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## Isolation and Culture of Murine Hepatic Stellate Cells

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**[Abstract]** Hepatic stellate cells (HSCs), alternatively known as liver pericytes, can differentiate into myofibroblasts and secrete extra-cellular matrix components, thereby promoting wound healing and fibrosis. Studying HSCs can provide insights into the pathological mechanisms governing these processes. HSC isolation methods typically comprise of enzymatic digestion followed by density gradient centrifugation and/or Fluorescent Activated Cell Sorting (FACS) mediated sorting. In this protocol, we describe a step-wise method for HSC isolation that utilizes Pronase and Collagenase for enzymatic tissue dissociation, followed by an Optiprep based density gradient centrifugation. The isolation can be performed using common media and buffers, and without the use of any special equipment for liver perfusion and HSC isolation. The technique yields *ex vivo* HSCs, suitable for use in assays.

**Keywords:** Hepatic stellate cells, extracellular matrix, Enzymatic digestion, Pronase, Optiprep

**[Background]** Hepatic stellate cells (HSCs) harbor in the perisinusoidal space amidst the endothelial cells and hepatocytes. Under homeostatic conditions, HSCs assume a quiescent state comprising about 15% of the liver resident cells (Friedman, 2008). Upon activation, HSCs can differentiate into myofibroblasts producing copious amount of extra-cellular matrix components, such as collagen.

Originally, rat livers were utilized for HSC isolation during the 1980s (Knook *et al.*, 1982; Friedman *et al.*, 1985). Owing to the ability of quiescent HSCs to store vitamin A in lipid droplets, isolation was based on density gradient separation. On similar lines, vitamin A rich mouse HSCs localize in the upper layers in density gradients (Chang *et al.*, 2014, Maschmeyer *et al.*, 2011). Subsequently developed FACS based methods explore the inherent vitamin A fluorescence to sort HSCs (Mederacke *et al.*, 2015).

Our method was implemented in our recent studies on pericyte induced wound healing (Minutti *et al.*, 2019) and is advantageous as a rapid, inexpensive method to yield around  $2 \times 10^5$  HSCs and takes about 3.5 h. HSCs isolated using our method can be used in gene expression studies, *in vitro* differentiation assays and can be applied to wild type as well as genetically modified mice.

### Materials and Reagents

1. Culture hood for sterile work
2. Dissection scissors (Any available dissection scissors can be used)

3. Forceps- 2 pairs (preferably blunt forceps from any manufacturer are preferred)
4. Ice box
5. Tissue culture treated T75 flask (Corning, catalog number: S430641)
6. 0.22  $\mu\text{m}$  sterile syringe filters (Millex/Millipore Sigma, catalog number: SLGP033RS)
7. 10 ml syringes (BD Plastipak, catalog number: 302188)
8. 30 ml Universal Containers (Thermo Fisher Scientific, catalog number: 12511299)
9. 27 G needles, disposable,  $\frac{1}{2}$  " (BD, catalog number: 305109)
10. Nybolt mesh or 70  $\mu\text{m}$  Cell Strainer (Falcon, catalog number: 352350)
11. 15 ml tubes (Sarstedt 62.554.502)
12. 50 ml tubes (Sarstedt 62.548.004)
13. Disposable 5 ml and 10 ml pipettes
14. 100 mm x 20 mm (D x H) Tissue-culture treated culture dishes (Corning, catalog number: 430167)
15. Taqman probes, used for *Acta2* (Mm00725412\_s1) and *Rn18s* (Mm03928990\_g1) from Applied Biosystems for validation in Figure 4
16. 4 x Mice
17. DNase 1, Grade II, from bovine pancreas (Roche, catalog number: 10104159001)
18. Collagenase B (Roche, catalog number: 11088807001)
19. Protease from *Streptomyces griseus* (Pronase) (Sigma, catalog number: P5147)
20. Hanks Balanced Salt Solution (Sigma-Aldrich, catalog number: H9394)
21. Axis-Shield Density Gradient Media OptiPrep™ (Abbott Diagnostics Technologies AS, catalog number: 1114542)
22. Dulbecco's Modified Eagle Medium (DMEM) with high glucose, pyruvate (Thermo Fisher Scientific, catalog number: 41966)
23. Fetal Bovine serum (FBS) (Gibco, catalog number: 10500064)
24. Penicillin-Streptomycin (Thermo Fisher Scientific, catalog number: 15140122)
25. L-Glutamine (Thermo Fisher Scientific, catalog number: 25030081)
26. Trypsin 0.25% EDTA (Gibco, catalog number: 25200056)
27. Dulbecco's Phosphate Buffered Saline (PBS) (Sigma-Aldrich, catalog number: D8537)
28. Red Blood Cell Lysing Buffer Hybri-Max (Sigma, catalog number: R7757)
29. Ethanol absolute > 99.8% (VWR, 20821.330P)
30. Enzymes (see Recipes)
31. Complete DMEM (see Recipes)

### **Equipment**

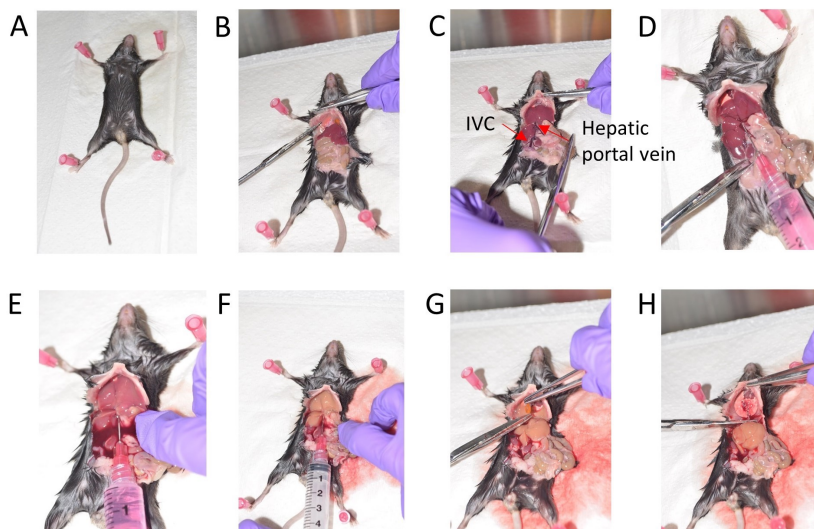
1. Milligram Balance
2. CO<sub>2</sub> chamber for mouse asphyxiation (Any available apparatus approved by the animal ethics committee and the animal facility may be used.)

3. Incubator-shaker or water bath (Grant W28)
4. Refrigerated centrifuge for 15- and 50-ml tubes (Eppendorf, model: 5810R)
5. Inverted microscope (Zeiss Primovert)
6. Incubator for culturing cells (Wolflabs, Binder CB160)

## **Procedure**

### A. Liver perfusion and isolation (Figure 1 and Video 1)

1. Pool the livers from 4 mice for setting up HSC isolation and culture.
2. Euthanize 1-2 mice at a time according to a method approved in your Institutional Animal Care and Use Committee (IACUC) Protocol.
3. After confirming death, cut the mouse skin along the midline from the abdomen to the thoracic cavity. Cut the diaphragm, effectively exposing the visceral organs (Figures 1A and 1B).
4. Gently move the liver lobes where the diaphragm was cut and then move the intestine to your right side to expose the inferior vena cava (Figure 1C).



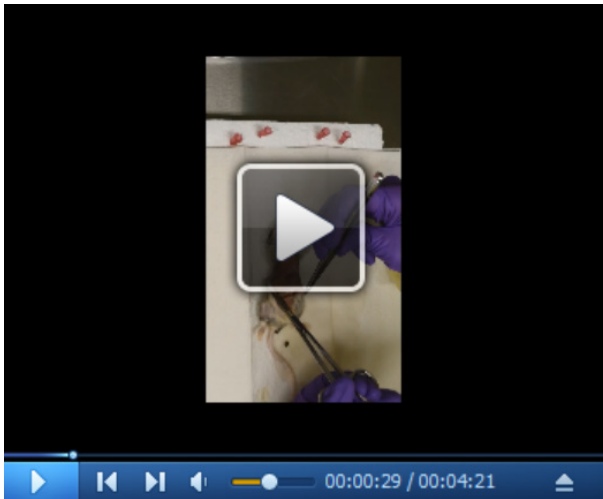
**Figure 1. Stepwise depiction of mouse liver perfusion.** A. Position the mouse for dissection. B. Expose the visceral organs and cut the diaphragm. C. Gently move the intestine to the right side. D. Inject 1x HBSS via the Hepatic portal vein. E. Cut the Inferior vena cava (IVC). F. Wash with 1x HBSS. G. Remove the gall bladder. H. Dissect out the liver.

5. Fill a 10 ml syringe with ice cold HBSS and use a 27G needle to inject this HBSS into the hepatic portal vein to perfuse the liver (Figure 1D).
6. Immediately cut off the inferior vena cava (IVC) to release the pressure, once you begin washing with HBSS via the portal vein (Figures 1E and 1F).
7. Use at least 10 ml HBSS to wash each liver.

*Note: Euthanizing 1-2 mice at a time helps preserve tissue integrity and leads to efficient flushing*

*of the livers resulting in a cleaner HSC preparation.*

8. Gently cut off the gall bladder amidst the liver lobes taking care to avoid puncturing it (Figure 1G).
9. Collect the lobes of the liver in a 30 ml universal container with 10 ml ice cold HBSS (Figure 1H).
10. Proceed to enzymatic digestion.



**Video 1. Mouse liver perfusion.** This video was made at Oregon Health and Science University (OHSU), Portland according OHSU Institutional Animal Care and Use Committee (IACUC) Protocol guidelines and approved by the Department of Comparative Medicine.

#### B. Enzymatic digestion

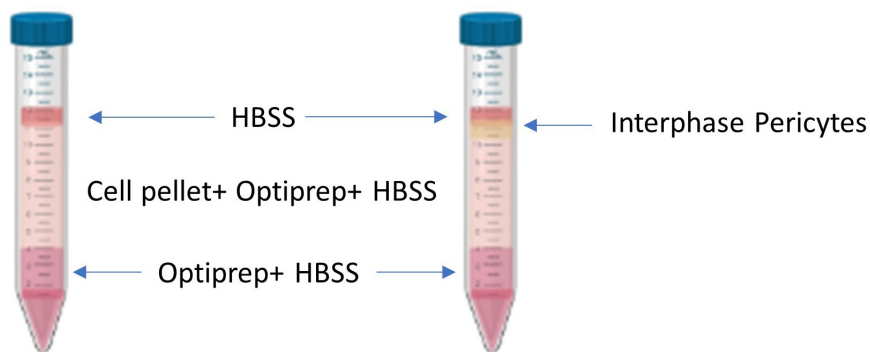
1. Under a sterile culture hood, drain the HBSS from the universal containing the livers.
2. Mince the livers in the same 30 ml universal container using sterile scissors and forceps (sprayed with 70% ethanol) and add 10 ml of Collagenase B and 10 ml of Pronase to the minced livers.

*Note: Pre-warm the Collagenase B and Pronase for 15 min in a 37 °C water bath.*

3. Add 2 ml DNase 1 solution to prevent clumping of cells.
4. Mix the contents well by inverting the tube 3-4 times and place the universal in an incubator-shaker at 37 °C for 20 min.
5. Pour the digest through a 70 µm cell strainer for collection into a 50 ml tube. Use the flat side of a 10 ml syringe plunger to gently press and force the liver digest from the cell strainer into the 50 ml tube and top up with cold HBSS to stop the enzymatic reaction.
6. Centrifuge at 600 x g (around 1,800 rpm for a benchtop centrifuge), at room temperature for 7 min.
7. Pre-cool the centrifuge to 4 °C for the density gradient step.

#### C. Assembling the density gradient (Figure 2)

1. For the bottom-most layer of the gradient, add 3.2 ml Optiprep to 4.8 ml HBSS. Split the 8 ml into two 15 ml tubes.
2. For the intermediate phase, homogenize the cell pellet from Step B6 by gently tapping the 50 ml tube at the base and add HBSS to make the total volume 8.9 ml.
3. Add 5.9 ml of Optiprep to the 8.9 ml cell pellet resuspended in HBSS. Mix using a 10 ml pipette.
4. Using a 5 ml pipette, gently add an equal volume dropwise on top of the 4 ml bottom phase to each 15 ml tube using the slowest setting on the pipette-aid. The volume in each 15 ml tube should be around 10.5-11 ml combining both phases.
5. Finally, for the top phase, gently add 0.5 ml HBSS over the intermediate phase.
6. Centrifuge the two 15 ml tubes at  $1,400 \times g$  (around 2,500 rpm) for 20 min at  $4^\circ\text{C}$  without break and at a slow acceleration.



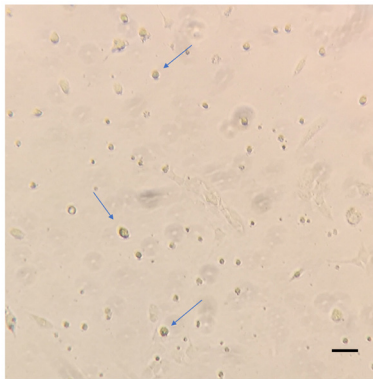
**Figure 2. Density gradient pre and post centrifugation.** A depiction of the layers before centrifugation (left) and after centrifugation (right). The isolated pericytes localize at the phase between the top layered HBSS and the middle layer containing the cell pellet homogenized with HBSS and Optiprep.

#### D. Washing and plating

1. The HSCs are in the second layer from the top, beneath the HBSS and appearing as a thick white band.
2. Gently remove this layer comprising about 0.5 to 1 ml, using a 1 ml micropipette, from each 15 ml tube and put in a new 50 ml tube. Tap and mix cells and add 2 ml DNase. Top up with HBSS up to 50 ml and spin down at  $600 \times g$  for 7 min at room temperature.
3. Aspirate the supernatant and mix the pellet. Add 3 ml of RBC lysis buffer and incubate at  $37^\circ\text{C}$  for 4-5 min.
4. Add 2 ml DNase solution to prevent clumping of cells. Wash with HBSS and spin down at  $600 \times g$  for 7 min.
5. Aspirate the HBSS, mix the pellet and resuspend in about 10 ml prewarmed complete DMEM (see Recipes).
6. Plate 10 ml of the cell suspension in a 100 mm plate or in a T75 flask as preferred. The

preparation looks messy immediately after isolation.

7. Incubate at 37 °C in 5% CO<sub>2</sub> environment.
8. After 24 h, gently rotate the plate in the palm of your hand to displace the thick sticky layer. To obtain a clear preparation, aspirate the media and wash 3-4 times with 3 ml warm PBS. Resuspend in complete DMEM or proceed to trypsinization to set up a new experiment.



**Figure 3. Representative image of HSCs 24 h post isolation.** Isolated HSCs cultured in 100 mm plates were washed with warm PBS and imaged on the Zeiss primover at 10x magnification. The blue arrows indicate the adhered HSCs. Scale bar = 200 μm.

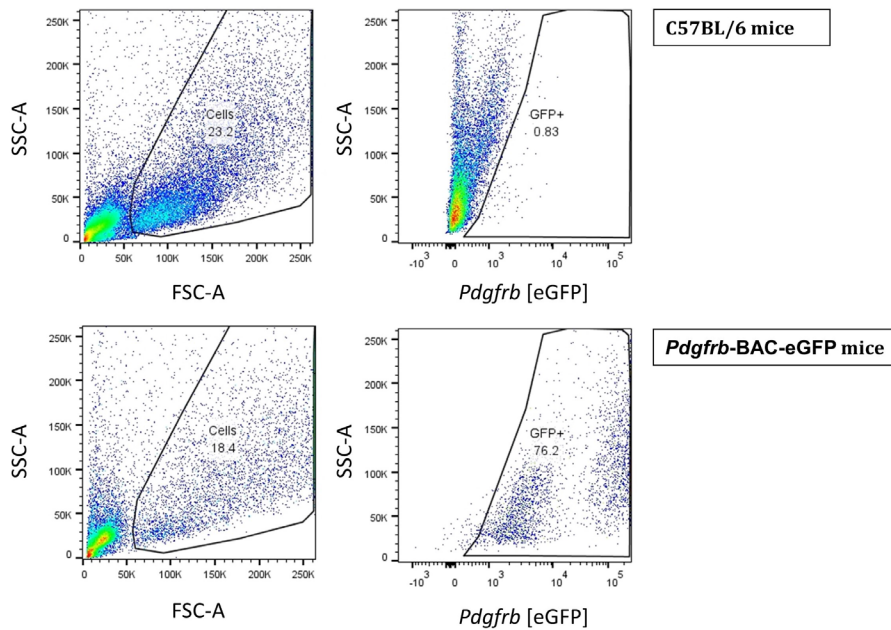
9. About 2-2.5 x 10<sup>5</sup> HSCs can be obtained by this method.

#### E. Trypsinization

1. Trypsinization of HSCs during the early stages after first wash is a delicate process. After 3 washes with pre-warmed PBS, add 3 ml trypsin and incubate at 37 °C for 4 min with pipetting up and down about 5 times during the incubation.
2. Stop the reaction with ice cold HBSS and spin down at 600 x g for 7 min to count cells and re-plate them.
3. For trypsinization at later stages, incubate for 5-6 min and proceed in a similar way.

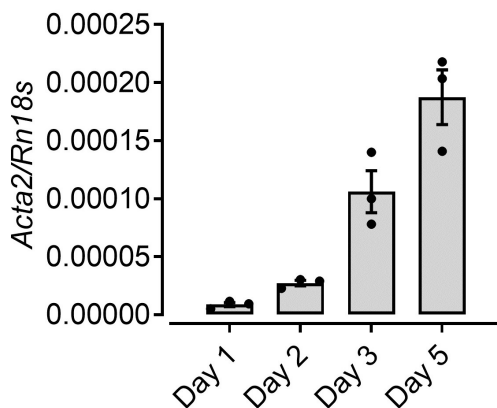
#### F. Validation

1. Let the HSCs isolated using the optiprep method to adhere overnight. After 24 h, wash, trypsinize and analyze the cells using flow cytometry (Figure 4). Since *Pdgfrb* is a prominent marker of HSCs (Henderson *et al.*, 2013), we used *Pdgfrb*-BAC-eGFP reporter mice to validate *Pdgfrb*-eGFP expressing cells alongside cells isolated from WT C57BL/6 mice used as a negative control.



**Figure 4. Flow cytometry of isolated pericytes.** HSCs from WT C57BL/6 mice *Pdgfrb*-BAC-eGFP mice were acquired on the BD LSRII flow cytometer and analyzed using FlowJo Software Version 10.

- As an additional validation of HSC differentiation, we cultured the isolated cells in complete DMEM for 5 days and harvested the samples intermittently for the analysis of *Acta2* gene expression (Figure 5). Alpha smooth muscle actin (*Acta2*) is a reliable marker of HSC differentiation.



**Figure 5. Increased *Acta2* expression upon differentiation of HSCs culture.** mRNA levels of  $\alpha$ -smooth muscle actin (*Acta2*) from HSCs isolated and cultured in complete DMEM over 5 days analyzed by RT-qPCR.



## **Notes**

For a high yield of HSCs, use old mice for isolation or mice subjected to overnight fasting.

## **Recipes**

### 1. Enzymes

DNase 1: 0.025% in 10 ml HBSS

Collagenase B solution: 0.2% in 10 ml HBSS

Pronase (Protease from *Streptomyces griseus*): 1% in 10 ml HBSS

*Note: Filter all the enzyme solutions using a 10 ml syringe and a 0.22  $\mu$ M filter into a new sterile universal tube.*

### 2. Complete DMEM (for culture)

20% FBS

1% Penicillin-Streptomycin

1% L-Glutamine

## **Acknowledgments**

We thank Prof. Neil Henderson for provision of *Pdgfrb*-BAC-eGFP mice. The work was supported by the European Union grant CIG-631413 (“EGF-R for Immunity”) and the Medical Research Council (MR/M011755/1). This protocol has been modified from Graham, 2002.

## **Competing interests**

The authors declare no conflicts of interests.

## **Ethics**

Animal experiments were approved by University of Edinburgh Animal Welfare and Ethical Review Body and were performed under the project license PPL70/8470 granted by the UK government Home Office.

Imaging and videography of the mouse dissection was conducted with the approval of Oregon Health and Science University–Institutional Animal Care and Use Committee (IACUC) with the kind support of Dr. Kim Saunders and Dr. Anupriya Agarwal.

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