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The Processes Of Anterior Pituitary Hormone Pulse

Generation

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1 Abstract

2 More than 60 years ago, Geoffrey Harris described his "Neurohumoral theory" in which the 3 regulation of pituitary hormone secretion was a "simple" hierarchal relationship, with the hypothalamus as the controller. In models based on this theory, the electrical activity of 4 5 hypothalamic neurons determines the release of hypophysiotropic hormones into the portal 6 circulation and the pituitary simply responds with secretion of a pulse of hormone into the 7 bloodstream. The development of methodologies allowing monitoring of the activities of 8 members of the hypothalamic-vascular-pituitary unit is increasingly allowing dissection of the 9 mechanisms generating hypothalamic and pituitary pulses. These have revealed that whilst 10 hypothalamic input is required, its role as a driver of pulsatile pituitary hormone secretion 11 varies between pituitary axes. The organisation of pituitary cells has a key role in modifying 12 their response to hypophysiotropic factors, which can lead to a memory of previous demand 13 and enhanced function. Feedback can lead to oscillatory hormone output that is independent of 14 pulses of hypohysiotropic factors and instead results from the temporal relationship between 15 pituitary output and target organ response. Thus, the mechanisms underlying the generation of 16 pulses can not be generalised and the circularity of feedforward and feedback interactions must 17 be considered to understand both normal physiological function and pathology. We describe 18 some examples of the clinical implications of the recognition of the importance of the pituitary 19 and target organs in pulse generation and suggest avenues for future research in both the short 20 and long-term.

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22 Précis

Le Tissier and his colleagues revisit Harris's "Neurohumoral theory" to reassess the
 contribution of individual components of hypothalamic-pituitary-target organ axes in hormone
 pulse generation.

1 Introduction

2 Understanding the origin of anterior pituitary hormone pulses in health and how they are 3 disturbed in disease is a long-standing question (1). The accepted "textbook" view has been that 4 hypothalamic hormones are the dominant factors generating these pulses, based largely on the 5 seminal experiments of Geoffrey Harris and colleagues that led him to develop the 6 "Neurohormonal Theory" (2). Over sixty years later, the importance of hypothalamic factors is 7 still unquestioned, however, it is apparent that their role in pituitary pulse generation is more 8 complex than previously assumed. It is now clear that no single model system exists and that for 9 each pituitary axis pulses of hormones are generated by a combination of hypothalamic input 10 (3), pituitary response (4), short loop feedback (5) and target organ feedback (6). A clearer 11 understanding of these interactions allows definition of their orchestration, essential for 12 understanding the circuitries underlying physiology and behaviour (7). 13 In this review we will principally consider ultradian pulses of anterior pituitary hormones and 14 divide their generation into two components: the regulatory inputs to the pituitary, both from 15 the hypothalamus and peripheral organs; and the response of the pituitary gland. We will use 16 specific examples to describe the processes and interactions involved and how their 17 modification may lead to pathology. We will focus on the mechanisms underlying the generation 18 of ultradian pulses. This review will not address the circadian pattern of pituitary hormone 19 output: instead the reader is referred to three relevant articles/reviews (8-10).

Hypothalamic and target organ input in the generation of pituitary hormone pulses.

3 Hypophysiotropic neurons share features with many other neuronal cell types

4 The parvocellular hypothalamic neurons, which store and secrete hypophysiotropic hormones, 5 have largely been considered as a separate class of neuron from those in other regions of the 6 brain that have traditionally been classified by their small neurotransmitters. This view has 7 now changed with studies of neurotransmitter contribution to the regulation of neurohormone 8 release (11,12) and the realisation that many other neuronal circuits can be classified by their 9 secretion of neuropeptides whether they have (e.g. somatostatin (SST) (13)) or do not have (e.g. 10 kisspeptin (14) and orexin (15)) a neuroendocrine role. A unique feature of parvocellular 11 neurons is that they lack post-synaptic targets, however, it has been shown that neuronal (16) 12 and endothelial cell (17) inputs can modulate GnRH nerve terminal activity at the median 13 eminence, similar to retrograde signalling at synaptic terminals. This highlights that the 14 mechanisms and interactions which regulate hypothalamic parvocellular neurons should not be 15 considered in any way distinct from those of other brain regions. A further recent realisation is 16 that neurons considered as single populations may exist as multiple subtypes, which has been 17 shown by single cell transcriptomics (18,19) and this is also true of hypophysiotropc neurons: 18 regionally distinct gonadotrophin releasing hormone (GnRH) neurons have differential roles in 19 GnRH pulse and surge generation (reviewed in (20)); and functional studies have identified two 20 types of tuberoinfundibular dopamine (TIDA) neurons, only one of which is responsible for 21 regulation of prolactin release (21). Further studies are likely to reveal heterogeneity in other 22 hypophysiotropic neuronal populations with distinct roles in both pituitary regulation and 23 modification of other hypothalamic functions.

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1 Pulsatile hypothalamic output is not required for pulsatility in all pituitary axes

2 It is becoming increasingly clear that the previously accepted concept of the hypothalamus as 3 the source of anterior pituitary hormone pulse generation is not applicable to all axes (Figure 4 1). This concept, of a simple hierarchal pulse generating relationship, is based on seminal 5 studies in multiple species showing a concordance of the pattern of GnRH output and that of 6 pituitary luteinising hormone (LH) and follicle stimulating hormone (FSH) (22-24). Afferent 7 inputs to GnRH neurons are the origin of pulse generation (25), as demonstrated by a series of 8 extensive and elegant studies of the GnRH system (20). As a consequence, the GnRH system has 9 provided a paradigm for pulse generation in other pituitary axes, especially where the pattern 10 of hypothalamic output can not be robustly measured. Identification of the factors regulating 11 other axes show a further level of complexity, with multiple hypothalamic factors having 12 synergistic (eg corticotrophin releasing hormone (CRH) and vasopressin (26)) or antagonistic 13 (eg growth hormone releasing hormone (GHRH) and SST (27)) actions that affect the amplitude 14 or duration of a pituitary hormone pulse but do not contradict the hierarchal relationship. 15 However, recent studies in the adrenal axis has questioned the requirement for pulsatile 16 hypothalamic input for a corresponding pulsatile pituitary output: constant CRH stimulation in 17 conscious, freely moving rats resulted in pulsatile adrenocorticotrophic hormone (ACTH) and 18 corticosterone release with a frequency unaltered from endogenous pulses (28). Similarly, in 19 the thyroid axis constant infusion of thyrotrophin releasing hormone (TRH) in humans has been 20 shown to result in pulses of TSH (29). This is not to say that TRH and CRH are not released into 21 the portal circulation in pulses, where measurement has been made release is pulsatile (30-33) 22 but this may be more related to maintaining responsiveness of target cells rather than pulse 23 generation per se (34). Thus the paradigm established by the GnRH-gonadotrophin-sex 24 hormone relationship (Figure 1, left) may not hold for other axes, such as CRH-ACTH-cortisol 25 (Figure 1, right), or indeed fully account for the relationship of GnRH and gonadotroph output at 26 the time of the LH surge (20). Measurement of other hypophysiotropic factors with sufficient

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2 optogenetic manipulation of their hypothalamic neurons are required to determine this. 3 The electrical activity required for neurohormone release can be defined but is modified 4 with physiological status 5 In those pituitary axes where pituitary hormones are released in pulses with a frequency of 10s-6 100s of minutes, it has not (to date) been possible to directly correlate the patterns of 7 hypothalamic neuron electrical activity with their hypophysiotropic secretion. Calcium imaging 8 and optogenetic manipulation to impose electrical activity with concurrent monitoring of 9 pituitary hormone output (assumed to reflect hypothalamic factor release) have been used as 10 alternative approaches to determine the minimal frequency and duration required to drive 11 neurohormone secretion. This has been successfully applied to GnRH neurons, demonstrating 12 that stimulation at 10 Hz (but not at frequencies below 5 Hz) are required for a duration of 2 13 minutes (but not 30 seconds) for generation of LH pulses (35), and to kisspeptin neurons, 14 identifying them as a source of the GnRH neuron pulse generator (25,36). These studies assume 15 that electrical activity and neurohormone release are correlated, which is likely in the short 16 term, however studies of TIDA neuron electrical activity with simultaneous recording of 17 dopamine output has demonstrated that this may not be true with changes in physiological 18 status (37). In lactation, prolactin feedback no longer leads to dopamine release from TIDA 19 neurons, which maintains the high level of the lactogen, but unexpectedly still leads to increased 20 electrical activity. A similar disconnect between electrical activity and neurohormone release 21 may occur in kisspeptin neurons which have an altered optogenetic stimulatory requirement in 22 diestrus and ovariectiomized females (36) and a loss of neurohormone expression in lactation 23 (38).

temporal resolution to determine their relationship with pituitary hormone output or

24 Hypothalamic neuron coordination is required for pituitary regulation

Whatever the requirement for pulsatile hypothalamic output to generate pituitary hormonal
pulses, there is an absolute requirement for hypophysiotropic regulation of the pituitary for

1 normal physiological function. This requires coordinated release from multiple neurons to 2 ensure a sufficient concentration of neurohormone in the portal circulation to elicit a response 3 from pituitary cells; for example, it has been shown that a minimum of 60 GnRH neurons are 4 required for pulsatile LH release in mice (35,39) but five times that number are required for 5 surge generation (40). More direct evidence for coordination has been shown by monitoring 6 pairs of TIDA neurons, where the electrical activity of a proportion of cells are coordinated over 7 a period of minutes (37). In both cases, there is an implication that a subset of the neuronal 8 population is active at any one time, which may be important in avoiding fatigue. This provides 9 a rationale for a large reserve population but also a requirement for interneuron coordination 10 over both space and time; for example, a multi-layered spatial and temporal coordination of 11 TIDA neurons remains stable over a period of days, which may underlie the sustained dopamine 12 release required for inhibition of prolactin secretion (41). Such multi-layered organization of 13 neuronal spiking frequencies are widely used for other brain-body functions, such as sleep, in 14 both mammalian animal models and humans (42).

15 Intrinsic and extrinsic mechanisms coordinate hypothalamic neuron activity

16 The coordination of hypothalamic populations regulating pituitary secretion can occur through 17 a number of mechanisms. In other brain systems, including those of other parvocellular 18 neuronal systems, negative feedback and feedforward loops act as relatively simple networks to 19 coordinate population activities (43) and there is evidence for a similar network-driven 20 coordination of the hypophysiotropic neurons. These can be divided into intrinsic interactions 21 within a population and extrinsic coordination requiring input from other neuronal cell types. 22 Whilst there is evidence for both (as described below), a combination is likely to ensure the 23 coordination required for robust pituitary regulation.

24 Intrinsic coordination

Coordination of the electrical activity of hypophystiotropic neurons has been reported, in
particular for dopamine (37,44) and cultured GnRH neurons (45). Whilst this may suggest a role

1 for electrical coupling through gap junctions, these have been shown to be absent in both mouse 2 TIDA (37) and GnRH (46) neurons. There may be species differences, however, since in rat TIDA 3 neurons electrical coupling mediated by gap junctions has been described (44). An alternative 4 mechanism underlying intrinsic coordination is chemical coupling and there is evidence for this 5 regulating TIDA neurons via negative feedback loops, with TIDA neurons both releasing and 6 responding to GABA (47). In addition, dopamine 2 receptor (D2R) at the TIDA neuron cell body 7 mediates an ultrashort feedback loop leading to oscillatory activity in rats (48). A similar 8 ultrashort autoregulatory loop has been described for GnRH neurons, which express GnRH 9 receptors and have altered electrical activity in response to GnRH (49).

10 Extrinsic coordination

11 Input from the higher brain centres regulating hypophysiotropic neurons will obviously 12 coordinate their activity, however, there is a clear role for intrahypothalamic regulation (50) 13 and it is well recognised that SST and kisspeptin have important regulatory roles in GHRH (51) 14 and GnRH output (14) respectively. Recent studies have determined specific roles for these 15 extrinsic factors and defined key steps in their regulation of neurohormone output. The inhibitory action of SST has been shown to counterintuitively lead to stimulation of GHRH 16 17 neurons as a result of an initial fast and transient direct inhibition of the GHRH neuron itself, 18 followed by a delayed inhibition of both excitatory glutamate and inhibitory GABA inputs (52). 19 Optogenetic manipulation has identified kisspeptin as the GnRH pulse generator (25) and other 20 studies have shown that firing of kisspeptin neurons is modulated by steroid feedback (53). 21 Thus, feedforward loops are key features of both of these hypophysiotropic systems. A further 22 complexity in the extrinsic inputs regulating hypophysiotropic ouput may be their subcellular 23 location. Kisspeptin has been shown to have differential effects at the GnRH cell body compared 24 with the nerve terminals at the median eminence (16), where a role for local endothelial nitric 25 oxide production has been suggested as a local synchronising signal (54).

26 The median eminence plays a role in coordinating and modifying hypothalamic output

1 The final step in the output of hypophysiotropic hormones is their release at the median 2 eminence (ME). Release from a large number of neurons into this richly vascularised structure, 3 with convoluted loops collecting output from a large release area, optimises both the amplitude 4 and duration of neurohormone pulses in the portal circulation, whilst avoiding neuronal fatigue 5 and exhaustion (41). In addition to roles in the coordination of hypophysiotropic factor release 6 (see above), the ME may actively modify output by alteration of access of nerve terminals to the 7 rich capillary bed by either changes in localisation, which has been shown to vary with age for 8 GHRH neurons (55), or tanycyte ensheathment, shown for GnRH neurons to vary at different 9 stages of the oestrous cycle (56).

10 Peripheral inputs can generate pituitary hormone pulses

The importance of target organ feedback in the regulation of hypothalamic-pituitary axes is well recognised and incontrovertible, balancing the feedforward regulation by hypothalamic and pituitary factors. An excellent example of this is the differential regulation of LH and FSH by ovarian inputs, with reduced inhibin and increased progesterone feedback actions on the pituitary generating a second phase of FSH (but not LH) at proestrus and estrus (reviewed in (22)) (Figure. 1, left panel).

17 Remarkably, recent studies inspired by mathematical modelling have shown that target organ 18 feedback itself can act as a pituitary hormone pulse generator, as the fast feedforward action of 19 ACTH on the adrenal gland and delayed feedback of glucocorticoids can generate pulses of both 20 hormones with invariant CRH (reviewed in(6)) (Figure 1, right panel). Since an intra-adrenal 21 glucocorticoid feedback loop has recently been suggested from modelling (57), this suggests 22 that the adrenal gland itself may be the primary pulse generator in the hypothalamic-pituitary-23 adrenal axis in the absence of stress. The extent to which similar temporal relationships of 24 hypothalamic-pituitary regulation and feedback exist for other axes is currently unclear, 25 although a delayed feedback of prolactin on dopamine neurons (41) may have an important

1 impact on dopamine tone, facilitate increased secretion of prolactin and lead to the reported

2 ultradian pulses of basal prolactin secretion (58).

3 The potential interactions whereby feedback can generate or modulate pulsatile pituitary
4 hormone secretion are complex and may include:

the temporal relationship between the feedforward and feedback regulation, which is
 complicated by the feedback occurring at multiple levels; for example, the differential
 feedback actions of ovarian steroids are mediated by rapid non-genomic and classical
 steroid receptor actions in both the hypothalamus and pituitary during the oestrous
 cycle, with effects that are dependent on receptor isoform expression and downstream
 signalling (reviewed in (22,24)).

the sensitivity of the system to feedback, exemplified in the thyroid axis, where
 differential expression of thyroid hormone receptor beta isoforms results in its relative
 increase in sensitivity to thyroid hormones (59), providing an anticipatory mechanism
 to protect peripheral organs from overexposure to these hormones (34).

differential feedback at the level of the hypothalamus and pituitary. Again the thyroid
axis provides an excellent example of this, since feedback to the hypothalamus is
dependent on active transport of thyroid hormone at the level of the median eminence
(60) but is enhanced by post-translational modification of type 2 deiodinase (61).
multiple factors feeding back on a single cell type; for example, dopamine neurons will

20 be exposed to GABA, dopamine and prolactin feedback, with different time scales
21 (41,47,48).

Thus, it is possible that feedback occurs at both pituitary and hypothalamic levels and at multiple sites within each organ, through the action of multiple factors on a single cell type, or both. This is further complicated when consideration of feedback to higher brain centres is included; for example, glucocorticoid feedback on the limbic system and brain stem (62).

1 Intrapituitary regulation of hormone pulse generation

2 It is perfectly feasible that the anterior pituitary gland would simply passively respond to 3 hypothalamic and peripheral inputs, with its cells simply acting as an amplifier of hypothalamic 4 regulation that is modulated by feedback from target organs. However, it is now apparent that 5 this is not the case, with an active role for the pituitary mediated by the structural organisation 6 of its component cell types affecting how they receive, interpret and translate hypothalamic and 7 peripheral signals into highly ordered hormone pulses. This was previously suggested by a 8 disconnect in the output of dispersed pituitary cells compared with those in the intact gland 9 (63) but is being increasingly demonstrated and dissected using a combination of mouse models 10 and technological innovation which allows both temporal and structural imaging (64,65).

11 Pituitary cells are organised as intermingled homotypic cell networks.

12 Large-scale 3D imaging of genetically-modified mouse models expressing fluorescent proteins 13 under hormone promoter control has revealed the structural and functional organisation of the 14 pituitary gland and its rich vascularisation (4). This has described the developmental program 15 of the topological organisation of differentiated cells throughout the gland (63,66) from early 16 fetal life to adulthood, and has demonstrated a role for early-differentiated cells (e.g. 17 corticotrophs) in controlling both the positioning and expression of late-differentiated cells (e.g. 18 gonadotrophs) (67). Contact between homotypic cells and organisation of characteristic 19 morphological features occurs soon after endocrine cell differentiation, before the onset of 20 hormone secretion and leads to cell network formation (67,68). Among the signal molecules 21 involved in cell network architecture and plasticity, detailed analysis of the cadherin family has 22 revealed a 'bar-coding' expression of cadherins within distinct pituitary cell populations from 23 both mouse models (69) and humans (70), which has also been proposed as a marker rule for 24 discriminating invasiveness in GH and PRL adenomas.

Pituitary cell networks share fundamental properties with other biological networks including
metabolic signalling networks in yeast and bacteria (71), where a prevalent feature is that of

1 simple assembles of elements (so-called network motifs) which recur within the population 2 (72). The organisation of the various pituitary cell types into distinct motifs suggests that there 3 will be differences and similarities in their role in axes function. The spatial organisation, and 4 its plasticity throughout life, is exemplified by the GH and prolactin cell networks. Upon sexual 5 maturation there is a transient increase in the generation of multiple clusters of contacting GH 6 cells (illustrated in Figure 2) in males but not females in the wings of the pituitary, coincident 7 with an increase in the highly ordered GH pulses which control liver insulin-like growth factor 1 8 production (64,68). The importance of these GH cell clusters as network motifs which lead to 9 increased body growth is suggested by the correlation of their formation with growth rate (68) 10 and the finding that GH-deficient animals are normal in size if the GH cell clusters are preserved 11 (73). In contrast, PRL cells are organised as multiple honeycomb network motifs (like an orange 12 peel) which are more prominent in lactating females and display experience-dependant 13 plasticity as they remain after weaning (74). This altered network organisation has been shown 14 to result in enhanced prolactin output in subsequent lactations (74) and may have a role in 15 reducing the tonic output of prolactin in reproductively experienced rats (75) through an 16 enhanced response to dopamine inhibition (76).

17 The vasculature has a role in signal input to pituitary cell networks

18 Networks of endocrine cells do not work alone but form a functional continuum with other 19 elements within the pituitary gland, including the vasculature. Network motifs and the rich 20 plexus of fenestrated capillaries are topologically organized in a manner which is distinct for 21 each endocrine cell type (Figure 2), and may therefore reflect their different secretory temporal 22 dynamics (63). Initial cell network formation begins before the first capillaries invade 23 embryonic pituitary tissue (67) and loss of the pituitary cell transcription factor Prop1 leads to a failure of organ vascularization (77). Thus, endocrine cell networks have a stimulatory and 24 25 organisational role in patterning capillary invasion.

1 The organisation of the vasculature with pituitary endocrine networks may have a significant 2 impact on the amplitude and timing of exposure of pituitary cells to hypothalamic regulatory 3 factors since seminal studies of the portal vasculature have shown that hypophysiotropic nerve 4 terminals specifically abut portal vessels which irrigate specific pituitary regions (78,79). In 5 addition, there is a highly dynamic regulation of the distribution of incoming secretagogues 6 within the pituitary through altered blood flow dynamics within the capillary bed of the 7 pituitary (80) but rapid transit of signalling molecules (in a range of seconds) throughout portal 8 fenestrated capillaries (81). This suggests differential timing of exposure of different regions of 9 the pituitary to hypophysiotropic factors, resulting in a complex dynamic of sequential 10 stimulation, with "scout cells" stimulated before other cells within a homotypic network. Since 11 networks have a functionally coordinating role (see below), this pattern of exposure may lead to 12 synergistic interactions and potential role(s) for specific subsets of cells ensuring robust 13 responses to stimulation. The contribution of the blood system to pituitary hormone pulsatility 14 also involves the fate of hormones from their releasing site towards the bloodstream, which will 15 ultimately deliver the appropriate pattern of hormone pulses to the peripheral target, as well as 16 coordination of oxygen and nutrient supply with the metabolically demanding processes of (80).

17 Pituitary networks coordinate response to regulation

18 A functional role for homotypic pituitary cell networks in determining endocrine output is 19 suggested by their formation before the onset of hormone secretion and stimulation by 20 secretagogues and functional reorganization in response to altered demand (68,74). This has 21 been confirmed by ex vivo analysis of calcium and gene expression dynamics in homotypic cell 22 networks, with coordinated responses to stimulation that are severely dysregulated when 23 networks are disrupted (64,73,82,83). Gap junction coupling contributes to this network 24 coordination (64,74,82), however, this does not preclude roles for paracrine factors between 25 both homotypic and heterotypic cells (reviewed extensively in (84)), such as secreted TSH 26 which exerts an ultrashort negative feedback which could drive ultradian TSH pulses (34). It is

1 also possible that pituitary networks mediate predictive programming, or priming, of axis 2 function since in prolactin cells, the increased organisation associated with lactation persists 3 for months after weaning and leads to enhanced function (74). Similarly, the increased 4 clustering of somatotrophs at puberty could be considered as a priming event for increased GH 5 release, although in this case the effect is transient (64). It is possible that similar transient 6 changes in organisation enhance the altered sensitivity and self-priming of gonadotrophs to 7 GnRH stimulation (85) since increased cell movement and number of cell processes have been 8 described in this cell type in response to GnRH (86) and estradiol (87). Thus both structural and 9 functional organisation of pituitary endocrine cells as intermingled 3D cell networks have 10 important roles in the amplitude and dynamics of hormone secretion which can be modified 11 throughout life.

12 Pituitary cells are heterogeneous

13 Whilst pituitary networks mediate a coordination of cell activity, individual cells also show 14 functional heterogeneity, which may reflect transient or permanent differences in cell activity. 15 This is exemplified by the identification of a small subset of prolactin cells which act as pace-16 makers, or network nodes, synchronizing the activity of nearby homotypic network cells (65). It 17 is these pace-making cells which mediate the altered function of prolactin cells between first 18 and second lactation, showing an ability to store a cellular memory of previous demand that 19 also leads to an enhanced output when rechallenged (akin to learning). Over a timescale which 20 is an order of magnitude longer than that of secretory activity, prolactin gene expression in 21 lactotrophs has also been shown to be heterogeneous (82). A continuous distribution of both 22 transcription rates and switches were found in this study, although interestingly this was locally 23 spatially coordinated by the prolactin cell network, suggesting that the mechanisms underlying 24 homotypic cell coordination can act over a wide range of timescales. Similar functional 25 heterogeneity has been described for other pituitary cell types, which can result in stereotypic 26 variable responses to stimulation which have previously been considered to be stochastic (88).

15

1 Further studies are required to determine if these heterogeneous responses identify a distinct 2 sub-population of pituitary cells or transient activity states, which will likely be identified by 3 high throughput sequencing technologies and/or photolabeling of individual cells in situ (89). 4 Pituitary cell secretion is integrated to shape pulsatile circulating hormone 5 The rate of entry of a secreted pulse of pituitary hormone to the bloodstream and exit from the 6 gland will be determined by the relationship of cell networks with the pituitary 7 microvasculature, where perivascular spaces act as gate-keepers for hormone transfer to the 8 capillary lumen (80). Once in the systemic circulation, a pulse of hormone will be combined with 9 that released in previous secretory events, resulting in the concentration of circulating hormone 10 is an integration of basal and pulsatile release which is dependent on hormone half-life (1,4). 11 Since the half-life of pituitary hormones can be modified by circulating binding proteins (90) 12 and post-translational modification (34), both of which also modify their bioavailability, the 13 pattern of exposure of a receptor to a hormone pulse is complex and will not simply mirror that 14 of pituitary release.

15

16 Implications for health and disease

17 A recognition that pulses of pituitary hormone are generated and modified at multiple levels has 18 important implications for the study of normal axes function but, importantly, also for how 19 dysregulation occurs and for identification of therapeutic targets. This is particularly relevant to 20 pituitary tumours, where hormone output is largely independent of hypothalamic stimulation. 21 In Cushing's disease, for example, there is a marked increase in basal secretion of both ACTH 22 and cortisol and pulsatility is preserved (91) but becomes less ordered (92). Significantly when 23 considering the interaction between glands, there is a decrease in the potency of cortisol 24 stimulation by ACTH (93) and a reduction in the pulse correlation of the two hormones (91). 25 Similar changes in the orderliness of pituitary hormones has been described for other types of

pituitary adenoma, which importantly is largely normalised by surgical but not medical
 treatment (94).

3 It is possible that the interactions between the hypothalamus, pituitary and target organs in 4 generating pulses may have a significant role in a number of endocrine disorders, and thus 5 should be considered as a potential mechanism leading to disease as well as new targets for 6 therapy. For example, multiple studies to identify defects leading to polycystic ovarian 7 syndrome (PCOS) have focused on dysregulation at the levels of the hypothalamus (95,96) and 8 ovary (97). Aspects of the disorder, such as the potential role of hyperinsulinemia in loss of 9 fertility have been studied at the level of the pituitary (eg (98)) but overall there has been a 10 paucity of studies of the role of the pituitary. It is possible that increased LH pulse amplitude 11 (but not pulse frequency) found in PCOS patients may be a result of an alteration of pituitary 12 function and further research into a potential role of the pituitary in this disorder is warranted. 13 Given that PCOS is a heterogeneous syndrome, it is possible that there are multiple aetiologies 14 that involve all levels of the HPG axis some of which may be secondary to the primary defect 15 but nevertheless require improved understanding and may be targets for therapy. 16 The multi-level regulation of pulsatile hormone output also has important implications for 17 diagnosis of dysfunction. An excellent example of this is provided by the thyroid axis, where the

"normal" concentration of circulating TSH can vary between individuals (34) and may be
altered for prolonged periods following normalisation of axis function following hyper- or
hypothyroidism, referred to as hysteresis (99). This may occur as a result of differential rates of
feedback regulation at different levels of the axis; for example, a reduction in hypothalamic TRH
gene expression in response to normalisation of hypothyroidism (100) would be expected to be
rapid in comparison with any change in pituitary thyrotroph cell mass (101), resulting in an

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26 Future research/perspective

alteration of the set point for each level of the axis.

1 Models of pituitary pulse generation:

2 It is clear that there has recently been substantial progress in understanding the contribution of 3 individual components of hypothalamic-pituitary-target organ axes to pulse generation. However, the 4 challenge is to dissect the temporal interactions of these individual components, which requires in 5 *vivo* studies with simultaneous monitoring of system inputs and/or outputs. The use of imaging 6 technology to observe cell activity has been instrumental in much of the recent progress, since it 7 allows both temporal and spatial resolution of activities. Further development of *in vivo* imaging 8 technology, such as the use of gradient index lenses (102), is required to facilitate this. An important 9 consideration in these studies is the silencing of multiple hypothalamic neuron populations by anaesthesia (80), meaning that imaging in awake, freely moving animals is required. Furthermore. 10 monitoring of cell stimulation and activity currently relies on imaging of specific cell signals, such as 11 12 calcium as a surrogate for monitoring both inputs and outputs (eg (64)). The development of 13 methodologies to specifically monitor receptor activation, such as "sniffer cells" (103) or luciferase 14 monitoring of G-protein coupled (104) and cytokine receptor (105) activation may allow more direct 15 measurement of both stimulatory inputs and hormonal output.

16 The heterogeneity of both hypothalamic and pituitary cell populations in generating hormone pulses 17 has been a notable feature throughout this review. Whilst it is possible that this may represent 18 stochastic cell activity in some cases, in others it has been found to be deterministic (88). Since 19 multiple studies have demonstrated that only a small proportion of hypothalamic (eg GnRH (40)) or 20 pituitary (eg GH (73)) cells are required for apparent normal function, the question remains whether 21 the heterogeneous responses reflect sub-populations with specific physiological functions. Identifying 22 the differences in protein expression and post-translational modification that may underlie 23 heterogeneity may be suggested by single cell transcriptomics of cells with specific activities. These 24 studies will not define whether the heterogeneity reflects transient activity or specific sub-population 25 of cells, however they will suggest factors that define sub-populations of cells currently primarily 26 defined by the hormone they produce. The use of cell tracing methodologies, optogenetics and 27 Designer Receptors Exclusively Activated by Designer Drugs (DREADDS), which have already

made dramatic contributions to understanding of hypothalamic-pituitary axis function (eg (25,106)),
will allow confirmation of sub-population identity and study of their function. This will be facilitated
by the use of CRISPR/Cas technology in combination with adeno-associated viral delivery of factors
to manipulate cell function (107). Such single cell transcriptomic approaches have been successfully
applied in other systems, with consequences for identifying novel therapeutic targets (108) and for

6 sub-population identification in the brain (109).

7 Our identification of features akin to memory and learning in the pituitary suggests a potential role of 8 the gland in physiological programming. For example, a persistent alteration in corticotroph activity 9 has been described in adult sheep exposed to a brief period of maternal perinatal undernutrition (110). 10 There is a clear requirement for further investigation of epigenetic alteration of gene expression in such models, however, persistent changes in pituitary cell organisation leading to altered network 11 12 functions are also possible. This will require *in vivo* analysis, as well as a clearer understanding of the 13 mechanisms underlying network-mediated regulation of axis function. The single cell transcriptomics 14 and cell manipulations described above may allow identification of potential mechanisms underlying 15 network function. Mathematical modelling and tissue engineering may aid understanding of how different network motifs affect cell-cell coordination. 16

Finally, the role of the vasculature in modifying temporal and spatial regulation of pituitary function and clearance of secreted hormone from the gland is an area that requires further analysis which should be possible with optogenetic manipulation or the use of DREADDS. These may also be used to determine how the relationship of pituitary cell networks and the vasculature is altered in adenoma formation (111), as well as in other axes dysfunctions such as PCOS.

22 Translation to the clinic:

The mechanisms leading to pituitary hormone pulse generation that are currently being elucidated in rodent models are likely to generally translate to those in humans, however, there are clear species differences in the physiology of pituitary axes (eg prolactin (112)). Analysis of post-mortem pituitary tissue will allow comparison of network organisation and their relationship with the vasculature, as

well as the expression patterns of factors identified as intermediates in network function. It is also
possible that fresh post-mortem tissue and adenomas from patients will allow some functional
analysis of human pituitary function and correlation with that of rodents. The analysis of organoids of
pituitary tissue differentiated from induced pluripotent stem cells (113) will most likely establish
whether mechanisms underlying rodent network function are recapitulated in humans, as well as the
consequences of mutations identified in patients presenting at clinic with pituitary dysfunction.

7 Many of the current protocols for diagnosis of pituitary dysfunction may not fully interrogate the 8 complex interactions leading to pulse generation, which may explain why, for example, current 9 provocation tests misdiagnose GH axis function in a proportion of patients (114). Rodent models will 10 allow the development of tests which can more fully define hypothalamic and pituitary functionality and determination of parameters that are affected when physiology is altered; for example, at puberty 11 12 and in obesity. It is also possible that an *in vivo* assessment of pituitary function may be possible 13 through an improved understanding of how pituitary blood flow relates to function, as this may be assessed in patients through, for example, ultra-fast ultrasound imaging (115). 14

15 Identification of pituitary cell networks may also affect whether and how stem cell therapy could be used for treatment of hypopituitarism, which would be simplified in humans through transphenoidal 16 17 access to the pituitary. Whilst there has been substantial progress in identifying pituitary stem cells 18 (116,117) and developing protocols for differentiation of embryonic and pluripotent stem cells to 19 pituitary tissue (118,119), it is currently unclear whether stem cells would be capable of self-20 organisation or integration into existing pituitary cell networks. This is further complicated by the 21 identification of functional heterogeneity and programming of cell function by previous demand. A 22 naïve stem cell may be capable of differentiation to a lactotroph and integration into an existing 23 network, for example, but may not be functionally equivalent to a cell exposed to the demands of 24 lactation. Furthermore, the prevalence of pituitary adenomas with aberrant network function reflected 25 in disorganised pulsatile output (94) suggests that a failure to fully integrate or recapitulate normal network function may be a risk for the development of pathology. Injection of lineage traced stem 26

- 1 cells into the pituitaries of rodent models of hypopituitarism and functional imaging of their function
- 2 may establish the potential for stem cell therapy in humans.

3 Conclusion

- 4 The elegant and ground-breaking experiments of Harris and colleagues were prescient in their
- 5 use of *in vivo* models which allow multi-organ interactions. It is now clear that in such an
- 6 interactive system the concept of a hierarchy is not appropriate except in the identification of a
- 7 pulse generator, which in the case of the HPA axis, at least, may not be the hypothalamus. This
- 8 does not suggest that the mechanisms and principles underlying the relationship of the
- 9 hypothalamus, pituitary and target organs differ between axes but the strength and timing of
- 10 inputs lead to unique features. Thus, the concepts underlying Harris's Neurohormone Theory of
- 11 regulation of pituitary axes have borne the test of time but new levels of complexity have
- 12 emerged that require consideration of interactions between multiple components of the axes.

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1 References

2 1. Veldhuis JD, Keenan DM, Pincus SM. Motivations and methods for analyzing pulsatile 3 hormone secretion. Endocrine reviews 2008; 29:823-864 4 2. Harris GW. Neural control of the pituitary gland. London: Eward Arnold (Publishers) LTD. 5 3. Clarke IJ. Hypothalamus as an endocrine organ. Compr Physiol 2015; 5:217-253 6 4. Le Tissier P, Campos P, Lafont C, Romano N, Hodson DJ, Mollard P. An updated view of 7 hypothalamic-vascular-pituitary unit function and plasticity. Nat Rev Endocrinol 2017; 8 13:257-267 9 5. Grattan DR, Le Tissier P. Chapter 12 - Hypothalamic Control of Prolactin Secretion, and the 10 Multiple Reproductive Functions of Prolactin. Knobil and Neill's Physiology of Reproduction 11 (Fourth Edition). San Diego: Academic Press; 2015:469-526. 12 6. Spiga F, Walker JJ, Gupta R, Terry JR, Lightman SL. 60 YEARS OF NEUROENDOCRINOLOGY: 13 Glucocorticoid dynamics: insights from mathematical, experimental and clinical studies. The 14 Journal of endocrinology 2015; 226:T55-66 15 7. Kohl J, Babayan BM, Rubinstein ND, Autry AE, Marin-Rodriguez B, Kapoor V, Miyamishi K, 16 Zweifel LS, Luo L, Uchida N, Dulac C. Functional circuit architecture underlying parental 17 behaviour. Nature 2018; 556:326-331 18 8. Bur IM, Zouaoui S, Fontanaud P, Coutry N, Molino F, Martin AO, Mollard P, Bonnefont X. The 19 comparison between circadian oscillators in mouse liver and pituitary gland reveals different 20 integration of feeding and light schedules. PloS one 2010; 5:e15316 21 9. Bonnefont X. Circadian timekeeping and multiple timescale neuroendocrine rhythms. 22 Journal of neuroendocrinology 2010; 22:209-216 23 10. Kalsbeek A, Fliers E. Circadian and endocrine rhythms. Best Pract Res Clin Endocrinol Metab 24 2017; 31:443 25 van den Pol AN, Wuarin JP, Dudek FE. Glutamate, the dominant excitatory transmitter in 11. 26 neuroendocrine regulation. Science 1990; 250:1276-1278 27 12. Decavel C, Van den Pol AN. GABA: a dominant neurotransmitter in the hypothalamus. J 28 Comp Neurol 1990; 302:1019-1037 29 13. Urban-Ciecko J, Barth AL. Somatostatin-expressing neurons in cortical networks. Nat Rev 30 Neurosci 2016; 17:401-409 31 Navarro VM, Tena-Sempere M. Kisspeptins and the neuroendocrine control of reproduction. 14. 32 Front Biosci (Schol Ed) 2011; 3:267-275 33 15. Haghparast A, Fatahi Z, Arezoomandan R, Karimi S, Taslimi Z, Zarrabian S. Functional roles of 34 orexin/hypocretin receptors in reward circuit. Prog Brain Res 2017; 235:139-154 35 16. Iremonger KJ, Porteous R, Herbison AE. Spike and Neuropeptide-Dependent Mechanisms 36 Control GnRH Neuron Nerve Terminal Ca(2+) over Diverse Time Scales. The Journal of 37 neuroscience : the official journal of the Society for Neuroscience 2017; 37:3342-3351 38 17. Bellefontaine N, Hanchate NK, Parkash J, Campagne C, de Seranno S, Clasadonte J, 39 d'Anglemont de Tassigny X, Prevot V. Nitric oxide as key mediator of neuron-to-neuron and 40 endothelia-to-glia communication involved in the neuroendocrine control of reproduction. 41 Neuroendocrinology 2011; 93:74-89 42 18. Chen R, Wu X, Jiang L, Zhang Y. Single-Cell RNA-Seq Reveals Hypothalamic Cell Diversity. Cell 43 reports 2017; 18:3227-3241 44 19. Campbell JN, Macosko EZ, Fenselau H, Pers TH, Lyubetskaya A, Tenen D, Goldman M, 45 Verstegen AM, Resch JM, McCarroll SA, Rosen ED, Lowell BB, Tsai LT. A molecular census of 46 arcuate hypothalamus and median eminence cell types. Nat Neurosci 2017; 20:484-496 47 20. Herbison AE. Control of puberty onset and fertility by gonadotropin-releasing hormone 48 neurons. Nat Rev Endocrinol 2016; 12:452-466

1 2	21.	Brown RS, Kokay IC, Phillipps HR, Yip SH, Gustafson P, Wyatt A, Larsen CM, Knowles P, Ladyman SR, LeTissier P, Grattan DR. Conditional Deletion of the Prolactin Receptor Reveals
3 4 5		Functional Subpopulations of Dopamine Neurons in the Arcuate Nucleus of the Hypothalamus. The Journal of neuroscience : the official journal of the Society for Neuroscience 2016: 26:0172-0185
5	22	Neuroscience 2010, 50.91/5-9165
7	22.	Neill's Physiology of Reproduction (Fourth Edition) San Diego: Academic Press: 2015:1199-
2 2		1257
9	23	Goodman RL Inskeen FK Chapter 27 - Control of the Ovarian Cycle of the Sheen Knobil and
10	23.	Neill's Physiology of Reproduction (Fourth Edition) San Diego: Academic Press: 2015:1259-
11		1305
12	24	Herbison AF Chanter 11 - Physiology of the Adult Gonadotronin-Releasing Hormone
13	24.	Neuronal Network Knobil and Neill's Physiology of Reproduction (Fourth Edition) San
14		Diego: Academic Press: 2015:399-467
15	25.	Clarkson I. Han SY. Piet R. McLennan T. Kane GM. Ng I. Porteous RW. Kim IS. Colledge WH.
16		Iremonger KI. Herbison AF. Definition of the hypothalamic GnRH pulse generator in mice.
17		Proceedings of the National Academy of Sciences of the United States of America 2017:
18		114:E10216-E10223
19	26.	Herman JP, Tasker JG. Paraventricular Hypothalamic Mechanisms of Chronic Stress
20		Adaptation. Frontiers in endocrinology 2016; 7:137
21	27.	Robinson ICAF, Hindmarsh PC. The Growth Hormone Secretory Pattern and Statural Growth.
22		Comprehensive Physiology: John Wiley & Sons, Inc.; 2010.
23	28.	Walker JJ, Spiga F, Waite E, Zhao Z, Kershaw Y, Terry JR, Lightman SL. The origin of
24		glucocorticoid hormone oscillations. PLoS Biol 2012; 10:e1001341
25	29.	Samuels MH, Henry P, Luther M, Ridgway EC. Pulsatile TSH secretion during 48-hour
26		continuous TRH infusions. Thyroid 1993; 3:201-206
27	30.	Thomas GB, Cummins JT, Yao B, Gordon K, Clarke IJ. Release of prolactin is independent of
28		the secretion of thyrotrophin-releasing hormone into hypophysial portal blood of sheep. The
29		Journal of endocrinology 1988; 117:115-122
30	31.	Mershon JL, Sehlhorst CS, Rebar RW, Liu JH. Evidence of a corticotropin-releasing hormone
31		pulse generator in the macaque hypothalamus. Endocrinology 1992; 130:2991-2996
32	32.	Engler D, Pham T, Fullerton MJ, Ooi G, Funder JW, Clarke IJ. Studies of the secretion of
33		corticotropin-releasing factor and arginine vasopressin into the hypophysial-portal
34		circulation of the conscious sheep. I. Effect of an audiovisual stimulus and insulin-induced
35		hypoglycemia. Neuroendocrinology 1989; 49:367-381
36	33.	Ixart G, Barbanel G, Nouguier-Soule J, Assenmacher I. A quantitative study of the pulsatile
37		parameters of CRH-41 secretion in unanesthetized free-moving rats. Exp Brain Res 1991;
38		87:153-158
39	34.	Hoermann R, Midgley JE, Larisch R, Dietrich JW. Homeostatic Control of the Thyroid-Pituitary
40		Axis: Perspectives for Diagnosis and Treatment. Frontiers in endocrinology 2015; 6:1/7
41	35.	Campos P, Herbison AE. Optogenetic activation of GnRH neurons reveals minimal
4Z		requirements for pulsatile luteinizing normone secretion. Proceedings of the National
43	20	Academy of Sciences of the United States of America 2014; 111:18387-18392
44 15	36.	Han SY, MICLENNAN I, CZIESEISKY K, HERDISON AE. SELECTIVE OPTOGENETIC ACTIVATION OF ARCUATE
43 16		Risspeptin neurons generates puisatile internizing normone secretion. Proceedings of the
40 1.7	27	Rational Academy of Sciences of the Office States of America 2015, 112:15109-13114
т/ 48	57.	Tissier D. Bunn SI. Grattan DR. Mollard D. Martin AO. Diastisity of hypothalamic denomina
то 40		neurons during lactation results in dissociation of electrical activity and release. The Journal
50		of neuroscience - the official journal of the Society for Neuroscience 2013- 33-M2A-M33
		or near observer i the official joannal of the bolicity for mear observe 2013, 33,7727 7733

1	38.	Liu X, Brown RS, Herbison AE, Grattan DR. Lactational anovulation in mice results from a
2	••	selective loss of kisspeptin input to GnRH neurons. Endocrinology 2014; 155:193-203
3 ⊿	39.	Kokoris GJ, Lam NY, Ferin M, Silverman AJ, Gibson MJ. Transplanted gonadotropin-releasing
4 r		hormone neurons promote puisatile internizing normone secretion in congenitally
5	40	nypogonadai (npg) male mice. Neuroendocrinology 1988; 48:45-52
6 7	40.	Herbison AE, Porteous R, Pape JR, Mora JM, Hurst PR. Gonadotropin-releasing normone
/		neuron requirements for puberty, ovulation, and fertility. Endocrinology 2008; 149:597-604
8	41.	Romano N, Guillou A, Hodson DJ, Martin AO, Mollard P. Multiple-scale neuroendocrine
9 10		signals connect brain and pituitary normone rhythms. Proceedings of the National Academy
10	40	of Sciences of the United States of America 2017; 114:2379-2382
11	42.	Buzsaki G, Logothetis N, Singer W. Scaling brain size, keeping timing: evolutionary
12		preservation of brain rhythms. Neuron 2013; 80:751-764
13	43.	Ludwig M, Leng G. Dendritic peptide release and peptide-dependent behaviours. Nat Rev
14		Neurosci 2006; 7:126-136
15	44.	Stagkourakis S, Perez CT, Hellysaz A, Ammari R, Broberger C. Network oscillation rules
16		imposed by species-specific electrical coupling. eLife 2018; 7
17	45.	Constantin S. Progress and Challenges in the Search for the Mechanisms of Pulsatile
18		Gonadotropin-Releasing Hormone Secretion. Frontiers in endocrinology 2017; 8:180
19	46.	Campbell RE, Ducret E, Porteous R, Liu X, Herde MK, Wellerhaus K, Sonntag S, Willecke K,
20		Herbison AE. Gap junctions between neuronal inputs but not gonadotropin-releasing
21		hormone neurons control estrous cycles in the mouse. Endocrinology 2011; 152:2290-2301
22	47.	Zhang X, van den Pol AN. Dopamine/Tyrosine Hydroxylase Neurons of the Hypothalamic
23		Arcuate Nucleus Release GABA, Communicate with Dopaminergic and Other Arcuate
24		Neurons, and Respond to Dynorphin, Met-Enkephalin, and Oxytocin. The Journal of
25		neuroscience : the official journal of the Society for Neuroscience 2015; 35:14966-14982
26	48.	Stagkourakis S, Kim H, Lyons David J, Broberger C. Dopamine Autoreceptor Regulation of a
27		Hypothalamic Dopaminergic Network. Cell reports 2016; 15:735-747
28	49.	Moenter SM. Identified GnRH neuron electrophysiology: a decade of study. Brain Res 2010;
29		1364:10-24
30	50.	Plant TM, Krey LC, Moossy J, McCormack JT, Hess DL, Knobil E. The arcuate nucleus and the
31		control of gonadotropin and prolactin secretion in the female rhesus monkey (Macaca
32		mulatta). Endocrinology 1978; 102:52-62
33	51.	MacGregor DJ, Leng G. Modelling the hypothalamic control of growth hormone secretion.
34		Journal of neuroendocrinology 2005; 17:788-803
35	52.	Osterstock G, Mitutsova V, Barre A, Granier M, Fontanaud P, Chazalon M, Carmignac D,
36		Robinson IC, Low MJ, Plesnila N, Hodson DJ, Mollard P, Mery PF. Somatostatin triggers
37		rhythmic electrical firing in hypothalamic GHRH neurons. Sci Rep 2016; 6:24394
38	53.	Adams C, Stroberg W, DeFazio RA, Schnell S, Moenter SM. Gonadotropin-Releasing Hormone
39		(GnRH) Neuron Excitability Is Regulated by Estradiol Feedback and Kisspeptin. The Journal of
40		neuroscience : the official journal of the Society for Neuroscience 2018; 38:1249-1263
41	54.	Clasadonte J, Poulain P, Beauvillain JC, Prevot V. Activation of neuronal nitric oxide release
42		inhibits spontaneous firing in adult gonadotropin-releasing hormone neurons: a possible
43		local synchronizing signal. Endocrinology 2008; 149:587-596
44	55.	Alonso G, Sanchez-Hormigo A, Loudes C, El Yandouzi T, Carmignac D, Faivre-Bauman A,
45		Recolin B, Epelbaum J, Robinson IC, Mollard P, Mery PF. Selective alteration at the growth-
46		hormone- releasing-hormone nerve terminals during aging in GHRH-green fluorescent
47		protein mice. Aging Cell 2007; 6:197-207
48	56.	King JC, Rubin BS. Dynamic alterations in luteinizing hormone-releasing hormone (LHRH)
49		neuronal cell bodies and terminals of adult rats. Cellular and molecular neurobiology 1995:
50		15:89-106

1 57. Spiga F, Zavala E, Walker JJ, Zhao Z, Terry JR, Lightman SL. Dynamic responses of the adrenal 2 steroidogenic regulatory network. Proceedings of the National Academy of Sciences of the 3 United States of America 2017; 114:E6466-E6474 4 58. Wiersma J, Kastelijn J. A chronic technique for high frequency blood sampling/transfusion in 5 the freely behaving rat which does not affect prolactin and corticosterone secretion. The 6 Journal of endocrinology 1985; 107:285-292 7 59. Lee S, Young BM, Wan W, Chan IH, Privalsky ML. A mechanism for pituitary-resistance to 8 thyroid hormone (PRTH) syndrome: a loss in cooperative coactivator contacts by thyroid 9 hormone receptor (TR)beta2. Molecular endocrinology 2011; 25:1111-1125 10 60. Visser WE, Friesema EC, Visser TJ. Minireview: thyroid hormone transporters: the knowns 11 and the unknowns. Molecular endocrinology 2011; 25:1-14 12 61. Werneck de Castro JP, Fonseca TL, Ueta CB, McAninch EA, Abdalla S, Wittmann G, Lechan 13 RM, Gereben B, Bianco AC. Differences in hypothalamic type 2 deiodinase ubiquitination 14 explain localized sensitivity to thyroxine. J Clin Invest 2015; 125:769-781 15 62. Herman JP, McKlveen JM, Ghosal S, Kopp B, Wulsin A, Makinson R, Scheimann J, Myers B. 16 Regulation of the Hypothalamic-Pituitary-Adrenocortical Stress Response. Compr Physiol 17 2016; 6:603-621 18 63. Le Tissier PR, Hodson DJ, Lafont C, Fontanaud P, Schaeffer M, Mollard P. Anterior pituitary 19 cell networks. Front Neuroendocrinol 2012; 33:252-266 20 64. Sanchez-Cardenas C, Fontanaud P, He Z, Lafont C, Meunier AC, Schaeffer M, Carmignac D, 21 Molino F, Coutry N, Bonnefont X, Gouty-Colomer LA, Gavois E, Hodson DJ, Le Tissier P, 22 Robinson IC, Mollard P. Pituitary growth hormone network responses are sexually dimorphic 23 and regulated by gonadal steroids in adulthood. Proceedings of the National Academy of 24 Sciences of the United States of America 2010; 107:21878-21883 25 65. Hodson DJ, Romano N, Schaeffer M, Fontanaud P, Lafont C, Fiordelisio T, Mollard P. 26 Coordination of calcium signals by pituitary endocrine cells in situ. Cell calcium 2012; 51:222-27 230 28 66. Mollard P, Hodson DJ, Lafont C, Rizzoti K, Drouin J. A tridimensional view of pituitary 29 development and function. Trends in endocrinology and metabolism: TEM 2012; 23:261-269 30 67. Budry L, Lafont C, El Yandouzi T, Chauvet N, Conejero G, Drouin J, Mollard P. Related 31 pituitary cell lineages develop into interdigitated 3D cell networks. Proceedings of the 32 National Academy of Sciences of the United States of America 2011; 108:12515-12520 33 68. Bonnefont X, Lacampagne A, Sanchez-Hormigo A, Fino E, Creff A, Mathieu MN, Smallwood S, 34 Carmignac D, Fontanaud P, Travo P, Alonso G, Courtois-Coutry N, Pincus SM, Robinson IC, 35 Mollard P. Revealing the large-scale network organization of growth hormone-secreting 36 cells. Proceedings of the National Academy of Sciences of the United States of America 2005; 37 102:16880-16885 38 69. Chauvet N, El-Yandouzi T, Mathieu MN, Schlernitzauer A, Galibert E, Lafont C, Le Tissier P, 39 Robinson IC, Mollard P, Coutry N. Characterization of adherens junction protein expression 40 and localization in pituitary cell networks. The Journal of endocrinology 2009; 202:375-387 41 70. Chauvet N, Romano N, Meunier AC, Galibert E, Fontanaud P, Mathieu MN, Osterstock G, 42 Osterstock P, Baccino E, Rigau V, Loiseau H, Bouillot-Eimer S, Barlier A, Mollard P, Coutry N. 43 Combining Cadherin Expression with Molecular Markers Discriminates Invasiveness in 44 Growth Hormone and Prolactin Pituitary Adenomas. Journal of neuroendocrinology 2016; 45 28:12352 46 71. Milo R, Shen-Orr S, Itzkovitz S, Kashtan N, Chklovskii D, Alon U. Network motifs: simple 47 building blocks of complex networks. Science 2002; 298:824-827 48 72. Alon U. Network motifs: theory and experimental approaches. Nat Rev Genet 2007; 8:450-49 461 50 73. Waite E, Lafont C, Carmignac D, Chauvet N, Coutry N, Christian H, Robinson I, Mollard P, Le 51 Tissier P. Different degrees of somatotroph ablation compromise pituitary growth hormone

1 2		cell network structure and other pituitary endocrine cell types. Endocrinology 2010; 151:234-243
3 1	74.	Hodson DJ, Schaeffer M, Romano N, Fontanaud P, Lafont C, Birkenstock J, Molino F, Christian
5		lasting experience-dependent plasticity in endocrine cell networks. Nat Commun 2012;
6		3:605
7	75.	Byrnes EM, Bridges RS. Lactation reduces prolactin levels in reproductively experienced
8		female rats. Horm Behav 2005; 48:278-282
9	76.	Byrnes EM, Bridges RS. Reproductive experience and expression of dopamine D(2) receptor
10 11		mRNA: a possible mechanism for reduced prolactin secretion in primiparous rats. Journal of neuroendocrinology 2007; 19:773-778
12	77.	Ward RD, Stone BM, Raetzman LT, Camper SA. Cell proliferation and vascularization in
13		mouse models of pituitary hormone deficiency. Molecular endocrinology 2006; 20:1378-
14		1390
15	78.	Porter JC, Reymond MJ, Arita J, Sissom JF. Secretion of hypothalamic dopamine into the
16		hypophysial portal vasculature: an overview. Methods Enzymol 1983; 103:607-618
17 18	79.	Adams JH, Daniel PM, Prichard MML. Observations on the Portal Circulation of the Pituitary
10	<u>ە</u> م	Lafont C. Decarmonion MG. Cassou M. Molino E. Lacon L. Hodson D. Lacampagno A
20	80.	Monnossior G. El Vandouzi T. Carmignas D. Fontanaud D. Christian H. Coutry N. Fornandoz
20 21		Function M. Charmark S. La Tissiar D. Pohlanan IC. Mallard D. Collular in vivo imaging royaals
21 22		coordinated regulation of nituitany microcirculation and GH cell network function
22 73		Proceedings of the National Academy of Sciences of the United States of America 2010:
23 24		
2 1 25	Q1	Schaeffer M. Langlet F. Lafont C. Molino F. Hodson DI. Roux T. Lamarque I. Verdie P.
26	01.	Bourrier F. Debouck B. Baneres II. Martinez I. Mery PF. Marie I. Trinquet F. Febrentz IA
27		Prevot V Mollard P. Banid sensing of circulating ghrelin by hypothalamic appetite-modifying
28		neurons. Proceedings of the National Academy of Sciences of the United States of America
29		2013; 110:1512-1517
30	82.	Featherstone K, Hey K, Momiji H, McNamara AV, Patist AL, Woodburn J, Spiller DG, Christian
31		HC, McNeilly AS, Mullins JJ, Finkenstadt BF, Rand DA, White MR, Davis JR. Spatially
32		coordinated dynamic gene transcription in living pituitary tissue. eLife 2016; 5
33	83.	Bargi-Souza P, Kucka M, Bjelobaba I, Tomic M, Janjic MM, Nunes MT, Stojilkovic SS. Loss of
34		basal and TRH-stimulated Tshb expression in dispersed pituitary cells. Endocrinology 2015;
35		156:242-254
36	84.	Denef C. Paracrinicity: the story of 30 years of cellular pituitary crosstalk. Journal of
37		neuroendocrinology 2008; 20:1-70
38	85.	Fink G. 60 YEARS OF NEUROENDOCRINOLOGY: MEMOIR: Harris' neuroendocrine revolution:
39		of portal vessels and self-priming. The Journal of endocrinology 2015; 226:T13-24
40	86.	Navratil AM, Knoll JG, Whitesell JD, Tobet SA, Clay CM. Neuroendocrine plasticity in the
41		anterior pituitary: gonadotropin-releasing hormone-mediated movement in vitro and in
42		vivo. Endocrinology 2007; 148:1736-1744
43	87.	Alim Z, Hartshorn C, Mai O, Stitt I, Clay C, Tobet S, Boehm U. Gonadotrope plasticity at
44		cellular and population levels. Endocrinology 2012; 153:4729-4739
45	88.	Romano N, McClafferty H, Walker JJ, Le Tissier P, Shipston MJ. Heterogeneity of Calcium
46		Responses to Secretagogues in Corticotrophs From Male Rats. Endocrinology 2017;
47	• -	158:1849-1858
48	89.	Johnston NR, Mitchell RK, Haythorne E, Pessoa MP, Semplici F, Ferrer J, Piemonti L,
49 50		Marchetti P, Bugliani M, Bosco D, Berishvili E, Duncanson P, Watkinson M, Broichhagen J,
50 51		I rauner D, Kutter GA, Hodson DJ. Beta Cell Hubs Dictate Pancreatic Islet Responses to
51		Glucose. Cell Metab 2016; 24:389-401

1 90. Schilbach K, Bidlingmaier M. Growth hormone binding protein - physiological and analytical 2 aspects. Best Pract Res Clin Endocrinol Metab 2015; 29:671-683 3 91. van den Berg G, Frolich M, Veldhuis JD, Roelfsema F. Combined amplification of the pulsatile 4 and basal modes of adrenocorticotropin and cortisol secretion in patients with Cushing's 5 disease: evidence for decreased responsiveness of the adrenal glands. J Clin Endocrinol 6 Metab 1995; 80:3750-3757 7 92. van den Berg G, Pincus SM, Veldhuis JD, Frolich M, Roelfsema F. Greater disorderliness of 8 ACTH and cortisol release accompanies pituitary-dependent Cushing's disease. Eur J 9 Endocrinol 1997; 136:394-400 10 Roelfsema F, Keenan DM, Veldhuis JD. Endogenous ACTH concentration-cortisol secretion 93. 11 dose analysis unmasks decreased ACTH potency in Cushing's disease with restoration after 12 successful pituitary adenomectomy. J Clin Endocrinol Metab 2011; 96:3768-3774 13 94. Roelfsema F, Pereira AM, Biermasz NR, Veldhuis JD. Hormone secretion by pituitary 14 adenomas is characterized by increased disorderliness and spikiness but more regular 15 pulsing. J Clin Endocrinol Metab 2014; 99:3836-3844 16 95. Roland AV, Moenter SM. Reproductive neuroendocrine dysfunction in polycystic ovary 17 syndrome: insight from animal models. Front Neuroendocrinol 2014; 35:494-511 18 96. Moore AM, Campbell RE. Polycystic ovary syndrome: Understanding the role of the brain. 19 Front Neuroendocrinol 2017; 46:1-14 20 97. Baskind NE, Balen AH. Hypothalamic-pituitary, ovarian and adrenal contributions to 21 polycystic ovary syndrome. Best Pract Res Clin Obstet Gynaecol 2016; 37:80-97 22 98. Brothers KJ, Wu S, DiVall SA, Messmer MR, Kahn CR, Miller RS, Radovick S, Wondisford FE, 23 Wolfe A. Rescue of obesity-induced infertility in female mice due to a pituitary-specific 24 knockout of the insulin receptor. Cell Metab 2010; 12:295-305 25 99. Leow MK. A Review of the Phenomenon of Hysteresis in the Hypothalamus-Pituitary-Thyroid 26 Axis. Frontiers in endocrinology 2016; 7:64 27 100. Herwig A, Campbell G, Mayer CD, Boelen A, Anderson RA, Ross AW, Mercer JG, Barrett P. A 28 thyroid hormone challenge in hypothyroid rats identifies T3 regulated genes in the 29 hypothalamus and in models with altered energy balance and glucose homeostasis. Thyroid 30 2014; 24:1575-1593 31 101. Roelfsema F, Pereira AM, Adriaanse R, Endert E, Fliers E, Romijn JA, Veldhuis JD. Thyrotropin 32 secretion in mild and severe primary hypothyroidism is distinguished by amplified burst 33 mass and Basal secretion with increased spikiness and approximate entropy. J Clin 34 Endocrinol Metab 2010; 95:928-934 35 102. Betley JN, Xu S, Cao ZF, Gong R, Magnus CJ, Yu Y, Sternson SM. Neurons for hunger and 36 thirst transmit a negative-valence teaching signal. Nature 2015; 521:180-185 37 103. Pinol RA, Jameson H, Popratiloff A, Lee NH, Mendelowitz D. Visualization of oxytocin release 38 that mediates paired pulse facilitation in hypothalamic pathways to brainstem autonomic 39 neurons. PloS one 2014; 9:e112138 40 104. Hattori M, Ozawa T. Split luciferase complementation for analysis of intracellular signaling. 41 Anal Sci 2014; 30:539-544 42 105. Liu Y, Berry PA, Zhang Y, Jiang J, Lobie PE, Paulmurugan R, Langenheim JF, Chen WY, Zinn KR, 43 Frank SJ. Dynamic analysis of GH receptor conformational changes by split luciferase 44 complementation. Molecular endocrinology 2014; 28:1807-1819 45 106. Manfredi-Lozano M, Roa J, Ruiz-Pino F, Piet R, Garcia-Galiano D, Pineda R, Zamora A, Leon S, 46 Sanchez-Garrido MA, Romero-Ruiz A, Dieguez C, Vazquez MJ, Herbison AE, Pinilla L, Tena-47 Sempere M. Defining a novel leptin-melanocortin-kisspeptin pathway involved in the 48 metabolic control of puberty. Mol Metab 2016; 5:844-857 49 107. Swiech L, Heidenreich M, Banerjee A, Habib N, Li Y, Trombetta J, Sur M, Zhang F. In vivo 50 interrogation of gene function in the mammalian brain using CRISPR-Cas9. Nat Biotechnol 51 2015; 33:102-106

1	108.	Villani AC, Satija R, Reynolds G, Sarkizova S, Shekhar K, Fletcher J, Griesbeck M, Butler A,
2		Zheng S, Lazo S, Jardine L, Dixon D, Stephenson E, Nilsson E, Grundberg I, McDonald D, Filby
3		A, Li W, De Jager PL, Rozenblatt-Rosen O, Lane AA, Haniffa M, Regev A, Hacohen N. Single-
4		cell RNA-seq reveals new types of human blood dendritic cells, monocytes, and progenitors.
5		Science 2017; 356
6	109.	Morel L, Chiang MSR, Higashimori H, Shoneye T, Iyer LK, Yelick J, Tai A, Yang Y. Molecular
7		and Functional Properties of Regional Astrocytes in the Adult Brain. The Journal of
8		neuroscience : the official journal of the Society for Neuroscience 2017; 37:8706-8717
9	110.	Bloomfield FH, Oliver MH, Giannoulias CD, Gluckman PD, Harding JE, Challis JR. Brief
10		undernutrition in late-gestation sheep programs the hypothalamic-pituitary-adrenal axis in
11		adult offspring. Endocrinology 2003; 144:2933-2940
12	111.	Chauvet N, Romano N, Lafont C, Guillou A, Galibert E, Bonnefont X, Le Tissier P, Fedele M,
13		Fusco A, Mollard P, Coutry N. Complementary actions of dopamine D2 receptor agonist and
14		anti-vegf therapy on tumoral vessel normalization in a transgenic mouse model. Int J Cancer
15		2017; 140:2150-2161
16	112.	Ben-Jonathan N, LaPensee CR, LaPensee EW. What can we learn from rodents about
17		prolactin in humans? Endocrine reviews 2008; 29:1-41
18	113.	Zimmer B, Piao J, Ramnarine K, Tomishima MJ, Tabar V, Studer L. Derivation of Diverse
19		Hormone-Releasing Pituitary Cells from Human Pluripotent Stem Cells. Stem Cell Reports
20		2016; 6:858-872
21	114.	Alatzoglou KS, Webb EA, Le Tissier P, Dattani MT. Isolated growth hormone deficiency (GHD)
22		in childhood and adolescence: recent advances. Endocrine reviews 2014; 35:376-432
23	115.	Demene C, Mairesse J, Baranger J, Tanter M, Baud O. Ultrafast Doppler for neonatal brain
24		imaging. Neuroimage 2018;
25	116.	Andoniadou CL, Matsushima D, Mousavy Gharavy SN, Signore M, Mackintosh AI, Schaeffer
26		M, Gaston-Massuet C, Mollard P, Jacques TS, Le Tissier P, Dattani MT, Pevny LH, Martinez-
27		Barbera JP. Sox2(+) stem/progenitor cells in the adult mouse pituitary support organ
28		homeostasis and have tumor-inducing potential. Cell stem cell 2013; 13:433-445
29	117.	Rizzoti K, Akiyama H, Lovell-Badge R. Mobilized adult pituitary stem cells contribute to
30		endocrine regeneration in response to physiological demand. Cell stem cell 2013; 13:419-
31		432
32	118.	Suga H, Kadoshima T, Minaguchi M, Ohgushi M, Soen M, Nakano T, Takata N, Wataya T,
33		Muguruma K, Miyoshi H, Yonemura S, Oiso Y, Sasai Y. Self-formation of functional
34		adenohypophysis in three-dimensional culture. Nature 2011; 480:57-62
35	119.	Dincer Z, Piao J, Niu L, Ganat Y, Kriks S, Zimmer B, Shi SH, Tabar V, Studer L. Specification of
36		functional cranial placode derivatives from human pluripotent stem cells. Cell reports 2013;
37		5:1387-1402
20		

1 Figure legends:

2 Figure 1. A simplified schematic showing the contrasting regulation of pituitary hormone 3 pulse generation between the hypothalamic-pituitary-gonadal (HPG) and hypothalamic-4 pituitary-adrenal (HPA) axes. In the HPG axis, the pulse generator is localised in the 5 hypothalamus, where afferent inputs from kisspeptin neurons (25) and neuronal feedforward 6 loops (53) lead to pulsatile release of gonadotrophin releasing hormone (GnRH). This results on 7 the release of pulses of luteinizing (LH) and follicle stimulating (FSH) hormone, stimulating 8 secretion of steroids from the gonads which feedback on a relatively slow timescale to both the 9 pituitary and hypothalamus. In contrast, in the HPA axis the rapid actions of adrenocorticotrophic hormone on the adrenal gland and delayed feedback of glucocorticoids on 10 11 the anterior pituitary is the source of pulse generation, with corticotrophin releasing hormone 12 (CRH) having a modulatory role (28). 13 14 Figure 2. The growth hormone (GH) cell network and its relationship with the 15 **vasculature.** Two-photon imaging of the pituitary of a GH-GFP transgenic mouse with

capillaries labelled with gelatine-rhodamine (red). GH cells are organised into a homotypic
topologically organised network of cell clusters which are linked by strings of single cells. The
cell network is closely associated with capillaries, which are aligned with strings of cells and
surround the clusters.



