

Faculty Scholarship

2018

Mammal communities are larger and more diverse in moderately developed areas

Arielle Waldstein Parsons

Tavis Forrester

Megan C. Baker-Whatton

William J. McShea

CHristopher T. Rota

See next page for additional authors

Follow this and additional works at: https://researchrepository.wvu.edu/faculty_publications

Part of the Forest Sciences Commons, and the Plant Sciences Commons

Authors

Arielle Waldstein Parsons, Tavis Forrester, Megan C. Baker-Whatton, William J. McShea, CHristopher T. Rota, Stephanie G. Schuttler, Joshua J. Millspaugh, and Roland Kays



Mammal communities are larger and more diverse in moderately developed areas

Arielle Waldstein Parsons^{1,2*}, Tavis Forrester^{3,4}, Megan C Baker-Whatton⁵, William J McShea⁴, Christopher T Rota⁶, Stephanie G Schuttler¹, Joshua J Millspaugh⁷, Roland Kays^{1,2}

¹North Carolina Museum of Natural Sciences, Raleigh, United States; ²Department of Forestry & Environmental Resources, North Carolina State University, Raleigh, United States; ³Oregon Department of Fish and Wildlife, Gekeler Lane, United States; ⁴Smithsonian Conservation Biology Institute, Front Royal, United State; ⁵The Nature Conservancy, Fairfax Drive Arlington, Virginia; ⁶Division of Forestry and Natural Resources, Wildlife and Fisheries Resources Program, West Virginia University, Morgantown, United States; ⁷Wildlife Biology Program, Department of Ecosystem and Conservation Sciences, College of Forestry and Conservation, University of Montana, Missoula, United States

Abstract Developed areas are thought to have low species diversity, low animal abundance, few native predators, and thus low resilience and ecological function. Working with citizen scientist volunteers to survey mammals at 1427 sites across two development gradients (wild-rural-exurbansuburban-urban) and four plot types (large forests, small forest fragments, open areas and residential yards) in the eastern US, we show that developed areas actually had significantly higher or statistically similar mammalian occupancy, relative abundance, richness and diversity compared to wild areas. However, although some animals can thrive in suburbia, conservation of wild areas and preservation of green space within cities are needed to protect sensitive species and to give all species the chance to adapt and persist in the Anthropocene.

*For correspondence: arielle.parsons@naturalsciences. orq

Competing interests: The authors declare that no competing interests exist.

Funding: See page 10

Received: 02 May 2018 Accepted: 21 August 2018 Published: 02 October 2018

Reviewing editor: Bernhard Schmid, University of Zurich, Switzerland

© Copyright Parsons et al. This article is distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use and redistribution provided that the original author and source are

credited.

DOI: https://doi.org/10.7554/eLife.38012.001

Introduction

Global loss of biodiversity leads to disruption of ecosystem services around the world, ultimately threatening human well-being (Cardinale et al., 2012). Vertebrate species loss is typically considered to be worst in the most developed landscapes, where urbanization serves as an intense and long-term disturbance that permanently alters habitat and truncates food webs (Lombardi et al., 2017; McKinney, 2006). However, for some species, urbanization can offer abundant nutrient-rich food that is less ephemeral compared to wild areas (Bateman and Fleming, 2012; Wang et al., 2017). Whether this food is enough to counteract the negative effects of disturbance (i.e. higher road mortality, fragmentation) depends on a species' ability to adapt to the stressors of urban living (Witte et al., 1982). Mammal species, especially those with large home ranges, are arguably most at risk from development, leading some to suggest that developed areas have a dearth of predators, and that prey species could benefit by using humans as a shield (Crooks, 2002; Ordeñana et al., 2010). Previous studies have shown cities to be depauperate of bird life, supporting the traditional view that development and biodiversity cannot coexist (Keast, 1995; Strohbach et al., 2014).

However, recent evidence has shown that some mammal species previously thought mal-adapted to urban landscapes (i.e. mountain lion [Puma concolor], fisher [Martes pennanti]) are thriving in them (Bateman and Fleming, 2012; LaPoint et al., 2013), suggesting an evolutionary trend that could be important for conservation in the Anthropocene. Existing research on mammal

CC

Ecology

become ecologically unbalanced as they lose biodiversity. However, the preservation of urban green space and lack of hunting could also open the door for some species to thrive in the midst of large human populations. Indeed, some animals, mammals in particular, have grown more tolerant of humans and appear to have adapted to suburban landscapes around the world. Some species that have been exclusively living in the wilderness, such as a small carnivore called the fisher, are even moving back into cities.

Research into how mammals are coping with the urbanization of their habitats has produced conflicting results. Studies that explore a variety of cities and habitats would help to clear up this confusion.

Parsons et al. worked with citizen scientist volunteers to survey the mammals at 1,427 sites across Washington DC and Raleigh, North Carolina. The volunteers set up motion-triggered cameras in these sites, which covered a full range of urban and wild habitats, including back yards and large nature preserves.

The cameras detected similar or higher numbers of mammal species in suburban sites compared to wild areas. Indeed, most species appear to use suburban areas at least as much as wild land. Urban green space is especially important; it is used by less urban-adapted species like coyotes to navigate areas that are densely populated by humans.

The results presented by Parsons et al. suggest that many mammals have indeed adapted to the suburban environment over the last few decades, resulting in more balanced urban ecosystems. More testing in other cities will help to determine how general this pattern of adaptation is, and provide us with knowledge that could help us to conserve many different species. However, some species were still most abundant in wild areas, emphasizing the need to also conserve wildlands and to minimize our impact on natural ecosystems.

DOI: https://doi.org/10.7554/eLife.38012.002

communities across urbanization gradients has focused on single cities, yielding conflicting results, perhaps due to variation in city structure and characteristics (*Lombardi et al., 2017; Saito and Koike, 2013*). Given the rapid expansion of urban areas worldwide, and the recent case studies of urban adaptations by wildlife (*LaPoint et al., 2013; Riley et al., 2014; Wang et al., 2017*), more large-scale studies are needed to evaluate the response of wildlife communities to urban development if we are to understand urban ecology, conservation, and evolution in the Anthropocene.

Here, we present the results of a large-scale mammal survey of two urban-wild gradients. Our objectives were to determine how diversity, richness, detection rate, and occupancy of the mammal community change as a function of human disturbance. We hypothesized that the availability of supplemental food at higher levels of development would positively affect mammalian populations and outweigh the negative effects of disturbance, except for the most sensitive species. Specifically, we predicted that mammalian relative abundance would increase with developmental level but that species richness and diversity would decrease. Furthermore, we predicted that occupancy of the most sensitive species (i.e. large and medium carnivores) would be highest in wild areas both in our study area and around the world.

Materials and methods

Study sites

Washington, District of Columbia, USA (hereafter DC) is a city of approximately 177 km² with an estimated human population size of 681,000, thus a density of 3847 people/km². Our study spanned a 56,023.7 km² area around the city with a mean of 4.4 houses/km² and matrix of agriculture (~21.3%) and forest (~54.1%). Raleigh, North Carolina, USA (hereafter Raleigh) is approximately 375 km² with an estimated human population size of 459,000, thus a density of 1278 people/km². Our study spanned a 66,640 km^2 area around the city with a mean of 17.7 houses/km² and matrix of agriculture (~24.3%) and forest (~52.3%).

Citizen science camera trap surveys

From 2012_2016, 557 trained volunteers deployed 1427 unbaited camera traps across an urban-wild gradient around Raleigh and DC. Each individual camera was considered a 'camera site' and volunteers ran cameras at an average of two sites each. Following Hammer et al. (2004), we used the Silvis housing density dataset with 1km grid cells to define five development levels of the gradient for sampling stratification (excluding open water): urban (>1000 houses/km²), suburban (147.048-1000 houses/km²), exurban (12.64–147.047 houses/km²), rural (0.51–12.63 houses/km²) and wild (<0.5 houses/km²). Within those gradient levels, camera placement was also stratified between residential vards, open areas (>0.001 km² absent of trees), small forest fragments (< 21 km²) and large forest fragments (>1 km²) Supplementary file 1. Forest fragment size was verified using the 2006 US National Landcover Dataset (NLCD) and Landscape Fragmentation Tool v2.0 (Vogt et al., 2007) in ArcMap (Version 10.1, ESRI, Redlands, CA, USA) which defines forest fragments by size. Yards were not available for sampling in the urban or wild levels of the gradient. Urban areas were not sampled in Raleigh and open areas were not sampled in DC. All adjacent cameras were spaced at least 200 m apart. Camera placement was randomized as much as possible using ArcMap (Version 10.1) to randomly generate points within polygons while following certain rules. For example, we selected sites within forests that volunteers were permitted to access and were within a reasonable hiking distance (i.e. < 11 km hike round trip) with terrain that was not too steep to traverse safely (i.e. <45degree slope). Within yards, cameras were placed as randomly as possible while avoiding the highest human traffic areas (i.e. walkways, doors, gates and driveways).

No explicit power analysis was used to predetermine sample size. Our sample size goal was 20 spatial replicates (equating to ~420 trap nights), which has been found to maximize precision for estimating detection rate (*Kays et al., 2010; Rowcliffe et al., 2008*). Camera sites are biological replicates, parallel measurements capturing random biological variation. This study did not include technical replicates.

Volunteers used Reconyx (RC55, PC800, and PC900, Reconyx, Inc. Holmen, WI) and Bushnell (Trophy Cam HD, Bushnell Outdoor Products, Overland Park, KS) camera traps attached to trees at 40 cm above the ground. Cameras were deployed for three weeks and then moved to a new location without returning, with sampling taking place continuously throughout the year. Cameras recorded multiple photographs per trigger, at a rate of 1 frame/s, re-triggering immediately if the animal was still in view. We grouped consecutive photos into on sequence if they were <60 s apart, and used these sequences as independent records, counting animals in the sequence, not individual photos (*Parsons et al., 2016*). We then collapsed these independent records into daily detection/nondetection for occupancy modeling. Initial species identifications were made by volunteers using customized software (available freely from eMammal.org, source code proprietary) and all were subsequently reviewed for accuracy before being archived at the Smithsonian Digital Repository (*McShea et al., 2016*).

Diversity

We used package iNEXT (*Hsieh et al., 2016*) in R (Version 3.1.0; *R Development Core Team., 2008*) via R Studio (*RStudio Team, 2015*) to calculate Hill numbers (i.e. the effective number of species, incorporating relative abundance and richness) of species richness and Shannon diversity (*Chao et al., 2014*) between gradient levels (urban-suburban-exurban-rural-wild) and plot types (yard, open, small forest, large forest). iNEXT calculates the Shannon diversity as Hill number q = 1, equal to the exponential of Shannon's entropy index, thus the natural log of those results was used for display purposes. We used detection/non-detection data to compute diversity estimates and the associated 95% confidence intervals via rarefaction, plotting the diversity estimates while accounting for sample size. We fit a curve to diversity estimates between gradient levels using a generalized additive model with a polynomial term.

Model covariates

We modeled variation in occupancy (ψ) and detection rate using 13 covariates (Supplementary file 2) representing development level, the amount of core forest, small scale forest cover, prey relative abundance and whether hunting was allowed. We added year as a covariate to account for population changes over time. We used the Landscape Fragmentation Tool v2.0 (Vogt et al., 2007) and the NLCD (2006) land use dataset in ArcMap (Version 10.1) to create a landcover layer representing the percent of large core forest (forest patches larger than 1 km²) in a 5 km radius around camera locations which we considered best approximated the home range size of our target species (Bekoff, 1977; Fritzell and Haroldson, 1982; Lariviere and Pasitschniak-Arts, 1996; Lariviere and Walton, 1997). Forest patches did not necessarily fall entirely within the buffer. We considered road density as an additional covariate at the 5 km scale but initial evaluations showed it to be highly correlated with housing density (87.1%) so we chose to eliminate it from the analysis. We used a 100 m radius for small-scale forest cover to best represent small forest patches within suburban neighborhoods (e.g. small vacant lots with trees, greenways). We represented deer and rodent+lagomorph relative abundance using site-specific detection rate (the number of detections divided by the total number of camera-nights). We included an indicator (0/1, no hunting/hunting) to categorize whether a site allowed hunting or not. We modeled detection probability (p) using five covariates (Supplementary file 2). Because both ambient temperature and undergrowth can affect the camera's ability to detect an animal, we included daily covariates for temperature and NDVI (Moderate Resolution Imaging Land Terra Vegetation Indices 1 km monthly, an average value over the month(s) the camera ran) obtained from Env-DATA (Dodge et al., 2013). To complement NDVI, we also considered site-specific detection distance, a measure of how far away the camera was able to detect a human, which is influenced by both understory and site topography. We included an indicator (0/1, not yard/yard) to categorize whether a site was a residential yard or not. In Raleigh, two different camera models were used (both Reconyx and Bushnell) so we added a 0/1 (Bushnell/Reconyx) covariate to account for potential difference in detection probability between the two brands. We diagnosed univariate correlations between covariates using a Pearson correlation matrix, and used a restrictive prior for beta coefficients where correlation was >0.60 (i.e. logistic(0,1); a prior with reduced variance to induce shrinkage, similar to ridge regression; Hooten and Hobbs, 2015). All covariates were mean-centered.

Detection rate models

We used a Poisson count model (e.g. Kays et al., 2017) to assess differences in total mammal detection rate (i.e. the intensity with which a site was used, count/day) between the five gradient levels (urban, suburban, exurban, rural, wild) and four plot types (large forest, small forest, open, yard). We fit a curve to total detection rate estimates between gradient levels using a generalized additive model. No other covariates were used in this model. We then ran separate count models for four predator species (coyote (Canis latrans), gray fox (Urocyon cinereoargenteus), red fox (Vulpes vulpes) and bobcat [Lynx rufus]) to evaluate covariates of detection rate, running one fully-parameterized model (Supplementary file 2) to evaluate which explained the most variation in detection rate. We assessed model fit with posterior predictive checks (PPC) (Gelman et al., 2014; Kery and Schaub, 2012) by calculating the sum of squared Pearson residuals from observed data (T(y)) and from data simulated assuming the fully parameterized model was the data-generating model ($T(y_{sim})$). We calculated a Bayesian p-value as $p_B = Pr(T(y_{sim}) > T(y))$ from posterior simulations and assumed adequate fit if $0.1 < p_B < 0.9$ (Supplementary file 3). We fit the detection rate model in OpenBUGS v3.2.3 (Lunn et al., 2009) via R2OpenBUGS v3.2 (Sturtz et al., 2005) in R (Version 3.1.0) via R Studio. We based inference on posterior samples generated from three Markov chains, using trace plots to determine an adequate burn-in phase. All models achieved adequate convergence (R < 1.1) (Gelman et al., 2014) by running for 50,000 iterations following a burn-in phase of 1000 iterations, thinning every 10 iterations. We based significance on whether parameter 95% credible intervals overlapped zero.

Occupancy models

We used the multispecies occupancy model of **Rota et al. (2016)** to estimate the probability of occupancy of four predator species: bobcat, coyote, red fox and gray fox. Although we are using

the term occupancy, because data were collected from camera traps estimates are more analogous to 'use' than true occupancy (**Burton et al., 2015**). This model is distinct from the classic multispecies community models (**Dorazio and Royle, 2005;Dorazio et al., 2006; Gelfand et al., 2005**) and is rather a generalization of the single-season occupancy model (**MacKenzie et al., 2002**) to accommodate two or more interacting species. It contains single-species (first order) occupancy models for each interacting species alone as well as pairwise (second order) models for the co-occurrence of each pair of species (**Rota et al., 2016**). For each species and pairwise interaction, the model estimates detection probability (*p*), defined as the probability of detecting an occurring species at a camera site, and occupancy (ψ), defined as the probability that a given camera site is occupied, for each species. The latent occupancy state of each species at a site is modeled as a multivariate Bernoulli random variable such that (assuming 2 interacting species):

$Z \sim MVB(\psi_{11}, \psi_{10}, \psi_{01}, \psi_{00})$

Where ψ_{11} is the probability that both species occupy a site, ψ_{10} is the probability that only species 1 occupies a site, ψ_{01} is the probability that only species 2 occupies a site and ψ_{00} is the probability that neither species occupies a site. We assumed all species occurred independently and considered the same set of five covariates for the detection probability models and 13 covariates in the occupancy model of each species (**Supplementary file 2**). We considered interactions (i.e. city*covariate) between each occupancy covariate and city (0/1, DC/Raleigh). We estimated occupancy for each species across levels of the development gradient (urban, suburban, exurban, rural, wild) and plot types (yard, open, small forest, large forest) within each city separately by including development level and plot type as categorical covariates in our model.

We fit models in STAN (Version 2.15.1; Stan Development Team, 2015b) via the RSTAN (Version 2.15.1; Stan Development Team, 2015a) interface in R (Version 3.4.0) via R Studio (Version 1.0.143). We based inference on posterior samples generated from two Markov chains, using trace plots to determine an adequate burn-in phase and subsequently running chains until they reached adequate convergence (R>1.1) (Gelman et al., 2014). All models achieved adequate convergence by running for 3000 iterations following a burn-in phase of 1000 iterations. We based predictor significance on whether beta coefficient 95% credible intervals overlapped zero. We assessed model fit with posterior predictive checks (PPC) (Gelman et al., 2014; Kery and Schaub, 2012) by calculating the sum of squared Pearson residuals from observed data (T(y)) and from data simulated assuming the fully parameterized model was the data-generating model ($T(y_{sim})$). We calculated a Bayesian pvalue as $p_B = \Pr(T(y_{sim}) > T(y))$ from posterior simulations and assumed adequate fit if $0.1 < p_B < 0.9$. To our knowledge, the squared Pearson's residual has not been derived in the context of occupancy models, so we present our derivation of this test statistic in Supplementary file 4. We added a random effect on detection/non-detection for the covote portion of the model since initial assessments of fit for this species were inadequate (i.e. $p_B > 0.9$). We assessed differences in occupancy between gradient levels for each species using overlapping 95% confidence intervals.

Comparison with global occupancy data

We removed omnivores from the dataset of **Rich et al. (2017)** to better compare with carnivore occupancy from our own dataset. Where species occupancy was estimated from multiple studies in the Rich et al. dataset, we calculated averages to compare to occupancy estimates from our own study. We summarized occupancy estimates of Rich et al. and our own study within each developmental level using a box and whisker plot and assessed statistically significant differences based on whether or not interquartile ranges overlapped.

Data accessibility

Raw detections data have been deposited in Data Dryad, doi:10.5061/dryad.11rf64v. The software used for initial species identifications is available via eMammal.org. To download and use the software, users must first create an account on eMammal and become associated with an existing project. This can be done by using the 'Join' button on the project's homepage, or by emailing the contact person, also listed on the project homepage. Usually the user will also have to pass an online or in person training, depending on the project requirements, to be approved to download the software.

Results and discussion

Working with citizen scientist volunteers, we obtained 53,273 detections of 19 mammal species at 1427 sites along an urban-wild gradient in Washington, DC and Raleigh, NC, USA, sampling both private and public lands. In DC, we detected 17 mammal species with mean naïve occupancy of 0.19 (min = 0, max = 0.93) and mean detection rate of 0.09 detections/day (min = 0, max = 1.05). In Raleigh, we detected 17 mammal species with mean naïve occupancy of 0.14 (min = 0, max = 0.79) and mean detections/day (min = 0, max = 0.09).

We found no significant decline of species diversity or richness from suburban to wild gradient levels (*Figure 2—figure supplement 1, Figure 1*). However, Shannon diversity was significantly lower at the urban level in DC, possibly due to low sampling (*Figure 2, Supplementary file 1*). Diversity in yards was significantly higher or not statistically different from large and small forest fragments in both cities (*Figure 2—figure supplements 2,3*). Most (92.3%) of the 13 mammal species

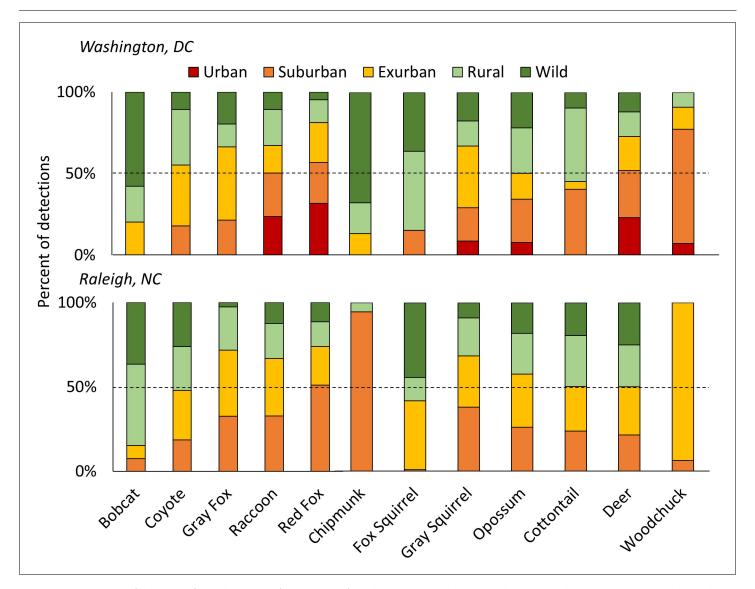


Figure 1. The percent of detections for each species of carnivores (left) and herbivores (right) in each development level along the urban-wild gradient in Washington, DC and Raleigh, NC, USA accounting for the effort (i.e. camera nights) within each level, sorted from lowest to highest proportion urban/suburban in DC. The dashed line shows 50% of total detections. Some species were predominantly rural/wild (i.e. bobcats and fox squirrels) while others were mainly detected in urban/suburban habitats (i.e. red fox, raccoon). Patchy distributions at different gradient levels were seen for species at the edge of their ranges (i.e. chipmunks and woodchucks in Raleigh). Urban habitats were not sampled in Raleigh. DOI: https://doi.org/10.7554/eLife.38012.003

eLIFE Short report

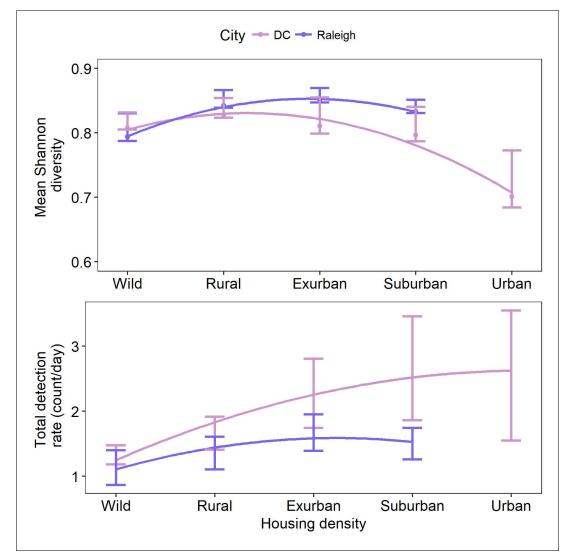


Figure 2. Mean Shannon diversity and total detection rate along a gradient of housing density in two cities, Washington, DC and Raleigh, NC USA taken from camera traps. Bars show 95% confidence intervals, lines are fit using a generalized additive model with a polynomial term. Diversity peaked at intermediate levels of urbanization (exurban in DC and suburban in Raleigh). Total detection rate peaked at the urban level in DC and exurban level in Raleigh.

DOI: https://doi.org/10.7554/eLife.38012.004

The following figure supplements are available for figure 2:

Figure supplement 1. Rarefaction curves estimating species richness in five development levels (urban, suburban, exurban, rural, wild) in two cities, Washington, DC and Raleigh, NC, USA, using camera traps between 2012 and 2016.

DOI: https://doi.org/10.7554/eLife.38012.005

Figure supplement 2. Shannon diversity index estimates from camera trapping in two cities, Washington, DC and Raleigh, NC, USA, across five development levels (urban, suburban, exurban, rural, wild).

DOI: https://doi.org/10.7554/eLife.38012.006

Figure supplement 3. Rarefaction curve estimating species richness in three plot types (residential yard, small forest, large forest) in two cities, Washington, DC and Raleigh, NC, USA, using camera traps between 2012 and 2016.

DOI: https://doi.org/10.7554/eLife.38012.007

detected >20 times occupied all levels of development below the urban level. Two of the largest predators, coyotes and bobcats, were absent from the highest development level (urban) but were detected at all other levels in both cities. Black bears (*Ursus americanus*), which are actively discouraged from colonizing central North Carolina (*North Carolina Wildlife Resources Commission, 2011*), were not detected in Raleigh and were detected in DC at all levels of the gradient except

suburban and urban, though were predominantly in the wild level. These results indicate that the extant mammal guild exploits all levels of the urban-wild gradient and that no species are entirely relegated to the wild gradient level. However, some species appear less adapted to habitation in human-dominated areas, spending most of their time at the wild levels of the gradient (i.e. bobcat, bear; *Figure 1*). We recognize that the current community represents species that survived the initial arrival of high-density human settlement. In particular, two large predators (wolves (*Canis lupus*) and cougars [*Puma concolor*]) were extirpated from our study area a century ago. However, even cougars and wolves have recently shown surprising adaptability in the face of development at other sites (*Bateman and Fleming, 2012; Wang et al., 2017*) suggesting that, given enough time and protection from persecution, many of the most 'wild' of species may adapt to human development.

Predators are thought to be the most at risk from urbanization (*Crooks, 2002*), therefore, we evaluated predictors for occupancy (*MacKenzie et al., 2002*) and detection rate (*Kays et al., 2017*) for four carnivores: coyote, gray fox, red fox, and bobcat. Both of our models fit well, with Bayesian p-values between 0.1 and 0.9 (*Supplementary file 3*). Suburban and urban occupancy probabilities were not statistically different from wild for any of the species (*Figure 3—figure supplement 1*) and we noted a decreasing trend in occupancy from urban to wild (*Figure 3*). We compared the occupancy estimates from our study to those reported for carnivores in protected areas around the world (*Rich et al., 2017*) and found no significant difference (*Figure 3*), suggesting that the ecological function of predators in this urban system is not substantially reduced from the current wild state, excepting the historical extirpation of the two largest native predators from the region.

Our occupancy and detection rate models yielded similar results (**Supplementary file 5–7**) demonstrating that green space is important to carnivore species that are less-adapted to human-altered landscapes. These models show a greater association of carnivores with green space when housing density is high (e.g. coyote and gray fox, **Supplementary file 6**, **7**), consistent with other studies finding urban green space important in maintaining biodiversity in cities (**Gallo et al., 2017**; **Lombardi et al., 2017**; **Matthies et al., 2017**). It is likely that shyer species are not avoiding regions of high human density, but require patches of forest to navigate residential areas that are freely used by more commensal species, such as red foxes (**Tigas et al., 2002**), which we frequently detected in yards. Indeed, we found a gradient of responses in carnivore use of human-dominated environments, from red fox which are the most urban adapted (i.e. negatively associated with local large forest fragments and the only species to have a positive association with yards) to bobcats which appear to be the most human-averse (i.e. rarely detected in the suburban level of the gradient) (**Figure 1; Figure 3—figure supplement 1**).

Contrary to expectations, we found no evidence for a negative impact of suburban and exurban development on extant native mammal diversity, richness, and occupancy and detection rate of carnivores. In fact, all metrics were significantly greater than, or equal to, wild areas. We suspect that developed areas offer good food resources for wildlife through direct and indirect feeding (i.e. bird feeders supplementing prey, pets), accidental feeding (i.e. garbage), and ornamental plantings (for herbivores), but testing this hypothesis will require additional diet studies in urban landscapes (*Contesse et al., 2004*). Furthermore, the structure of mature suburbia (i.e. older, established neighborhoods with large trees, wooded riparian areas, small parks) contributes to a more diverse and varied landscape than wild areas with more homogenous forest cover, which is potentially beneficial for many generalist species. Developed areas where hunting is limited or prohibited also offer a safe haven for game species, presuming they can navigate the road networks (*Collins and Kays, 2011*) and avoid direct human conflict.

Our discovery of a wild suburbia suggests high levels of adaptation by mammals to developed landscapes over the last few decades, including predators and prey. The resilience of these species gives hope for wildlife in the Anthropocene, but the generality of this pattern needs to be tested in other cities to understand how habitat type, development patterns, apex predators, and hunting regulations influence urban mammal communities, as there are examples of far more drastic impacts of urbanization on other taxa and in other places around the globe (*Keast, 1995; McKinney, 2008*). Indeed, in Tokyo, Japan, the relative abundance of mammals declined with urbanization (*Saito and Koike, 2013*) and avian communities in Quebec, Canada and Rennes, France showed a similar decline in richness (*Clergeau et al., 1998; Saito and Koike, 2013*). This suggests that city structure, size and human density may influence mammalian distribution along urban-wild gradients with large, sprawling New World cities showing different patterns than the smaller more concentrated cities of

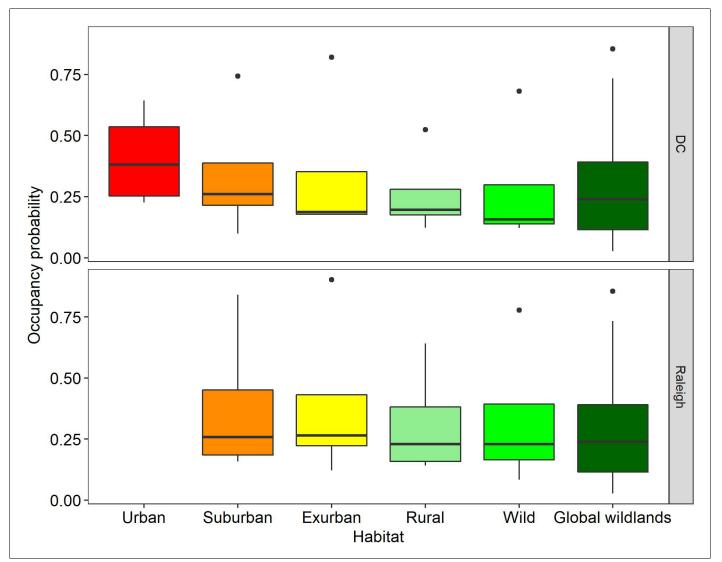


Figure 3. Comparison of carnivore (i.e.bobcat, coyote, gray fox, red fox) occupancy probabilities at each developmental level in two cities with global values from Rich et al. (*Rich et al., 2017*), representing 93 carnivores from 13 protected areas on five continents (Global Wildlands). Each box for our dataset represents the distribution of marginal occupancy probabilities for each of four carnivore species in that city (i.e. four probabilities). The boxes for Global Wildlands represent the distribution of marginal occupancy probabilities for 93 species. We found no statistically significant differences between any habitat levels in our study or between our study and global wildland occupancy probabilities but noted a decreasing trend in occupancy from urban-wild. We included only predators from *Rich et al. (2017)* and removed omnivores (i.e. raccoon, coati) to better reflect our data. DOI: https://doi.org/10.7554/eLife.38012.008

The following figure supplement is available for figure 3:

Figure supplement 1. Occupancy estimates from single season occupancy model for four carnivore species (bobcat, coyote, gray fox and red fox) in five development levels (urban, suburban, exurban, rural, wild) in two cities, Washington, DC and Raleigh, NC, USA, using camera traps between 2012 and 2016.

DOI: https://doi.org/10.7554/eLife.38012.009

the Old World. Although our study provides a less dire picture of urban ecosystem function than previously thought, we do not suggest abandoning mitigation of urbanization's negative impacts, or conservation of completely wild areas. Factors such as urban green space, connectivity and availability of completely wild areas give species the time and space to adapt to changing habitats and climates. Further understanding of how urban wildlife navigates human-dominated areas and factors that contribute to the adaptation of species to the Anthropocene will be critical to maintaining diversity in a rapidly urbanizing world.

Acknowledgements

We thank our 557 volunteers for their hard work collecting camera trap data for this study. For their field assistance and volunteer coordination we thank the staff of the NPS, USFWS, USFS, TNC, NC, VA, and MD State Parks, NCWRC and VDGIF. We thank A Mash, N Fuentes, S Higdon, T Perkins, L Gatens, R Owens, R Gayle, C Backman, K Clark, J Grimes and J Simkins for their help reviewing photographs. This work was conducted with funding from the National Science Foundation [grant #1232442 and #1319293], the USDA National Institute of Food and Agriculture, the VWR Foundation, the US Forest Service, the North Carolina Museum of Natural Sciences and the Smithsonian Institution. We thank M Katti and R Dunn for comments on early versions of this manuscript.

Additional information

Funding

Funder	Grant reference number	Author		
National Science Foundation	#1232442	William J McShea Roland Kays		
National Science Foundation	#1319293	William J McShea Roland Kays		
VWR Foundation		William J McShea Roland Kays		
International Programs, US Forest Service		William J McShea Roland Kays		
Smithsonian Institution		William J McShea		
National Institute of Food and Agriculture	McIntire Stennis project , WVA00124	Christopher T Rota		
North Carolina Museum of Natural Sciences		Roland Kays		

The funders had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

Author contributions

Arielle Waldstein Parsons, Conceptualization, Data curation, Formal analysis, Validation, Investigation, Visualization, Methodology, Writing—original draft, Project administration, Writing—review and editing; Tavis Forrester, Conceptualization, Supervision, Funding acquisition, Investigation, Methodology, Project administration, Writing—review and editing; Megan C Baker-Whatton, Conceptualization, Supervision, Investigation, Methodology, Project administration, Writing—review and editing; William J McShea, Conceptualization, Data curation, Supervision, Funding acquisition, Investigation, Methodology, Project administration, Writing—review and editing; Christopher T Rota, Formal analysis, Visualization, Writing—review and editing; Stephanie G Schuttler, Investigation, Writing—review and editing; Joshua J Millspaugh, Formal analysis, Writing—review and editing; Roland Kays, Conceptualization, Supervision, Funding acquisition, Investigation, Methodology, Writing—review and editing

Author ORCIDs

Arielle Waldstein Parsons in http://orcid.org/0000-0003-1076-2896

Decision letter and Author response

Decision letter https://doi.org/10.7554/eLife.38012.021 Author response https://doi.org/10.7554/eLife.38012.022

Additional files

Supplementary files

• Supplementary file 1. Effort expressed as camera nights with spatial replicates in parentheses for camera traps run in Washington, DC and Raleigh, NC from 2012 to 2016 between different levels along the urban-wild gradient around each city.

DOI: https://doi.org/10.7554/eLife.38012.010

• Supplementary file 2. Covariates used in the detection rate and occupancy analyses. DOI: https://doi.org/10.7554/eLife.38012.011

• Supplementary file 3. Results of goodness-of-fit tests for occupancy and Poisson count models assessed by posterior predictive check with adequate fit if 0.1 < pB < 0.9. DOI: https://doi.org/10.7554/eLife.38012.012

• Supplementary file 4. Calculating goodness of fit statistics DOI: https://doi.org/10.7554/eLife.38012.013

• Supplementary file 5. Results of a Poisson regression to determine differences in detection rate between the wild gradient level (reference level) and all other levels of the development gradient (above dotted line). Also presented are results of a separate Poisson regression to determine differences in detection rate between yards and all other plot types. Significant results (95% CIs not overlapping zero) are in bold.

DOI: https://doi.org/10.7554/eLife.38012.014

• Supplementary file 6. Results of Poisson regression for single species using camera trapping over two cities between 2012–2016. Posterior mean and posterior standard deviation for each predictor are shown with bold entries indicating predictors with 95% credible intervals that did not overlap zero. Predictors in bold were used for modeling occupancy for that species. Housing density was used as a predictor in all occupancy models, regardless of whether it was significant in the preliminary count analysis.

DOI: https://doi.org/10.7554/eLife.38012.015

• Supplementary file 7. Beta coefficients and 95% credible intervals (parentheses) for an occupancy model based on camera trapping data in Washington, DC, USA and Raleigh, NC, USA from 2012 to 2016. Those coefficients with 95% CIs not overlapping zero are shown in bold. DOI: https://doi.org/10.7554/eLife.38012.016

• Transparent reporting form

DOI: https://doi.org/10.7554/eLife.38012.017

Data availability

Raw detections data have been deposited in Data Dryad, doi:10.5061/dryad.11rf64v. The software used for initial species identification is available via eMammal.org. To download and use the software, users must first create an account on eMammal and become associated with an existing project. This can be done by using the 'Join' button on the project's homepage, or by emailing the contact person, also listed on the project homepage. Usually the user will also have to pass an online or in person training, depending on the project requirements, and they will then become approved to download the software.

The following dataset was generated:

Author(s)	Year	Dataset title	Dataset URL	Database, license, and accessibility information
Arielle Waldstein Parsons, Tavis For- rester, Megan C Baker-Whatton, Wil- liam McShea, Chris- topher T Rota, Stephanie G Schut- tler, Joshua J Mill- spaugh, Roland Kays	2018	Wild suburbia: mammal communities are larger and more diverse in moderately developed areas	http://dx.doi.org/10. 5061/dryad.11rf64v	Available at Dryad Digital Repository under a CC0 Public Domain Dedication

References

- Bateman PW, Fleming PA. 2012. Big City life: carnivores in urban environments. Journal of Zoology 287:1–23. DOI: https://doi.org/10.1111/j.1469-7998.2011.00887.x
- Bekoff M. 1977. Canis latrans. Mammalian Species 79:1-9. DOI: https://doi.org/10.2307/3503817
- Burton AC, Neilson E, Moreira D, Ladle A, Steenweg R, Fisher JT, Bayne E, Boutin S. 2015. REVIEW: Wildlife camera trapping: a review and recommendations for linking surveys to ecological processes. *Journal of Applied Ecology* 52:675–685. DOI: https://doi.org/10.1111/1365-2664.12432
- Cardinale BJ, Duffy JE, Gonzalez A, Hooper DU, Perrings C, Venail P, Narwani A, Mace GM, Tilman D, Wardle DA, Kinzig AP, Daily GC, Loreau M, Grace JB, Larigauderie A, Srivastava DS, Naeem S. 2012. Biodiversity loss and its impact on humanity. Nature 486:59–67. DOI: https://doi.org/10.1038/nature11148, PMID: 22678280
- Chao A, Gotelli NJ, Hsieh TC, Sander EL, Ma KH, Colwell RK, Ellison AM. 2014. Rarefaction and extrapolation with Hill numbers: a framework for sampling and estimation in species diversity studies. *Ecological Monographs* 84:45–67. DOI: https://doi.org/10.1890/13-0133.1
- Clergeau P, Savard J-PL, Mennechez G, Falardeau G. 1998. Bird abundance and diversity along an Urban-Rural gradient: a comparative study between two cities on different continents. *The Condor* **100**:413–425. DOI: https://doi.org/10.2307/1369707
- Collins C, Kays R. 2011. Causes of mortality in north american populations of large and medium-sized mammals. Animal Conservation 14:474–483. DOI: https://doi.org/10.1111/j.1469-1795.2011.00458.x
- Contesse P, Hegglin D, Gloor S, Bontadina F, Deplazes P. 2004. The diet of urban foxes (Vulpes vulpes) and the availability of anthropogenic food in the city of Zurich, Switzerland. Mammalian Biology - Zeitschrift Für Säugetierkunde 69:81–95. DOI: https://doi.org/10.1078/1616-5047-00123
- Crooks KR. 2002. Relative sensitivities of mammalian carnivores to habitat fragmentation. Conservation Biology 16:488–502. DOI: https://doi.org/10.1046/j.1523-1739.2002.00386.x
- Dodge S, Bohrer G, Weinzierl R, Davidson SC, Kays R, Douglas D, Cruz S, Han J, Brandes D, Wikelski M. 2013. The environmental-data automated track annotation (Env-DATA) system: linking animal tracks with environmental data. *Movement Ecology* 1:3. DOI: https://doi.org/10.1186/2051-3933-1-3, PMID: 25709817
- Dorazio RM, Royle JA, Söderström B, Glimskär A. 2006. Estimating species richness and accumulation by modeling species occurrence and detectability. *Ecology* 87:842–854. DOI: https://doi.org/10.1890/0012-9658 (2006)87[842:ESRAAB]2.0.CO;2, PMID: 16676528
- **Dorazio RM**, Royle JA. 2005. Estimating size and composition of biological communities by modeling the occurrence of species. *Journal of the American Statistical Association* **100**:389–398. DOI: https://doi.org/10. 1198/016214505000000015
- Fritzell EK, Haroldson KJ. 1982. Urocyon cinereoargenteus. Mammalian Species 189:1–8. DOI: https://doi.org/ 10.2307/3503957
- Gallo T, Fidino M, Lehrer EW, Magle SB. 2017. Mammal diversity and metacommunity dynamics in urban green spaces: implications for urban wildlife conservation. *Ecological Applications* 27:2330–2341. DOI: https://doi.org/10.1002/eap.1611, PMID: 28833978
- Gelfand AE, Schmidt AM, Wu S, Silander JA, Latimer A, Rebelo AG. 2005. Modelling species diversity through species level hierarchical modelling. *Journal of the Royal Statistical Society: Series C* 54:1–20. DOI: https://doi.org/10.1111/j.1467-9876.2005.00466.x
- Gelman A, Carlin JB, Stern HS, Dunson DB, Vehtari A, Rubin DB. 2014. Bayesian Data Analysis. Third edition. CRC Press.
- Hammer RB, Stewart SI, Winkler RL, Radeloff VC, Voss PR. 2004. Characterizing dynamic spatial and temporal residential density patterns from 1940–1990 across the North Central United States. *Landscape and Urban Planning* **69**:183–199. DOI: https://doi.org/10.1016/j.landurbplan.2003.08.011
- Hooten MB, Hobbs NT. 2015. A guide to Bayesian model selection for ecologists. *Ecological Monographs* **85**:3–28. DOI: https://doi.org/10.1890/14-0661.1
- Hsieh TC, Ma KH, Chao A. 2016. iNEXT: iNterpolation and EXTrapolation for Species Diversity . (Version 2.0.12).
- Kays R, Parsons AW, Baker MC, Kalies EL, Forrester T, Costello R, Rota CT, Millspaugh JJ, McShea WJ. 2017. Does hunting or hiking affect wildlife communities in protected areas? *Journal of Applied Ecology* 54:242–252. DOI: https://doi.org/10.1111/1365-2664.12700
- Kays R, Tilak S, Kranstauber B, Jansen PA, Carbone C, Rowcliffe MJ, He Z. 2010. Monitoring wild animal communities with arrays of motion sensitive Camera traps. *arXiv*.
- Keast A. 1995. Habitat loss and species loss: the birds of Sydney 50 years ago and now. Australian Zoologist **30**: 3–25. DOI: https://doi.org/10.7882/AZ.1995.002
- Kery M, Schaub M. 2012. Bayesian Population Analysis Using Win-BUGS: A Hierarchical Perspective. Waltham: Academic Press.
- Kéry M, Royle JA. 2015. Applied Hierarchical Modeling in Ecology: Analysis of Distribution, Abundance and Species Richness in R and BUGS: Volume 1: Prelude and Static Models. Amsterdam: Academic Press.
- LaPoint 5, Gallery P, Wikelski M, Kays R. 2013. Animal behavior, cost-based corridor models, and real corridors. Landscape Ecology 28:1615–1630. DOI: https://doi.org/10.1007/s10980-013-9910-0
- Lariviere S, Pasitschniak-Arts M. 1996. Vulpes vulpes. Mammalian Species 537:1–11. DOI: https://doi.org/10. 2307/3504236
- Lariviere S, Walton LR. 1997. Lynx rufus. Mammalian Species 563:1–8. DOI: https://doi.org/10.2307/3504533

- Lombardi JV, Comer CE, Scognamillo DG, Conway WC. 2017. Coyote, fox, and bobcat response to anthropogenic and natural landscape features in a small urban area. *Urban Ecosystems* **20**:1239–1248. DOI: https://doi.org/10.1007/s11252-017-0676-z
- Lunn D, Spiegelhalter D, Thomas A, Best N. 2009. The BUGS project: Evolution, critique and future directions. Statistics in Medicine 28:3049–3067. DOI: https://doi.org/10.1002/sim.3680, PMID: 19630097
- MacKenzie DI, Nichols JD, Lachman GB, Droege S, Andrew Royle J, Langtimm CA. 2002. Estimating site occupancy rates when detection probabilities are less than one. *Ecology* 83:2248–2255. DOI: https://doi.org/ 10.1890/0012-9658(2002)083[2248:ESORWD]2.0.CO;2
- Matthies SA, Rüter S, Schaarschmidt F, Prasse R. 2017. Determinants of species richness within and across taxonomic groups in urban green spaces. Urban Ecosystems 20:897–909. DOI: https://doi.org/10.1007/s11252-017-0642-9
- McKinney ML. 2006. Urbanization as a major cause of biotic homogenization. *Biological Conservation* **127**:247–260. DOI: https://doi.org/10.1016/j.biocon.2005.09.005
- McKinney ML. 2008. Effects of urbanization on species richness: A review of plants and animals. Urban Ecosystems 11:161–176. DOI: https://doi.org/10.1007/s11252-007-0045-4
- McShea WJ, Forrester T, Costello R, He Z, Kays R. 2016. Volunteer-run cameras as distributed sensors for macrosystem mammal research. Landscape Ecology 31:55–66. DOI: https://doi.org/10.1007/s10980-015-0262-9

North Carolina Wildlife Resources Commission. 2011. North Carolina Black Bear Managementplan 2012-2022. Ordeñana MA, Crooks KR, Boydston EE, Fisher RN, Lyren LM, Siudyla S, Haas CD, Harris S, Hathaway SA,

- Turschak GM, Miles AK, Van Vuren DH. 2010. Effects of urbanization on carnivore species distribution and richness. *Journal of Mammalogy* **91**:1322–1331. DOI: https://doi.org/10.1644/09-MAMM-A-312.1
- Parsons AW, Bland C, Forrester T, Baker-Whatton MC, Schuttler SG, McShea WJ, Costello R, Kays R. 2016. The ecological impact of humans and dogs on wildlife in protected areas in eastern North America. *Biological Conservation* 203:75–88. DOI: https://doi.org/10.1016/j.biocon.2016.09.001
- **R Development Core Team.** 2008. R: A language and environment for statistical computing. *R Foundation for Statistical Computing*. 3.4.0. Vienna, Austria:
- Rich LN, Davis CL, Farris ZJ, Miller DAW, Tucker JM, Hamel S, Farhadinia MS, Steenweg R, Di Bitetti MS, Thapa K, Kane MD, Sunarto S, Robinson NP, Paviolo A, Cruz P, Martins Q, Gholikhani N, Taktehrani A, Whittington J, Widodo FA, et al. 2017. Assessing global patterns in mammalian carnivore occupancy and richness by integrating local camera trap surveys. *Global Ecology and Biogeography* 26:918–929. DOI: https://doi.org/10. 1111/geb.12600
- Riley SP, Serieys LE, Pollinger JP, Sikich JA, Dalbeck L, Wayne RK, Ernest HB. 2014. Individual behaviors dominate the dynamics of an urban mountain lion population isolated by roads. *Current Biology* 24:1989–1994. DOI: https://doi.org/10.1016/j.cub.2014.07.029, PMID: 25131676
- Rota CT, Ferreira MAR, Kays RW, Forrester TD, Kalies EL, McShea WJ, Parsons AW, Millspaugh JJ, Warton D. 2016. A multispecies occupancy model for two or more interacting species. *Methods in Ecology and Evolution* 7:1164–1173. DOI: https://doi.org/10.1111/2041-210X.12587
- **Rowcliffe JM**, Field J, Turvey ST, Carbone C. 2008. Estimating animal density using camera traps without the need for individual recognition. *Journal of Applied Ecology* **45**:1228–1236. DOI: https://doi.org/10.1111/j. 1365-2664.2008.01473.x
- RStudio Team. 2015. RStudio: Integrated Development for R. 1.0.143. Boston, RStudio, Inc.
- Saito M, Koike F. 2013. Distribution of wild mammal assemblages along an urban-rural-forest landscape gradient in warm-temperate east asia. *PLoS ONE* 8:e65464. DOI: https://doi.org/10.1371/journal.pone.0065464, PMID: 23741495
- Stan Development Team. 2015b. Stan: a C++ library for probability and sampling.
- Strohbach MW, Hrycyna A, Warren PS. 2014. 150 years of changes in bird life in Cambridge, Massachusetts from 1860 to 2012. The Wilson Journal of Ornithology **126**:192–206. DOI: https://doi.org/10.1676/13-127.1
- Sturtz S, Ligges U, Gelman A. 2005. R2WinBUGS: a package for running WinBUGS from R. *Journal of Statistical* Software **12**:1–16.
- Stan Development Team. 2015a. Rstan: the R interface to Stan.
- Tigas LA, Van Vuren DH, Sauvajot RM. 2002. Behavioral responses of bobcats and coyotes to habitat fragmentation and corridors in an urban environment. *Biological Conservation* **108**:299–306. DOI: https://doi.org/10.1016/S0006-3207(02)00120-9
- Vogt P, Riitters KH, Estreguil C, Kozak J, Wade TG, Wickham JD. 2007. Mapping spatial patterns with morphological image processing. *Landscape Ecology* 22:171–177. DOI: https://doi.org/10.1007/s10980-006-9013-2
- Wang Y, Smith JA, Wilmers CC. 2017. Residential development alters behavior, movement, and energetics in an apex predator, the puma. *PLoS One* **12**:e0184687. DOI: https://doi.org/10.1371/journal.pone.0184687, PMID: 29020087
- Witte R, Diesing D, Godde M. 1982. Urbanophobe, urbanoneutral, urbanophile behavior of species concerning the urban habitat. Flora : The Journal of Infectious Diseses and Clinical Microbiology = Infeksiyon Hastalıkları Ve Klinik Mikrobiyoloji Dergisi 177:265–282.