

Title: Starch replacement in gluten free bread by cellulose and fibrillated cellulose

Author names and affiliations:

Yi Ren¹ yi.ren@nottingham.ac.uk +44(0)1159516012

Bruce R. Linter² Bruce.Linter@pepsico.com

Tim J. Foster¹ Tim.Foster@nottingham.ac.uk

¹ Division of Food Sciences, School of Biosciences, University of Nottingham, Sutton Bonington Campus, Loughborough, LE12 5RD, UK

² PepsiCo International Ltd, 4 Leycroft Rd, Leicester, LE4 1ET, UK

Highlights:

- The incorporation of (fibrillated) cellulose strengthened gluten free doughs
- The generalised Maxwell model was applied to analyse dough rheology
- The incorporation of (fibrillated)cellulose influenced the later stage of proofing
- Volume and water competition dominates the thermal rheological behaviours
- Loaves with (fibrillated)cellulose were smaller with harder, denser but finer crumb

Keywords:

Fibrillated cellulose, cellulose, gluten free bread, dough rheology, bread quality

Abstract

This study investigated starch reduction and replacement by purified cellulose (FC0) and fibrillated cellulose (FC60) which included a comprehensive investigation on dough properties, proofing behaviours, cooking performance, and bread qualities. Replacing flour with FC0 and FC60 was found to strength the doughs without, however, altering the extension of the structural network according to the weak gel model. The relaxation time calculated by the generalised Maxwell model was found to be shorter than the deformation rate during proofing which suggests that the doughs behave like fluids during proofing. The relaxation time was less influenced by the additions of FC0 and FC60. Although the initial stage of proofing was less influenced, the later stage was significantly affected by the additions of FC0 and FC60 which increased dough rigidity and restrained the volume growth. The pasting properties were significantly influenced by the competition for water and volume between FC0/FC60 and flour. The bread qualities were characterised in terms of loaf volume and crumb properties. Loaves containing FC0 and FC60 had smaller specific volume and harder crumb. However, the addition of FC0 and FC60 is beneficial to the generation of a finer crumb structure. Fibrillation process is detrimental to maximise the addition of fibres. However, a small amount of fibrillated cellulose is beneficial to workability and crumb structure.

1 **1. Introduction**

2 Wheat flour is the main ingredient of bread, the staple food in many regions. However, many
3 people around the world are intolerant or sensitive to gluten, which leads to coeliac disease and
4 other gluten-triggered health problems (Catassi et al., 2012; Czaja-Bulsa, 2015; Sapone et al.,
5 2012). In 2014, Aziz et al. (2014) conducted a survey in Sheffield, UK, and found that 13% of
6 the population reported gluten sensitivity and the prevalence of diagnosed Celiac disease was
7 0.8%. An early study reported that the prevalence of the coeliac disease in the world is
8 approximately 1 in every 100 people (Zandonadi, Botelho & Araujo, 2009). Celiac disease, the
9 most widely studied gluten-triggered disease, is an autoimmune genetically determined chronic
10 inflammatory intestinal disorder (Fasano & Catassi, 2001; Schuppan, 2000) which can be
11 influenced by both genetic (intrinsic) and environmental (extrinsic) factors (Di Sabatino &
12 Corazza, 2009; Schuppan, 2000; Wahab, Meijer, Goerres & Mulder, 2002). On the other hand,
13 ‘free from’ is becoming a global trend of a healthy lifestyle choice, which further promotes the
14 market of gluten free products. The production and improvement of gluten free bread have
15 been widely studied and the current methodologies can be categorised as the application of
16 alternative flour blends, enzyme treatments and sourdough applications, and other treatments.
17 The application of alternative flour blends is the most investigated which includes
18 investigations on formulations (Haque & Morris, 1994; Lazaridou, Duta, Papageorgiou, Belc
19 & Biliaderis, 2007; Nishita, Roberts, Bean & Kennedy, 1976; Ronda, Pérez-Quirce, Angioloni
20 & Collar, 2013) and structured gel and colloids (Van Riemsdijk, van der Goot, Hamer & Boom,
21 2011). Both dough rheological properties and bread qualities have been widely investigated. In
22 addition, the development of the dough microstructure and during baking, which can be
23 influential in the production of bread and other bakery goods, have also been studied (Babin et
24 al., 2006; Baldino, Laitano, Lupi, Curcio & Gabriele, 2018; Bousquière, Michon & Bonazzi,
25 2017; Wagner, Quellec, Trystram & Lucas, 2008).

26 In addition, the proportion of overweight adults and children has increased around the world
27 and obesity has become a global health challenge during the past three decades (Ng et al., 2014).
28 The prevalence of obesity around the world is one of the main reasons for the increased
29 morbidity rate of type-2 diabetes (Shaw, Sicree & Zimmet, 2010). However, the removal of
30 gluten from the diet easily lead to a significant decrease in carbohydrate oxidation rate, increase
31 in body fat stores, and weight gain in celiac patients (Capristo et al., 2000; Hager et al., 2012).

32 Therefore, food containing low calories and/or glycaemic index is recommended and the
33 production of gluten-free bread with low-calorie content and glycaemic index will be beneficial
34 to deal with gluten intolerance, obesity and type 2 diabetes simultaneously. The additions of
35 dietary fibres or fibre enriched materials, most by-product or cell wall materials, have been
36 studied to improve the nutritional profiles of gluten free bread. Demirkesen, Mert, Sumnu and
37 Sahin (2010) produced gluten free bread with desirable qualities in terms of loaves, texture,
38 colour and sensory perception by adding an intermediate amount of chestnut flour. Gluten free
39 bread with enhanced fibre contents was achieved by incorporating apple fibres and sugar beet
40 fibres with hydroxypropyl methylcellulose (HPMC) (Djordjevic et al., 2019). The addition of
41 rice bran in gluten free bread has been investigated by Phimolsiripol, Mukprasirt and
42 Schoenlechner (2012). The investigations on the addition of fibres were mainly based on the
43 materials with a defined processing procedure or commercial products but the influence of fibre
44 treatment, such as fibrillation, is less understood.

45 The fibres added in the bread formulation can be enriched in cellulose, hemicellulose, and/or
46 pectin. These materials, either soluble or insoluble, usually have high water absorbability which
47 significantly alters the rheological properties of doughs. The insoluble cellulose enriched
48 materials can also be expected to play a role as fillers.

49 Cellulose is one of the main components in the plant cell wall, which structures, with
50 hemicellulose, pectin, and lignin, to maintain the mechanical property of cell walls.
51 Microfibrillated cellulose (MFC) was first processed using high pressure and shearing by
52 Herrick, Casebier, Hamilton and Sandberg (1983) and Turbak, Snyder and Sandberg (1983a),
53 where the native cellulose fibres are physically unwound and highly entangled cellulose fibrils
54 are generated with high surface area, liquid retention, and reactivity to chemical treatments. In
55 food productions, MFC can be used as a thickener, compound carriers, and suspension &
56 emulsion stabilisers (Turbak, Snyder & Sandberg, 1982, 1983b). Currently, the applications of
57 high pressure homogeniser and microfluidiser as mechanical cellulose fibrillation treatments
58 are widely investigated to produce MFC (López-Rubio et al., 2007; Nakagaito & Yano, 2004;
59 Pääkkö et al., 2007; Stenstad, Andresen, Tanem & Stenius, 2008; Zimmermann, Pöhler &
60 Geiger, 2004).

61 In this study, a colloid mill was used which has lower energy input. Therefore, the cellulose
62 was fibrillated to a less extend comparing to the production of MFC. The water absorbability

63 and fibre configuration and rigidity were greatly altered by the fibrillation process. The aim of
64 this study was to investigate the starch/flour reduction of rice flour based gluten free bread by
65 the addition of cellulose and fibrillated cellulose. A high replacement level of up to 20% was
66 included in the investigation. Dough properties, proofing behaviours, cooking performance,
67 and bread qualities were of the interests to achieve a comprehensive understanding.

68 **2. Materials and methods**

69 **2.1. Materials**

70 Rice flour was purchased from Doves Farm online store. Allinson Easy Bake Yeast (Allinson
71 Flour, Peterborough, UK), sugar (Sainsbury's, UK), sunflower oil (Sainsbury's, UK), and salt
72 (Sainsbury's, UK) were purchased from local supermarkets. Methylcellulose (Methocel[®] A4M)
73 was supplied by The Dow Chemical Company (Bomlitz, Germany). Vitacel[®] Psyllium seed
74 husk powder was kindly donated by the JRS (J. Rettenmaier & Söhne Group, Rosenberg,
75 Germany). Pure amylose from potato and amylopectin from corn were purchased from Sigma-
76 Aldrich (Dorset, UK). Pure powdered cellulose, Solka floc 900FCC, was supplied by
77 International Fiber Corporation, US.

78 **2.2. Biochemical analysis**

79 Nitrogen content was measured using a Nitrogen Analyser NA 2000 (Fisons Scientific
80 Equipment, Loughborough, UK) and the protein contents of rice flour and psyllium seed husk
81 powder were converted with a factor of 6.25. Lipid was extracted with a chloroform-methanol
82 mixture (2:1) and the contents were calculated. Moisture contents were obtained by drying at
83 105 °C and ash contents were measured using a muffle furnace at 550°C.

84 AACC method (61-03) and the method from Kaufman, Wilson, Bean, Herald and Shi (2015)
85 were modified to determine the amylose content of rice flour. Rice flour was defatted using
86 85% methanol. Defatted rice flour, potato amylose and corn amylopectin (100 mg) were
87 dispersed in 1 ml of 95% ethanol and then dissolved in 9 ml of 1 M NaOH in a boiling water
88 bath for 10 min. After standing at room temperature for 3 h, the starch solutions were diluted
89 to 100 ml. The standard curve was obtained from mixtures of potato amylose and corn
90 amylopectin in a series of different ratios. Samples (5ml) were diluted by 20 times with
91 additions of 1 ml of 1M acetic acid and 2 ml of iodine solution (0.2% I₂ and 2% KI w/v) and

92 were incubated at room temperature for 20 min. The absorbance at 720 nm was determined
93 using a spectrophotometer. The amylose contents were calculated against the regression
94 equation determined from the standard curve and verified by a standard starch sample with 66%
95 amylose content.

96 **2.3. Cellulose fibrillation**

97 Five grams of cellulose was dispersed in 500 ml of reverse osmosis (RO) water and fibrillated
98 by a colloid mill (Winkworth, Basingstoke, UK) for 30 minutes (60 min L⁻¹) and labelled as
99 FC60. The fibrillated cellulose was then centrifuged at 4000 g, 4 °C, for 15 min and the
100 supernatants (residue less than 0.01%) were discarded. The concentration of sediments was
101 verified by drying at 105 °C for each batch and stored at 4 °C, which were then diluted to the
102 required concentrations for dough preparations.

103 Table 1. Addition levels of rice flour, FC0, and FC60.

Sample code	Rice flour	FC0	FC60
F(100) (control)	100	0	0
F(98)+FC60(2)	98	0	2
F(90)+FC0(10)	90	10	0
F(90)+FC0(8)+FC60(2)	90	8	2
F(80)+FC0(20)	80	20	0
F(80)+FC0(18)+FC60(2)	80	18	2
F(80)+FC0(16)+FC60(4)	80	16	4

104

105 **2.4. Dough formulation and preparation**

106 The basic formulation (control) was decided according to preliminary tests and included 100g
107 of rice flour, 5g of sugar, 2g of salt, 1.5g of yeast, 5g of vegetable oil, 1g of methylcellulose
108 (MC), 1g of psyllium, and 120g of water. Rice flour was partially replaced with pure cellulose
109 powder (FC0) and/or fibrillated cellulose (FC60) as shown in Table 1. Apart from F(100)
110 (control) and F(98)+FC60(2), all other FC0/FC60 incorporated formulas are expected to be
111 claimed 'high fibre' according to Regulation (EC) No 1924/2006 based on a rough calculation
112 that the fibre content is higher than 3 g per 100 kcal. FC0 was directed used to replace rice
113 flour and mixed with other dry ingredients. FC60 was redispersed in water required in the
114 formulation and added with oil into dry ingredients. Doughs were mixed based on 200g of rice
115 flour or the mixture of flour and cellulose per batch by a stand mixer (Kenwood, UK) equipped

116 with a CHEF flexible beater (AT501, Kenwood, Havant, UK). Dry ingredients were mixed
117 thoroughly and then mixed with water/FC60 dispersion and oil for 7 min at speed 1. Doughs
118 prepared for rheological measurements did not contain yeast and were allowed for hydration
119 at room temperature for 1 hour.

120 **2.5. Dough evaluation**

121 **2.5.1. Fundamental rheological measurements and thermal behaviour of doughs**

122 Doughs without yeast, which were hydrated for 1 hour at room temperature, were subjected to
123 shear stress ramp tests and small amplitude oscillatory shear tests by MRC 301 rheometer
124 (Anton Paar, Austria) equipped with serrated parallel plate geometry (PP25/P2-SN15766,
125 Anton Paar). The measuring gap was 2 mm. The temperature was controlled by a Peltier system
126 with the assistance of a water bath (R1, Grant, Shepreth, UK). The extra sample was trimmed
127 by a spatula during sample loading and the edge was covered by low viscosity mineral oil
128 (Sigma, USA) to prevent drying. Doughs rested at 30 °C for 500 seconds before measurements.
129 Logarithmic increase of shear stress from 0.03 to 30000 Pa in 18.5 min was performed to obtain
130 the yield point or yield zone. Frequency sweep tests were performed in a logarithmic decrease
131 from 600 to 0.06 rad s⁻¹ with 0.02% strain (in the linear viscoelastic region). The slopes of lgG'
132 versus lgω in the middle frequency range (0.881 to 40.9 s⁻¹) were calculated. The dynamic
133 oscillatory measurement data were also fitted into the 'weak gel' model (equation 1) proposed
134 by Gabriele, de Cindio and D'Antona (2001) where z represents the number of the interacting
135 rheological units of 'weak gel' structure and A refers to the interaction strength between the
136 rheological units. G*(ω) is the complex modulus.

$$137 \quad G^*(\omega) = \pi A \omega^{\frac{1}{z}} \quad (1)$$

138 Additionally, the obtained mechanical spectra were fitted to generalised Maxwell model as
139 shown in equation (2) with individual relaxation time (λ_i) and individual relaxation moduli (G_i)
140 describing the discrete relaxation time spectrum of the sample where $G(t)$ is the relaxation
141 modulus at time t.

$$142 \quad G(t) = \sum_{i=1}^N G_i e^{-t/\lambda_i} \quad (2)$$

143 The relaxation times of each component (λ_i) were arbitrarily decided as shown in Table 2. The
 144 dynamic moduli $G'(\omega)$ and $G''(\omega)$ were calculated from angular frequency (ω) by equation (3)
 145 and (4) (Baumgaertel & Winter, 1992; Ferry, 1980; Laun, 1986). Individual relaxation moduli
 146 G_i , which is listed in Table 2, was varied to minimise the sum of square differences between
 147 calculated $G'(\omega)$, $G''(\omega)$ and experimentally obtained G' , G'' . G_e represents a pure elastic
 148 component connected in parallel with other Maxwell elements in the model.

$$149 \quad G'(\omega) = \sum_i G_i \frac{\omega^2 \lambda_i^2}{1 + \omega^2 \lambda_i^2} + G_e \quad (3)$$

$$150 \quad G''(\omega) = \sum_i G_i \frac{\omega \lambda_i}{1 + \omega^2 \lambda_i^2} \quad (4)$$

151 Temperature sweep tests were performed at a constant strain (0.02%) and angular frequency
 152 (10 rad s⁻¹) with the temperature increased from 20 °C to 98 °C with a heating rate of 2.6 °C
 153 min⁻¹ mimicking the temperature profile during baking.

154 Table 2. Individual relaxation time and relaxation moduli for Generalised Maxwell Model fitting on the
 155 mechanical spectra shown in Figure 2.

individual relaxation time λ_i (s)	0.0001	0.001	0.01	0.1	1	10	100	1000	10000	100000	R ²
F(100) (control)	123755	0	1201	1146	724	1934	1237	0	0.010	0.001	0.991
F(98)+FC60(2)	0	14804	6066	3950	3113	1925	8856	0	0.012	0.001	0.996
F(90)+FC0(10)	255156	0	3575	3270	2264	6034	3748	0	0.011	0.001	0.989
F(90)+FC0(8)+ FC60(2)	279077	0	5451	6761	3917	11764	6455	0	0.013	0.002	0.980
F(80)+FC0(20)	354470	18597	23156	18045	12357	22615	25966	0	0.010	0.002	0.998
F(80)+FC0(18) +FC60(2)	904692	0	31077	29132	18275	37525	42383	0	0.010	0.000	0.994
F(80)+FC0(16) +FC60(4)	1271	88716	36796	29398	18524	32671	47364	0	0.010	0.017	0.998

156 2.5.2. Empirical rheological measurements

157 Doughs were also characterised on a TA-Xt plus texture analyser (Stable Micro systems,
 158 Surrey, UK) equipped with a 5 kg loading cell. Backward extrusion tests and SMS/Chen-
 159 Hosney Dough Stickiness tests (Stable Micro Systems, UK) were performed. For the

160 backward extrusion tests, doughs were loaded into a container with a diameter of 42 mm to a
161 height of 36 mm avoiding big air pockets. A disc with a diameter of 30 mm and thickness of 5
162 mm was centred and extruded into the dough by 22.5 mm and then returned to the height of
163 120 mm from the base at a speed of 1 mm s⁻¹. The positive peak force and positive area during
164 downward stroke indicated firmness and consistency of doughs respectively. The negative peak
165 force and area during upward stroke indicated cohesiveness and index of viscosity. The
166 returning distance before the force reached a constant negative value indicated dough
167 extensibility. As for the SMS/Chen-Hoseney Dough Stickiness tests (Chen & Hoseney, 1995),
168 doughs were loaded into the equipped cell (A/DSC) and a uniform surface was generated. The
169 surface was compressed by an aluminium cylinder probe (P/25) by 40 g for 0.1 s. The probe
170 compressed the sample at the speed of 0.5 mm s⁻¹ and withdrew at 10 mm s⁻¹. The maximum
171 force, area, and travel distance during probe withdrawing indicated dough stickiness, work of
172 adhesion, and strength individually. Data recording and analysis were performed by the
173 equipped software Exponent 6.1.14.

174 **2.6. Pasting properties of flour blends**

175 The pasting profiles of flour blends were measured by Rapid Visco Analyser (RVA) (Newport
176 Scientific Pty. Ltd., Warriewood, New South Wales, Australia) equipped with a water bath
177 (Thermo scientific C10, Karlsruhe, Germany). The solid levels of rice flour, MC, psyllium,
178 FC0, and FC60 are listed in Table 3, which were in the same ratio as in the dough formulation
179 without sugar, salt, yeast, and oil. More specifically, flour, MC, psyllium and FC0 were mixed
180 thoroughly. Flour blends (2.55 g) prepared according to the formula which does not include
181 FC60 were dispersed in 24 ml of RO water. As for FC60-containing formula, FC60 stock
182 suspensions were diluted to 24.05 g or 24.1 g with FC60 concentration of 0.208% and 0.415%
183 respectively for the formulations of 2% and 4% replacement by FC60. Flour blends (2.5 or
184 2.45 g respectively) were dispersed into diluted FC60 suspensions. The test started with high
185 shear rate mixing (960 rpm) for 60 seconds at 25 °C followed by 60 seconds mixing at 160
186 rpm. The temperature then increased to 95 °C in 350 s, held at 95 °C for 150 s, and decreased
187 to 25 °C in 350 s. The pasting profiles were analysed by ThermoLine where pasting
188 temperature (the temperature of the onset of viscosity increase), peak (the highest viscosity),
189 peak time (the time when the highest viscosity was reached), trough (lowest viscosity after the
190 peak), trough time (the time when the trough occurred), final viscosity, breakdown (difference

191 between peak and trough), and setback (difference between final viscosity and trough viscosity)
192 were reported.

193 Table 3. Addition levels of rice flour, MC, psyllium, FC0, and FC60 in pasting property analysis.

Sample code	Rice flour	MC	psyllium	FC0	total dry blends	FC60
F(100) (control)	2.5	0.025	0.025	0	2.55	0
F(98)+FC60(2)	2.45	0.025	0.025	0	2.5	0.05
F(90)+FC0(10)	2.25	0.025	0.025	0.25	2.55	0
F(90)+FC0(8)+FC60(2)	2.25	0.025	0.025	0.2	2.5	0.05
F(80)+FC0(20)	2	0.025	0.025	0.5	2.55	0
F(80)+FC0(18)+FC60(2)	2	0.025	0.025	0.45	2.5	0.05
F(80)+FC0(16)+FC60(4)	2	0.025	0.025	0.4	2.45	0.1

194

195 2.7. Baking tests

196 Doughs prepared according to section 2.3 were loaded in a baking pan with a dimension of 7.5
197 x 7.5 x 10 cm (W×L×H). Each batch generated two doughs with weights of 200 to 210 g.
198 Baking pan was shaken to expel big air pockets in doughs. The doughs were covered by cling
199 film and proofed in an incubator (Binder, Tuttlingen, Germany) at 30 °C for 85 min and then
200 baked in a deck oven (Tom Chandley, Manchester, UK) for 40 min at 230 °C. The loaves were
201 cooled on a rack at the atmosphere environment for 1 hour before further evaluation.

202 2.7.1. Basic analysis and calculations

203 Proofing behaviour was monitored by incubating approximately 10 ml of doughs in a measuring
204 cylinder at 30 °C for 85 min during which the volume was recorded. Doughs were weighted
205 before baking and the loaves were weighted after baking and cooling. Loaf volumes were
206 measured by rapeseed replacement. The baking loss was calculated by *baking loss =*
207 *(dough weight before proving – loaf weight after cooling)/dough weight before proving.*
208 Specific volume was the ratio of loaf volume to loaf weight. The moisture content of the loaf centre
209 was measured by drying a piece of crumb cut from the centre at 105 °C.

210 The loaves were sliced perpendicularly to a thickness of 1.25 cm and the middle surface was
211 scanned using a C-Cell imaging system (Calibre Control International LTD, Warrington, UK).
212 The obtained imaged were analysed using C-Cell software version 2.0.

213 2.7.2. Bread textural evaluation

214 The textural property of gluten free bread was evaluated by texture profile analysis (TPA) on
215 a TA-XT plus texture analyser (Stable Micro systems, Surrey, UK) equipped with 30 kg
216 loading cell. A cylinder piece of bread crumb was cut from the centre of every loaf slice by a
217 diameter of 3 cm and two pieces from the middle four slices of each loaf were stacked together.
218 Therefore, two pieces of the sample were obtained from the middle of each loaf with the shape
219 of a cylinder with a height and a diameter of 2.5 cm and 3 cm individually. The samples were
220 65% compressed by a 100 mm plate twice at a speed of 1 mm s⁻¹ with 5 s between two
221 compresses. Hardness, springiness, cohesiveness, chewiness, and resilience were obtained
222 from the TPA profiles.

223 **2.8. Statistical analysis**

224 All measurements were repeated at least three times. Plots are shown as representative curves.
225 Results are shown as the mean ± standard deviation in bar charts and tables. The C-Cell
226 parameters were analysed and compared using one-way analysis of variance (ANOVA) and
227 Turkey's test at a significance level of $p < 0.05$ using IBM SPSS statistics version 26 (IBM
228 Corp., Armonk, NY, USA).

229 **3. Results and discussion**

230 **3.1. Biochemical properties**

231 The moisture content, protein content, ash content, and lipid content of rice flour were 11.1 %,
232 7.23 %, 0.42 %, and 2.8 % respectively. That of PSY was 7.23 %, 3.40 %, 2.89 %, and 3.30 %
233 respectively (Ren, Yakubov, Linter, MacNaughtan & Foster, Unpublished results-b). The
234 amylose content of rice flour was 28.8 %.

235 **3.2. Dough rheological properties**

236 **3.2.1. Fundamental rheological analysis of dough properties**

237 The stress ramp data are shown in Figure 1. It can be seen (Figure 1a) that the shear strain
238 increased with shear stress with different rates in different stress ranges. The replacement of
239 rice flour by FC0 and FC60 significantly decreases the shear strain caused by a certain applied
240 shear stress in the higher stress range (>10 Pa). The control dough (F100) shows a relatively
241 short transition from the linear-elastic deformation behaviour (shear strain increases linearly
242 with shear stress) to flow behaviour and a yield point can be defined by calculating the

243 deviation from the linearity at low shear stress range. However, when FC0 or FC60 is added,
244 it shows a long yield zone and yield point cannot be defined by this method. The longer yield
245 zone could be due to the long cellulose fibres with various sizes. The viscosity from the same data
246 sets is plotted versus shear stress in Figure 1b. The viscosity plateau at low shear stress which
247 is similar to the Newtonian plateau is, however, possibly due to the fact that the doughs did not
248 reach a steady state. However, the start of viscosity decrease also indicates yield stress (Walls,
249 Caines, Sanchez & Khan, 2003). Although it is still difficult to define the yield point due to a
250 long yield-flow transition, it can be seen that the replacement by FC0 and FC60 increased the
251 stress at which doughs start to flow (significant decrease in viscosity). Similar enhancing
252 effects on viscosity and yield stress by the addition of chestnut flour, which has high fibre
253 content, in gluten free rice bread has been reported by Demirkesen et al. (2010) which can be
254 assigned to the high entanglement and water binding ability of fibres.

255 The mechanical spectra of doughs are shown in Figure 2. In the mechanical spectra, it can be
256 seen that all doughs show solid like property with G' higher than G'' . The additions of FC0 or
257 FC60 increased both moduli and decreased $\tan\delta$ suggesting a more solid like property, which
258 is in accordance with the strengthening effects of doughs evidenced by yield stress and
259 viscosity as discussed above.

260 The similarities in the general structure and rheological behaviours of different foods has been
261 concluded and a weak gel model has been proposed by Gabriele et al. (2001). Foods can be
262 described to be structured by strands constituted by weakly interacting flow units, and strongly
263 interacting topological points, which is analogous to the classic 'true gel' with permanent cross-
264 linked three dimensional network (Gabriele et al., 2001). The complex moduli (G^*) of gluten
265 free doughs and angular frequencies (ω) were fitted with the weak gel model (equation (1))
266 with R^2 higher than 0.95 and two model parameters, A and z , were obtained and are listed in
267 Table 4. It can be seen that A increased significantly with the addition of FC0 and FC60 which
268 is in accordance with the increase of moduli. However, there is no significant difference in the
269 value of z between all flour-replaced doughs, which suggests that there are no significant
270 effects on the extension of the network of strands and rheological units. The increase in both
271 moduli and parameter A and the decrease of $\tan\delta$ are more likely to be attributed to the increase
272 in the strength of the topologically interactions and strand (rheological units) upon the additions
273 of FC0 or FC60 fibres. In other words, it can be speculated that both flour particles and FC

274 fibres are rigid fillers regardless of the difference in shapes. FC fibres are more rigid than the
 275 hydrated flour particles and they compete for water with starch which further increases the
 276 rigidity of the flour particles. Therefore, the doughs are strengthened by the addition of FC0 or
 277 FC60.

278 The slopes of $\lg G'$ versus $\lg \omega$ have a similar value of approximately 0.105 in all cases
 279 regardless of the addition of FC0 or FC60, which is in accordance with the uninfluenced z
 280 values. However, it is lower compared with most gluten free and wheat doughs which range
 281 from 0.11 to 0.37 (Georgopoulos, Larsson & Eliasson, 2004; Ronda et al., 2013; Tanner, Qi &
 282 Dai, 2008; Upadhyay, Ghosal & Mehra, 2012; Villanueva, Harasym, Muñoz & Ronda, 2019).
 283 The low $\lg G'$ versus $\lg \omega$ slope suggests approaching a true gel at the angular frequency range
 284 studied. It was less influenced by FC0 and FC60 at the addition levels studied.

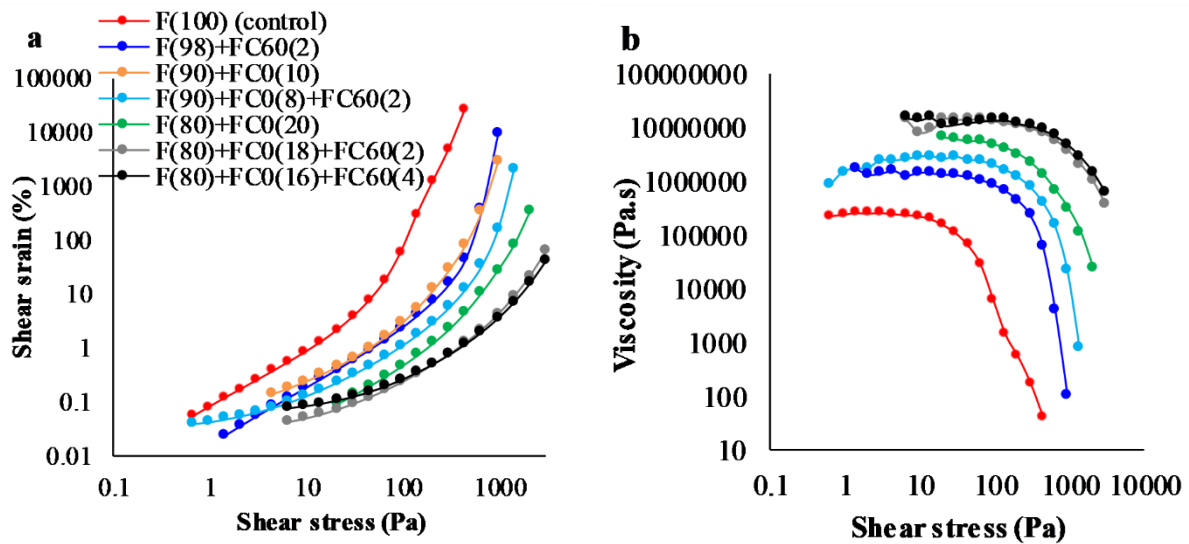
285 FC60 has more pronounced influence than FC0 even with very low addition levels (2 or 4%
 286 replacement). In fact, the effect of 2% replacement by FC60 is similar to 10% replacement by
 287 FC0 shown as similar shear strains and viscosities plotted versus shear stress, and similar
 288 mechanical spectra and parameter A in the weak gel model. This could be explained by the
 289 higher water holding ability of FC60 due to fibrillation treatment (Agarwal, Hewson & Foster,
 290 2018a; Ren, Linter & Foster, 2020). Additionally, an increase in fibre entanglement can also
 291 be expected as an outcome of fibrillation and FC60 appears as flocculates or aggregates (Ren
 292 et al., 2020). It is likely that FC60 exists as highly entangled fibre aggregates which can be
 293 considered as individual rheological units with enhanced properties. However, considering that
 294 there was no significant difference in the value of the weak gel model parameter z between
 295 FC0 and FC60 doughs, the interaction and entanglement between FC60 aggregates and FC60
 296 aggregates and flour particles are limited therefore the influence on the extension of the
 297 structural network of doughs are limited.

298 Table 4. Weak gel model parameters.

	A (Pa.s ^{1/z})	z
F(100) (control)	3216±538 ^a	8.90±1.12 ^a
F(98)+FC60(2)	10915±2055 ^a	9.06±1.04 ^a
F(90)+FC0(10)	11486±493 ^a	10.47±0.10 ^a
F(90)+FC0(8)+FC60(2)	21971±1733 ^{ab}	10.85±0.46 ^a
F(80)+FC0(20)	52447±4287 ^{bc}	9.34±0.35 ^a
F(80)+FC0(18)+FC60(2)	82416±11993 ^c	9.69±0.34 ^a
F(80)+FC0(16)+FC60(4)	80984±18218 ^c	9.08±0.27 ^a

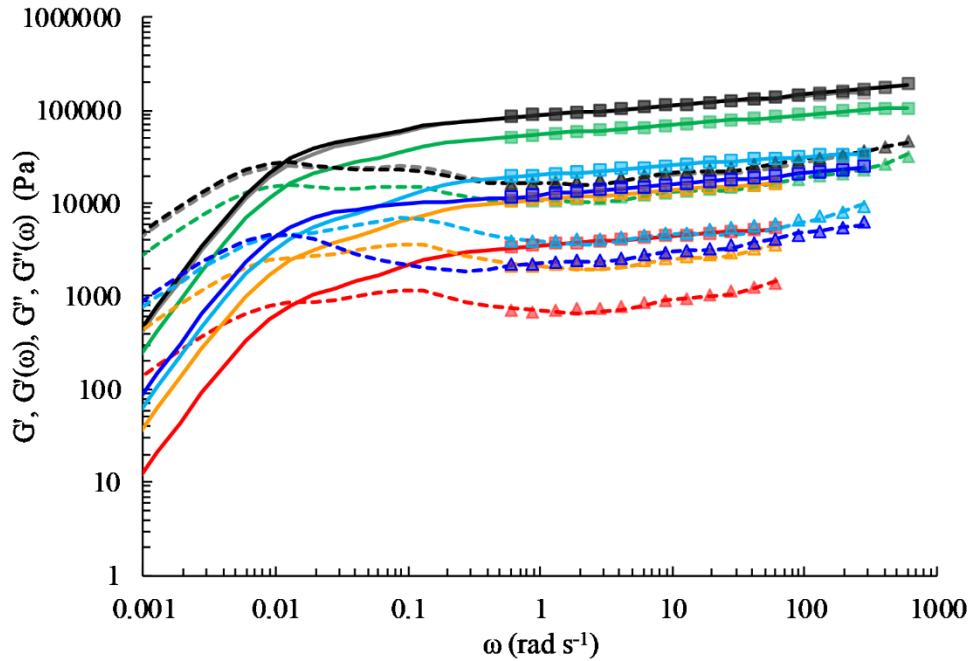
Data are shown as mean \pm standard deviation. Values with different letters in the same column were significantly different ($p < 0.05$).

299 The generalised Maxwell model was applied to estimate the dough rheological behaviours at a
300 longer time scale (slow deformation) (Figure 2). The arbitrarily decided λ_i and calculated
301 corresponding relaxation moduli G_i are listed in Table 2. Ten pairs of λ_i and G_i along with a G_e
302 representing a single pure elastic component were used to calculate $G'(\omega)$ and $G''(\omega)$ by
303 equation (3) and (4). The value of G_e of each sample is either 0 or very low therefore it is not
304 listed in Table 2. A zero value of G_e is typical for viscoelastic liquid of uncross-linked polymers
305 (Ferry, 1980). Therefore, gluten free bread doughs are structurally and rheologically analogous
306 to viscoelastic fluids instead of a solid.



307

308 Figure 1. Shear stress ramp data (a) and viscosities plotted versus shear stress (b) of gluten free doughs.
309 Plots are shown as representative curves from experiments run in triplicates.
310



311

312 Figure 2. Mechanical spectra and curves calculated from generalised Maxwell model of F(100) (control)
 313 (red), F(98)+FC60(2) (dark blue), F(90)+FC0(10) (yellow), F(90)+FC0(8)+FC60(2) (light blue),
 314 F(80)+FC0(20) (green), F(80)+FC0(18)+FC60(2) (grey), and F(80)+FC0(16)+FC60(4) (black).
 315 Experimental storage moduli (G') and loss moduli (G'') are shown by square and triangle symbols
 316 respectively and calculated $G'(\omega)$ and $G''(\omega)$ are presented by solid lines and dashed lines respectively.
 317 Plots are shown as representative curves from experiments run in triplicates.

318 The generalised Maxwell model expands the evaluable mechanical spectra to a lower
 319 frequency range with R^2 higher than 0.98 (Table 2). It can be seen in Figure 2 that all doughs
 320 have a relaxation frequency of nearly 0.01 rad s^{-1} . The influences of FC0 and FC60 additions
 321 are insignificant which is in accordance with the uninfluenced z value in the weak gel model
 322 that FC0 and FC60 do not significantly alter the extension of the rheological network. The
 323 porous structure of loaves mainly forms, develops, and sometimes collapses during proofing
 324 with low strain rates (10^{-4} to 10^{-3} s^{-1} as reported by Babin et al. (2006) and Grenier, Lucas and
 325 Le Ray (2010)). The time scale is longer than the relaxation time (reciprocal of relaxation
 326 frequency) of gluten free. In other words, doughs behave like fluids ($G'' > G'$) in this low strain
 327 rate range as found during proofing, which could be beneficial to the development of the loaf
 328 structure. However, this also implies that only strengthening doughs might not be able to
 329 provide efficient stability to maintain the porous structure. This could explain from, one aspect,
 330 why air pockets and big voids in crumb structure is a common and difficult issue in the gluten
 331 free bread production which have been widely reported (Haque & Morris, 1994; McCarthy,

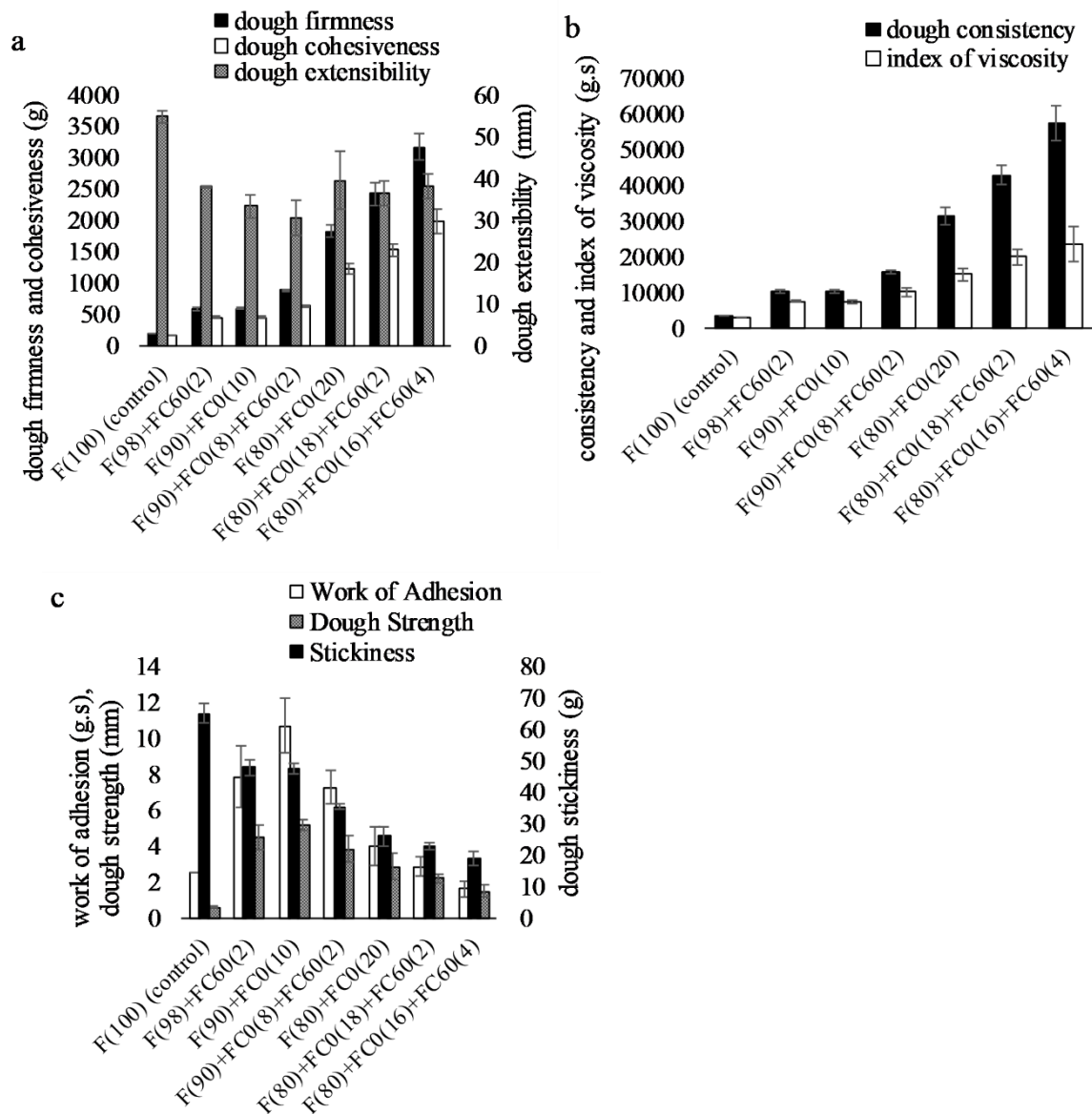
332 Gallagher, Gormley, Schober & Arendt, 2005; Nishita et al., 1976; Paciulli, Rinaldi, Cirlini,
333 Scazzina & Chiavaro, 2016; Schober, Bean & Boyle, 2007).

334 **3.2.2. Empirical rheological analysis of dough properties**

335 The influence of FC0 and FC60 addition on the behaviour of doughs under large deformation
336 were analysed by backward extrusion experiments. Dough stickiness was measured using an
337 SMS/Chen-Hoseney Dough Stickiness rig. As shown in Figure 3, the addition of FC0 and FC60
338 significantly increased dough firmness, cohesiveness, consistency, and index of viscosity,
339 which is in agreement with the increasing effects on yield stress and dynamic moduli. Similar
340 to what was observed in the fundamental rheological analysis, 2% replacement by FC60 and
341 10% replacement by FC0 show similar values for these four parameters which indicate that
342 fibrillated cellulose is more efficient in strengthening doughs than untreated cellulose. The
343 additions of FC0 and FC60 generally decreased dough extensibility by approximately 20 mm
344 while the effects of FC0 fibres tend to be quadratic as 20% replacement doughs have higher
345 extensibility than 10% replacement doughs by 6 mm. Extensibility describes the flowability of
346 doughs in extensional flow. The negative effects of FC0 and FC60 are likely to be caused by
347 their strengthening effect which leads to higher resistance to flow. However, the slight increase
348 in extensibility by FC0 at higher addition level might be because of its fibrous structure where the
349 fibres are much longer (intact) than FC60 (fibrillated).

350 The addition of both FC0 and FC60 increased dough strength and work of adhesion while
351 further increasing the addition levels decreased these two parameters. The stickiness was
352 decreased by the addition of both FC0 and FC60. The reduction of dough stickiness is in
353 agreement with the additions of fibres in wheat doughs (Collar, Santos & Rosell, 2007;
354 Sangnark & Noomhorm, 2004). Minimised stickiness is a desirable dough textural property in
355 the manufacture of wheat bread (Collar et al., 2007), where it is also beneficial to the handling
356 property. Generally, stickiness of a material is affected by both adhesive force and cohesive
357 (rheological) force which could oppose each other (Hoseney & Smewing, 1999). The
358 SMS/Chen-Hoseney Dough Stickiness method minimises the interference from the bulk
359 rheology (Chen & Hoseney, 1995). Therefore, it can be expected that the bulk rheology
360 property have less influence on dough stickiness measured in this experiment. Additionally,
361 the adhesive force is influenced by water surface tension (Hoseney & Smewing, 1999). The
362 influence of water absorption on the stickiness of wheat doughs has also been highlighted

363 (Armero & Collar, 1997; Heddleson, Hamann, Lineback & Slade, 1994). The influence on
364 dough stickiness is contrary to the influences on dough firmness and cohesiveness. In fact, they
365 show close negative correlations when fitted by power equations with R^2 of 0.97 and 0.94
366 respectively. Therefore, the high water absorbability of FC0 and FC60 contribute both to the
367 strengthening of doughs, in addition to the contribution of the fibrous structure, and reduction
368 of stickiness. However, dough strength and work of adhesion reflect both the adhesive force
369 (stickiness measured in this experiment) and cohesive force (bulk rheological, dough
370 strengthening effect). Therefore, they increased with the replacements by FC0 and FC60 at
371 lower addition levels but decreased coincidentally with stickiness with further additions. A lower
372 stickiness and work of adhesion suggests better workability and handling properties of the
373 doughs.



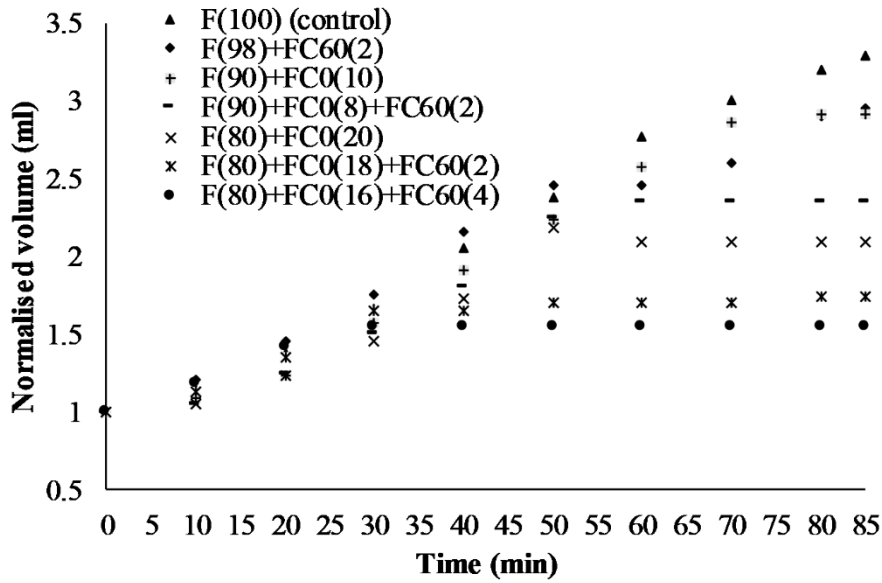
374

375 Figure 3. Effects of cellulose and FC fibres additions on dough firmness, Cohesiveness, extensibility
 376 (a), consistency, and index of viscosity (b) in backward extrusion measurements, and work of adhesion,
 377 dough strength, and dough stickiness (c) measured by SMS/Chen-Hoseney Dough Stickiness rig. Error
 378 bars represent the standard deviations averaging at least five replicates.

379 3.3. Proofing behaviours

380 The proofing behaviour of doughs are monitored by recording the volume increase of a piece
 381 of dough sample in a measuring cylinder and the proofing profiles are shown in Figure 4. The
 382 differences in volume between gluten free doughs are less significant during the first 30
 383 minutes. However, with increasing addition of FC0 or FC60, volume growth stopped earlier
 384 during proofing which leads to lower final volume. Considering the correlations between low

385 molecular weight sugars, yeast activity, and porosity kinetics (Romano, Toraldo, Cavella &
386 Masi, 2007; Sahlström, Park & Shelton, 2004), sucrose in the formula might promote the
387 volume increase during the early stage of proofing but be less influential after being consumed
388 by yeasts during the later stage. The reduction of flour, instead, limits the further fermentation
389 by yeast during the later stage of proofing. However, the difference between F(100) (control),
390 F(90)+FC0(10), and F(98)+FC60(2) are less pronounced although the flour was replaced by
391 only 2% and 10% respectively. Hence, the answer could also lie in rheological properties. As
392 shown by the generalised Maxwell model, doughs behave like fluids during proofing and
393 dough strengthening by FC0 and FC60 is less influential on the relaxation time. In addition,
394 according to the model derived by Shah, Campbell, McKee and Rielly (1998), dough rheology
395 is less influential during the early stage of proofing while it becomes critical during the later
396 stage (Mills, Wilde, Salt & Skeggs, 2003). A rigid dough is highly resistant to deformation
397 which limits the expansion of gas cells during proofing (Lazaridou et al., 2007; Van Vliet,
398 Janssen, Bloksma & Walstra, 1992). Therefore, the influences of FC0 and FC60 on proofing
399 behaviour during different stages can be also assigned to their strengthening effects on dough
400 rheology. There is no difference between maximum volume increase and final volume increase
401 indicating that doughs did not collapse during the proofing process even for
402 F(80)+FC0(16)+FC60(4), whose volume did not change after 40 min of proofing. This
403 stabilising effect is attributed to the dough strengthening by FC0 and FC60 additions as seen by
404 the rheological measurements.



405

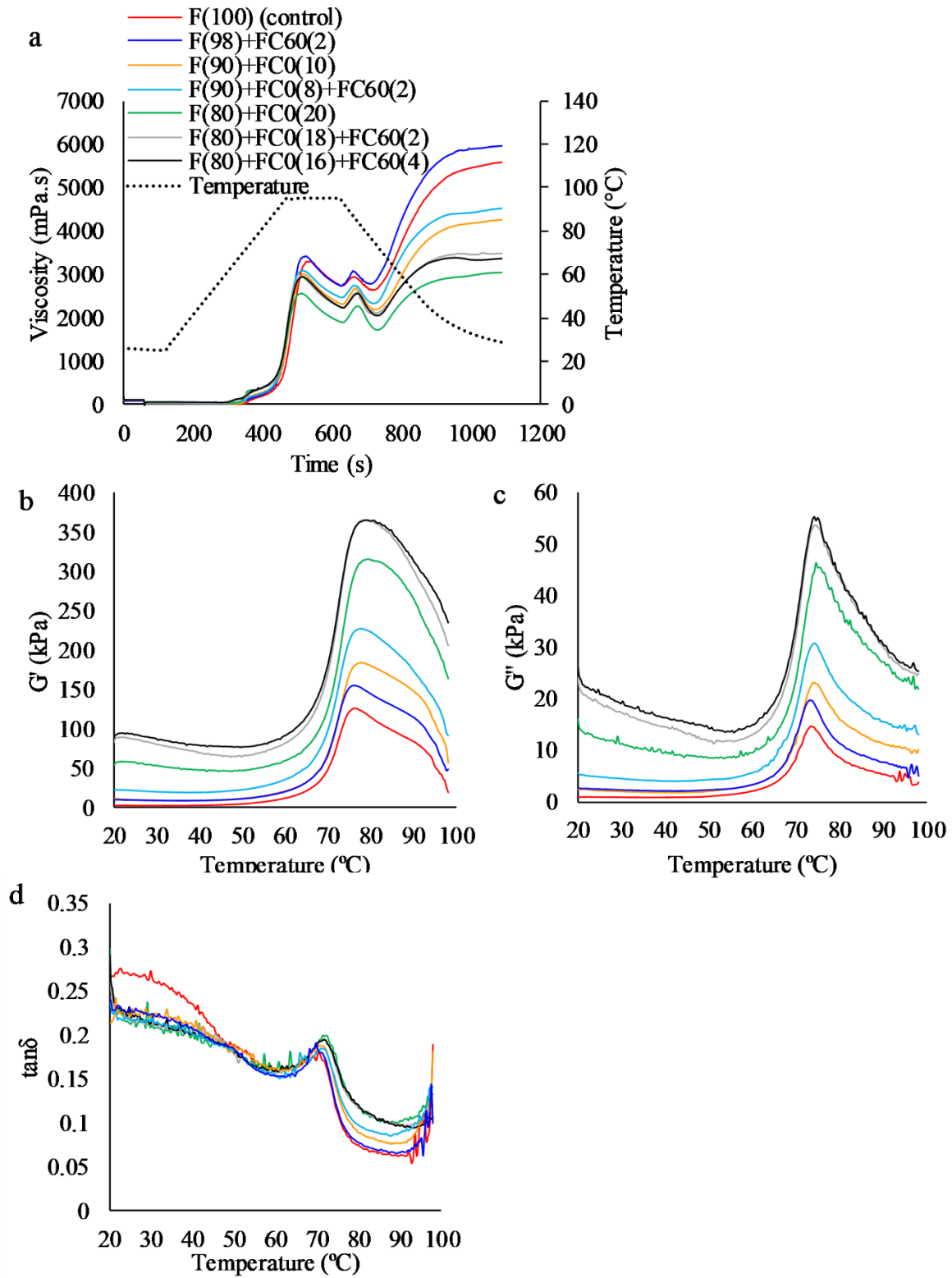
406 Figure 4. Proofing profiles (normalised) of gluten free doughs. Plots are shown as representative profiles
 407 from experiments run in triplicates.

408 3.4. Rheological properties during heating and cooling

409 Pasting profiles describe viscosity changes of flour blends during heating and cooling which
 410 typically describe the gelatinisation and retrogradation. The profiles are shown in Figure 5a. A
 411 secondary peak at 85 °C during cooling is observed at all curves due to the addition of psyllium,
 412 and possible to a balance between interaction with amylose (James M. Cowley, personal
 413 communication), formation of weak PSY gel particles, PSY particle interactions, and PSY
 414 particle breakdown (Ren, Linter & Foster, Unpublished results-a). The addition of FC0 and
 415 FC60 both significantly decreased the pasting temperature (the onset of viscosity increase)
 416 shown as a small shoulder before the main viscosity peak. The reduction of pasting temperature
 417 is widely observed for most starch/hydrocolloid mixtures (BeMiller, 2011;
 418 Naruenartwongsakul, Chinnan, Bhumiratana & Yoovidhya, 2004; Sullo & Foster, 2010). It is
 419 also observed for starch and bacterial cellulose mixtures (Díaz-Calderón et al., 2018). In the
 420 mixture of MC and starch, the thermal gelation of MC, which occurs at a lower temperature
 421 than the significant swelling of starch granules, leads to an increase in starch concentration and,
 422 therefore, enhanced starch granule interactions and increase in viscosity (Sullo & Foster, 2010).
 423 Naruenartwongsakul et al. (2004) suggested that the concentration increase of starch is due to
 424 water competition with MC. Therefore, water and volume competition might also explain the
 425 reduction of pasting temperature of FC0(FC60)/flour mixtures. When 20% of flour was

426 replaced by FC0, the further addition of FC60 lead to another smaller shoulder at a lower
427 temperature, which suggests that FC60 is more powerful than FC0 in the competition with
428 starch for volume and water due to its higher abilities to hold water and to occupy volume
429 (Agarwal et al., 2018a; Ren et al., 2020).

430 Replacement by FC0 decreased the overall viscosity, peak time and setback while it increased
431 breakdown. The decrease of overall viscosity, including peak, and setback is, due to the
432 decrease in starch concentration, as it is replaced by FC0 which does not swell as starch
433 granules nor retrograde as amylose. The reduction of swellable starch granules leads to a
434 reduction of the ability to be closely packed which appears as early onset of breakdown. The
435 increased breakdown is likely to be due to the enhancement of shear force exerted on starch
436 granules by the fibrous structure of FC0. However, the substitution by FC60 result in an
437 increase in the overall viscosity and decrease in breakdown. Fibrillated cellulose (FC60)
438 appears as flocculates or aggregates (Agarwal et al., 2018b; Ren et al., 2020) which can be
439 considered similar to swollen starch granules or granule fragments with freed fibrils similar to
440 leaked amylose. The increase of viscosity indicates that FC60 is similar to or even more
441 efficient than starch in increasing viscosity while it does not breakdown as starch granules.
442 Therefore, the functionality of fibrillated cellulose has an effective enhancement of overall
443 composite properties beyond the effects of the 'inert' unfibrillated filler.



444

445 Figure 5. Pasting profiles (a), storage moduli G' (b), loss moduli G'' (c), and loss factor $\tan\delta$ of flours
 446 blends. G' and G'' were recorded with a heating rate of $2.6\text{ }^{\circ}\text{C min}^{-1}$.

447 The rheological property of doughs during cooking was monitored by temperature sweep tests
448 (Figure 5b, c, and d) with a temperature profile mimicking the temperature changes during
449 baking. It can be seen that G' and G'' decreased slightly with the temperature increasing up to
450 approximately 50 °C, especially with 20% replacement of flour by FC0 and FC60, which could
451 be attributed to the softening induced by the increase of temperature. The initial G' and G''
452 decrease was accompanied by a moderate decrease in $\tan\delta$ until approximately 60 °C, which
453 can be attributed to slow swelling of starch granules which was not influenced by the
454 replacement by FC0 and FC60. It can be speculated that the slow granule swelling overcomes
455 the softening effect due to heating, therefore the temperature-induced softening (G' and G''
456 decrease) became more pronounced when flour was replaced by FC0 and FC60. With further
457 increase in temperature, G' and G'' significantly increased to a peak because of a dramatic
458 swelling and volume filling of starch granules which are eventually close-packed at the peaks
459 in moduli. In contrast to pasting temperature, the onsets of G' and G'' increase shifted to a
460 higher temperature when FC0 and/or FC60 were added. FC0/FC60 and flour were already
461 closely packed with the higher concentration in doughs compared to the experimental condition
462 of RVA. When the starch content was reduced, the starch granules needed to swell to a larger
463 volume at a higher temperature to overcome the initial temperature-controlled softening
464 (moduli decrease) and to contribute to the overall rigidity (G' increase). It was also different
465 from pasting properties that both the addition of FC0 or FC60 increased the peaks of G' and
466 G'' . This could also be due to the closely packed structure where the swelling and rigidity
467 changes of starch granules can be detected and, as demonstrated by the rheological properties
468 in both fundamental and empirical experiments, the fibrous structure and high water binding
469 ability of FC0 and FC60. FC0 and FC60 competed for water with starch, which restrained the
470 swelling of the granules, increased their rigidity, and, hence, further increased moduli. As for
471 $\tan\delta$, it showed a peak at about 70 °C. Baldino et al. (2018) also observed the peaks of phase
472 angle at 60.9 °C when gluten free doughs contained HPMC which is related to the phase
473 separation of HPMC. However, comparing to their observation, $\tan\delta$ peaks in Figure 5d had
474 lower value and occurred at a higher temperature but MC has a lower gelation temperature than
475 HPMC. Moreover, phase separation between amylose and amylopectin in baked wheat bread
476 crumb has been reported (Hug-Iten, Handschin, Conde-Petit & Escher, 1999). Therefore, the
477 $\tan\delta$ peaks are possibly attributed to the phase separation between amylose and amylopectin.
478 The reasons for G' and G'' decreasing after peaks might be the melting of remaining crystallites,

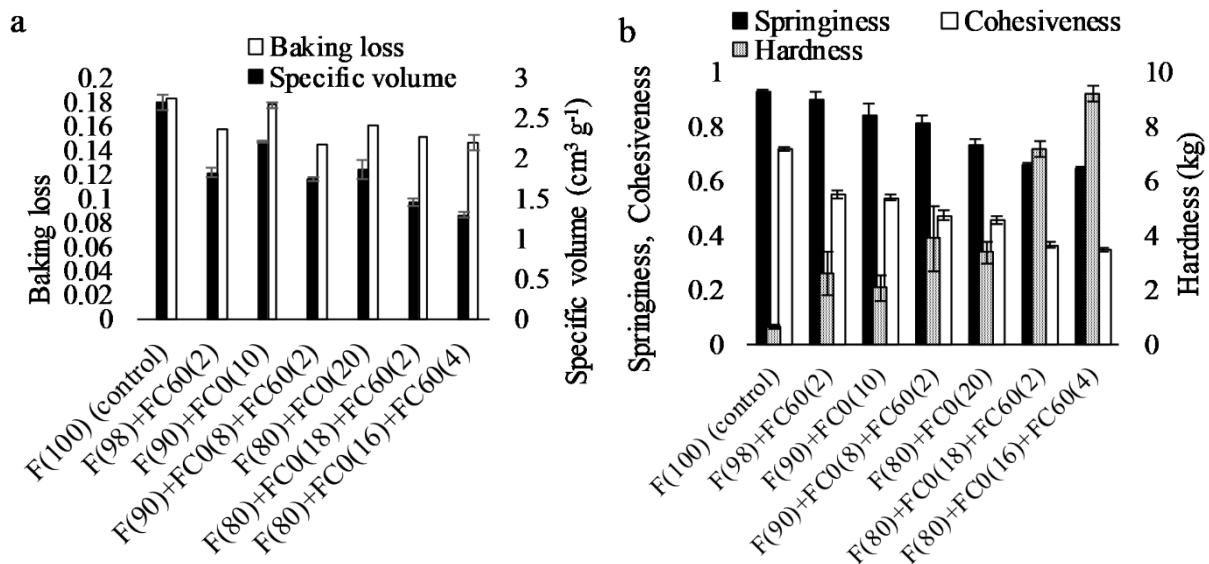
479 separation of amylose and amylopectin, amylopectin matrix breakdown, and disentanglement
480 of amylopectin chains, which lead to granule softening (Keetels, vanVliet & Walstra, 1996).

481 **3.5. Bread qualities**

482 The influence of the addition of FC0 and/or FC60 on specific volumes and baking loss of starch
483 reduced gluten free bread are shown in Figure 6a. The influence on the moisture content of the
484 centre crumb is insignificant which is not shown. The baking loss is significantly reduced upon
485 the additions of FC0 and/or, especially, FC60, which could be attributed to their high water
486 holding ability. It could also be because of the denser crumb structure where water evaporation
487 within the gas cells was restrained but water diffusion in the crumb matrix became more
488 dominant. The FC0/FC60 additions significantly reduced the specific volume, which is the
489 same as their influences on final proofing volume. The close correlation between loaf volume
490 and final proofing volume is also documented in a previous study (Ren et al., Unpublished
491 results-a), which suggests that loaves are stable during both the later stage of proofing and oven
492 rising. The detrimental effect on specific volume of fibre addition and generation of denser
493 crumb structure have also been observed in studies on both wheat bread and gluten free bread
494 (Demirkesen et al., 2010; Gómez, Oliete, Caballero, Ronda & Blanco, 2008; Gomez, Ronda,
495 Blanco, Caballero & Apesteguía, 2003).

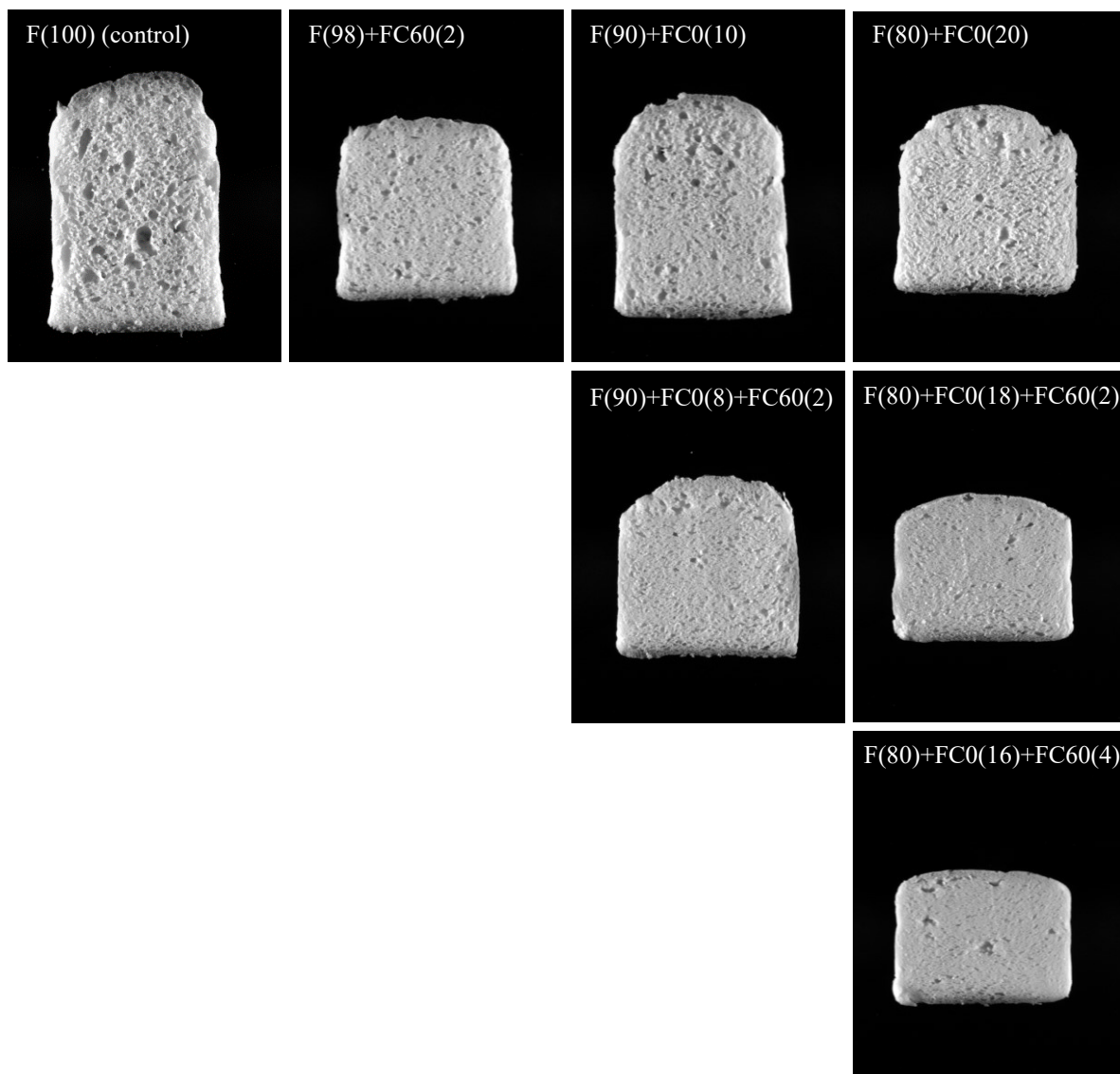
496 The crumb was evaluated by C Cell and images are shown in Figure 7. Six C Cell parameters
497 are chosen to describe the crumb structures (Table 5). The area of cells indicates the percentage
498 of the cells of the total slice area and the number of cells is the number of discrete gas cells.
499 The addition of FC0 and/or FC60 increased the number of cells but decreased the area of cells
500 and wall thickness, and, consequently, reduced cell diameters. With the fact that the specific
501 volume was decreased, which led to a smaller slice area, a finer crumb structure was obtained.
502 Comparing to the addition of FC0, FC60 was more effective in altering these parameters. It
503 can be attributed to the fibrous structure of FC0 and, especially, FC60, which formed a fine
504 fibrous framework. The fibrous framework stabilised the porous structure of doughs and loaves
505 and allowed the gelatinised starch to attach on and, therefore, reinforce the starch matrix.
506 Additionally, area of holes and top concavity, which reflect the structural instability are not
507 significant among all loaves.

508 The texture properties of bread were evaluated by TPA. Hardness, springiness, and
 509 cohesiveness are shown in Figure 6b. Specific volume is negatively correlated with hardness
 510 due to a denser crumb structure. As expected, loaves with FC0 and FC60, which have lower
 511 specific volume, have a harder crumb. They also show lower springiness and cohesiveness,
 512 which indicates that the crumb is less resistant to the applied large deformation. Good wheat
 513 bread is expected to have a thinner cell wall and uniform cells which, therefore, has softer and
 514 more elastic texture providing good mouth feel (Scanlon & Zghal, 2001). However, due to the
 515 absence of gluten and the more compact crumb structure, the starch reduced gluten free loaves
 516 show the opposite that they have smaller cells, thinner cell walls, but harder and less springy
 517 crumb.



518

519 Figure 6. Baking lose, specific volume (a), and textural properties (b) of starch replaced gluten free
 520 breads. Error bars represent the standard deviations averaging four replicates.



521

522 Figure 7. Images of starch replaced gluten free breads.

523

524 Table 5. C-Cell parameters of starch redplaced gluten free bread.

	Top Concavity (%)	Number of Cells	Area of Cells (%)	Area of Holes (%)	Wall Thickness (mm)	Cell Diameter (mm)
F(100) (control)	0.070.02 ^a	3523.25±27.90 ^a	50.83±0.96 ^a	2.41±0.39 ^{ac}	0.50±0.01 ^a	2.61±0.07 ^a
F(98)+FC60(2)	0.78±0.38 ^b	3986.75±91.51 ^{ab}	44.73±0.84 ^b	1.33±0.88 ^{abc}	0.44±0.01 ^{cd}	1.53±0.07 ^b
F(90)+FC0(10)	0.45±0.14 ^{bc}	3781.50±84.82 ^{ab}	48.20±0.57 ^c	1.43±0.81 ^{abc}	0.46±0.01 ^b	1.88±0.09 ^c
F(90)+FC0(8)+FC60(2)	0.56±0.07 ^{bc}	4033.50±69.00 ^b	45.00±0.43 ^b	0.85±0.48 ^{ab}	0.43±0.00 ^d	1.44±0.02 ^b
F(80)+FC0(20)	0.47±0.07 ^{bc}	3559.75±246.75 ^a	47.08±0.49 ^c	0.26±0.28 ^b	0.46±0.01 ^{bc}	1.76±0.13 ^c
F(80)+FC0(18)+FC60(2)	0.27±0.04 ^{ac}	4084.50±362.24 ^b	43.88±0.96 ^{bd}	1.29±0.82 ^{abc}	0.39±0.01 ^c	1.14±0.10 ^d
F(80)+FC0(16)+FC60(4)	0.21±0.04 ^{ac}	4065.00±274.07 ^b	42.73±0.78 ^d	2.56±0.94 ^c	0.38±0.01 ^c	1.09±0.06 ^d

Dara are shown as mean ± standard deviation. Values with different letters in the same column were significantly different ($p < 0.05$).

525 4. Conclusion

526 The conducted study aimed to evaluate the starch reduction of gluten free doughs and bread by
527 FC0 and FC60 fibres. The fundamental dough rheological properties were analysed using the
528 weak gel model and the generalised Maxwell model. The addition of FC0 and/or FC60
529 significantly increased the dough strength shown as increased viscosity, yield zone, and storage
530 and loss moduli measured by fundamental rheological measurements, and dough firmness,
531 cohesiveness, consistency and index of viscosity measured by empirical measurements. As the
532 outcome of the fibrillation treatment, with high water absorbability and entangled fibrous
533 aggregate structure, FC60 is more efficient than FC0 in altering the rheological properties.
534 Further analysis of the mechanical spectra by generalised Maxwell model suggests that gluten
535 free doughs are structurally and rheologically analogous to a viscoelastic fluid instead of a solid.
536 It also suggests that doughs are flowable at the time scales of structure developing during
537 proofing which is longer than their relaxation times. Generally, a comprehensive design of
538 experiments including both fundamental and empirical analysis would be necessary to
539 maximise the characterisation of doughs. Rheological properties of doughs influence the
540 proofing behaviour where the addition of FC0/FC60 mainly restrained the volume increase
541 during the later stage of proofing (after approximately 30 min). The fibrillated cellulose has an
542 effective enhancement of overall composite properties beyond the effects of the 'inert'
543 unfibrillated filler. Fibrillation of cellulose increased its similarity of pasting properties to flour
544 in a cellulose/flour blend. Volume and water competition of FC0 and FC60 restricted the
545 hydration and swelling of starch granules significantly influencing the pasting and thermal-
546 mechanical behaviours of the blends. The additions of FC0 and FC60 decrease the specific
547 volume of gluten free loaves, which is correlated with the decrease of final volume during
548 proofing. The additions of FC0 and FC60 generate denser and harder but finer crumb. FC0 and,
549 especially, FC60, play a role as a framework which stabilise the porous dough/crumb structure
550 and reinforce the starch matrix. The further improvement of cellulose enriched gluten free
551 bread might rely on the structuring of added hydrocolloids and fibrillated cellulose and
552 optimisation of formulation including water addition levels to increase specific volume by
553 obtaining desired rheological properties of doughs.

554 **Acknowledgement**

555 This work was supported by the University of Nottingham (Vice-Chancellor's Scholarship for
556 Research Excellence (International)) and PepsiCo. The views and opinions expressed in this
557 manuscript are those of the author and do not necessarily reflect the position or policy of
558 PepsiCo.

Reference

- Agarwal, D., Hewson, L. & Foster, T. J. (2018a). A comparison of the sensory and rheological properties of different cellulosic fibres for food. *Food & Function*, 9(2), 1144-1151.
- Agarwal, D., MacNaughtan, W. & Foster, T. J. (2018b). Interactions between microfibrillar cellulose and carboxymethyl cellulose in an aqueous suspension. *Carbohydrate Polymers*, 185, 112-119.
- Armero, E. & Collar, C. (1997). Texture properties of formulated wheat doughs Relationships with dough and bread technological quality. *Zeitschrift für Lebensmitteluntersuchung und -Forschung A*, 204(2), 136-145.
- Aziz, I., Lewis, N. R., Hadjivassiliou, M., Winfield, S. N., Rugg, N., Kelsall, A., Newrick, L. & Sanders, D. S. (2014). A UK study assessing the population prevalence of self-reported gluten sensitivity and referral characteristics to secondary care. *European Journal of Gastroenterology & Hepatology*, 26(1), 33-39.
- Babin, P., Della Valle, G., Chiron, H., Cloetens, P., Hoszowska, J., Pernot, P., Réguerre, A. L., Salvo, L. & Dendievel, R. (2006). Fast X-ray tomography analysis of bubble growth and foam setting during breadmaking. *Journal of Cereal Science*, 43(3), 393-397.
- Baldino, N., Laitano, F., Lupi, F. R., Curcio, S. & Gabriele, D. (2018). Effect of HPMC and CMC on rheological behavior at different temperatures of gluten-free bread formulations based on rice and buckwheat flours. *European Food Research and Technology*, 244(10), 1829-1842.
- Baumgaertel, M. & Winter, H. H. (1992). Interrelation between continuous and discrete relaxation time spectra. *Journal of Non-Newtonian Fluid Mechanics*, 44, 15-36.
- BeMiller, J. N. (2011). Pasting, paste, and gel properties of starch–hydrocolloid combinations. *Carbohydrate Polymers*, 86(2), 386-423.
- Bousquières, J., Michon, C. & Bonazzi, C. (2017). Functional properties of cellulose derivatives to tailor a model sponge cake using rheology and cellular structure analysis. *Food Hydrocolloids*, 70, 304-312.
- Capristo, E., Addolorato, G., Mingrone, G., De Gaetano, A., Greco, A. V., Tataranni, P. A. & Gasbarrini, G. (2000). Changes in body composition, substrate oxidation, and resting metabolic rate in adult celiac disease patients after a 1-y gluten-free diet treatment. *The American Journal of Clinical Nutrition*, 72(1), 76-81.
- Catassi, C., Anderson, R. P., Hill, I. D., Koletzko, S., Lionetti, E., Mouane, N., Schumann, M. & Yachha, S. K. (2012). World perspective on celiac disease. *Journal of Pediatric Gastroenterology and Nutrition*, 55(5), 494-499.
- Chen, W. Z. & Hosney, R. C. (1995). Development of an objective method for dough stickiness. *LWT - Food Science and Technology*, 28(5), 467-473.

- Collar, C., Santos, E. & Rosell, C. M. (2007). Assessment of the rheological profile of fibre-enriched bread doughs by response surface methodology. *Journal of Food Engineering*, 78(3), 820-826.
- Czaja-Bulsa, G. (2015). Non coeliac gluten sensitivity - A new disease with gluten intolerance. *Clinical Nutrition*, 34(2), 189-194.
- Demirkesen, I., Mert, B., Sumnu, G. & Sahin, S. (2010). Utilization of chestnut flour in gluten-free bread formulations. *Journal of Food Engineering*, 101(3), 329-336.
- Di Sabatino, A. & Corazza, G. R. (2009). Coeliac disease. *The Lancet*, 373(9673), 1480-1493.
- Díaz-Calderón, P., MacNaughtan, B., Hill, S., Foster, T., Enrione, J. & Mitchell, J. (2018). Changes in gelatinisation and pasting properties of various starches (wheat, maize and waxy maize) by the addition of bacterial cellulose fibrils. *Food Hydrocolloids*, 80, 274-280.
- Djordjevic, M., Soronja-Simovic, D., Nikolic, I., Djordjevic, M., Seres, Z. & Milasinovic-Seremesic, M. (2019). Sugar beet and apple fibres coupled with hydroxypropylmethylcellulose as functional ingredients in gluten-free formulations: Rheological, technological and sensory aspects. *Food Chemistry*, 295, 189-197.
- Fasano, A. & Catassi, C. (2001). Current approaches to diagnosis and treatment of celiac disease: An evolving spectrum. *Gastroenterology*, 120(3), 636-651.
- Ferry, J. D. (1980). *Viscoelastic properties of polymers* (3rd ed.). New York: John Wiley & Sons.
- Gabriele, D., de Cindio, B. & D'Antona, P. (2001). A weak gel model for foods. *Rheologica Acta*, 40(2), 120-127.
- Georgopoulos, T., Larsson, H. & Eliasson, A.-C. (2004). A comparison of the rheological properties of wheat flour dough and its gluten prepared by ultracentrifugation. *Food Hydrocolloids*, 18(1), 143-151.
- Gómez, M., Oliete, B., Caballero, P. A., Ronda, F. & Blanco, C. A. (2008). Effect of nut paste enrichment on wheat dough rheology and bread volume. *Food Science and Technology International*, 14(1), 57-65.
- Gomez, M., Ronda, F., Blanco, C. A., Caballero, P. A. & Apesteguia, A. (2003). Effect of dietary fibre on dough rheology and bread quality. *European Food Research and Technology*, 216(1), 51-56.
- Grenier, D., Lucas, T. & Le Ray, D. (2010). Measurement of local pressure during proving of bread dough sticks: Contribution of surface tension and dough viscosity to gas pressure in bubbles. *Journal of Cereal Science*, 52(3), 373-377.
- Hager, A.-S., Wolter, A., Czerny, M., Bez, J., Zannini, E., Arendt, E. K. & Czerny, M. (2012). Investigation of product quality, sensory profile and ultrastructure of breads made from a range of commercial gluten-free flours compared to their wheat counterparts. *European Food Research and Technology*, 235(2), 333-344.

- Haque, A. & Morris, E. R. (1994). Combined use of ispaghula and HPMC to replace or augment gluten in breadmaking. *Food Research International*, 27(4), 379-393.
- Heddleson, S. S., Hamann, D. D., Lineback, D. R. & Slade, L. (1994). Pressure-sensitive adhesive properties of wheat-flour dough and the influence of temperature, separation rate, and moisture-content. *Cereal Chemistry*, 71(6), 564-570.
- Herrick, F. W., Casebier, R. L., Hamilton, J. K. & Sandberg, K. R. (1983). Microfibrillated cellulose: morphology and accessibility. In *Journal of Applied Polymer Science: Applied Polymer Symposium; (United States)* (Vol. 37): ITT Rayonier Inc., Shelton, WA.
- Hoseney, R. C. & Smewing, J. (1999). Instrumental measurement of stickiness of doughs and other foods. *Journal of Texture Studies*, 30(2), 123-136.
- Hug-Iten, S., Handschin, S., Conde-Petit, B. & Escher, F. (1999). Changes in starch microstructure on baking and staling of wheat bread. *Food Science and Technology-Lebensmittel-Wissenschaft & Technologie*, 32(5), 255-260.
- Kaufman, R. C., Wilson, J. D., Bean, S. R., Herald, T. J. & Shi, Y. C. (2015). Development of a 96-well plate iodine binding assay for amylose content determination. *Carbohydrate Polymers*, 115, 444-447.
- Keetels, C. J. A. M., vanVliet, T. & Walstra, P. (1996). Gelation and retrogradation of concentrated starch systems .1. Gelation. *Food Hydrocolloids*, 10(3), 343-353.
- Laun, H. M. (1986). Prediction of elastic strains of polymer melts in shear and elongation. *Journal of Rheology*, 30(3), 459-501.
- Lazaridou, A., Duta, D., Papageorgiou, M., Belc, N. & Biliaderis, C. G. (2007). Effects of hydrocolloids on dough rheology and bread quality parameters in gluten-free formulations. *Journal of Food Engineering*, 79(3), 1033-1047.
- López-Rubio, A., Lagaron, J. M., Ankerfors, M., Lindström, T., Nordqvist, D., Mattozzi, A. & Hedenqvist, M. S. (2007). Enhanced film forming and film properties of amylopectin using micro-fibrillated cellulose. *Carbohydrate Polymers*, 68(4), 718-727.
- McCarthy, D. F., Gallagher, E., Gormley, T. R., Schober, T. J. & Arendt, E. K. (2005). Application of response surface methodology in the development of gluten-free bread. *Cereal Chemistry*, 82(5), 609-615.
- Mills, E. N. C., Wilde, P. J., Salt, L. J. & Skeggs, P. (2003). Bubble formation and stabilization in bread dough. *Food and Bioproducts Processing*, 81(3), 189-193.
- Nakagaito, A. N. & Yano, H. (2004). The effect of morphological changes from pulp fiber towards nano-scale fibrillated cellulose on the mechanical properties of high-strength plant fiber based composites. *Applied Physics a-Materials Science & Processing*, 78(4), 547-552.

- Naruenartwongsakul, S., Chinnan, M. S., Bhumiratana, S. & Yoovidhya, T. (2004). Pasting characteristics of wheat flour-based batters containing cellulose ethers. *LWT - Food Science and Technology*, 37(4), 489-495.
- Ng, M., Fleming, T., Robinson, M., Thomson, B., Graetz, N., Margono, C., Mullany, E. C., Biryukov, S., Abbafati, C., Abera, S. F., Abraham, J. P., Abu-Rmeileh, N. M. E., Achoki, T., AlBuhairan, F. S., Alemu, Z. A., Alfonso, R., Ali, M. K., Ali, R., Guzman, N. A., Ammar, W., Anwari, P., Banerjee, A., Barquera, S., Basu, S., Bennett, D. A., Bhutta, Z., Blore, J., Cabral, N., Nonato, I. C., Chang, J.-C., Chowdhury, R., Courville, K. J., Criqui, M. H., Cundiff, D. K., Dabhadkar, K. C., Dandona, L., Davis, A., Dayama, A., Dharmaratne, S. D., Ding, E. L., Durrani, A. M., Esteghamati, A., Farzadfar, F., Fay, D. F. J., Feigin, V. L., Flaxman, A., Forouzanfar, M. H., Goto, A., Green, M. A., Gupta, R., Hafezi-Nejad, N., Hankey, G. J., Harewood, H. C., Havmoeller, R., Hay, S., Hernandez, L., Hussein, A., Idrisov, B. T., Ikeda, N., Islami, F., Jahangir, E., Jassal, S. K., Jee, S. H., Jeffreys, M., Jonas, J. B., Kabagambe, E. K., Khalifa, S. E. A. H., Kengne, A. P., Khader, Y. S., Khang, Y.-H., Kim, D., Kimokoti, R. W., Kinge, J. M., Kokubo, Y., Kosen, S., Kwan, G., Lai, T., Leinsalu, M., Li, Y., Liang, X., Liu, S., Logroscino, G., Lotufo, P. A., Lu, Y., Ma, J., Mainoo, N. K., Mensah, G. A., Merriman, T. R., Mokdad, A. H., Moschandreas, J., Naghavi, M., Naheed, A., Nand, D., Narayan, K. M. V., Nelson, E. L., Neuhouser, M. L., Nisar, M. I., Ohkubo, T., Oti, S. O., Pedroza, A., Prabhakaran, D., Roy, N., Sampson, U., Seo, H., Sepanlou, S. G., Shibuya, K., Shiri, R., Shiue, I., Singh, G. M., Singh, J. A., Skirbekk, V., Stapelberg, N. J. C., Sturua, L., Sykes, B. L., Tobias, M., Tran, B. X., Trasande, L., Toyoshima, H., van de Vijver, S., Vasankari, T. J., Veerman, J. L., Velasquez-Melendez, G., Vlassov, V. V., Vollset, S. E., Vos, T., Wang, C., Wang, X., Weiderpass, E., Werdecker, A., Wright, J. L., Yang, Y. C., Yatsuya, H., Yoon, J., Yoon, S.-J., Zhao, Y., Zhou, M., Zhu, S., Lopez, A. D., Murray, C. J. L. & Gakidou, E. (2014). Global, regional, and national prevalence of overweight and obesity in children and adults during 1980–2013: A systematic analysis for the Global Burden of Disease Study 2013. *The Lancet*, 384(9945), 766-781.
- Nishita, K. D., Roberts, R. L., Bean, M. M. & Kennedy, B. M. (1976). Development of a yeast-leavened rice-bread formula. *Cereal Chemistry*, 53(5), 626-635.
- Pääkkö, M., Ankerfors, M., Kosonen, H., Nykänen, A., Ahola, S., Österberg, M., Ruokolainen, J., Laine, J., Larsson, P. T., Ikkala, O. & Lindström, T. (2007). Enzymatic hydrolysis combined with mechanical shearing and high-pressure homogenization for nanoscale cellulose fibrils and strong gels. *Biomacromolecules*, 8(6), 1934-1941.
- Paciulli, M., Rinaldi, M., Cirlini, M., Scazzina, F. & Chiavaro, E. (2016). Chestnut flour addition in commercial gluten-free bread: A shelf-life study. *LWT - Food Science and Technology*, 70, 88-95.
- Phimolsiripol, Y., Mukprasirt, A. & Schoenlechner, R. (2012). Quality improvement of rice-based gluten-free bread using different dietary fibre fractions of rice bran. *Journal of Cereal Science*, 56(2), 389-395.
- Ren, Y., Linter, B. R. & Foster, T. J. (2020). Cellulose fibrillation and interaction with psyllium seed husk heteroxylan. *Food Hydrocolloids*, 104, 105725.

- Ren, Y., Linter, B. R. & Foster, T. J. (Unpublished results-a). A comprehensive investigation of gluten free bread dough rheology, proving and baking performance and bread qualities by response surface design and principal component analysis. *Manuscript submitted for publication*.
- Ren, Y., Yakubov, G., Linter, B. R., MacNaughtan, B. & Foster, T. J. (Unpublished results-b). Temperature fractionation, and physicochemical and rheological analysis of psyllium. *Manuscript submitted for publication*.
- Romano, A., Toraldo, G., Cavella, S. & Masi, P. (2007). Description of leavening of bread dough with mathematical modelling. *Journal of Food Engineering*, 83(2), 142-148.
- Ronda, F., Pérez-Quirce, S., Angioloni, A. & Collar, C. (2013). Impact of viscous dietary fibres on the viscoelastic behaviour of gluten-free formulated rice doughs: A fundamental and empirical rheological approach. *Food Hydrocolloids*, 32(2), 252-262.
- Sahlström, S., Park, W. & Shelton, D. R. (2004). Factors influencing yeast fermentation and the effect of LMW sugars and yeast fermentation on hearth bread quality. *Cereal Chemistry*, 81(3), 328-335.
- Sangnark, A. & Noomhorm, A. (2004). Effect of dietary fiber from sugarcane bagasse and sucrose ester on dough and bread properties. *LWT - Food Science and Technology*, 37(7), 697-704.
- Sapone, A., Bai, J. C., Ciacci, C., Dolinsek, J., Green, P. H., Hadjivassiliou, M., Kaukinen, K., Rostami, K., Sanders, D. S., Schumann, M., Ullrich, R., Villalta, D., Volta, U., Catassi, C. & Fasano, A. (2012). Spectrum of gluten-related disorders: consensus on new nomenclature and classification. *BMC Medicine*, 10(1), 1-12.
- Scanlon, M. G. & Zghal, M. C. (2001). Bread properties and crumb structure. *Food Research International*, 34(10), 841-864.
- Schober, T. J., Bean, S. R. & Boyle, D. L. (2007). Gluten-free sorghum bread improved by sourdough fermentation: Biochemical, rheological, and microstructural background. *Journal of Agricultural and Food Chemistry*, 55(13), 5137-5146.
- Schuppan, D. (2000). Current concepts of celiac disease pathogenesis. *Gastroenterology*, 119(1), 234-242.
- Shah, P., Campbell, G. M., McKee, S. L. & Rielly, C. D. (1998). Proving of bread dough: Modelling the growth of individual bubbles. *Food and Bioproducts Processing*, 76(2), 73-79.
- Shaw, J. E., Sicree, R. A. & Zimmet, P. Z. (2010). Global estimates of the prevalence of diabetes for 2010 and 2030. *Diabetes Research and Clinical Practice*, 87(1), 4-14.
- Stenstad, P., Andresen, M., Tanem, B. S. & Stenius, P. (2008). Chemical surface modifications of microfibrillated cellulose. *Cellulose*, 15(1), 35-45.
- Sullo, A. & Foster, T. J. (2010). Characterisation of starch/cellulose blends. *Annual Transactions of the Nordic Rheology Society*, 18, 1-7.

- Tanner, R. I., Qi, F. & Dai, S.-C. (2008). Bread dough rheology and recoil: I. Rheology. *Journal of Non-Newtonian Fluid Mechanics*, 148(1), 33-40.
- Turbak, A. F., Snyder, F. W. & Sandberg, K. R. (1982). Food products containing microfibrillated cellulose. US4341807.
- Turbak, A. F., Snyder, F. W. & Sandberg, K. R. (1983a). Microfibrillated cellulose. US4374702A.
- Turbak, A. F., Snyder, F. W. & Sandberg, K. R. (1983b). Suspensions containing microfibrillated cellulose. US4378381A.
- Upadhyay, R., Ghosal, D. & Mehra, A. (2012). Characterization of bread dough: Rheological properties and microstructure. *Journal of Food Engineering*, 109(1), 104-113.
- Van Riemsdijk, L. E., van der Goot, A. J., Hamer, R. J. & Boom, R. M. (2011). Preparation of gluten-free bread using a meso-structured whey protein particle system. *Journal of Cereal Science*, 53(3), 355-361.
- Van Vliet, T., Janssen, A. M., Bloksma, A. H. & Walstra, P. (1992). Strain hardening of dough as a requirement for gas retention. *Journal of Texture Studies*, 23(4), 439-460.
- Villanueva, M., Harasym, J., Muñoz, J. M. & Ronda, F. (2019). Rice flour physically modified by microwave radiation improves viscoelastic behavior of doughs and its bread-making performance. *Food Hydrocolloids*, 90, 472-481.
- Wagner, M., Quellec, S., Trystram, G. & Lucas, T. (2008). MRI evaluation of local expansion in bread crumb during baking. *Journal of Cereal Science*, 48(1), 213-223.
- Wahab, P. J., Meijer, J. W., Goerres, M. S. & Mulder, C. J. (2002). Coeliac disease: changing views on gluten-sensitive enteropathy. *Scandinavian Journal of Gastroenterology*, 37(236), 60-65.
- Walls, H. J., Caines, S. B., Sanchez, A. M. & Khan, S. A. (2003). Yield stress and wall slip phenomena in colloidal silica gels. *Journal of Rheology*, 47(4), 847-868.
- Zandonadi, R. P., Botelho, R. B. A. & Araujo, W. M. C. (2009). Psyllium as a substitute for gluten in bread. *Journal of the American Dietetic Association*, 109(10), 1781-1784.
- Zimmermann, T., Pöhler, E. & Geiger, T. (2004). Cellulose fibrils for polymer reinforcement. *Advanced Engineering Materials*, 6(9), 754-761.