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1 **Title:** Synaptic integrative mechanisms for spatial cognition

2

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4

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8

9

10 **Abstract**

11 Synaptic integrative mechanisms have profound impacts on electrical signaling in the brain that,
12 while largely hidden from recording methods that observe spiking activity of neurons, may be
13 critical for how information is encoded, stored and retrieved. Here, we review roles for synaptic
14 integrative mechanisms in selection, generation and plasticity of place and grid fields, and in
15 related temporal codes for representation of space. We outline outstanding questions and
16 challenges in testing hypothesized models for spatial computation and memory.

17

18

19 **Main text**

20 The spatial firing patterns of neurons in the hippocampal formation are central to neurobiological
21 theories of spatial cognition^{1,2}. How the spatial modulation of place, grid, head-direction, border
22 and other spatial cell types emerges from their synaptic input, while largely hidden from view
23 when observing spike firing, is likely to be critical for spatial computation. In simple models the
24 details of this process of synaptic integration are of limited importance - spike output is assumed
25 to be a stable, linear function of synaptic input. In contrast, considerable experimental evidence
26 demonstrates that synaptic integration is often non-linear, can be spatially compartmentalised
27 within a cell and is controlled by diverse mechanisms, suggesting it has key computational
28 roles³⁻⁵. Here, we consider evidence that specific mechanisms for integration of synaptic input
29 are critical for spatial cognition. We will focus on aspects of hippocampal spatial firing fields and
30 temporal codes for which recent experiments give insights into roles of these integrative
31 mechanisms.

32

33 What cellular mechanisms does a neuron have available to determine integration of its synaptic
34 input? To influence spatial firing a synaptic event must influence action potential initiation.

35 Several cellular properties determine the impact of a synaptic event³⁻⁵. First, neuronal
36 excitability is established by ion channels that set a neuron's resting membrane potential,
37 voltage threshold for triggering an action potential and membrane conductance. The difference
38 between the resting potential and threshold potential gives the voltage change that must be
39 achieved to trigger a spike. The membrane conductance, in conjunction with the capacitance
40 established by the lipid bilayer of the cell membrane, determines how easily and rapidly a
41 synaptic input can change the membrane potential. Second, various voltage-dependent ionic
42 currents, including those mediated by Na⁺, Ca²⁺, and NMDA receptor (NMDAR) channels, can
43 amplify synaptic responses, while other ion channels and inhibitory synaptic receptors suppress
44 synaptic responses. Third, most synaptic inputs are made onto dendrites, which can extend

45 hundreds of microns from a neuron's soma. All other things being equal, more distant synapses
46 are less effective because attenuation of synaptic responses increases as they propagate
47 further. Finally, spatially extended dendrites enable compartmentalisation. For example,
48 different dendritic domains may be endowed with distinct combinations of voltage-gated ion
49 channels and the particular signalling mechanisms that modify synaptic responses may be
50 directed to specific locations.

51
52 These synaptic integrative mechanisms suggest considerable cellular complexity, but why
53 should we consider them when trying to understand spatial computation? Powerful artificial
54 neural networks can be assembled from simple neurons that linearly sum their synaptic inputs⁶.
55 Why then do networks for spatial cognition employ neurons with diverse and complex
56 integrative properties? One possibility is that specialisation enables neurons to adapt to
57 fundamental limits imposed by their cellular hardware⁷. For example, to integrate many synaptic
58 inputs a neuron requires an extensive dendritic tree, but this comes at a cost in that distal inputs
59 will evoke smaller and less temporally precise somatic responses. This cost can be
60 compensated by integrative mechanisms that boost the strength of distal synapses and that
61 normalise their time course at the soma. A second possibility is that diversity in subthreshold
62 properties reflects the selection of distinct building blocks for specialised computations^{5,7}.
63 According to this view, specific mechanisms for synaptic integration may be necessary to the
64 cognitive function implemented in a circuit. While these viewpoints are not mutually exclusive,
65 we will focus here primarily on computational roles for synaptic integrative mechanisms.

66
67 For which aspects of spatial cognition might synaptic integrative mechanisms be important? We
68 will address roles in key elements of spatial computation in the hippocampus and medial
69 entorhinal cortex (MEC). We focus on hippocampal place and entorhinal grid cells, which use
70 their spike firing rate to encode locations with a high signal to noise ratio; firing within fields
71 usually peaks at frequencies > 10 Hz, whereas firing rates outside of fields are less than 1 Hz^{8,9}.
72 We will first consider recent evidence that integrative mechanisms determine selection of active
73 place cells and how this might form a basis for allocation of memory engrams.

74
75 In both place and grid cells the approximate Gaussian firing rate distribution of a single firing
76 field is driven by a ramp-like membrane potential depolarization (Fig. 1a-b)¹⁰⁻¹². This is at first
77 glance consistent with models in which straightforward linear integration of excitatory drive is
78 sufficient to explain place firing¹³⁻¹⁵. However, more recent observations that we discuss below
79 argue that active integrative mechanisms are essential for the emergence of ramp-like
80 depolarizations driving place fields, and may influence the spacing and stability of grid fields.

81
82 Beyond moment to moment computation, synaptic integrative mechanisms may contribute to
83 spatial memory by influencing the induction of synaptic plasticity. Specifically, encoding and
84 recall of spatial memories are associated with plasticity in the spatial firing of hippocampal
85 neurons^{16,17}. A critical issue is how patterns of spatially modulated synaptic input couple
86 appropriately to plasticity mechanisms. We will consider evidence that synaptic integrative
87 mechanisms establish rules for plasticity of spatial firing, by both promoting and suppressing
88 synaptic plasticity.

89
90 Finally, the relative timing of action potentials fired by place and grid cells may be of particular
91 importance for spatial memories. In particular, phase precession of action potentials relative to
92 the network theta rhythm (Fig. 1c) leads to the emergence of population level spike sequences
93 that may be structured to support associative memory storage^{18–21}. We will consider how
94 synaptic integrative mechanisms may contribute to temporal codes that are linked to theta
95 activity and that may be important for episodic memory.

96
97

98 **Excitability, place cell selection and memory allocation**

99 In a given environment, only subsets of CA1 pyramidal cells have place fields. Estimates range
100 from as low as 20% rising to 65% for larger environments^{22–24}. While additional cells are
101 recruited to encode larger environments, the number of silent cells and the number of cells with
102 multiple firing fields is greater than expected if all cells have a similar probability of generating a
103 place field²³. Rather, the probability that cells will have place fields is described by a gamma
104 distribution, suggesting a population-level code for environmental context²³. Such a code
105 requires mechanisms to determine which cells within the population become active. Recent
106 studies point to pyramidal cell excitability as critical for selection of active cells and suggest how
107 this population-level code may contribute to temporal components of episodic memories.

108
109 How are active place cells selected? Differences in excitable properties are an attractive
110 candidate mechanism, but it has been difficult to directly relate excitable integrative properties of
111 neurons to their firing fields during a behaviour. Technically demanding experiments involving
112 patch-clamp recordings from behaving rodents of the membrane potential of CA1 pyramidal
113 cells, and simultaneous measurement and manipulation of their electrical integrative properties,
114 have met this challenge. These studies reveal two differences in excitability between silent cells
115 and CA1 pyramidal cells that go on to have place fields in a novel environment; future place
116 cells have a lower threshold for action potential firing and a greater likelihood of firing bursts of
117 action potentials^{25,26} (Fig. 2a). By making action potential firing in response to synaptic input
118 more likely, both differences should promote the emergence of firing fields. Therefore, whether
119 a pyramidal cell becomes a place cell may in part be determined a priori by its intrinsic electrical
120 properties.

121
122 What consequences might a priori selection of place cells have for memory functions of the
123 hippocampus? Selection of active cells through differences in excitability has been suggested to
124 underpin temporal features of episodic memories²⁷. Elegant investigations of memory allocation
125 in the amygdala provide support for this general idea^{28–30}, but whether hippocampal-dependent
126 memories employ similar mechanisms has only recently been explored. When the activity of
127 populations of CA1 pyramidal cells are imaged in the same environment over multiple days, the
128 ensemble representation slowly evolves; some cells leave the ensemble, whereas others
129 join^{31,32}. If this subset of place cells is predetermined by differences in their excitability, then
130 memories formed on the same day are likely to be allocated to overlapping groups of neurons,
131 whereas memories formed on different days should be allocated to different populations.
132 Consistent with this prediction, cell populations tagged with a genetically encoded activity-

133 dependent reporter in a first context overlapped with cells labelled by a second activity-
134 dependent marker following exposure to a second context several hours later, but not several
135 days later³². Moreover, memories formed by exposure to contexts several hours apart, but not
136 several days apart, interact with one another³². Thus, the temporal properties of active place cell
137 assemblies, and of contextual memory storage, are consistent with there being an active subset
138 of excitable CA1 pyramidal cells that changes over a time-scale of days.

139
140 Selection of active place cells provides a potentially powerful mechanism for encoding temporal
141 components of memory, but what drives the subset of active neurons to change over time? One
142 possibility is that during memory formation neuronal activity leads in itself to transient (hours
143 long) increases in excitability, providing a mechanism for association of a second memory
144 formed within a time window defined by increases in excitability^{32,33}(Fig. 2b). This general
145 scheme is supported by investigations in which virally mediated expression, in subsets of
146 amygdala neurons, of the transcription factor CREB increases their excitability causing them to
147 be selected to participate in the engram of fear memories^{28,29}. Evidence that in hippocampal
148 neurons synaptic activity or spike firing activate CREB^{34,35}, and that CREB activation increases
149 excitability³⁶, is consistent with this idea. In this scheme, later periods of lowered activity that
150 facilitate memory dissociation may be established by self-regulatory mechanisms that come into
151 play after initial activation of CREB^{27,30}. A complementary possibility is that the identity of active
152 subsets of neurons provides a code from which the timeline of events can be read out³¹ (Fig.
153 2c). This idea is supported by observations that ensemble place field maps of different
154 environments on the same day share representations, and that decoders trained on one
155 environment can infer the day on which ensemble patterns were recorded from a second
156 environment³¹. According to this view, the active subset of place cells could be established
157 independently from neural activity, either through network wide coordination of the excitable set
158 of CA1 pyramidal cells, or perhaps through stochastic switching of CA1 pyramidal cells between
159 more and less excitable states.

160
161 Together these observations are consistent with excitability of CA1 pyramidal neurons selecting
162 place cell firing and memory allocation. Nevertheless, important questions remain to be
163 addressed. What is the ionic mechanism that controls which cells become excitable? The
164 difference in spike threshold between place cells and inactive CA1 pyramidal cells points to
165 voltage-gated ion channels that control action potential initiation^{25,26}. In contrast, activation of
166 CREB^{28,36}, and recent learning³⁷, both reduce afterhyperpolarization currents in CA1 neurons.
167 These differences may reflect multiple mechanisms acting across different timescales. Does
168 excitability predict ensemble membership over days? Correlating changes in a cell's excitability
169 with its firing fields will be critical here. Is the probability of a CA1 pyramidal cell forming a place
170 field a cell autonomous property, or does it depend on whether other cells are active? When
171 excitability of subsets of cells in the amygdala is increased, these cells predominate in engrams
172 that are formed, but the overall number of engram cells does not increase, implying that the
173 proportion of cells that form an engram is fixed by reciprocal inhibition³⁰. Similar mechanisms
174 may be present in hippocampal circuits (e.g. ^{38,39}). Does selection of place cells through
175 differences in excitability extend to other hippocampal areas? Unlike CA1, ensemble codes in
176 CA3 appear to be stable over days⁴⁰, whereas ensemble codes in CA2 evolve even more

177 rapidly than in CA1⁴¹. If stability of intrinsic excitability is used for place cell selection, then we
178 expect this to be reflected in differential control of excitability in each area.

179
180

181 **Membrane potential dynamics driving spatial firing**

182 How are synaptic inputs converted into action potential outputs that form a neuron's spatial firing
183 field? In vitro studies demonstrate that dendritic active conductances can either amplify or
184 suppress synaptic responses in hippocampal neurons (e.g. ⁴²⁻⁴⁵). Recent experiments probing
185 the membrane potential of spatial cells in awake animals, in the real world and using virtual
186 environments, show that ramp-like depolarizations drive spatial firing and have begun to reveal
187 roles for synaptic integrative mechanisms.

188

189 **Membrane potential ramps in CA1 pyramidal cells.** CA1 pyramidal cells provide a striking
190 example of how computation emerges through interaction between synaptic integrative
191 mechanisms, neuronal morphology and circuit connectivity. Excitatory inputs from layer 3 of
192 entorhinal cortex target distal dendrites of CA1 pyramidal cells, whereas local inputs from CA3
193 target their proximal dendrites⁴⁶ and diverse interneuron populations provide spatially restricted
194 inhibition⁴⁷. Either excitatory pathway appears to be sufficient to drive place firing^{48,49}, and active
195 integrative mechanisms may control responses to either or both pathways^{3,50-53}. How then is
196 spatial firing in place cells shaped by active synaptic integration?

197

198 Direct evidence that non-linear integrative mechanisms contribute to place firing comes from
199 experiments in which the membrane potential of silent CA1 pyramidal cells was continuously
200 depolarized while rats navigated an oval track⁵⁴. This manipulation caused place cells to
201 emerge. Importantly, the location of the induced field could not be predicted from the membrane
202 potential prior to injection of the depolarizing current. This finding argues against simple models
203 for the membrane potential ramp in which synaptic inputs within the firing field are stronger than
204 those outside the field, as these models predict that prior to continuous depolarization there
205 should be subthreshold ramps at the location of the firing field (Fig. 3a). Instead, in these
206 experimental conditions the emergence of place fields appears to be determined by a voltage-
207 dependent gating mechanism.

208

209 What might be the nature of this mechanism? One possibility is that excitatory synaptic inputs
210 within the field are indeed stronger than those outside, but that in silent cells the depolarization
211 they generate is insufficient to produce a measureable change in the somatic membrane
212 potential (Fig. 3b). This implies substantial attenuation of EPSPs as they propagate along
213 dendrites towards the soma, as has for example been reported for the basal dendrites of
214 neocortical pyramidal cells⁵⁵. In this scenario, continuous somatic depolarization may activate
215 voltage-dependent dendritic Na⁺, Ca²⁺, or NMDAR channels to amplify the local EPSPs, or
216 cause inactivation of K⁺ channels that would otherwise suppress EPSPs, either way enabling
217 the EPSPs to propagate to the soma. Another possibility is that synapses active within the firing
218 field have similar strength to those outside, but face a lower threshold to engage amplifying
219 dendritic conductances (Fig. 3c). This situation may be favored by clustering of synapses with
220 similar spatial preferences onto CA1 pyramidal cell dendrites⁵⁶. Although the signals encoded

221 by individual synapses on place cell dendrites are not yet clear, recent *in vivo* spine imaging
222 studies in visual cortex support the hypothesis that functionally similar inputs preferentially
223 target nearby locations on the dendritic tree of a neuron^{57,58}. Future experiments might address
224 this by imaging, during behavior, of synaptic terminals on identified place cells.

225

226 The voltage dependence of firing fields that emerges during prolonged membrane potential
227 depolarization provides strong evidence for functional engagement of integrative mechanisms
228 during place cell firing. However, whether active integrative mechanisms are also essential for
229 the generation of place fields under more physiological conditions is unknown. Recent
230 experiments in which the membrane potential of place cells was recorded in novel and familiar
231 environments suggest that new place fields emerge in the absence of a sustained
232 depolarization²⁶. Whether place fields in these conditions require voltage-dependent gating
233 mechanisms is not yet clear.

234

235 Regardless of the role of active integration, additional mechanisms are likely to shape the
236 membrane potential ramp driving place firing. For example, the membrane potential ramps
237 underlying receptive fields in other brain regions are substantially shaped by synaptic
238 inhibition⁵⁹. Specifically, orientation tuning curves in visual cortex are transformed linearly, with a
239 threshold, by inhibition from parvalbumin expressing interneurons^{60,61}. Input from local inhibitory
240 interneurons to CA1 place cells affects the shape of sub- and suprathreshold place fields in a
241 strikingly similar manner (Fig. 4), likely by suppressing firing and opposing active mechanisms
242 that amplify synaptic responses outside of the place field⁶². In visual cortex, different interneuron
243 subpopulations are thought to play specific roles in shaping orientation selectivity⁶³⁻⁶⁵. Similarly,
244 the effects of inhibition on the rising and falling parts of the place field ramp may be respectively
245 mediated by parvalbumin and somatostatin expressing interneurons⁶⁶.

246

247 **Membrane potential ramps in medial entorhinal cortex.** While grid firing fields of entorhinal
248 neurons are also driven by slow ramp-like depolarizations^{11,12}, the underlying integrative
249 mechanisms may be fundamentally different. For example, in contrast to hippocampal place
250 cells, the relative location of grid cell firing fields is stable across environmental manipulations,
251 suggesting circuit level interactions constrain grid cell firing fields^{67,68}. At the cellular level, the
252 dendritic morphology of hippocampal pyramidal cells differs substantially from stellate and
253 pyramidal cells in layer 2 of the MEC^{69,70}. Proposed mechanisms for generation of ramp
254 depolarizations also differ. Thus, the ramp depolarization recorded from grid cells is consistent
255 with predictions of continuous attractor network models^{11,12}. When these models are
256 implemented so that they reflect evidence that stellate cells interact primarily via local inhibitory
257 neurons^{71,72}, they predict that the depolarizing ramp results from disinhibition⁷².

258

259 Although network mechanisms are good candidates for generation of the membrane potential
260 ramp underlying grid firing, there is evidence that synaptic integrative mechanisms contribute to
261 the spacing and stability of grid fields. First, deletion of HCN1 channels, which mediate a major
262 component of the hyperpolarization-activated currents (I_h) in entorhinal stellate cells⁷³, increases
263 the width and spacing of grid cell firing fields⁷⁴. I_h is a mixed Na^+ and K^+ current that is unusual
264 in that it is activated by membrane hyperpolarization⁷⁵. Along with leak K^+ channels, I_h

265 generates a dorsoventral gradient in synaptic integration by stellate cells⁷⁶. At more dorsal
266 locations, where grid cells have closely spaced firing fields, a high density of both currents
267 reduces the width of synaptic potentials and opposes their temporal summation, whereas at
268 more ventral locations where grid cells typically have widely spaced firing fields, synaptic
269 potentials are broader and temporal summation is greater because the density of each current
270 is lower⁷⁶. Gradients in I_h are also associated with dorsoventral differences in intrinsic oscillatory
271 properties of stellate cells⁷⁷, which we discuss further below. Second, entorhinal stellate and
272 pyramidal cells are endowed with active conductances that produce a supralinear
273 transformation of synaptic inputs into action potential output⁷⁸. Simulations suggest that a slow,
274 NMDAR-mediated supralinear integration mechanism can promote the robustness of the grid
275 cell rate code. While direct recordings of NMDAR-mediated responses have not yet been
276 obtained from grid cells *in vivo*, NMDARs have been shown to be engaged during behaviour in
277 other brain regions⁵, where they contribute to receptive field tuning of somatosensory^{79,80} and
278 visual responses⁸¹.

279

280 What are the implications of these biophysical data for computations carried out by place and
281 grid cells? In place cells, non-linear synaptic integrative mechanisms may enable gating of place
282 firing⁵⁴, and maximize memory storage capacity⁸². For grid cells, differences in grid scale may
283 maximise the representational capacity of grid networks⁸³, but whether dorsoventral tuning of
284 synaptic integration plays a necessary or a modulatory role is unclear.

285

286

287 **Active synaptic integration and plasticity of spatial representations**

288 Successful learning requires plasticity of behaviorally relevant connections between neurons,
289 which in the case of spatial memory is thought to lead to stabilisation of place fields⁸³⁻⁸⁵.
290 Considerable evidence supports a necessary role for NMDAR-dependent synaptic plasticity in
291 this process⁸⁵. For example, pharmacological and genetic manipulations of NMDARs disrupt
292 long-term potentiation (LTP) of synaptic responses⁸⁵, spatial learning^{86,87}, the stability of place
293 cells⁸⁸, and spatial representation by place cells^{87,89}. Plasticity driven by activation of voltage-
294 gated Ca^{2+} channels may also play important roles (e.g. ⁹⁰). By determining the effects of
295 synaptic inputs on the membrane potential, active synaptic integration may interact with several
296 proposed mechanisms for recruitment of NMDARs and voltage-gated Ca^{2+} channels to either
297 facilitate or oppose induction of synaptic plasticity.

298

299 **Synaptic plasticity during place field formation and stabilization.** If NMDARs are indeed
300 instrumental for the stabilization of place cells, the Ca^{2+} influx that is associated with
301 postsynaptic depolarization and NMDAR channel opening should be detectable in the dendritic
302 tree during crossings of future or existing place fields. In support of this hypothesis, regenerative
303 Ca^{2+} events occur in basal dendritic branches during place field crossings and are associated
304 with the precision and stability of place fields⁹¹, suggesting that they represent postsynaptic
305 plasticity signals. A second type of regenerative calcium event is generated in the apical
306 dendrites of CA1 pyramidal cells. Precisely timed, coincident entorhinal cortex and CA3 inputs
307 evoke NMDAR-dependent dendritic plateau potentials *in vitro* and *in vivo*^{92,93} that can trigger
308 synaptic plasticity at least *in vitro*⁹². These complex spikes are associated with stabilization of

309 membrane potential maps in novel environments²⁶. However, while evoked plateau potentials
310 may be sufficient to induce place fields under some conditions⁹³, they do not appear to be
311 necessary for new place field generation in novel environments²⁶.

312
313 Further clues to the forms of plasticity promoting place field formation and stability come from
314 intracellular recordings from CA1 pyramidal cells in novel and familiar virtual environments²⁶. In
315 these experiments place field formation appears not to require firing of action potentials,
316 suggesting that the initial place field ramp is generated by sub-threshold forms of plasticity^{94,95}.
317 For example, isolated dendritic spikes in conjunction with presynaptic activity are sufficient to
318 induce LTP of the CA3 input to CA1 pyramidal cells⁹⁴. This form of spike-independent, localized
319 plasticity could explain why place cells appear rapidly in a novel environment^{96,97}. A similar
320 spike-independent LTP mechanism has also been described for CA3 pyramidal cells: powerful
321 proximal inputs from mossy fiber axons can induce synaptic plasticity even in the absence of
322 postsynaptic somatic spikes^{98,99}. This may enable sparse inputs from dentate gyrus granule
323 cells to efficiently generate active assemblies of CA3 pyramidal cells.

324
325 **Constraints on synaptic plasticity.** Distinct and spatially localised integrative mechanisms
326 may oppose synaptic plasticity. For example, HCN1 channels, which are highly enriched in the
327 distal dendrites of CA1 pyramidal neurons¹⁰⁰, suppress LTP of distal synaptic inputs⁵¹. By
328 depolarizing distal dendrites HCN1 channels prevent synaptically driven calcium transients
329 mediated by T-type Ca²⁺ channels, suggesting a mechanism to account for their actions on
330 LTP⁵². At a behavioural level deletion of HCN1 from forebrain neurons enhances hippocampal-
331 dependent forms of learning⁵¹, and increases the size and stability of CA1 place cell firing
332 fields¹⁰¹. Conversely, cannabinoid mediated enhancement of HCN1 channels reduces LTP and
333 suppresses hippocampal-dependent learning¹⁰². Together, these observations reinforce the idea
334 that compartmentalisation of synaptic integration contributes to spatial computations, and
335 suggest that HCN1 channels in distal dendrites control spatial firing and memory by gating
336 plasticity of direct cortical inputs.

337
338 A challenge in establishing roles of synaptic integrative mechanisms in memory is that the ion
339 channels implicated in control of synaptic plasticity may also influence membrane potential
340 ramps that drive spatial firing fields. For example, while HCN1 channels oppose distal synaptic
341 plasticity through their contribution to the resting membrane potential, HCN1 channels also
342 affect the waveform and temporal summation of distally originating post-synaptic potentials as
343 they propagate to the soma^{43,51,103}. Similarly, voltage-dependent gating of NMDARs contributes
344 directly to postsynaptic integration as well as providing a Ca²⁺ source for induction of plasticity.

345
346
347 **Theta oscillations and temporal codes**
348 The rate coded representations provided by place and grid fields are multiplexed with codes that
349 represent location through the timing of action potentials relative to the network theta (4-10 Hz)
350 rhythm^{20,104,105}. The theta rhythm is entrained by GABAergic projections from the medial septum
351 to interneurons in the hippocampus and entorhinal cortex¹⁰⁶⁻¹¹⁰. Because the relative delay
352 between theta cycles in the hippocampus and entorhinal cortex is greater than expected from

353 the synaptic delays between each area, the theta rhythm may establish temporal windows for
354 local circuit interactions¹¹¹. We will focus here on mechanisms by which the hippocampal
355 formation responds to theta modulated inputs and generates population level theta sequences.

356
357 **Responses to theta modulated signals.** How do neurons in the hippocampus and entorhinal
358 cortex respond to theta frequency synaptic inputs? The membrane potential response of
359 hippocampal CA1 pyramidal cells and stellate cells in the MEC to oscillating current inputs are
360 largest for oscillation frequencies in the theta range, whereas fast spiking interneurons may
361 prefer higher input frequencies¹¹²⁻¹¹⁵(Fig. 5a). At resting potentials the theta frequency
362 selectivity, or resonance, of pyramidal and stellate cells requires HCN1 channel mediated I_h
363 currents^{51,73,113,116}, whereas at depolarized potentials around spike threshold M-type K^+ channels
364 appear to be critical^{116,117}. Relatively slow voltage-dependent gating of both types of ion channel
365 leads to the appearance of resonance by opposing responses to input currents with frequencies
366 < 5 Hz. Resonance mechanisms directly affect spike output, by causing neurons to generate
367 greater numbers of spikes in response to inputs active near a cell's resonant frequency¹¹², and
368 may also modify the timing of action potentials driven by synaptic inputs at different phases of
369 the theta cycle¹¹⁸.

370
371 Do these single cell resonance phenomena manifest in vivo? Two lines of evidence suggest that
372 HCN1-dependent resonance is engaged during theta states. First, the amplitude of theta
373 frequency field potential oscillations recorded from CA1 is increased following genetic deletion
374 of HCN1^{51,101}. This is consistent with models of the contribution of dendritic HCN channels to the
375 local field potential¹¹⁹. Second, in behaving animals CA1 pyramidal cells respond preferentially
376 to activation of PV interneurons at theta frequencies¹²⁰ (Fig. 5b). This resonance effect is
377 abolished by pharmacological block of HCN channels¹²⁰. Interestingly, pyramidal cells did not
378 show theta frequency resonance upon direct optogenetic activation, suggesting that HCN
379 channel-dependent resonance, which would more effectively be engaged by hyperpolarizing
380 inhibition, may be more prevalent in behaving animals than peri-threshold resonance, which in
381 vitro does not require HCN channels¹¹⁶.

382
383 How does membrane potential resonance affect spatial computation? Ion channels contributing
384 to membrane potential resonance participate in sub-threshold theta frequency membrane
385 potential oscillations observed during in vitro recordings^{121,122}. These intrinsic oscillations have
386 been suggested to contribute to rate and temporal codes through oscillatory interference
387 mechanisms^{77,105,123}. However, this intrinsic oscillatory activity is suppressed by background
388 synaptic activity¹²⁴ and has not been observed in recordings from hippocampal or entorhinal
389 neurons in awake animals^{10-12,25}. Alternatively, by filtering signals with frequency outside the
390 theta band, resonance mechanisms may promote the emergence of temporal computations
391 within windows defined by theta oscillations¹¹¹.

392
393 **Theta phase precession and theta sequences.** As an animal moves through a cell's firing
394 field, an advance in the timing of the cell's action potentials relative to the theta rhythm (phase
395 precession) leads to the emergence of population level theta sequences²⁰. How theta rhythms
396 interact with synaptic inputs to cause phase precession and sequences is unresolved¹²⁵.

397 Several classes of model include components implemented by active integration mechanisms
398 (e.g. ^{21,105,126–128}). For example, ion channels that mediate spike frequency adaptation promote
399 symmetry of firing in models in which phase precession involves asymmetric ramp-like synaptic
400 inputs^{10,127,129,130}. In these models place fields are driven by an input current that rises slowly
401 and falls rapidly, with adaptation causing the spike rate to fall before the peak of the ramp. In a
402 detailed pyramidal cell model, sub-threshold membrane potential oscillations and resonance
403 promote phase precession that is generated by shifting the balance between oscillating
404 excitatory and inhibitory synaptic inputs¹²⁶. In simulations of grid cell firing a fast supralinear
405 dendritic integration mechanism sharpens phase precession by restricting time windows for
406 spike firing⁷⁸.

407
408 Experiments that focus on model predictions at the level of synaptic integration may help
409 distinguish between these various models and test roles for phase precession in spatial
410 behaviors. Because numerous models generate phase precession, an approach to evaluate
411 model predictions may be to also consider dependence of spatial codes on factors in addition to
412 location¹³¹. In this spirit, a recent model suggests integrative properties may be tuned to account
413 for dorsoventral differences in theta phase, and to maintain phase precession when running
414 speed varies²¹. While experiments with knockout mice show that HCN1 channels are not
415 required for either theta oscillations or for phase precession^{51,101,132}, identifying which ion
416 channels do play roles in phase precession may provide targets for testing contributions of theta
417 sequences to spatial memory.

418
419

420 **Concluding remarks**

421 The phenomena of rate and temporally coded spatial firing, when considered at the level of
422 membrane potential dynamics, appear to arise from complex and multi-layered mechanisms,
423 with synaptic integration playing critical roles at multiple key points. For example, in CA1
424 pyramidal cells, active integrative mechanisms contribute to selection of active cells, membrane
425 potential dynamics driving firing, plasticity of synaptic inputs, and responses to oscillatory
426 network activity. Entorhinal grid cells also appear to engage specific integrative mechanisms,
427 but intriguingly these may be distinct from those used in CA1. In our view, experiments to date
428 may only be scratching the surface of a rich diversity of dynamic integrative mechanisms
429 underlying the well defined spatial firing properties of neurons in the hippocampal formation. We
430 end by outlining areas that may be of importance to future investigation of mechanisms for
431 spatial computation in these circuits.

432
433 Spatial cognition involves numerous cell types not considered here. For example, principal cells
434 in CA3 and the dentate gyrus employ distinct integrative mechanisms. Dendritic regenerative
435 events can be readily evoked in CA3 pyramidal neurons^{133,134}. In contrast, while distinct
436 regenerative events have not been observed in direct recordings from granule cell dendrites¹³⁵,
437 their NMDAR-dependent sensitivity to sequences of synaptic inputs¹³⁶ and pronounced dendritic
438 Ca²⁺ transients during backpropagating action potentials¹³⁷ suggest that nonlinear dendritic
439 conductances can be recruited. In agreement with this view, selective deletion of NMDARs in
440 the dentate gyrus causes deficits in rapidly producing a unique memory of a novel context, and

441 discriminating it from previously encountered contexts¹³⁸. The dendrites of granule cells may
442 support this “pattern separation” function by increasing the sparsity of firing; assuming the same
443 number and weights of synaptic inputs, a granule cell is less likely to fire a spike if it has more
444 dendrites¹³⁹. This sparsification of firing may further be enhanced by short coincidence detection
445 windows for EPSPs in granule cell dendrites¹⁴⁰.

446

447 How synaptic integrative properties contribute to the spatial codes of border, head-direction and
448 other spatially modulated neurons is an open target for future investigation. Likewise, network
449 activity patterns such as sharp wave ripples and associated spike sequences, and gamma
450 oscillations that co-occur with theta states, may also be shaped by active synaptic integrative
451 mechanisms. For example, during sharp wave ripples synaptic inhibition may dynamically re-
452 configure synaptic integration by CA1 pyramidal cells^{141,142}.

453

454 Synaptic integrative properties are dynamically regulated by neuromodulatory systems
455 according to brain state and behavioural demands¹⁴³. Ion channels that mediate integration of
456 synaptic responses are prime targets for neuromodulators¹⁴⁴, raising questions about how these
457 systems influence spatial computations in the behaving animal. Recent studies indicate highly
458 selective roles for certain neuromodulators. For example, cholinergic inputs increase excitability
459 of dentate gyrus granule cells by stimulating interactions between axonal T-type Ca²⁺ channels
460 and Kv7 channels¹⁴⁵. At a systems level, computational models incorporating neuromodulatory
461 systems provide frameworks for predicting how modulation of ion channels important for
462 synaptic integration contributes to circuit computations and behaviour¹⁴⁶. Future investigation of
463 interactions between neuromodulation and synaptic integration may lead to important insights
464 into control mechanisms for spatial cognition.

465

466 New tools will be critical to the twin challenges of selective experimental manipulation of
467 synaptic integrative mechanisms and observation of the subcellular membrane potential
468 dynamics on which they act. Promising strategies for manipulation include optical control of
469 native or engineered light-sensitive ion channels¹⁴⁷. For example, rapid optical block of HCN1
470 channels in distal dendrites of CA1 pyramidal cells may help resolve the question of whether
471 their influence on spatial firing fields is through control of synaptic plasticity, or by effects on the
472 waveform of synaptic responses that propagate to the soma. Novel imaging approaches,
473 including miniaturization of microscope technologies¹⁴⁸ and development of fluorescent voltage-
474 sensors¹⁴⁹ will facilitate exploration of the impacts of synaptic integrative mechanisms on sub-
475 cellular and network level computations. For example, measurement of resting membrane
476 potential and spike threshold across populations of neurons in behaving animals will facilitate
477 direct testing of the contributions of excitability to selection of active place cells.

478

479 Finally, it is intriguing to consider whether synaptic integrative mechanisms contributing to
480 spatial computations are similarly used in other neural systems. On the one hand, the evidence
481 we have considered points towards diversity in strategies for synaptic integrative mechanisms to
482 influence neural computation. On the other hand, nonlinear synaptic integrative mechanisms
483 engaged to drive receptive fields in visual cortex⁸¹ appear similar to those used by CA1
484 pyramidal cells. As we highlight above, the effect of inhibition on the shape of sub- and

485 suprathreshold spatially receptive fields in place cells^{62,66} also bears resemblance with
486 observations from orientation-sensitive neurons in visual cortex^{60,63,64}. Establishing how
487 common synaptic integrative mechanisms are adapted to the specific computations carried out
488 by different circuits would be a major achievement for cellular and systems neuroscience.

489
490

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499 **References**

- 500 1. O'Keefe, J. & Nadel, L. *The hippocampus as a cognitive map*. (Oxford: Clarendon Press,
501 1978).
- 502 2. Redish, A. D. *Beyond the Cognitive Map: From Place Cells to Episodic Memory*. (MIT
503 Press, 1999).
- 504 3. Spruston, N. Pyramidal neurons: dendritic structure and synaptic integration. *Nat. Rev.*
505 *Neurosci.* **9**, 206–221 (2008).
- 506 4. Häusser, M., Spruston, N. & Stuart, G. J. Diversity and dynamics of dendritic signaling.
507 *Science* **290**, 739–744 (2000).
- 508 5. Chadderton, P., Schaefer, A. T., Williams, S. R. & Margrie, T. W. Sensory-evoked synaptic
509 integration in cerebellar and cerebral cortical neurons. *Nat. Rev. Neurosci.* **15**, 71–83
510 (2014).
- 511 6. LeCun, Y., Bengio, Y. & Hinton, G. Deep learning. *Nature* **521**, 436–444 (2015).
- 512 7. Häusser, M. & Mel, B. Dendrites: bug or feature? *Curr. Opin. Neurobiol.* **13**, 372–383
513 (2003).
- 514 8. O'Keefe, J., Burgess, N., Donnett, J. G., Jeffery, K. J. & Maguire, E. A. Place cells,
515 navigational accuracy, and the human hippocampus. *Philos. Trans. R. Soc. Lond. B Biol.*
516 *Sci.* **353**, 1333–1340 (1998).

- 517 9. Hafting, T., Fyhn, M., Molden, S., Moser, M.-B. & Moser, E. I. Microstructure of a spatial
518 map in the entorhinal cortex. *Nature* **436**, 801–806 (2005).
- 519 10. Harvey, C. D., Collman, F., Dombeck, D. A. & Tank, D. W. Intracellular dynamics of
520 hippocampal place cells during virtual navigation. *Nature* **461**, 941–946 (2009).
- 521 11. Schmidt-Hieber, C. & Häusser, M. Cellular mechanisms of spatial navigation in the medial
522 entorhinal cortex. *Nat. Neurosci.* **16**, 325–331 (2013).
- 523 12. Domnisoru, C., Kinkhabwala, A. A. & Tank, D. W. Membrane potential dynamics of grid
524 cells. *Nature* **495**, 199–204 (2013).
- 525 13. Hartley, T., Burgess, N., Lever, C., Cacucci, F. & O’Keefe, J. Modeling place fields in terms
526 of the cortical inputs to the hippocampus. *Hippocampus* **10**, 369–379 (2000).
- 527 14. Burgess, N. & O’Keefe, J. Models of place and grid cell firing and theta rhythmicity. *Curr.*
528 *Opin. Neurobiol.* **21**, 734–744 (2011).
- 529 15. Solstad, T., Moser, E. I. & Einevoll, G. T. From grid cells to place cells: a mathematical
530 model. *Hippocampus* **16**, 1026–1031 (2006).
- 531 16. Eichenbaum, H., Dudchenko, P., Wood, E., Shapiro, M. & Tanila, H. The hippocampus,
532 memory, and place cells: is it spatial memory or a memory space? *Neuron* **23**, 209–226
533 (1999).
- 534 17. Burgess, N., Maguire, E. A. & O’Keefe, J. The human hippocampus and spatial and
535 episodic memory. *Neuron* **35**, 625–641 (2002).
- 536 18. Chrobak, J. J., Lörcincz, A. & Buzsáki, G. Physiological patterns in the hippocampo-
537 entorhinal cortex system. *Hippocampus* **10**, 457–465 (2000).
- 538 19. O’Keefe, J. Place units in the hippocampus of the freely moving rat. *Exp. Neurol.* **51**, 78–
539 109 (1976).
- 540 20. Skaggs, W. E., McNaughton, B. L., Wilson, M. A. & Barnes, C. A. Theta phase precession
541 in hippocampal neuronal populations and the compression of temporal sequences.
542 *Hippocampus* **6**, 149–172 (1996).

- 543 21. Chadwick, A., van Rossum, M. C. & Nolan, M. F. Flexible theta sequence compression
544 mediated via phase precessing interneurons. *Elife* **5**, (2016).
- 545 22. Ziv, Y. *et al.* Long-term dynamics of CA1 hippocampal place codes. *Nat. Neurosci.* **16**,
546 264–266 (2013).
- 547 23. Rich, P. D., Liaw, H.-P. & Lee, A. K. Place cells. Large environments reveal the statistical
548 structure governing hippocampal representations. *Science* **345**, 814–817 (2014).
- 549 24. Thompson, L. T. & Best, P. J. Place cells and silent cells in the hippocampus of freely-
550 behaving rats. *J. Neurosci.* **9**, 2382–2390 (1989).
- 551 25. Epsztein, J., Brecht, M. & Lee, A. K. Intracellular determinants of hippocampal CA1 place
552 and silent cell activity in a novel environment. *Neuron* **70**, 109–120 (2011).
- 553 26. Cohen, J. D., Bolstad, M. & Lee, A. K. Experience-dependent shaping of hippocampal CA1
554 intracellular activity in novel and familiar environments. *Elife* **6**, e23040 (2017).
- 555 27. Silva, A. J., Zhou, Y., Rogerson, T., Shobe, J. & Balaji, J. Molecular and cellular
556 approaches to memory allocation in neural circuits. *Science* **326**, 391–395 (2009).
- 557 28. Zhou, Y. *et al.* CREB regulates excitability and the allocation of memory to subsets of
558 neurons in the amygdala. *Nat. Neurosci.* **12**, 1438–1443 (2009).
- 559 29. Yiu, A. P. *et al.* Neurons are recruited to a memory trace based on relative neuronal
560 excitability immediately before training. *Neuron* **83**, 722–735 (2014).
- 561 30. Rashid, A. J. *et al.* Competition between engrams influences fear memory formation and
562 recall. *Science* **353**, 383–387 (2016).
- 563 31. Rubin, A., Geva, N., Sheintuch, L. & Ziv, Y. Hippocampal ensemble dynamics timestamp
564 events in long-term memory. *Elife* **4**, (2015).
- 565 32. Cai, D. J. *et al.* A shared neural ensemble links distinct contextual memories encoded close
566 in time. *Nature* **534**, 115–118 (2016).
- 567 33. Kastellakis, G., Silva, A. J. & Poirazi, P. Linking Memories across Time via Neuronal and
568 Dendritic Overlaps in Model Neurons with Active Dendrites. *Cell Rep.* **17**, 1491–1504

- 569 (2016).
- 570 34. Dudek, S. M. & Fields, R. D. Somatic action potentials are sufficient for late-phase LTP-
571 related cell signaling. *Proc. Natl. Acad. Sci. U. S. A.* **99**, 3962–3967 (2002).
- 572 35. Deisseroth, K., Bitto, H. & Tsien, R. W. Signaling from synapse to nucleus: postsynaptic
573 CREB phosphorylation during multiple forms of hippocampal synaptic plasticity. *Neuron* **16**,
574 89–101 (1996).
- 575 36. Lopez de Armentia, M. *et al.* cAMP response element-binding protein-mediated gene
576 expression increases the intrinsic excitability of CA1 pyramidal neurons. *J. Neurosci.* **27**,
577 13909–13918 (2007).
- 578 37. Disterhoft, J. F. & Oh, M. M. Learning, aging and intrinsic neuronal plasticity. *Trends*
579 *Neurosci.* **29**, 587–599 (2006).
- 580 38. Lovett-Barron, M. *et al.* Dendritic inhibition in the hippocampus supports fear learning.
581 *Science* **343**, 857–863 (2014).
- 582 39. Stefanelli, T., Bertollini, C., Lüscher, C., Muller, D. & Mendez, P. Hippocampal Somatostatin
583 Interneurons Control the Size of Neuronal Memory Ensembles. *Neuron* **89**, 1074–1085
584 (2016).
- 585 40. Mankin, E. A. *et al.* Neuronal code for extended time in the hippocampus. *Proc. Natl. Acad.*
586 *Sci. U. S. A.* **109**, 19462–19467 (2012).
- 587 41. Mankin, E. A., Diehl, G. W., Sparks, F. T., Leutgeb, S. & Leutgeb, J. K. Hippocampal CA2
588 activity patterns change over time to a larger extent than between spatial contexts. *Neuron*
589 **85**, 190–201 (2015).
- 590 42. Lipowsky, R., Gillessen, T. & Alzheimer, C. Dendritic Na⁺ channels amplify EPSPs in
591 hippocampal CA1 pyramidal cells. *J. Neurophysiol.* **76**, 2181–2191 (1996).
- 592 43. Magee, J. C. Dendritic Ih normalizes temporal summation in hippocampal CA1 neurons.
593 *Nat. Neurosci.* **2**, 508–514 (1999).
- 594 44. Ngo-Anh, T. J. *et al.* SK channels and NMDA receptors form a Ca²⁺-mediated feedback

- 595 loop in dendritic spines. *Nat. Neurosci.* **8**, 642–649 (2005).
- 596 45. Losonczy, A., Makara, J. K. & Magee, J. C. Compartmentalized dendritic plasticity and
597 input feature storage in neurons. *Nature* **452**, 436–441 (2008).
- 598 46. Amaral, D. G. & Witter, M. P. The three-dimensional organization of the hippocampal
599 formation: a review of anatomical data. *Neuroscience* **31**, 571–591 (1989).
- 600 47. Freund, T. F. & Buzsáki, G. Interneurons of the hippocampus. *Hippocampus* **6**, 347–470
601 (1996).
- 602 48. Middleton, S. J. & McHugh, T. J. Silencing CA3 disrupts temporal coding in the CA1
603 ensemble. *Nat. Neurosci.* **19**, 945–951 (2016).
- 604 49. Schlesiger, M. I. *et al.* The medial entorhinal cortex is necessary for temporal organization
605 of hippocampal neuronal activity. *Nat. Neurosci.* **18**, 1123–1132 (2015).
- 606 50. Magee, J. C. Dendritic integration of excitatory synaptic input. *Nat. Rev. Neurosci.* **1**, 181–
607 190 (2000).
- 608 51. Nolan, M. F. *et al.* A behavioral role for dendritic integration: HCN1 channels constrain
609 spatial memory and plasticity at inputs to distal dendrites of CA1 pyramidal neurons. *Cell*
610 **119**, 719–732 (2004).
- 611 52. Tsay, D., Dudman, J. T. & Siegelbaum, S. A. HCN1 channels constrain synaptically evoked
612 Ca²⁺ spikes in distal dendrites of CA1 pyramidal neurons. *Neuron* **56**, 1076–1089 (2007).
- 613 53. Cai, X. *et al.* Unique roles of SK and Kv4.2 potassium channels in dendritic integration.
614 *Neuron* **44**, 351–364 (2004).
- 615 54. Lee, D., Lin, B.-J. & Lee, A. K. Hippocampal place fields emerge upon single-cell
616 manipulation of excitability during behavior. *Science* **337**, 849–853 (2012).
- 617 55. Nevian, T., Larkum, M. E., Polsky, A. & Schiller, J. Properties of basal dendrites of layer 5
618 pyramidal neurons: a direct patch-clamp recording study. *Nat. Neurosci.* **10**, 206–214
619 (2007).
- 620 56. Druckmann, S. *et al.* Structured synaptic connectivity between hippocampal regions.

621 *Neuron* **81**, 629–640 (2014).

622 57. Wilson, D. E., Whitney, D. E., Scholl, B. & Fitzpatrick, D. Orientation selectivity and the
623 functional clustering of synaptic inputs in primary visual cortex. *Nat. Neurosci.* **19**, 1003–
624 1009 (2016).

625 58. Iacaruso, M. F., Gasler, I. T. & Hofer, S. B. Synaptic organization of visual space in primary
626 visual cortex. *Nature* (2017). doi:10.1038/nature23019

627 59. Haider, B., Häusser, M. & Carandini, M. Inhibition dominates sensory responses in the
628 awake cortex. *Nature* **493**, 97–100 (2013).

629 60. Atallah, B. V., Bruns, W., Carandini, M. & Scanziani, M. Parvalbumin-expressing
630 interneurons linearly transform cortical responses to visual stimuli. *Neuron* **73**, 159–170
631 (2012).

632 61. Atallah, B. V., Scanziani, M. & Carandini, M. Atallah et al. reply. *Nature* **508**, E3 (2014).

633 62. Grienberger, C., Milstein, A. D., Bittner, K. C., Romani, S. & Magee, J. C. Inhibitory
634 suppression of heterogeneously tuned excitation enhances spatial coding in CA1 place
635 cells. *Nat. Neurosci.* (2017). doi:10.1038/nn.4486

636 63. Wilson, N. R., Runyan, C. A., Wang, F. L. & Sur, M. Division and subtraction by distinct
637 cortical inhibitory networks in vivo. *Nature* **488**, 343–348 (2012).

638 64. Lee, S.-H. *et al.* Activation of specific interneurons improves V1 feature selectivity and
639 visual perception. *Nature* **488**, 379–383 (2012).

640 65. Cottam, J. C. H., Smith, S. L. & Häusser, M. Target-specific effects of somatostatin-
641 expressing interneurons on neocortical visual processing. *J. Neurosci.* **33**, 19567–19578
642 (2013).

643 66. Royer, S. *et al.* Control of timing, rate and bursts of hippocampal place cells by dendritic
644 and somatic inhibition. *Nat. Neurosci.* **15**, 769–775 (2012).

645 67. Yoon, K. *et al.* Specific evidence of low-dimensional continuous attractor dynamics in grid
646 cells. *Nat. Neurosci.* **16**, 1077–1084 (2013).

- 647 68. Fyhn, M., Hafting, T., Treves, A., Moser, M.-B. & Moser, E. I. Hippocampal remapping and
648 grid realignment in entorhinal cortex. *Nature* **446**, 190–194 (2007).
- 649 69. Pastoll, H., Ramsden, H. L. & Nolan, M. F. Intrinsic electrophysiological properties of
650 entorhinal cortex stellate cells and their contribution to grid cell firing fields. *Front. Neural*
651 *Circuits* **6**, 17 (2012).
- 652 70. Ray, S. *et al.* Grid-layout and theta-modulation of layer 2 pyramidal neurons in medial
653 entorhinal cortex. *Science* **343**, 891–896 (2014).
- 654 71. Couey, J. J. *et al.* Recurrent inhibitory circuitry as a mechanism for grid formation. *Nat.*
655 *Neurosci.* **16**, 318–324 (2013).
- 656 72. Pastoll, H., Solanka, L., van Rossum, M. C. W. & Nolan, M. F. Feedback inhibition enables
657 θ -nested γ oscillations and grid firing fields. *Neuron* **77**, 141–154 (2013).
- 658 73. Nolan, M. F., Dudman, J. T., Dodson, P. D. & Santoro, B. HCN1 channels control resting
659 and active integrative properties of stellate cells from layer II of the entorhinal cortex. *J.*
660 *Neurosci.* **27**, 12440–12451 (2007).
- 661 74. Giocomo, L. M. *et al.* Grid cells use HCN1 channels for spatial scaling. *Cell* **147**, 1159–
662 1170 (2011).
- 663 75. Robinson, R. B. & Siegelbaum, S. A. Hyperpolarization-activated cation currents: from
664 molecules to physiological function. *Annu. Rev. Physiol.* **65**, 453–480 (2003).
- 665 76. Garden, D. L. F., Dodson, P. D., O'Donnell, C., White, M. D. & Nolan, M. F. Tuning of
666 synaptic integration in the medial entorhinal cortex to the organization of grid cell firing
667 fields. *Neuron* **60**, 875–889 (2008).
- 668 77. Giocomo, L. M., Zilli, E. A., Fransén, E. & Hasselmo, M. E. Temporal frequency of
669 subthreshold oscillations scales with entorhinal grid cell field spacing. *Science* **315**, 1719–
670 1722 (2007).
- 671 78. Schmidt-Hieber, C. *et al.* Active dendritic integration as a mechanism for robust and precise
672 grid cell firing. *Nat. Neurosci.* (2017). doi:10.1038/nn.4582

- 673 79. Palmer, L. M. *et al.* NMDA spikes enhance action potential generation during sensory input.
674 *Nat. Neurosci.* **17**, 383–390 (2014).
- 675 80. Lavzin, M., Rapoport, S., Polsky, A., Garion, L. & Schiller, J. Nonlinear dendritic processing
676 determines angular tuning of barrel cortex neurons in vivo. *Nature* **490**, 397–401 (2012).
- 677 81. Smith, S. L., Smith, I. T., Branco, T. & Häusser, M. Dendritic spikes enhance stimulus
678 selectivity in cortical neurons in vivo. *Nature* **503**, 115–120 (2013).
- 679 82. Poirazi, P. & Mel, B. W. Impact of active dendrites and structural plasticity on the memory
680 capacity of neural tissue. *Neuron* **29**, 779–796 (2001).
- 681 83. Buzsáki, G. & Moser, E. I. Memory, navigation and theta rhythm in the hippocampal-
682 entorhinal system. *Nat. Neurosci.* **16**, 130–138 (2013).
- 683 84. Martin, S. J., Grimwood, P. D. & Morris, R. G. Synaptic plasticity and memory: an
684 evaluation of the hypothesis. *Annu. Rev. Neurosci.* **23**, 649–711 (2000).
- 685 85. Bliss, T. V. & Collingridge, G. L. A synaptic model of memory: long-term potentiation in the
686 hippocampus. *Nature* **361**, 31–39 (1993).
- 687 86. Morris, R. G., Anderson, E., Lynch, G. S. & Baudry, M. Selective impairment of learning
688 and blockade of long-term potentiation by an N-methyl-D-aspartate receptor antagonist,
689 AP5. *Nature* **319**, 774–776 (1986).
- 690 87. Tsien, J. Z., Huerta, P. T. & Tonegawa, S. The essential role of hippocampal CA1 NMDA
691 receptor-dependent synaptic plasticity in spatial memory. *Cell* **87**, 1327–1338 (1996).
- 692 88. Kentros, C. *et al.* Abolition of long-term stability of new hippocampal place cell maps by
693 NMDA receptor blockade. *Science* **280**, 2121–2126 (1998).
- 694 89. McHugh, T. J., Blum, K. I., Tsien, J. Z., Tonegawa, S. & Wilson, M. A. Impaired
695 hippocampal representation of space in CA1-specific NMDAR1 knockout mice. *Cell* **87**,
696 1339–1349 (1996).
- 697 90. Moosmang, S. *et al.* Role of hippocampal Cav1.2 Ca²⁺ channels in NMDA receptor-
698 independent synaptic plasticity and spatial memory. *J. Neurosci.* **25**, 9883–9892 (2005).

- 699 91. Sheffield, M. E. J. & Dombeck, D. A. Calcium transient prevalence across the dendritic
700 arbour predicts place field properties. *Nature* **517**, 200–204 (2015).
- 701 92. Takahashi, H. & Magee, J. C. Pathway interactions and synaptic plasticity in the dendritic
702 tuft regions of CA1 pyramidal neurons. *Neuron* **62**, 102–111 (2009).
- 703 93. Bittner, K. C. *et al.* Conjunctive input processing drives feature selectivity in hippocampal
704 CA1 neurons. *Nat. Neurosci.* **18**, 1133–1142 (2015).
- 705 94. Golding, N. L., Staff, N. P. & Spruston, N. Dendritic spikes as a mechanism for cooperative
706 long-term potentiation. *Nature* **418**, 326–331 (2002).
- 707 95. Dudman, J. T., Tsay, D. & Siegelbaum, S. A. A role for synaptic inputs at distal dendrites:
708 instructive signals for hippocampal long-term plasticity. *Neuron* **56**, 866–879 (2007).
- 709 96. Frank, L. M. Hippocampal Plasticity across Multiple Days of Exposure to Novel
710 Environments. *Journal of Neuroscience* **24**, 7681–7689 (2004).
- 711 97. Hill, A. J. First occurrence of hippocampal spatial firing in a new environment. *Exp. Neurol.*
712 **62**, 282–297 (1978).
- 713 98. Brandalise, F. & Gerber, U. Mossy fiber-evoked subthreshold responses induce timing-
714 dependent plasticity at hippocampal CA3 recurrent synapses. *Proc. Natl. Acad. Sci. U. S.*
715 *A.* **111**, 4303–4308 (2014).
- 716 99. Brandalise, F., Carta, S., Helmchen, F., Lisman, J. & Gerber, U. Dendritic NMDA spikes are
717 necessary for timing-dependent associative LTP in CA3 pyramidal cells. *Nat. Commun.* **7**,
718 13480 (2016).
- 719 100. Lörincz, A., Notomi, T., Tamás, G., Shigemoto, R. & Nusser, Z. Polarized and
720 compartment-dependent distribution of HCN1 in pyramidal cell dendrites. *Nat. Neurosci.* **5**,
721 1185–1193 (2002).
- 722 101. Hussaini, S. A., Kempadoo, K. A., Thuault, S. J., Siegelbaum, S. A. & Kandel, E. R.
723 Increased size and stability of CA1 and CA3 place fields in HCN1 knockout mice. *Neuron*
724 **72**, 643–653 (2011).

- 725 102. Maroso, M. *et al.* Cannabinoid Control of Learning and Memory through HCN Channels.
726 *Neuron* **89**, 1059–1073 (2016).
- 727 103. Magee, J. C. Dendritic Ih normalizes temporal summation in hippocampal CA1 neurons.
728 *Nat. Neurosci.* **2**, 848 (1999).
- 729 104. Buzsáki, G. Theta oscillations in the hippocampus. *Neuron* **33**, 325–340 (2002).
- 730 105. O’Keefe, J. & Recce, M. L. Phase relationship between hippocampal place units and the
731 EEG theta rhythm. *Hippocampus* **3**, 317–330 (1993).
- 732 106. Bender, F. *et al.* Theta oscillations regulate the speed of locomotion via a hippocampus to
733 lateral septum pathway. *Nat. Commun.* **6**, 8521 (2015).
- 734 107. Freund, T. F. & Antal, M. GABA-containing neurons in the septum control inhibitory
735 interneurons in the hippocampus. *Nature* **336**, 170–173 (1988).
- 736 108. Boyce, R., Glasgow, S. D., Williams, S. & Adamantidis, A. Causal evidence for the role of
737 REM sleep theta rhythm in contextual memory consolidation. *Science* **352**, 812–816
738 (2016).
- 739 109. Gonzalez-Sulser, A. *et al.* GABAergic projections from the medial septum selectively inhibit
740 interneurons in the medial entorhinal cortex. *J. Neurosci.* **34**, 16739–16743 (2014).
- 741 110. Tóth, K., Freund, T. F. & Miles, R. Disinhibition of rat hippocampal pyramidal cells by
742 GABAergic afferents from the septum. *J. Physiol.* **500 (Pt 2)**, 463–474 (1997).
- 743 111. Mizuseki, K., Sirota, A., Pastalkova, E. & Buzsáki, G. Theta oscillations provide temporal
744 windows for local circuit computation in the entorhinal-hippocampal loop. *Neuron* **64**, 267–
745 280 (2009).
- 746 112. Pike, F. G. *et al.* Distinct frequency preferences of different types of rat hippocampal
747 neurones in response to oscillatory input currents. *J. Physiol.* **529 Pt 1**, 205–213 (2000).
- 748 113. Zemankovics, R., Káli, S. & Paulsen, O. Differences in subthreshold resonance of
749 hippocampal pyramidal cells and interneurons: the role of h⁺ current and passive
750 membrane characteristics. *The Journal of* (2010).

- 751 114. Leung, L. S. & Yu, H. W. Theta-frequency resonance in hippocampal CA1 neurons in vitro
752 demonstrated by sinusoidal current injection. *J. Neurophysiol.* **79**, 1592–1596 (1998).
- 753 115. Erchova, I., Kreck, G., Heinemann, U. & Herz, A. V. M. Dynamics of rat entorhinal cortex
754 layer II and III cells: characteristics of membrane potential resonance at rest predict
755 oscillation properties near threshold. *J. Physiol.* **560**, 89–110 (2004).
- 756 116. Hu, H., Vervaeke, K. & Storm, J. F. Two forms of electrical resonance at theta frequencies,
757 generated by M-current, h-current and persistent Na⁺ current in rat hippocampal pyramidal
758 cells. *J. Physiol.* **545**, 783–805 (2002).
- 759 117. Peters, H. C., Hu, H., Pongs, O., Storm, J. F. & Isbrandt, D. Conditional transgenic
760 suppression of M channels in mouse brain reveals functions in neuronal excitability,
761 resonance and behavior. *Nat. Neurosci.* **8**, 51–60 (2005).
- 762 118. Borel, M., Guadagna, S., Jang, H. J., Kwag, J. & Paulsen, O. Frequency dependence of
763 CA3 spike phase response arising from h-current properties. *Front. Cell. Neurosci.* **7**, 263
764 (2013).
- 765 119. Ness, T. V., Remme, M. W. H. & Einevoll, G. T. Active subthreshold dendritic conductances
766 shape the local field potential. *J. Physiol.* **594**, 3809–3825 (2016).
- 767 120. Stark, E. *et al.* Inhibition-induced theta resonance in cortical circuits. *Neuron* **80**, 1263–1276
768 (2013).
- 769 121. Alonso, A. & Llinás, R. R. Subthreshold Na⁺-dependent theta-like rhythmicity in stellate
770 cells of entorhinal cortex layer II. *Nature* **342**, 175–177 (1989).
- 771 122. Leung, L. W. & Yim, C. Y. Intrinsic membrane potential oscillations in hippocampal neurons
772 in vitro. *Brain Res.* **553**, 261–274 (1991).
- 773 123. O'Keefe, J. & Burgess, N. Dual phase and rate coding in hippocampal place cells:
774 theoretical significance and relationship to entorhinal grid cells. *Hippocampus* **15**, 853–866
775 (2005).
- 776 124. Fernandez, F. R. & White, J. A. Artificial synaptic conductances reduce subthreshold

777 oscillations and periodic firing in stellate cells of the entorhinal cortex. *J. Neurosci.* **28**,
778 3790–3803 (2008).

779 125. Jaramillo, J. & Kempter, R. Phase precession: a neural code underlying episodic memory?
780 *Curr. Opin. Neurobiol.* **43**, 130–138 (2017).

781 126. Leung, L. S. A model of intracellular θ phase precession dependent on intrinsic
782 subthreshold membrane currents. *J. Neurosci.* **31**, 12282–12296 (2011).

783 127. Harris, K. D. *et al.* Spike train dynamics predicts theta-related phase precession in
784 hippocampal pyramidal cells. *Nature* **417**, 738–741 (2002).

785 128. Magee, J. C. Dendritic mechanisms of phase precession in hippocampal CA1 pyramidal
786 neurons. *J. Neurophysiol.* **86**, 528–532 (2001).

787 129. Mehta, M. R., Lee, A. K. & Wilson, M. A. Role of experience and oscillations in transforming
788 a rate code into a temporal code. *Nature* **417**, 741–746 (2002).

789 130. Mehta, M. R., Quirk, M. C. & Wilson, M. A. Experience-dependent asymmetric shape of
790 hippocampal receptive fields. *Neuron* **25**, 707–715 (2000).

791 131. Chadwick, A., van Rossum, M. C. W. & Nolan, M. F. Independent theta phase coding
792 accounts for CA1 population sequences and enables flexible remapping. *Elife* **4**, (2015).

793 132. Eggink, H., Mertens, P., Storm, E. & Giocomo, L. M. Hyperpolarization-activated cyclic
794 nucleotide-gated 1 independent grid cell-phase precession in mice. *Hippocampus* **24**, 249–
795 256 (2014).

796 133. Kim, S., Guzman, S. J., Hu, H. & Jonas, P. Active dendrites support efficient initiation of
797 dendritic spikes in hippocampal CA3 pyramidal neurons. *Nat. Neurosci.* **15**, 600–606
798 (2012).

799 134. Makara, J. K. & Magee, J. C. Variable dendritic integration in hippocampal CA3 pyramidal
800 neurons. *Neuron* **80**, 1438–1450 (2013).

801 135. Krueppel, R., Remy, S. & Beck, H. Dendritic Integration in Hippocampal Dentate Granule
802 Cells. *Neuron* **71**, 512–528 (2011).

- 803 136. Branco, T., Clark, B. A. & Häusser, M. Dendritic discrimination of temporal input sequences
804 in cortical neurons. *Science* **329**, 1671–1675 (2010).
- 805 137. Stocca, G., Schmidt-Hieber, C. & Bischofberger, J. Differential dendritic Ca²⁺ signalling in
806 young and mature hippocampal granule cells. *J. Physiol.* **586**, 3795–3811 (2008).
- 807 138. McHugh, T. J. *et al.* Dentate gyrus NMDA receptors mediate rapid pattern separation in the
808 hippocampal network. *Science* **317**, 94–99 (2007).
- 809 139. Chavlis, S., Petrantonakis, P. C. & Poirazi, P. Dendrites of dentate gyrus granule cells
810 contribute to pattern separation by controlling sparsity. *Hippocampus* **27**, 89–110 (2017).
- 811 140. Schmidt-Hieber, C., Jonas, P. & Bischofberger, J. Subthreshold dendritic signal processing
812 and coincidence detection in dentate gyrus granule cells. *J. Neurosci.* **27**, 8430–8441
813 (2007).
- 814 141. Valero, M. *et al.* Determinants of different deep and superficial CA1 pyramidal cell
815 dynamics during sharp-wave ripples. *Nat. Neurosci.* **18**, 1281–1290 (2015).
- 816 142. Gan, J., Weng, S.-M., Pernía-Andrade, A. J., Csicsvari, J. & Jonas, P. Phase-Locked
817 Inhibition, but Not Excitation, Underlies Hippocampal Ripple Oscillations in Awake Mice In
818 Vivo. *Neuron* **93**, 308–314 (2017).
- 819 143. Cobb, S. & Lawrence, J. J. Neuromodulation of Hippocampal Cells and Circuits. in
820 *Hippocampal Microcircuits* 187–246 (2010).
- 821 144. Marder, E. Neuromodulation of neuronal circuits: back to the future. *Neuron* **76**, 1–11
822 (2012).
- 823 145. Martinello, K. *et al.* Cholinergic afferent stimulation induces axonal function plasticity in
824 adult hippocampal granule cells. *Neuron* **85**, 346–363 (2015).
- 825 146. Hasselmo, M. E., Hay, J., Ilyn, M. & Gorchetchnikov, A. Neuromodulation, theta rhythm and
826 rat spatial navigation. *Neural Netw.* **15**, 689–707 (2002).
- 827 147. Kramer, R. H., Mouro, A. & Adesnik, H. Optogenetic pharmacology for control of native
828 neuronal signaling proteins. *Nat. Neurosci.* **16**, 816–823 (2013).

- 829 148. Hamel, E. J. O., Grewe, B. F., Parker, J. G. & Schnitzer, M. J. Cellular level brain imaging
830 in behaving mammals: an engineering approach. *Neuron* **86**, 140–159 (2015).
- 831 149. Gong, Y. *et al.* High-speed recording of neural spikes in awake mice and flies with a
832 fluorescent voltage sensor. *Science* **350**, 1361–1366 (2015).
- 833 150. Koenig, J., Linder, A. N., Leutgeb, J. K. & Leutgeb, S. The spatial periodicity of grid cells is
834 not sustained during reduced theta oscillations. *Science* **332**, 592–595 (2011).

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837 **Figure 1.** Membrane potential ramp and intracellular phase precession during place and grid
838 field crossings. (a) Top panels show example recordings from a place cell (left, adapted from
839 ref. ¹⁵⁰) and from a grid cell (right, adapted from ref. ⁹). Bottom panels show simultaneous
840 membrane potential and LFP recordings during firing field crossings. In both CA1 pyramidal
841 cells (left, adapted from ref. ⁶²) and in MEC stellate cells (right, adapted from ref. ¹¹), firing during
842 field crossings is driven by a sustained membrane potential depolarization^{10–12}. (b) Average
843 firing rate (upper) and membrane potential (lower) of CA1 pyramidal cells (left, adapted from ref.
844 ¹⁰) and MEC stellate cells (right, adapted from ref. ¹¹) during field crossings. (c) Action potential
845 phase relative to the local field potential, membrane potential theta oscillation phase with
846 respect to the local field potential and action potential phase with respect to the membrane
847 potential theta oscillation, are each plotted as a function of position within the rate coded firing
848 field, for a CA1 pyramidal cell (left, adapted from ref. ¹⁰) and MEC stellate cells (right, adapted
849 from ref. ¹¹).

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852 **Figure 2.** Excitability and place cell selection. (a) Schematised differences between excitable
853 and silent cells recorded in a novel environment²⁵. Excitable cells are more likely to acquire
854 place firing fields as the threshold depolarization required to trigger an action potential is
855 reduced, enabling them to respond with action potentials to a ramp depolarization that is
856 insufficient to trigger output from a silent cell. When activated the excitable cells tend to fire
857 spike bursts, whereas the silent cells do not. (b) Hypothesized model for roles of excitability
858 differences in linking of memories^{27,33}. The excitability of neurons storing information about a
859 recent event is selectively increased. Because spikes required for activity-dependent
860 associative plasticity are more likely to occur in excitable neurons, subsequent events occurring
861 within a period determined by the duration of the increase in excitability are captured to the
862 same neurons. Events occurring after excitability of the previously activated neurons has
863 decayed to baseline are stored by different neurons. (c) Hypothesized model for roles of
864 excitability differences in establishing a temporal context for spatial codes. In this scheme the
865 identity of excitable CA1 pyramidal cells evolves on a time scale of days. The probability of a
866 neuron firing action potentials within its place field is greatest during the periods in which it is
867 most excitable. In this way, the set of a neurons representing a location on a particular day can

868 be used to generate a timestamp for when an event takes place³¹.

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871 **Figure 3.** Candidate models of nonlinear integration during firing field crossings in
872 hyperpolarized or depolarized neurons. (a) Schematised synaptic integration in a model that
873 computes the arithmetic (“linear”) sum of its inputs. Synaptic inputs are stronger inside (IN) than
874 outside (OUT) of the field. While this model produces a firing field when the cell is depolarized
875 (right), it predicts a subthreshold membrane potential field in a hyperpolarized neuron (left),
876 contradicting experimental data⁵⁴. (b) Same scheme as in (a) for a model neuron that integrates
877 inputs nonlinearly, and strongly attenuates EPSPs as they propagate along the dendritic tree.
878 The strong attenuation produces subthreshold membrane potential fields that are
879 indistinguishable inside and outside of the field, while the nonlinear mechanism boosts EPSPs
880 sufficiently to produce a firing field when the cell is depolarized, consistent with experimental
881 recordings⁵⁴. (c) Same scheme as in (a) for a model neuron with distinct nonlinear integration
882 functions for inputs arriving inside or outside of the field. Although synaptic weights are the
883 same inside and outside the field, EPSPs inside the field are boosted when the neuron is
884 depolarized because of a lower threshold for engaging nonlinear mechanisms. In
885 hyperpolarized neurons, synaptic inputs inside and outside the field produce similar somatic
886 depolarization, resulting in a lack of a distinct membrane potential field, consistent with
887 experimental data⁵⁴.

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890 **Figure 4.** Comparison of the effect of local inhibition on visual and spatial receptive fields. (a)
891 Left panel, experimentally recorded IPSCs (blue) and EPSCs (red) during presentation of six
892 different grating orientations. Inhibition (blue curve) is less tuned to orientation than excitation
893 (red curve), leading to an increase in the excitation-inhibition ratio in the center of the receptive
894 field. Middle and right panels, membrane potential (middle) and firing rate (right) tuning to
895 grating orientation in a model of a L2/3 visual cortex pyramidal cell. Model responses are
896 derived from experimental recordings of IPSCs and EPSCs shown in the left panel. Black:
897 control; green: suppression of PV+ interneurons by light-activating archeorhodopsin. The model
898 reproduces an experimentally observed linear-threshold transformation of firing rate tuning
899 curves by inhibition from PV+ interneurons. Adapted from ⁶⁰. (b) Left panel, ratio of excitatory
900 and inhibitory currents (E-I ratio) during a firing field crossing in a place cell model constrained
901 by experimental data. Spatially uniform inhibition leads to an increase in the E-I ratio during the
902 field crossing. Middle and right panels, experimentally determined spatial tuning of the
903 membrane potential (middle) and firing rate (right) to spatial position in place cells. Data are
904 aligned to the place field center. Black: control; orange: suppression of GAD2+ or VGAT+
905 interneurons by light-activating archeorhodopsin. Adopted from ⁶². Note the similarity of the
906 effect of suppressing local inhibition on visual (a) and spatial (b) tuning curves.

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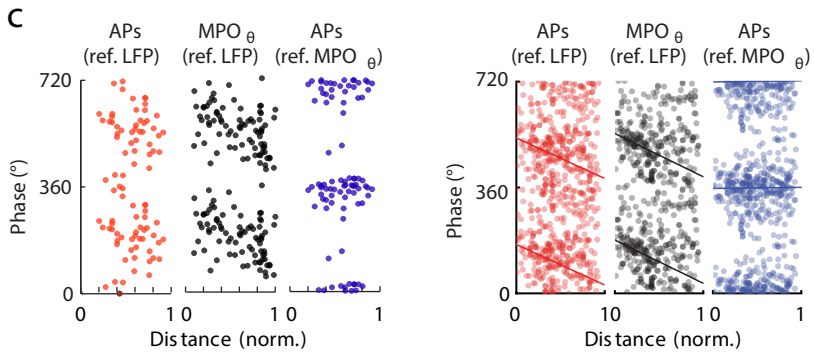
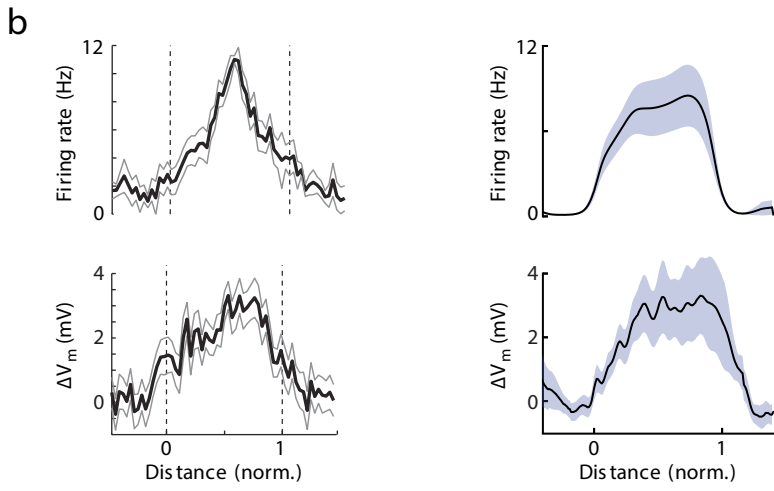
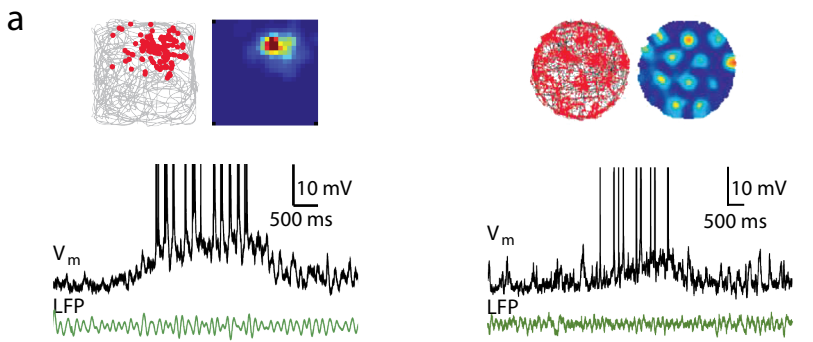
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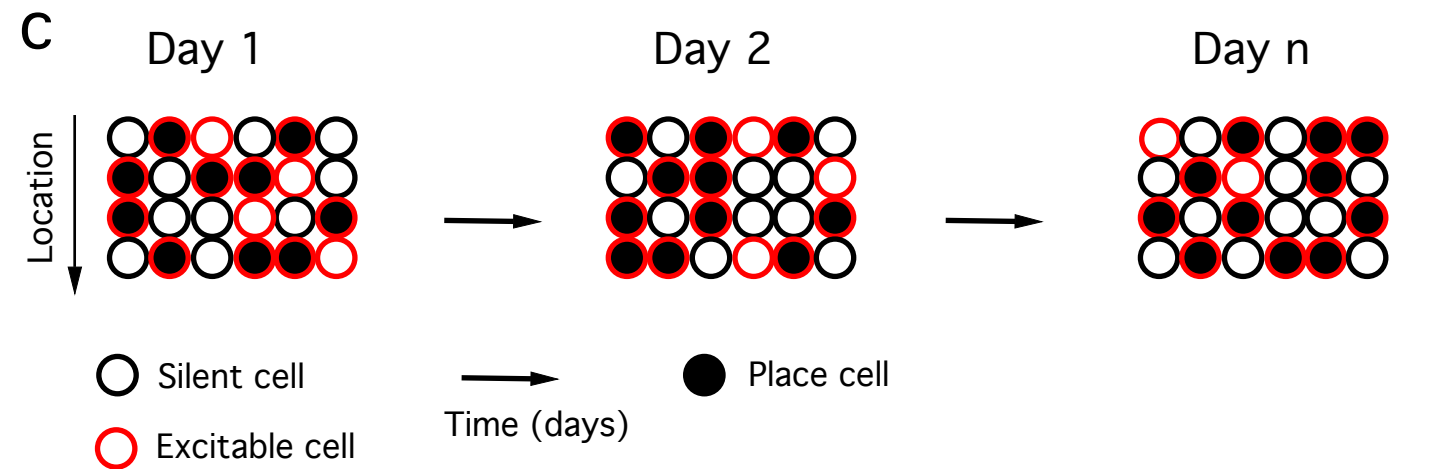
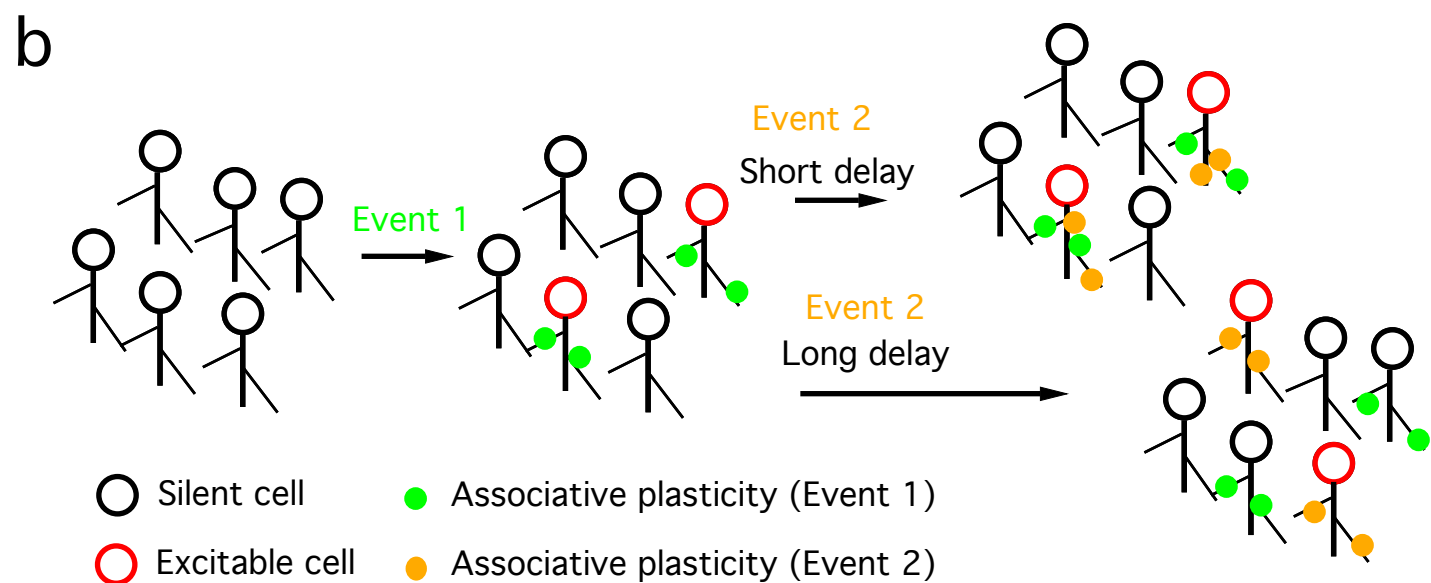
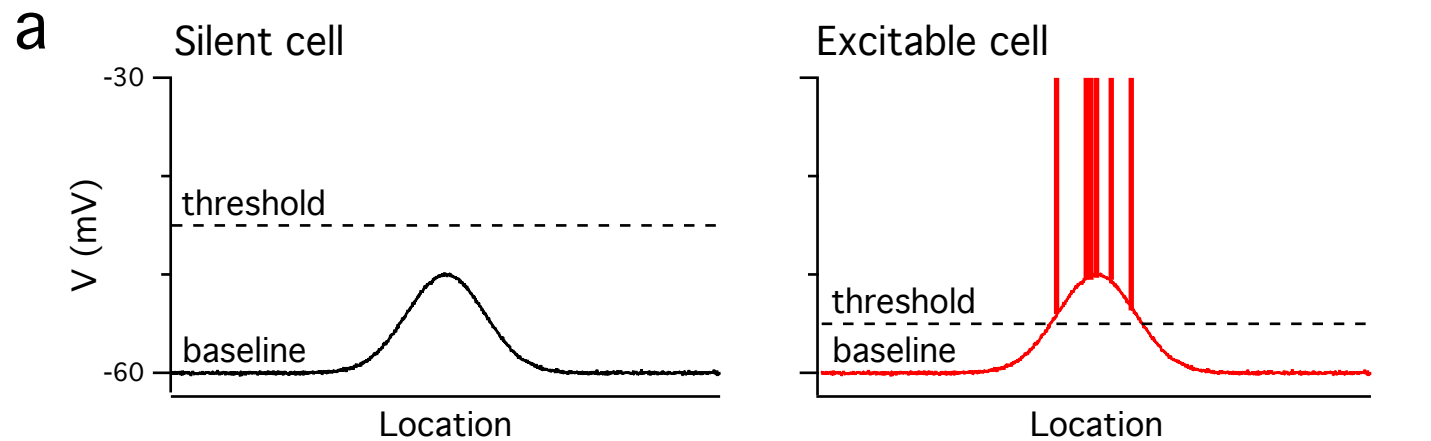
909 **Figure 5.** Theta resonant responses of hippocampal neurons. (a) When recorded in brain slices
910 CA1 pyramidal neurons preferentially respond to inputs oscillating at theta frequencies, whereas
911 fast spiking interneurons prefer higher frequency oscillatory inputs (left panels)¹¹². (b) To

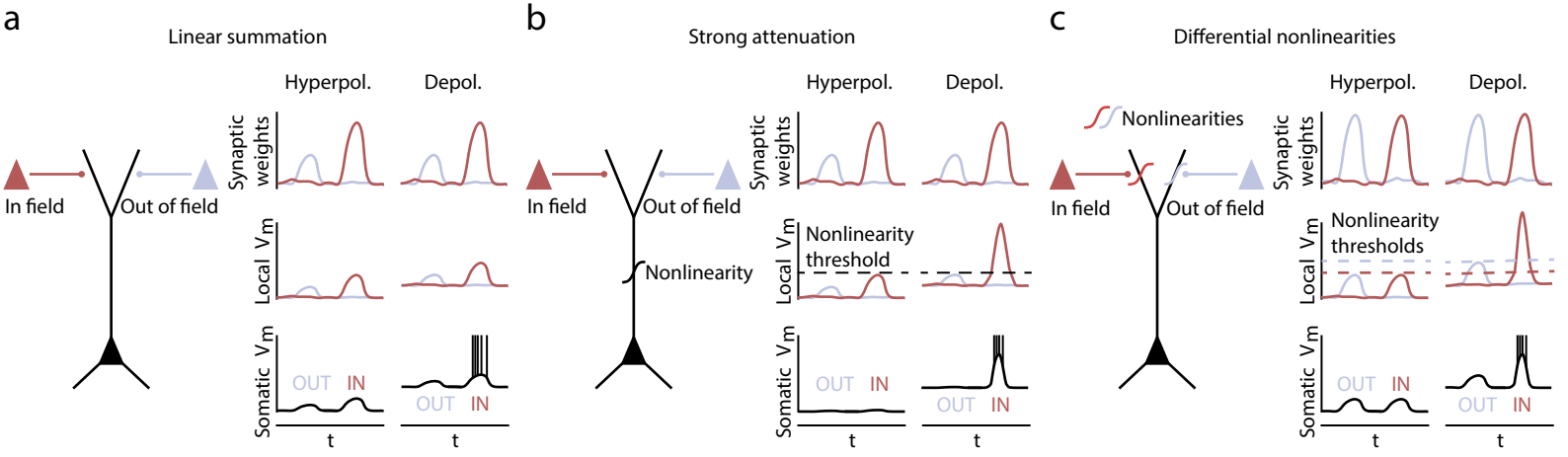
912 investigate resonant firing in behaving animals parvalbumin interneurons expressing
913 channelrhodopsin were stimulated at frequencies up to 30 Hz¹²⁰. The coherence between the
914 stimulus and the spiking response is plotted as a function of frequency for CA1 pyramidal cells
915 (upper right) and interneurons (lower right). Consistent with in vitro data CA1 pyramidal cells
916 prefer inputs in the theta frequency band.

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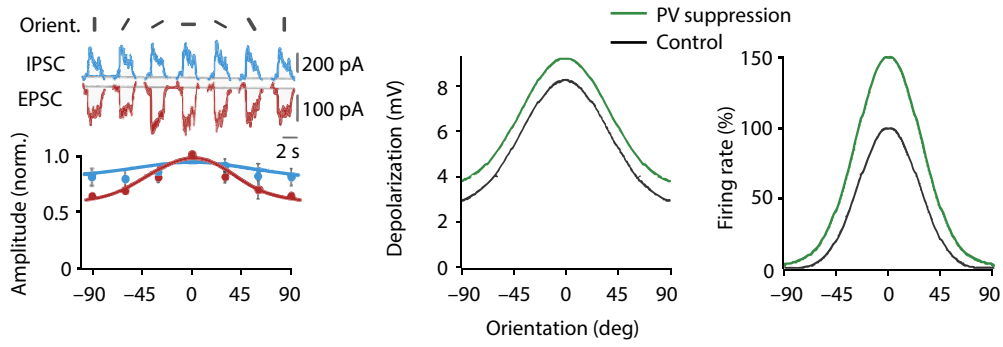
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a Orientation tuning in visual cortex
(Atallah et al., 2012)



b Spatial tuning in hippocampus
(Grienberger et al., 2017)

