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Diagnostic accuracy, risk assessment, and cost-effectiveness of component-resolved diagnostics for food allergy: a systematic review

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Abstract

Background: Component-resolved diagnostics (CRD) are promising tools for diagnosing food allergy, offering the potential to determine specific phenotypes and to develop patient-tailored risk profiles. Nevertheless, the diagnostic accuracy of these tests varies across studies; thus, their clinical utility remains unclear. Therefore, we synthesized the evidence from studies investigating the diagnostic accuracy, risk assessment ability, and cost-effectiveness of CRD for food allergy.

Methods: We systematically searched 10 electronic databases and four clinical trial registries for studies published January 2000-February 2017. The quality of included studies was assessed using QUADAS-2. Due to heterogeneity, we narratively synthesized the evidence.

Results: Eleven studies met inclusion criteria, altogether recruiting 1,098 participants. The food allergies investigated were cow's milk, hen's egg, peanut, hazelnut, and shrimp. The components with the highest diagnostic accuracy for each allergen, along with their sensitivity-specificity pairs, were: Bos d 4 for cow's milk (62.0% and 87.5%), Gal d 1 for hen's egg (84.2% and 89.8% for heated egg, and 60.6% and 97.1% for raw egg), Ara h 2 for peanut (80.3% and 95.1%), Cor a 14 for hazelnut (100% and 93.8%), and Lit v 1 for shrimp (82.8% and 56.3%) allergy.

Conclusion: Selected components of cow's milk, hen's egg, peanut, hazelnut, and shrimp allergen showed high specificity, but lower sensitivity. However, few studies exist for each component, and studies vary widely regarding the cut-off values used, making it challenging to synthesize findings across studies. Further research is needed to determine clinically appropriate cut-off values, risk assessment abilities, and cost-effectiveness of CRD approaches.

Key words

Component-resolved diagnostics, cost-effectiveness, diagnostic test accuracy systematic review, food allergy, risk assessment.

Abbreviations

aa	Amino acid
APT	Atopy patch test
CRD	Component-resolved diagnostics
DBPCFC	Double-blind, placebo controlled food challenge
DTA	Diagnostic test accuracy
HSROC	Hierarchical summary receiver operating characteristic
IgE	Immunoglobulin E

sIgE	Specific-immunoglobulin E
NPV	Negative predictive value
PPV	Positive predictive value
QUADAS-2	Quality Assessment of Diagnostic Accuracy Studies-2
ROB	Risk of bias
ROC	Receiver operating characteristic
SPT	Skin prick test

Introduction

The high prevalence of food allergy is now an emerging global public health concern (1). Estimates of the prevalence of food allergy vary, but overall lifetime prevalence has been estimated to be between 4% and 7% for children and between 3% and 6% for adults in economically-developed countries (2, 3). The quality of life of patients with food allergy is often severely affected, resulting in considerable morbidity and healthcare utilization, including risk of accidental exposure leading to life-threatening anaphylactic reactions (4).

An accurate diagnosis of food allergy is essential to provide appropriate, potentially life-saving advice on how to prevent and manage allergic reactions and prevent unnecessary dietary restrictions (1, 4). The diagnosis of food allergy is dependent on a thorough clinical history as well as an objective marker of allergic sensitization and, in some cases, oral food challenge tests (5). Current first-line tests to assess allergen sensitization are skin prick tests (SPT) and/or immunoassays of serum food specific-IgE (sIgE) levels. However, these approaches have a high rate of false positive results and are poor predictors of the severity of allergic reactions (4). Thus, diagnostic confirmation with (ideally) a double-blinded placebo-controlled food challenge (DBPCFC) is often required (5). While DBPCFCs are considered the gold standard diagnostic tests, they are costly, technically challenging, time-consuming, labor-intensive, and are associated with important safety risks, as they can trigger anaphylactic reactions (4).

Given the limitations of conventional methods for diagnosing food allergy, new molecular-based diagnostic techniques—collectively referred to as component-resolved diagnostics (CRD)—have emerged as promising diagnostic tools (6). Whilst current approaches evaluate patients' reactivity to whole food extracts, CRD involves detecting sIgE levels to individual allergenic molecules or the epitopes of those allergens (7). This approach may enhance determination of specific food allergy phenotypes, assist in the development of patient-tailored risk profiles for specific food allergens, and improve detection of possible cases of cross-reactivity between different allergenic molecules (8).

Over the last decade, researchers have compared CRD to conventional diagnostic approaches for food allergy (9). Through this work, the major allergen components in different food allergies have been identified. However, the diagnostic accuracy of identified components varies across studies, and thus the diagnostic value and clinical utility of CRD remains unclear (9, 10). CRD approaches are also expensive, which raises questions about their cost-effectiveness (11).

While the diagnostic accuracy of various tests for food allergy was evaluated in our previous systematic review, CRD was not included (12). A health technology assessment was carried out to evaluate multiplex CRD assays, but clinical effectiveness (rather than diagnostic accuracy) was investigated (13). To the best of our knowledge, only one CRD-specific diagnostic test accuracy (DTA) review has been conducted, but it focused solely on peanut allergy diagnosis (14). This review concluded that Ara h 2 showed superior diagnostic accuracy than SPT and sIgE tests, and therefore has the potential to replace first-line tests for the diagnosis of peanut allergy. Given the increasing body of work, there is a need to undertake a more comprehensive evidence synthesis on the diagnostic accuracy of CRD. We therefore conducted a systematic review to: i). determine the accuracy of CRD for the diagnosis of food allergy, focusing on the 'big eight' food allergies (i.e. cow's milk, wheat, hen's egg, peanut, soy, tree nuts, fish, and shellfish allergy); ii). estimate the effectiveness and cost-effectiveness of CRD in comparison to conventional techniques for the diagnosis of these food allergies; and iii). summarize the evidence on the ability of CRD to predict the severity of allergic reactions. We focused on these eight food allergies in order to align with the foods considered in our previous systematic reviews for the European Academy of Allergy and Clinical Immunology (2, 12).

Methods

The Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) checklist guided the reporting of this systematic review (see online supplement Table S1) (15). Our protocol was published (16) and pre-registered (PROSPERO:CRD42016053512).

Eligibility criteria

We included prospective, retrospective, cross-sectional, and case-control studies that examined the accuracy of CRD in diagnosing cow's milk, hen's egg, wheat, soybean, peanut, tree nuts, fish, or shellfish allergy in children or adults. Studies were required to have sufficient data to calculate the following four relevant diagnostic measures: sensitivity, specificity, positive predicted value (PPV), and negative predictive value (NPV). Additionally, all studies were required to have a defined study population with either consecutive or random sampling of participants. Studies in which the recruitment technique used to select participants was not indicated were included, and the lack of information regarding their sampling methodology was noted during the quality assessment process. The reference standard was DBPCFC used in at least 50% of the participants.

Search strategy

Although CRD methods were originally described in the 1990s (17), their application to food allergy diagnosis was not clinically implemented until the 2000s (1). Hence, we chose the beginning of 2000 as the starting time for the literature search. We searched the following databases from January 2000 to February 2017: AMED (Ovid), CAB Abstracts (Ovid), the Cochrane Library, CINAHL (EBSCO), Embase (Ovid), Global Health (Ovid), PsycINFO (Ovid), Web of Science Core Collection (Thomson Reuters), WHO's Global Health Library and the Health Economic Evaluations Database. Our full search strategy is included in the online supplement (S2). We also contacted international experts who have published in the field, screened the references cited in identified studies, and used the citation-tracking feature of Google Scholar to find any additional studies. The list of contacted experts can be found in the online supplement (S3). Additionally, the International Standard Randomized Controlled Trial Number (ISRCTN) Registry, ClinicalTrials.gov, the Australian and New Zealand Clinical Trials Registry, and WHO's International Clinical Trials Registry Platform (ICTRP) were searched to identify relevant on-going studies. No language restrictions were applied.

Study selection and data collection

Two reviewers (JFK and NM) independently screened titles and abstracts and then reviewed full-texts to identify eligible studies. Authors of studies for which further details were required to determine inclusion or exclusion were contacted to obtain further information to enable a decision. For papers in languages other than English, speakers of the language in question were contacted to determine eligibility. Both reviewers independently extracted data from included studies using a form developed specifically for this systematic review. Study characteristics, DTA measures (i.e. sensitivity, specificity, PPV and NPV) and 2x2 contingency tables (reflecting the number of true positives, true negatives, false positives, and false negatives) were extracted. DTA measures and 95% confidence intervals were calculated from 2x2 contingency tables when not provided by authors (18-20). The two reviewers assessed the quality of the included studies using the Quality Assessment of Diagnostic Accuracy Studies (QUADAS-2) tool (21). Discrepancies were resolved by discussion and consensus, or when necessary, arbitrated by a third reviewer (BN).

Data synthesis, analysis, and reporting

Diagnostic accuracy measures (sensitivity, specificity, PPV and NPV) of individual studies were summarized in tables, and presented by allergy type and individual allergen component. We had planned to conduct meta-analyses of the evidence with respect to each allergen component by fitting a bivariate model (when included studies used a common threshold) or a hierarchical summary receiver operating characteristic (HSROC) model (when included studies used multiple thresholds). However, we were unable to do this, as the number of studies for each component was too small to permit quantitative syntheses. In a simulated analysis based on the Bayesian approach, it was recommended that a minimum of four studies were required in order to reasonably fit these models (22). In a very few cases we had a maximum of three studies per allergen component; the most common was two studies. The statistical programs we tried to use to fit the models (R and Stata) indicated that the models lacked convergence, as a result of containing too few studies. For these reasons, we narratively synthesized the evidence.

Results

Study selection

A total of 10,380 articles were identified through the literature search carried out on June 15, 2016. After excluding duplicate articles, 6,853 titles and abstracts were screened against the inclusion and exclusion criteria; of these, 195 full-text papers were assessed. Thirteen articles reporting 11 studies met our criteria and were thus included (23-35). Additionally, one potentially relevant on-going clinical trial was found in ClinicalTrials.gov (details can be found in Table S4). The literature search was updated on February 9, 2017 to incorporate newly published papers. No additional relevant studies were identified in the updated search. The study screening and selection processes are summarized in Figure 1. A list of potentially relevant studies can be found in the Online supplement (S5 and S6). The authors of these studies did not reply to a request for further information.

Characteristics of included studies

Table 1 summarizes the main characteristics of the 11 studies included. The studies altogether recruited a total of 1,098 participants. Nine studies were carried out in Western Europe (23-28, 30, 32, 33), and two multi-center studies analyzed data from multiple countries (31, 34). Two studies used a case-control design (32, 34), while nine were cross-sectional studies (23-28, 30, 31, 33). Two of the cross-sectional studies used consecutive sampling to recruit participants (26, 28), while the sampling strategy used in the remaining seven cross-sectional studies was unclear (23-25, 27, 30, 31, 33). Taking all studies together, 87% of participants underwent DBPCFC to verify their food allergy status.

The included studies analyzed the diagnostic accuracy of CRD for the following types of food allergy: cow's milk (n=2) (23, 33), hen's egg (n=3) (24, 25, 33), peanut (n=3) (28, 30, 31), hazelnut (n=2) (27, 32), and shrimp (n=2) (26, 34). No studies investigated the other allergies of interest (i.e. wheat, soybean, and fish). All but one study analyzed a single type of allergy; this study analyzed the diagnostic accuracy of CRD for both cow's milk and hen's egg (33). Five studies used the ImmunoCAP test to measure sIgE levels (23, 25-27, 31, 32), three studies used microarray techniques (ISAC CRD 51, ISAC 103, and ISAC 112) (24, 28, 33), one study used a combination of ImmunoCAP and a microarray technique (ISAC 112) (30), and one study used an immunoblotting technique for sIgE detection (34). Two studies additionally analyzed the DTA of individual component epitopes using immunoblotting techniques (26, 34).

Quality assessment of included studies

Table 1 includes a summary of the QUADAS-2 quality assessment for each study. Table S7 in the supplementary information provides the detailed QUADAS-2 assessments.

Patient selection

Two studies were rated as high risk of bias (ROB) in this domain because of the use of a case-control design (32, 34). Eight other studies were found to have an unclear ROB, mainly because they did not explicitly indicate their sampling methodology and/or did not avoid inappropriate exclusions (23-25, 27, 28, 30, 31, 33). The remaining study had a low ROB (26).

Index test

Two studies had high ROB in this domain because they did not use pre-specified thresholds for determining positive results (27, 33). Eight studies had unclear ROB because they did not report whether index test results were interpreted without knowledge of DBPCFC results (23-26, 28, 31, 32, 34). Only one study had low ROB in this domain (30). Two studies used immunoblotting assays to analyze sIgE reactivity against allergens; because these assays do not provide quantitative sIgE levels, these two studies were scored as unsure in terms of their applicability to this review's research question (26, 34).

Reference standard

One study did not specify the criteria used to classify DBPCFC results and was thus scored as having an unclear ROB in this domain (26). Two studies were scored as unsure in terms of their applicability to this review's research question (30, 32). The reason for this appraisal was that the purpose of this review was to assess the accuracy of CRD and its ability to predict allergy severity (both of which are assessed through objective symptoms in a DBPCFC), and the two aforementioned studies included patients with mild or subjective DBPCFC symptoms in the same group as patients with negative DBPCFC.

Patient flow and timing

Two studies had a high ROB in this domain because less than 100% of patients underwent DBPCFCs, and not all patient data were included in their data analysis (26, 34). Three studies specified the time interval between index and reference tests, compared all patients against the same reference standard, and included all patients in data analysis, and were thus ranked as low ROB in this domain (24, 25, 27). The remaining six studies failed to meet at least one of those criteria, and were thus scored as having an unclear ROB (23, 28, 30-33).

Diagnostic accuracy of CRD

DTA measures for all the 11 studies are presented in Table 2. The information in this table includes data points for all sIgE cut-off values that the included studies used to define test positivity, as some studies used multiple values. In the following narrative synthesis, we present the results of the diagnostic accuracy of all components per food allergy type for only cut-off values with the highest diagnostic potential as defined in each study.

Cow's milk allergy

Two studies evaluated CRD for cow's milk allergy (23, 33), and the following components were assessed: Bos d 4 (α -lactalbumin), Bos d 5 (β -lactoglobulin), Bos d 8 (caseins), and the caseins (α -, β -, and κ -) separately. The reported sensitivity-specificity for these components were: for Bos d 4, 62.0% and 87.5% (with a cut-off values defining a positive test of >0.01 kUa/L) (23), and 50.0% and 93.0% (at >0.1 FI) (33); for Bos d 5, 82.0% and 62.5% (at >0.35 kUa/L) (23), and 23.8% and 95.3% (at >0.1 FI) (33); for Bos d 8, 88.0% and 56.3% (at >0.35 kUa/L) (23); and the casein with the highest DTA was κ -casein with a sensitivity-specificity pair of 38.1% and 88.4% (at >0.1 FI) (33).

Hen's egg allergy

Three studies evaluated CRD for hen's egg allergy (24, 25, 33), and the following components were assessed: Gal d 1 (ovomucoid), Gal d 2 (ovalbumin), Gal d 3 (ovotransferrin), and Gal d 4 (lysozyme). Two studies investigated heated egg and raw egg allergy separately (24, 25), while the third study analyzed only raw egg allergy (33). For heated egg allergy, the reported sensitivity-specificity for these components were: for Gal d 1, 84.2% and 89.8% (at >0.01 kUa/L) (24), and 76.3% and 81.4% (at >4.4 kUa/L) (25); for Gal d 2, 52.6% and 83.7% (at >0.01 kUa/L) (24), and 73.7% and 72.9% (at >6.33 kUa/L);(25) and for Gal d 3, 21.1% and 93.9% (at >0.01 kUa/L) (24). For raw egg allergy, the reported sensitivity-specificity for these components were: for Gal d 1, 60.6% and 97.1% (at >0.01 kUa/L) (24), 73.1% and 82.9% (at >2.26 kUa/L) (25), and 57.8% and 86.7% (at >0 FI) (33); for Gal d 2, 42.4% and 88.6% (at >0.01 kUa/L) (24), and 76.1% and 82.9% (at >3.88 kUa/L) (25), and 57.8% and 80.0% (at >0 FI) (33); for Gal d 3, 18.2% and 97.1% (at >0.01 kUa/L) (24); and for Gal d 4, 17.8% and 100% (at >0 FI) (33).

Peanut allergy

Three studies evaluated the diagnostic accuracy of CRD for peanut allergy (28, 30, 31), and the following components were assessed: Ara h 1 (cupin, a 7S globulin), Ara h 2 (conglutinin, a 2S albumin), Ara 3 (cupin, a 11S globulin), Ara h 6 (conglutinin, a 2S albumin), Ara h 8 (Bet v 1 homologue), and Ara h 9 (LTP). The reported sensitivity-specificity for these components were: for Ara h 1, 56.6% and 86.9% (at >0.35 kUa/L) (31), and 60.7% and 95.1% (at >0.8 kUa/L) (30); for Ara h 2, 69.2% and 90.5% (at >0.3 ISU/L) (28), 80.2% and 91.8% (at >0.35 kUa/L) (31), and 80.3% and 95.1% (at >1.8 kUa/L) (30); for Ara h 3, 48.1% and 90.2% (at >0.35 kUa/L) (31), and 55.7% and 95.1% (at >0.8 kUa/L) (30); for Ara h 6, 61.5% and 95.2% (at >1.0 ISU/L) (28); for Ara h 8, 34.9% and 42.6% (at >0.35 kUa/L) (31), and 78.7% and 14.6% (at >0.35 kUa/L) (30); and for Ara h 9, 14.8% and 85.4% (at >0.35 kUa/L) (30).

Hazelnut allergy

Two studies evaluated the diagnostic accuracy of CRD for hazelnut allergy (27, 32), and the following components were assessed: Cor a 1 (PR-10 protein), Cor a 8 (LTP), Cor a 9 (11S seed storage globulin), and Cor a 14 (2S albumin). Additionally, one of the studies investigated whether sensitization to the allergens Bet v 1 (PR-10 protein) and Bet v 2 (profilin) (from the European White Birch) could also predict hazelnut allergy (32). The reported sensitivity-specificity for these

components were: for Cor a 1, 79.7% and 7.3% (at >0.35 kUa/L (32); for Cor a 8, 6.3% and 96.3% (at >0.35 kUa/L) (32); for Cor a 9, 100% and 71.9% (at >0.65 kUa/L) (27), and 54.4% and 97.6% (at >1 kUa/L) (32); for Cor a 14, 100% and 93.8% (at >0.64 kUa/L) (27), and 54.4% and 85.4% (at >0.35 kUa/L) (32); for Bet v 1, 81.0% and 7.3% (at 0.35 kUa/L) (32); and for Bet v 2, 10.1% and 81.7% (at 0.35 kUa/L) (32).

Shrimp allergy

Two studies reported data on CRD for shrimp allergy (26, 34). One study tested the component Pen a 1 (tropomyosin) using the ImmunoCAP test (26), and the other study investigated the components Lit v 1 (tropomyosin) and Lit v 4 (sarcoplasmic calcium-binding protein) through an immunoblotting technique (34). Additionally, both studies investigated the diagnostic value of several individual epitopes in shrimp through an immunoblotting technique. The components (and their epitopes) tested in this manner were: Lit v 1, Lit v 2 (arginine kinase), Lit v 3 (myosin light chain), and Lit v 4. The reported sensitivity-specificity pair of Lit v 1 was 82.8% and 56.3%, and of Lit v 4 was 34.5% and 93.8% (34), and the reported sensitivity-specificity pair of Pen a 1 was 88.2% and 23.8% (at >0.35 kUa/L) (26). With regards to epitope data, the epitopes with highest DTA were found on Lit v 1 and Lit v 2 (26, 34). Table S8 in the online supplement presents the full DTA data by epitope.

Cost-effectiveness of CRD

None of the studies meeting our inclusion criteria evaluated the cost-effectiveness of CRD or made mention of any economic considerations. The only relevant evidence identified were two manufacturer-authored abstracts featuring Markov simulation-based cost-utility models for CRD versus DBPCFC for peanut allergy and one with unspecified methodology featuring multiple food allergies; contacting the authors confirmed there were no accompanying peer reviewed papers (36-38).

Risk assessment ability of CRD

Two studies assessing CRD for hen's egg allergy found that sIgE levels for all components tested were higher in patients with more severe allergies (24, 25). Another study found similar results for cow's milk allergic patients (23). For peanut allergy, one study found that patients with more severe food challenge reactions had higher sIgE levels to Ara h 1, 2, 3, and 6 than did patients with no reaction or mild symptoms (30). Furthermore, this study found that all severe allergic patients were sensitized to Ara h 2 or Ara h 6, and none of them were sensitized to Ara h 1, 3, or 9 without Ara h 2 (30). For hazelnut allergy, one study found that higher sIgE levels to Cor a 9 and Cor a 14 were associated with more severe reactions in food challenges, but found no correlation between sIgE levels to Cor a 1 or Cor a 8 and reaction severity (32).

Discussion

Summary of key findings

This systematic review included 11 studies that assessed the accuracy of CRD in diagnosing cow's milk, hen's egg, peanut, hazelnut, and shrimp allergies. Overall, the components tested by the studies included in this review were: Bos d 4, Bos d 5, Bos d 8, and the caseins for cow's milk allergy; Gal d 1, Gal d 2, Gal d 3, and Gal d 4 for hen's egg allergy; Ara h 1, Ara h 2, Ara 3, Ara h 6, Ara h 8, and Ara h 9 for peanut allergy; Cor a 1, Cor a 8, Cor a 9, Cor a 14, Bet v 1, and Bet v 2 for hazelnut allergy; and Pen a 1, Lit v 1, and Lit v 4 for shrimp allergy. No studies meeting our inclusion criteria investigated CRD for diagnosing wheat, soy, and fish allergies. The components with the highest diagnostic accuracy reported, along with their sensitivity-specificity pairs, were: Bos d 4 for cow's milk allergy (62.0% and 87.5%), (23) Gal d 1 for hen's egg allergy (84.2% and 89.8% for heated egg, and 60.6% and 97.1% for raw egg) (24), Ara h 2 for peanut allergy (80.3% and 95.1%) (30), Cor a 14 for hazelnut allergy (100% and 93.8%) (27), and Lit v 1 for shrimp (82.8% and 56.3%) (34). Additionally, two studies found that individual epitopes in shrimp's Lit v 1 and Lit v 2 could potentially have high diagnostic accuracy measures (26, 34).

Of the included studies, one study had a high ROB score in two of the four QUADAS-2 domains (34), and four studies had one such score (26, 27, 32, 33). The remaining six studies were scored low or unclear ROB in all four domains (23-25, 28, 30, 31).

Strengths and limitations

To the best of our knowledge, this is the first systematic review analyzing the evidence on the diagnostic accuracy of CRD for a range of food allergies. The strengths of this study include the use of a highly sensitive search strategy with no language restrictions, which allowed a comprehensive literature search, conducted across several databases and clinical trial registries. The inclusion criteria for this review were carefully selected to provide clinically relevant information (16). Similar work in the field, including a RAND report and a systematic review by the European Academy of Allergy and Clinical Immunology (EAACI) Food Allergy and Anaphylaxis Guidelines Group (12, 39), were used as bases for the inclusion criteria of this review. Furthermore, the internal validity of the studies included in this review was strong, since they all used DBPCFC as the reference standard in at least 50% of participants. A limitation of this review is that due to the large degree of heterogeneity between studies that met the inclusion criteria (in terms of the components tested, the particular CRD assay employed, and the cut-off values used), a quantitative synthesis of data could not be undertaken. Additionally, we acknowledge that food allergies other than the ones we focused on are becoming increasingly important. Since our search strategy was broadly formulated, we were, in response to expert peer-review feedback, able to check for any other potentially relevant studies for other foods. We were however unable to find any such studies. In the future, in the context of planned updates to this review, we plan to formally include terms for other foods (e.g. apple, cherry and peach) as this may impact on the sensitivity of our searches.

Comparison of findings with the wider literature

The results of this review corroborate the findings of a previous DTA systematic review on CRD for peanut allergy (14), suggesting that sIgE levels to the component Ara h 2 can provide diagnostic measures with very high accuracy. Our review, however, used more rigorous inclusion criteria than the previous one, strengthening the DTA evidence in relation to this component. The studies included in our review that analyzed peanut components also found that sIgE levels to the components Ara h 1, 3, 8, and 9 showed varying results, most with underperforming diagnostic values (14). Nevertheless, this is the first review to present evidence on the diagnostic value of Ara h 6 as a CRD component, which was found to have higher sensitivity and specificity values than Ara h 2.

When comparing the diagnostic accuracy of CRD found in our review with that of first-line diagnostic tests for food allergy (atopy patch tests (APT), SPT, and sIgE) reported in a previous systematic review (12), results vary by allergy. For cow's milk allergy, Bos d 4 and Bos d 8 have similar DTA results to APT and sIgE (these tests showed sensitivity-specificity pairs of 52.8% and 88.1%, and 87.3% and 47.7%, respectively); additionally, these components displayed lower sensitivity and higher specificity than SPT (which showed a sensitivity-specificity pair of 87.9% and 67.5%) (12). For hen's egg allergy, Gal d 1 had lower sensitivity and higher specificity than SPT and sIgE (these tests showed sensitivity-specificity pairs of 92.4% and 58.1%, and 93.4% and 49.2%, respectively) for both raw and heated egg (12). For peanut allergy, Ara h 6 showed higher DTA measures than SPT and sIgE (these tests showed sensitivity-specificity pairs of 94.7% and 61.0%, and 96.3% and 59.3%, respectively) (12). Although the previous systematic review did not carry out meta-analysis for hazelnut and shrimp allergies, results from individual studies suggest that Cor a 9 and Cor a 14 may have higher DTA measures than SPT and sIgE for hazelnut allergy, and that Lit v 1 shows marginally lower sensitivity and higher sensitivity than SPT and sIgE (12).

Implications for research

We have identified important research gaps in this field. First, there is a limited body of methodologically robust evidence to assess the accuracy of CRD in diagnosing food allergies. From potentially relevant studies identified through database searches, 21 were excluded because they did not carry out DBPCFC in at least 50% of participants, and 27 were excluded because their food challenges were not double-blind placebo-controlled. Furthermore, we did not identify any methodologically strong studies assessing the diagnostic accuracy of CRD for wheat, soy, or fish allergies. Therefore, there is a need for more DTA studies using DBPCFC as the reference standard in >50% of participants in order to better assess the diagnostic accuracy of CRD for food allergies. Alternatively, because of the challenges that DBPCFC pose for researchers and patients, there is a need to systematically assess whether other types of diagnoses, such as a combination of open food challenges and other markers of sensitization as recently proposed (40), could be used as a reference standard for DTA studies.

Second, this review found that at present, a quantitative synthesis of CRD diagnostic accuracy data is not possible because of the paucity of studies for each of the components that have been studied. Given that CRD is still in development, there is a need for more studies and a consensus reached on

the optimal cut-off values that will facilitate quantitative evidence synthesis and data pooling across studies. Alternatively, the heterogeneity of cut-off values could be alleviated if all DTA studies reported appendices with sIgE concentration values for each participant. This would allow reviewers to obtain DTA summary measures from all studies in a transparent and homogenous manner.

Third, there is a need to standardize all CRD assays to ensure that results are comparable between different tests. This includes the individual allergen components used, the results obtained from assays from different manufacturers, and the results obtained from microarray and single-component tests.

Finally, there is a dearth of evidence on the cost-effectiveness and the risk assessment ability of CRD relative to current care models. In principle, it may be possible to utilize some of the data obtained here for use in economic modeling to facilitate comparisons of the relative value of any trade-off between sensitivity and specificity in a more formal manner. Such analyses were beyond the scope of this review.

Implications for patient care

Selected CRD components have the potential to diagnose food allergies with a higher specificity, but lower sensitivity than current first-line tests. Furthermore, risk assessments carried out by five of the included studies suggest that quantitative measurements of sIgE levels to key components have the potential to identify patients with more severe allergic phenotypes. Such is the case for Ara h 2 and Ara h 6 for peanut allergy, and Cor a 9 and Cor a 14 for hazelnut allergy. Nevertheless, further research is necessary to draw stronger conclusions and to determine the components with the highest diagnostic value, as well as the clinically appropriate cut-off values.

Importantly, all studies included in this review recruited patients with suspected allergies. Therefore, it is likely that the prevalence of allergies in the study populations is considerably higher than in more population-based settings, rendering the tests' PPVs higher and NPVs lower than they would be in populations with lower allergy prevalence.

Conclusions

The findings of this review suggest that some CRD components have the potential to diagnose cow's milk, hen's egg, peanut, hazelnut, and shrimp allergies with high specificity, but low sensitivity. Nevertheless, at present there is not enough methodologically robust evidence to draw definite conclusions. Further studies employing DBPCFC as the reference standard are urgently needed to effectively evaluate the DTA and cost-effectiveness of CRD, as well as standardization of the components assessed and CRD assays used, and consensus on study reporting.

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Conflict of interest

The authors declare no conflict of interest.

Authors' contributions

AS conceived this study and together with BN, NM, and ASt secured the funding for the work. The systematic review was led by JFK and NM with ASt advising on the health economics. JFK drafted the manuscript, which was commented on by other authors. All authors approved the final version of the manuscript.

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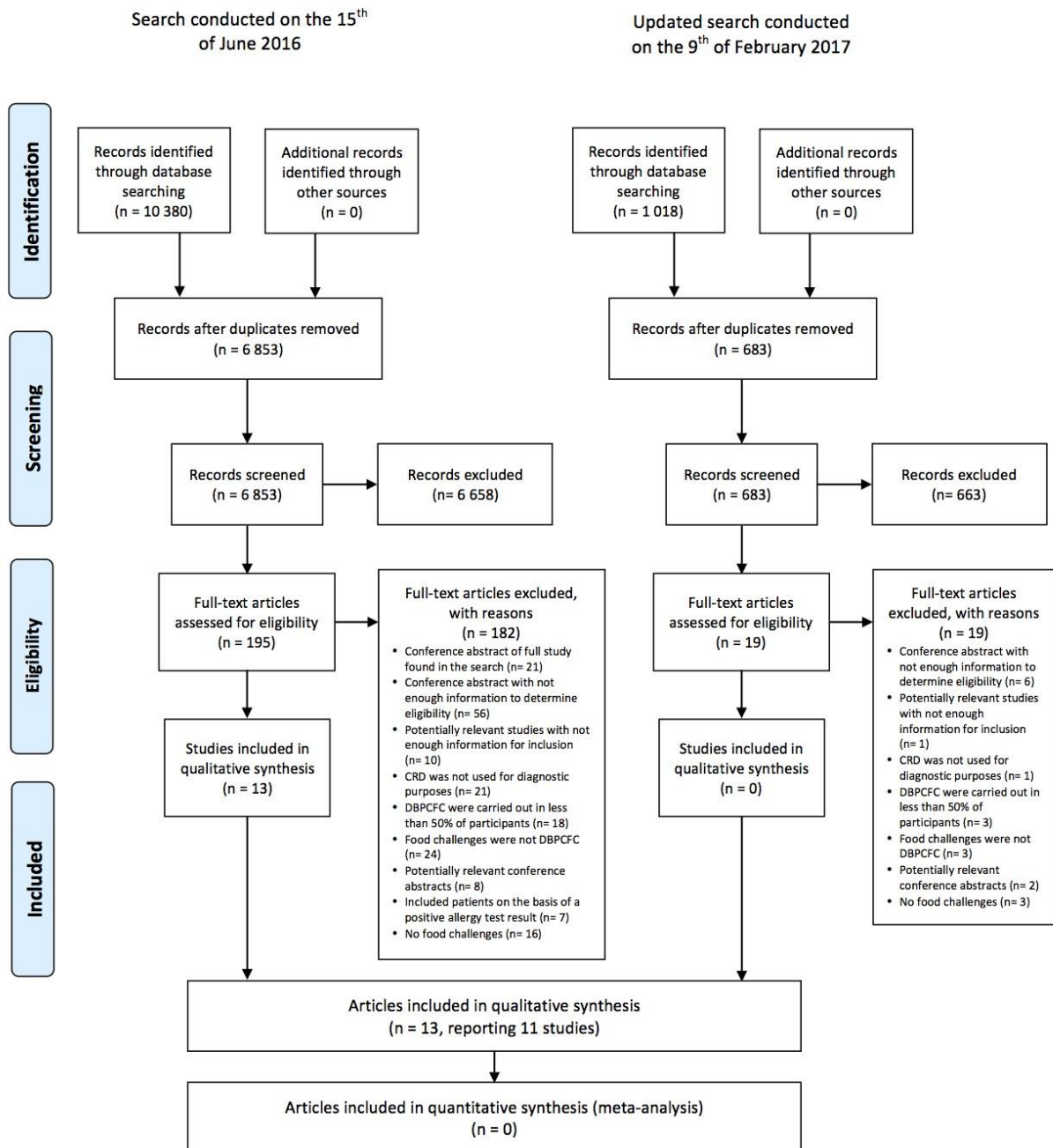
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Figure 1: PRISMA Diagram for the literature search



Adapted from: Moher D, Liberati A, Tetzlaff J, Altman DG, The PRISMA Group (2009). Preferred Reporting Items for Systematic Reviews and Meta-Analyses: The PRISMA Statement. PLoS Med 6(7): e1000097. doi:10.1371/journal.pmed1000097

Table 1: Main characteristics of included studies

First Author, Year	Country of Origin	Reference No.	Study Design	Food Allergy Studied	Population	Sample Size	Components Tested	Index Test	Number of Patients That Underwent DBPCFC	QUADAS-2 Risk of Bias Domains 1, 2, 3, 4 Applicability domains 1, 2, 3
Alessandri 2012a	Italy	23	Cross-sectional	Cow Milk	Children with suspected cow's milk allergy	66	Bos d 4, 5, and 8	ImmunoCAP	66 (100%)	
Alessandri 2012b	Italy	24	Cross-sectional	Hen's Egg	Children with suspected hen's egg allergy	68	Gal d 1, 2, 3, and 5	ISAC 103	68 (100%)	
Ando 2008	Japan	25	Cross-sectional	Hen's Egg	Children with suspected hen's egg allergy	108	Gal d 1, and 2	ImmunoCAP	108 (100%)	
Ayuso 2012	Spain	26	Cross-sectional	Shrimp	Patients with reported immediate shrimp allergic reactions	37	Pen a 1; and Lit v 1, 2, 3, and 4	ImmunoCAP and Immunoblot	31 (83.8%)	
Brandstrom 2015	Sweden	27	Cross-sectional	Hazelnut	Children referred for oral challenge for hazelnut allergy suspicion	40	Cor a 1, 8, 9, and 14; and Bet v 1	ImmunoCAP	40 (100%)	
Klemans 2014 (& Klemans 2013)	The Netherlands	28 (29)	Cross-sectional	Peanut	Adults who underwent a DBPCFC for peanut allergy suspicion	107	Ara h 2 and 6	ISAC 112	107 (100%)	

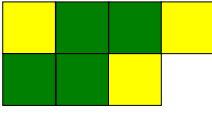
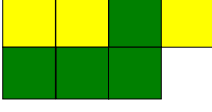
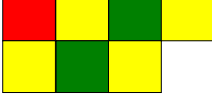
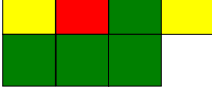
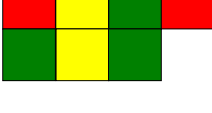
Kukkonen 2015	Finland	30	Cross-sectional	Peanut	Children with suspected peanut allergy	102	Ara h 1, 2, 3, 6, 8, and 9	ImmunoCap and ISAC112	102 (100%)	
Lieberman 2013	USA and Sweden	31	Cross-sectional	Peanut	Children referred to an allergy centre for evaluation of peanut allergy	167	Ara h 1, 2, 3, and 8	ImmunoCAP	119 (71.3%)	
Masthoff 2013 (& Masthoff 2015)	The Netherlands	32 (35)	Case-Control	Hazelnut	Patients sensitized to hazelnut extract	161	Cor a 1, 8, 9, and 14; and Bet v 1	ImmunoCAP	137 (85.1%)	
Ott 2008	Germany	33	Cross-sectional	Hen's Egg and Cow's Milk	Children with suspected hen's egg or cow's milk allergy	130	Gal d 1, 2 and 4; Bos d 4, and 5, and Caseins	ISAC version CRD 51	103 (71%)	
Pascal 2015	Brazil, Spain, and USA	34	Case-control	Shrimp	Patients sensitized to shrimp	103	Lit v 1, Lit v 2, Lit v 3, and Lit v 4	Immunoblot	78 (76%)	

Table 2: Summary DTA measures of CRD components tested by allergy type

Food-allergy	Component	Study	Year	Reference No.	Country	Index-test	Cut-off value	Sensitivity (95%CI)	Specificity (95%CI)	PPV (95%CI)	NPV (95%CI)
Cow's mik	Bos d 4	Alessandri	2012a	23	Italy	ISAC 103	>0 (kUa/L)	56.0 (41.3-70.0)	87.5 (61.7-98.5)	93.3 (78.9-98.1)	38.9 (38.9-47.8)
Cow's mik	Bos d 4	Alessandri	2012a	23	Italy	ISAC 103	>0.01 (kUa/L)	62.0 (47.2-75.4)	87.5 (61.7-98.5)	93.9 (80.6-98.3)	42.4 (33.1-52.4)
Cow's mik	Bos d 4	Ott	2008	33	Germany	ISAC CRD51	>0.1 (FI)	50.0 (34.2-65.8)	93.0 (80.9-98.5)	87.5 (69.3-95.6)	65.6 (58.2-72.3)
Cow's mik	Bos d 4	Alessandri	2012a	23	Italy	ImmunoCAP	>0.35 (kUa/L)	80.0 (66.3-90.0)	50.0 (24.7-75.4)	83.3 (75.0-89.3)	44.4 (27.6-62.6)
Cow's mik	Bos d 4	Alessandri	2012a	23	Italy	ImmunoCAP	>1.02 (kUa/L)	58.0 (43.2-71.8)	81.3 (54.4-96.0)	90.6 (77.2-96.5)	38.2 (29.3-48.1)
Cow's mik	Bos d 5	Alessandri	2012a	23	Italy	ImmunoCAP	>0 (kUa/L)	90.0 (78.2-96.7)	50.0 (24.7-75.4)	84.9 (77.4-90.3)	61.5 (37.9-80.8)
Cow's mik	Bos d 5	Alessandri	2012a	23	Italy	ISAC 103	>0 (kUa/L)	40.0 (26.4-54.8)	93.8 (69.8-99.8)	95.2 (74.4-99.3)	33.3 (27.8-39.3)
Cow's mik	Bos d 5	Alessandri	2012a	23	Italy	ISAC 103	>0.01 (kUa/L)	44.0 (30.0-58.8)	93.8 (69.8-99.8)	95.7 (76.3-99.3)	34.9 (28.9-41.4)
Cow's mik	Bos d 5	Ott	2008	33	Germany	ISAC CRD 51	>0.1 (FI)	23.8 (12.1-39.5)	95.3 (84.2-99.4)	83.3 (53.8-95.6)	56.2 (51.7-60.6)

Cow's mik	Bos d 5	Alessandri	2012a	23	Italy	ImmunoCAP	>0.35 (kUa/L)	82.0 (68.6-91.4)	62.5 (35.4-84.8)	87.2 (78.2-92.9)	52.6 (35.5-69.2)
Cow's mik	Bos d 8	Alessandri	2012a	23	Italy	ISAC 103	>0 (kUa/L)	54.0 (39.3-68.2)	81.3 (54.4-96.0)	90.0 (75.9-96.3)	36.1 (27.9-45.3)
Cow's mik	Bos d 8	Alessandri	2012a	23	Italy	ISAC 103	>0.01 (kUa/L)	56.0 (41.3-70.0)	81.3 (54.4-96.0)	90.3 (76.6-96.4)	37.1 (28.6-46.6)
Cow's mik	Bos d 8	Alessandri	2012a	23	Italy	ImmunoCAP	>0.35 (kUa/L)	88.0 (75.7-95.5)	56.3 (29.9-80.3)	86.3 (78.1-91.7)	60.0 (38.7-78.1)
Cow's mik	Bos d 8	Alessandri	2012a	23	Italy	ImmunoCAP	>0.44 (kUa/L)	82.0 (68.6-91.4)	62.5 (35.4-84.8)	87.2 (78.2-92.9)	52.6 (35.5-69.2)
Cow's mik	α -casein	Ott	2008	33	Germany	ISAC CRD 51	>0.1 (FI)	26.2 (13.9-42.0)	97.7 (87.7-99.9)	91.7 (59.8-98.8)	57.5 (52.9-62.0)
Cow's mik	β -casein	Ott	2008	33	Germany	ISAC CRD 51	>0.1 (FI)	26.2 (13.9-42.0)	93.0 (80.9-98.5)	78.6 (52.4-92.4)	56.3 (51.4-61.1)
Cow's mik	κ -casein	Ott	2008	33	Germany	ISAC CRD 51	>0.1 (FI)	38.1 (23.6-54.4)	88.4 (74.9-96.1)	76.2 (56.3-88.8)	59.4 (53.0-65.5)
Heated Hen's Egg	Gal d 1	Alessandri	2012b	24	Italy	ISAC 103	>0.01 (kUa/L)	84.2 (60.4-96.6)	89.8 (77.8-96.6)	76.2 (57.7-88.3)	93.6 (83.8-97.7)
Heated Hen's Egg	Gal d 1	Ando	2008	25	Japan	ImmunoCAP	>0.37 (kUa/L)	97.4 (86.2-99.9)	35.7 (24.6-48.1)	45.1 (40.7-49.7)	96.2 (77.9-99.4)
Heated Hen's Egg	Gal d 1	Ando	2008	25	Japan	ImmunoCAP	>4.4 (kUa/L)	76.3	81.4	69.0	86.4

								(59.8-88.6)	(70.3-89.8)	(57.0-79.0)	(78.0-91.9)
Heated Hen's Egg	Gal d 2	Alessandri	2012b	24	Italy	ISAC 103	>0.01 (kUa/L)	52.6 (28.9-75.6)	83.7 (70.3-92.7)	55.6 (36.8-72.9)	82.0 (73.6-88.1)
Heated Hen's Egg	Gal d 2	Ando	2008	25	Japan	ImmunoCAP	>0.37 (kUa/L)	100.0 (90.8-100)	21.4 (12.5-32.9)	40.9 (37.9-43.9)	100.0 (NA)
Heated Hen's Egg	Gal d 2	Ando	2008	25	Japan	ImmunoCAP	>6.33 (kUa/L)	73.7 (56.9-86.6)	72.9 (60.9-82.8)	59.6 (49.0-69.3)	83.6 (76.6-89.9)
Heated Hen's Egg	Gal d 3	Alessandri	2012b	24	Italy	ISAC 103	>0.01 (kUa/L)	21.1 (6.1-45.6)	93.9 (83.1-98.7)	57.1 (24.7-84.4)	75.4 (70.6-79.6)
Raw Hen's Egg	Gal d 1	Ott	2008	33	Germany	ISAC CRD 51	>0 (FI)	57.8 (42.2-72.3)	86.7 (59.5-98.3)	92.9 (77.7-98.0)	40.6 (31.6-50.4)
Raw Hen's Egg	Gal d 1	Alessandri	2012b	24	Italy	ISAC 103	>0.01 (kUa/L)	60.6 (42.1-77.1)	97.1 (85.1-99.9)	95.2 (74.0-99.3)	72.3 (63.1-80.0)
Raw Hen's Egg	Gal d 1	Ando	2008	25	Japan	ImmunoCAP	>0.37 (kUa/L)	86.6 (76.0-93.7)	41.5 (26.3-57.9)	70.7 (64.8-76.1)	65.4 (48.2-79.3)
Raw Hen's Egg	Gal d 1	Ando	2008	25	Japan	ImmunoCAP	>2.26 (kUa/L)	73.1 (60.9-83.2)	82.9 (67.9-92.9)	87.5 (77.8-93.3)	65.4 (55.4-74.2)
Raw Hen's Egg	Gal d 2	Ott	2008	33	Germany	ISAC CRD 51	>0 (FI)	57.8 (42.2-72.3)	80.0 (51.9-95.7)	89.7 (75.3-96.1)	38.7 (29.2-49.1)
Raw Hen's Egg	Gal d 2	Alessandri	2012b	24	Italy	ISAC 103	>0.01 (kUa/L)	42.4 (25.5-60.8)	88.6 (73.3-96.8)	77.8 (56.2-90.5)	62.0 (54.3-69.1)

Raw Hen's Egg	Gal d 2	Ando	2008	25	Japan	ImmunoCAP	>0.37 (kUa/L)	97.0 (89.6-99.6)	31.7 (18.1-48.1)	69.9 (65.2-74.2)	86.7 (60.7-96.5)
Raw Hen's Egg	Gal d 2	Ando	2008	25	Japan	ImmunoCAP	>3.88 (kUa/L)	76.1 (64.1-85.7)	82.9 (67.9-92.9)	87.9 (78.6-93.6)	68.0 (57.6-76.9)
Raw Hen's Egg	Gal d 3	Alessandri	2012b	24	Italy	ISAC 103	>0.01 (kUa/L)	18.2 (7.0-35.5)	97.1 (85.1-99.9)	85.7 (43.3-97.9)	55.7 (51.5-59.9)
Raw Hen's Egg	Gal d 4	Ott	2008	33	Germany	ISAC CRD 51	>0 (FI)	17.8 (8.0-32.1)	100.0 (78.2-100)	100.0 (NA)	28.8 (26.1-31.7)
Peanut	Ara h 1	Kukkonen	2015	30	Finland	ImmunoCAP	>0.35 kUa/l	63.9 (50.6-75.8)	87.8 (73.8-95.9)	88.6 (77.1-94.8)	62.1 (53.5-70.0)
Peanut	Ara h 1	Lieberman	2013	31	USA and Sweden	ImmunoCAP	>0.35 kUa/L	56.6 (46.6-66.2)	86.9 (75.8-94.2)	88.2 (79.4-93.6)	53.5 (47.6-59.4)
Peanut	Ara h 1	Kukkonen	2015	30	Finland	ImmunoCAP	>0.8 kUa/L	60.7 (47.3-72.9)	95.1 (83.5-99.4)	94.9 (82.5-98.6)	61.9 (54.2-69.1)
Peanut	Ara h 2	Klemans	2014	28	The Netherlands	ISAC 112	>0.3 ISU/L	69.2 (56.6-80.1)	90.5 (77.4-97.3)	91.8 (81.4-96.7)	65.5 (56.6-73.5)
Peanut	Ara h 2	Kukkonen	2015	30	Finland	ImmunoCAP	>0.35 kUa/L	95.1 (86.3-99.0)	73.2 (57.1-85.8)	84.1 (76.0-89.8)	90.9 (76.6-96.8)
Peanut	Ara h 2	Lieberman	2013	31	USA and Sweden	ImmunoCAP	>0.35 kUa/L	80.2 (71.3-87.3)	91.8 (81.9-97.3)	94.4 (88.0-97.5)	72.7 (64.4-79.8)
Peanut	Ara h 2	Klemans	2014	28	The Netherlands	ISAC 112	>1.0 ISU/L	58.5	95.2 (83.8-99.4)	95.0	59.7

								(45.6-70.6)		(82.9-98.7)	(52.4-66.6)
Peanut	Ara h 2	Kukkonen	2015	30	Finland	ImmunoCAP	>1.8 kUa/L	80.3 (68.2-89.4)	95.1 (83.5-99.4)	96.1 (86.3-99.0)	76.5 (66.1-84.4)
Peanut	Ara h 3	Kukkonen	2015	30	Finland	ImmunoCAP	>0.35 kU/L	57.4 (46.1-70.0)	90.2 (76.9-97.3)	89.7 (77.1-95.8)	58.7 (51.1-66.0)
Peanut	Ara h 3	Lieberman	2013	31	USA and Sweden	ImmunoCAP	>0.35 kUa/L	48.1 (38.3-58.1)	90.2 (79.8-96.3)	89.5 (79.5-94.9)	50.0 (45.0-55.0)
Peanut	Ara h 3	Kukkonen	2015	30	Finland	ImmunoCAP	>0.8 kUa/L	55.7 (42.5-68.5)	95.1 (83.5-99.4)	94.4 (81.2-98.5)	59.1 (51.9-65.9)
Peanut	Ara h 6	Kukkonen	2015	30	Finland	ISAC 112	>0.3 ISU	98.3 (90.9-100)	90.2 (76.9-97.3)	93.5 (85.1-97.4)	97.4 (84.1-99.6)
Peanut	Ara h 6	Klemans	2014	28	The Netherlands	ISAC 112	>0.3 ISU/L	70.8 (58.2-81.4)	85.7 (71.5-94.6)	88.5 (78.2-94.2)	65.5 (56.0-73.8)
Peanut	Ara h 6	Kukkonen	2015	30	Finland	ISAC 112	>0.8 ISU	94.9 (85.9-98.9)	95.1 (83.5-99.4)	96.6 (87.9-99.1)	92.9 (81.2-97.5)
Peanut	Ara h 6	Klemans	2014	28	The Netherlands	ISAC 112	>1.0 ISU/L	61.5 (48.6-73.4)	95.2 (83.8-99.4)	95.2 (83.6-98.7)	61.5 (53.9-68.7)
Peanut	Ara h 8	Kukkonen	2015	30	Finland	ImmunoCAP	>0.35 kU/l	78.7 (66.3-88.1)	14.6 (5.6-29.2)	57.8 (53.3-62.2)	31.6 (16.0-52.7)
Peanut	Ara h 8	Lieberman	2013	31	USA and Sweden	ImmunoCAP	>0.35 kUa/L	34.9 (25.9-44.8)	42.6 (30.0-55.9)	51.4 (43.0-59.7)	27.4 (21.4-34.2)

Peanut	Ara h 9	Kukkonen	2015	30	Finland	ImmunoCAP	>0.35 kUa/L	14.8 (7.0-26.2)	85.4 (70.8-94.4)	60.0 (36.6-79.6)	40.2 (36.4-44.2)
Hazelnut	Cor a 1	Masthoff	2012	32	The Netherlands	ImmunoCAP	>0.35 kUa/L	79.7 (69.2-88.0)	7.3 (2.7-15.3)	45.3 (42.2-48.5)	27.3 (13.4-47.6)
Hazelnut	Cor a 8	Masthoff	2013	32	The Netherlands	ImmunoCAP	>0.35 kUa/L	6.3 (2.1-14.2)	96.3 (89.7-99.2)	62.5 (29.2-87.1)	51.6 (49.9-53.4)
Hazelnut	Cor a 9	Masthoff	2013	32	The Netherlands	ImmunoCAP	>0.35 kUa/L	59.5 (47.9-70.4)	87.8 (78.7-94.0)	82.5 (71.9-89.6)	69.2 (63.0-74.8)
Hazelnut	Cor a 9	Brandsstrom	2015	27	Sweden	ImmunoCAP	>0.65 kUa/L	100.0 (63.1-100)	71.9 (53.3-86.3)	47.1 (33.8-60.7)	100.0 (NA)
Hazelnut	Cor a 9	Masthoff	2013	32	The Netherlands	ImmunoCAP	>1 kUa/L	54.4 (42.8-65.7)	97.6 (91.7-99.7)	95.6 (84.4-98.9)	69.0 (63.5-73.9)
Hazelnut	Cor a 9	Masthoff	2013	32	The Netherlands	ImmunoCAP	>5 kUa/L	30.4 (20.5-41.8)	98.8 (93.4-100)	96.0 (76.9-99.4)	59.6 (56.0-63.1)
Hazelnut	Cor a 14	Masthoff	2013	32	The Netherlands	ImmunoCAP	>0.35 kUa/L	54.4 (42.8-65.7)	85.4 (75.8-92.2)	78.2 (67.2-86.3)	66.0 (60.1-71.6)
Hazelnut	Cor a 14	Brandsstrom	2015	27	Sweden	ImmunoCAP	>0.64 kUa/L	100.0 (63.1-100)	93.8 (79.2-99.2)	80.0 (51.1-93.9)	100.0 (NA)
Hazelnut	Cor a 14	Masthoff	2013	32	The Netherlands	ImmunoCAP	>1 kUa/L	50.6 (39.1-62.1)	86.6 (77.3-93.1)	78.4 (66.8-86.8)	64.5 (58.9-69.8)
Hazelnut	Cor a 14	Masthoff	2013	32	The Netherlands	ImmunoCAP	>5 kUa/L	39.2	98.8	96.9	62.8

								(28.4-50.9)	(93.4-100)	(81.3-100)	(58.5-66.9)
Hazelnut	Bet v 1	Masthoff	2013	32	The Netherlands	ImmunoCAP	>0.35 kUa/L	81.0 (70.6-89.0)	7.3 (2.7-15.3)	45.7 (42.7-48.8)	28.6 (14.1-49.5)
Hazelnut	Bet v 2	Masthoff	2013	32	The Netherlands	ImmunoCAP	>0.35 kUa/L	10.1 (4.5-19.0)	81.7 (71.6-89.4)	34.8 (19.3-54.3)	48.6 (45.4-51.7)
Shrimp	Lit v 1	Pascal	2015	34	Brazil, Spain and USA	Immunoblot	NA	82.8 (70.6-91.4)	56.3 (29.9-80.3)	87.3 (79.5-92.4)	47.4 (30.7-64.7)
Shrimp	Lit v 4	Pascal	2015	34	Brazil, Spain and USA	Immunoblot	NA	34.5 (22.5-48.1)	93.8 (69.8-99.8)	95.2 (74.4-99.3)	28.3 (24.0-33.1)
Shrimp	rPen a 1	Ayuso	2012	26	Spain	ImmunoCAP	>0.35 kUa/L	88.2 (63.6-98.5)	23.8 (8.2-47.2)	48.4 (41.1-55.8)	71.4 (35.6-91.9)