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**Citation for published version:**

Naval-Sanchez, M, McWilliam, S, Evans, BJ, Yáñez, JM, Houston, R & Kijas, JW 2020, 'Changed Patterns of Genomic Variation Following Recent Domestication: Selection Sweeps in Farmed Atlantic Salmon', *Frontiers in genetics*. <https://doi.org/10.3389/fgene.2020.00264>

**Digital Object Identifier (DOI):**

[10.3389/fgene.2020.00264](https://doi.org/10.3389/fgene.2020.00264)

**Link:**

[Link to publication record in Edinburgh Research Explorer](#)

**Document Version:**

Publisher's PDF, also known as Version of record

**Published In:**

Frontiers in genetics

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# Changed Patterns of Genomic Variation Following Recent Domestication: Selection Sweeps in Farmed Atlantic Salmon

Marina Naval-Sanchez<sup>1†</sup>, Sean McWilliam<sup>1</sup>, Bradley Evans<sup>2</sup>, José M. Yáñez<sup>3</sup>, Ross D. Houston<sup>4</sup> and James W. Kijas<sup>1\*</sup>

## OPEN ACCESS

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### \*Correspondence:

James W. Kijas  
James.Kijas@csiro.au

### † Present address:

Marina Naval-Sanchez,  
Institute for Molecular Biosciences,  
The University of Queensland,  
Brisbane, QLD, Australia

### Specialty section:

This article was submitted to  
Livestock Genomics,  
a section of the journal  
Frontiers in Genetics

**Received:** 03 December 2019

**Accepted:** 05 March 2020

**Published:** 03 April 2020

### Citation:

Naval-Sanchez M, McWilliam S,  
Evans B, Yáñez JM, Houston RD and  
Kijas JW (2020) Changed Patterns  
of Genomic Variation Following  
Recent Domestication: Selection  
Sweeps in Farmed Atlantic Salmon.  
*Front. Genet.* 11:264.  
doi: 10.3389/fgene.2020.00264

<sup>1</sup> CSIRO Agriculture and Food, Brisbane, QLD, Australia, <sup>2</sup> Salmon Enterprises of Tasmania Pty. Limited, Wayatinah, TAS, Australia, <sup>3</sup> Faculty of Veterinary and Animal Sciences, University of Chile, Santiago, Chile, <sup>4</sup> The Roslin Institute and Royal (Dick) School of Veterinary Studies, University of Edinburgh, Midlothian, United Kingdom

The introduction of wild Atlantic salmon into captivity, and their subsequent artificial selection for production traits, has caused phenotypic differences between domesticated fish and their wild counterparts. Identification of regions of the genome underlying these changes offers the promise of characterizing the early biological consequences of domestication. In the current study, we sequenced a population of farmed European Atlantic salmon and compared the observed patterns of SNP variation to those found in conspecific wild populations. This identified 139 genomic regions that contained significantly elevated SNP homozygosity in farmed fish when compared to their wild counterparts. The most extreme was adjacent to *versican*, a gene involved in control of neural crest cell migration. To control for false positive signals, a second and independent dataset of farmed and wild European Atlantic salmon was assessed using the same methodology. A total of 81 outlier regions detected in the first dataset showed significantly reduced homozygosity within the second one, strongly suggesting the genomic regions identified are enriched for true selection sweeps. Examination of the associated genes identified a number previously characterized as targets of selection in other domestic species and that have roles in development, behavior and olfactory system. These include *arvcf*, *sema6*, *errb4*, *id2-like*, and *6n1-like* genes. Finally, we searched for evidence of parallel sweeps using a farmed population of North American origin. This failed to detect a convincing overlap to the putative sweeps present in European populations, suggesting the factors that drive patterns of variation under domestication and early artificial selection were largely independent. This is the first analysis on domestication of aquaculture species exploiting whole-genome sequence data and resulted in the identification of sweeps common to multiple independent populations of farmed European Atlantic salmon.

**Keywords:** Atlantic salmon, selection sweep, domestication, evolution, whole genome sequence

## INTRODUCTION

Comparison between domestic animals and their wild counterparts reveals a spectrum of fundamental phenotypic and behavioral differences that includes body conformation, reproductive seasonality and life history traits. The genomic consequences of domestication, whereby previously wild populations are progressively brought under captive management and controlled breeding, has been the subject of evolutionary investigation in dog (*Canis familiaris*) (Axelsson et al., 2013; Wang et al., 2013), pig (*Sus scrofa*) (Larson et al., 2005; Frantz et al., 2015), sheep (*Ovis aries*) (Naval-Sanchez et al., 2018), sheep and goat (Alberto et al., 2018), chicken (*Gallus gallus*) (Xiang et al., 2014), rabbit (*Oryctolagus cuniculus*) (Carneiro et al., 2014), and several other species (Koenig et al., 2013; Lemmon et al., 2014). Most mammalian domesticates have a population history spanning 10,000 years or more, creating significant challenges to distinguishing evolutionary events that arose early during domestication and selection from those caused during subsequent strong positive selection and the formation of breeds. This prompted us to explore the genomic consequence of domestication in farmed Atlantic salmon (*Salmo salar*) for three key reasons. Firstly, the onset of commercial Atlantic salmon farming for human consumption commenced as recently as the 1960s in Norway. This offers the opportunity to access the early evolutionary consequences of domestication. Since the 1960's, the industry has expanded rapidly and marked phenotypic differences now exist between farmed and wild Atlantic Salmon. These differences take many forms as extensively reviewed by Glover et al. (2017), with the two largest and most consistently observed being growth rate and survival in the wild (Fleming et al., 2000; McGinnity et al., 2003; Glover et al., 2009, 2017; Skaala et al., 2012, 2019; Solberg et al., 2013). Other differences include disease resistance, age at sexual maturation and product quality traits (Gjedrem et al., 2012; Gutierrez et al., 2014, 2016). Secondly, both European and North America wild Atlantic salmon have been independently domesticated to initiate a range of breeding programs around the world. A comparison of selection sweeps in farmed animals of European and North American origin offers the opportunity to identify genes and gene networks potentially involved in the phenotypic changes common to both farmed populations. Finally, the identification of genetic changes underlying aquaculture-associated phenotypes offers the opportunity to identify DNA markers with potential for use in selective breeding to expedite improvement of key production traits, and to understand how domestication mediated changes in escaped farmed animals may impact the population dynamics of wild populations (Glover et al., 2013; Wringe et al., 2018). Domestication-associated genetic changes are not well understood in aquaculture species. In Atlantic salmon, several studies have compared aquaculture strains against wild populations (Bourret et al., 2011; Vasemägi et al., 2012; Gutierrez et al., 2014; Mäkinen et al., 2015; Liu et al., 2017; López et al., 2019a). These have used SNP arrays that generate genome-wide allele frequency data for tens or hundreds of thousands of loci. Comparison of the outlier loci identified between studies has shown little or no overlap. This may change

as additional studies emerge, and as the size of the populations under investigation and the scale and precision of genomic datasets improve.

In the current study, we utilized whole-genome sequence data obtained from populations of both farmed and wild European salmon to call SNPs and search for evidence of selection sweeps associated with domestication. Genomic regions that displayed low heterozygosity were identified in each of the groups, and a number were subsequently identified using independent populations of salmon. This increased the likelihood they reflect real selection events as opposed to neutral evolutionary forces such as random drift. Genes within regions under selection were involved in neurogenesis or previously associated with domestication in other species.

## MATERIALS AND METHODS

### Samples and Sequencing

Three datasets containing genome sequence from wild and domestic Atlantic salmon were used of either European or North American origin (**Table 1**). Dataset 1 contained whole genome sequence derived from four pools of DNA sampled from a commercial breeding program (Landcatch, United Kingdom). Each of the four pools were created by mixing individually extracted genomic DNA from 20 to 22 individuals at equimolar concentrations, with the individuals being sampled from two different year classes of the breeding program. Each pool was sequenced (pool-seq) to a depth of 20 – 24 fold coverage using paired-end libraries with an Illumina HiSeq 2500 (Edinburgh Genomics, United Kingdom). The origin of this farmed population is composed of a mixture of wild fish from a range of Scottish rivers and introductions from Norway, including from the Norwegian river Namsen and the Mowi program. The population underwent approximately 10 generations of domestication and selection prior to sampling and sequencing. Dataset 1 also contained pool-seq data from European derived wild Atlantic salmon from previously published work (Ayllon et al., 2015, **Table 1**). The fastq files from six pools, each containing 20 individuals from a different river of Western Norway, were downloaded from the NCBI SRA (Bioproject number PRJNA293012). Dataset 2 sought to mirror Dataset 1 in that it contained both wild and domesticated European derived Atlantic salmon. Dataset 2 differed, however, in that it comprised individually sequenced genomes rather than pooled data (**Table 1**). The farmed population in Dataset 2 included fastq files of individual whole-genome sequences from 13 Chilean animals originating either from Scotland (Lochy and Landcatch strains) or Norway (Fanad and Mowi strains) (SRA deposition SRP059652) (Yáñez et al., 2016). Briefly, each individual was sequenced to a depth of 6 fold coverage using 100 bp paired-end libraries with an Illumina HiSeq 2000 machine (Macrogen, South Korea). The wild population comprised published fastq files of individual whole-genome sequences from 12 wild salmon from the Norwegian Atlantic clade rivers - Jølstra (3), Naustra (3), Namsen (3) and Argadvassdraget (3) downloaded from SRA (Bioproject PRJEB10744) (Barson et al., 2015). Dataset 3

**TABLE 1** | Origin of the whole genome sequences used.

Dataset	Origin	Type	Data	Population	n	Sex	References
1	EU	Farmed	Pool	Scotland	4*22	M/F	This Study
	EU	Wild	Pool	Eidselva	4*20	M	Ayllon et al., 2015
	EU	Wild	Pool	Aardselva	4*20	M	Ayllon et al., 2015
	EU	Wild	Pool	Flekke	4*20	M	Ayllon et al., 2015
	EU	Wild	Pool	Gloppenelva	4*20	M	Ayllon et al., 2015
	EU	Wild	Pool	Sulsdagen	4*20	M	Ayllon et al., 2015
	EU	Wild	Pool	Vorma	4*20	M	Ayllon et al., 2015
2	EU	Farmed	Indiv	Chile	13	M/F	Yáñez et al., 2016
	EU	Wild	Indiv	Jølstra	3	2M/1F	Barson et al., 2015
	EU	Wild	Indiv	Naustra	3	2M/1F	Barson et al., 2015
	EU	Wild	Indiv	Namsen	3	2M/1F	Barson et al., 2015
	EU	Wild	Indiv	Argadvassdraget	3	2M/1F	Barson et al., 2015
3	NA	Farmed	Indiv	Tasmanian	20	M/F	Kijas et al., 2018

Dataset 1 consisted of pooled samples. Dataset 2 and 3 consisted of samples sequenced individually. EU: European origin. NA, North American origin.

contained domesticated animals of North America origin, farmed in Tasmania, Australia (Kijas et al., 2018). A total of 19 animals from the SALTAS breeding program from different year classes and sex lineages were downloaded from NCBI Bio Project ID PRJNA403334, with individual animal raw sequence accession numbers SRR6019467 – SRR6019464.

## Mapping, Variant Calling, Annotation and PCA

Variant calling was performed independently for each Dataset (1 – 3) using the same pipeline and parameters. Sequence reads were first mapped to the salmon reference genome ICSASG\_v2 (Lien et al., 2016) using Burrows–Wheeler algorithm (bwa) (Li and Durbin, 2009), with default parameters. Duplicate reads were removed using Picard tools<sup>1</sup> and local realignment around INDELS was performed in accordance to GATK best practices (DePristo et al., 2011). Variants were called using GATK haplotypeCaller (v3.7) to produce joint genotyping calls within each dataset. Variants were filtered using bcftools filter (version 1.3.1) to remove variants (i) with mapping quality < 50 (ii) read depth < 5 and (iii) variants other than biallelic SNPs. Only variants with genotype calls in more than 90% of the samples per set were retained. The variant effect predictor tool from ensembl (version 78) was used to identify 24 separate SNP classifications in relation to the gene model annotated reference assembly ICSASG\_v2 (Lien et al., 2016). Classifications included coding, missense and non-synonymous substitutions, intronic and intergenic variants. In preparation for principal component analysis (PCA), SNP subsets were identified by removal of loci in linkage disequilibrium using the `-indep-pairwise 500 100 0.9` command implemented in PLINK v1.9<sup>2</sup> (Chang et al., 2015). This assessed genomic bins containing 500 SNP with a 100 SNP set size and removed loci with squared correlation exceeding 0.9. PCA was also performed using PLINK v1.9 before the eigenvector values were visualized by plotting in R v3.1.2.

<sup>1</sup><http://broadinstitute.github.io/picard/>

<sup>2</sup><https://www.cog-genomics.org/plink2>

## Selection Sweep Detection

To identify signatures of selection in sequence data we calculated a window based heterozygosity score  $H_p$  as in Rubin et al. (2010, 2012). In brief, we determined the number of reads corresponding to the most and least abundant allele ( $n_{MAJ}$  and  $n_{MIN}$ ) per SNP separately for farmed and wild fish populations. The  $H_p$  metric was estimated in 150 kb genome windows with 75 kb overlap, before windows containing less than 20 SNP were discarded. For each window we calculated  $H_p = 2 \sum n_{MAJ} \sum n_{MIN} / (\sum n_{MAJ} + \sum n_{MIN})^2$ , where  $\sum n_{MAJ}$  and  $\sum n_{MIN}$  are the sums of  $n_{MAJ}$  and, respectively,  $n_{MIN}$  for all SNPs in the window. Individual  $H_p$  values were then Z-transformed as follows:  $ZH_p = (H_p - \mu_{H_p}) / \sigma_{H_p}$ . To derive directionality of selection intensity, we calculated  $\Delta ZH_p = ZH_{p_{Wild}} - ZH_{p_{Farmed}}$ . Positive  $\Delta ZH_p$  values reflect genome regions containing loss of heterozygosity in farmed fish when compared to wild fish, consistent with positive selection post domestication. Multiple testing was performed by Bonferroni correction to account for evaluation of 150 Kb genome bins with a step size of 75 Kb. A second method to detect selection sweeps exploited allele frequency differences between populations measured as  $F_{ST}$  (reviewed by Luikart et al., 2003).  $F_{ST}$  was calculated using the same 150 kb windows as  $H_p$  and compared farmed and wild populations in accordance to the Cockerham and Weir (1984) implemented in vcftools (Danecek et al., 2011).

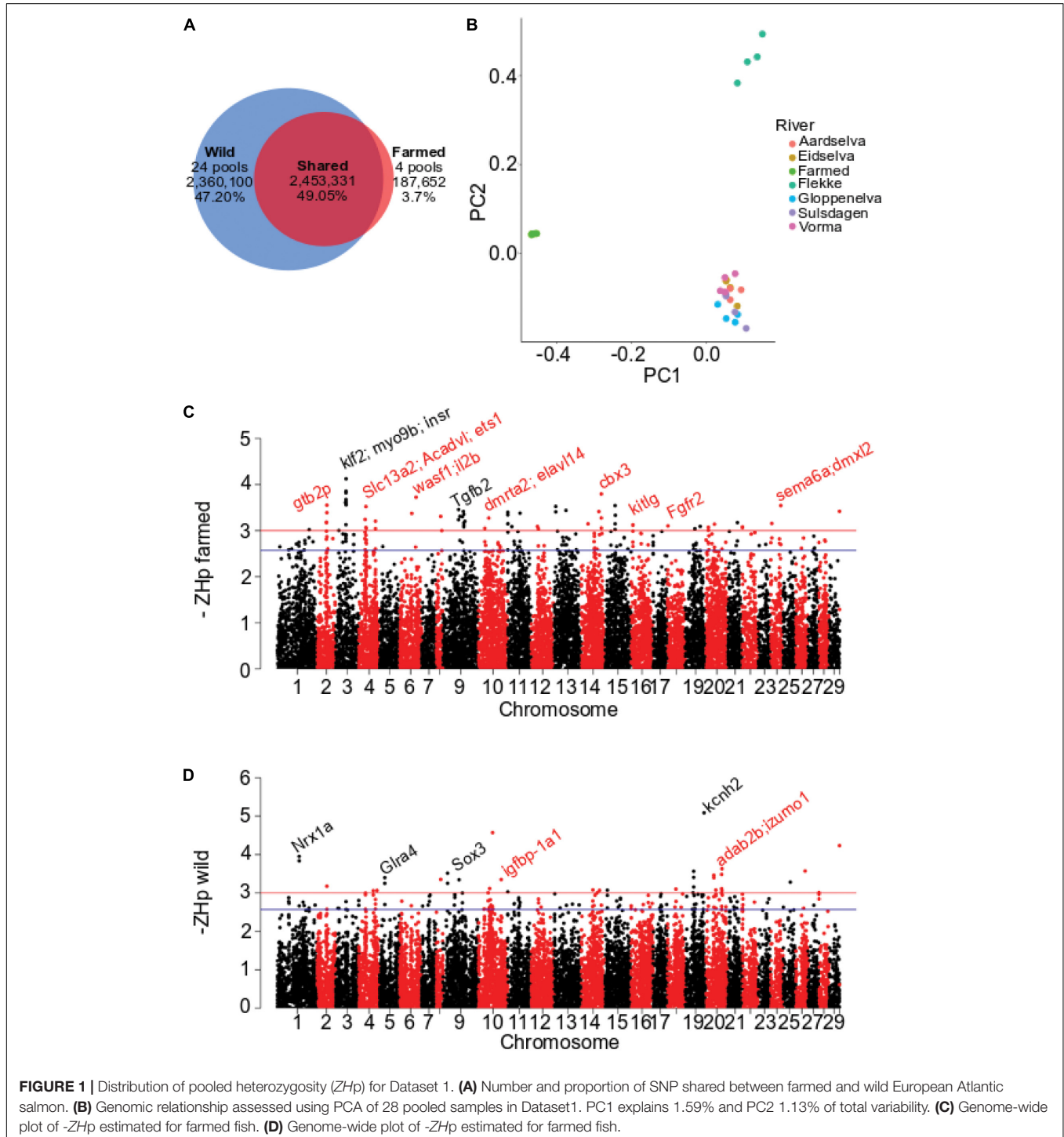
## RESULTS

### Genome Sequencing and SNP Calling

To commence analysis of domesticated Atlantic salmon, four pools of genomic DNA containing 22 animals per pool were each sequenced to 20 – 24 fold coverage (Dataset 1, Table 1). These farmed fish have founders originating from ancestral populations of both the Norwegian and Scottish Atlantic clade of wild Atlantic salmon. We therefore selected previously published whole genome sequence data from wild Norwegian populations for comparison (Ayllon et al., 2015) (Table 1 and Supplementary

**Table S1).** Variant calling across all pools identified 5,001,083 high-quality SNPs. Around half were independently identified in both the wild and domestic DNA pools (2.45 M or 49%, **Figure 1A**). A large proportion of the SNPs were found only in wild Atlantic salmon (2.36 M or 47%) while a comparatively small number (187,652 or 3.7%) were specific to farmed fish (**Figure 1A** and **Supplementary Figure S1**). Comparison against

the available protein coding gene annotation for Atlantic salmon (Lien et al., 2016) revealed less than 1% of variants were in exons (232,030). The majority of exonic SNPs were synonymous substitutions (116,228, **Supplementary Table S2**). Prior to assessing patterns of variation that characterized farmed and wild populations, we checked the genomic relationship between pools using PCA and allele frequency divergence using  $F_{ST}$  (**Figure 1B**





and **Supplementary Figure S2**). The pools of farmed fish clustered together and separately from wild populations using both PC1 and PC2 (**Figure 1B** and **Supplementary Figure S2**). The population wise  $F_{ST}$  values were consistent with the PCA analysis, with the highest values (0.052 – 0.073) found in pairwise comparison of farmed pools versus wild pools. The reduction in SNP frequency detected within the farmed pools, together with their divergence from wild populations, suggested Dataset 1 and the methods applied to identify genomic variation were well suited to subsequent analysis.

## Genomic Regions Putatively Under Selection Within Farmed or Wild Populations

Genomic regions impacted by positive selection are likely to display reduced heterozygosity in response to selection for beneficial alleles. To search for regions exhibiting depressed polymorphism, or an excess of homozygosity, pooled whole-genome sequence data (Dataset 1) was analyzed in 150 kb genomic windows using the  $H_p$  metric (Rubin et al., 2010, 2012) (**Supplementary Figure S3**). When applied to genome data from farmed fish, 71 windows were identified as outliers from the 29,614 tested genome wide ( $ZHp < -3$ ,  $p\text{-val} = 0.001$ ; **Figure 1C** and **Supplementary Table S3**). The windows contained a small number of genes previously identified by genome scans for selection or with interesting functional roles. A prominent example is *kitlg*, a target of selection in multiple livestock species associated with melanocyte migration and pigmentation changes (Wilkinson et al., 2013; Qanbari et al., 2014). The same  $ZHp$  analysis of wild populations identified only 37 outlier windows ( $ZHp < -3$ ,  $p\text{-val} = 0.001$ ) (**Figure 1D**). The collection of genes contained within the outlier windows is given in **Supplementary Table S4**. The total number of windows that displayed significantly elevated homozygosity (71 and 37) is low when compared to similar studies conducted in livestock species. For example, analysis for domestic dog (Axelsson et al., 2013), chicken (Rubin et al., 2010), and pig (Rubin et al., 2012) all revealed generally higher rates of outlier window detection with more extreme  $-ZHp$  values than observed here. This likely reflects the very recent domestication of Atlantic salmon, in comparison to domesticated livestock and companion animals that have vastly different population histories characterized by much longer periods of artificial selection following domestication events that may have commenced more than 10,000 years ago (Larson et al., 2005).

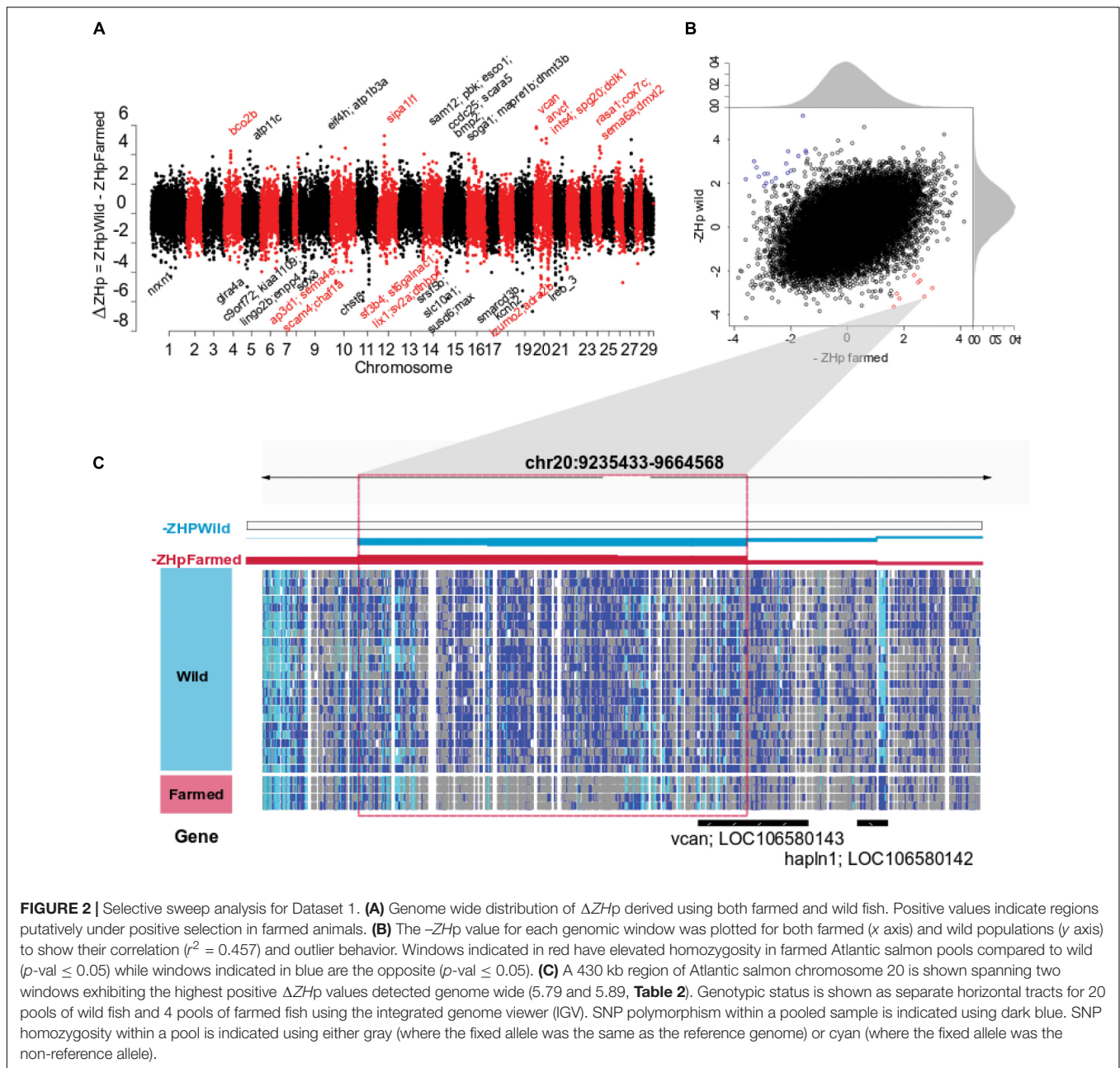
## Selection Sweeps Identified by Comparison Between Domestic With Wild Populations

We sought to identify regions undergoing selection in a population specific manner by searching for regions with a marked difference in heterozygosity ( $\Delta ZHp$ ) between wild and farmed populations ( $\Delta ZHp = ZHp_{\text{Wild}} - ZHp_{\text{Farmed}}$ ). Windows with increasingly positive  $\Delta ZHp$  values exhibit elevated homozygosity in farmed fish while maintaining heterozygosity in wild populations, consistent with positive

selection in response to domestication (**Figure 2**). This approach identified surprisingly few regions as  $ZHp_{\text{Wild}}$  and  $ZHp_{\text{Farmed}}$  values were significantly positively correlated ( $r^2 = 0.457$ ,  $p\text{-val} < 2.2 \cdot 10^{-16}$ , **Figure 2B**). A total of 139 regions had  $\Delta ZHp > 3$  (**Figure 2A** and **Supplementary Table S5**). After adjustment for multiple testing ( $\Delta ZHp \geq 4.55$ ,  $p\text{-adj} 0.05$ ), only 8 windows were identified showing a strong reduction in heterozygosity in farmed fish compared with their wild counterparts (**Table 2**). The most extreme window ( $\Delta ZHp = 5.89$ ) contained *Versican* (*vcan*), a gene involved in the control of neural crest cell migration (Dutt et al., 2006). This is of particular interest given a broad range of neural crest cell regulatory genes have demonstrated roles in domestication related traits in other animals (Wilkinson et al., 2013; Pendleton et al., 2018). The sweep region is located upstream of *vcan* where each of the four pools of farmed fish have sharply reduced heterozygosity across a region spanning  $> 100$  kb (**Figure 2C**). The sweep is noticeably absent in any of the 20 pools evaluated containing wild fish, giving rise to the most extreme outlier region genome wide. A further 131 regions had  $\Delta ZHp < -3$  and are indicative of sweeps specific to the wild populations (**Supplementary Table S6**).

## Sweep Regions Assessed Using an Independent Dataset

The putative selection sweeps identified using Dataset 1 provide an interesting basis for discussion, but verification within independent populations are required to provide strong evidence that they are the result of domestication. The wild populations assayed may not be strongly enriched for ancestor fish recruited to initiate the farmed populations, or the pool-seq approach may introduce bias, each leading to the potential for the detection of false positive selection sweeps. Therefore, we repeated aspects of the analysis using Dataset 2, an independent collection of individually sequenced farmed and wild European derived Atlantic salmon (Dataset 2, **Table 1**). Variant calling for Dataset 2 yielded 2,251,066 SNPs, before genome-wide  $ZHp$  scores were calculated as performed for Dataset 1. Comparative analysis revealed the collection of outlier windows identified in Dataset 1 (139 genomic windows with  $\Delta ZHp \geq 3$ ,  $p\text{-val} = 0.001$ ) displayed increased homozygosity compared with their genome-wide mean within the commercially farmed salmon in Dataset 2 (Wilcoxon test,  $p\text{-val} = 4.038 \times 10^{-5}$ ). Importantly, this was not the case within the wild salmon individuals in Dataset 2 (**Figure 3A**). This strongly suggested that the sweeps identified using Dataset 1 are enriched for true selection sweeps and are unlikely to be entirely due to drift or the result of the populations used in the genome wide comparisons. We next sought to identify the number and distribution of genome windows that displayed significantly positive  $\Delta ZHp$  values within the farmed fish populations in both Datasets 1 and 2 (**Figure 3B**). This yielded 81 regions ( $p\text{-adj} < 0.05$ ) with significantly reduced heterozygosity in farmed salmon compared to wild fish in both datasets (**Supplementary Table S7**). Investigation of the genes contained in these windows revealed clear relevance to domestication and/or genes with established roles in phenotypic



variation. At least three genes are related to neurogenesis and neural function suggesting a relationship with behavioral traits (*arvcf*, *sema6a*, and *erbb4*; **Figures 2, 3** and **Supplementary Table S7**). Others relate to neural crest cell migration (*txbl-a*) and olfactory mechanisms (*id2-like*, *6nl-like*), both of which have plausible roles in processes associated with domestication and adaptation to a captive environment.

## Sweep Regions in North American Derived Farmed Atlantic Salmon

The final component of the analysis sought to identify evidence for parallel selection sweeps founded independently within

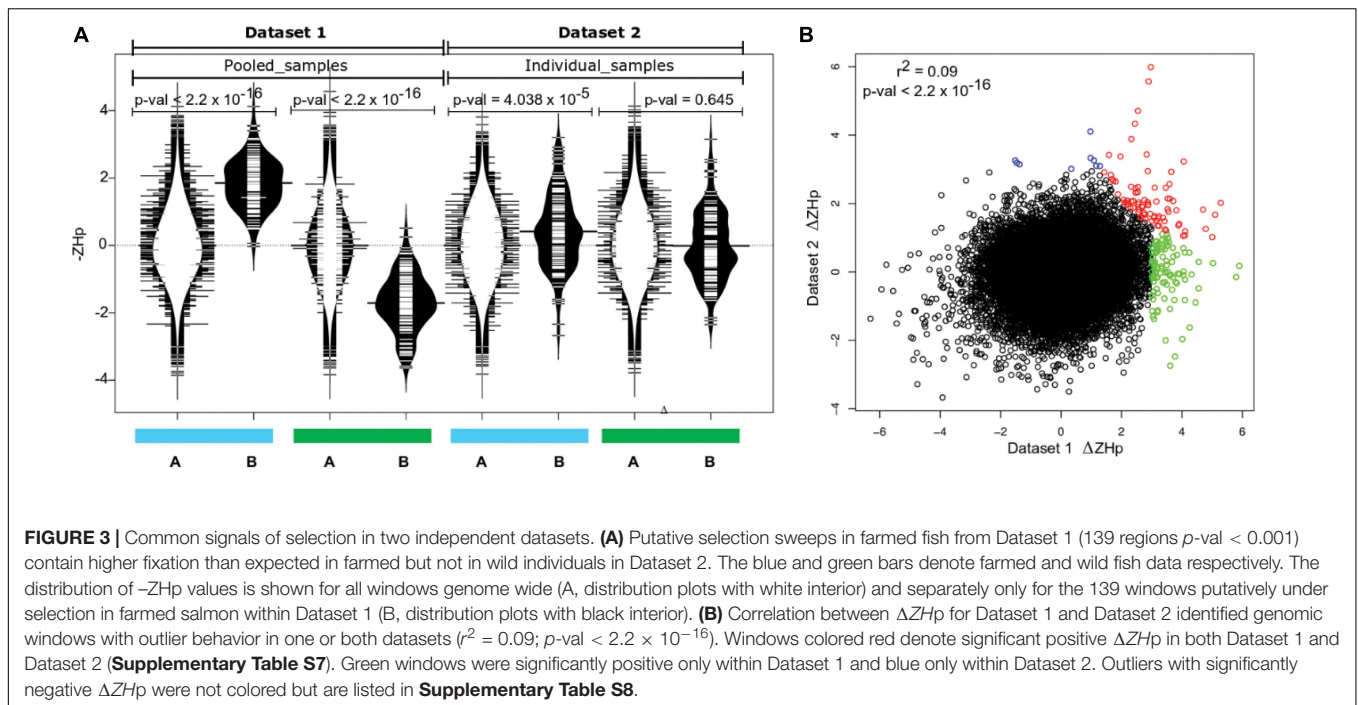
farmed stocks established from both European and North American wild populations. Dataset 3 comprised 19 genomes of farmed fish from Tasmania that originate from Canadian stocks (**Table 1**). Variant calling yielded 7,623,909 high quality SNPs before heterozygosity was assessed as  $ZHp$  and plotted as a function of genomic location (**Figure 4A**). Few genomic regions had evidence for either mild (79 windows,  $-ZHp > 2.57$ ;  $p\text{-val} < 0.01$ ) or strong increases in homozygosity (9 windows,  $-ZHp > 3$ ;  $p\text{-val} < 0.001$ , **Table 3**). The number of identified outliers was lower than detected within the European derived farmed population in Dataset 1 (**Figure 4A**). The most extreme outlier window was found on Ssa15, and spanned a region containing more than 10 genes making it difficult to identify the

**TABLE 2** | Putative selective sweeps detected in farmed Atlantic salmon (Dataset 1).

chr	Start	End	ZHp <sub>dom</sub>	ZHp <sub>wild</sub>	Δ ZHp	Alias	Genes product description	Function	References	Other species
20	9375001	9525000	-2.72	3.18	5.89	Vcan	LOC106580143 versican core protein-like	Cell adhesion, proliferation, migration. Neural crest cells migration. Wound healing	Sotoodehnejadnematalahi and Burke, 2013	<b>Great apes</b> (Cagan et al., 2016) <b>Human</b> (Pääbo, 2014)
20	9300001	9450000	-3.00	2.79	5.79	-	-			
<b>12</b>	<b>27975001</b>	<b>28125000</b>	<b>-1.64</b>	<b>3.64</b>	<b>5.28</b>	<b>Sipa111</b>	<b>LOC106564813 uncharacterized LOC106564812 signal-induced proliferation-associated 1-like protein</b>	<b>Contributes to the regulation of dendritic spine morphogenesis</b>	Maruoka et al., 2005; Chutabhakdikul and Surakul, 2013	<b>+ Sheep</b> (Fariello et al., 2014) <b>Human</b> (Racimo, 2016; Peyrégne et al., 2017)
<b>20</b>	<b>53625001</b>	<b>53775000</b>	<b>-1.84</b>	<b>3.26</b>	<b>5.10</b>	-	<b>LOC106580979 LOC106580980 LOC106580981 olfactory receptor 1D2-like LOC106580982 olfactory receptor 6N1-like</b>	<b>Neuronal response that triggers the perception of a smell</b>		
27	26325001	26475000	-2.50	2.53	5.02	-	-			
<b>20</b>	<b>32775001</b>	<b>32925000</b>	<b>-2.57</b>	<b>2.44</b>	<b>5.00</b>	<b>arvcf</b>	<b>LOC106580542 LOC106580487 armadillo repeat protein deleted in velo-cardio-facial syndrome-like</b>	<b>Neurocognitive and neuroanatomical, craniofacial morphology</b>	Cho et al., 2011; Tran et al., 2011; Sim et al., 2012	<b>+</b> <b>Dog breeds</b> (Persson et al., 2016)
<b>20</b>	<b>53700001</b>	<b>53850000</b>	<b>-1.74</b>	<b>3.04</b>	<b>4.79</b>	<b>gsonmt000 21581001 gsonmt 00046282001 ints4 spg20 dckl1 panx1</b>	<b>LOC106580981 olfactory receptor 1D2-like; LOC106580982 olfactory receptor 6N1-like; LOC106580972; LOC106580980; LOC106580973 40S ribosomal protein S3-like; LOC106580983 mth938 domain-containing protein-like; LOC106580975; LOC106580974 spartin-like%2C transcript variant; LOC106580976 serine/threonine-protein kinase DCLK1-like%2C transcript variant</b>	<b>Spg20; synaptic growth and neuronal survival, linked to spastic paraplegia DCLK1: neuronal migration, neurogenesis Panx1: inflammation; myoblast differentiation; neural precursor and maintenance</b>	Nahm et al., 2013; Butler et al., 2016; Dardour et al., 2017 Nawabi et al., 2015; Wicki-Stordeur et al., 2016; Langlois and Cowan, 2017	<b>+</b> <b>(* DCX:</b> (Huang et al., 2015)
9	95175001	95325000	-2.36	2.20	4.55	Eif4h gsonmt 00055384001 atp1b3a	if4h Eukaryotic translation initiation factor 4H LOC106612405 E3 ubiquitin-protein ligase rifylin-like LOC106612413 sodium/potassium-transporting ATPase subunit beta-3-like	Atp1b3:osmoregulation		

150 kb regions chr, start, end, ZHpvalue in Atlantic salmon farmed pools, ZHp scores in Atlantic salmon wild pools, ΔZHpPools= ZHp<sub>wild</sub> - ZHp<sub>Farmed</sub>, Overlapping gene Alias as from SalmoBase Gene Browser track, Overlapping Genes product LOC identifier and gene product description. Regions in bold confirmed in an independent farmed and wild set (Dataset 2).



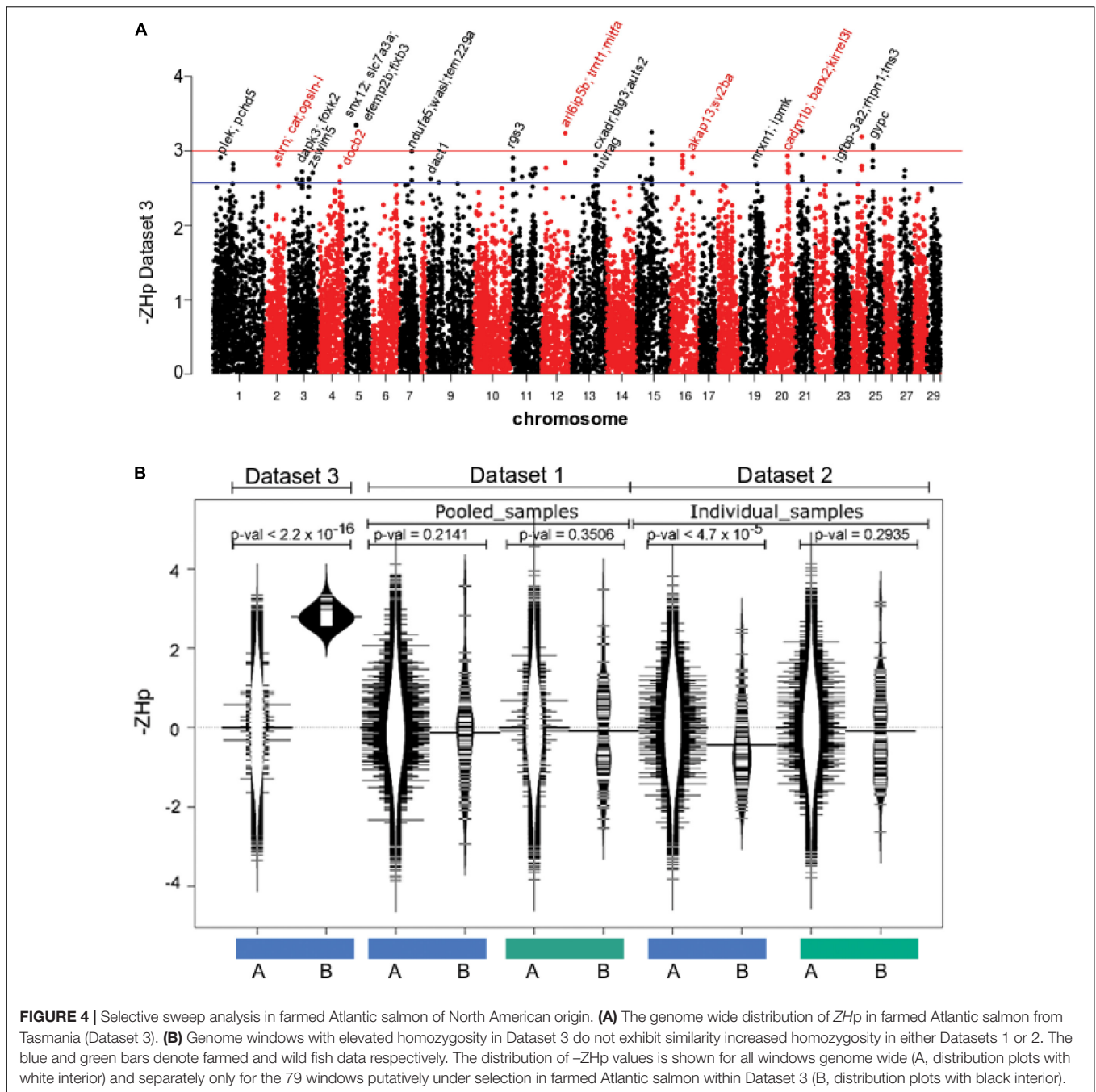


likely target(s) of selection. We assessed if the small collection of outlier regions identified from Atlantic salmon in Tasmania (79 windows with  $-ZHp > 2.57$ ) showed evidence for increased homozygosity within the farmed European fish in either Datasets 1 or 2. No evidence was present (**Figure 4B**), suggesting little or no overlap appears to be present between putative sweep regions found in European and North American derived farmed populations. On the basis of the analysis performed here, parallel selection events do not appear to be a significant factor shaping patterns of genetic variability.

## DISCUSSION

The primary objective of the study was to deepen our understanding of how the captive farming environment and artificial selection have impacted patterns of variation across the genome of Atlantic salmon. A number of previous studies have addressed this question, resulting in the identification of largely non-overlapping genomic regions and gene sets (Bourret et al., 2011; Vasemägi et al., 2012; Martinez et al., 2013; Mäkinen et al., 2015; López et al., 2019b). We designed this experiment to be differentiated from earlier work in at least two important ways. First, we exploited the availability of whole genome data which holds the promise of increased precision to detect sweeps over low and medium-density SNP arrays. Genome sequencing data also opens the possibility of detecting functional variants that directly underpin trait variation. Secondly, the experimental design incorporated analysis of multiple independent populations to address the issue of false positive sweep signals. This was important given the farmed and wild population comparisons used were not

optimal. Specifically, ancestors of the farmed fish were not sourced from the rivers sampled to generate the wild fish genome data. This leaves open the possibility that phylogenetic structure may contribute to the observed patterns of SNP variation in addition to selection following domestication. The finding that outlier regions with reduced heterozygosity in Dataset 1 also showed significantly lower heterozygosity in Dataset 2 provided confidence the shared regions have biological validity. Analysis of the genes present in putative sweep regions revealed a number have established roles relevant to domestication, with functions relating to brain function and behavioral traits prominent. This is consistent with findings from other species such as rabbit which have a comparatively short domestication history compared with most livestock species (Carneiro et al., 2014). Reduction in fear and changes to social behavior are cases in point (Jensen, 2014). In this study, *arvcf* was identified in a sweep region on *Ssa20* in both Dataset 1 and 2. The gene was previously associated with dog sociability to humans (Persson et al., 2016), while a paralog of *dclk1* (doublecortin like) termed *dcx* (doublecortin) was found in a separate sweep on chromosome 20 that has been associated with tameness in foxes (Huang et al., 2015). A selective sweep on *Ssa24* spans *sema6a* that is under selection in domestic cattle (Qanbari et al., 2014). It has been directly associated with exploration behavior, with knockout mice expressing marked behavioral differences (Håkansson et al., 2017). A recent study has also identified the related gene *sema6b* as a candidate underlying selection in farmed Atlantic salmon (López et al., 2019b). It has a major role in axon guidance and both peripheral and central nervous system development. A fourth behavioral gene, *erbb4*, was identified on *Ssa21* that has evidence of selection in domestic cattle (Qanbari et al., 2014). Expression changes in *erbb4* in mice regulated fear and mania-like



behaviors (hyperactivity, reduced anxiety and depression and increased sucrose preference) (Chen et al., 2017; Cao et al., 2018). Each of these findings suggest behavioral adaptations to the farmed environment, including altered response to threats, and differences in aggression and feeding behavior may be an early consequence of domesticating Atlantic salmon (Huntingford, 2004; Glover et al., 2017). Interestingly, a selective sweep on *Ssa20* contained two olfactory receptor genes (olfactory receptor *id2-like* and *6n1-like*) (Table 2). The olfactory system plays a key role in the 'homing' instinct of wild salmon (Nevitt and Dittman, 2004), and multiple studies have demonstrated that offspring of

domesticated Atlantic salmon display reduced survival in the marine phase of early life (McGinnity et al., 2003; Skaala et al., 2019). The homing instinct is no longer relevant in the farmed environment and may be associated with behaviors or physiology with negative consequences for survival or growth in culture. Further, olfactory receptors are associated with domestication selection in several other species, including dogs (Chen et al., 2012), cats (Montague et al., 2014), and pigs (Amaral et al., 2011). The gene *sipa11l* identified in a sweep region on *Ssa12*, which is involved in the regulation of dendritic spine morphogenesis, and therefore, a key role in synaptic process, has also been identified as

**TABLE 3** | Candidate Selective sweeps detected in North American Atlantic Salmon samples from Tasmania (Dataset 3).

chr	Start	End	ZHp	Alias	Genes product description
5	32325001	32475000	3.34	ubl3 at13 dipa fibp slc7a3a snx12 fosl2 prs23 efemp2b foxn3 cof2	ubl3; ubiquitin-like 3LOC106604707; serine protease 23-like%2C transcript variant LOC106604709; atlastin-3-likeLOC106604711; protein Y1F1A-like%2C transcript variant LOC106604713; coiled-coil domain-containing protein 85B-like%2C transcript variant LOC106604715; acidic fibroblast growth factor intracellular-binding protein-likesnx12; sorting nexin 12%2C transcript variant slc7a3; solute carrier family 7 (cationic amino acid transporter%2C y+ system)%2C member 3%2C transcript variant LOC106604708; cofilin-2-likeLOC106604710; forkhead box protein N2-like%2C transcript variant LOC106604712; fos-related antigen 1-like%2C transcript variant LOC106604716; EGF-containing fibulin-like extracellular matrix protein 2%2C transcript variant
21	17625001	17775000	3.26	i20ra orf2p i10r2 crfb2 btn1a1 crfb1a	20ra; Interleukin-20 receptor alpha chainLOC106581740; uncharacterized LOC106581740%2C transcript variant LOC106581741; interleukin-10 receptor subunit beta-likeLOC106581720; CD276 antigen homolog%2C transcript variant LOC106581739; uncharacterized protein C21orf62-like%2C transcript variant
15	45075001	45225000	3.25		LOC106571716; uncharacterized protein K02A2.6-likeLOC106571620; C-C chemokine receptor type 6-like
12	72000001	72150000	3.24	gsonmt00065759001 arl6ip5b trnt1 nft-vtr mitfa gsonmt00065604001	
24	30900001	31050000	3.19	cep78 naa35 gnaq vps13a agtpbp1 gna14	naa35; N(alpha)-acetyltransferase 35%2C NatC auxiliary subunit %2C transcript variant LOC106585634; guanine nucleotide-binding protein subunit alpha-14-likeLOC106585689; guanine nucleotide-binding protein G(q) subunit alphacep78; centrosomal protein 78 kDa %2C transcript variant golm1; golgi membrane protein 1vps13a; vacuolar protein sorting 13 homolog A ( <i>S. cerevisiae</i> ) %2C transcript variant agtpbp1; ATP/GTP binding protein 1%2C transcript variant
15	45150001	45300000	3.09		LOC106571620; C-C chemokine receptor type 6-like
25	16725001	16875000	3.07	gypc	LOC106586280; glycophorin-C-like%2C transcript variant
25	16800001	16950000	3.05	Unknown	LOC106586279; contactin-associated protein-like 5
25	17100001	17250000	3.03	gsonmt00061935001	

a gene potentially subjected to domestication and selection in two independent studies in Atlantic salmon (López et al., 2019a,b). The final gene of interest is versican (*vcan*), which displayed the most extreme outlier behavior in Dataset 1. The gene acts to control neural crest cell migration (Landolt et al., 1995; Dutt et al., 2006), a fundamental developmental process whereby multipotent stem cells emerge from the neural crest before differentiating into a wide variety of cell types and physiology. They play a central role in body conformation, pigmentation, neuronal changes and craniofacial development, each of which are often impacted in animals by domestication. A recent study that compared village dog and gray wolf genomes illustrates this well, as a range of genes were implicated in domestication that have clear roles in neural crest cell pathways and their control (Pendleton et al., 2018). These findings have given rise to the hypothesis that neural crest cell migration is a key mechanism by which domestication acts to alter multiple animal characteristics (Wilkinson et al., 2013). Our results indicate this may also be the case in Atlantic salmon, although it remains unclear by what mechanism. It is also noteworthy *vcan* was significantly associated to human lean body mass in meta-analysis of more than sixty thousand individuals (Zillikens et al., 2017), suggesting

it may have undergone change as farmed Atlantic salmon are selectively bred for improved growth rate and weight.

Independent domestication events, whereby farmed fish stocks were founded from either European or North American wild stocks, offered the opportunity to search for evidence of parallel selection sweeps. It is important to note that earlier analysis of genome sequence comparing European and North America Atlantic salmon revealed them to be highly genetically divergent (Kijas et al., 2017). Comparing outlier regions in this study (from Dataset3 with those from either Datasets 1 or 2) failed to detect an overlap. This is consistent with the latest findings on parallel evolution in Atlantic salmon from López et al. (2019a). The authors used a 200K SNP array and found only four regions of overlap between populations with North American and European origins. This suggests parallel genetic changes resulting from domestication are rare, and our results in this study obtained using whole genome sequencing datasets provide additional evidence. This may arise due to differences in nutrition, management or environmental conditions. Alternatively, largely distinct sets of genes may have undergone change between populations and result in similar phenotypic changes to complex traits. In addition, it is worth

noting that unlike European farmed Atlantic salmon populations under active selection for economic traits since the 1970s, family selection commenced for the Tasmanian population in 2005. Even where artificial selection has acted to promote a trait in common, for example growth rate, the growing body of evidence suggest selection has acted on different subsets of genes.

## DATA AVAILABILITY STATEMENT

The datasets generated for this study can be found in NCBI PRJNA293012, PRJEB10744, SRP059652, PRJNA403334, and PRJNA614520.

## ETHICS STATEMENT

Ethical review and approval was not required for the animal study because Animals used in this study were part of the commercial operations of Salmon Enterprises of Tasmania Pty. Ltd. Their use was in accordance with authorized management practices of the company and compliant with the Tasmanian Animal Welfare Act (1993) which is under the jurisdiction of Biosecurity Tasmania, Department of Primary Industries, Parks, Water and Environment. Under this Act, those animals that are expressly killed for purposes other than research, such as abattoir specimens, do not need specific approval of an Animal Ethics

## REFERENCES

- Alberto, F. J., Boyer, F., Orozco-terWengel, P., Streeter, I., Servin, B., de Villemereuil, P., et al. (2018). Convergent genomic signatures of domestication in sheep and goats. *Nat. Commun.* 9:813. doi: 10.1038/s41467-018-03206-y
- Amaral, A. J., Ferretti, L., Megens, H.-J., Crooijmans, R. P. M. A., Nie, H., Ramos-Onsins, S. E., et al. (2011). Genome-wide footprints of pig domestication and selection revealed through massive parallel sequencing of pooled DNA. *PLoS One* 6:e14782. doi: 10.1371/journal.pone.0014782
- Axelsson, E., Ratnakumar, A., Arendt, M. L., Maqbool, K., Webster, M. T., Perloski, M., et al. (2013). The genomic signature of dog domestication reveals adaptation to a starch-rich diet. *Nature* 495, 360–364. doi: 10.1038/nature11837
- Ayllon, F., Kjærner-Semb, E., Furmanek, T., Wennevik, V., Solberg, M. F., Dahle, G., et al. (2015). The *vgl3* locus controls age at maturity in wild and domesticated atlantic salmon (*Salmo salar* L.) males. *PLoS Genet.* 11:e1005628. doi: 10.1371/journal.pgen.1005628
- Barson, N. J., Aykanat, T., Hindar, K., Baranski, M., Bolstad, G. H., Fiske, P., et al. (2015). Sex-dependent dominance at a single locus maintains variation in age at maturity in salmon. *Nature* 528, 405–408. doi: 10.1038/nature16062
- Bourret, V., O'Reilly, P. T., Carr, J. W., Berg, P. R., and Bernatchez, L. (2011). Temporal change in genetic integrity suggests loss of local adaptation in a wild Atlantic salmon (*Salmo salar*) population following introgression by farmed escapees. *Heredity* 106, 500–510. doi: 10.1038/hdy.2010.165
- Butler, S., Helbig, K. L., Alcaraz, W., Seaver, L. H., Hsieh, D. T., and Rohena, L. (2016). Three cases of Troyer syndrome in two families of Filipino descent. *Am. J. Med. Genet. A* 170, 1780–1785. doi: 10.1002/ajmg.a.37658
- Cagan, A., Theunert, C., Laayouni, H., Santpere, G., Pybus, M., Casals, F., et al. (2016). Natural selection in the great apes. *Mol. Biol. Evol.* 33, 3268–3283.
- Cao, S.-X., Zhang, Y., Hu, X.-Y., Hong, B., Sun, P., He, H.-Y., et al. (2018). ErbB4 deletion in noradrenergic neurons in the locus coeruleus induces mania-like behavior via elevated catecholamines. *eLife* 7:e39907.
- Carneiro, M., Rubin, C. J., Di Palma, F., Albert, F. W., Alfvöldi, J., Martinez Barrio, A., et al. (2014). Rabbit genome analysis reveals a polygenic basis for phenotypic

change during domestication. *Science* 345, 1074–1079. doi: 10.1126/science.1253714

Chang, C. C., Chow, C. C., Tellier, L. C., Vattikuti, S., Purcell, S. M., and Lee, J. J. (2015). Second-generation PLINK: rising to the challenge of larger and richer datasets. *Gigascience* 4:7. doi: 10.1186/s13742-015-0047-8

Chen, R., Irwin, D. M., and Zhang, Y.-P. (2012). Differences in selection drive olfactory receptor genes in different directions in dogs and wolf. *Mol. Biol. Evol.* 29, 3475–3484. doi: 10.1093/molbev/mss153

Chen, Y.-H., Lan, Y.-J., Zhang, S.-R., Li, W.-P., Luo, Z.-Y., Lin, S., et al. (2017). ErbB4 signaling in the prefrontal cortex regulates fear expression. *Transl. Psychiatry* 7:e1168. doi: 10.1038/tp.2017.139

Cho, K., Lee, M., Gu, D., Munoz, W. A., Ji, H., Kloc, M., et al. (2011). Kazrin, and its binding partners ARVCF- and delta-catenin, are required for *Xenopus laevis* craniofacial development. *Dev. Dyn. Off. Publ. Am. Assoc. Anat.* 240, 2601–2612. doi: 10.1002/dvdy.22721

Chutabhakdikul, N., and Surakul, P. (2013). Prenatal stress increased Snk Polo-like kinase 2, SCF  $\beta$ -TrCP ubiquitin ligase and ubiquitination of SPAR in the hippocampus of the offspring at adulthood. *Int. J. Dev. Neurosci. Off. J. Int. Soc. Dev. Neurosci.* 31, 560–567. doi: 10.1016/j.ijdevneu.2013.06.011

Cockerham, C. C., and Weir, B. S. (1984). Covariances of relatives stemming from a population undergoing mixed self and random mating. *Biometrics* 40, 157–164.

Danecek, P., Auton, A., Abecasis, G., Albers, C. A., Banks, E., DePristo, M. A., et al. (2011). The variant call format and VCFtools. *Bioinform. Oxf. Engl.* 27, 2156–2158. doi: 10.1093/bioinformatics/btr330

Dardour, L., Roelens, F., Race, V., Souche, E., Holvoet, M., and Devriendt, K. (2017). SPG20 mutation in three siblings with familial hereditary spastic paraplegia. *Cold Spring Harb. Mol. Case Stud.* 3:a001537. doi: 10.1101/mcs.a001537

DePristo, M. A., Banks, E., Poplin, R., Garimella, K. V., Maguire, J. R., Hartl, C., et al. (2011). A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nat. Genet.* 43, 491–498. doi: 10.1038/ng.806

## AUTHOR CONTRIBUTIONS

MN-S, RH, and JK conceived the study and contributed to the experimental design. RH and BE contributed to animal samples, genome data or both. MN-S, SM, and JK performed the analysis. MN-S and JK wrote the manuscript. All authors contributed to the final version.

## FUNDING

MN-S was funded by the CSIRO Science Excellence Research Office. RH receives funding from BBSRC Institute Strategic Program Grants (BB/P013759/1 and BB/P013740/1).

## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fgene.2020.00264/full#supplementary-material>



- Dutt, S., Kléber, M., Matasci, M., Sommer, L., and Zimmermann, D. R. (2006). Versican V0 and V1 guide migratory neural crest cells. *J. Biol. Chem.* 281, 12123–12131. doi: 10.1074/jbc.m510834200
- Fariello, M.-I., Servin, B., Tosser-Klopp, G., Rupp, R., Moreno, C., International Sheep Genomics Consortium, et al. (2014). Selection signatures in worldwide sheep populations. *PLoS One* 9:e103813. doi: 10.1371/journal.pone.0103813
- Fleming, I. A., Hindar, K., Mjølnerød, I. B., Jonsson, B., Balstad, T., and Lamberg, A. (2000). Lifetime success and interactions of farm salmon invading a native population. *Proc. Biol. Sci.* 267, 1517–1523. doi: 10.1098/rspb.2000.1173
- Frantz, L. A., Schraiber, J. G., Madsen, O., Megens, H. J., Cagan, A., Bosse, M., et al. (2015). Evidence of long-term gene flow and selection during domestication from analyses of Eurasian wild and domestic pig genomes. *Nat. Genet.* 47, 1141–1148. doi: 10.1038/ng.3394
- Gjedrem, T., Robinson, N., and Rye, M. (2012). The importance of selective breeding in aquaculture to meet future demands for animal protein: a review. *Aquaculture* 350–353, 117–129. doi: 10.1016/j.aquaculture.2012.04.008
- Glover, K. A., Otterå, H., Olsen, R. E., Slinde, E., Taranger, G. L., and Skaala, Ø (2009). A comparison of farmed, wild and hybrid Atlantic salmon (*Salmo salar* L.) reared under farming conditions. *Aquaculture* 286, 203–210. doi: 10.1016/j.aquaculture.2008.09.023
- Glover, K. A., Pertoldi, C., Besnier, F., Wennevik, V., Kent, M., and Skaala, Ø (2013). Atlantic salmon populations invaded by farmed escapees: quantifying genetic introgression with a Bayesian approach and SNPs. *BMC Genet.* 14:74. doi: 10.1186/1471-2156-14-74
- Glover, K. A., Solberg, M. F., McGinnity, P., Hindar, K., Verspoor, E., Coulson, M. W., et al. (2017). Half a century of genetic interaction between farmed and wild Atlantic salmon: status of knowledge and unanswered questions. *Fish Fish.* 18, 890–927. doi: 10.1111/faf.12214
- Gutierrez, A. P., Lubieniecki, K. P., Fukui, S., Withler, R. E., Swift, B., and Davidson, W. S. (2014). Detection of quantitative trait loci (QTL) related to grilising and late sexual maturation in Atlantic salmon (*Salmo salar*). *Mar. Biotechnol. N. Y.* 16, 103–110. doi: 10.1007/s10126-013-9530-3
- Gutierrez, A. P., Yáñez, J. M., and Davidson, W. S. (2016). Evidence of recent signatures of selection during domestication in an Atlantic salmon population. *Mar. Genomics* 26, 41–50. doi: 10.1016/j.margen.2015.12.007
- Håkansson, K., Runker, A. E., O'Sullivan, G. J., Mitchell, K. J., Waddington, J. L., and O'Tuathaigh, C. M. P. (2017). Semaphorin 6A knockout mice display abnormalities across ethologically-based topographies of exploration and in motor learning. *Neurosci. Lett.* 641, 70–76. doi: 10.1016/j.neulet.2017.01.043
- Huang, S., Slomianka, L., Farmer, A. J., Kharlamova, A. V., Gulevich, R. G., Herbeck, Y. E., et al. (2015). Selection for tameness, a key behavioral trait of domestication, increases adult hippocampal neurogenesis in foxes. *Hippocampus* 25, 963–975. doi: 10.1002/hipo.22420
- Huntingford, F. A. (2004). Implications of domestication and rearing conditions for the behaviour of cultivated fishes. *J. Fish Biol.* 65, 122–142. doi: 10.1111/j.0022-1112.2004.00562.x
- Jensen, P. (2014). Behavior genetics and the domestication of animals. *Annu. Rev. Anim. Biosci.* 2, 85–104. doi: 10.1146/annurev-animal-022513-114135
- Kijas, J., Elliot, N., Kube, P., Evans, B., Botwright, N., King, H., et al. (2017). Diversity and linkage disequilibrium in farmed Tasmanian Atlantic salmon. *Anim. Genet.* 48, 237–241. doi: 10.1111/age.12513
- Kijas, J., McWilliam, S., Naval Sanchez, M., Kube, P., King, H., Evans, B., et al. (2018). Evolution of sex determination *Loci* in Atlantic salmon. *Sci. Rep.* 8:5664. doi: 10.1038/s41598-018-23984-1
- Koenig, D., Jiménez-Gómez, J. M., Kimura, S., Fulop, D., Chitwood, D. H., Headland, L. R., et al. (2013). Comparative transcriptomics reveals patterns of selection in domesticated and wild tomato. *Proc. Natl. Acad. Sci. U.S.A.* 110, e2655–e2662. doi: 10.1073/pnas.1309606110
- Landolt, R. M., Vaughan, L., Winterhalter, K. H., and Zimmermann, D. R. (1995). Versican is selectively expressed in embryonic tissues that act as barriers to neural crest cell migration and axon outgrowth. *Development* 121, 2303–2312.
- Langlois, S., and Cowan, K. N. (2017). Regulation of skeletal muscle myoblast differentiation and proliferation by pannexins. *Adv. Exp. Med. Biol.* 925, 57–73. doi: 10.1007/5584\_2016\_53
- Larson, G., Dobney, K., Albarella, U., Fang, M., Matisoo-Smith, E., Robins, J., et al. (2005). Worldwide phylogeography of wild boar reveals multiple centers of pig domestication. *Science* 307, 1618–1621. doi: 10.1126/science.1106927
- Lemmon, Z. H., Bukowski, R., Sun, Q., and Doebley, J. F. (2014). The role of cis regulatory evolution in maize domestication. *PLoS Genet.* 10:e1004745. doi: 10.1371/journal.pgen.1004745
- Li, H., and Durbin, R. (2009). Fast and accurate short read alignment with Burrows–Wheeler transform. *Bioinformatics* 25:1754. doi: 10.1093/bioinformatics/btp324
- Lien, S., Koop, B. F., Sandve, S. R., Miller, J. R., Kent, M. P., Nome, T., et al. (2016). The Atlantic salmon genome provides insights into rediploidization. *Nature* 533, 200–205. doi: 10.1038/nature17164
- Liu, L., Ang, K. P., Elliot, J. A. K., Kent, M. P., Lien, S., MacDonald, D., et al. (2017). A genome scan for selection signatures comparing farmed Atlantic salmon with two wild populations: Testing colocalization among outlier markers, candidate genes, and quantitative trait loci for production traits. *Evol. Appl.* 10, 276–296. doi: 10.1111/eva.12450
- López, M. E., Benestan, L., Moore, J.-S., Perrier, C., Gilbey, J., Genova, A. D., et al. (2019a). Comparing genomic signatures of domestication in two Atlantic salmon (*Salmo salar* L.) populations with different geographical origins. *Evol. Appl.* 12, 137–156. doi: 10.1111/eva.12689
- López, M. E., Linderth, T., Norris, A., Lhorente, J. P., Neira, R., and Yáñez, J. M. (2019b). Multiple selection signatures in farmed Atlantic salmon adapted to different environments across hemispheres. *Front. Genet.* 10:901. doi: 10.3389/fgene.2019.00901
- Luikart, G., England, P. R., Tallmon, D., Jordan, S., and Taberlet, P. (2003). The power and promise of population genomics: from genotyping to genome typing. *Nat. Rev. Genet.* 12, 981–994. doi: 10.1038/nrg1226
- Mäkinen, H., Vasemägi, A., McGinnity, P., Cross, T. F., and Primmer, C. R. (2015). Population genomic analyses of early-phase Atlantic salmon (*Salmo salar*) domestication/captive breeding. *Evol. Appl.* 8, 93–107. doi: 10.1111/eva.12230
- Martinez, V., Dettleff, P., Lopez, P., Fernandez, G., Jedlicki, A., Yáñez, J. M., et al. (2013). Assessing footprints of selection in commercial Atlantic salmon populations using microsatellite data. *Anim. Genet.* 44, 223–226. doi: 10.1111/j.1365-2052.2012.02387.x
- Maruoka, H., Konno, D., Hori, K., and Sobue, K. (2005). Collaboration of PSD-Zip70 with its binding partner, SPAR, in dendritic spine maturity. *J. Neurosci. Off. J. Soc. Neurosci.* 25, 1421–1430. doi: 10.1523/jneurosci.3920-04.2005
- McGinnity, P., Prodöhl, P., Ferguson, A., Hynes, R., Maoiléidigh, N. O., Baker, N., et al. (2003). Fitness reduction and potential extinction of wild populations of Atlantic salmon, *Salmo salar*, as a result of interactions with escaped farm salmon. *Proc. Biol. Sci.* 270, 2443–2450. doi: 10.1098/rspb.2003.2520
- Montague, M. J., Li, G., Gandolfi, B., Khan, R., Aken, B. L., Searle, S. M. J., et al. (2014). Comparative analysis of the domestic cat genome reveals genetic signatures underlying feline biology and domestication. *Proc. Natl. Acad. Sci. U.S.A.* 111, 17230–17235. doi: 10.1073/pnas.1410083111
- Nahm, M., Lee, M.-J., Parkinson, W., Lee, M., Kim, H., Kim, Y.-J., et al. (2013). Spartin regulates synaptic growth and neuronal survival by inhibiting BMP-mediated microtubule stabilization. *Neuron* 77, 680–695. doi: 10.1016/j.neuron.2012.12.015
- Naval-Sanchez, M., Nguyen, Q., McWilliam, S., Porto-Neto, L. R., Tellam, R., Vuocolo, T., et al., (2018). Sheep genome functional annotation reveals proximal regulatory elements contributed to the evolution of modern breeds. *Nat. Commun.* 9:859. doi: 10.1038/s41467-017-02809-1
- Nawabi, H., Belin, S., Cartoni, R., Williams, P. R., Wang, C., Latremolière, A., et al. (2015). Doublecortin-like kinases promote neuronal survival and induce growth cone reformation via distinct mechanisms. *Neuron* 88, 704–719. doi: 10.1016/j.neuron.2015.10.005
- Neveit, G., and Dittman, A. (2004). “Olfactory imprinting in salmon: new models and approaches,” in *The Senses of Fish*, eds G. von der Emde, J. Mogdans, B.G. Kapoor (Dordrecht: Springer), 109–127. doi: 10.1007/978-94-007-1060-3\_5
- Pääbo, S. (2014). The human condition—a molecular approach. *Cell* 157, 216–226. doi: 10.1016/j.cell.2013.12.036
- Pendleton, A. L., Shen, F., Taravella, A. M., Emery, S., Veeramah, K. R., Boyko, A. R., et al. (2018). Comparison of village dog and wolf genomes highlights the role of the neural crest in dog domestication. *BMC Biol.* 16:64. doi: 10.1186/s12915-018-0535-2
- Persson, M. E., Wright, D., Roth, L. S. V., Batakis, P., and Jensen, P. (2016). Genomic regions associated with interspecies communication in dogs contain genes related to human social disorders. *Sci. Rep.* 6:33439. doi: 10.1038/srep33439

- Peyrégne, S., Boyle, M. J., Dannemann, M., and Prüfer, K. (2017). Detecting ancient positive selection in humans using extended lineage sorting. *Genome Res.* 27, 1563–1572. doi: 10.1101/gr.219493.116
- Qanbari, S., Pausch, H., Jansen, S., Somel, M., Strom, T. M., Fries, R., et al. (2014). Classic selective sweeps revealed by massive sequencing in cattle. *PLoS Genet.* 10:e1004148. doi: 10.1371/journal.pgen.1004148
- Racimo, F. (2016). Testing for ancient selection using cross-population allele frequency differentiation. *Genetics* 202, 733–750. doi: 10.1534/genetics.115.178095
- Rubin, C.-J., Megens, H.-J., Barrio, M. A., Maqbool, K., Sayyab, S., Schwochow, D., et al. (2012). Strong signatures of selection in the domestic pig genome. *Proc. Natl. Acad. Sci. U.S.A.* 109, 19529–19536. doi: 10.1073/pnas.1217149109
- Rubin, C.-J., Zody, M. C., Eriksson, J., Meadows, J. R. S., Sherwood, E., Webster, M. T., et al. (2010). Whole-genome resequencing reveals loci under selection during chicken domestication. *Nature* 464, 587–591. doi: 10.1038/nature08832
- Sim, K., Chan, W.-Y., Woon, P.-S., Low, H.-Q., Lim, L., Yang, G.-L., et al. (2012). ARVCF genetic influences on neurocognitive and neuroanatomical intermediate phenotypes in Chinese patients with schizophrenia. *J. Clin. Psychiatry* 73, 320–326. doi: 10.4088/JCP.10m06491
- Skaala, Ø, Besnier, F., Borgström, R., Barlaup, B., Sørvik, A. G., Normann, E., et al. (2019). An extensive common-garden study with domesticated and wild Atlantic salmon in the wild reveals impact on smolt production and shifts in fitness traits. *Evol. Appl.* 12, 1001–1016. doi: 10.1111/eva.12777
- Skaala, Ø, Glover, K. A., Barlaup, B., Svasand, T., Besnier, F., Hansen, M. M., et al. (2012). Performance of farmed, hybrid, and wild Atlantic salmon (*Salmo salar*) families in a natural river environment. *Can. J. Fish Aquat. Sci.* 69, 1994–2006. doi: 10.1139/f2012-118
- Solberg, M. F., Skaala, Ø, Nilsen, F., and Glover, K. A. (2013). Does domestication cause changes in growth reaction norms? A study of farmed, wild and hybrid Atlantic salmon families exposed to environmental stress. *PLoS One* 8:e54469. doi: 10.1371/journal.pone.0054469
- Sotoodehnejadnematlahi, F., and Burke, B. (2013). Structure, function and regulation of versican: the most abundant type of proteoglycan in the extracellular matrix. *Acta Med. Iran* 51, 740–750.
- Tran, H. T., Delvaeye, M., Verschuere, V., Descamps, E., Crabbe, E., Van Hoorebeke, L., et al. (2011). ARVCF depletion cooperates with Tbx1 deficiency in the development of 22q11.2DS-like phenotypes in *Xenopus*. *Dev. Dyn. Off. Publ. Am. Assoc. Anat.* 240, 2680–2687. doi: 10.1002/dvdy.22765
- Vasemägi, A., Nilsson, J., McGinnity, P., Cross, T., O'Reilly, P., Glebe, B., et al. (2012). Screen for footprints of selection during domestication/captive breeding of Atlantic salmon. *Comp. Funct. Genomics* 2012:628204. doi: 10.1155/2012/628204
- Wang, G. D., Zhai, W., Yang, H. C., Fan, R. X., Cao, X., Zhong, L., et al. (2013). The genomics of selection in dogs and the parallel evolution between dogs and humans. *Nat. Commun.* 4:1860. doi: 10.1038/ncomms2814
- Wicki-Stordeur, L. E., Sanchez-Arias, J. C., Dhaliwal, J., Carmona-Wagner, E. O., Shestopalov, V. I., Lagace, D. C., et al. (2016). Pannexin 1 differentially affects neural precursor cell maintenance in the ventricular zone and perinatal cortex. *J. Neurosci. Off. J. Soc. Neurosci.* 36, 1203–1210. doi: 10.1523/JNEUROSCI.0436-15.2016
- Wilkinson, S., Lu, Z. H., Megens, H. J., Archibald, A. L., Haley, C., Jackson, I. J., et al. (2013). Signatures of diversifying selection in European pig breeds. *PLoS Genet.* 9:e1003453. doi: 10.1371/journal.pgen.1003453
- Wringe, B. F., Jeffery, N. W., Stanley, R. R. E., Hamilton, L. C., Anderson, E. C., Fleming, I. A., et al. (2018). Extensive hybridization following a large escape of domesticated Atlantic salmon in the Northwest Atlantic. *Commun. Biol.* 1:108. doi: 10.1038/s42003-018-0112-9
- Xiang, H., Gao, J., Yu, B., Zhou, H., Cai, D., Zhang, Y., et al. (2014). Early Holocene chicken domestication in northern China. *Proc. Natl. Acad. Sci. U.S.A.* 111, 17564–17569. doi: 10.1073/pnas.1411882111
- Yáñez, J. M., Naswa, S., López, M. E., Bassini, L., Correa, K., Gilbey, J., et al. (2016). Genomewide single nucleotide polymorphism discovery in Atlantic salmon (*Salmo salar*): validation in wild and farmed American and European populations. *Mol. Ecol. Resour.* 16, 1002–1011. doi: 10.1111/1755-0998.12503
- Zillikens, M. C., Demissie, S., Hsu, Y. H., Yerges-Armstrong, L. M., Chou, W. C., Stolk, L., et al. (2017). Large meta-analysis of genome-wide association studies identifies five loci for lean body mass. *Nat. Commun.* 8:80.

**Conflict of Interest:** BE was employed by Salmon Enterprises of Tasmania (Saltas) and Tassal.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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