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Chapter

Clinical Manifestations in Pregnant Women and Congenital Abnormalities in Fetus and Newborns during a Zika Transmission Period in South Mexico

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Abstract

Dengue, Chikungunya and Zika are arboviruses transmitted by *Ae. aegypti* with significant public health impact. In the first trimester of 2015, autochthonous Zika transmission was reported in Mexico. The state of Yucatan is an endemic region where pregnant women with acute infection and related congenital abnormalities in fetus and newborns were observed. We describe results from a cohort of pregnant women and their babies followed up in Yucatan during the first Zika transmission outbreak (2016–2018). Clinical manifestations of acute ZIKV infection, persistence of viral RNA in pregnant women, as well as congenital abnormalities were observed. In addition, we describe the phenotype of newborns from confirmed or suspected ZIKV prenatal infection.

Keywords: Zika infection, pregnant women birth defects, pregnancy, congenital Zika syndrome

1. Introduction

Dengue (DEN), Chikungunya (CHIK) and Zika (ZIKV) are arboviruses transmitted by the mosquito *Aedes aegypti* and known as *Aedes*-borne-diseases. These diseases are associated with high morbidity and low mortality and considered a public health problem [1]. In the 2015, the ZIKV outbreak was considered an international emergency because infection in pregnant women was related to the increase of congenital abnormalities in the fetuses [2–4]. Vertical transmission of ZIKV was demonstrated by the RNA viral detection in placenta, amniotic fluid, serum and fetal brain in products with microcephaly, abortions or in autopsies of affected newborns and offspring of symptomatic or asymptomatic mothers [5, 6].

The clinical manifestations of Zika in general population and pregnant women were mild rash, conjunctivitis and low fever, although up to 80% remain asymptomatic, higher than DEN (19%) and CHIK (45%) [1, 2, 7, 8].

ZIKV is a Flavivirus with an Asian and African lineages [9, 10]. Its RNA genome (10.8 kb) encodes for a 3419-amino acid polyprotein which form a capsid (C), a membrane precursor (prM), a wrap (E) and seven non-structural proteins (NS1, NS2A, NS2B, NS3, NS4A, NS4B and NS5) [11]. ZIKV interferes with the neural development through decreased neural progenitor cells, arrest in neuronal migration and/or disruption of the maturation process of the fetus central nervous system (CNS) [12, 13]. The congenital Zika virus syndrome (CZVS) is a pattern of congenital defects associated with ZIKV infection during the pregnancy so ZIKV pathogenicity and virulence is currently studied [14–18].

The surveillance of ZIKV infection during pregnancy in endemic regions requires screening and detection of fetal morphological abnormalities [19]. An integrated intervention model for the prevention of Zika and *Aedes*-borne diseases, that includes primary health care services, gynecologists, obstetricians, pediatricians, geneticists and neurologists should be mandatory. Strategies to prevent and control the vectors and reduce the risk for diseases transmission should be strengthened, particularly for protection of women in reproductive ages [20, 21].

Here we report final the clinical manifestations observed in a cohort of pregnant women and the congenital abnormalities in fetus and newborns during a Zika transmission period (2016–2018) in South Mexico.

2. Methods

We developed a prospective study to quantify the incidence of disease and infection in a cohort of pregnant women and newborns during an epidemic period of Zika (2016–2018). One of the main objectives of the study was to know the effect of prenatal exposure to ZIKV. The cohort included pregnant women, preferably in the first trimester of pregnancy. The follow up included clinical and molecular detection of ZIKV, DEN and CHIK. Obstetric ultrasound was performed to recognize morphological abnormalities in the fetuses.

During the development of this study, information on health care was provided to pregnant women and their partners, highlighting the importance of family planning and the use of condoms as a method to prevent the transmission of ZIKV, in addition to the implementation of measures to prevent the breeding of the *Aedes aegypti* and mosquito bites at home. We also provided information about general healthy habits, family planning and prevention of sexually transmitted diseases.

2.1 Study population

After the informed consent was signed, women were interviewed, and their medical records including periconceptual and pregnancy history, were collected. The recruitment included 884 families (3993 people) from the cities of Merida, Ticul, and Progreso de Castro in the Yucatan State, South-east Mexico [16]. Merida and its metropolitan area (≈ 1 million inhabitants), comprises $\approx 50\%$ of the Yucatan population. Progreso de Castro (37,400) and Ticul (32,000) are smaller urban areas. We enrolled consenting pregnant women from these areas from July 1, 2016 to June 2018 including pregnant women referred by physicians in primary care facilities or hospitals within the areas of our cohort study.

2.2 Clinical follow-up of pregnant women

Patient monitoring included a monthly visit for clinical assessment and sample collection (blood and urine for RTC DEN/CHIK/ZIKV), weekly follow-up by text messages, and complete access to a telephone to report any clinical signs in pregnant women, their newborns, or any family contact. Tissues as umbilical cord blood, placental, amniotic fluid and breastmilk were collected, when possible for assessment of RT-PCR RNA ZIKV (TRIOPLEX) [22]. Other biochemical tests were performed to rule out Toxoplasma, Rubella, Cytomegalovirus, Herpes, Syphilis and HIV [23]. Depending of the mother or/and fetus risks, other test such as biochemical serum test and karyotype in amniotic fluid were taken. Ultrasound scanning was performed at the first contact and every 2 months. At the first visit, a questionnaire was fill up to establish the clinical-epidemiologic profile. The follow-up ended when the pregnancy was completed by delivery or fetal loss, or the participant withdrew from the study.

2.3 Clinical follow-up of newborns

Newborn follow up included: clinical evaluation (anthropometric measurements, APGAR score and physical exam) and sample collection for RT-PCR for DEN/CHIK/ZIKV [20–22, 24]. Patients were followed up for 24 months of life for early recognition of morphological anomalies and recorded the neurological development. These evaluations included genetic, neurologic, ophthalmologic, and audiologic evaluation. Microcephaly was defined as a cranial circumference ≥ 2 SDs below the mean for the age and sex of the baby, following the recommendation of the World Health Organization [25–28].

2.4 Patients not initially integrated into the cohort

At the time when the cohort study was ongoing, in the Genetic Service in the center of Investigations Dr. Hideyo Noguchi/UADY received pregnant women with morphological abnormalities detected in their fetuses, or newborn patients in whom ZIKV prenatal infection was suspected. These patients were not integrated to the cohort, but a clinical follow-up was granted. Sample collections for RT-PCR for DEN/CHIK/ZIKV were offered.

3. Results

3.1 Pregnant women cohort

The study integrated 130 pregnant women with average age of 25 years. No major differences in age distribution and socioeconomic status between ZIKV-positive and ZIKV-negative mothers were observed. Of all women, 40 (30%) were in the first trimester of pregnancy upon admission to the study, 62 (48%) in the second trimester and 28 (22%) in the third trimester (**Table 1**).

Positive results in *blood/urine* for ZIKV were found in 39 pregnant women, 31% (n = 13/39) at the first trimester, 52% (n = 20/39) in the second trimester and 15% (n = 6/39) in the third trimester.

Of 130 pregnant women, 39 (30%) were RNA-ZIKV positive at the time of the recruitment and 91 were negative (70%). From these, 11 (12% of initially RNA-ZIKV negative) became positive during the surveillance.

	Total pregnant women	Positive for ZIKV in blood/urine	Negative for ZIKV in blood/urine	Trimester of pregnancy when enrolled pregnant women (130)		
	N:130	N:39	N:91	1st N:40	2nd N:62	3rd N:28
Mérida	75 (58%)	27 (69%)	47 (52%)	16 (40%)	37 (60%)	22 (79%)
Progreso	4 (3%)	0	4 (4)	0	2 (3%)	2 (7%)
Ticul	51 (39%)	12 (31%)	41 (44%)	24 (60%)	23 (37%)	4 (14%)
15–19 years	29 (22%)	8 (28%)	21 (72%)	6 (15%)	16 (26%)	7 (25%)
20–29 years	70 (54%)	22 (31%)	48 (69)	22 (55%)	34 (55%)	15 (54%)
30–39 years	30 (23%)	9 (23%)	21 (%)	12 (30%)	12 (19%)	6 (21%)
40–49 years	1 (1%)	0	1 (%)	0	0	1
Signs and symptoms		21/39 (%)				
Exanthema		21/21 (100%)				
Pruritus		11/21 (52%)				
Joint edema		7/21 (33%)				
Conjunctivitis		16/21 (76%)				
Retro-orbital pain		10/21 (55%)				
Hyperemia		5/21 (22%)				
Clinical profile in persistence in serum samples		Total N:15	Symptomatic N:9	Asymptomatic		p
Age (years)		24.7 ± 4.4	24.2 ± 3.3	23.8 ± 6		0.874
Persistence (days) (interval)		45.93 ± 24.4 (17–19)	50.11 ± 30.58 (17–97)	39.67 ± 9.89 (29–52)		0.363
Pregnancy trimester						
First		6 (40%)	3	3		
Second		8 (53%)	6	2		
Third		1 (7%)		1		

Table 1. Summary of health, medical and laboratory data collected from pregnant women in the cohort.

Of the 28 symptomatic patients (negative or positive ZIKV), the most common symptoms were exanthema (75%), pruritus (39%) and conjunctivitis (57%). Of the 21 patients with RT-PCR ZIKV-positive, 75% were symptomatic and 25% were asymptomatic. Even so, more than half (64%) of the women had at least, more than one sign or symptom compatible with an *Aedes*-borne acute infection. Most prevalent symptoms were exanthema (100%), conjunctivitis (76%), pruritus (52%) headache (50%), retro-orbital pain (55%), arthralgia (33%), hyperemia (22%) and joint edema (6%). No hemorrhagic or systemic complications were observed in any patient (**Table 1**). Differences in the distribution of ZIKV-positive vs. ZIKV-negative women between the studied cities were not founded.

3.2 Persistence of ZIKV viral RNA in serum and urine in pregnant women

Of the 39 ZIKV-positive pregnant women, persistent RNA-ZIKV was detected in 38.5% (15/39) of the patients during 14 days after the initial symptoms or the last

PCR detection. Of these, six women were in the first trimester of gestation, eight in the second and one in the third. Within the group of symptomatic women (n = 28), nine (9/28) presented persistence of RNA-ZIKV, three in the first trimester and six

Newborn residence	Merida N:67		Progreso N:2		Ticul N:46		Total births N:115
Percentile	Percentile <3 N (%)		Percentile 3–97 N (%)		Percentile >97 N (%)		Total
	At birth	End of the study	At birth	End of the study	At birth	End of the study	
Birth weight percentiles	5 (4)	1 (1)	110 (96)	114 (99)	0 (0)	0 (0)	115 (100)
Birth height percentiles	3 (3)	0 (0)	112 (97)	115 (100)	0 (0)	0 (0)	115 (100)
Birth head circumference percentiles	2 (2)	1 (1)	112 (97)	114 (99)	0 (0)	0 (0)	115 (100)
Newborn evaluations							
	From mothers ZIKV negative during pregnancy N:80			From mothers ZIKV positive during pregnancy N:35			
	N (%)		Alterations N (%)		N (%)		Alterations N (%)
Newborn blood	29 (36)				27 (77)		
Newborn urine	32 (40)				14 (40)		
Pediatrics	39 (49)				30 (86)		
Ophthalmology	19 (24)				31 (89)		3 (10)
Auditory screening	51 (64)				30 (86)		2 (7)
Placenta	NA		NA		17 (49)		
Umbilical cord	NA		NA		17 (49)		
Genetics	NA		NA		35 (100)		
Metabolic screening	NA		NA		34 (97)		
Transfontanelar and abdominal ultrasound	NA		NA		22 (63)		
Age of newborns who completed the study							
Age	Mother ZIKV –		Mother ZIKV +		Total		
1–6 months	16 (22%)		0		16 (16%)		
7–12 months	25 (34%)		3 (10%)		28 (27%)		
13–18 months	20 (27%)		17 (59%)		37 (36%)		
19–23 months	12 (17%)		9 (31%)		21 (21%)		
Total	73		29		102		

Table 2.
Newborn cohort follow-up.

Patient/ sex	Mother Zika symptoms (MoG ¹)	WoG ²	Neonatal head circumference (z score)	Birth weight for gestational age	Phenotype	Intracranial calcification	Zika IgM InBios newborn/ mother ⁶	Final diagnosis
1/Male	No	38	Microcephaly (-3)	Small	Microcephaly MMC³	Yes	-/-	CZS/neural tube defect
2/Male	Yes (2)	37	Normocephaly (10)	Small	IUGR ⁴	NR ⁵	-/-	IUGR
3/Male	No	38	Microcephaly (-3)	Appropriated	Microcephaly	Yes	-/-	CZS
4/Male	Yes (2)	37	Normocephaly (3)	Small	Asymptomatic	NR	-/-	IgM positive for toxoplasma
5/Male	No	33	Normocephaly (10)	Appropriated	Microcephaly Arthrogyposis	Yes	Positive in serum/-	CZS
6/Male	No	39	Microcephaly (-3)	Appropriated	Microcephaly	Yes	-/-	CZS
7/Female	Yes (2)	40	Normocephaly (3)	Small	Postnatal microcephaly	NR	-/-	CZS
8/Female	No	38	Macrocephaly (97)	Appropriated	Macrocephaly Arthrogyposis	Normal	-/-	Amyoplasia congenital

¹Month of gestation.

²Weeks of gestation.

³Myelomeningocele.

⁴Intrauterine growth retardation.

⁵NR: not reported.

⁶Serum and urine were taken at born in newborns and theirs mothers.

Table 3.
Main findings of eight patients evaluated for suspected ZIKV prenatal infection.

in the second. Of these, 100% (9/9) presented rash, 55% (5/9) conjunctivitis and 33.3% (2/9) fever. Only 22.2% (2/28) reported having three symptoms, fever, rash and conjunctivitis (**Table 1**).

3.3 Newborn evaluations

All pregnancies of women in the cohort have ended the pregnancy. Two fetal losses (one in the first trimester and one in the third trimester) occurred in ZIKV-negative mothers. Of all newborns alive, 3% were preterm (two in ZIKV-negative mothers and one in a ZIKV-positive moth group). No newborns or products of conception were positive for RNA-ZIKV. Clinical evaluation of Apgar scores in the newborns did not show significant differences between positive/negative ZIKV mothers. Also, microcephaly was not founded in any newborn. One newborn of Zika-positive mother died the first days of life due to gastroschisis (**Table 2**).

During the ZIKV epidemiological period in Yucatan, 10 patients attended the medical genetics clinic in CIR Hideyo Noguchi with a reference diagnosis of microcephaly, arthrogryposis and/or ventriculomegaly. Two of them were excluded from the follow up because ZIKV prenatal symptoms were absent in the mothers and microcephaly and intracranial calcifications were discarded in the patients. Preconceptional, prenatal and perinatal backgrounds were investigated in all patients. Only one woman took folic acid 6 months before the conception and none used mosquito repellent during pregnancy, even they were living in an endemic region.

Of the newborns evaluated, clinical symptoms of Zika were reported only in two mothers in the first trimester of gestation, but only one was tested positive for RT-PCR Zika. One more woman reported symptoms of Zika 1 month before conception. During the pregnancy, intrauterine growth restriction in 4/8 (50%) and oligohydramnios in 2/8 (25%) were reported. Only one patient was reported prenatally with microcephaly, intracranial calcifications and ventriculomegaly (patient 5) (**Table 3**). Five pregnancies were ended by caesarean section and three by vaginal delivery between second semester of 2016 and the second semester of 2017.

All newborns were at term, except one. Of them, six were males and two were females. Three males had microcephaly with less than three standard deviations and one male with microcephaly less than two standard deviations. One female had macrocephaly. All babies from the three mothers with positive ZIKV symptoms during or before pregnancy were born with normocephaly, but were small for gestational age. One of these babies, a female, developed microcephaly within the first 6 months of life. Of all babies follow up, two had arthrogryposis, one with microcephaly and one with macrocephaly. Both of them were child from non-symptomatic mothers. Only five patients were evaluated with axial computerized tomography with positive intracranial calcifications founded in four. All of them were from asymptomatic mothers.

All patients and their mothers were tested for RT-PCR for ZIKV/DEN/CHIK in serum and urine. TORCH was also performed. RNA-ZIKV was obtained only in serum of one male with microcephaly, arthrogryposis and intracranial calcifications. This male was from an asymptomatic mother. Antibodies IgM of toxoplasma were detected in one asymptomatic male whom has clinical symptoms of Zika during first trimester of pregnancy.

4. Discussion

The WHO declared the ZIKV outbreak in South America and the associated increase in neurological disorders and neonatal malformations a “*public health*

emergency of international concern” [2], and the CDC issued the ZIKV epidemiological alert, recommendations of high-risk Mexican territories was recognized by the National Health Service. Even so, the recognition of the ZIKV infection symptoms by patients and health workers, were underestimated [29]. In Mexico, first patients with ZIKV infection were documented early in 2016 even so, captured mosquitoes in early 2015 were recognized with ZIKV infection [30].

After the observed relationship between prenatal Zika infection and the risk of congenital defects in Brazil and Colombia, we realized a cohort with detailed evaluation of pregnant women searching ZIKV infection and congenital abnormalities [1, 31, 32].

The women in this study with the highest proportion of symptoms with ZIKV were those from 20 to 29 years, different from other studies that have reported the highest symptomatic disease ratio among women >30 years of age [33, 34]. In our studies this can be explained because the highest incidence of pregnancies in women was aged 20–29 years [1]. In clinically affected women, univariate analyzes showed that the most sensitive clinical sign was the exanthema, but it was also the least specific. The triad, conjunctival hyperemia, joint edema and exanthema had the highest level of specificity [35].

In pregnant women with exanthema without fever or other symptoms should to suspect ZIKV infection. In this study, ZIKV/DEN/CHIK co-infection was not identified neither [36].

The persistence of RNA viral in pregnant women has been described previously, reported a patient positive for RNA-ZIKV until 10 weeks after the onset of symptoms. In this study, the maximum viremia persistence was 97 days, but a quarter of positive mothers had viremia for more than 8 weeks [37]. From all RNA ZIKV-positive mothers, only 50% of the symptomatic group had persistence of ZIKV for more than 8 weeks. Other series reported longer RNA ZIKV viremia in symptomatic women than asymptomatic and other study obtained similar results in serum, in a range of 14–63 days; although in this study, we detected two pregnant women who had viremia older than 90 days [33, 34]. A study reported viral persistence in five pregnant women, of which 80% were symptomatic and only one case was asymptomatic. In this study, a higher percentage of asymptomatic viral persistence cases were found with 40%. In the symptomatic group, the average number of days of persistence after the onset of symptoms was greater than that reported, no relationship was observed between viral persistence and the presence of abnormalities in pregnancy products [38, 39].

The confirmatory tests which provide evidence for prenatal infection by ZIKV were conducted based on tests in the mother and the newborn were made with viral RNA isolated from biological fluids and placental tissues performed with the Trioplex kit of the CDC. The ZIKV genome was detected in cerebrospinal fluid, cardiac fluid, chorionic villi, fetal face of the placenta, serum and urine [26]. Vertical transmission studying the placental and fetus tissue also failed been demonstrated [40, 41].

Clinical variability in patients in whom ZIKV infection were prenatally suspected was described previously, in this cohort, congenital abnormalities associated to ZIKV were not observed in fetus, stillborn or newborns [1, 17]. Only one patient with gastroschisis was detected but the prenatal ZIKV infection in her mother was after detection of this abnormality.

Congenital Zika syndrome (CSZ) was observed in the patients evaluated from the Genetics Service out of the initial cohort. In them, Zika symptoms in mothers, microcephaly, arthrogryposis and intracranial calcifications were observed as in other reports [1, 17, 18]. Establishing the final diagnosis of ZIKV prenatal infection was difficult especially in asymptomatic mothers or in mild affected babies [31].

In two patients, the final diagnosis was toxoplasma and congenital amyoplasia. Asymptomatic or mild symptomatic women would be a serious limitation from early ZIKV infection. To define diagnostic of prenatal ZIKV is important: (1) to establish the follow up of the affected patient, (2) to limit the tests related to others genetic diseases that share symptoms with CSZ and (3) to establish the specific risk of recurrences of congenital abnormalities in subsequent pregnancies in the mother.

5. Conclusions

In this study, symptomatic or asymptomatic pregnant women with a high prevalence for ZIKV are reported, however there was no positive newborn or with malformations associated with ZIKV, despite the genetic consultation, the presence of CSZ, laboratory-confirmed if present so doctors should maintain a realistic perspective of the impact of ZIKV on pregnancy. And despite the uncertainties, future mothers should receive adequate, systematic advice and the best planned obstetric surveillance, always considering the gestational moment of maternal ZIKV infection and accept that there is gestational risk for different elements of the CZS phenotype in risk areas ZIKV transmission.

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References

- [1] Brasil P, Pereira JP, Raja Gabaglia C, Damasceno L, Wakimoto M, Ribeiro Nogueira RM, et al. Zika virus infection in pregnant women in Rio de Janeiro—Preliminary report. *The New England Journal of Medicine*. 2016. DOI: 10.1056/NEJMoa1602412
- [2] Reynolds MR, Jones AM, Petersen EE, Lee EH, Rice ME, Bingham A, et al. Vital signs: Update on Zika virus-associated birth defects and evaluation of all U.S. infants with congenital Zika virus exposure—U.S. Zika pregnancy. *MMWR*. 2017;**66**:1-9
- [3] Venancio FA, Quilião Bernal ME, Vieira Ramos MCB, Rocha Chaves N, Hedges MV, Marques Rodrigues de Souza M, et al. Congenital Zika syndrome in a Brazil-Paraguay-Bolivia border region: Clinical features of cases diagnosed between 2015 and 2018. *PLoS ONE*. 2019;**14**(10):e0223408. DOI: 10.1371/journal.pone.0223408
- [4] World Health Organization. Zika situation report. [Accessed: 5 February 2016]. Available at: <http://www.who.int/emergencies/zika-virus/situation-report/5-february-2016/en/>
- [5] Besnard M, Lastère S, Teissier A, Cao-Lormeau VM, Musso D. Evidence of perinatal transmission of Zika virus, French Polynesia, December 2013 and February 2014. *Euro Surveill*. 2014;**19**:1-4
- [6] Calvet G, Aguiar RS, Melo ASO, Sampaio SA, de Filippis I, Fabri A, et al. Detection and sequencing of Zika virus from amniotic fluid of fetuses with microcephaly in Brazil: A case study. *The Lancet Infectious Diseases*. 2016;**16**:653-660
- [7] Rojas DP, Barrera-Fuentes GA, Pavia-Ruz N, Salgado-Rodriguez M, Che-Mendoza A, Manrique-Saide P, et al. Epidemiology of dengue and other arboviruses in a cohort of school children and their families in Yucatan, Mexico: Baseline and first year follow-up. *PLoS Neglected Tropical Diseases*. 2018;**12**(11):e0006847. DOI: 10.1371/journal.pntd.0006847
- [8] Pavía-Ruz N, Villanueva S, Granja P, Balam-May A, Longini IM, Halloran ME, et al. Seroprevalence of dengue antibodies in three urban settings in Yucatan, Mexico. *American Journal of Tropical Medicine and Hygiene*. 2018;**98**:1202-1208. DOI: 10.4269/ajtmh.17-0382
- [9] Haddow AD, Schuh AJ, Yasuda CY, Kasper MR, Heang V, Huy R, et al. Genetic characterization of Zika virus strains: Geographic expansion of the Asian lineage. *PLoS Neglected Tropical Diseases*. 2012;**6**(2):e1477. DOI: 10.1371/journal.pntd.0001477
- [10] Rinckenberger N, Schoggins JW. Comparative analysis of viral entry for Asian and African lineages of Zika virus. *Virology*. 2019;**533**:59-67
- [11] Xing H, Xu S, Jia F, et al. Zika NS2B is a crucial factor recruiting NS3 to the ER and activating its protease activity. *Virus Research*. 2020;**275**:197793. DOI: 10.1016/j.virusres.2019.197793
- [12] Annamalai AS, Pattnaik A, Sahoo BR, Muthukrishnan E, Natarajan SK, Steffen D, et al. Zika virus encoding nonglycosylated envelope protein is attenuated and defective in neuroinvasion. *Journal of Virology*. 2017;**91**:e01348-e01317. DOI: 10.1128/JVI.01348-17
- [13] Leda AR, Bertrand L, Andras IE, El-Hage N, Nair M, Toborek M. Selective disruption of the blood-brain barrier by Zika virus. *Frontiers in Microbiology*. 2019;**10**:2158. DOI: 10.3389/fmicb.2019.02158

- [14] Beys-da-Silva WO, Rosa RL, Santi L, Berger M, Park SK, Campos AR, et al. Zika virus infection of human mesenchymal stem cells promotes differential expression of proteins linked to several neurological diseases. *Molecular Neurobiology*. 2019;**56**(7):4708-4717. DOI: 10.1007/s12035-018-1417-x
- [15] Shao Q, Herrlinger S, Yang S-L, Lai F, Moore JM, Brindley MA, et al. Zika virus infection disrupts neurovascular development and results in postnatal microcephaly with brain damage. *Development*. 2016;**143**(22):4127-4136. DOI: 10.1242/dev.143768
- [16] Acosta-Reyes J, Navarro E, Herrera MJ, Goenaga E, Ospina ML, Parra E, et al. Severe neurologic disorders in 2 fetuses with Zika virus infection, Colombia. *Emerging Infectious Diseases*. 2017;**23**:982-984. DOI: 10.3201/eid2306.161702
- [17] Del Campo M, Feitosa IM, Ribeiro EM, Horovitz DD, Pessoa AL, França GV, et al. The phenotypic spectrum of congenital Zika syndrome. *American Journal of Medical Genetics*. 2017;**173**:841-857. DOI: 10.1002/ajmg.a.38170
- [18] Moore CA, Stables JE, Dobyans WB, Pessoa A, Ventura CV, Fonseca EB, et al. Characterizing the pattern of anomalies in congenital Zika syndrome for pediatric clinicians. *JAMA Pediatrics*. 2017;**171**(3):288-2995. DOI: 10.1001/jamapediatrics.2016.3982
- [19] Lebov JF, Arias JF, Balmaseda A, Britt W, Cordero JF, Galvão LA, et al. International prospective observational cohort study of Zika in infants and pregnancy (ZIP study): Study protocol. *BMC Pregnancy and Childbirth*. 2019;**19**:282. DOI: 10.1186/s12884-019-2430-4
- [20] Romer Y, Valadez-Gonzalez N, Contreras-Capetillo S, Manrique-Saide P, Vazquez-Prokopec G, Pavia-Ruz N. Zika virus infection in pregnant women, Yucatan, Mexico. *Emerging Infectious Diseases*. 2019;**25**:1452-1460
- [21] Pavía-Ruz N, Contreras-Capetillo S, Valadez-González N, Villegas-Chim J, Carcaño-Castillo R, Valencia-Pacheco G, et al. Current Topics in Zika: An Integrated Intervention Model for the Prevention of Zika and Other Aedes-Borne Diseases in Women and Their Families in Mexico. IntechOpen; 2018. ISBN: 978953.51-0317-2
- [22] Colombo TE, Versiani AF, Dutra KR, et al. Performance of CDC Trioplex qPCR during a dengue outbreak in Brazil. *Journal of Clinical Virology*. 2019;**121**:104208. DOI: 10.1016/j.jcv.2019.104208
- [23] Oduyabo T, Petersen EE, Rasmussen AA, Mead PS, Meaney-Delman D, Renquist CM, et al. Update: Interim guidelines for health care providers caring for pregnant women and women of reproductive age with possible Zika virus exposure—United States, 2016. *MMWR*. 2016;**65**(5):122-127
- [24] Casey BM, McIntire DD, Leveno KJ. The continuing value of the Apgar score for the assessment of newborn infants. *The New England Journal of Medicine*. 2001;**344**:46771. DOI: 10.1056/NEJM200102153440701
- [25] Lanciotti RS, Kosoy OL, Laven JJ, Velez JO, Lambert AJ, Johnson AJ, et al. Genetic and serologic properties of Zika virus associated with an epidemic, Yap State, Micronesia, 2007. *Emerging Infectious Diseases*. 2008;**4**:1232-1239. DOI: 10.3201/eid1408.080287
- [26] Rabe IB, Staples JE, Villanueva J, Hummel KB, Johnson JA, Rose L, et al. Interim guidance for interpretation of Zika virus antibody test results. *MMWR. Morbidity and Mortality*

Weekly Report. 2016;**65**:543-546. DOI: 10.1555_85/mmwr.mm6521e1

[27] Bhatnagar J, Rabeneck DB, Martines RB, Reagan-Steiner S, Ermias Y, Estetter B, et al. Zika virus RNA replication and persistence in brain and placental tissue. *Emerging Infectious Diseases*. 2017;**23**:405-414. DOI: 10.3201/eid2303.161499

[28] Centers for Disease Control and Prevention. Revised diagnostic testing for Zika, Chikungunya, and dengue viruses in US public health laboratories. [Cited: 31 August 2017]. Available from: <https://www.cdc.gov/zika/transmission/index.html>

[29] Flemming-Dutra KE, Nelson JM, Fischer M, Staples E, Karwowski MP, Mead O, et al. Update: Interim guidelines for health care provider caring for infants and children with possible Zika virus infection-United States, February 2016. *MMWR*. 2016;**65**:1-6

[30] Díaz-Quinonez JA, López-Martínez I, Torres-Longoria B, Vázquez-Pichardo M, Cruz-Ramírez E, Ramírez-González JE, et al. Evidence of the presence of the Zika virus in Mexico since early 2015. *Virus Genes*. 2016;**52**(6):855-857

[31] Contreras-Capetillo SN, Valadéz-González N, Manrique-Saide P, Carcaño-Castillo RE, Pacheco-Tugores F, Barrera-Pérez HAM, et al. Birth defects associated with congenital Zika virus infection in Mexico. *Clinical Pediatrics*. 2017;**10**:1-10

[32] Alvarado-Socarras JL, Ocampo-González M, Vargas-Soler JA, Rodríguez-Morales AJ, Franco-Paredes C. Congenital and neonatal chikungunya in Colombia. *Journal of the Pediatric Infectious Diseases Society*. 2016;**5**:e17-e20

[33] Duffy MR, Chen T-H, Hancock WT, Powers AM, Kool JL, Lanciotti RS,

et al. Zika virus outbreak on Yap Island, Federated States of Micronesia. *The New England Journal of Medicine*. 2009;**360**:2536-43. DOI: 10.1056/NEJMoa0805715

[34] Flamand C, Fritzell C, Matheus S, Dueymes M, Carles G, Favre A, et al. The proportion of asymptomatic infections and spectrum of disease among pregnant women infected by Zika virus: Systematic monitoring in French Guiana, 2016. *Euro Surveill*. 2017;**22**. DOI: 10.2807/1560-7917.ES.2017.22.44.17-00102

[35] Nicastrì E, Pisapia R, Corpolongo A, Fusco FM, Cicalini S, Scognamiglio P, et al. Three cases of Zika virus imported in Italy: Need for a clinical awareness and evidence-based knowledge. *BMC Infectious Diseases*. 2016;**16**:66960

[36] Vogels CBF, Rückert C, Cavany SM, Perkins TA, Ebel G, Grubaugh ND. Arbovirus coinfection and co-transmission: A neglected public health concern? *PLoS Biology*. 2019;**17**(1):e3000130. DOI: 10.1371/journal.pbio.3000130

[37] Salehuddin AR, Haslan H, Mamikutty N, Zaidun NH, Azmi MF, Senin MM, et al. Zika virus infection and its emerging trends in Southeast Asia. *Asian Pacific Journal of Tropical Medicine*. 2017;**10**(3):211-219

[38] Driggers RW, Korhonen EM, Kuivanen S, Jääskeläinen AJ, Smura T, Rosenberg A, et al. Zika virus infection with prolonged maternal viremia and fetal brain abnormalities. *The New England Journal of Medicine*. 2016. DOI: 10.1056/NEJMoa1601824

[39] Meaney-Delman D, Oduyebo T, Polen KN, White JL, Bingham AM, Slavinski SA, et al. Prolonged detection of Zika virus RNA in pregnant women. *Obstetrics and Gynecology*. 2016;**128**(4):724-730

[40] Paz-Bailey G, Rosenberg ES, Doyle K, Munoz-Jordan J, Santiago GA, Klein L, et al. Persistence of Zika virus in body fluids—Final report. *The New England Journal of Medicine*. 2017;**379**(13):1234-1243. DOI: 10.1056/NEJMoa1613108

[41] Besnard M, Lastère S, Teissier A, Cao-Lourmeau VM, Musso D. Evidence of perinatal transmission of Zika virus, French Polynesia, December 2013 and February 2014. *Euro Surveill*. 2014;**19**(13):1-4

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