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# Ovarian tissue cryopreservation for fertility preservation: clinical and research perspectives

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**BACKGROUND:** Small case series have reported successful live births after ovarian tissue cryopreservation and orthotopic transplantation, demonstrating that it can be of value in increasing the chance of successful pregnancy after treatment for cancer and other fertility-impacting diseases in adult women.

**OBJECTIVE AND RATIONALE:** This review is intended to set out the current clinical issues in the field of ovarian tissue cryopreservation, and elucidate the status of laboratory studies to address these.

**SEARCH METHODS:** We reviewed the English-language literature on ovarian tissue cryopreservation and in vitro maturation (IVM) of ovarian follicles.

**OUTCOMES:** Ovarian tissue cryopreservation is increasingly used for fertility preservation and, whilst areas for development remain (optimal patient selection, minimizing risk of contamination by malignant cells and IVM protocols), there are emerging data as to its efficacy. We review the current status of ovarian tissue cryopreservation in girls and young women facing loss of fertility from treatment of cancer and other serious diseases. Increasingly large cohort studies are reporting on success rates from ovarian tissue cryopreservation giving an indication of likely success rates. Patient selection is necessary to ensure the safety and effectiveness of this approach, especially in the very experimental situation of its application to prepubertal girls. There are continuing developments in supporting follicle development *in vitro*.

**LIMITATIONS, REASONS FOR CAUTION:** The evidence base consists largely of case series and cohort studies, thus there is the possibility of bias in key outcomes. *In vitro* development of human ovarian follicles remains some way from clinical application.

**WIDER IMPLICATIONS OF THE FINDINGS:** Ovarian tissue cryopreservation is becoming established as a valuable approach to the preservation of fertility in women. Its application in prepubertal girls may be of particular value, as it offers the only approach in this patient group. For both girls and young women, more accurate data are needed on the likelihood of successful childbirth after this procedure and the factors that underpin successful application of this approach, which will lead to its more effective use.

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**Key words:** cryopreservation / gonadotoxicity / fertility preservation / tissue culture / ivm: artificial ovary

## WHAT DOES THIS MEAN FOR PATIENTS?

Freezing part of an ovary before treatment for cancer and other diseases that might lower fertility can provide women with a better chance of having a baby once they have recovered. Approximately 100 successful live births have occurred after replacing this ovarian tissue.

The procedure of freezing ovarian tissue (cryopreservation) and later transplanting it back to the body raises a number of important clinical questions. This review identified what these main issues are and what studies have been, and still need to be, carried out.

Increasing the use of ovarian cryopreservation is providing results on the likely successful birth rates but more information is required. It may be more suitable for some women than others and careful patient selection will be needed, especially to get a more accurate prediction of the risk of loss of fertility to the individual patient. Researchers are also working on better ways to support growth of follicles (where immature eggs, or oocytes, are found) in the laboratory, for future use in the IVF clinic.

Studies show that freezing of ovarian tissue for later transplantation is a major advance for fertility preservation. However, it is not suitable for all patients as there may be concerns about transplanting cancer cells into disease-free patients. The key areas for further study are optimal patient selection, minimizing risk of contamination by cancer cells and improving laboratory protocols for oocyte maturation.

## Introduction

Advances in treatment of many cancers with consequent dramatic improvements in long-term survival (Miller et al., 2016) have led to increasing awareness of the issues in what is sometimes termed 'survivorship' to encompass the wide range of medical and social issues faced by cancer survivors. Amongst medical issues, the potential effects on fertility are an important concern of young women facing therapy for cancer and other conditions treated with similar chemotherapeutic regimens (Peate et al., 2011), and the field of fertility preservation has grown rapidly over the last two decades (Ataman et al., 2016). The most common cancers in girls and young women are those of the breast, cervix, central nervous system, leukaemias and lymphomas. Treatment of all these conditions may compromise fertility in girls and young women as a result of the surgery, chemotherapy and radiotherapy required for treatment.

Ovarian tissue cryopreservation is one of several options available for female fertility preservation, together with oocyte and embryo cryopreservation (Loren et al., 2013; Anderson et al., 2015) (Fig. 1). Other approaches include administration of agents that might protect the ovary, such as GnRH analogues (Lambertini et al., 2015), and surgical transposition of the ovary to reduce the radiation dose (Gubbala et al., 2014). This review will focus on ovarian tissue cryopreservation and address some of the outstanding issues involved in the development of this approach, including issues surrounding patient selection, the evidence for its efficacy and the current status of attempts to circumvent one of the significant issues with this technique, i.e. the potential presence of malignant cells contaminating the cryopreserved tissue.

The potential value of cryopreservation of ovarian tissue was first demonstrated in a series of experiments in the sheep where autografts of cryopreserved ovarian cortex were sutured on to the ovarian pedicle, with subsequent restoration of ovarian cyclicity and natural conception (Baird et al., 1999). These key experiments demonstrated the potential of this approach for clinical application but the detailed histological analysis that accompanied them also showed one of the major issues, which is that the majority of primordial follicles in the cryopreserved tissues do not survive. This is thought to be due to ischaemia prior to revascularization of the graft rather than direct cryo-damage and there is, as yet, no clear way of markedly improving this. The advent of vitrification, which has transformed the cryopreservation of

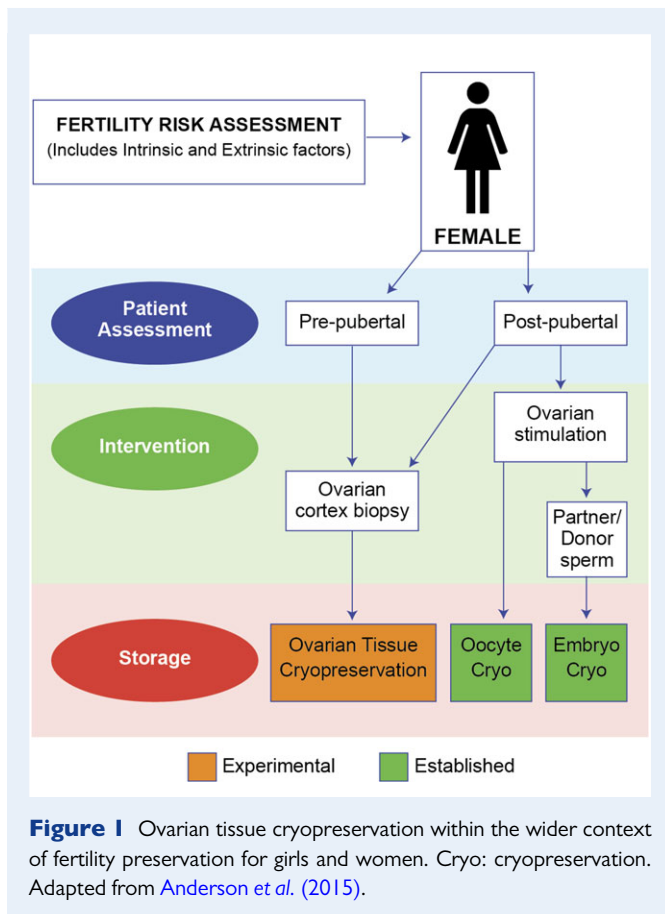
mature oocytes, may offer some advantages in reducing the cryo-damage that does occur (Ting et al., 2011; Amorim et al., 2012; Zhou et al., 2016), and indeed has been successfully used clinically in this context, but it does not address the problem of revascularization. One approach to that problem is the cryopreservation of the whole ovary with subsequent vascular re-anastomosis, and whilst there have been developments towards successful use of this approach in animal models (Campbell et al., 2014), it remains some way from clinical practice.

## Search Methods

We reviewed the English-language literature on ovarian tissue cryopreservation and IVM of ovarian follicles using conventional search methods. This was supplemented with searching cited publications in identified reports, and personal contacts with several key researchers.

## When and for whom is ovarian tissue cryopreservation appropriate?

This question raises much broader issues than strictly medical ones, and includes assessment of risk of loss of fertility, patient autonomy, decision-making at a time of stress and potential information overload, and cost (either to the patient or the health service). This can, in part, be perceived as the need to strike a balance between offering fertility preservation to all at risk, or only to those with a clear need, i.e. at high risk. Part of the evidence base relevant to this is analysis of which cancer treatments clearly do or do not impact on female fertility. The most detailed information on this is derived from the US Childhood Cancer Survivor Study (CCSS) which has for many years published analyses of reproductive outcomes in a cohort of cancer survivors using their siblings as the control group (Chemaitilly et al., 2006; Signorello et al., 2006; Green et al., 2009). A recent report from this study analysed chemotherapy exposures in relation to the prevalence of pregnancy and live birth in women who had been exposed to chemotherapy: females who had received cranial or pelvic radiotherapy were excluded from this analysis (Chow et al., 2016). The overall hazard ratio for pregnancy was 0.87 (95% CI: 0.81–0.94) compared with the control group of their siblings, demonstrating that most women who have had chemotherapy for cancer treatment in



childhood will be able to have children. Detailed analysis of the particular agents involved identified that alkylating agents were only associated with a decrease in the risk of pregnancy and live birth at the highest doses. A similar result was reported from a recent analysis of survivors of Hodgkin lymphoma in childhood and adolescence from Germany, in which the overall chance of parenthood of Hodgkin lymphoma survivors was very similar to that of the German population (Bramswig *et al.*, 2015). They showed that procarbazine in cumulative doses up to 11 400 mg/m<sup>2</sup> and cyclophosphamide in cumulative doses up to 6000 mg/m<sup>2</sup> had no significant or only minor effects on parenthood. The chance of parenthood was however significantly reduced in survivors receiving pelvic radiation. These studies also highlight that it is the treatment rather than the diagnosis that determines the risk, and also that the majority of young cancer survivors will go on to have the opportunity to have a family.

This information underpins the other aspect of the argument, whether this approach should be primarily aimed at those with a clear need, i.e. where a significant risk of loss of fertility is predicted based on the planned treatment. Ovarian tissue cryopreservation for restoration of fertility remains widely regarded as an experimental procedure (Loren *et al.*, 2013), although increasing experience with it has led to it becoming established practise in Israel (Meirow *et al.*, 2016). In our view, patients having ovarian tissue cryopreservation should be counselled and have their data collected in a research context, to improve the evidence base underpinning this approach. This is particularly important for children and younger adolescents where proxy consent from parents and carers is often the norm (Wallace *et al.*, 2016). It

remains important to avoid unnecessary intervention or procedures of uncertain benefit in unwell children at a time of extreme stress to the patient and their parents and carers. To provide a structure for discussions surrounding whether or not to proceed with ovarian tissue cryopreservation, we have developed a risk assessment tool, dividing the key issues into those intrinsic to the patient, and those extrinsic (Table 1)(Wallace *et al.*, 2012).

The intrinsic factors include whether the patient is well enough to have the necessary laparoscopic surgery to remove ovarian tissue. This should include assessment of coagulation disorders, infection risk, issues regarding the risk of anaesthesia and the potential risk of contamination of ovarian tissue by malignant disease (the latter is discussed more fully below). Issues regarding informed consent have been mentioned above. Whilst there is no theoretical lower age limit for the potential value of ovarian tissue cryopreservation, the decline in the ovarian reserve (Wallace and Kelsey, 2010), and in natural fertility in the later 30s and the consequent chances of success of the procedure in terms of restoring fertility should be clearly discussed. Assessment of the ovarian reserve using biomarkers such as anti-Müllerian hormone (AMH) or antral follicle count are routinely performed prior to assisted reproduction to predict the patient's response (Iliodromiti *et al.*, 2014). Many studies have shown a reduction in these biomarkers after cancer treatments indicating their potential to quantify ovarian toxicity and distinguish between the effects of different regimens (Bath *et al.*, 2003; Lutchman Singh *et al.*, 2007; Lie Fong *et al.*, 2008; Decanter *et al.*, 2010). Pretreatment levels of AMH have also been shown to predict the risk of amenorrhoea in women subsequently treated with chemotherapy for breast cancer (Anderson and Cameron, 2011; Su *et al.*, 2014), substantiating the hypothesis that women with a higher ovarian reserve pretreatment are more likely to have ongoing ovarian function thereafter. Such studies have not been performed with fertility as an outcome but this approach does seem to have some promise to individualize more accurately the risk of specific therapies. It would seem likely that these biomarkers could be used to give some prediction of remaining postchemotherapy ovarian function and reproductive lifespan. Similar analyses have been performed in healthy women, but accurate prediction of remaining reproductive lifespan seems only poorly predicted by, for example, AMH, except at particularly low levels in younger women (van Disseldorp *et al.*, 2008; Tehrani *et al.*, 2013). Even then, sporadic ovulation in young women will allow some to conceive. The limited value of these markers for prediction reflects our poor understanding of ovarian compensation after such damage, and this is an area where research could have a marked impact on clinical practice.

The key extrinsic factor is the nature of the predicted treatment and, as mentioned above, the key risk factors are the administration of whole body, pelvic (or abdominopelvic in children) radiotherapy and high-dose alkylating agents. The patient's age is probably the most important determinant of how damaging these therapies are, as reflected in the dramatic increase in risk of amenorrhoea after chemotherapy in women treated for breast cancer with older age at treatment (Petrek *et al.*, 2006). However, a significant problem is that even therapies that have low risk, such as the commonly administered ABVD regimen for early stage Hodgkin lymphoma, may have some impact on ovarian function (potentially in those with lower pretreatment ovarian reserve) as women treated with this regimen have recently been shown to have a reduced oocyte yield when oocyte collection for cryopreservation is subsequently

**Table 1** Intrinsic and extrinsic factors that should be taken into account when considering fertility preservation strategies for girls and young women undergoing treatment.

Intrinsic factors	
Health status of the patient	
Psycho-social factors	
Consent (patient/parent)	
Assessment of pubertal status	
Assessment of ovarian reserve	
Extrinsic factors	
Nature of predicted treatment (high/medium/low/uncertain risk)	
Time available	
Expertise/technical options available	

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performed following relapse of the disease ([Sonigo et al., 2016](#)), although no effect on risk of early menopause (before the age of 40 years) has been reported ([Swerdlow et al., 2014](#)). This also highlights that whereas initial therapy for the primary malignancy may carry a low risk of premature ovarian insufficiency (POI) and infertility, treatment may change depending on initial response (e.g. in lymphoma: [Barrington et al., 2016](#)) and regimens used for relapse will generally carry a high risk. Ovarian tissue cryopreservation and restoration of fertility potential is likely to be more effective if those patients had undergone fertility preservation procedures at the time of initial diagnosis. Treatment protocols to minimize the risk of recurrence in combination with reducing general, including gonadal, toxicity are continually evolving.

The potential value of careful prediction of risk of treatment to ovarian function has been supported by an analysis of ovarian function in a population of children and young adults aged up to 18 years, some of whom were offered ovarian tissue cryopreservation ([Wallace et al., 2014](#)). This showed that those who met the criteria and were therefore offered ovarian tissue cryopreservation had a substantial risk of POI during the course of the period of follow-up, of 35% over the 15 year follow-up period. This contrasted with a prevalence of only 1% (1 individual in a cohort of 141) in those not offered ovarian tissue cryopreservation giving a hazard ratio of 56.8 (95% CI: 6.2–521.6) at 10 years follow-up. The young age of the population studied precluded the use of fertility as the key outcome but this clearly shows the potential for focussed patient selection. In contrast, in a relatively unselected population of women who underwent pre-chemotherapy unilateral oophorectomy for fertility preservation, follow-up at a mean of 58 months showed a prevalence of POI of 9% in women with breast cancer, and 18% in those with lymphoma, but 87% of those with leukaemia, reflecting the widespread use of bone marrow transplantation in that group ([Schmidt et al., 2013](#)). Overall, 41 of the 57 women in the group who had tried to conceive had been successful, i.e. a retained fertility rate of 72%. Whilst the issues surrounding the provision of all aspects of fertility preservation require a much more complete evidence base than is currently available, it seems clear that one aspect that should be addressed is the need for more accurate identification of those who are very likely to retain their fertility after their cancer

treatment, which will guide both patients and medical services in the provision of this treatment option.

## How effective is ovarian tissue cryopreservation?

The first successful pregnancy after replacement of cryopreserved ovarian tissue was reported in 2004 ([Donnez et al., 2004](#)) and at present there are ~100 babies born following this procedure, although this number is increasing all the time. An analysis of 60 tissue replacements carried out in Belgium, Denmark and Spain demonstrated that over 90% of women showed some evidence of ovarian activity, first seen a median of 4 months after transplantation ([Donnez et al., 2013](#)). Eighteen per cent of these women achieved a pregnancy, the majority after natural conception, and 12 live births were recorded from 6 women. An updated analysis including additional centres (total 111 women) indicated a 29% pregnancy rate. An analysis of 41 transplantations in Denmark showed that 31% of those wishing to conceive had a successful pregnancy ([Jensen et al., 2015](#)): 100 replacements have now been performed there (C.Y. Andersen, personal report). A recent report has presented data on 95 orthotopic cryopreserved ovarian tissue transplantations in 74 women in the FertiProtekt Network ([Van der Ven et al., 2016](#)). These women had an average age of 30 years at cryopreservation and 34 years at transplantation, with the two most common diagnoses being breast cancer and Hodgkin lymphoma. Amongst those undergoing their first transplantation who had clear evidence of POI at the time of transplantation, 62.5% showed evidence of ovarian activity 1 year after transplantation, and 27.5% achieved a pregnancy with delivery in 22.5%. These case series provide, at present, the best estimate of the likely success of this approach, although no doubt additional pregnancies will be achieved, increasing the overall success rate. Indeed, there are case reports of individual women having up to three successful pregnancies after ovarian tissue replacement, with graft function lasting up to 10 years ([Jensen et al., 2015](#)). Calculation of success rate is however complicated by a range of issues including that some transplantations are not being performed to restore fertility but for endocrine reasons, and that some women have some residual ovarian activity at the time of transplantation ([Andersen, 2015](#)).

The majority of these pregnancies were after natural conception, and the potential for this is one of the key positive aspects of ovarian tissue cryopreservation compared with oocyte or embryo cryopreservation. A more aggressive approach with repeated cycles of ovarian stimulation to either achieve pregnancy with fresh embryo replacement ([Meirow et al., 2016](#)) or, indeed, to bank cryopreserved embryos whilst the ovarian graft remains functional ([Oktay et al., 2016](#)) has also been reported. In one series of 20 women, some of whom had previously received chemotherapy for leukaemia or lymphoma, 32% had a baby after up to 11 cycles of IVF ([Meirow et al., 2016](#)). However, in a total of 56 IVF cycles, a total of only 84 oocytes were obtained, but a fertilization rate of 58% was achieved with embryo transfer in 68% of cycles.

In addition to the potential for natural conception, endogenous hormone production by the graft is also a key benefit of this approach. The great majority of ovarian replacements showed evidence of hormonal activity, although for a variable duration, occasionally up to several years. It is unclear at present what factors predict the duration of the graft, and how much can be related to surgical technique at

replacement compared with the intrinsic functional potential of the tissue. Related to this, replacement has also been proposed as a method for inducing puberty (Poirot *et al.*, 2012; Ernst *et al.*, 2013). This, however, is controversial for a number of reasons (Anderson *et al.*, 2013). These include that it results in rapid escalation of oestrogen levels to adult concentrations, in contradiction to the established principles of pubertal induction with slow progression and escalation of oestrogen dose. Additionally, the early onset of ovulatory cycles will expose the patient to progesterone earlier than would occur during normal puberty, which may impact on normal breast development. The tissue also contains a finite number of oocytes, thus this could be argued to be a wasteful use of this scarce resource. There is also the potential risk of malignant contamination, discussed more fully below. However, there is also clear value to women from endogenous and physiological hormone production. The adverse health effects of POI are increasingly recognized and whilst it is very likely that many of these are reduced by appropriate exogenous hormone replacement, there is a paucity of robust evidence regarding the risks and benefits of HRT in women with POI (Langrish *et al.*, 2009; Crofton *et al.*, 2010), with much of it being extrapolated from women taking HRT after a physiological menopause (ESHRE Guideline Group on POI *et al.*, 2016). Replacement of cryopreserved ovarian tissue purely for hormone production rather than fertility has been proposed (Andersen and Kristensen, 2015). This may well be appropriate for some women who have completed their family or no longer wish to pursue fertility, taking into account the risk of malignant contamination, the uncertain duration of function of replaced ovarian tissue, and issues surrounding risks and benefits of hormone 'replacement' beyond the age of natural menopause.

## The risk of malignant contamination

The storage of ovarian tissue before cancer treatment is started carries with it the risk that the tissue will contain malignant cells, which will survive cryopreservation and could potentially be transplanted back into the patient with the risk of recurrence of the malignancy. A range of methods specific to the diagnosis can be used to detect malignant contamination, including immunohistochemistry and molecular analysis, and a sample of tissue should be sent for such analysis at the time it is stored. These tests are all, however, destructive, and therefore cannot be applied to the actual pieces of tissue that will be replaced in the patient. In general, it is considered that where there is no evidence of metastatic disease of solid cancers, then the risk of ovarian contamination is low. However, for leukaemias before treatment has started there is a high risk that circulating malignant cells will be present in the ovary, and this has been confirmed in a number of studies specifically investigating this (Greve *et al.*, 2012; Dolmans *et al.*, 2013; Rosendahl *et al.*, 2013). This also confirms the potential of these tests to identify malignant contamination although there may be issues of sensitivity to pick up minimal residual disease, which will be expected to improve with time. A high level of vigilance is required to detect contamination even when the diagnosis and staging tests indicate a low risk. In our own series of patients, we have identified a case of ovarian contamination with Ewing's sarcoma that had arisen in a thoracic (T7) vertebra and conventional staging showed no evidence of metastatic disease (Fig. 2). The identification of this contamination will, of course, preclude

replacement of the tissue and alternative approaches to the restoration of fertility in such patients are required. These are discussed below. The application of ovarian tissue cryopreservation in patients with leukaemia remains controversial because of this risk of contamination. It has been proposed that ovarian tissue can be taken following initial courses of chemotherapy, potentially sufficient to reduce the risk of cryopreservation of circulating malignant cells but without excessively compromising ovarian function (Greve *et al.*, 2012). Whilst successful cases have been reported (Meirow *et al.*, 2016), the benefits and risks of this approach have yet to be clearly quantified and undertaking ovarian tissue cryopreservation in these patients remains contentious.

## Developing methodologies to optimize the potential of cryopreserved tissue

Whilst reimplantation of ovarian tissue has been a major advance for fertility preservation, it is not suitable for all patients as there may be concerns about transplanting malignant cells into disease-free patients. With these concerns in mind, a great deal of research has focused on first, isolating follicles from tissue at specific stages of development either before or after cryopreservation, removing any contaminating cells and recombining the follicles within a supportive matrix to form an 'artificial ovary' for subsequent reimplantation or *in vitro* development, and second, developing oocytes from cryopreserved tissue completely *in vitro*.

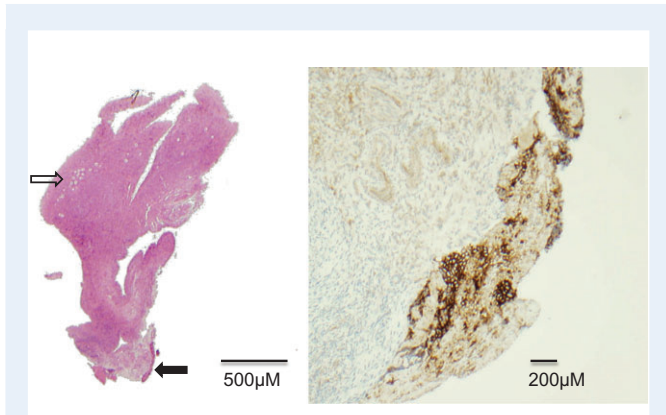
## Making an artificial ovary

The aim of developing an 'artificial ovary' is to produce a structure that would be free of any potential malignant cells and contain healthy immature follicles (and stromal cells). Groups working in this area have focused on isolation methods and in developing matrices to support isolated follicles (Skory *et al.*, 2015; Paulini *et al.*, 2016). Developing the optimal matrix to support follicle/oocyte development has been slow but some advances have been made. The optimal matrix needs to support growth, development and recovery of human ovarian follicles as well as control rigidity. Bioengineering has led to the development of many materials to support follicle development (Shea *et al.*, 2014). Alginate has given positive results with mouse and human follicles (Yin *et al.*, 2016) and fibrin has also been demonstrated to be a viable material to support isolated murine primordial follicles after transplantation (Kniazeva *et al.*, 2015). Decellularized ovary has been used to support the growth and development of mouse follicles (Laronda *et al.*, 2015). Recent work with isolated preantral human follicles has demonstrated that a matrix formed by combining fibrin with fibrinogen and thrombin improves the health and development of the encapsulated follicles (Paulini *et al.*, 2016). Work in this area is promising but there is still much to be tested before an 'artificial' ovary could reach the clinic.

The avoidance of a transplantation procedure would be preferred for some patients. To obtain fully grown competent oocytes from cryopreserved tissue without a transplantation step would require the immature oocytes to be grown completely *ex vivo*.

## Oocyte growth and development *in vitro* using cryopreserved tissue

Growth *in vitro* from the most immature oocytes (primordial stages) with subsequent IVF and production of live young has only been achieved in mouse (Eppig and O'Brien, 1996; O'Brien et al., 2003). The complete process of murine oogenesis from the primordial germ cell stage to the production of developmentally competent oocytes can be recapitulated *in vitro* (Morohaku et al., 2016) and the resultant offspring have been shown to be healthy. Whilst there are major differences between human and mouse oocyte development, the mouse work provides proof of concept that complete oocyte development can be



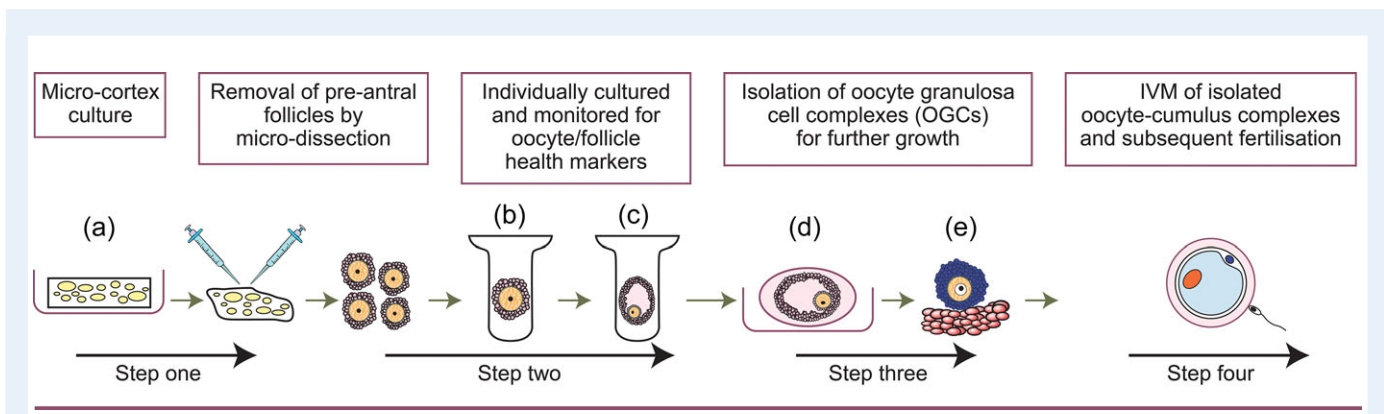
**Figure 2** Biopsy of ovary from a girl with Ewing's sarcoma in a vertebra. A 12-year-old with Ewing's sarcoma localized to the T7 vertebra and no evidence of metastatic disease underwent ovarian biopsy for fertility preservation. A biopsy sent for pathological examination showed presumed malignant contamination, with CD99 positive cells present consistent with metastatic Ewing's sarcoma. Left panel, solid arrow indicates area of CD99 positive cells in a haematoxylin and eosin stained section, open arrow indicates primordial follicles in cortex of ovary. Right panel: CD99 (brown stain) immunohistochemistry result.

achieved *in vitro* and has driven the development of culture systems that could be applied to other species, particularly human. Advances in culturing follicles from humans, non-human primates and domestic species have been made, bringing the prospect of achieving an *in vitro* system that supports complete human oocyte development closer (Smits et al., 2010; Telfer and Zelinski, 2013; Xiao et al., 2015).

Several culture systems have been developed that have been optimized for a range of different animal models (Smits et al., 2010; Telfer and McLaughlin, 2012; Telfer and Zelinski, 2013). Whilst some are more advanced than others, there is not a single system that has been fully optimized to support the complete *in vitro* growth and maturation of human follicles and oocytes. All of the published systems have strengths and weaknesses that may be usefully exploited to meet the challenges of human follicle culture but there is agreement that to achieve complete *in vitro* growth, a dynamic culture system is required to support the three main transition steps in oocyte/follicle development, i.e. growth activation, ongoing culture of isolated follicles and final oocyte maturation.

## *In vitro* activation of primordial follicles

The regulation of follicle activation involves a combination of inhibitory, stimulatory and maintenance factors (Nelson et al., 2013; Hsueh et al., 2015). Work using knockout mouse models has highlighted the importance of the phosphatidylinositol-3'-kinase (PI3K-AKT) signalling pathway within the oocyte in regulating follicle activation (Reddy et al., 2008). Using pharmacological inhibitors of phosphatase and tensin homologue *in vitro* leads to increased activation of human primordial follicles (Li et al., 2010; McLaughlin et al., 2014), whereas treatment with rapamycin (an inhibitor of mammalian target of rapamycin complex 1: mTORC) results in decreased activation (McLaughlin et al., 2011). A human live birth has been reported following the treatment of ovarian tissue for 48 h *in vitro* with bpV(HOpic) and 740YP, an Akt stimulant, followed by replacement and IVF (Kawamura et al., 2013). This is an extremely encouraging development but needs to be treated with caution as similar inhibitors *in vitro*, whilst increasing activation of



**Figure 3** Multistep culture system for activation of human primordial follicles and subsequent follicle/oocyte development. Primordial follicles can be activated or held dormant within ovarian cortical tissue (a). Once activated preantral stages can be isolated (b), they are grown individually to antral stages (c). These can be further grown, for example, within an alginate bead (d) to obtain fully grown oocytes (e), then competence to undergo maturation and fertilization is tested.

primordial follicles, result in poor quality oocytes at later stages (McLaughlin *et al.*, 2014).

## Isolation and culture of growing follicles to achieve oocyte growth and development

Once follicles have reached the multi-laminar preantral stage of development, there are several culture systems that support further growth. Placing preantral follicles in v-shaped micro-well plates allows maintenance of three-dimensional follicular architecture *in vitro* whilst promoting growth and differentiation of human follicles (Telfer *et al.*, 2008; Anderson *et al.*, 2014; McLaughlin *et al.*, 2014) with antral formation occurring within 10 days. Follicle encapsulation in alginate hydrogels has also been used to support secondary human (Xu *et al.*, 2009; Xiao *et al.*, 2015) and rhesus monkey (Ting *et al.*, 2011) follicle growth *in vitro*. Alginate encapsulation is believed to mimic the extracellular matrix *in vivo* in terms of its ability to facilitate molecular exchange between the follicle and the culture medium, and whilst its flexibility can accommodate cell proliferation, its rigidity prevents dissociation of the follicular unit. The progression of human follicles following isolation from the cortex is remarkable. In the presence of FSH, enzymatically isolated secondary human follicles can differentiate, become steroidogenically active and complete oocyte growth in 30 days (Xu *et al.*, 2009) and have been shown to be capable of meiotic maturation (Xiao *et al.*, 2015).

## Aspiration and maturation of oocyte cumulus complexes

The final goal of an *in vitro* system is to produce oocytes that can be fertilized and produce developmentally normal embryos. In order to achieve this, *in vitro* grown human oocytes need to be meiotically matured *in vitro* and several groups have made important steps towards this for clinical use over the last two decades (Alak *et al.*, 1998; Cha and Chian, 1998; Mikkelsen *et al.*, 1999; Cavilla *et al.*, 2008; Chian *et al.*, 2013). It is widely accepted that whilst 40–80% of immature human oocytes can successfully complete IVM and fertilization, the rate of maturation of immature oocytes is still well below that of oocytes harvested from stimulated ovaries, indicating that the protocols are suboptimal or many of the harvested oocytes are intrinsically unable to undergo maturation (Chang *et al.*, 2014).

The challenge in developing a complete *in vitro* growth system is to combine each of these stages to produce a multistep system that supports the complete process of oocyte development (Fig. 3).

## Conclusion

Preservation of fertility in girls and young women with cancer who are at risk of loss of fertility remains an important priority to improve their long-term health and well-being. Oocyte and embryo cryopreservation are established but in most countries in the world, ovarian tissue cryopreservation remains experimental, particularly in younger adolescents and prepubertal girls. Cryopreservation of ovarian cortical tissue with later replacement has now resulted in at least 100 live births but requires an

additional invasive procedure at a time of emotional distress and uncertainty, and there is an uncertain risk of tissue contamination in haematological and other malignancies. *In vitro* growth and maturation of immature ovarian follicles is developing but further research is required before it can be used for clinical benefit. Girls and young women having ovarian tissue cryopreservation should be counselled and have their data collected in a research context, to improve the evidence base underpinning this approach. This field requires more basic and clinical research to provide accurate information for patients and their medical teams and to promote the development of ovarian cryopreservation as an effective and cost-efficient treatment option for female fertility preservation.

## Authors' roles

R.A.A. and E.E.T. drafted the manuscript; all authors reviewed and edited it; all authors approved the final version.

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## Conflict of interest

The authors declare that there is no conflict of interest that could prejudice the impartiality of the present research.

## References

- Alak BM, Coskun S, Friedman CI, Kennard EA, Kim MH, Seifer DB. Activin A stimulates meiotic maturation of human oocytes and modulates granulosa cell steroidogenesis *in vitro*. *Fertil Steril* 1998;**70**:1126–1130.
- Amorim CA, Dolmans MM, David A, Jaeger J, Vanacker J, Camboni A, Donnez J, Van Langendonck A. Vitrification and xenografting of human ovarian tissue. *Fertil Steril* 2012;**98**:1291–1298.e1–2.
- Andersen CY. Success and challenges in fertility preservation after ovarian tissue grafting. *Lancet* 2015;**385**:1947–1948.
- Anderson RA, Cameron DA. Pretreatment serum anti-Mullerian hormone predicts long-term ovarian function and bone mass after chemotherapy for early breast cancer. *J Clin Endocrinol Metab* 2011;**96**:1336–1343.
- Anderson RA, Hindmarsh PC, Wallace WH. Induction of puberty by autograft of cryopreserved ovarian tissue in a patient previously treated for Ewing sarcoma. *Eur J Cancer* 2013;**49**:2960–2961.
- Andersen CY, Kristensen SG. Novel use of the ovarian follicular pool to postpone menopause and delay osteoporosis. *Reprod Biomed Online* 2015;**31**:128–131.
- Anderson RA, McLaughlin M, Wallace WH, Albertini DF, Telfer EE. The immature human ovary shows loss of abnormal follicles and increasing follicle developmental competence through childhood and adolescence. *Hum Reprod* 2014;**29**:97–106.
- Anderson RA, Mitchell RT, Kelsey TW, Spears N, Telfer EE, Wallace WH. Cancer treatment and gonadal function: experimental and established strategies for fertility preservation in children and young adults. *Lancet Diabetes Endocrinol* 2015;**3**:556–567.
- Ataman LM, Rodrigues JK, Marinho RM, Caetano JP, Chehin MB, Alves da Motta EL, Serafini P, Suzuki N, Furui T, Takae S *et al.* Creating a global community of practice for oncofertility. *J Glob Oncol* 2016;**2**:83–96.



- Baird DT, Webb R, Campbell BK, Harkness LM, Gosden RG. Long-term ovarian function in sheep after ovariectomy and transplantation of autografts stored at -196 C. *Endocrinology* 1999;**140**:462–471.
- Barrington SF, Kirkwood AA, Franceschetto A, Fulham MJ, Roberts TH, Almquist H, Brun E, Hjorthaug K, Viney ZN, Pike LC et al. PET-CT for staging and early response: results from the response-adapted therapy in advanced Hodgkin lymphoma study. *Blood* 2016;**127**:1531–1538.
- Bath LE, Wallace WH, Shaw MP, Fitzpatrick C, Anderson RA. Depletion of ovarian reserve in young women after treatment for cancer in childhood: detection by anti-Mullerian hormone, inhibin B and ovarian ultrasound. *Hum Reprod* 2003;**18**:2368–2374.
- Bramswig JH, Riepenhausen M, Schellong G. Parenthood in adult female survivors treated for Hodgkin's lymphoma during childhood and adolescence: a prospective, longitudinal study. *Lancet Oncol* 2015;**16**:667–675.
- Campbell BK, Hernandez-Medrano J, Onions V, Pincott-Allen C, Aljaser F, Fisher J, McNeilly AS, Webb R, Picton HM. Restoration of ovarian function and natural fertility following the cryopreservation and auto-transplantation of whole adult sheep ovaries. *Hum Reprod* 2014;**29**:1749–1763.
- Cavilla JL, Kennedy CR, Byskov AG, Hartshorne GM. Human immature oocytes grow during culture for IVF. *Hum Reprod* 2008;**23**:37–45.
- Cha KY, Chian RC. Maturation in vitro of immature human oocytes for clinical use. *Hum Reprod Update* 1998;**4**:103–120.
- Chang EM, Song HS, Lee DR, Lee WS, Yoon TK. In vitro maturation of human oocytes: its role in infertility treatment and new possibilities. *Clin Exp Reprod Med* 2014;**41**:41–46.
- Chian RC, Uzelac PS, Nargund G. In vitro maturation of human immature oocytes for fertility preservation. *Fertil Steril* 2013;**99**:1173–1181.
- Chemaitilly W, Mertens AC, Mitby P, Whitton J, Stovall M, Yasui Y, Robison LL, Sklar CA. Acute ovarian failure in the Childhood Cancer Survivor Study. *J Clin Endocrinol Metab* 2006;**91**:1723–1728.
- Chow EJ, Stratton KL, Leisenring WM, Oeffinger KC, Sklar CA, Donaldson SS, Ginsberg JP, Kenney LB, Levine JM, Robison LL et al. Pregnancy after chemotherapy in male and female survivors of childhood cancer treated between 1970 and 1999: a report from the Childhood Cancer Survivor Study cohort. *Lancet Oncol* 2016;**17**:567–576.
- Crofton PM, Evans N, Bath LE, Warner P, Whitehead TJ, Critchley HO, Kelnar CJ, Wallace WH. Physiological versus standard sex steroid replacement in young women with premature ovarian failure: effects on bone mass acquisition and turnover. *Clin Endocrinol (Oxf)* 2010;**73**:707–714.
- Decanter C, Morschhauser F, Pigny P, Lefebvre C, Gallo C, Dewailly D. Anti-Mullerian hormone follow-up in young women treated by chemotherapy for lymphoma: preliminary results. *Reprod Biomed Online* 2010;**20**:280–285.
- Dolmans MM, Luyckx V, Donnez J, Andersen CY, Greve T. Risk of transferring malignant cells with transplanted frozen-thawed ovarian tissue. *Fertil Steril* 2013;**99**:1514–1522.
- Donnez J, Dolmans MM, Demyle D, Jadoul P, Pirard C, Squifflet J, Martinez-Madrid B, Van Langendonck A. Livebirth after orthotopic transplantation of cryopreserved ovarian tissue. *Lancet* 2004;**364**:1405–1410.
- Donnez J, Dolmans MM, Pellicer A, Diaz-Garcia C, Sanchez Serrano M, Schmidt KT, Ernst E, Luyckx V, Andersen CY. Restoration of ovarian activity and pregnancy after transplantation of cryopreserved ovarian tissue: a review of 60 cases of reimplantation. *Fertil Steril* 2013;**99**:1503–1513.
- Eppig JJ, O'Brien MJ. Development in vitro of mouse oocytes from primordial follicles. *Biol Reprod* 1996;**54**:197–207.
- Ernst E, Kjaersgaard M, Birkebaek NH, Clausen N, Andersen CY. Case report: stimulation of puberty in a girl with chemo- and radiation therapy induced ovarian failure by transplantation of a small part of her frozen/thawed ovarian tissue. *Eur J Cancer* 2013;**49**:911–914.
- ESHRE Guideline Group on POI, Webber L, Davies M, Anderson R, Bartlett J, Braat D, Cartwright B, Cifkova R, de Muinck Keizer-Schrama S, Hogervorst E et al. ESHRE guideline: management of women with premature ovarian insufficiency. *Hum Reprod* 2016;**31**:926–937.
- Green DM, Kawashima T, Stovall M, Leisenring W, Sklar CA, Mertens AC, Donaldson SS, Byrne J, Robison LL. Fertility of female survivors of childhood cancer: a report from the Childhood Cancer Survivor Study. *J Clin Oncol* 2009;**27**:2677–2685.
- Greve T, Clasen-Linde E, Andersen MT, Andersen MK, Sorensen SD, Rosendahl M, Ralfkiaer E, Andersen CY. Cryopreserved ovarian cortex from patients with leukemia in complete remission contains no apparent viable malignant cells. *Blood* 2012;**120**:4311–4316.
- Gubbala K, Laios A, Gallos I, Pathiraja P, Haldar K, Ind T. Outcomes of ovarian transposition in gynaecological cancers; a systematic review and meta-analysis. *J Ovarian Res* 2014;**7**:69.
- Hsueh AJ, Kawamura K, Cheng Y, Fauser BC. Intraovarian control of early folliculogenesis. *Endocr Rev* 2015;**36**:1–24.
- Iliodromiti S, Anderson RA, Nelson SM. Technical and performance characteristics of anti-Mullerian hormone and antral follicle count as biomarkers of ovarian response. *Hum Reprod Update* 2014;**21**:698–710.
- Jensen AK, Kristensen SG, Macklon KT, Jeppesen JV, Fedder J, Ernst E, Andersen CY. Outcomes of transplantations of cryopreserved ovarian tissue to 41 women in Denmark. *Hum Reprod* 2015;**30**:2838–2845.
- Kawamura K, Cheng Y, Suzuki N, Deguchi M, Sato Y, Takae S, Ho CH, Kawamura N, Tamura M, Hashimoto S et al. Hippo signaling disruption and Akt stimulation of ovarian follicles for infertility treatment. *Proc Natl Acad Sci USA* 2013;**110**:17474–17479.
- Kniazeva E, Hardy AN, Boukaidi SA, Woodruff TK, Jeruss JS, Shea LD. Primordial follicle transplantation within designer biomaterial grafts produce live births in a mouse infertility model. *Sci Rep* 2015;**5**:17709.
- Lambertini M, Ceppi M, Poggio F, Peccatori FA, Azim HA Jr, Ugolini D, Pronzato P, Loibl S, Moore HC, Partridge AH et al. Ovarian suppression using luteinizing hormone-releasing hormone agonists during chemotherapy to preserve ovarian function and fertility of breast cancer patients: a meta-analysis of randomized studies. *Ann Oncol* 2015;**26**:2408–2419.
- Langrish JP, Mills NL, Bath LE, Warner P, Webb DJ, Kelnar CJ, Critchley HO, Newby DE, Wallace WH. Cardiovascular effects of physiological and standard sex steroid replacement regimens in premature ovarian failure. *Hypertension* 2009;**53**:805–811.
- Laronda MM, Jakus AE, Whelan KA, Wertheim JA, Shah RN, Woodruff TK. Initiation of puberty in mice following decellularized ovary transplant. *Biomaterials* 2015;**50**:20–29.
- Li J, Kawamura K, Cheng Y, Liu S, Klein C, Liu S, Duan EK, Hsueh AJ. Activation of dormant ovarian follicles to generate mature eggs. *Proc Natl Acad Sci USA* 2010;**107**:10280–10284.
- Lie Fong S, Lugtenburg PJ, Schipper I, Themmen AP, de Jong FH, Sonneveld P, Laven JS. Anti-Mullerian hormone as a marker of ovarian function in women after chemotherapy and radiotherapy for haematological malignancies. *Hum Reprod* 2008;**23**:674–678.
- Loren AW, Mangu PB, Beck LN, Brennan L, Magdalinski AJ, Partridge AH, Quinn G, Wallace WH, Oktay K. Fertility preservation for patients with cancer: American Society of Clinical Oncology clinical practice guideline update. *J Clin Oncol* 2013;**31**:2500–2510.
- Lutchman Singh K, Muttukrishna S, Stein RC, McGarrigle HH, Patel A, Parikh B, Groome NP, Davies MC, Chatterjee R. Predictors of ovarian reserve in young women with breast cancer. *Br J Cancer* 2007;**96**:1808–1816.
- McLaughlin M, Kinnell HL, Anderson RA, Telfer EE. Inhibition of phosphatase and tensin homologue (PTEN) in human ovary in vitro results in increased activation of primordial follicles but compromises development of growing follicles. *Mol Hum Reprod* 2014;**20**:736–744.

- McLaughlin M, Patrizio P, Kayisli U, Luk J, Thomson TC, Anderson RA, Telfer EE, Johnson J. mTOR kinase inhibition results in oocyte loss characterized by empty follicles in human ovarian cortical strips cultured in vitro. *Fertil Steril* 2011;**96**:1154–1159.
- Meirow D, Ra'anani H, Shapira M, Brenghausen M, Derech Chaim S, Aviel-Ronen S, Amariglio N, Schiff E, Orvieto R, Dor J. Transplantations of frozen-thawed ovarian tissue demonstrate high reproductive performance and the need to revise restrictive criteria. *Fertil Steril* 2016;**106**:467–474.
- Mikkelsen AL, Smith SD, Lindenberg S. In-vitro maturation of human oocytes from regularly menstruating women may be successful without follicle stimulating hormone priming. *Hum Reprod* 1999;**14**:1847–1851.
- Miller KD, Siegel RL, Lin CC, Mariotto AB, Kramer JL, Rowland JH, Stein KD, Alteri R, Jemal A. Cancer treatment and survivorship statistics, 2016. *CA Cancer J Clin* 2016;**66**:271–289.
- Morohaku K, Tanimoto R, Sasaki K, Kawahara-Miki R, Kono T, Hayashi K, Hirao Y, Obata Y. Complete in vitro generation of fertile oocytes from mouse primordial germ cells. *Proc Natl Acad Sci USA* 2016;**113**:9021–9026.
- Nelson SM, Telfer EE, Anderson RA. The ageing ovary and uterus: new biological insights. *Hum Reprod Update* 2013;**19**:67–83.
- O'Brien MJ, Pendola JK, Eppig JJ. A revised protocol for in vitro development of mouse oocytes from primordial follicles dramatically improves their developmental competence. *Biol Reprod* 2003;**68**:1682–1686.
- Oktay K, Bedoschi G, Pacheco F, Turan V, Emirdar V. First pregnancies, live birth, and in vitro fertilization outcomes after transplantation of frozen-banked ovarian tissue with a human extracellular matrix scaffold using robot-assisted minimally invasive surgery. *Am J Obstet Gynecol* 2016;**214**:94.e1–94.e9.
- Paulini F, Vilela JM, Chiti MC, Donnez J, Jadoul P, Dolmans MM, Amorim CA. Survival and growth of human preantral follicles after cryopreservation of ovarian tissue, follicle isolation and short-term xenografting. *Reprod Biomed Online* 2016;**33**:425–432.
- Peate M, Meiser B, Friedlander M, Zorbas H, Rovelli S, Sansom-Daly U, Sangster J, Hadzi-Pavlovic D, Hickey M. It's now or never: fertility-related knowledge, decision-making preferences, and treatment intentions in young women with breast cancer—an Australian fertility decision aid collaborative group study. *J Clin Oncol* 2011;**29**:1670–1677.
- Petrek JA, Naughton MJ, Case LD, Paskett ED, Naftalis EZ, Singletary SE, Sukumvanich P. Incidence, time course, and determinants of menstrual bleeding after breast cancer treatment: a prospective study. *J Clin Oncol* 2006;**24**:1045–1051.
- Poirot C, Abirached F, Prades M, Coussieu C, Bernaudin F, Piver P. Induction of puberty by autograft of cryopreserved ovarian tissue. *Lancet* 2012;**379**:588.
- Reddy P, Liu L, Adhikari D, Jagarlamudi K, Rajareddy S, Shen Y, Du C, Tang W, Hamalainen T, Peng SL et al. Oocyte-specific deletion of PTEN causes premature activation of the primordial follicle pool. *Science* 2008;**319**:611–613.
- Rosendahl M, Greve T, Andersen CY. The safety of transplanting cryopreserved ovarian tissue in cancer patients: a review of the literature. *J Assist Reprod Genet* 2013;**30**:11–24.
- Schmidt KT, Nyboe Andersen A, Greve T, Ernst E, Loft A, Yding Andersen C. Fertility in cancer patients after cryopreservation of one ovary. *Reprod Biomed Online* 2013;**26**:272–279.
- Shea LD, Woodruff TK, Shikanov A. Bioengineering the ovarian follicle microenvironment. *Annu Rev Biomed Eng* 2014;**16**:29–52.
- Signorello LB, Cohen SS, Bosetti C, Stovall M, Kasper CE, Weathers RE, Whitton JA, Green DM, Donaldson SS, Mertens AC et al. Female survivors of childhood cancer: preterm birth and low birth weight among their children. *J Natl Cancer Inst* 2006;**98**:1453–1461.
- Skory RM, Xu Y, Shea LD, Woodruff TK. Engineering the ovarian cycle using in vitro follicle culture. *Hum Reprod* 2015;**30**:1386–1395.
- Smits J, Dolmans MM, Donnez J, Fortune JE, Hovatta O, Jewgenow K, Picton HM, Plancha C, Shea LD, Stouffer RL et al. Current achievements and future research directions in ovarian tissue culture, in vitro follicle development and transplantation: implications for fertility preservation. *Hum Reprod Update* 2010;**16**:395–414.
- Sonigo C, Seroka A, Cedrin-Dumerin I, Sermondade N, Sifer C, Grynberg M. History of ABVD alters the number of oocytes vitrified after in vitro maturation in fertility preservation candidates. *Future Oncol* 2016;**12**:1713–1719.
- Su HC, Haunschild C, Chung K, Komrokian S, Boles S, Sammel MD, DeMichele A. Prechemotherapy anti-Mullerian hormone, age, and body size predict timing of return of ovarian function in young breast cancer patients. *Cancer* 2014;**120**:3691–3698.
- Swerdlow AJ, Cooke R, Bates A, Cunningham D, Falk SJ, Gilson D, Hancock BW, Harris SJ, Horwich A, Hoskin PJ et al. Risk of premature menopause after treatment for Hodgkin's lymphoma. *J Natl Cancer Inst* 2014;**106**:pii: dju207.
- Tehrani FR, Solymani-Dodaran M, Tohidi M, Gohari MR, Azizi F. Modeling age at menopause using serum concentration of anti-Mullerian hormone. *J Clin Endocrinol Metab* 2013;**98**:729–735.
- Telfer EE, McLaughlin M. Strategies to support human oocyte development in vitro. *Int J Dev Biol* 2012;**56**:901–907.
- Telfer EE, McLaughlin M, Ding C, Thong KJ. A two step serum free culture system supports development of human oocytes from primordial follicles in the presence of activin. *Hum Reprod* 2008;**23**:1151–1158.
- Telfer EE, Zelinski MB. Ovarian follicle culture: advances and challenges for human and nonhuman primates. *Fertil Steril* 2013;**99**:1523–1533.
- Ting AY, Yeoman RR, Lawson MS, Zelinski MB. In vitro development of secondary follicles from cryopreserved rhesus macaque ovarian tissue after slow-rate freeze or vitrification. *Hum Reprod* 2011;**26**:2461–2472.
- Van der Ven H, Liebenthron J, Beckmann M, Toth B, Korell M, Krussel J, Frambach T, Kupka M, Hohl MK, Winkler-Crepaz K et al. Ninety-five orthotopic transplantations in 74 women of ovarian tissue after cytotoxic treatment in a fertility preservation network: tissue activity, pregnancy and delivery rates. *Hum Reprod* 2016;**31**:2031–2041.
- van Disseldorp J, Faddy MJ, Themmen AP, de Jong FH, Peeters PH, van der Schouw YT, Broekmans FJ. Relationship of serum anti-Mullerian hormone concentration to age at menopause. *J Clin Endocrinol Metab* 2008;**93**:2129–2134.
- Wallace WH, Critchley HO, Anderson RA. Optimizing reproductive outcome in children and young people with cancer. *J Clin Oncol* 2012;**30**:3–5.
- Wallace WH, Kelsey TW. Human ovarian reserve from conception to the menopause. *PLoS One* 2010;**5**:e8772.
- Wallace WH, Kelsey TW, Anderson RA. Fertility preservation in prepubertal girls with cancer: the role of ovarian tissue cryopreservation. *Fertil Steril* 2016;**105**:6–12.
- Wallace WH, Smith AG, Kelsey TW, Edgar AE, Anderson RA. Fertility preservation for girls and young women with cancer: population-based validation of criteria for ovarian tissue cryopreservation. *Lancet Oncol* 2014;**15**:1129–1136.
- Xiao S, Zhang J, Romero MM, Smith KN, Shea LD, Woodruff TK. In vitro follicle growth supports human oocyte meiotic maturation. *Sci Rep* 2015;**5**:17323.
- Xu M, Barrett SL, West-Farrell E, Kondapalli LA, Kiesewetter SE, Shea LD, Woodruff TK. In vitro grown human ovarian follicles from cancer patients support oocyte growth. *Hum Reprod* 2009;**24**:2531–2540.
- Yin H, Kristensen SG, Jiang H, Rasmussen A, Andersen CY. Survival and growth of isolated pre-antral follicles from human ovarian medulla tissue during long-term 3D culture. *Hum Reprod* 2016;**31**:1531–1539.
- Zhou XH, Zhang D, Shi J, Wu YJ. Comparison of vitrification and conventional slow freezing for cryopreservation of ovarian tissue with respect to the number of intact primordial follicles: a meta-analysis. *Medicine (Baltimore)* 2016;**95**:e4095.