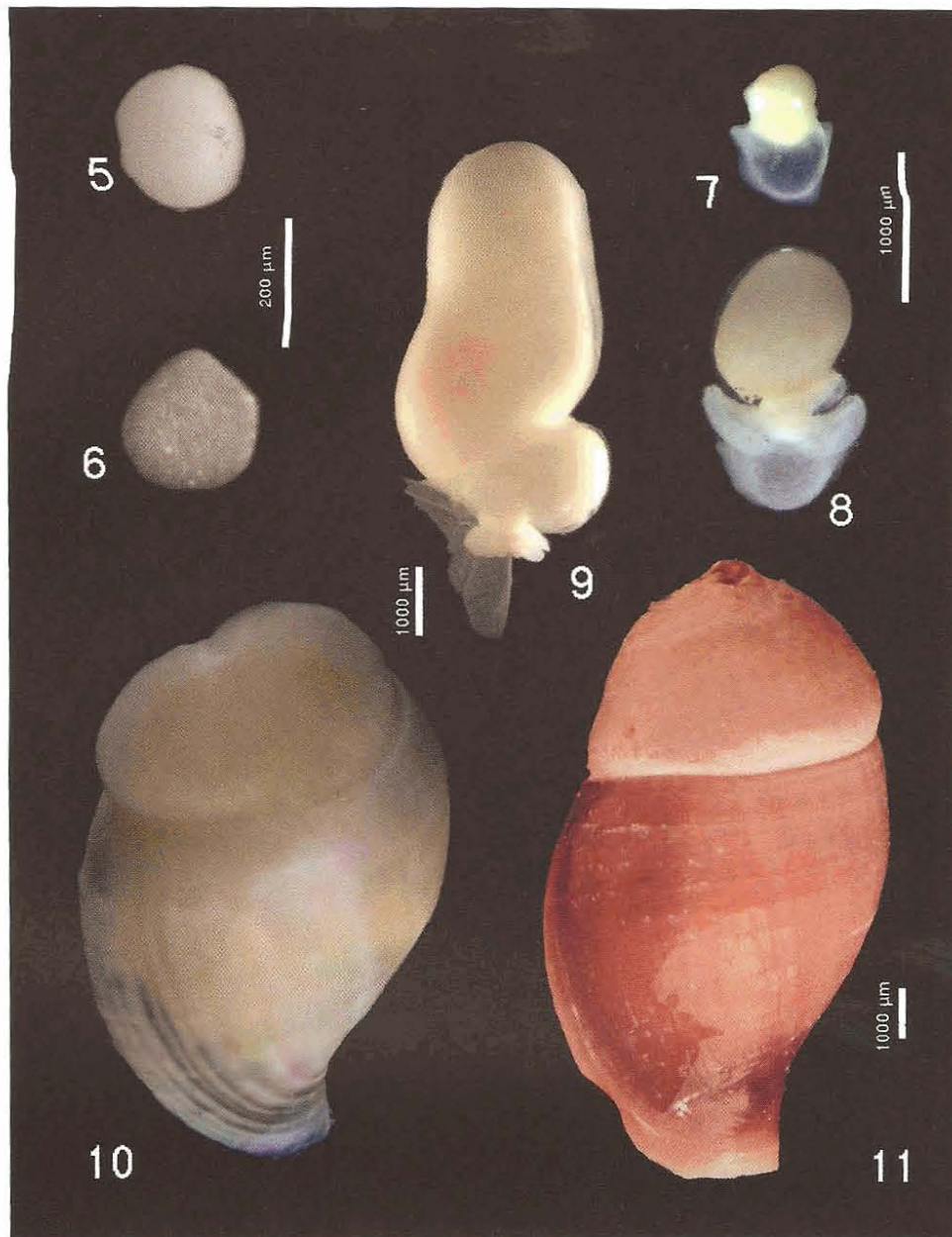
A total of 95 egg capsules of *Adelomelon ferussacii* were collected during the austral summer by free-diving during low tides (2–3 m depth) at La Cascada in January 2005, and manually from areas exposed during an extraordinary low tide event in February 2006 at La Mina Beach, both located in San Julián, Santa Cruz Province (respectively 67°43' W, 49°19' S and 67°40' W, 49°15' S). The water temperature at the time of collection was 15°C. Egg capsules were collected from the rocky bottom by hand with the help of a spatula, preserved in individual jars in 70 % ethanol, and examined under dissecting and transmission optical microscopes, as needed. The diameter and height of each egg capsule was measured using a Vernier caliper, and each internal volume was measured by carefully extracting the intracapsular liquid with a Pasteur pipette. Egg capsules were opened by cutting along their base line using a small surgical scissors. The number and stage of each





**Figures 1–4.** Habitat and egg capsules of *Adelomelon ferussacii*. 1. Adult at low tide. 2. Panoramic view of “Playa La Mina”; egg capsules of *A. ferussacii* are exposed during low tide. Circles indicate clusters of egg capsules. 3. Detail of an egg capsule cluster. 4. Egg capsules with juveniles close to hatching. Photographed by Natalie Collin.





**Figures 5–11.** Development of *Adelomelon ferussacii*. 5. Uncleaved egg. 6. Morula stage. 7. First “veliger” stage. 8. Second “veliger” stage. 9. Late embryo without shell. 10. Late embryo with calcified shell. 11. Pre-juvenile close to hatching.

embryo was recorded and photographs of each stage were obtained through the microscope.

## RESULTS

**SPAWNING SITES:** *Adelomelon ferussacii* (Figure 1) lays egg capsules on rocky bottoms. At La Cascada, where there is sandy-mud bottom, they were attached to flagstone slabs. At La Mina Beach, egg capsules were attached to the flat rocky bottom. Communal spawning was observed at both sites. The aggregation of spawning females results in a spawn cluster of more than 20 egg

capsules (Figure 2), which indicates a gregarious behavior for spawning. Within the spawn cluster, individual egg capsules showed different developmental stages.

**CHARACTERISTICS OF THE EGG CAPSULE:** The spawn consists of a single egg capsule attached to hard substrate, either a flagstone slab or another type of flat, rocky substrate. The egg capsule is globose, hemispherical and flexible, with a white opaque color (Figures 3, 4). It had a basal minor axis measuring 15–18 mm ( $N=95$ ), basal major axis measuring 29–31 mm ( $N=95$ ), and height of 11–21 mm ( $N=95$ ). The internal volume of egg capsules was 1.2–6.0 ml ( $N=95$ ) (Table 1). No exit plug or escape aperture was observed in



**Table 1.** Dimensions of the egg capsule of *Adelomelon ferussacii*.

	N	Mean	Max.	Min.	SD
Diameter (mm)	95	21 × 23	29 × 31	15 × 18	2.30 × 2.52
Height (mm)	95	15.52	21	11	2.25
Int. volume (ml)	95	2.74	6	1.2	0.89

any capsule, only a suture line on one side of the capsules. The base is round with a narrow margin (~3 mm). No external calcareous layer was present.

#### CHARACTERISTICS OF THE EARLY DEVELOPMENTAL STAGES:

Out of the 95 egg capsules collected, only 61 contained embryos. The majority of the embryos were found in late developmental stages. Between one and six embryos per egg capsule were found, with a mode of three (mean = 2.8; SD = 1.1; N = 61) (Table 2). The following stages of development were identified: uncleaved egg; eight-cells; morula; "veliger I"; "veliger II"; late embryo without shell; late embryo with shell; and pre-juveniles close to hatching (Figures 5–11). The uncleaved egg diameter was 220 µm (N = 1), the eight-cell diameter was 220 µm (N = 1), the embryos in the morula stage measured 210–240 µm diameter (mean = 224 µm; N = 5). Those embryos in "veliger I" measured 750–950 µm in length (mean = 810 µm; N = 4); "veliger II" 1250–3500 µm in length (mean = 1860 µm; N = 21); embryos without shell 5–15 mm in length (mean = 8.3 mm; N = 98). The embryos presenting calcified shells ranged between 7.5–12.5 mm total shell length (mean = 9.7 mm; N = 34) and embryos close to hatching between 11.2–14.8 mm in total length (mean = 13.1 mm; N = 9) (Table 3).

#### DISCUSSION

Information on the spawning of volutids is scarce not only for South American species, but also for those from other regions of the world. As a general rule, South American volutids show little variation with regard to their reproductive patterns. Commonly, the egg capsules are attached to hard substrates; the fact that *Adelomelon brasiliiana* spawns free eggs capsules is a remarkable adaptation to shallow sandy bottoms, given that they may

**Table 2.** Frequency of number of embryos per egg capsule in *Adelomelon ferussacii* collected in January 2005 and February 2006 in "La Cascada" and "La Mina" beach, San Julian, Argentina (N = 95).

N° embryos	Frequency
0	34
1	7
2	15
3	24
4	12
5	2
6	1
more	0

**Table 3.** Size at different stages of development identified for *Adelomelon ferussacii*.

Stages	n	Mean
Uncleaved egg	1	220 µm
8 cells	1	220 µm
Morule	5	224 µm
Veliger I	4	0.8 mm
Veliger II	21	1.8 mm
Embryo without shell	98	8.3 mm
Embryo with shell	34	9.7 mm
Close to hatching	9	13.0 mm

be carried away by the currents but are never buried in the sand (Penchaszadeh and De Mahieu, 1976).

*Adelomelon ferussacii* lives in shallow water, on mixed or soft bottoms along the Magellanic biogeographical province. The only available information on this species is based on very few specimens and mainly on shell features (e.g., Clench and Turner, 1964; Weaver and du Pont, 1970). As with all the other studied South American volutids, except for a single report on *Voluta virescens* (Bandel, 1976), *Adelomelon ferussacii* spawns egg capsules containing few eggs. The embryos feed on substances contained in the intracapsular fluid. Development is direct (intracapsular metamorphosis) and crawling hatchlings may have a shell length of more than 10 mm (Carcelles, 1944; De Mahieu et al., 1974; Penchaszadeh and De Mahieu, 1976; Penchaszadeh, 1988; Hain, 1992; Penchaszadeh et al., 1999).

The diameter of the eggs of *Adelomelon ferussacii*, including the uncleaved egg, eight-cells and morula stages, is about 220 µm. This size is smaller than the egg sizes reported for *Voluta musica* Linnaeus, 1758 (330 µm) (Penchaszadeh and Miloslavich, 2001). However, the egg size we measured is similar to those sizes reported for *Adelomelon brasiliiana* (Lamarck, 1811) (240 µm), *A. ancilla* (Lightfoot, 1786) (200–220 µm) (Penchaszadeh and De Mahieu, 1976), and *Odontocymbiola magellanica* (Gmelin, 1791) (210 µm) (Bigatti, 2005), but larger than those of *Zidona dufresnei* (Donovan, 1823) (90 µm) (Penchaszadeh and de Mahieu, 1976).

The embryological development is similar to those described for *A. brasiliiana* and *A. ancilla* by Penchaszadeh and de Mahieu (1976), with presence of a poorly developed velum. This contrasts with *Voluta musica* Linnaeus, 1758, which has a well-developed and wide intracapsular velum, the largest of the studied volutids (Penchaszadeh and Miloslavich, 2001).

Gastropod egg capsules are morphologically and chemically complex; they provide mostly protection against bacterial attacks, environmental stress, and predation (Pechenik 1979, 1986; Miloslavich 1996). Despite this, studies show that gastropod egg capsules are targets for predation by fish, crustaceans, polychaetes, and even other gastropods (D'Asaro, 1970). In this study, preyed-upon *Adelomelon ferussacii* egg capsules were observed. These were found lacerated mainly on their upper portions, probably by sea birds such as *Larus dominicanus* (Lich-



tenstein, 1823) and *Haematopus ater* Vieillot and Oudart, 1825, which were observed pecking on the egg capsules when these were exposed at low tides. Bird predation on volute stranded free egg capsules (*Adelomelon brasiliense*) was studied by Penchaszadeh et al. (2000).

*Adelomelon ferussacii* egg capsules lack an external calcium carbonate cover such as found in the common Patagonian *Odontocymbiola magellanica* (see Bigatti, 2005); this would increase their susceptibility to predation.

Gregarious behavior was observed for the spawning of *A. ferussacii*, as has been reported for several caenogastropod species such as *Engoniophos uncinatus* Say, 1825 (Miloslavich and Penchaszadeh, 1994), *Fusinus closter* Philippi, 1850 (Miloslavich and Penchaszadeh, 1997), and *Chicoreus margaritensis* (Abbott, 1958) (Cipriani, 1990). This conclusion is based on observations of presence of patches of egg capsules in different developmental stages along the shore. This is, to our knowledge, the first report of this behavior in the family Volutidae.

#### ACKNOWLEDGMENTS

We are grateful to Dr. Bayden Russell, University of Adelaide, and Guido Pastorino (MACN) for suggestions that improved the manuscript. We appreciate the assistance of Natalie Collm and Juan Pablo Livore in the field, and of Carlos Sanchez Antelo and Diego Urteaga in the laboratory. This work benefited from grants PICT 10975, PICT 14419 and GEF AC-56.

#### LITERATURE CITED

- Bandel, K. 1976. Spawning, development and ecology of some higher Neogastropoda from the Caribbean Sea of Colombia. *The Veliger* 19: 176–193.
- Bigatti, G. 2005. Reproducción y Ecología del caracol rojo *Odontocymbiola magellanica* (Gastropoda: Volutidae) en Golfo Nuevo, Patagonia. PhD thesis. Universidad de Buenos Aires.
- Carcelles, A.R. 1944. Catálogo de los moluscos marinos de Puerto Quequén. *Revista Museo de La Plata* 3: 233–309, 15 pls.
- Carcelles, A.R. and S.I. Williamson. 1951. Catálogo de los Moluscos Marinos de la Provincia Magallánica. *Revista del Instituto de Investigaciones de las Ciencias Naturales, Zoology* 2: 225–383.
- Cipriani, R. 1990. Aspectos de la reproducción de *Murex margaritensis* (Mollusca, Gasterópoda) en Venezuela. Tesis de Licenciatura. Universidad Simón Bolívar.
- Clench, W.J. and R.D. Turner. 1964. The subfamilies Volutinae, Zidoninae, Odontocymbiolinae and Calliotelectinae in the Western Atlantic. *Johnsonia*, 4(43): 129–180.
- D'Asaro, C.N. 1970. Egg capsules of prosobranch mollusks from South Florida and the Bahamas and notes on spawning in the laboratory. *Bulletin of Marine Science* 20: 414–440.
- De Mahieu, G.C., P.E. Penchaszadeh, and A. Casal. 1974. Algunos aspectos de las variaciones de proteínas y aminoácidos libres totales del líquido intracapsular en relación al desarrollo embrionario en *Adelomelon brasiliense* (Lamarek, 1811). *Cahiers de Biologie Marine* 15: 215–227.
- Hain, S. 1992. Maintenance and culture of living benthic molluscs from high Antarctic shelf areas. *Aquaculture and Fisheries Management* 23: 1–11.
- Miloslavich, P. and P.E. Penchaszadeh. 1994. Spawn and development of *Engoniophos uncinatus* (Say, 1825) (Gastropoda: Prosobranchia) from the southern Caribbean Sea. *The Veliger* 37: 425–429.
- Miloslavich, P. 1996. Nurse-egg feeding prosobranchs: a comparative biochemical and electrophoretic analysis of eggs and hatchlings. *American Malacological Bulletin* 13: 37–46.
- Miloslavich, P. 1996. Biochemical composition of prosobranch egg capsules. *Journal of Molluscan Studies* 62: 133–136.
- Miloslavich, P. and P.E. Penchaszadeh. 1997. Spawn and development of *Fusinus closter* Philippi 1850 (Gastropoda, Prosobranchia) from Venezuelan Caribbean. *The Veliger* 40: 93–100.
- Miloslavich, P. 1999. Nutritional value of the intracapsular liquid of *Engoniophos uncinatus* Say, 1825 (Caenogastropoda: Buccinidae). *Journal of Molluscan Studies* 65: 502–503.
- Pechenik, J.A. 1979. Role of encapsulation in invertebrate life histories. *American Naturalist* 114: 859–870.
- Pechenik, J.A. 1986. The encapsulation of eggs and embryos by molluscs: an overview. *American Malacological Bulletin* 4: 165–172.
- Penchaszadeh, P.E. 1988. Reproductive patterns of some South American Prosobranchia as a contribution to classification. *Malacological Review, suppl.* 4: 284–287.
- Penchaszadeh, P.E., F. Botto, and O. Iribarne. 2000. Shorebird feeding on stranded giant Gastropod egg capsules of *Adelomelon brasiliense* (Volutidae) in coastal Argentina. *Journal of Shellfish Research* 15: 901–904.
- Penchaszadeh, P.E. and G.C. De Mahieu. 1976. Reproducción de gasterópodos Prosobranchios del Atlántico suroccidental. Volutidae. *Physis, sección A* 35(91): 145–153.
- Penchaszadeh, P.E., P. Miloslavich, M. Lasta, and P.M.S. Costa. 1999. Egg capsules in the genus *Adelomelon* (Caenogastropoda: Volutidae) from the Atlantic coast of South America. *The Nautilus* 113: 56–63.
- Penchaszadeh, P.E. and P. Miloslavich. 2001. Embryonic stages and feeding substances of the South American volutid *Voluta musica* (Caenogastropoda) during intracapsular development. *American Malacological Bulletin* 16: 21–31.
- Weaver, C.S. and J.E. du Pont. 1970. Living Volutes: A Monograph of the Recent Volutidae of the World. Delaware Museum of Natural History. Monograph Series 1, Greenville, xv+375 pp.