



Clayton, N. S., & Ridley, A. J. (2020). Targeting Rho GTPase Signaling Networks in Cancer. *Frontiers in Cell and Developmental Biology*, 8, [222]. <https://doi.org/10.3389/fcell.2020.00222>

Publisher's PDF, also known as Version of record

License (if available):  
CC BY

Link to published version (if available):  
[10.3389/fcell.2020.00222](https://doi.org/10.3389/fcell.2020.00222)

[Link to publication record in Explore Bristol Research](#)  
PDF-document

This is the final published version of the article (version of record). It first appeared online via Frontiers Media at <https://www.frontiersin.org/articles/10.3389/fcell.2020.00222/full>. Please refer to any applicable terms of use of the publisher.

## **University of Bristol - Explore Bristol Research**

### **General rights**

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available:  
<http://www.bristol.ac.uk/pure/about/ebr-terms>



# Targeting Rho GTPase Signaling Networks in Cancer

Natasha S. Clayton and Anne J. Ridley\*

School of Cellular and Molecular Medicine, University of Bristol, Bristol, United Kingdom

## OPEN ACCESS

### Edited by:

Peter J. Parker,  
King's College London,  
United Kingdom

### Reviewed by:

Reinhard Wetzker,  
Friedrich Schiller University Jena,  
Germany  
Lee Bardwell,  
University of California, Irvine,  
United States

### \*Correspondence:

Anne J. Ridley  
anne.ridley@bristol.ac.uk

### Specialty section:

This article was submitted to  
Signaling,  
a section of the journal  
Frontiers in Cell and Developmental  
Biology

**Received:** 15 January 2020

**Accepted:** 16 March 2020

**Published:** 03 April 2020

### Citation:

Clayton NS and Ridley AJ (2020)  
Targeting Rho GTPase Signaling  
Networks in Cancer.  
*Front. Cell Dev. Biol.* 8:222.  
doi: 10.3389/fcell.2020.00222

As key regulators of cytoskeletal dynamics, Rho GTPases coordinate a wide range of cellular processes, including cell polarity, cell migration, and cell cycle progression. The adoption of a pro-migratory phenotype enables cancer cells to invade the stroma surrounding the primary tumor and move toward and enter blood or lymphatic vessels. Targeting these early events could reduce the progression to metastatic disease, the leading cause of cancer-related deaths. Rho GTPases play a key role in the formation of dynamic actin-rich membrane protrusions and the turnover of cell-cell and cell-extracellular matrix adhesions required for efficient cancer cell invasion. Here, we discuss the roles of Rho GTPases in cancer, their validation as therapeutic targets and the challenges of developing clinically viable Rho GTPase inhibitors. We review other therapeutic targets in the wider Rho GTPase signaling network and focus on the four best characterized effector families: p21-activated kinases (PAKs), Rho-associated protein kinases (ROCKs), atypical protein kinase Cs (aPKCs), and myotonic dystrophy kinase-related Cdc42-binding kinases (MRCKs).

**Keywords:** cancer, GTPase, invasion, metastasis, Rho

## INTRODUCTION

Rho GTPases are a family of highly conserved GTPases that are encoded by 20 genes in humans and regulate a range of cellular functions, including vesicular transport, gene expression, neuronal development, and cell division. By far the best characterized of Rho GTPase functions is the organization of the actin cytoskeleton into structures required for cell migration. In cancer, the adoption of a pro-migratory phenotype enables tumor cells to invade the stroma surrounding the primary tumor, migrate toward blood vessels and enter the circulation (**Figure 1A**). Migrating cells extend membrane protrusions at the leading edge, which are driven forward by localized actin polymerization (**Figure 1B**). In general, the formation of lamellipodia is regulated by Rac and Rho, whilst Cdc42 activity drives the formation of filopodia and maintains the cellular polarization required for directional migration. Rho regulates actomyosin contractility, which can lead to the formation of actin stress fibers and focal adhesions on rigid substrata, which are required to generate the traction forces needed to pull the cell body in the direction of movement. Migration of cells in 3D often requires the degradation of matrix components by proteases released from invadopodia, which are actin-rich structures regulated by RhoA, RhoC, and Cdc42 (Lawson and Ridley, 2018).

Altered expression of several Rho GTPases has been reported in a variety of human tumors (reviewed in Svensmark and Brakebusch, 2019). Whilst mutations in genes encoding Rho GTPases are rare in cancer, somatic mutations have been reported in *RHOA*, *RHOB*, *RAC1*, *RAC2*, and *CDC42* (Hurst et al., 2017; Aspenström, 2018). Here, we comment on the recent advances in

targeting Rho GTPase activation and focus on inhibition of Rho GTPase effectors as a viable therapeutic strategy in cancer treatment.

## TARGETING RHO GTPASE ACTIVATION

The binding of GTP to the nucleotide binding pocket of membrane anchored Rho GTPases leads to a conformational change that stimulates their interaction with various downstream effectors, including protein kinases and scaffold/adaptor-like proteins. The activity of most Rho GTPases is terminated by the hydrolysis of bound GTP to GDP, and the dissociation of GDP is required before another GTP molecule can bind. For the classical Rho GTPases, this cycling between GTP- and GDP-bound states is tightly regulated by guanine nucleotide exchange factors (GEFs) and GTPase-activating proteins (GAPs) (Figure 2). Rho GTPases can also be regulated by guanine nucleotide dissociation inhibitor (GDI) binding, which sequesters Rho GTPases in the cytoplasm (Golding et al., 2019).

Significant effort has been made to develop compounds that modulate multiple stages of the GDP/GTP cycle (Jansen et al., 2018; Maldonado and Dharmawardhane, 2018). These include compounds that inhibit GEF-GTPase binding, which have produced promising results in preclinical cancer models. For example, EHop-016 blocks the interaction of Rac1 with Vav2 (Montalvo-Ortiz et al., 2012), and has been shown to enhance the anti-tumor effect of cisplatin in xenograft models of esophageal squamous cell carcinoma (Zeng et al., 2019). ZCL278 blocks the interaction of Cdc42 with intersectin (Friesland et al., 2013) and can inhibit the growth of lung cancer xenografts (Aguilar et al., 2019). It is important to note that the activity of atypical Rho GTPases is unlikely to be regulated by GEF/GAP interactions (Aspenstrom et al., 2007; Hodge and Ridley, 2017), and therefore alternative strategies will be required to target these family members effectively.

In contrast to the prevailing view that the GDP-bound forms of Rho GTPases are inactive, recent data have identified a role for Rho-GDP in oncogenic signaling. The RhoA driver mutation G17V, which disrupts GTP binding, is frequently identified in T-cell lymphoma (Yoo et al., 2014) and studies in *Dictyostelium* cells demonstrate that phosphorylation of RhoA-GDP at S192 leads to increased mTORC2 activity and phosphorylation of AKT (Senoo et al., 2019). These findings suggest that, in certain cancers, targeting GTP-bound Rho GTPases may offer little therapeutic benefit.

Membrane localization of Rho GTPases following post-translational C-terminal lipid modifications, including prenylation and palmitoylation, is crucial for their function (Mitin et al., 2012). The therapeutic potential of prenylation inhibitors is currently being investigated. The geranylgeranyltransferase type I inhibitor GGTI-2418 (PTX-100) has shown anti-tumor effects in breast cancer xenograft models (Aspenstrom et al., 2007) and was well tolerated in an initial phase I clinical trial (Karasic et al., 2019), although further phase I trials are required to optimize dosing and assess responses in patients with advanced malignancies

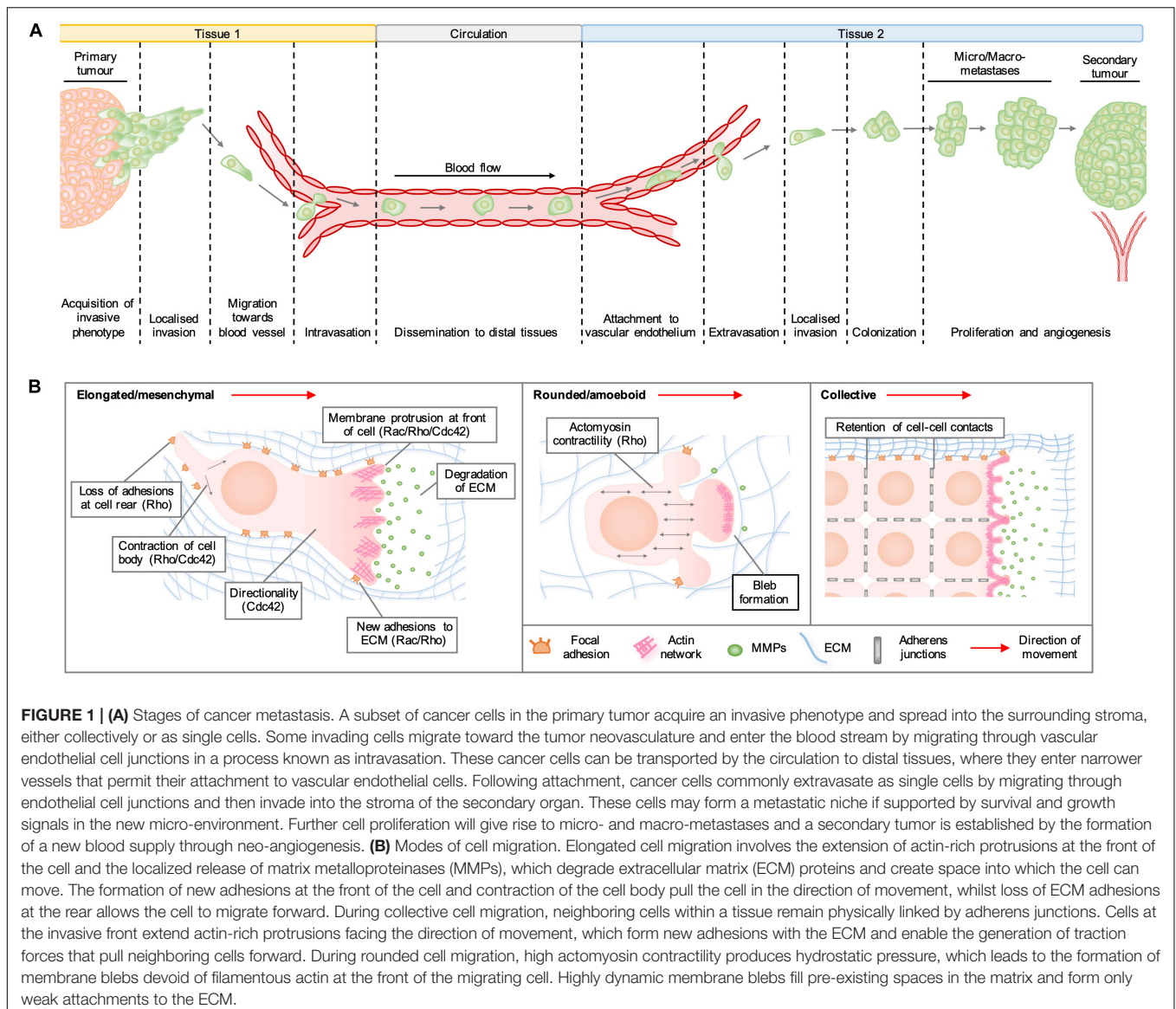
(NCT03900442). Rho GTPase membrane localization can also be inhibited by treatment with statins, which suppress cholesterol biosynthesis, reducing the abundance of farnesyl pyrophosphate and geranylgeranyl pyrophosphate (Denoyelle et al., 2001). Statins are widely prescribed for treatment of cardiovascular disease (Tobert, 2003) and have been reported to reduce cancer recurrence and mortality in several cancer types (Beckwitt et al., 2018). However, this likely reflects the modulation of multiple pathways by statins, including inhibition of Rho GTPases.

A more promising approach to targeting Rho GTPase signaling is through the direct inhibition of Rho GTPase effectors. Here, we discuss the roles of PAKs, ROCKs, MRCKs, and aPKCs in Rho GTPase signaling and review recent advances in targeting these proteins in cancer.

## P21-ASSOCIATED KINASES (PAKS)

The best characterized Rho GTPase effectors are the highly conserved p21-activated kinases (PAKs), which are found in almost all eukaryotes except plants (Hofmann et al., 2004). In humans, the PAKs comprise a family of six serine/threonine kinases which can be divided into two subgroups based on sequence homology and structural similarity. All PAKs possess an N-terminal Cdc42- and Rac-Interactive Binding (CRIB) domain, which mediates their interaction with GTP-bound Rho GTPases, and a conserved kinase domain at the C-terminus (Figure 3). Group I PAKs (PAK1-3) possess an autoinhibitory domain which interacts with the kinase domain in a cis-autoinhibitory interaction. GTPase binding disrupts this autoinhibition and leads to the dimerization and trans-autophosphorylation of two PAK monomers, resulting in their full activation (Sorrell et al., 2019). In contrast, group II PAKs (PAK4-6) show constitutive activation loop autophosphorylation, which is not affected by the binding of Rho GTPases. Group II PAKs are instead activated by the dissociation of a proline-rich pseudosubstrate region from the kinase domain, which may be triggered by the binding of an SH3-domain containing protein (Ha et al., 2012). According to this model, Rho GTPases may promote the activation of group II PAKs by facilitating these protein-protein interactions. PAKs were initially identified as hits in a screen for Cdc42 and Rac1 binding partners (Manser et al., 1994) and have since been shown to interact with a number of other Rho GTPases. PAK signaling downstream of Rho GTPase activity contributes to multiple cellular responses including cell migration, cell survival and sensitivity to certain chemotherapeutic agents (Figure 4; Rane and Minden, 2019).

PAKs are involved in membrane protrusion and focal adhesion turnover by promoting the phosphorylation of proteins involved in actin dynamics (Figure 4). Following activation by Cdc42/Rac1, PAK1 phosphorylates LIM Kinase-1 (LIMK1), which in turn phosphorylates and inactivates cofilin, suppressing the disassembly of actin filaments at the leading edge. Interaction of Rac1 with the WAVE regulatory complex leads to activation of the actin nucleation complex Arp2/3, which when combined with localized inactivation of cofilin, leads to formation of the branched actin network characteristic of lamellipodia (Jansen

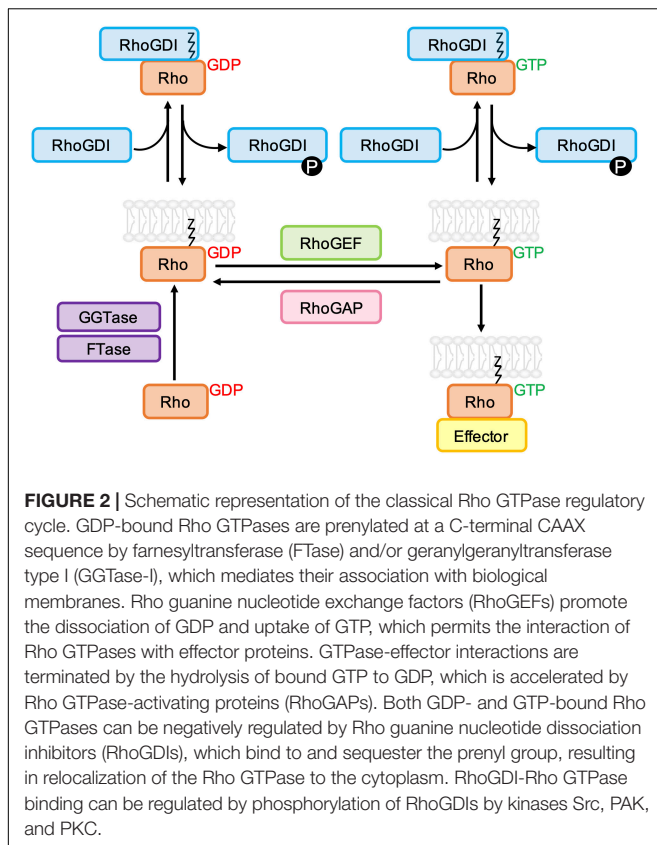


et al., 2018). In breast cancer cell lines, PAK4 has been shown form a complex with RhoU, providing a scaffold leading to increased phosphorylation of paxillin at S272, a crucial step in focal adhesion turnover (Nayal et al., 2006; Dart et al., 2015).

PAKs also contribute to cell migration and invasion by influencing the spatiotemporal activation of Rho GTPases. PAK4 is known to directly bind and inhibit PDZ-RhoGEF (Barac et al., 2004), leading to an indirect inhibition of RhoA. In melanoma cell lines, depletion of PAK4 was associated with an increase in RhoA activity and a decrease in the percentage of cells with mature degradative invadopodia. Expression of PDZ-RhoGEF dominant negative mutants rescued invadopodia in PAK4-depleted melanoma cells, which suggests that a PAK4-mediated reduction in RhoA activity is required for invadopodium maturation. In the same cell lines, depletion of PAK1 was associated with a decrease in RhoA activity and fewer nascent invadopodial actin puncta, suggesting that PAK1 acts upstream

of PAK4 to drive the early stages of invadopodium formation (Nicholas et al., 2016). This suggests that, in addition to the tight spatiotemporal control of RhoC activity (Bravo-Cordero et al., 2011), co-ordinated activation of RhoA at different stages of the invadopodium lifecycle is required for efficient cancer cell invasion.

Group I PAKs can bind directly to the GEFs  $\alpha$ -PIX and  $\beta$ -PIX and influence their activity and/or localization (Manser et al., 1998). PAK1 binding has been shown to stimulate the GEF activity of  $\alpha$ -PIX, resulting in an increase in GTP-bound Cdc42 (Daniels et al., 1999). Other work has shown that PAK1 and Rac1 compete for binding to  $\beta$ -PIX and that the PAK1- $\beta$ -PIX interaction is negatively regulated by PAK1 autophosphorylation. This may provide a mechanism through which other Rho GTPases, such as Cdc42, influence the spatio-temporal activation of Rac1 (ten Klooster et al., 2006). However, PAK1 activation at the leading edge of fibroblasts has been shown to recruit  $\beta$ -PIX



and promote Rac1 activation (Cau and Hall, 2005). Thus, the interplay between PAK1,  $\beta$ -PIX, and Rac1 is likely to be complex and context specific (Jansen et al., 2018).

PAK1 activation can contribute to chemoresistance in melanoma through the inhibition of pro-apoptotic signaling pathways. For example, RhoJ is overexpressed in metastatic melanoma (Ho et al., 2012) and treatment of melanoma cell lines with cisplatin was found to promote PAK1 autophosphorylation in a RhoJ-dependent manner (Figure 4). PAK1 activation led to an uncoupling of the kinase ATR from its effectors Chk1 and ATF2, which resulted in an impaired DNA damage response and reduced apoptosis. The observation that PAK1 depletion sensitized melanoma cell lines to cisplatin suggests that combination treatments of PAK1 inhibitors and DNA-damaging agents may improve chemoresponsiveness in certain human tumors (Ho et al., 2012). In addition, BRAF mutant melanoma cell lines, PAK1 has been shown to phosphorylate the pro-apoptotic protein Bad (Figure 4), preventing its association with Bcl-2 and the subsequent release of cytochrome C from mitochondria (Schurmann et al., 2000; Ruiz et al., 2017). Treatment with the PAK inhibitor FRAX597 led to a decrease in Bad phosphorylation and induced more apoptosis within 24 h in BRAF mutant RhoJ positive melanoma cell lines than the BRAF inhibitors Vemurafenib and Trametinb (Ruiz et al., 2017).

Several ATP-competitive PAK inhibitors have been developed, although specificity has proven challenging, due to the structural similarity of PAK catalytic domains, as well as the relative

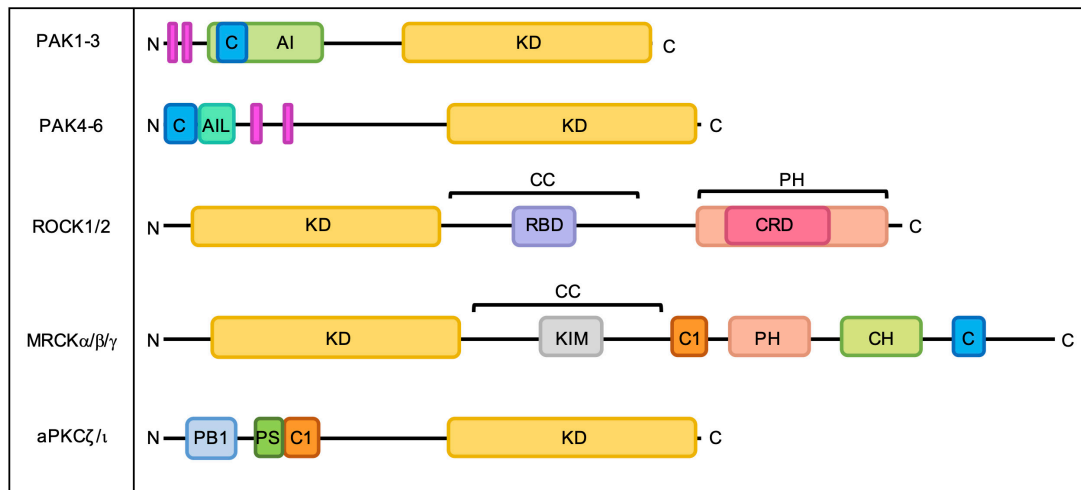
flexibility and “open” conformation of the ATP-binding cleft (Semenova and Chernoff, 2017). FRAX597 is an orally available group I PAK inhibitor which has been shown to inhibit cancer cell growth *in vitro* (Chow et al., 2012; Semenova et al., 2017; Tan et al., 2017; Araiza-Olivera et al., 2018) and to sensitize orthotopic tumors to gemcitabine in a murine model of pancreatic cancer (Yeo et al., 2016). The closely related inhibitor FRAX486 showed anti-proliferative effects in a mouse model of childhood acute lymphoblastic leukemia (Siekmann et al., 2018) and the analog FRAX1036 was recently found to inhibit tumor growth in murine models of breast (Korobeynikov et al., 2019) and thyroid (Knippler et al., 2019) cancer. However, FRAX1036 has been found to adversely inhibit hERG potassium channels, suggesting that FRAX1036 and its analogs may not be suitable for clinical use (Ndubaku et al., 2015). Glaucarubinone, first developed as an antimalarial, was found to inhibit the activation of both PAK1 and PAK4, which may prove advantageous in treating cancers where the co-ordinated activity of these isoforms contributes to disease progression (Yeo et al., 2014; Nicholas et al., 2016). Glaucarubinone reduced the growth of pancreatic cancer xenografts, and glaucarubinone and gemcitabine were found to have a synergistic effect on the inhibition of PAK1/4 activation and tumor growth (Yeo et al., 2014). Although originally developed as a PAK4 inhibitor, the Pfizer compound PF-3758309 inhibits all PAK family kinases and is the only ATP-competitive PAK inhibitor to have reached clinical trials (Semenova and Chernoff, 2017). Following a phase I trial in patients with advanced solid tumors, PF-3758309 was withdrawn from clinical investigation due to poor bioavailability, adverse effects and lack of tumor response (Mileshkin et al., 2011).

KPT-9274 is an allosteric PAK4 inhibitor which binds to and destabilizes PAK4 (Rane et al., 2017). KPT-9274 and its analogs may therefore prove superior to kinase inhibitors in treating certain cancers as they could provide dual blockade of PAK4 kinase-dependent and independent functions. KPT-9274 has been shown to inhibit tumor growth of a number of pre-clinical cancer models (Rane and Minden, 2019) and is currently in phase I clinical trials for non-Hodgkin lymphoma and advanced solid malignancies (NCT02702492).

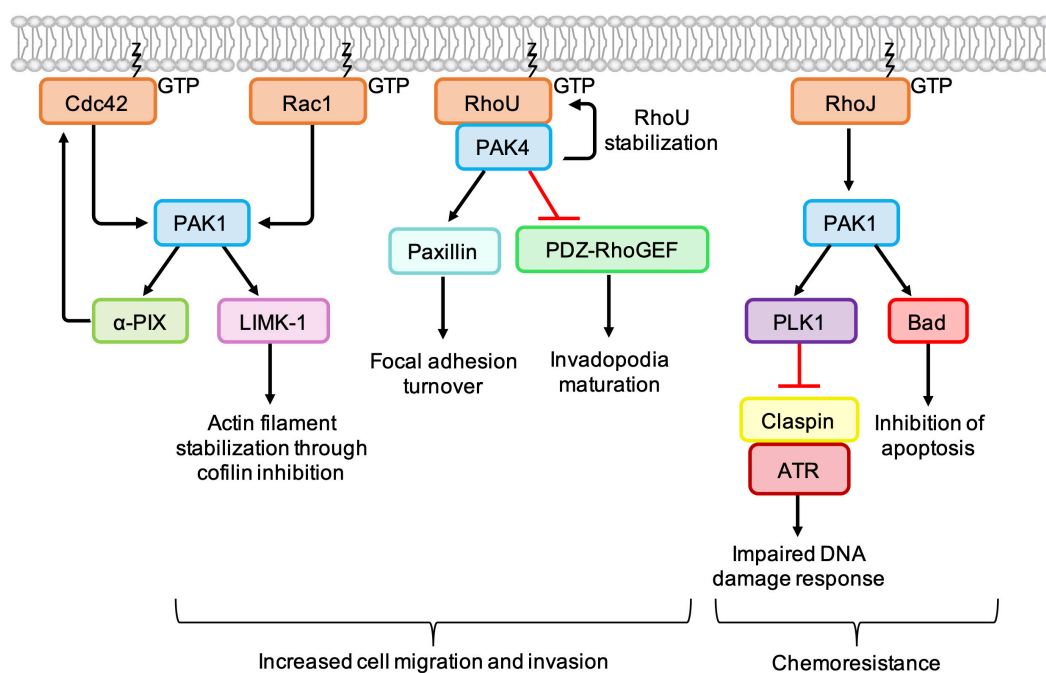
## RHO-ASSOCIATED PROTEIN KINASES (ROCKS)

The Rho GTPase effectors ROCK1 and ROCK2 are AGC-family serine/threonine kinases that are involved in a diverse range of cellular processes, including cell motility, cell survival and proliferation, gene transcription, differentiation, and angiogenesis (Wei et al., 2016). Here, we focus on the role of ROCK in promoting actin polymerization and actomyosin contractility.

Both ROCKs have an N-terminal kinase domain, a coiled-coil region containing a Rho-binding domain (RBD) and a C-terminal pleckstrin homology (PH) domain (Figure 3; Morgan-Fisher et al., 2013). ROCKs exist as constitutive parallel homodimers (Truebestein et al., 2015) and interaction with membrane lipids (Yoneda et al., 2005), cleavage by granzyme



**FIGURE 3** | Domain organization of PAKs, ROCKs, MRCKs, and aPKCs. C, Cdc42- and Rac-Interactive Binding (CRIB) domain; AI, autoinhibitory domain; KD, kinase domain; AIL, autoinhibitory-like domain; RBD, Rho-binding domain; CC, coiled-coil region; PH, pleckstrin homology domain; CRD, cysteine-rich domain; C1, C1 domain; CH, citron homology domain; PB1, Phox Bem1 domain; PS, pseudosubstrate motif. Proline-rich regions are indicated by pink bars.

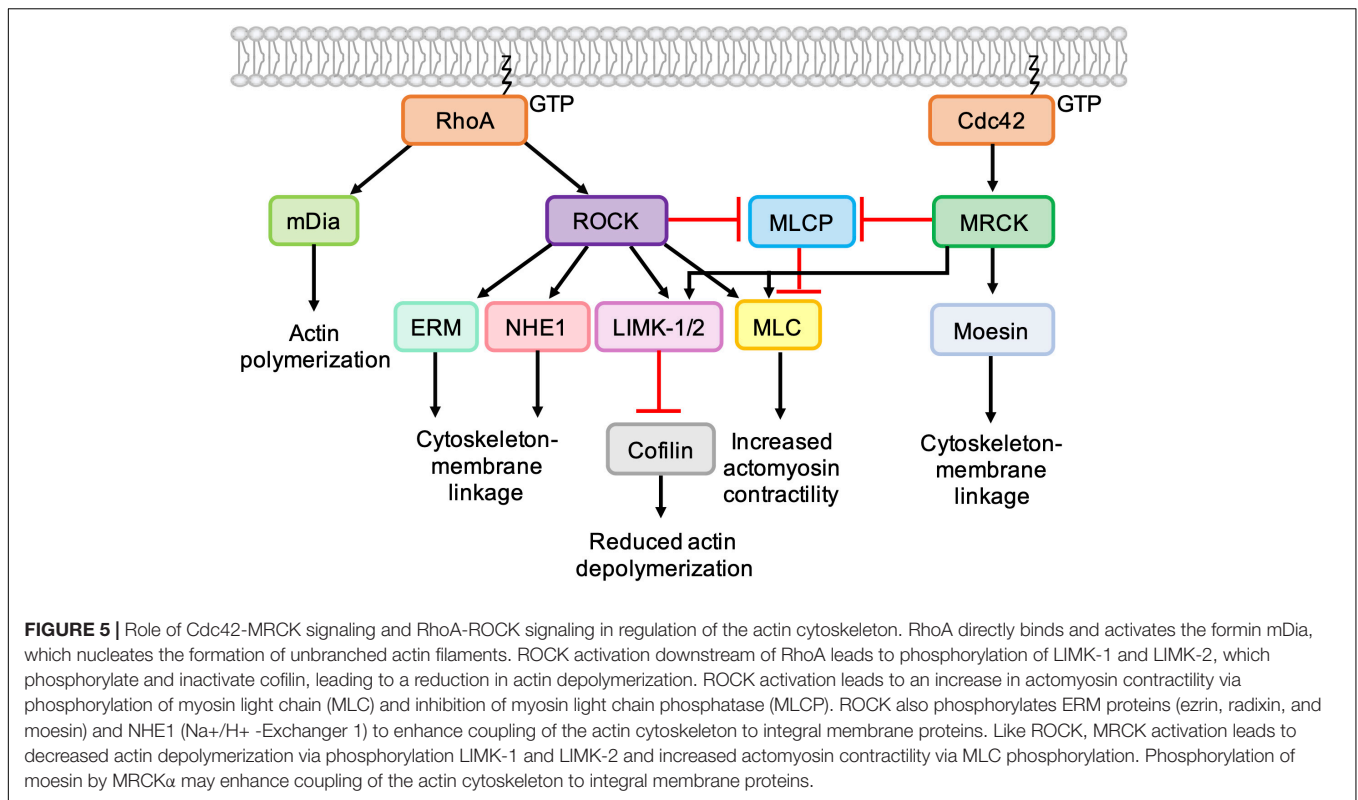


**FIGURE 4** | Cancer-associated PAK signaling pathways activated downstream of Rho GTPases. PAK1 phosphorylates LIM Kinase-1 (LIMK-1), which stabilizes actin filaments through inhibition of cofilin. PAK1 also binds to and stimulates the GEF  $\alpha$ -PIX, leading to an increase in Cdc42 activation. PAK1 activates PLK1, which leads to phosphorylation and degradation of claspin. In the absence of claspin, ATR is uncoupled from its effectors, resulting in an impaired DNA damage response. PAK1 also phosphorylates the pro-apoptotic protein Bad, which prevents its association with Bcl-2 and inhibits apoptosis. PAK4 binding protects RhoU from proteasomal degradation and promotes focal adhesion turnover by providing a scaffold that promotes the phosphorylation of paxillin. PAK4 also inhibits PDZ-RhoGEF, which suppresses the activation of RhoA and promotes invadopodia maturation.

B or caspases (Sebbagh et al., 2001; Sebbagh et al., 2005) and binding of RhoA, RhoB, and RhoC (Amano et al., 2000) have been shown to stimulate ROCK signaling. ROCK1 and ROCK2 share 92% sequence homology within their kinase domains (Nakagawa et al., 1996) and have many common substrates

(Yoneda et al., 2005), yet are not functionally redundant (Jerrell et al., 2017).

During cell migration, ROCK promotes the extension of actin-rich membrane protrusions through the activation of LIMK1/2 and inhibition of cofilin (Ohashi et al., 2000) and also stimulates



cell body contraction through the generation of actomyosin contractility (Figure 5). Following activation by RhoA and RhoC, ROCK directly phosphorylates the myosin light chain (MLC) of non-muscle myosin II and inhibits MLC phosphatase through phosphorylation of the myosin binding subunit (Julian and Olson, 2014). Phosphorylation of MLC on Ser19 and Thr18 influences the ATPase activity of the myosin heavy chain (MHC) head groups, which move along actin filaments to produce contractile force (Mizutani et al., 2006). In a 3D environment, single cells have been shown to utilize two distinct modes of motility that are driven by different Rho signaling pathways (Figure 1B). Rac1 activity drives elongated cell motility, which is characterized by cell polarization, the extension of F-actin-rich protrusions and degradation of the ECM. In contrast, RhoA/C signaling through ROCK promotes a rounded bleb-based mode of motility driven by high levels of actomyosin contractility, which enables the cell to squeeze into pre-existing spaces and deform the ECM without the need for significant pericellular proteolysis (Wyckoff et al., 2006). To adopt a rounded mode of migration, Rho-ROCK signaling is often coupled to inhibition of Rac1 signaling, which can be achieved by stimulation of the Rac GAP ARHGAP22 downstream of ROCK activation. During elongated cell motility, active Rac1 suppresses the high levels of actomyosin contractility required for rounded motility via its effector WAVE2, which negatively regulates MLC phosphorylation (Sanz-Moreno et al., 2008). Cancer cells have been shown to convert between elongated and rounded modes of motility, suggesting that inhibition of both modes will be required to effectively prevent metastasis, for example

via dual inhibition of secreted proteases and ROCK activity (Yamada and Sixt, 2019).

Deregulation of Rho-ROCK signaling has been identified in a number of human cancers and correlated with disease progression (Rodriguez-Hernandez et al., 2016). The *ROCK1* gene is amplified in a number of lung and gastric cancers as well as in head-and-neck squamous cell carcinoma (Svensmark and Brakebusch, 2019), whilst *ROCK2* amplification has been associated with peripheral nerve sheath tumors (Upadhyaya et al., 2012). Three somatic mutations were identified in *ROCK1* that result in activating C-terminal truncations (Lochhead et al., 2010) and polymorphisms in both *ROCK1* and *ROCK2* have been associated with colorectal cancer development (Sari et al., 2013).

Although significant effort has been made to develop ROCK inhibitors for cancer intervention, the vast majority of compounds have not progressed to clinical trials. The ATP-competitive ROCK inhibitors Y-27632 and fasudil have been used extensively as tool compounds and have been shown to inhibit cancer cell migration in various *in vitro* cancer models (Wei et al., 2016). Although fasudil is clinically approved for the acute treatment of cerebral vasospasm in Japan and China, its pharmacokinetic profile makes it unsuitable for use in chemotherapy (Rath et al., 2018). More recently developed pan-ROCK inhibitors, such as OXA-06, PT262, RKI-1447, and CTT129253, show more potent ROCK inhibition and achieve anti-tumor effects in pre-clinical models (reviewed in Wei et al., 2016; Jansen et al., 2018). Several compounds displaying selectivity for *ROCK2* over *ROCK1* have also been reported, although the efficacy of these compounds has mostly been tested

in models of other diseases, such as glaucoma, hypertension and chronic kidney disease (Wei et al., 2016). To date, only one ROCK inhibitor has progressed into clinical trials for cancer treatment. AT13148 is an orally available multi-AGC kinase inhibitor that was identified through a fragment-based screen and was found to potently inhibit ROCK1 and ROCK2 (Yap et al., 2012). AT13148 was shown to have anti-tumor effects in pre-clinical models of pancreatic (Rath et al., 2018), breast, prostate and uterine cancer (Yap et al., 2012) and was well tolerated in a phase I clinical trial in patients with advanced solid tumors (NCT01585701) (Kumar et al., 2014).

It is important to note that pan-ROCK inhibition using Y-27632 has been shown to increase the proliferation and migration of a number of *in vitro* and *in vivo* cancer models (Adachi et al., 2011; Vishnubhotla et al., 2012; Yang and Kim, 2014; Chang et al., 2018). These effects may be explained by the observation that ROCK activation can contribute to negative feedback mechanisms that regulate pro-proliferative pathways. In a model of pancreatic cancer, EGF stimulation led to increased ROCK activation, which then negatively regulated EGFR signaling by affecting receptor trafficking. Treatment with Y-27632 removed this negative regulation and resulted in significantly increased AKT and MAPK signaling following EGF stimulation (Nakashima et al., 2011). Targeting ROCK may therefore only be appropriate in certain contexts and combination treatments may be required to avoid compensatory upregulation of other kinases involved in actomyosin function, such as LIMKs and MRCKs (Rath and Olson, 2012).

## MYOTONIC DYSTROPHY KINASE-RELATED CDC42-BINDING KINASES (MRCKS)

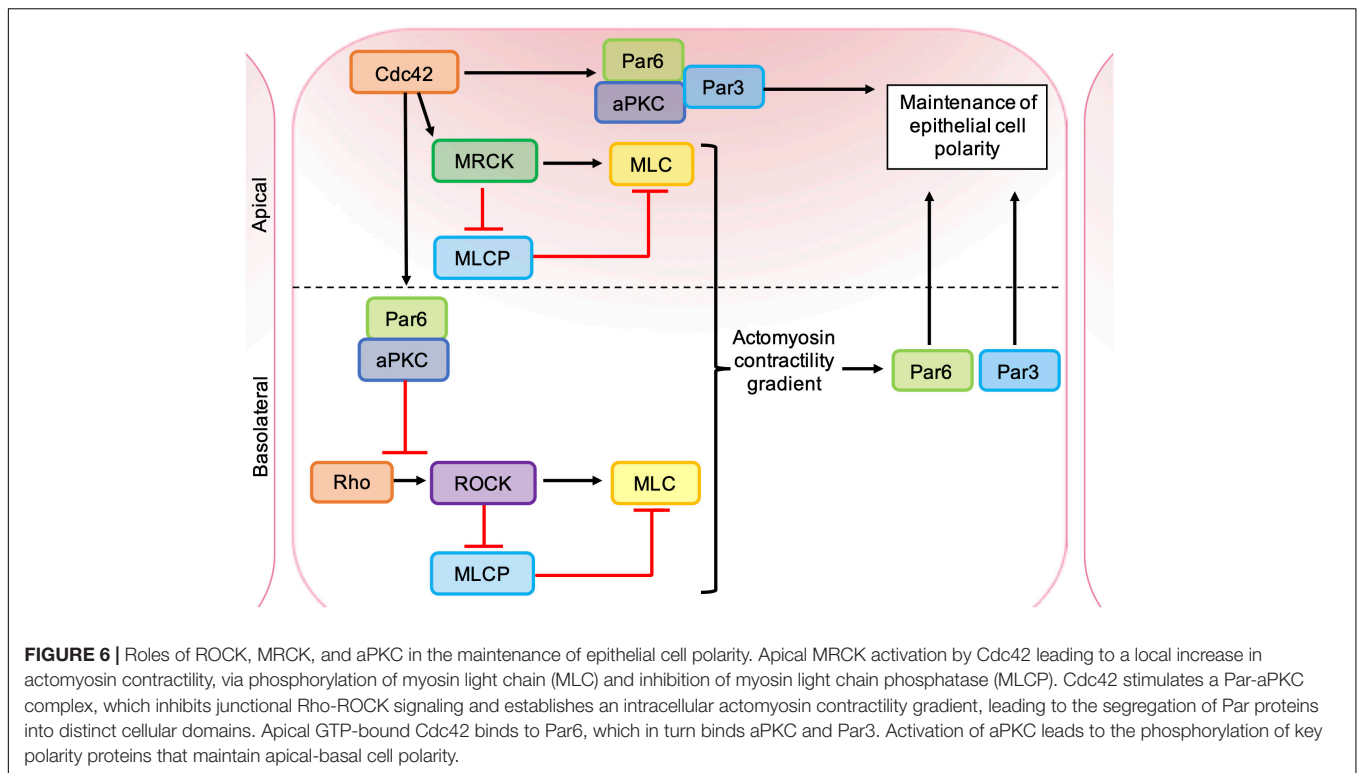
In humans, myotonic dystrophy kinase-related Cdc42-binding kinases (MRCKs) form a family of three serine/threonine protein kinases that are evolutionarily related to ROCKs and play a key role in the regulation of actomyosin contractility (Zhao and Manser, 2015; Leroux et al., 2018). MRCK $\alpha$  and MRCK $\beta$  are ubiquitously expressed and share 85% kinase domain homology, whilst MRCK $\gamma$  is more divergent and shows a more restricted tissue expression pattern (Unbekandt and Olson, 2014). All MRCK proteins have an N-terminal protein kinase domain followed by a C-terminal protein kinase C conserved region (C1), PH-like domain, citron homology (CH) domain and CRIB domain (Figure 3). Membrane localization of MRCKs may be mediated at least in part by the C1 domain, which has been shown to bind phorbol esters (Choi et al., 2008; Talman et al., 2014), and the PH-like domain, which may bind membrane lipids. The CH domain may facilitate substrate docking or engage in protein-protein interactions that specify protein localization (Unbekandt and Olson, 2014). The CRIB domains of MRCK $\alpha$  and MRCK $\beta$  bind GTP-loaded Cdc42 (Leung et al., 1998) and Rac1 (Ng et al., 2004; Schwarz et al., 2012), whilst the CRIB domain of MRCK $\gamma$  has been shown to bind to RhoQ with higher affinity than Cdc42 and Rac1 (Ng et al., 2004). Although the molecular mechanisms underlying MRCK activation are not

fully understood, Cdc42/Rac1 binding, membrane localization and the release of auto-inhibitory interactions appear to be required for MRCK signaling (Leung et al., 1998; Unbekandt and Olson, 2014; Zhao and Manser, 2015). Recent identification of an autophosphorylation site in MRCK $\alpha$  at Ser1003 led to the development of a phospho-site specific MRCK $\alpha$  antibody that can be used to assess levels of MRCK $\alpha$  activation and will likely prove useful for the further elucidation of MRCK $\alpha$  activation mechanisms (Unbekandt et al., 2018). Autophosphorylation of MRCK $\beta$  at Thr1108 can be used as a readout of MRCK $\beta$  kinase activity, although attempts to develop a suitable antibody tool have so far been unsuccessful (Unbekandt et al., 2019).

The catalytic domains of MRCK and ROCK are highly related and have several common substrates (Zhao and Manser, 2015). MRCK $\alpha$  is known to phosphorylate LIMK1/2 (Sumi et al., 2001), leading to phosphorylation and inhibition of cofilin (Ohashi et al., 2000), and also promotes MLC phosphorylation through the inhibition of MLC phosphatase (Tan et al., 2001; Wilkinson et al., 2005) and phosphorylation of MLC on Ser19 (Leung et al., 1998; Figure 5). However, ROCK and MRCK are not functionally redundant and the differential regulation of ROCK and MRCK activity plays an important role in establishing cell polarity (Figure 6). Epithelial polarization requires the segregation of PAR proteins into distinct cortical domains, which is driven at least in part by an intracellular actomyosin activity gradient. Apical actomyosin contractility is stimulated by Cdc42-MRCK activity, whilst RhoA-ROCK activity regulates contractility along cell-cell junctions. Apical Cdc42 activity leads to stimulation of a Par6-aPKC complex, which in turn inhibits junctional RhoA-ROCK signaling, establishing an actomyosin contractility gradient (Etienne-Manneville and Hall, 2001; Zihni et al., 2017). Differential regulation of ROCK and MRCK between different tumor cells may also be important for invasion. Studies of squamous cell carcinoma (SCC) cells and stromal fibroblasts in 3D co-cultures found that collective invasion required Rho-ROCK signaling in fibroblasts and Cdc42-MRCK signaling in SCC cells. Imaging revealed that fibroblasts always act as the “leading cell,” whilst SCC cells migrate through the channels in the ECM created by the fibroblast. This remodeling of the ECM by fibroblasts was dependent on both extracellular protease activity and the generation of contractile force by Rho-ROCK signaling. In contrast, the SCC cells were found to rely on Cdc42-MRCK-mediated regulation of MLC to follow leading the fibroblasts (Gaggioli et al., 2007).

MRCK $\alpha$  expression is increased in a number of human cancers (Unbekandt and Olson, 2014) and dual inhibition of ROCK and MRCK activity has been shown to produce a greater inhibition of cancer cell migration and invasion than blocking ROCK alone (Wilkinson et al., 2005; Kale et al., 2014). Widely used ROCK inhibitors such as Y-27632 and fasudil bind to MRCKs (Heikkila et al., 2011), although these compounds also inhibit several other kinases and are not clinically viable (Bain et al., 2007). Whilst dual ROCK/MRCK inhibition remains a valid therapeutic strategy, selective MRCK inhibitors may prove beneficial in overcoming the dose-limiting hypotension associated with long-term ROCK inhibitor treatment (Kale et al., 2015). The recently identified azaindole compounds BDP8900





and BDP9066 shown to have high selectivity for MRCK $\alpha$  and MRCK $\beta$  over ROCK1 and ROCK2 in *in vitro* kinase assays and were found to inhibit MRCK-mediated MLC phosphorylation more potently than ROCK-mediated MLC phosphorylation in a breast cancer cell line. MRCK $\alpha$  and MRCK $\beta$  are overexpressed in squamous cell carcinoma (SCC) (Nindl et al., 2006) and treatment of SCC cell lines with BDP9066 led to morphological changes and decreased bundling of filamentous actin, as well as reduced cell motility and invasion. In a mouse model of SCC, topical application of BDP9066 reduced MRCK $\alpha$  activation in the skin and resulted in a decrease in tumor volume relative to controls. In a screen of 757 human cancer cell lines from 40 different cancer types, BDP8900 and BDP9066 treatment produced consistent anti-proliferative effects, suggesting MRCK may be a valid therapeutic target in a number of cancers other than SCC (Unbekandt et al., 2018). It is important to note that the toxicity of systemic BDP8900 and BDP9066 administration has not been studied and information regarding the bioavailability of these compounds is not yet available. The development of these novel MRCK-selective inhibitors will nevertheless prove valuable for the further pre-clinical validation of MRCK as a therapeutic target and may inform the future development of MRCK inhibitors better suited for clinical use.

## ATYPICAL PROTEIN KINASE CS (APKCS)

The protein kinase C (PKC) family comprises three subfamilies of serine/threonine kinases; classic PKCs (cPKCs; PKC $\alpha$ , PKC $\beta$ ,

and PKC $\gamma$ ), novel PKCs (nPKC; PKC $\delta$ , PKC $\epsilon$ , PKC $\eta$ , and PKC $\theta$ ) and atypical PKCs (aPKC; PKC $\zeta$  and PKC $\iota$ ). All PKC proteins exist in a basally auto-inhibited conformation, which is mediated by an interaction between the conserved C-terminal kinase domain and an N-terminal regulatory region, which comprises a pseudosubstrate motif and other structural domains that vary between PKC subfamilies (Figure 3). Classic PKCs require diacylglycerol (DAG), phosphatidylserine (PS) and Ca<sup>2+</sup> for activation, whilst nPKCs require only interaction with DAG and PS (Isakov, 2018). In contrast, aPKCs are primarily activated by protein-protein interactions mediated by an N-terminal Phox Bem1 (PB1) domain and can also be regulated by PIP<sub>3</sub> as well as by specific phosphorylation events (Garg et al., 2014). Through forming a complex with Par6 and Par3, aPKCs play a key role in establishing both the apical-basal polarity of epithelial tissues and the front-to-rear polarity required for cell migration downstream of Cdc42.

Most human cancers arise from epithelial tissues and the disruption of apical-basal epithelial polarity is considered an early event in tumorigenesis (Fomicheva et al., 2019). During the establishment of epithelial cell polarity, GTP-bound Cdc42 binds to Par6 via its semi-CRIB domain, which in turn binds to PKC $\iota$  via a PB1-PB1 interaction (Hirano et al., 2005). The interaction of Cdc42 with Par6-PKC $\iota$  results in allosteric PKC $\iota$  activation and recruitment of Par3, which binds to PKC $\iota$  and Par6 via its PDZ domain. Par3 then targets the complex to the apical compartment of the cell membrane through its interaction with several cell adhesion molecules (Figure 6). Apical aPKC activity then maintains apical-basal polarity through the phosphorylation of a number of polarity proteins, including

MARCKS, LGN, Lgl, GSK3 $\beta$ , Par1, Crumbs, and Lin5/NuMA (Chen and Zhang, 2013). One mechanism by which human tumors lose polarity is through loss of Par3, which leads to mislocalization of PKC $\iota$  in the cytoplasm and nucleus. Mislocalized PKC $\iota$  can remain associated with Par6 and has been shown to contribute to maintenance of the transformed phenotype (reviewed in Parker et al., 2014).

The Par6-aPKC complex also plays a crucial role in establishing the front-back polarity required for directional cell migration and invasion. PKC $\zeta$  has been shown to promote invasion and metastasis of breast cancer xenografts in mice by promoting the nuclear localization of NF $\kappa$ B-p65 and suppressing the expression of junctional proteins E-cadherin and ZO1 (Paul et al., 2015). PKC $\iota$  has also been shown to stimulate cell migration by regulating the trafficking of protease-containing vesicles to the plasma membrane of invadopodia. In invasive breast cancer cell lines, the secretion of type 1-matrix metalloproteinase (MT1-MMP) was shown to be dependent on the interaction of cortactin and dynamin-2 on MT1-MMP containing endosomes. This interaction was found to be regulated by the PKC $\iota$ -mediated phosphorylation of cortactin on Ser261 and depletion of PKC $\iota$  led to a decrease in the release of MT1-MMP from invadopodia and a reduction in the degradative capacity of the cells (Rosse et al., 2014).

In non-small cell lung cancer (NSCLC), PKC $\iota$  has been shown to promote cell survival and invasion through its action on the Rho GEF Ect2. Binding of Ect2 to the Par6-PKC $\iota$  leads to phosphorylation of Ect2 on T328, which enhances its GEF activity toward Rac1 (Justilien et al., 2011). PKC $\iota$ -mediated Rac1 activation can stimulate MEK-ERK signaling in pancreatic cancer cell lines, leading to increased anchorage-independent growth *in vitro* and promoting tumor growth and metastasis in an orthotopic mouse model (Scotti et al., 2010).

Overexpression of PKC $\iota$  has been observed in several human cancers, including gastric (Hashimoto et al., 2019), pancreatic (Scotti et al., 2010), breast (Rosse et al., 2014), ovarian (Tsang et al., 2017), prostate (Apostolatos et al., 2018), and lung cancer (Kim et al., 2019) and has been shown to act as an oncogenic driver in numerous studies (reviewed in Parker et al., 2014). The role of PKC $\zeta$  in cancer is less clear, with both increased and decreased expression reported in human tumors (Garg et al., 2014). Targeting aPKCs in cancer may therefore only be appropriate in certain contexts. Several ATP-competitive inhibitors have been described that show selectivity for PKC $\zeta$ /PKC $\iota$  over cPKC and nPKC isoforms. For example, CRT0066854 potently inhibits PKC $\iota$  and PKC $\zeta$ , with some off-target inhibition of ROCK2 and PRK2, and was shown to inhibit anchorage-independent cell growth and migration *in vitro* (Kjaer et al., 2013). More recently, the selective aPKC inhibitors 2-acetyl-1,3-cyclopentanedione (ACPD) and 3,4-diaminonaphthalene-2,7-disulfonic acid (DNDA) were found

to suppress the proliferation of melanoma cell lines showing high levels of PKC $\iota$  and PKC $\zeta$  expression, whilst having little effect on the proliferation of normal melanocytes (Ratnayake et al., 2017). Compounds that block protein-protein interactions at the PB1 domain may provide greater selectivity for aPKCs over other protein kinases. The gold compound aurothiomalate (ATM) selectively inhibits the binding of Par6 to PKC $\iota$  and PKC $\zeta$  (Erdogan et al., 2006; Stallings-Mann et al., 2006; Butler et al., 2015) and has been shown to inhibit tumor growth in animal models (Butler et al., 2015; Tsang et al., 2017). ATM is approved for the treatment of rheumatoid arthritis (RA) and a phase I study found ATM to be well tolerated in patients with advanced NSCLC, ovarian cancer and pancreatic cancer (Mansfield et al., 2013). Another gold compound used to treat RA, auranofin (ANF), is reported to block the interaction of PKC $\iota$  with Par6 more potently than ATM (Parker et al., 2014) and phase I/II trials are ongoing to investigate the effect of ANF in combination with an mTOR inhibitor in lung cancer (NCT01737502).

## CONCLUDING REMARKS

The role of Rho GTPases as key regulators of cell migration and invasion has been recognized for decades, yet few compounds targeting Rho GTPase signaling networks have been developed beyond an early preclinical stage. Due to the challenges of inhibiting Rho GTPase activation directly, targeting Rho GTPase effectors remains the most promising approach. Whilst PAK and ROCK inhibitors have progressed to phase I clinical trials, further work is needed to elucidate the context-dependent roles of Rho GTPase effectors and to identify compensatory feedback networks which may limit the success of these targeted therapies. Genome sequencing of human tumors has identified several mutations in Rho GTPases, yet the functional and clinical significance of many of these mutants remain poorly understood. The generation of mouse models with tissue-specific Rho GTPase alterations will be crucial in understanding the role of these mutants in cancer and may lead to the identification of novel therapeutic targets within Rho GTPase signaling networks.

## AUTHOR CONTRIBUTIONS

NC and AR wrote, corrected, and reviewed the final manuscript.

## FUNDING

We thank the School of Cellular and Molecular Medicine (University of Bristol) for the research support.

## REFERENCES

- Adachi, S., Yasuda, I., Nakashima, M., Yamauchi, T., Yoshioka, T., Okano, Y., et al. (2011). Rho-kinase inhibitor upregulates migration by altering focal adhesion formation via the Akt pathway in colon cancer cells. *Eur. J. Pharmacol.* 650, 145–150. doi: 10.1016/j.ejphar.2010.10.014
- Aguilar, B. J., Zhao, Y., Zhou, H., Huo, S., Chen, Y. H., and Lu, Q. (2019). Inhibition of Cdc42-intersectin interaction by small molecule ZCL367

- impedes cancer cell cycle progression, proliferation, migration, and tumor growth. *Cancer Biol. Ther.* 20, 740–749. doi: 10.1080/15384047.2018.1564559
- Amano, M., Fukata, Y., Shimokawa, H., and Kaibuchi, K. (2000). Purification and in vitro activity of Rho-associated kinase. *Methods Enzymol.* 325, 149–155.
- Apostolatos, A. H., Ratnayake, W. S., Win-Piazza, H., Apostolatos, C. A., Smalley, T., Kang, L., et al. (2018). Inhibition of atypical protein kinase Ciota effectively reduces the malignancy of prostate cancer cells by downregulating the NF-kappaB signaling cascade. *Int. J. Oncol.* 53, 1836–1846. doi: 10.3892/ijo.2018.4542
- Araza-Olivera, D., Feng, Y., Semenova, G., Prudnikov, T. Y., Rhodes, J., and Chernoff, J. (2018). Suppression of RAC1-driven malignant melanoma by group A PAK inhibitors. *Oncogene* 37, 944–952. doi: 10.1038/onc.2017.400
- Aspenström, P. (2018). Activated Rho GTPases in cancer—the beginning of a new paradigm. *Int. J. Mol. Sci.* 19:3949.
- Aspenstrom, P., Ruusala, A., and Pacholsky, D. (2007). Taking Rho GTPases to the next level: the cellular functions of atypical Rho GTPases. *Exp. Cell Res.* 313, 3673–3679.
- Bain, J., Plater, L., Elliott, M., Shpiro, N., Hastie, C., McLauchlan, H., et al. (2007). The selectivity of protein kinase inhibitors: a further update. *Biochem. J.* 408, 297–315.
- Barac, A., Basile, J., Vazquez-Prado, J., Gao, Y., Zheng, Y., and Gutkind, J. S. (2004). Direct interaction of p21-activated kinase 4 with PDZ-RhoGEF, a G protein-linked Rho guanine exchange factor. *J. Biol. Chem.* 279, 6182–6189.
- Beckwith, C. H., Brufsky, A., Oltvai, Z. N., and Wells, A. (2018). Statin drugs to reduce breast cancer recurrence and mortality. *Breast Cancer Res.* 20:144. doi: 10.1186/s13058-018-1066-z
- Bravo-Cordero, J. J., Oser, M., Chen, X., Eddy, R., Hodgson, L., and Condeelis, J. (2011). A novel spatiotemporal RhoC activation pathway locally regulates cofilin activity at invadopodia. *Curr. Biol.* 21, 635–644. doi: 10.1016/j.cub.2011.03.039
- Butler, A. M., Scotti Buzhardt, M. L., Erdogan, E., Li, S., Inman, K. S., Fields, A. P., et al. (2015). A small molecule inhibitor of atypical protein kinase C signaling inhibits pancreatic cancer cell transformed growth and invasion. *Oncotarget* 6, 15297–15310.
- Cau, J., and Hall, A. (2005). Cdc42 controls the polarity of the actin and microtubule cytoskeletons through two distinct signal transduction pathways. *J. Cell Sci.* 118, 2579–2587.
- Chang, F., Zhang, Y., Mi, J., Zhou, Q., Bai, F., Xu, X., et al. (2018). ROCK inhibitor enhances the growth and migration of BRAF-mutant skin melanoma cells. *Cancer Sci.* 109, 3428–3437. doi: 10.1111/cas.13786
- Chen, J., and Zhang, M. (2013). The Par3/Par6/aPKC complex and epithelial cell polarity. *Exp. Cell Res.* 319, 1357–1364. doi: 10.1016/j.yexcr.2013.03.021
- Choi, S. H., Czifra, G., Kedei, N., Lewin, N. E., Lazar, J., Pu, Y., et al. (2008). Characterization of the interaction of phorbol esters with the C1 domain of MRCK (myotonic dystrophy kinase-related Cdc42 binding kinase) alpha/beta. *J. Biol. Chem.* 283, 10543–10549. doi: 10.1074/jbc.M707463200
- Chow, H. Y., Jubba, A. M., Koch, J. N., Jaffer, Z. M., Stepanova, D., Campbell, D. A., et al. (2012). p21-Activated kinase 1 is required for efficient tumor formation and progression in a Ras-mediated skin cancer model. *Cancer Res.* 72, 5966–5975. doi: 10.1158/0008-5472.CAN-12-2246
- Daniels, R. H., Zenke, F. T., and Bokoch, G. M. (1999). alphaPix stimulates p21-activated kinase activity through exchange factor-dependent and -independent mechanisms. *J. Biol. Chem.* 274, 6047–6050.
- Dart, A. E., Box, G. M., Court, W., Gale, M. E., Brown, J. P., Pinder, S. E., et al. (2015). PAK4 promotes kinase-independent stabilization of RhoU to modulate cell adhesion. *J. Cell Biol.* 211, 863–879. doi: 10.1083/jcb.201501072
- Denoyelle, C., Vasse, M., Korner, M., Mishal, Z., Ganne, F., Vannier, J. P., et al. (2001). Cerivastatin, an inhibitor of HMG-CoA reductase, inhibits the signaling pathways involved in the invasiveness and metastatic properties of highly invasive breast cancer cell lines: an in vitro study. *Carcinogenesis* 22, 1139–1148.
- Erdogan, E., Lamark, T., Stallings-Mann, M., Lee, J., Pellicchia, M., Thompson, E. A., et al. (2006). Aurothiomalate inhibits transformed growth by targeting the PB1 domain of protein kinase Ciota. *J. Biol. Chem.* 281, 28450–28459.
- Etienne-Manneville, S., and Hall, A. (2001). Integrin-mediated activation of Cdc42 controls cell polarity in migrating astrocytes through PKCzeta. *Cell* 106, 489–498.
- Fomicheva, M., Tross, E. M., and Macara, I. G. (2019). Polarity proteins in oncogenesis. *Curr. Opin. Cell Biol.* 62, 26–30. doi: 10.1016/j.ccb.2019.07.016
- Friesland, A., Zhao, Y., Chen, Y. H., Wang, L., Zhou, H., and Lu, Q. (2013). Small molecule targeting Cdc42-intersectin interaction disrupts Golgi organization and suppresses cell motility. *Proc. Natl. Acad. Sci. U.S.A.* 110, 1261–1266. doi: 10.1073/pnas.1116051110
- Gaggioli, C., Hooper, S., Hidalgo-Carcedo, C., Grosse, R., Marshall, J. F., Harrington, K., et al. (2007). Fibroblast-led collective invasion of carcinoma cells with differing roles for RhoGTPases in leading and following cells. *Nat. Cell Biol.* 9, 1392–1400.
- Garg, R., Benedetti, L. G., Abera, M. B., Wang, H., Abba, M., and Kazanietz, M. G. (2014). Protein kinase C and cancer: what we know and what we do not. *Oncogene* 33, 5225–5237. doi: 10.1038/onc.2013.524
- Golding, A. E., Visco, I., Bieling, P., and Bement, W. M. (2019). Extraction of active RhoGTPases by RhoGDI regulates spatiotemporal patterning of RhoGTPases. *eLife* 8:e50471.
- Ha, B. H., Davis, M. J., Chen, C., Lou, H. J., Gao, J., Zhang, R., et al. (2012). Type II p21-activated kinases (PAKs) are regulated by an autoinhibitory pseudosubstrate. *Proc. Natl. Acad. Sci. U.S.A.* 109, 16107–16112. doi: 10.1073/pnas.1214447109
- Hashimoto, I., Sakamaki, K., Oue, N., Kimura, Y., Hiroshima, Y., Hara, K., et al. (2019). Clinical significance of PRKCI gene expression in cancerous tissue in patients with gastric cancer. *Anticancer. Res.* 39, 5715–5720. doi: 10.21873/anticancer.13771
- Heikkilä, T., Wheatley, E., Crighton, D., Schroder, E., Boakes, A., Kaye, S. J., et al. (2011). Co-crystal structures of inhibitors with MRCKbeta, a key regulator of tumor cell invasion. *PLoS One* 6:e24825. doi: 10.1371/journal.pone.0024825
- Hirano, Y., Yoshinaga, S., Takeya, R., Suzuki, N. N., Horiuchi, M., Kohjima, M., et al. (2005). Structure of a cell polarity regulator, a complex between atypical PKC and Par6 PB1 domains. *J. Biol. Chem.* 280, 9653–9661.
- Ho, H., Aruri, J., Kapadia, R., Mehr, H., White, M. A., and Ganesan, A. K. (2012). RhoJ regulates melanoma chemoresistance by suppressing pathways that sense DNA damage. *Cancer Res.* 72, 5516–5528. doi: 10.1158/0008-5472.CAN-12-0775
- Hodge, R. G., and Ridley, A. J. (2017). Regulation and functions of RhoU and RhoV. *Small GTPases* 11, 1–8.
- Hofmann, C., Shepelev, M., and Chernoff, J. (2004). The genetics of Pak. *J. Cell Sci.* 117, 4343–4354.
- Hurst, C. D., Alder, O., Platt, F. M., Droop, A., Stead, L. F., Burns, J. E., et al. (2017). Genomic subtypes of non-invasive bladder cancer with distinct metabolic profile and female gender bias in KDM6A mutation frequency. *Cancer Cell* 32, 701.e7–715.e7.
- Isakov, N. (2018). Protein kinase C (PKC) isoforms in cancer, tumor promotion and tumor suppression. *Semin. Cancer Biol.* 48, 36–52. doi: 10.1016/j.semcancer.2017.04.012
- Jansen, S., Gosens, R., Wieland, T., and Schmidt, M. (2018). Paving the Rho in cancer metastasis: Rho GTPases and beyond. *Pharmacol. Ther.* 183, 1–21. doi: 10.1016/j.pharmthera.2017.09.002
- Jerrell, R. J., Leih, M. J., and Parekh, A. (2017). The ROCK isoforms differentially regulate the morphological characteristics of carcinoma cells. *Small GTPases* 11, 131–137.
- Julian, L., and Olson, M. F. (2014). Rho-associated coiled-coil containing kinases (ROCK): structure, regulation, and functions. *Small GTPases* 5:e29846. doi: 10.4161/sgtp.29846
- Justilien, V., Jameison, L., Der, C. J., Rossman, K. L., and Fields, A. P. (2011). Oncogenic activity of Ect2 is regulated through protein kinase C iota-mediated phosphorylation. *J. Biol. Chem.* 286, 8149–8157. doi: 10.1074/jbc.M110.196113
- Kale, V. P., Hengst, J. A., Desai, D. H., Amin, S. G., and Yun, J. K. (2015). The regulatory roles of ROCK and MRCK kinases in the plasticity of cancer cell migration. *Cancer Lett.* 361, 185–196. doi: 10.1016/j.canlet.2015.03.017
- Kale, V. P., Hengst, J. A., Desai, D. H., Dick, T. E., Choe, K. N., Colledge, A. L., et al. (2014). A novel selective multikinase inhibitor of ROCK and MRCK effectively blocks cancer cell migration and invasion. *Cancer Lett.* 354, 299–310. doi: 10.1016/j.canlet.2014.08.032
- Karasic, T. B., Chiorean, E. G., Sebti, S. M., and O'Dwyer, P. J. (2019). A phase I study of GGTI-2418 (geranylgeranyl transferase I inhibitor) in patients with advanced solid tumors. *Target Oncol.* 14, 613–618. doi: 10.1007/s11523-019-00661-5

- Kim, K. H., Chung, C., Kim, J. M., Lee, D., Cho, S. Y., Lee, T. H., et al. (2019). Clinical significance of atypical protein kinase C (PKC $\delta$  and PKC $\zeta$ ) and its relationship with yes-associated protein in lung adenocarcinoma. *BMC Cancer* 19:804. doi: 10.1186/s12885-019-5992-7
- Kjaer, S., Linch, M., Purkiss, A., Kostelecky, B., Knowles, P. P., Rosse, C., et al. (2013). Adenosine-binding motif mimicry and cellular effects of a thieno[2,3-d]pyrimidine-based chemical inhibitor of atypical protein kinase C isoenzymes. *Biochem. J.* 451, 329–342. doi: 10.1042/BJ20121871
- Knippler, C. M., Saji, M., Rajan, N., Porter, K., La Perle, K., and Ringel, M. D. (2019). MAPK- and AKT-activated thyroid cancers are sensitive to group I PAK inhibition. *Endocr. Relat. Cancer* 26, 699–712.
- Korobeynikov, V., Borakove, M., Feng, Y., Wuest, W. M., Koval, A. B., Nikonova, A. S., et al. (2019). Combined inhibition of Aurora A and p21-activated kinase 1 as a new treatment strategy in breast cancer. *Breast Cancer Res. Treat.* 177, 369–382. doi: 10.1007/s10549-019-05329-2
- Kumar, R., Mateo, J., Smith, A. D., Khan, K. H., Ruddle, R., Swales, K. E., et al. (2014). First-in-human, first-in-class phase 1 study of a novel oral multi-AGC kinase inhibitor AT13148 in patients (pts) with advanced solid tumors. *J. Clin. Oncol.* 32, 2554–2554.
- Lawson, C. D., and Ridley, A. J. (2018). Rho GTPase signaling complexes in cell migration and invasion. *J. Cell. Biol.* 217, 447–457.
- Leroux, A. E., Schulze, J. O., and Biondi, R. M. (2018). AGC kinases, mechanisms of regulation and innovative drug development. *Semin. Cancer Biol.* 48, 1–17. doi: 10.1016/j.semcancer.2017.05.011
- Leung, T., Chen, X. Q., Tan, I., Manser, E., and Lim, L. (1998). Myotonic dystrophy kinase-related Cdc42-binding kinase acts as a Cdc42 effector in promoting cytoskeletal reorganization. *Mol. Cell. Biol.* 18, 130–140.
- Lochhead, P. A., Wickman, G., Mezna, M., and Olson, M. F. (2010). Activating ROCK1 somatic mutations in human cancer. *Oncogene* 29, 2591–2598. doi: 10.1038/onc.2010.3
- Maldonado, M. D. M., and Dharmawardhane, S. (2018). Targeting Rac and Cdc42 GTPases in Cancer. *Cancer Res.* 78, 3101–3111. doi: 10.1158/0008-5472.CAN-18-0619
- Manser, E., Leung, T., Salihuddin, H., Zhao, Z. S., and Lim, L. (1994). A brain serine/threonine protein kinase activated by Cdc42 and Rac1. *Nature* 367, 40–46.
- Manser, E., Loo, T. H., Koh, C. G., Zhao, Z. S., Chen, X. Q., Tan, L., et al. (1998). PAK kinases are directly coupled to the PIX family of nucleotide exchange factors. *Mol. Cell.* 1, 183–192.
- Mansfield, A. S., Fields, A. P., Jatoti, A., Qi, Y., Adjei, A. A., Erlichman, C., et al. (2013). Phase I dose escalation study of the PKC $\delta$  inhibitor aurothiomalate for advanced non-small-cell lung cancer, ovarian cancer, and pancreatic cancer. *Anticancer Drugs* 24, 1079–1083. doi: 10.1097/CAD.0000000000000009
- Mileshkin, L. R., Rosen, L. S., Blumenkopf, T., Breazna, A., Darang, S., Davison, J., et al. (2011). Phase I, dose-escalation, safety, pharmacokinetic, and pharmacodynamic study of single-agent PF-03758309, an oral PAK inhibitor, in patients with advanced solid tumors. *J. Clin. Oncol.* 29:e13607.
- Mitin, N., Roberts, P. J., Chenette, E. J., and Der, C. J. (2012). Posttranslational lipid modification of Rho family small GTPases. *Methods Mol. Biol.* 827, 87–95. doi: 10.1007/978-1-61779-442-1\_6
- Mizutani, T., Haga, H., Koyama, Y., Takahashi, M., and Kawabata, K. (2006). Diphosphorylation of the myosin regulatory light chain enhances the tension acting on stress fibers in fibroblasts. *J. Cell. Physiol.* 209, 726–731.
- Montalvo-Ortiz, B. L., Castillo-Pichardo, L., Hernandez, E., Humphries-Bickley, T., De la Mota-Peynado, A., Cubano, L. A., et al. (2012). Characterization of EHop-016, novel small molecule inhibitor of Rac GTPase. *J. Biol. Chem.* 287, 13228–13238.
- Morgan-Fisher, M., Wewer, U. M., and Yoneda, A. (2013). Regulation of ROCK activity in cancer. *J. Histochem. Cytochem.* 61, 185–198. doi: 10.1369/0022155412470834
- Nakagawa, O., Fujisawa, K., Ishizaki, T., Saito, Y., Nakao, K., and Narumiya, S. (1996). ROCK-I and ROCK-II, two isoforms of Rho-associated coiled-coil forming protein serine/threonine kinase in mice. *FEBS Lett.* 392, 189–193.
- Nakashima, M., Adachi, S., Yasuda, I., Yamauchi, T., Kawaguchi, J., Hanamatsu, T., et al. (2011). Inhibition of Rho-associated coiled-coil containing protein kinase enhances the activation of epidermal growth factor receptor in pancreatic cancer cells. *Mol. Cancer* 10:79. doi: 10.1186/1476-4598-10-79
- Nayal, A., Webb, D. J., Brown, C. M., Schaefer, E. M., Vicente-Manzanares, M., and Horwitz, A. R. (2006). Paxillin phosphorylation at Ser273 localizes a GIT1-PIX-PAK complex and regulates adhesion and protrusion dynamics. *J. Cell Biol.* 173, 587–589.
- Ndubaku, C. O., Crawford, J. J., Drobnick, J., Aliagas, I., Campbell, D., Dong, P., et al. (2015). Design of selective PAK1 inhibitor G-5555: improving properties by employing an unorthodox low-pKa polar moiety. *ACS Med. Chem. Lett.* 6, 1241–1246. doi: 10.1021/acsmchemlett.5b00398
- Ng, Y., Tan, I., Lim, L., and Leung, T. (2004). Expression of the human myotonic dystrophy kinase-related Cdc42-binding kinase gamma is regulated by promoter DNA methylation and Sp1 binding. *J. Biol. Chem.* 279, 34156–34164.
- Nicholas, N. S., Pipili, A., Lesjak, M. S., Ameer-Beg, S. M., Geh, J. L., Healy, C., et al. (2016). PAK4 suppresses PDZ-RhoGEF activity to drive invadopodia maturation in melanoma cells. *Oncotarget* 7, 70881–70897. doi: 10.18632/oncotarget.12282
- Nindl, I., Dang, C., Forschner, T., Kuban, R. J., Meyer, T., Sterry, W., et al. (2006). Identification of differentially expressed genes in cutaneous squamous cell carcinoma by microarray expression profiling. *Mol. Cancer* 5:30.
- Ohashi, K., Nagata, K., Maekawa, M., Ishizaki, T., Narumiya, S., and Mizuno, K. (2000). Rho-associated kinase ROCK activates LIM-kinase 1 by phosphorylation at threonine 508 within the activation loop. *J. Biol. Chem.* 275, 3577–3582.
- Parker, P. J., Justilien, V., Riou, P., Linch, M., and Fields, A. P. (2014). Atypical protein kinase C $\delta$  as a human oncogene and therapeutic target. *Biochem. Pharmacol.* 88, 1–11. doi: 10.1016/j.bcp.2013.10.023
- Paul, A., Danley, M., Saha, B., Tawfik, O., and Paul, S. (2015). PKC $\zeta$  Promotes Breast Cancer Invasion by Regulating Expression of E-cadherin and Zonula Occludens-1 (ZO-1) via NF $\kappa$ B-p65. *Sci. Rep.* 5:12520. doi: 10.1038/srep12520
- Rane, C., Senapedis, W., Baloglu, E., Landesman, Y., Crochiere, M., Das-Gupta, S., et al. (2017). A novel orally bioavailable compound KPT-9274 inhibits PAK4, and blocks triple negative breast cancer tumor growth. *Sci. Rep.* 7:42555. doi: 10.1038/srep42555
- Rane, C. K., and Minden, A. (2019). P21 activated kinase signaling in cancer. *Semin. Cancer Biol.* 54, 40–49.
- Rath, N., Munro, J., Cutiongco, M. F., Jagiello, A., Gadegaard, N., McGarry, L., et al. (2018). Rho kinase inhibition by AT13148 blocks pancreatic ductal adenocarcinoma invasion and tumor growth. *Cancer Res.* 78, 3321–3336. doi: 10.1158/0008-5472.CAN-17-1339
- Rath, N., and Olson, M. F. (2012). Rho-associated kinases in tumorigenesis: reconsidering ROCK inhibition for cancer therapy. *EMBO Rep.* 13, 900–908. doi: 10.1038/embor.2012.127
- Ratnayake, W. S., Apostolatos, A. H., Ostrov, D. A., and Acevedo-Duncan, M. (2017). Two novel atypical PKC inhibitors; ACPD and DNDA effectively mitigate cell proliferation and epithelial to mesenchymal transition of metastatic melanoma while inducing apoptosis. *Int. J. Oncol.* 51, 1370–1382.
- Rodriguez-Hernandez, I., Cantelli, G., Bruce, F., and Sanz-Moreno, V. (2016). Rho, ROCK and actomyosin contractility in metastasis as drug targets. *F1000Research* 5:783.
- Rosse, C., Lodillinsky, C., Fuhrmann, L., Nourieh, M., Monteiro, P., Irondelle, M., et al. (2014). Control of MT1-MMP transport by atypical PKC during breast-cancer progression. *Proc. Natl. Acad. Sci. U.S.A.* 111, E1872–E1879. doi: 10.1073/pnas.1400749111
- Ruiz, R., Jahid, S., Harris, M., Marzese, D. M., Espitia, F., Vasudeva, P., et al. (2017). The RhoJ-BAD signaling network: an Achilles' heel for BRAF mutant melanomas. *PLoS Genet.* 13:e1006913. doi: 10.1371/journal.pgen.1006913
- Sanz-Moreno, V., Gadea, G., Ahn, J., Paterson, H., Marra, P., Pinner, S., et al. (2008). Rac activation and inactivation control plasticity of tumor cell movement. *Cell* 135, 510–523. doi: 10.1016/j.cell.2008.09.043
- Sari, I., Berberoglu, B., Ozkara, E., Oztuzcu, S., Camci, C., and Demiryurek, A. T. (2013). Role of rho-kinase gene polymorphisms and protein expressions in colorectal cancer development. *Pathobiology* 80, 138–145. doi: 10.1159/000341395
- Schurmann, A., Mooney, A. F., Sanders, L. C., Sells, M. A., Wang, H. G., Reed, J. C., et al. (2000). p21-activated kinase 1 phosphorylates the death agonist bad and protects cells from apoptosis. *Mol. Cell. Biol.* 20, 453–461.

- Schwarz, J., Proff, J., Hävemeier, A., Ladwein, M., Rottner, K., Barlag, B., et al. (2012). Serine-71 phosphorylation of Rac1 modulates downstream signaling. *PLoS One* 7:e44358. doi: 10.1371/journal.pone.0044358
- Scotti, M. L., Bamlet, W. R., Smyrk, T. C., Fields, A. P., and Murray, N. R. (2010). Protein kinase Ciota is required for pancreatic cancer cell transformed growth and tumorigenesis. *Cancer Res.* 70, 2064–2074. doi: 10.1158/0008-5472.CAN-09-2684
- Sebbagh, M., Hamelin, J., Bertoglio, J., Solary, E., and Breard, J. (2005). Direct cleavage of ROCK II by granzyme B induces target cell membrane blebbing in a caspase-independent manner. *J. Exp. Med.* 201, 465–471.
- Sebbagh, M., Renvoize, C., Hamelin, J., Riche, N., Bertoglio, J., and Breard, J. (2001). Caspase-3-mediated cleavage of ROCK I induces MLC phosphorylation and apoptotic membrane blebbing. *Nat. Cell Biol.* 3, 346–352.
- Semenova, G., and Chernoff, J. (2017). Targeting PAK1. *Biochem. Soc. Trans.* 45, 79–88. doi: 10.1042/BST20160134
- Semenova, G., Stepanova, D. S., Dubyk, C., Handorf, E., Deyev, S. M., Lazar, A. J., et al. (2017). Targeting group I p21-activated kinases to control malignant peripheral nerve sheath tumor growth and metastasis. *Oncogene* 36, 5421–5431. doi: 10.1038/onc.2017.143
- Senoo, H., Kamimura, Y., Kimura, R., Nakajima, A., Sawai, S., Sesaki, H., et al. (2019). Phosphorylated Rho-GDP directly activates mTORC2 kinase towards AKT through dimerization with Ras-GTP to regulate cell migration. *Nat. Cell Biol.* 21, 867–878. doi: 10.1038/s41556-019-0348-8
- Siekman, I. K., Dierck, K., Prall, S., Klokow, M., Strauss, J., Buhs, S., et al. (2018). Combined inhibition of receptor tyrosine and p21-activated kinases as a therapeutic strategy in childhood ALL. *Blood Adv.* 2, 2554–2567. doi: 10.1182/bloodadvances.2018020693
- Sorrell, F. J., Kilian, L. M., and Elkins, J. M. (2019). Solution structures and biophysical analysis of full-length group A PAKs reveal they are monomeric and auto-inhibited in cis. *Biochem. J.* 476, 1037–1051.
- Stallings-Mann, M., Jamieson, L., Regala, R. P., Weems, C., Murray, N. R., and Fields, A. P. (2006). A novel small-molecule inhibitor of protein kinase Ciota blocks transformed growth of non-small-cell lung cancer cells. *Cancer Res.* 66, 1767–1774.
- Sumi, T., Matsumoto, K., Shibuya, A., and Nakamura, T. (2001). Activation of LIM kinases by myotonic dystrophy kinase-related Cdc42-binding kinase alpha. *J. Biol. Chem.* 276, 23092–23096.
- Svensmark, J. H., and Brakebusch, C. (2019). Rho GTPases in cancer: friend or foe? *Oncogene* 38, 7447–7456.
- Talman, V., Gateva, G., Ahti, M., Ekokoski, E., Lappalainen, P., and Tuominen, R. K. (2014). Evidence for a role of MRCK in mediating HeLa cell elongation induced by the C1 domain ligand HMI-1a3. *Eur. J. Pharm. Sci.* 55, 46–57. doi: 10.1016/j.ejps.2014.01.002
- Tan, L., Ng, C. H., Lim, L., and Leung, T. (2001). Phosphorylation of a novel myosin binding subunit of protein phosphatase 1 reveals a conserved mechanism in the regulation of actin cytoskeleton. *J. Biol. Chem.* 276, 21209–21216.
- Tan, Y., Sementino, E., Chernoff, J., and Testa, J. R. (2017). Targeting MYC sensitizes malignant mesothelioma cells to PAK blockade-induced cytotoxicity. *Am. J. Cancer Res.* 7, 1724–1737.
- ten Klooster, J. P., Jaffer, Z. M., Chernoff, J., and Hordijk, P. L. (2006). Targeting and activation of Rac1 are mediated by the exchange factor beta-Pix. *J. Cell Biol.* 172, 759–769.
- Tobert, J. A. (2003). Lovastatin and beyond: the history of the HMG-CoA reductase inhibitors. *Nat. Rev. Drug Discov.* 2, 517–526.
- Truebestein, L., Elsner, D. J., Fuchs, E., and Leonard, T. A. (2015). A molecular ruler regulates cytoskeletal remodeling by the Rho kinases. *Nat. Commun.* 6:10029. doi: 10.1038/ncomms10029
- Tsang, T. Y., Wei, W., Itamochi, H., Tambouret, R., and Birrer, M. J. (2017). Integrated genomic analysis of clear cell ovarian cancers identified PRKCI as a potential therapeutic target. *Oncotarget* 8, 96482–96495. doi: 10.18632/oncotarget.19946
- Unbekandt, M., Belshaw, S., Bower, J., Clarke, M., Cordes, J., Crighton, D., et al. (2018). Discovery of potent and selective MRCK inhibitors with therapeutic effect on skin cancer. *Cancer Res.* 78, 2096–2114. doi: 10.1158/0008-5472.CAN-17-2870
- Unbekandt, M., Lilla, S., Zanivan, S., and Olson, M. F. (2019). The CDC42 effector protein MRCKbeta autophosphorylates on Threonine 1108. *Small GTPases* doi: 10.1080/21541248.2018.1564472 [Epub ahead of print].
- Unbekandt, M., and Olson, M. F. (2014). The actin-myosin regulatory MRCK kinases: regulation, biological functions and associations with human cancer. *J. Mol. Med.* 92, 217–225. doi: 10.1007/s00109-014-1133-6
- Upadhyaya, M., Spurllock, G., Thomas, L., Thomas, N. S., Richards, M., Mautner, V. F., et al. (2012). Microarray-based copy number analysis of neurofibromatosis type-1 (NF1)-associated malignant peripheral nerve sheath tumors reveals a role for Rho-GTPase pathway genes in NF1 tumorigenesis. *Hum. Mutat.* 33, 763–776. doi: 10.1002/humu.22044
- Vishnubhotla, R., Bharadwaj, S., Sun, S., Metlushko, V., and Glover, S. C. (2012). Treatment with Y-27632, a ROCK inhibitor, increases the proinvasive nature of SW620 cells on 3D collagen type 1 matrix. *Int. J. Cell Biol.* 2012:259142. doi: 10.1155/2012/259142
- Wei, L., Surma, M., Shi, S., Lambert-Cheatham, N., and Shi, J. (2016). Novel insights into the roles of Rho kinase in cancer. *Arch. Immunol. Ther. Exp.* 64, 259–278. doi: 10.1007/s00005-015-0382-6
- Wilkinson, S., Paterson, H. F., and Marshall, C. J. (2005). Cdc42-MRCK and Rho-ROCK signalling cooperate in myosin phosphorylation and cell invasion. *Nat. Cell Biol.* 7, 255–261.
- Wyckoff, J. B., Pinner, S. E., Gschmeissner, S., Condeelis, J. S., and Sahai, E. (2006). ROCK- and myosin-dependent matrix deformation enables protease-independent tumor-cell invasion in vivo. *Curr. Biol.* 16, 1515–1523.
- Yamada, K. M., and Sixt, M. (2019). Mechanisms of 3D cell migration. *Nat. Rev. Mol. Cell Biol.* 20, 738–752. doi: 10.1038/s41580-019-0172-9
- Yang, S., and Kim, H. M. (2014). ROCK inhibition activates MCF-7 cells. *PLoS One* 9:e88489. doi: 10.1371/journal.pone.0088489
- Yap, T. A., Walton, M. I., Grimshaw, K. M., Te Poele, R. H., Eve, P. D., Valenti, M. R., et al. (2012). AT13148 is a novel, oral multi-AGC kinase inhibitor with potent pharmacodynamic and antitumor activity. *Clin. Cancer Res.* 18, 3912–3923. doi: 10.1158/1078-0432.CCR-11-3313
- Yeo, D., He, H., Patel, O., Lowy, A. M., Baldwin, G. S., and Nikfarjam, M. (2016). FRAX597, a PAK1 inhibitor, synergistically reduces pancreatic cancer growth when combined with gemcitabine. *BMC Cancer* 16:24. doi: 10.1186/s12885-016-2057-z
- Yeo, D., Huynh, N., Beutler, J. A., Christophi, C., Shulkes, A., Baldwin, G. S., et al. (2014). Glaucaurubinone and gemcitabine synergistically reduce pancreatic cancer growth via down-regulation of P21-activated kinases. *Cancer Lett.* 346, 264–272. doi: 10.1016/j.canlet.2014.01.001
- Yoneda, A., Multhaupt, H. A., and Couchman, J. R. (2005). The Rho kinases I and II regulate different aspects of myosin II activity. *J. Cell Biol.* 170, 443–453.
- Yoo, H. Y., Sung, M. K., Lee, S. H., Kim, S., Lee, H., Park, S., et al. (2014). A recurrent inactivating mutation in RHOA GTPase in angioimmunoblastic T cell lymphoma. *Nat. Genet.* 46, 371–375. doi: 10.1038/ng.2916
- Zeng, R. J., Zheng, C. W., Gu, J. E., Zhang, H. X., Xie, L., Xu, L. Y., et al. (2019). RAC1 inhibition reverses cisplatin resistance in esophageal squamous cell carcinoma and induces downregulation of glycolytic enzymes. *Mol. Oncol.* 13, 2010–2030. doi: 10.1002/1878-0261.12548
- Zhao, Z., and Manser, E. (2015). Myotonic dystrophy kinase-related Cdc42-binding kinases (MRCK), the ROCK-like effectors of Cdc42 and Rac1. *Small GTPases* 6, 81–88. doi: 10.1080/21541248.2014.1000699
- Zihni, C., Vlassaks, E., Terry, S., Carleton, J., Leung, T. K. C., Olson, M., et al. (2017). An apical MRCK-driven morphogenetic pathway controls epithelial polarity. *Nat. Cell Biol.* 19, 1049–1060. doi: 10.1038/ncb3592

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Clayton and Ridley. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.