

1 **The addition of calcium chloride in combination with a lower draining pH to change the**
2 **microstructure and improve fat retention in Cheddar cheese**

3

4 Lydia Ong^{a, b, c}, Kevany Soodam^{a, b}, Sandra E. Kentish^{a, c}, Ian B. Powell^d and Sally L. Gras^{a, b, c, *}

5 ^aParticulate Fluid Processing Centre, Department of Chemical and Biomolecular Engineering,
6 The University of Melbourne, Parkville, Vic 3010, Australia.

7 ^bThe Bio21 Molecular Science and Biotechnology Institute, The University of Melbourne,
8 Parkville, Vic 3010, Australia.

9 ^cARC Dairy Innovation Hub, Department of Chemical and Biomolecular Engineering, The
10 University of Melbourne, Parkville, Vic 3010, Australia.

11 ^dDairy Innovation Australia Limited, Werribee, Vic 3030, Australia.

12

13 *Corresponding author: sgras@unimelb.edu.au (S.L. Gras)

14 Tel.: +61-3-8344 6281; fax: +61-3-8344 4153

15 **ABSTRACT**

16 Calcium chloride addition and the whey draining pH are known to impact on cheese making. The
17 effect of 100 or 300 mg kg⁻¹ calcium chloride (CaCl₂) and the whey draining pH (6.2 and 6.0)
18 on the microstructure of Cheddar cheese was assessed using confocal and cryo scanning electron
19 microscopy. The gel made with 300 mg kg⁻¹ CaCl₂ was found to have a denser protein network
20 and smaller pores than the gel with lower or no CaCl₂ addition. CaCl₂ addition reduced fat lost
21 to the sweet whey. The texture of the cheeses with a lower draining pH was harder and moisture
22 content lower. Our results show that the combination of calcium addition and lower draining pH
23 could be used to increase network formation at the early stages of cheese making to improve fat
24 retention while maintaining a similar level of total calcium in the final cheese.

25 1. Introduction

26 Calcium chloride (CaCl_2) is added to cheese milk to assist milk gelation (Okigbo,
27 Richardson, Brown & Ernstrom, 1985; Ustunol & Hicks, 1990), to improve the cheese making
28 process (Gastaldi, Pellegrini, Lagaude & Fuente, 1994) and/or to increase cheese yield
29 (Wolfschoon-Pombo, 1997). The effect of the concentration of calcium (Ca) on rennet induced
30 coagulation has been reported in various studies (Dalglish, 1983; Lucey & Fox, 1993; Udabage,
31 McKinnon & Augustin, 2001; Zoon, van Vliet & Walstra, 1988) where a higher calcium
32 concentration results in a faster rennet coagulation due to the combined effect of the increased
33 calcium ion activity and a drop in milk pH, leading to changes in cheese properties (Ong,
34 Dagastine, Kentish & Gras, 2013a).

35 The level of CaCl_2 added varies between manufacturers and regions. Typically, a
36 concentration of 100-200 mg CaCl_2 kg^{-1} of milk is added (Gastaldi et al., 1994; Okigbo et al.,
37 1985; Wolfschoon-Pombo, 1997), although this may reach up to 500 mg kg^{-1} in experimental
38 studies (Ustunol & Hicks, 1990). The United States Federal standard limits the addition of CaCl_2
39 up to a level of 200 mg kg^{-1} (Federal Register, 1982), while other codes such as the Australian
40 and New Zealand food standard or Codex Alimentarius do not specify limits. Addition of 200-
41 300 mg kg^{-1} CaCl_2 has been shown to produce a denser gel network, which significantly
42 increases the fat retained within the curd and decreases the fat lost to sweet whey (Ong et al.,
43 2013a). More calcium is also present in the final cheese when more CaCl_2 is added to the milk.
44 Increasing the concentration of calcium, however, may lead to undesirable properties such as the
45 formation of calcium lactate crystals (CLC), a common defect in Cheddar cheese (Phadungath &
46 Metzger, 2011).

47 The concentration of Ca in the final cheese can also affect the texture of the cheese
48 immediately after pressing and during ripening. A higher level of total Ca increases cross-linking
49 between casein micelles, leading to a harder gel (Choi, Horne, Johnson & Lucey, 2008).
50 Conversely, the loss of insoluble Ca may increase repulsion between the exposed casein micelles
51 phosphoserine residues resulting in a weaker gel network (Choi et al., 2008). The changes in gel
52 properties may then in turn affect the textural properties of the cheese. Consequently, cheese
53 with a low level of Ca, such as Cheshire cheese, tends to be crumbly, whereas cheese with high
54 level of Ca, such as Gouda or Swiss, tends to be more rubbery and elastic (Lucey & Fox, 1993).

55 The draining pH is known to have a large influence on the mineral content of a cheese
56 and decreases in draining pH can decrease the mineral content (Dalglish & Law, 1989; Guinee,
57 Feeney, Auty & Fox, 2002; Lucey & Fox, 1993). This change influences cheese texture and also
58 the amount of lactose remaining in the curd and consequently the concentration of lactic acid and
59 final pH, as well as the retention of some types of coagulant (e.g. retention of more chymosin at
60 lower pH). This retention of rennet has implications for the proteolysis of cheese during
61 maturation (Holmes, Duersch & Ernstrom, 1977). The analysis of mozzarella by scanning
62 electron microscopy has shown that lowering the draining pH from 6.4 to 6.15 or to 5.9 resulted
63 in an increase in the fusion of para-casein particles, producing a more continuous three
64 dimensional network (Kiely, Kindstedt, Hendricks, Levis, Yun & Barbano, 1992). A lower whey
65 draining pH in buffalo-milk mozzarella production (pH 5.3 vs pH 6.2) was also found to increase
66 the yield of the cheese but the decrease in the level of total calcium adversely affected the
67 melting properties of the cheese (Yazici & Akbulut, 2007).

68 The aim of this study was to investigate the effect of calcium chloride addition in
69 combination with the lowering of the draining pH. Although the effect of CaCl₂ addition or
70 draining pH has been well described for different cheeses, including Cheddar and Gouda cheese
71 (Ong et al., 2013a; Walstra, Wouters & Geurts, 2006), the combination of these two processing
72 parameters has not been investigated. These process adjustments may provide an approach to
73 improve fat retention and cheese yield whilst reducing the concentration of calcium in the final
74 cheese in order to avoid defect formation. The microstructure of the gel, cooked curd, milled
75 curd and the final cheese were a specific focus of this work. Studies investigating the effect of
76 draining pH on the microstructure of cheese are limited and have only focussed on mozzarella
77 cheese to date (Kiely et al., 1992). This study uses the combination of confocal laser scanning
78 microscopy (CLSM) and cryo scanning electron microscopy (cryo SEM) to examine the effect of
79 calcium and draining pH. The composition and the texture of the cheese are also investigated.

80

81 **2. Materials and methods**

82

83 **2.1. Cheese making**

84 Six batches of Cheddar cheeses (Batches 1-6) were made in random order over three days
85 at the Warrnambool Cheese and Butter pilot plant (WCB, Allansford, Australia) with different

86 levels of calcium chloride addition: 0 mg kg⁻¹ cheese milk (Batch 1 control cheese and Batch 4),
87 100 mg kg⁻¹ cheese milk (Batches 2 and 5) and 300 mg kg⁻¹ cheese milk (Batches 3 and 6) and
88 different whey draining pH (Batches 1-3 drained at pH 6.2 and Batches 4-6 drained at pH 6.0).
89 Raw milk was standardized with cream and ultrafiltration retentate to obtain cheese milk with a
90 protein to fat ratio of 0.79. The cheese milk was then pasteurized at 72°C for 15 s and cooled to
91 32°C. The cheese milk was weighed (20 kg) and transferred to a cheese vat maintained at 32°C
92 before inoculation with 1.2 % (w/w) of mixed-strain *Lactococcus lactis* starter (Dairy Innovation
93 Australia, Werribee, Australia) cultured in bulk by WCB. The CaCl₂ (Ajax Finechem, Taren
94 Point, Australia) was added to Batches 2, 3, 5 and 6 immediately after starter addition. The total
95 calcium concentration of the cheese milk was 1125 ± 35 mg kg⁻¹ (Batches 1 and 4), 1176 ± 35
96 mg kg⁻¹ (Batches 2 and 5) and 1268 ± 36 mg kg⁻¹ (Batches 3 and 6). The cheese-milk was stirred
97 at 20 revolutions per min (rpm) and ripened for 10-25 min (Table 1). The ripening time was
98 varied in order to achieve a similar pH at the point of rennet addition (~ pH 6.5). This is needed
99 due to the variation in the milk pH as a result of the addition of calcium chloride. The ripening
100 time of milk was shorter for samples with additional CaCl₂, as indicated in Table 1.

101 Rennet (Hannilase L; 690 IMCU mL⁻¹, Chr. Hansen, Bayswater, Australia) was added at
102 a concentration of 0.06 mL kg⁻¹ for all batches. The coagulated cheese-milk was then cut. The
103 total coagulation time, defined here as time from rennet addition to cutting, varied between
104 samples. This adjustment was made based on the results obtained from rheology experiments
105 conducted prior to the pilot scale trial. The storage modulus (G'), which is an indicator of curd
106 elasticity or stiffness during renneting was measured using a rheometer (TA Instruments, New
107 Castle, DE, USA), as reported in a previous study (Ong, Dagastine, Kentish & Gras, 2012) using
108 the same cheese milk used for the pilot scale cheese making. The gelation time, which is defined
109 as the time when the G' increased rapidly (Fig. 1b thick arrows), was significantly shorter for
110 milk where CaCl₂ was added (~500 s for milk with 100 and 300 mg kg⁻¹ calcium addition and
111 950 s for milk without calcium addition). The gel without calcium addition was cut after 45 min
112 of coagulation (2700 s) when the stiffness of the gel was 43 Pa (Fig. 1). The addition of CaCl₂
113 (100 or 300 mg kg⁻¹) significantly decreased ($P < 0.05$) the time for the gel to reach a G' of 43 Pa
114 to ~1750 s (Fig. 1, thin arrows). A shorter coagulation time of 30 min was therefore used for the
115 gel with CaCl₂ addition in the pilot scale trial to ensure a similar gel consistency, avoiding the
116 production of curd fines during cutting.

117 After cutting, the curd was cooked by raising the temperature from 32°C to 38°C in 60
118 min at a rate of 1°C per 10 min with stirring that gradually increased from 5-45 rpm. After 60
119 min, the cooking temperature was maintained at 38°C with stirring at 45 rpm until the curds and
120 whey reached the target draining pH. The whey was drained and collected for compositional
121 analysis when the pH reached pH 6.2 (Batches 1-3) or 6.0 (Batches 4-6). The coagulation and
122 cooking times are shown in Table 1. The cooked curd samples were combined into blocks of
123 about 15 cm in height and turned at 15 min intervals while the temperature was maintained at
124 38°C until the pH of the curds reached pH 5.4. The curd was then milled, 2.5 % (w/w) NaCl
125 added and the curd pressed at 689 kPa overnight.

126 Another six batches of cheeses were made in a second trial using cheese-milk
127 standardized to the same composition and the same conditions applied in the first trial, giving a
128 total of 12 cheeses for the study. Samples for microscopy analysis were collected after each
129 processing step: after gel formation (immediately prior to cutting), after curd cooking
130 (immediately prior to draining), after curd milling (immediately prior to salting) and after
131 pressing (i.e. the cheese).

132

133 **2.2. Confocal microscopy and cryo scanning electron microscopy**

134 The gel, curd and cheese samples were prepared for microstructure examination by
135 confocal laser scanning microscopy (CLSM; Leica Microsystems, Heidelberg, Germany) using a
136 method reported previously (Ong, Dagastine, Kentish & Gras, 2010). The sample preparation for
137 cryo scanning electron microscopy (cryo SEM) has also been reported (Ong, Dagastine, Kentish
138 & Gras, 2011). Briefly, samples 5 mm x 2 mm x 2 mm were fixed by rapid plunging into a liquid
139 nitrogen slush (*ca.* -210°C). The fixed sample was then freeze-fractured in a vacuum chamber,
140 etched at -95°C for 30 min, coated with a mixture of gold and palladium and transferred to a cryo
141 stage maintained at -140°C inside the SEM chamber. The sample was observed using a Quanta
142 SEM (Fei, Hillsboro, OR, USA). Two images were taken for each of the six treatments in each
143 of the two independent trials. The images shown are thus representative of the four images
144 obtained for each of the six treatments.

145

146 **2.3. Image analysis**

147 Image analysis and three dimensional image reconstruction were carried out using an
148 Imaris software package (Bitplane, South Windsor, CT, USA). For each 3D image, 40 adjacent
149 planes were recorded; each consisting of 2D layers 512 x 512 pixels in size and the separation
150 between the planes was 0.25 μm . The image analysis was performed following a previously
151 published method (Ong et al., 2012). The porosity of the sample was calculated from the pore
152 volume fraction, assessed from the volume within a sample that was not stained by fat or protein
153 specific dyes, with respect to total volume of the sample. The data presented are the mean of the
154 four images collected for each treatment (n = 4).

155

156 **2.4. Cheese composition, fat and protein retention and cheese yield**

157 The fat, protein, pH and moisture content of the cheese were determined as described
158 previously (Ong et al., 2012). Fat and protein lost in the whey (FL and PL, respectively) and fat
159 and protein retained in the cheese (FRet and PRet, respectively) were calculated on the basis of
160 the fat or protein concentrations in the cheese-milk. The fat or protein balance (the sum of fat or
161 protein lost in the whey and the fat or protein retained in the cheese), yield of cheese (Ya), dry
162 matter yield of cheese (YDM) were also determined as described in Ong et al. (2012). The
163 cheese yield data are presented as the mean of 2 data points obtained for the two independent
164 trials (n = 2). The fat, protein and moisture content analyses were performed in duplicate for each
165 of the six treatments in each trial. The data presented are the mean of 4 data points collected for
166 the two independent trials (n = 4).

167

168 **2.5. Texture measurement**

169 A texture analyser (Stable Micro System, Godalming, UK) was used to measure the
170 hardness and cohesiveness of the cheese with conditions described in a previous study (Ong et
171 al., 2012). Texture measurement were repeated six times for each of the six independent samples
172 from each batch of cheese, which were then averaged to give one data point for each treatment in
173 each trial. The results were presented as the average of two means for each treatment from the
174 two independent trials (n = 2).

175

176 **2.6. Calcium and phosphorus concentrations**

177 Inductively Coupled Plasma Optical Emission Spectroscopy (Varian, Palo Alto, CA,
178 USA) was used to measure the total calcium and phosphorous content as reported previously
179 (Ong et al., 2012). The analysis was performed in duplicate for each of the six treatments in each
180 trial. The data presented are the mean of 4 data points collected for each treatment for the two
181 independent trials (n = 4).

182

183 **2.7. *Statistical analysis***

184 Statistical analysis of the results was carried out using Minitab statistical package
185 (Minitab Inc., State College, PA, USA). A general linear model was used to study the effect of
186 treatments (calcium addition, draining pH and their interactions) with a significance level of $\alpha =$
187 0.05.

188

189 **3. Results and discussion**

190

191 **3.1. *Variation in cheese making parameters***

192 The addition of CaCl₂ and the alteration of the draining pH require changes to the timing
193 of the cheese making process to ensure the process remains consistent with the approach that
194 would be taken in a manufacturing setting. Specifically, the time allocated for ripening of the
195 milk was reduced for samples with additional CaCl₂ (Table 1) to ensure that all milk was
196 renneted at ~pH 6.5. A shorter coagulation time of 30 min (Table 1) was used for the gel with
197 CaCl₂ addition in the pilot scale trial to ensure a similar gel consistency (G' of 43 Pa) as
198 indicated in Fig 1. All gels were classified as 'medium set' by an experienced cheese maker prior
199 to cutting. The cooking time of the curd was 30-60 min longer for samples drained at pH 6.0, as
200 compared to pH 6.2. As a result, the addition of CaCl₂ shortened the overall processing time
201 when the whey draining pH was 6.2 but was not significantly different ($P > 0.05$) when the
202 draining pH was 6.0 (Table 1). This shorter processing time is desirable, as it allows an increase
203 in plant productivity and a reduction in labour and running costs.

204

205 **3.2. *Microstructure of the gel***

206 The cryo SEM micrographs of the gels (Fig. 2) show a protein network (Pr), which
207 entraps the round fat globules (FG) and starter bacteria (Bc). Some fat globules were dislodged

208 from the network during freeze-fracture, leaving behind a remnant structure that may be the milk
209 fat globule membrane. Both cryo SEM and CLSM micrographs (Fig. 2) show that the gel made
210 with 300 mg kg⁻¹ CaCl₂ addition had a denser protein network with smaller pores than the gel
211 without CaCl₂ addition. The fat globules were homogeneously distributed within all gels
212 regardless of CaCl₂ addition. The 3D image analysis of the gels confirmed the decreased
213 porosity in the gel sample with 300 mg kg⁻¹ CaCl₂ addition (Table 2). The denser protein
214 network obtained from CaCl₂ addition might be expected to mechanically trap more fat globules
215 during whey draining and subsequent processing steps, thus reducing the amount of fat loss
216 during processing. As the gel formed during Cheddar manufacture is somewhat similar to that
217 formed during the production of other cheeses, such as Gouda, these observations may be more
218 widely applicable.

219

220 **3.3. *Microstructure of the cooked curd***

221 During cooking, the fusion of curd particles progressed to a point where a more
222 continuous protein network with smaller pores could be observed (Fig. 3). Several coalesced fat
223 globules (CFG) were also observed as a result of cooking in all samples. The compact structure
224 of the cooked curd makes qualitative comparison of these images harder. The decrease in
225 porosity observed earlier within the gel to which 300 mg kg⁻¹ CaCl₂ was added could not be
226 observed qualitatively within the cooked curd images. The quantitative analysis of the CLSM
227 images, however, shows the porosity of this cooked curd sample was significantly lower than
228 samples lacking calcium addition at both draining pHs ($P < 0.05$, suppl. Fig. 1a).

229 It is difficult to see qualitative differences in the number of fat globules from the cryo
230 SEM micrographs of the cooked curd (Fig. 3). Quantitative analysis of the 3D CLSM images
231 shows a significantly higher number of fat globules with added CaCl₂ at draining pH of 6.2 ($P <$
232 0.05 , suppl. Fig. 1b). This trend was not evident for CaCl₂ addition at draining pH of 6.0,
233 possibly due to the variation in the cooking time (Fig. 1a) required to achieve the target draining
234 pH of 6.0. The large variation observed in the image analysis results also reflects the limitations
235 of this technique; it is difficult to detect differences in a compact curd structure such as what
236 occurs after cooking, where fat coalescence increases and the structure is less homogenous.
237 Compositional analysis of the whey was therefore employed to assess the effect of CaCl₂
238 addition and draining pH on the fat and protein levels in the sample.

239

240 **3.4. Fat and protein content of the sweet whey**

241 The sweet whey acts as a vehicle that carries the fat dislodged from the curd matrix
242 during cooking and draining. The volume of the whey released was not significantly different (P
243 > 0.05) between treatments. The fat concentration within this whey and hence the fat loss was
244 significantly reduced ($P < 0.05$), however, in samples with $300 \text{ mg kg}^{-1} \text{ CaCl}_2$ addition, as
245 compared to the control for both draining pHs (Fig. 4). These results highlight the potential
246 benefit of altering the protein microstructure by means of calcium addition. Interestingly, the
247 denser protein network did not affect the level of protein lost to the sweet whey ($P > 0.05$, Fig.
248 4). This result was consistent with a previous trial, where it was reported that the non-fat milk
249 solids was not significantly affected for Emmentaler cheese when $100 \text{ mg L}^{-1} \text{ CaCl}_2$ was added
250 (Wolfschoon-Pombo, 1997).

251

252 **3.5. Microstructure of the milled curd**

253 The microstructure of the curd after cheddaring and milling (Fig. 5) was more compact
254 than the microstructure of the cooked curd (Fig. 4). The cheddaring process involves the fusion
255 of curd particles, particularly once the curd reaches a pH of ~ 5.8 , as a consequence of the loss of
256 Ca and PO_4 from the protein matrix (Lucey & Fox, 1993). Addition of CaCl_2 did not affect the
257 microstructure of the milled curd, as observed using cryo SEM or CLSM. The differences in
258 porosity observed within the gel and cooked curd became less evident as the structure became
259 more compact. The draining pH, however, appeared to affect the microstructure of the milled
260 curd.

261 The continued cheddaring causes the curd to shrink, expelling more moisture. The pore
262 size reduced to approximately $1 \text{ }\mu\text{m}$. These micron size, circular pores were more obvious within
263 the protein strands of milled curds produced at a higher draining pH (Fig. 5a-c) than at a lower
264 draining pH (Fig. 5d-f). Micron size pores have also been observed within the milled curds of
265 samples made using cheese-milk with different milk protein concentrations (Ong, Dagastine,
266 Kentish & Gras, 2013b). These pores were more readily observed within the milled curd made
267 using milk with a lower protein concentration and the final moisture content of the cheese was
268 higher. Such pores may be important in regulating salt retention. The images reinforce that cryo
269 SEM can be used as a tool to capture structural information that can in turn be used to control the

270 quality of the final cheese product. The bacterial cells also appear in much larger clusters within
271 the milled curd at a draining pH of 6.0 compared to at pH 6.2, possibly due to the longer cooking
272 time for these samples. The difference in distribution was not quantified and the contribution to
273 cheese ripening is unclear and would require further investigation.

274 Suppl. Fig. 2 provides lower magnification cryo SEM images of the milled curd where
275 curd junctions can be observed. These junctions form when the fat depleted curd fines fuse
276 together during the cheddaring process, forming a protein rich seam within the network. Several
277 crystalline inclusions can be observed especially at curd junctions in samples where 300 mg kg^{-1}
278 CaCl_2 had been added at both draining pHs (suppl. Fig. 2a and b). Suppl. Fig. 2 also shows
279 fewer crystalline inclusions at a lower draining pH. The presence of such crystalline phases of
280 salts in the curd and cheese during processing has been reported in several studies (Brooker,
281 1975; Frau, Mulet, Simal, Massanet & Rossello, 1997). It is not clear, however, if the presence
282 of these crystalline salts relates to calcium lactate formation in cheese and this warrants further
283 investigation.

284 The micron size pores seen by cryo SEM (Fig. 5) could not be easily observed within the
285 CLSM images (data not shown) due to the presence of the aqueous serum phase and the lower
286 resolution of the CLSM technique. The porosity of the milled curds and the number of fat
287 globules were also not significantly different when assessed by CLSM ($P > 0.05$, suppl. Fig. 1c
288 and d).

289

290 **3.6. *Microstructure and texture of the Cheddar cheese***

291 Pressing reduced the size of the pores observed within the milled curd and the structure
292 became more compact (Fig. 6). The differences between treatments observed at the earlier stages
293 are less obvious at this stage. The 3D CLSM images, however, show the presence of large holes
294 randomly distributed within the cheeses (Fig. 6h, i and j, indicated by the arrows). It is not clear
295 why these holes form. They could be the result of the calcium addition (Ong et al., 2013a) or the
296 longer cooking time and the shorter cheddaring time applied for samples with a lower draining
297 pH, which may prevent the curd from completely fusing together. No clear trend is observed,
298 however, in the porosity of the cheese as a function of pH or calcium addition (suppl. Fig. 1e)
299 suggesting that the distribution of pores is altered rather than the total sample porosity.

300 Textural analysis of the cheeses indicated that the cheese with a lower draining pH was
301 significantly harder (Fig. 7), possibly due to the lower moisture content of the cheese (Fig. 8a).
302 Addition of CaCl_2 up to 300 mg kg^{-1} did not affect the hardness of the cheese ($P > 0.05$). There
303 was also no significant difference in cohesiveness between the treatments ($P > 0.05$) despite the
304 occurrence of holes observed in the cheese, as discussed above. Previous studies have shown that
305 the pH at whey draining influences the texture of a cheese due to changes in the mineral content
306 of the cheese (Lucey & Fox, 1993). Cheeses with a low draining pH, such as Cheshire cheese,
307 are known to be more crumbly than cheese such as Colby, which is drained at a higher pH (Hall
308 & Creamer, 1972). The low draining pH of 6.0 used in this study does not appear to be
309 sufficiently low to cause significant changes to the cohesiveness of the freshly-pressed Cheddar
310 cheese.

311

312 **3.7. Yield and composition of Cheddar cheese**

313 The fat retained in the cheese was significantly higher with calcium addition at both
314 draining pHs ($P < 0.05$, Fig. 8b), possibly due to the lower concentration of fat lost to sweet whey
315 (Fig. 4a). The protein retained in the cheese (Fig. 8b), the yield (Ya) and yield in dry matter
316 (YDM) of the cheeses, however, were not significantly different between treatments ($P > 0.05$).
317 The average Ya of the cheeses was $12.2 \pm 0.4 \%$ (w/w) and the YDM was $7.9 \pm 0.3 \%$ (w/w).
318 Our yield result is in apparent contrast to the findings of Wolfschoon-Pombo (1997) and Ustunol
319 et al. (1990), who both observed an increase in yield of Emmentaler-type cheese or stirred curd
320 cheese, when 100 mg L^{-1} or 200 mg L^{-1} of calcium was added to the cheese-milk. The $\sim 3.5 \%$
321 difference in fat retention observed in the cheese here is equal to 35 g of increased fat per vat of
322 cheese or increase in yield of $\sim 0.18 \%$ (Ya, estimated as kg cheese per kg of cheese-milk). The
323 variation in Ya observed between the trials conducted here was 0.4 %, which makes the increase
324 in fat retention not observable within the range of variation observed for the Ya data. The total
325 fat and protein mass balance during this cheese making trial was within the range of 94-97%
326 recovery and 90-92% recovery for fat and protein respectively.

327 The composition of the cheese was within the range usually observed for Cheddar cheese
328 (Fig. 8a, c). There were no significant differences in fat or protein composition with CaCl_2
329 addition ($P > 0.05$, Fig. 8c). The concentration of fat and protein in the cheese, however, was
330 significantly higher ($P < 0.05$) in cheeses with a lower draining pH (Fig. 8c) possibly due to the

331 lower moisture content of these cheeses (Fig. 8a). This is consistent with previously published
332 results, where cheese drained at a lower pH tends to have a lower moisture level (Kiely et al.,
333 1992; Yazici & Akbulut, 2007). The fat in dry matter of the cheeses was not significantly
334 different ($P > 0.05$) regardless of the different treatments (Fig. 8a), showing that moisture was
335 the only significant difference between treatments.

336

337 **3.8. Total and soluble calcium and phosphorus concentrations**

338 Special whey treatment is normally required if the sweet whey contains an excessive
339 level of calcium but our study shows that the addition of 300 mg kg^{-1} did not change the calcium
340 concentration in the whey and would not necessitate additional treatment. The total calcium in
341 the whey was $346\text{-}374 \text{ mg kg}^{-1}$ compared to $330\text{-}358 \text{ mg kg}^{-1}$ for whey from cheese with 300 mg
342 kg^{-1} added CaCl_2 vs no added calcium, respectively). Most of the added CaCl_2 was retained in
343 the curd. As a result there was an increasing trend in the level of total calcium in the cooked curd
344 and cheese with increasing CaCl_2 addition ($P < 0.05$) (Fig. 9). The effect of the whey draining pH
345 on the level of the total calcium concentration was more significant, however, than the addition
346 of CaCl_2 .

347 More calcium phosphate is solubilised at a lower pH resulting in the higher concentration
348 ($P < 0.05$) of total calcium in the whey ($325 \pm 24 \text{ mg kg}^{-1}$ and $366 \pm 8 \text{ mg kg}^{-1}$ in whey drained at
349 pH 6.2 or pH 6.0, respectively). As a result, the total calcium retained in the cooked curd and
350 cheese for samples drained at a lower pH was lower regardless of the CaCl_2 level (Fig. 9). The
351 difference in the total calcium content of the whey due to the lower draining pH is consistent
352 with a previous study for Cheddar cheese reported by Lucey and Fox (1993).

353 Fig. 9 also shows an interesting comparison between the different treatments. The level
354 of total calcium in the cheese with a draining pH of 6.2 without added CaCl_2 was similar to
355 cheese with a draining pH 6.0 where 300 mg kg^{-1} CaCl_2 was added. This suggests the
356 combination of calcium addition and lower draining pH could be used to produce cheese with a
357 similar concentration of calcium to the control (no added CaCl_2 , pH 6.2). The extra calcium
358 added at the beginning of cheese making would improve coagulation but is drained away by
359 lowering the draining pH. This minimizes potential defects associated with excess calcium in the
360 final cheese, although textural and microstructural analyses show these cheeses here differ in
361 hardness and the arrangement of pores within the microstructure

362 The total phosphorus concentration was not affected by the addition of CaCl₂. A lower
363 concentration of phosphorus was observed in cheese with a lower draining pH ($P < 0.05$, data not
364 shown), similar to observations for calcium.

365

366 **4. Conclusion**

367 This study supports evidence from past studies that the addition of calcium chloride has
368 the potential to reduce the fat lost to sweet whey and potentially improve the fat retention in
369 cheese. Advanced microscopy tools such as CLSM and cryo SEM can be used to provide
370 microstructural data to understand the mechanisms of fat retention within the gel during the early
371 stage of cheese making. Quantitative image analysis measures such as porosity, however, are less
372 able to differentiate between the process variables as the curd becomes more compact. The
373 moisture content of the cheese was lower at a lower draining pH and the fat concentration and
374 cheese hardness increased. Chemical and textural analyses were therefore needed in combination
375 with the microstructural data to assess the effect of calcium addition and draining pH.

376 This study shows that CaCl₂ addition in combination with a lower draining pH could be
377 used to improve coagulation at the early stage of cheese making, potentially improving fat
378 retention and maintaining a similar level of total calcium in the final cheese. Our study provides
379 further evidence that the addition of CaCl₂ can be used to shorten the processing time and
380 increase factory productivity. Our findings may also be relevant to the manufacture of Gouda
381 type cheeses, which share similar early steps of manufacture with Cheddar cheese.

382

383 **Acknowledgements**

384 This work was funded by the Australia Research Council (ARC- LP0883300) and Dairy
385 Innovation Australia Limited. LO, SEK and SLG are also supported by the ARC Dairy
386 Innovation Hub (IH120100005). The authors thank the Electron Microscopy Unit and the
387 Biological Optical Microscopy Platform at Bio21 Molecular Science & Biotechnology Institute
388 and the Particulate Fluids Processing Centre, which is a Special Research Centre of the ARC at
389 The University of Melbourne, for equipment access. We also thank Warrnambool Cheese and
390 Butter for their involvement in this project.

391

392 **References**

393 Brooker, B. E. (1975). Observations on the microscopic crystalline inclusions in Cheddar cheese.
394 *Journal of Dairy Research*, 42, 341-348.

395 Choi, J., Horne, D. S., Johnson, M. E., & Lucey, J. A. (2008). Effects of the concentration of
396 insoluble calcium phosphate associated with casein micelles on the functionality of
397 directly acidified cheese. *Journal of Dairy Science*, 91, 513-522.

398 Dalgleish, D. G. (1983). Coagulation of renneted bovine casein micelles-dependence on
399 temperature, calcium-ion concentration and ionic strength. *Journal of Dairy Research*,
400 50, 331-340.

401 Dalgleish, D. G., & Law, A. J. R. (1989). pH-Induced dissociation of bovine casein micelles. II.
402 Mineral solubilization and its relation to casein release. *Journal of Dairy Research*, 56,
403 727-735.

404 Federal Register (1982). Electronic code of federal regulations. Title 21: Food and Drugs. Part
405 184 - Direct food substances affirmed as generally recognize as safe. Retrieved on 21
406 May 2014. Last updated: 1 April 1996, from www.ecfr.gov.

407 Frau, M., Mulet, A., Simal, S., Massanet, J., & Rossello, C. (1997). Microscopic crystalline
408 inclusions in Mahón cheese / Inclusiones cristalinas microscópicas en el queso Mahón.
409 *Food Science and Technology International*, 3, 43-47.

410 Gastaldi, E., Pellegrini, O., Lagaude, A., & Fuente, B. T. (1994). Functions of added calcium in
411 acid milk coagulation. *Journal of Food science*, 59, 310-312.

412 Guinee, T. P., Feeney, E. P., Auty, M. A. E., & Fox, P. F. (2002). Effect of pH and calcium
413 concentration on some textural and functional properties of mozzarella cheese. *Journal of*
414 *Dairy Science*, 85, 1655-1669.

415 Hall, D. M., & Creamer, L. K. (1972). A study of the sub-microscopic structure of Cheddar,
416 Cheshire and Gouda cheese by electron microscopy. *New Zealand Journal of Dairy*
417 *Science and Technology*, 7, 95.

418 Holmes, D. G., Duersch, J. W., & Ernstrom, C. A. (1977). Distribution of milk clotting enzymes
419 between curd and whey and their survival during Cheddar cheese making. *Journal of*
420 *Dairy Science*, 60, 862-869.

421 Kiely, L. J., Kindstedt, P. S., Hendricks, G. M., Levis, J. E., Yun, J. J., & Barbano, D. M. (1992).
422 Effect of draining pH on the development of curd structure during the manufacture of
423 mozzarella cheese. *Food Structure*, *11*, 217-224.

424 Lucey, J. A., & Fox, P. F. (1993). Importance of calcium and phosphate in cheese manufacture: a
425 review. *Journal of Dairy Science*, *76*, 1714-1724.

426 Okigbo, L. M., Richardson, G. H., Brown, R. J., & Ernstrom, C. A. (1985). Effects of pH,
427 calcium chloride, and chymosin concentration on coagulation properties of abnormal and
428 normal milk. *Journal of Dairy Science*, *68*, 2527-2533.

429 Ong, L., Dagastine, R. R., Kentish, S. E., & Gras, S. L. (2010). The effect of milk processing on
430 the microstructure of the milk fat globule and rennet induced gel observed using confocal
431 laser scanning microscopy. *Journal of Food Science*, *75*, E135-E145.

432 Ong, L., Dagastine, R. R., Kentish, S. E., & Gras, S. L. (2011). Microstructure of gel and cheese
433 curd observed using cryo scanning electron microscopy and confocal microscopy. *LWT -*
434 *Food Science and Technology*, *44*, 1291-1302.

435 Ong, L., Dagastine, R. R., Kentish, S. E., & Gras, S. L. (2012). The effect of pH at renneting on
436 the microstructure, composition and texture of Cheddar cheese. *Food Research*
437 *International*, *48*, 119-130.

438 Ong, L., Dagastine, R. R., Kentish, S. E., & Gras, S. L. (2013a). The effect of calcium chloride
439 addition on the microstructure and composition of Cheddar cheese. *International Dairy*
440 *Journal*, *33*, 135-141.

441 Ong, L., Dagastine, R. R., Kentish, S. E., & Gras, S. L. (2013b). Microstructure and composition
442 of full fat Cheddar cheese made with ultrafiltered milk retentate. *Foods*, *2*, 310-331.

443 Phadungath, C., & Metzger, L. E. (2011). Effect of sodium gluconate on the solubility of calcium
444 lactate. *Journal of Dairy Science*, *94*, 4843-4849.

445 Udabage, P., McKinnon, I. R., & Augustin, M. A. (2001). Effects of mineral salts and calcium
446 chelating agents on the gelation of renneted skin milk. *Journal of Dairy Science*, *84*,
447 1569-1575.

448 Ustunol, Z., & Hicks, C. L. (1990). Effect of calcium addition on yield of cheese manufactured
449 with endothia parasitica Protease. *Journal of Dairy Science*, *73*, 17-25.

450 Walstra, P., Wouters, T. M., & Geurts, T. J. (2006). Cheese manufacture. In *Dairy Science and*
451 *Technology* 2nd ed., (pp. 583-638). Boca Raton, FL, USA.

- 452 Wolfschoon-Pombo, A. F. (1997). Influence of calcium chloride addition to milk on the cheese
453 yield. *International Dairy Journal*, 7, 249-254.
- 454 Yazici, F., & Akbulut, C. (2007). Impact of whey pH at drainage on the physicochemical,
455 sensory and functional properties of mozzarella cheese made from buffalo milk. *Journal*
456 *of Agricultural and Food Chemistry*, 55, 9993-10000.
- 457 Zoon, P., van Vliet, T., & Walstra, P. (1988). Rheological properties of rennet-induced skim
458 milk gels. 3. The effect of calcium and phosphate. *Netherlands Milk & Dairy Journal*, 42,
459 295-312.

460 **List of Figures**

461

462 **Fig. 1.** The storage modulus (G') measured from the time of rennet addition for cheese-milk
463 without CaCl_2 addition (dotted line), with 100 mg kg^{-1} CaCl_2 addition (thin solid line), or with
464 300 mg kg^{-1} CaCl_2 addition (thick solid line). The thick arrows indicate the gelation point when
465 G' reaches $\sim 5 \text{ Pa}$) and the thin arrows indicate the cutting point when G' reaches 43 Pa . Error
466 bars are the standard deviation of the mean from two independent trials ($n = 2$).

467

468 **Fig. 2.** Cryo SEM (first row, **a-c**) and CLSM (second row, **d-f** and third row, **g-i**) micrographs of
469 gels made using cheese-milk without CaCl_2 addition (left), with 100 mg kg^{-1} CaCl_2 addition
470 (middle) or 300 mg kg^{-1} CaCl_2 addition (right). FG = fat globules, Pr = protein network and Bc =
471 bacteria. The black areas correspond to pores. The 3D CLSM micrograph (second row) is
472 constructed from 40 layers of 2D images with a total depth of $10 \mu\text{m}$. The middle section of the
473 3D protein reconstruction (third row) is presented in three views: the X-Y image (top), X-Z
474 image (bottom) and Z-Y (right). Scale bars within the cryo SEM and CLSM images are $10 \mu\text{m}$
475 and $20 \mu\text{m}$ in length respectively. Nile red stained fat appears red and FCF fast green stained
476 protein appears green. Please refer to the online edition for a colour version of this figure.

477

478 **Fig. 3.** Cryo SEM micrographs of cooked curd made using cheese-milk without CaCl_2 addition
479 (**a, d**), with 100 mg kg^{-1} CaCl_2 addition (**b, e**) or 300 mg kg^{-1} CaCl_2 addition (**c, f**) where the
480 whey was drained at pH 6.2 (**a-c**) or at pH 6.0 (**d-f**). FG = fat globules, CFG = coalesced fat
481 globules, Pr = protein network and Bc = bacteria. The black areas correspond to pores. Scale bars
482 within the cryo SEM images are $10 \mu\text{m}$ in length.

483

484 **Fig. 4.** The effect of CaCl_2 addition on the percentage of fat lost (FL, \blacksquare or \square with solid line) or
485 protein lost (PL, \blacktriangle or \triangle with dotted line) to the sweet whey at pH 6.2 (closed markers, \blacksquare or \blacktriangle)
486 or pH 6.0 (open markers, \square or \triangle). Error bars are the standard error of the mean ($n = 4$).

487

488 **Fig. 5.** Cryo SEM micrographs of milled curd made using cheese-milk without CaCl_2 addition
489 (**a, d**), with 100 mg kg^{-1} CaCl_2 addition (**b, e**) or 300 mg kg^{-1} CaCl_2 addition (**c, f**) where the
490 whey was drained at pH 6.2 (a-c) or at pH 6.0 (d-f). FG = fat globules, CFG = coalesced fat

491 globules, Pr = protein network, Bc = bacteria and Cr = crystalline inclusion. The black areas
492 correspond to pores. Scale bars within the cryo SEM images are 10 μm in length.

493
494 **Fig. 6.** Cryo SEM micrographs of Cheddar cheese made using cheese-milk without CaCl_2
495 addition (**a, d**), with 100 mg kg^{-1} CaCl_2 addition (**b, e**) or 300 mg kg^{-1} CaCl_2 addition (**c, f**) where
496 the whey was drained at pH 6.2 (**a-c**) or at pH 6.0 (**d-f**). FG = fat globules, CFG = coalesced fat
497 globules, Pr = protein network and Bc = bacteria. The black areas correspond to pores.
498 Representative CLSM micrographs of Cheddar cheese made using cheese-milk without CaCl_2
499 addition (**g, i**) or with 300 mg kg^{-1} CaCl_2 addition (**h, j**) where the whey was drained at 6.2 (**g, h**)
500 or pH 6.0 (**i, j**). The 3D images each consist of 40 layers of 2D images with a total depth of 10
501 μm . Scale bars within the cryo SEM and CLSM images are 10 μm and 20 μm in length
502 respectively. Nile red stained fat appears red and FCF fast green stained protein appears green.
503 Please refer to the online edition for a colour version of this figure.

504
505 **Fig. 7.** The hardness (\blacksquare or \square with dotted line) or and cohesiveness (\blacktriangle or \triangle with solid line) of
506 Cheddar cheese made with different concentration of CaCl_2 addition at a draining pH of 6.2
507 (closed markers, \blacksquare or \blacktriangle) or pH 6.0 (open markers, \square or \triangle). Error bars are the standard error of
508 the mean ($n = 2$). Each mean is the average of 6 texture analyses from the same batch of cheese.

509
510 **Fig. 8.** The effect of CaCl_2 addition or whey draining pH on **a**) the concentration of moisture in
511 cheese (Moisture, \blacksquare or \square with dotted line) and the fat concentration in dry matter of the cheese
512 (FDM, \blacktriangle or \triangle with solid line); **b**) the percentage of fat (FRet, \blacksquare or \square with solid line) or protein
513 retained in the cheese (PRet, \blacktriangle or \triangle with dotted line); **c**) the concentration of fat (Fat, \blacksquare or \square
514 with solid line) or protein in cheese (Protein, \blacktriangle or \triangle with dotted line). The whey draining pH
515 was 6.2 (closed markers, \blacksquare or \blacktriangle) or pH 6.0 (open markers, \square or \triangle). The fat and protein retained
516 were calculated on the basis of fat and protein concentration in the cheese-milk. Error bars are
517 the standard error of the mean ($n = 4$).

518
519 **Fig. 9.** The effect of CaCl_2 addition or whey draining pH on the level of total calcium in the curd
520 collected after cooking (Cooked, \blacktriangle or \triangle with solid line) and after pressing (Cheese, \blacksquare or \square with

521 dotted line). The whey draining pH was 6.2 (closed markers, ■ or ▲) or pH 6.0 (open markers, □
522 or Δ). Error bars are the standard error of the mean (n = 4).

523

524 **Supplementary Fig. 1.** Physical properties of the cooked curd (a-b), milled curd (c-d) and
525 cheese (e-f) made with different levels of CaCl₂ addition. All properties were determined by
526 three dimensional image analysis of CLSM images. Results are presented as the mean ± the
527 standard error of the mean (n = 4). The whey draining pH was 6.2 (closed marker, ■, ● or ▲
528 with solid line) or pH 6.0 (open markers, □, ○ or Δ with dotted line).

529

530 **Supplementary Fig. 2.** Cryo SEM micrographs of milled curd made using cheese-milk with 300
531 mg kg⁻¹ CaCl₂ addition and with a whey draining pH of 6.2 (a) or pH 6.0 (b). The white circles
532 indicate the presence of crystalline inclusions. Arrows indicate the curd junction. Scale bars
533 within the images are 50 μm in length.

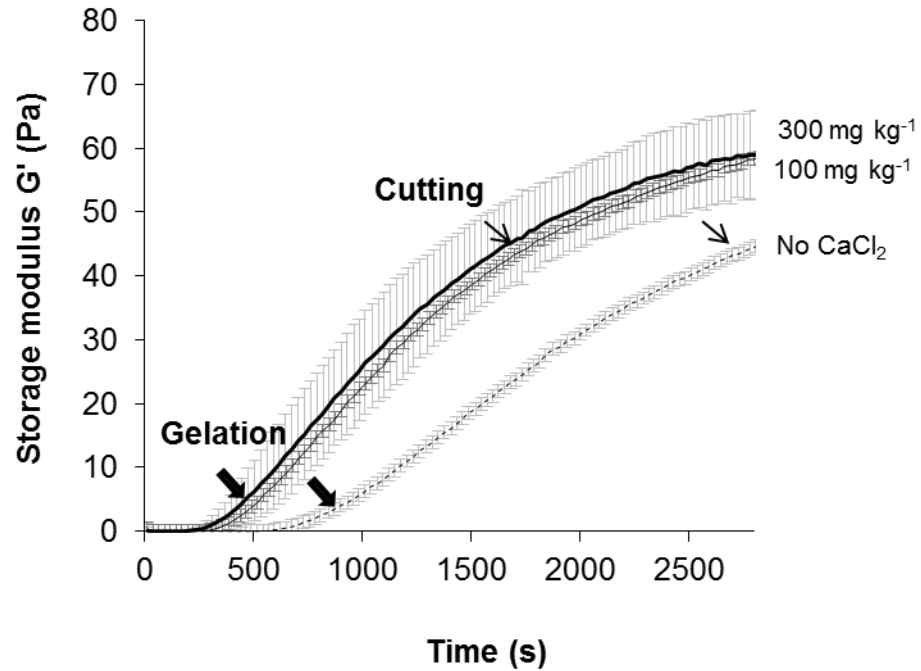


Fig. 1. The storage modulus (G') measured from the time of rennet addition for cheese-milk without CaCl₂ addition (dotted line), with 100 mg kg⁻¹ CaCl₂ addition (thin solid line), or with 300 mg kg⁻¹ CaCl₂ addition (thick solid line). The thick arrows indicate the gelation point when G' reaches ~5 Pa) and the thin arrows indicate the cutting point when G' reaches 43 Pa. Error bars are the standard deviation of the mean from two independent trials ($n = 2$).

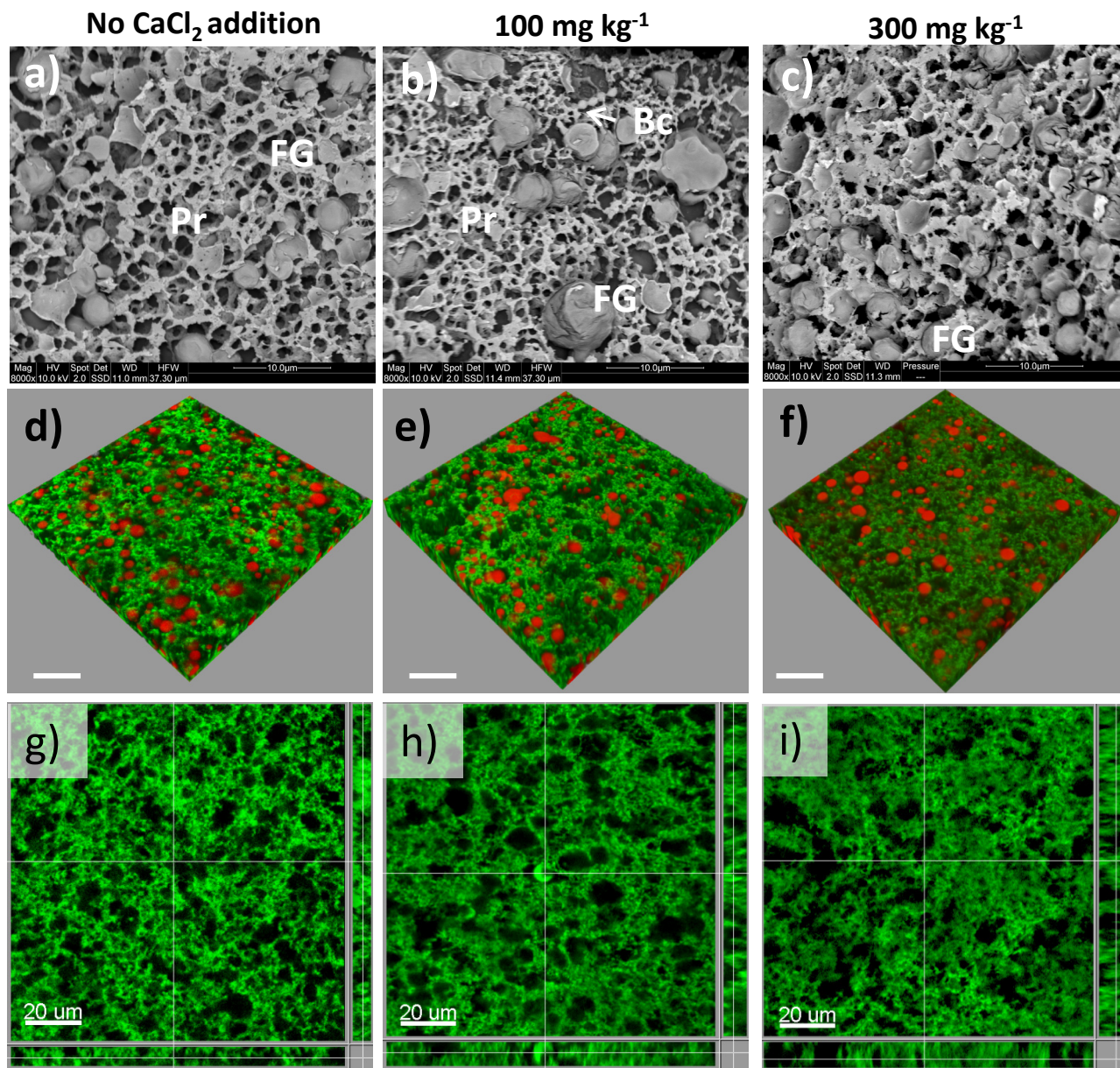


Figure 2. Cryo SEM (first row, a-c) and CLSM (second row, d-f and third row, g-i) micrographs of gels made using cheese-milk without CaCl_2 addition (left), with 100 mg kg^{-1} CaCl_2 addition (middle) or 300 mg kg^{-1} CaCl_2 addition (right). FG = fat globules, Pr = protein network and Bc = bacteria. The black areas correspond to pores. The 3D CLSM micrograph (second row) is constructed from 40 layers of 2D images with a total depth of 10 μm . The middle section of the 3D protein reconstruction (third row) is presented in three views: the X-Y image (top), X-Z image (bottom) and Z-Y (right). Scale bars within the cryo SEM and CLSM images are 10 μm and 20 μm in length respectively. Nile red stained fat appears red and FCF fast green stained protein appears green.

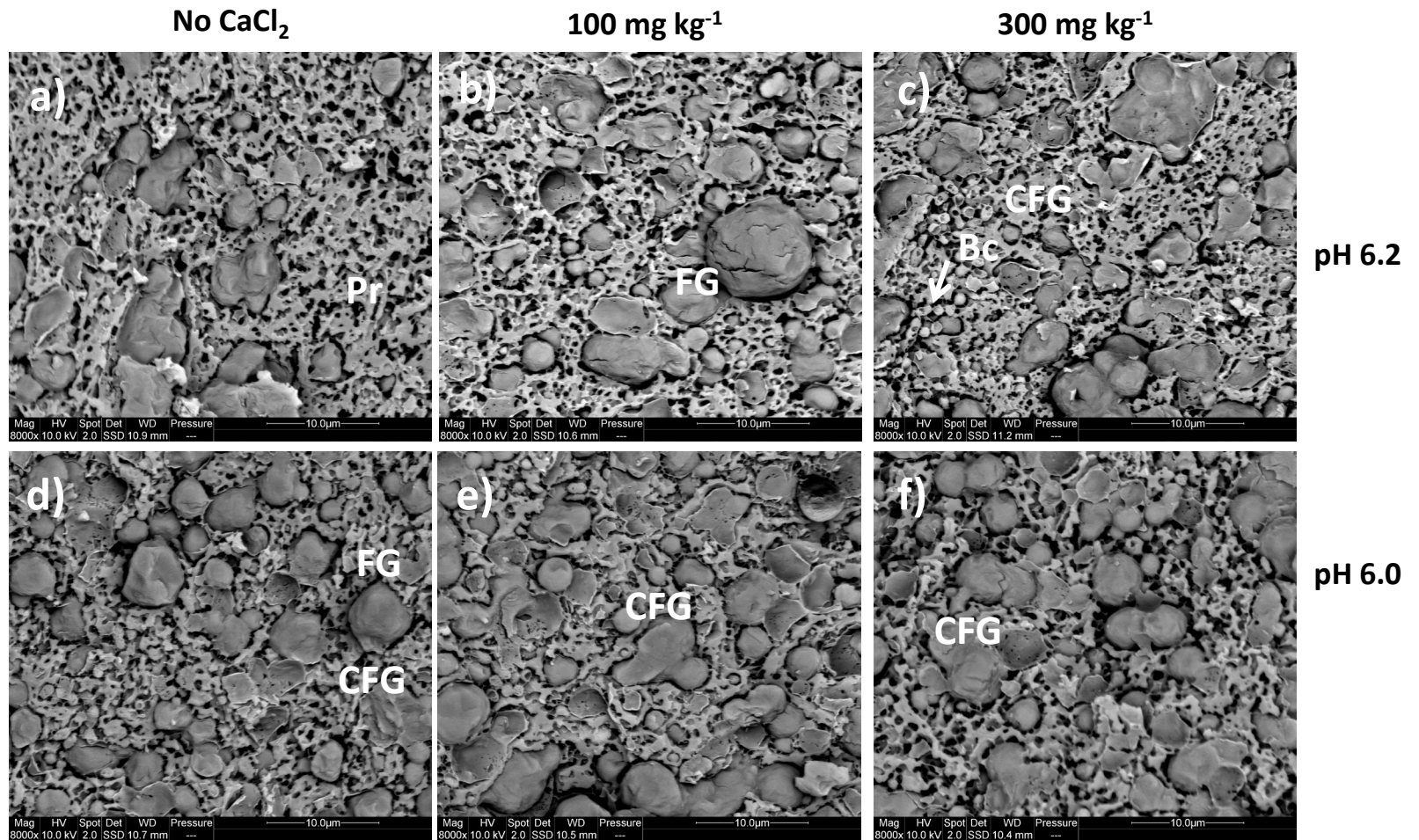


Fig. 3. Cryo SEM micrographs of cooked curd made using cheese-milk without CaCl₂ addition (**a, d**), with 100 mg kg⁻¹ CaCl₂ addition (**b, e**) or 300 mg kg⁻¹ CaCl₂ addition (**c, f**) where the whey was drained at pH 6.2 (**a-c**) or at pH 6.0 (**d-f**). FG = fat globules, CFG = coalesced fat globules, Pr = protein network and Bc = bacteria. The black areas correspond to pores. Scale bars within the cryo SEM images are 10 μm in length.

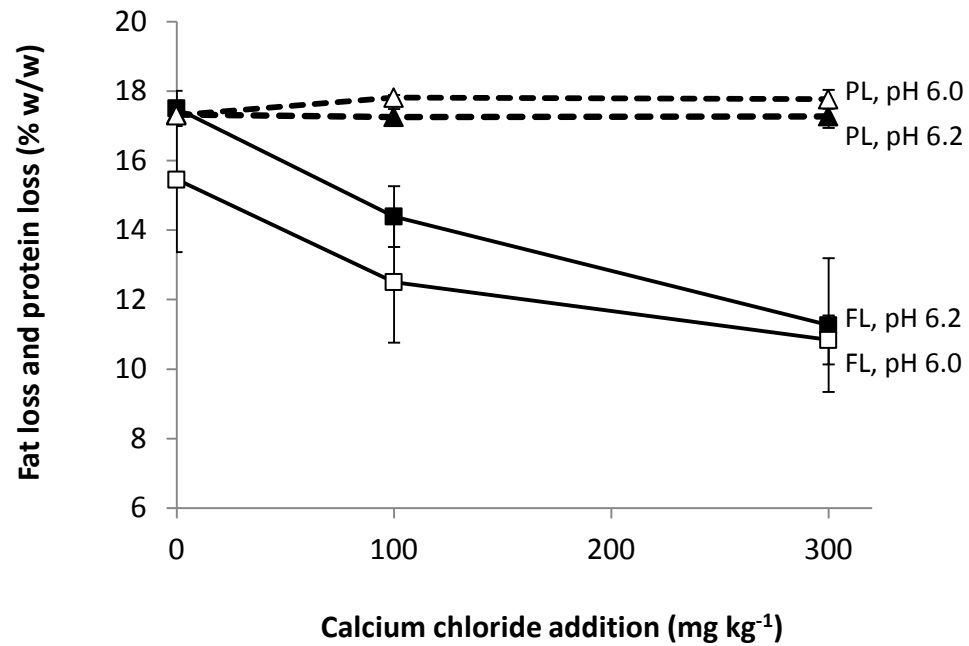


Fig. 4. The effect of CaCl₂ addition on the percentage of fat lost (FL, ■ or □ with solid line) or protein lost (PL, ▲ or Δ with dotted line) to the sweet whey at pH 6.2 (closed markers, ■ or ▲) or pH 6.0 (open markers, □ or Δ). Error bars are the standard error of the mean (n = 4).

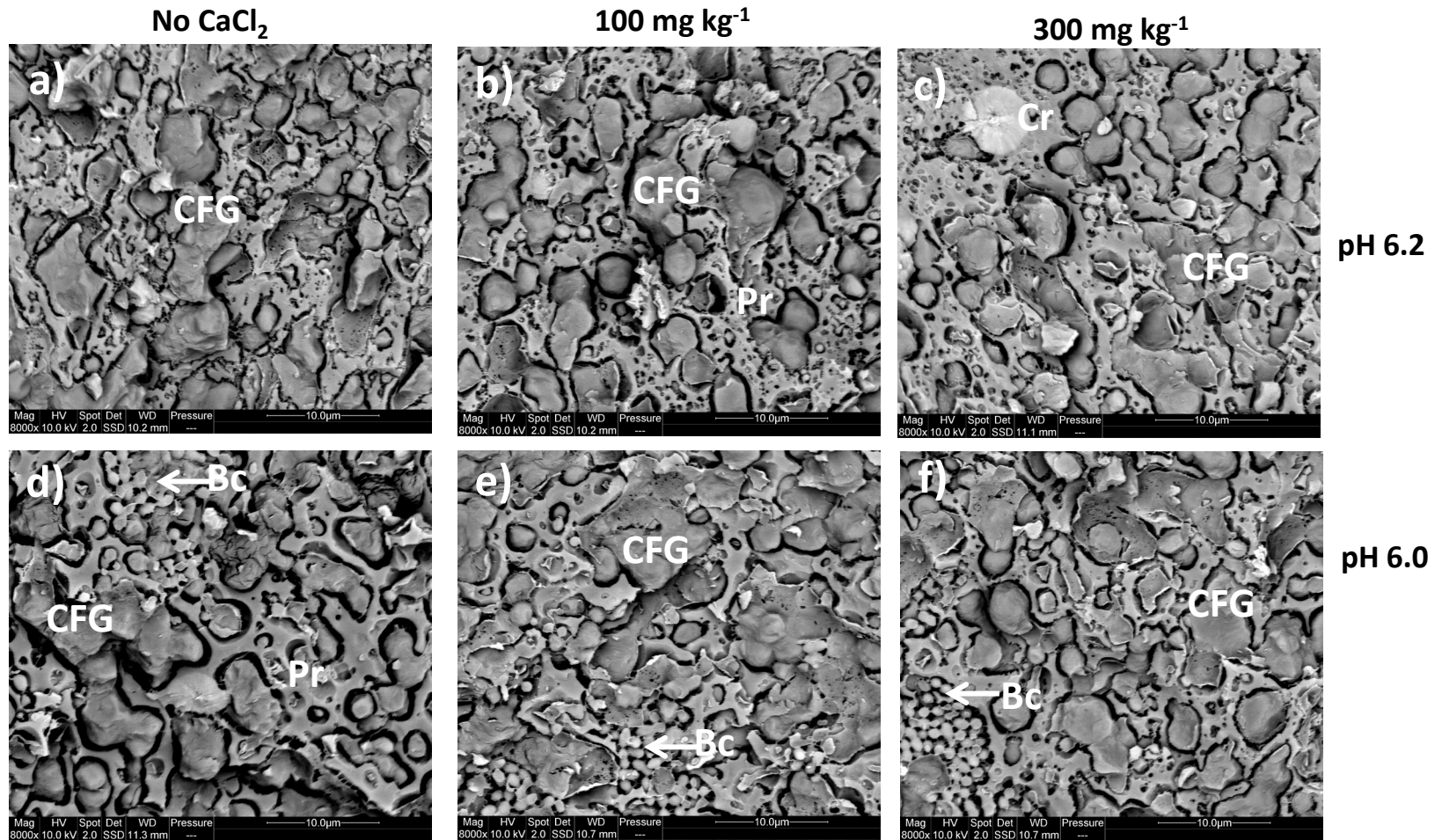


Fig. 5. Cryo SEM micrographs of milled curd made using cheese-milk without CaCl₂ addition (**a, d**), with 100 mg kg⁻¹ CaCl₂ addition (**b, e**) or 300 mg kg⁻¹ CaCl₂ addition (**c, f**) where the whey was drained at pH 6.2 (**a-c**) or at pH 6.0 (**d-f**). FG = fat globules, CFG = coalesced fat globules, Pr = protein network, Bc = bacteria and Cr = crystalline inclusion. The black areas correspond to pores. Scale bars within the cryo SEM images are 10 µm in length.

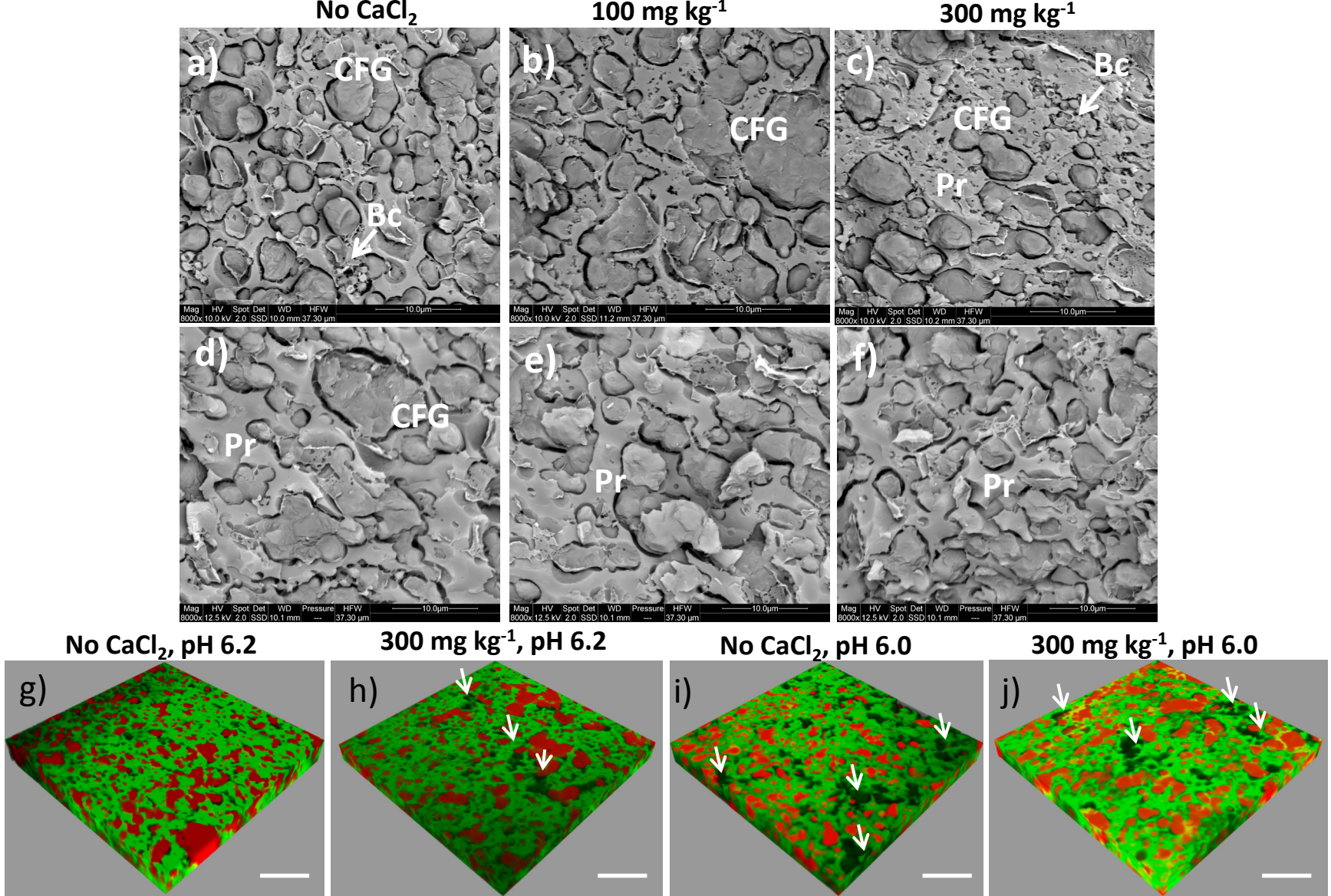


Fig. 6. Cryo SEM micrographs of Cheddar cheese made using cheese-milk without CaCl_2 addition (a, d), with 100 mg kg^{-1} CaCl_2 addition (b, e) or 300 mg kg^{-1} CaCl_2 addition (c, f) where the whey was drained at pH 6.2 (a-c) or at pH 6.0 (d-f). FG = fat globules, CFG = coalesced fat globules, Pr = protein network and Bc = bacteria. The black areas correspond to pores. Representative CLSM micrographs of Cheddar cheese made using cheese-milk without CaCl_2 addition (g, i) or with 300 mg kg^{-1} CaCl_2 addition (h, j) where the whey was drained at 6.2 (g, h) or pH 6.0 (i, j). The 3D images each consist of 40 layers of 2D images with a total depth of 10 μm . Nile red stained fat appears red and FCF fast green stained protein appears green. Scale bars within the cryo SEM and CLSM images are 10 μm and 20 μm in length respectively.

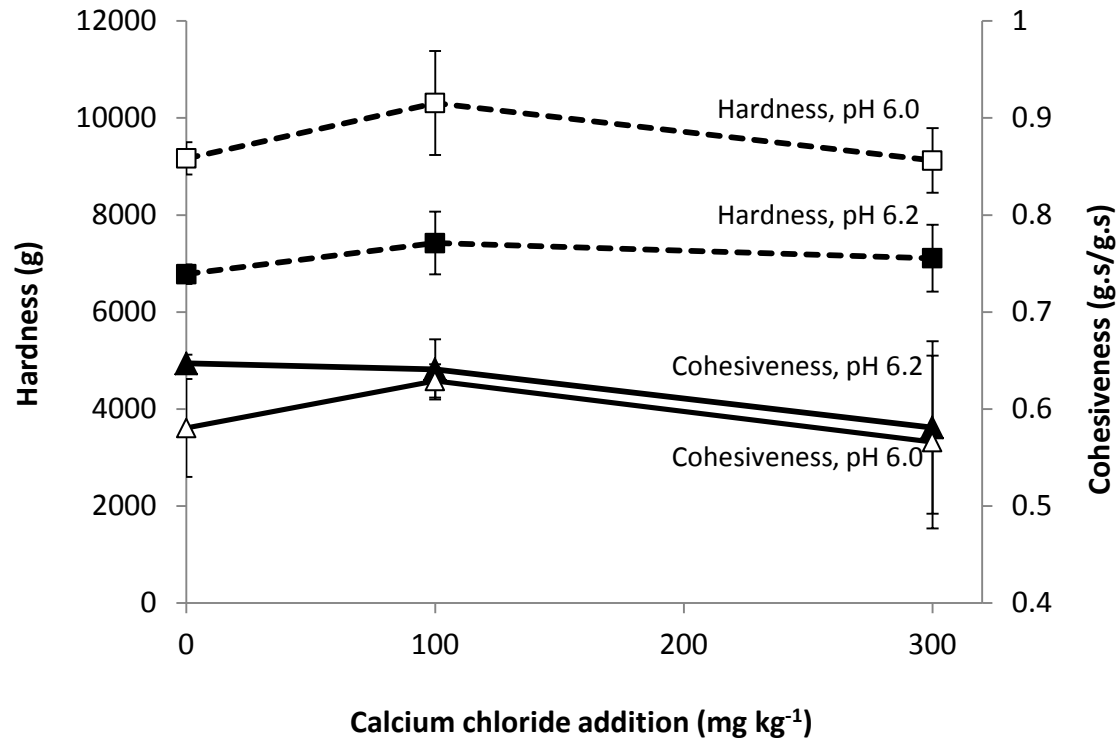


Fig. 7. The hardness (■ or □ with dotted line) or and cohesiveness (▲ or Δ with solid line) of Cheddar cheese made with different concentration of CaCl₂ addition at a draining pH of 6.2 (closed markers, ■ or ▲) or pH 6.0 (open markers, □ or Δ). Error bars are the standard error of the mean (n = 2). Each mean is the average of 6 texture analyses from the same batch of cheese.

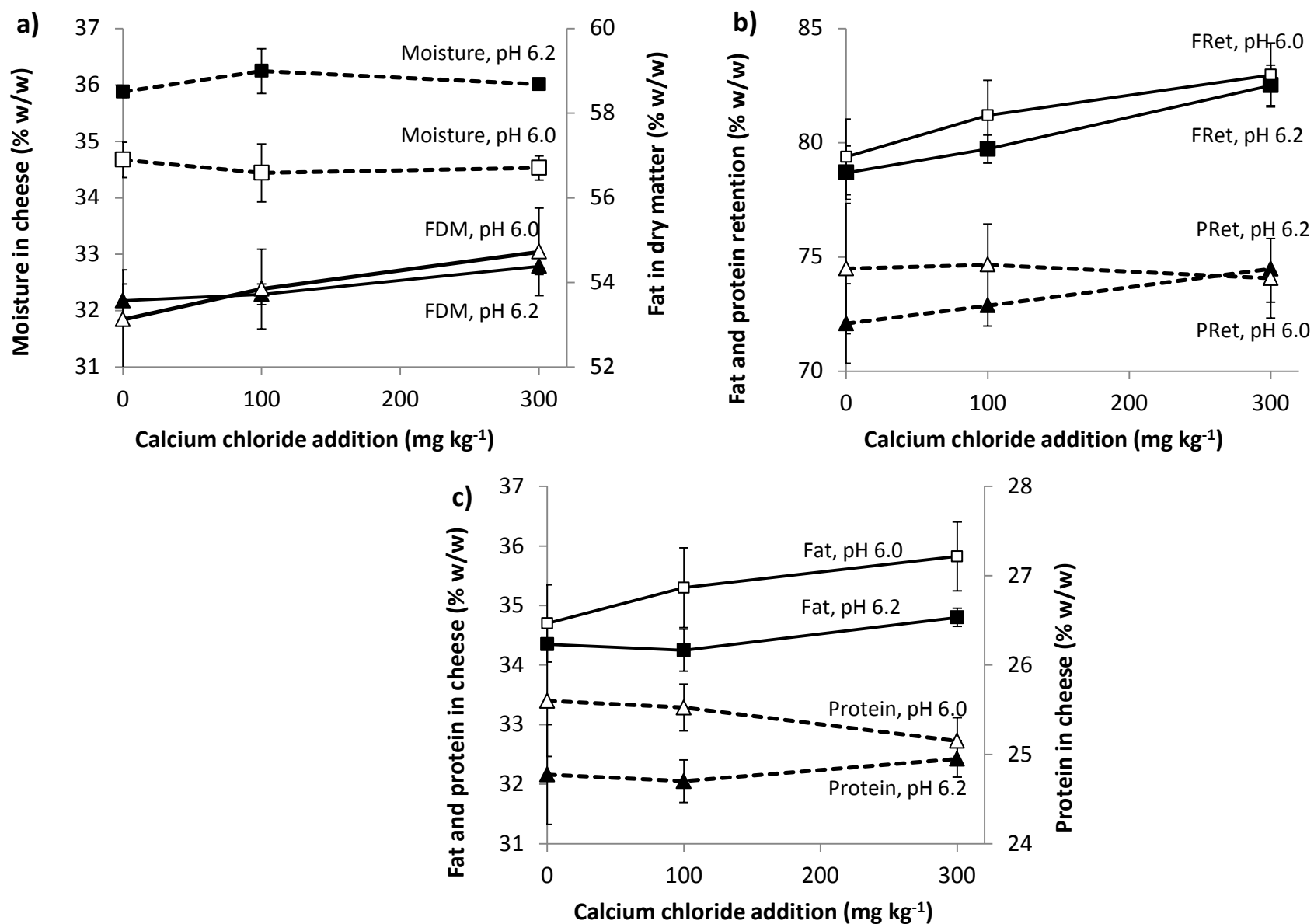


Fig. 8. The effect of CaCl₂ addition or whey draining pH on **a)** the concentration of moisture in cheese (Moisture, ■ or □ with dotted line) and the fat concentration in dry matter of the cheese (FDM, ▲ or Δ with solid line); **b)** the percentage of fat (FRet, ■ or □ with solid line) or protein retained in the cheese (PRet, ▲ or Δ with dotted line); **c)** the concentration of fat (Fat, ■ or □ with solid line) or protein in cheese (Protein, ▲ or Δ with dotted line). The whey draining pH was 6.2 (closed markers, ■ or ▲) or pH 6.0 (open markers, □ or Δ). The fat and protein retained were calculated on the basis of fat and protein concentration in the cheese-milk. Error bars are the standard error of the mean (n = 4).

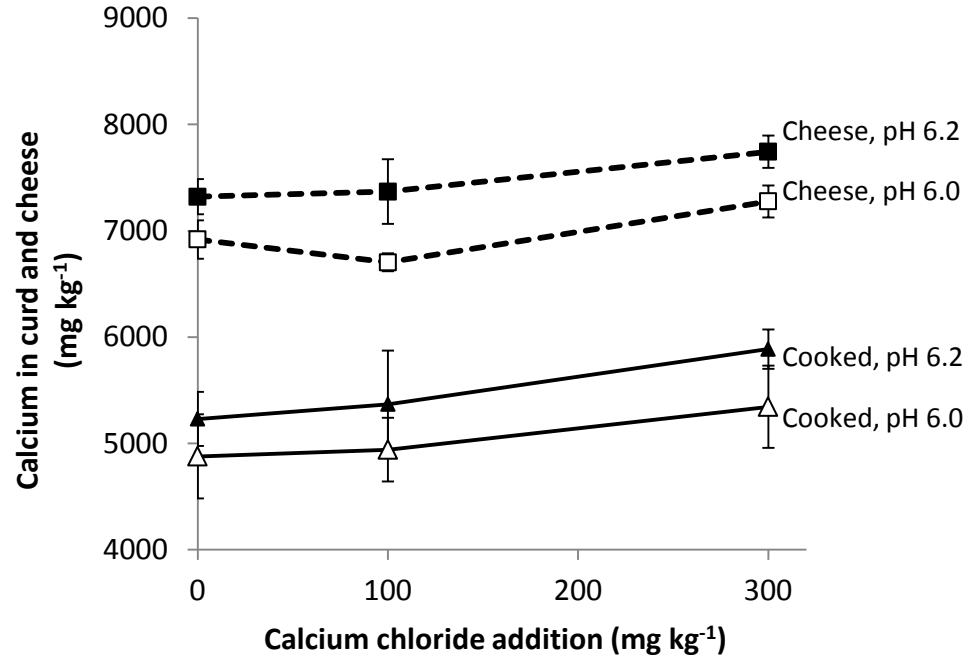
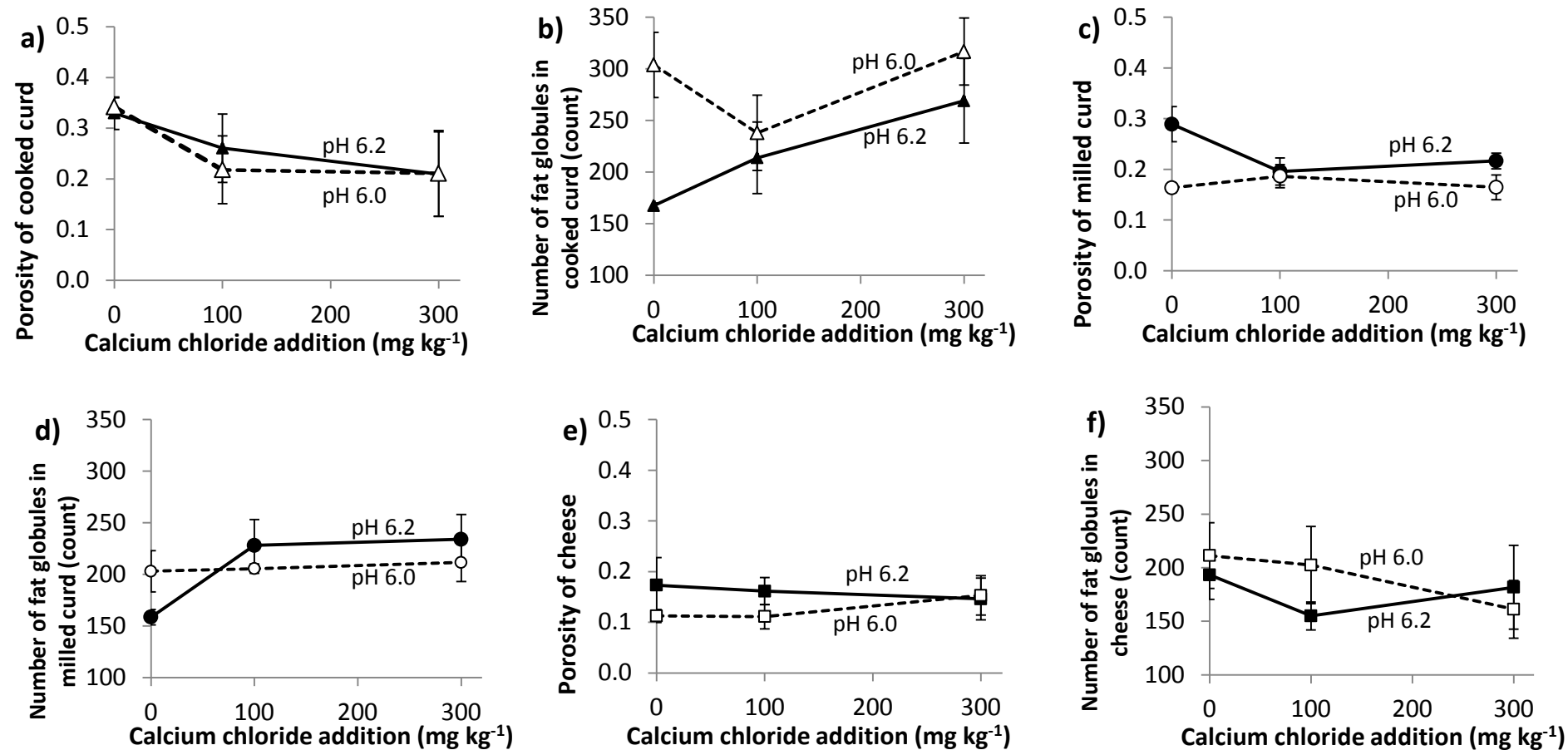
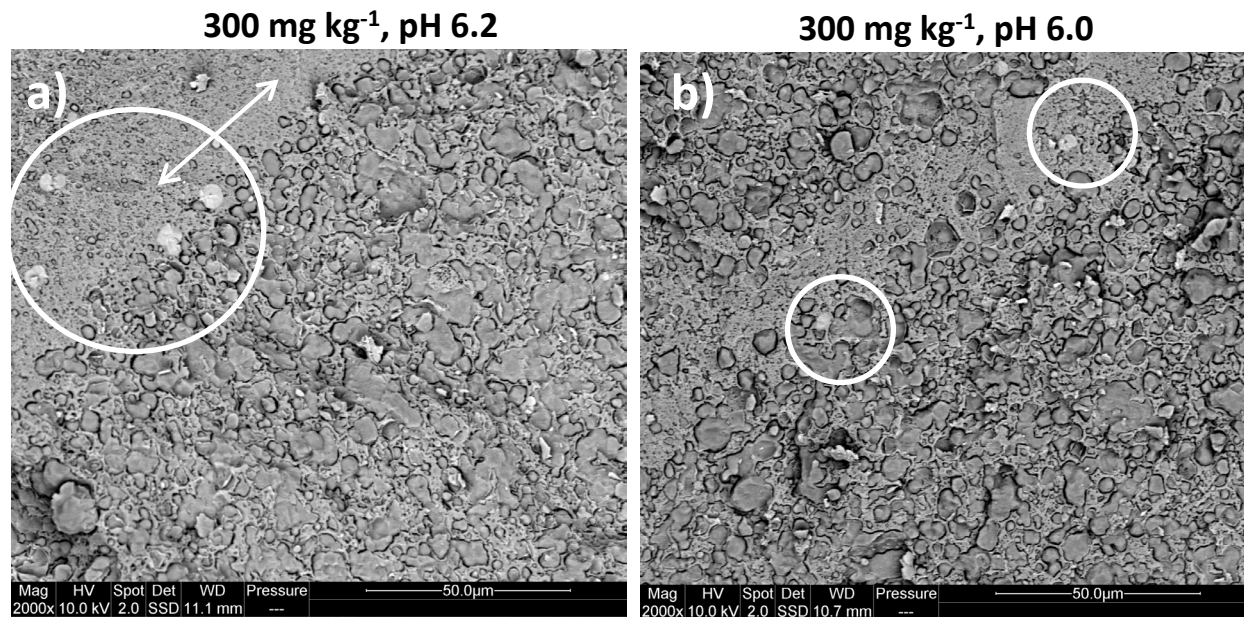


Fig. 9. The effect of CaCl₂ addition or whey draining pH on the level of total calcium in the curd collected after cooking (Cooked, ▲ or Δ with solid line) and after pressing (Cheese, ■ or □ with dotted line). The whey draining pH was 6.2 (closed markers, ■ or ▲) or pH 6.0 (open markers, □ or Δ). Error bars are the standard error of the mean (n = 4).



Supplementary Fig. 1. Physical properties of the cooked curd (a-b), milled curd (c-d) and cheese (e-f) made with different levels of CaCl₂ addition. All properties were determined by three dimensional image analysis of CLSM images. Results are presented as the mean ± the standard error of the mean (n = 4). The whey draining pH was 6.2 (closed marker, ■, ● or ▲ with solid line) or pH 6.0 (open markers, □, ○ or △ with dotted line).



Supplementary Fig. 2. Cryo SEM micrographs of milled curd made using cheese-milk with 300 mg kg⁻¹ CaCl₂ addition and with a whey draining pH of 6.2 (a) or pH 6.0 (b). The white circles indicate the presence of crystalline inclusions. Arrows indicate the curd junction. Scale bars within the images are 50 μm in length.



Minerva Access is the Institutional Repository of The University of Melbourne

Author/s:

Ong, L; Soodam, K; Kentish, SE; Powell, IB; Gras, SL

Title:

The addition of calcium chloride in combination with a lower draining pH to change the microstructure and improve fat retention in Cheddar cheese

Date:

2015-07-01

Citation:

Ong, L., Soodam, K., Kentish, S. E., Powell, I. B. & Gras, S. L. (2015). The addition of calcium chloride in combination with a lower draining pH to change the microstructure and improve fat retention in Cheddar cheese. *International Dairy Journal*, 46, pp.53-62.
<https://doi.org/10.1016/j.idairyj.2014.07.003>.

Persistent Link:

<http://hdl.handle.net/11343/238446>

File Description:

Accepted version