the world's leading publisher of Open Access books Built by scientists, for scientists

4,800

Open access books available

122,000

International authors and editors

135M

Downloads

154

TOP 1%

Our authors are among the

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.

For more information visit www.intechopen.com



Soybean Amino Acids in Health, Genetics, and Evaluation

William Monte Singer, Bo Zhang, M.A. Rouf Mian and Haibo Huang

Additional information is available at the end of the chapter

Abstract

Soybean is an important source of protein and amino acids for humans and livestock because of its well-balanced amino acid profile. This chapter outlines the strengths and weaknesses of soybean as a complete amino acid source as well as the relative importance of individual amino acids. Special attention is paid to the sulfur-containing amino acids, methionine and cysteine. Breeding and genetic engineering efforts are summarized to highlight previous accomplishments in amino acid improvement and potential avenues for future research. Agronomic properties and processing methods that affect amino acid levels in soybean food and feed are also explained. A brief introduction into current amino acid evaluation techniques is provided. By understanding the complexities of amino acids in soybean, protein quality for humans and livestock can be maximized.

Keywords: essential and nonessential amino acids, soybean meal, methionine, cysteine, sulfur-containing amino acids

1. Introduction

Soybean is one of the world's most economically and nutritionally important crops. In 2018, soybean were 61% of international oilseed production with 397.9 tons harvested worldwide [1]. The United States and Brazil were the largest producers at 4545 and 4299 million bushels, respectively, with China being the largest importer of U.S. whole soybeans valued over \$3 billion U.S. [2, 3]. Soybean products, namely meal and oil, are popular in a myriad of industries for their versatility and utility. Soybean oil provides the most versatility with uses in fuel, solvents, candles, cosmetics, construction, and foam. However, soybean meal is the driving



factor for 70% of the plant's value with 97% of all U.S. soybean meal being used for animal feed [4]. As such, an enormous portion of soybean's importance lies with its nutritional capabilities for livestock and humans.

Nutritionally speaking, soybeans are a highly valued protein source. Proteins are a crucial macromolecule needed in the diets of human and livestock. However, the true significance of soybean protein is due to its well-balanced amino acid profile that aligns with dietary needs of humans and animals [5]. Amino acids are the functional subunits of proteins that, when linked together in different orders, generate the variety of proteins critical to life. Amino acids are also important intermediates for many biosynthesis pathways [6]. Deficiencies in single or multiple amino acids can negatively impact an individual's growth and development [7, 8]. Intriguingly, an excess of certain amino acids has also been shown to worsen feed intake, nitrogen efficiency, and growth rate in livestock [9–11]. The importance of amino acid levels on human health has also been well documented [12, 13].

Amino acids are characterized by having amine (-NH₂) and carboxyl (-COOH) functional groups as well as a "R-group" that is unique to each amino acid [14]. Amino acids are abundant in both proteinogenic (protein-incorporated) and non-proteinogenic forms [15]. The 20 common, proteinogenic amino acids are generally the focus of research in soybeans as they are the defining nutritive feature. Of those twenty, nine amino acids are essential for humans to consume. Livestock usually require these same amino acids from feed and might require others because of their biological systems [16]. Soybeans contain some level of all nine essential amino acids which creates a suitable nutritional foundation for livestock feed and human food.

2. Essential amino acids

Essential amino acids are ones that living organisms are unable to biosynthesize themselves and must obtain from their food source [7, 12, 16]. Therefore, in this term, "essential" refers to the amino acid requirements in dietary ingredients. The nine standard essential amino acids for humans present in soybean are: histidine, isoleucine, leucine, lysine, methionine, phenylalanine, threonine, tryptophan, and valine [17]. Arginine is regularly considered an essential amino acid for fish, poultry and sometimes swine due to absent or deficient urea cycles [9, 16]. Poultry and reptiles also require dietary glycine because of differing waste excretion pathways [16]. While crude protein content is normally recognized as the driving nutritional factor for soybean meal, these essential amino acids provide true utility.

It has long been recommended that protein quality is based upon essential amino acid content. However, for many reasons, animal feed and human food markets have only recently begun assessing accordingly [5, 18, 19]. Equipment required for accurate amino acid measurement and the diversity of markets for amino acids makes it difficult for supply chain evaluators like elevator operators to appraise amino acid content on site. To some degree, the well-balanced soybean amino acid profile also devalues the need to measure individual amino acid levels. Since all essential amino acids are present, less attention is paid to deficient amino acids such as methionine and tryptophan [17, 20].

Deficiencies in soybean's essential amino acid profile has led to a large section of the livestock industry focusing on feed mixing and supplementation. Rationing with other feed sources such as cereal grain and synthetic amino acid augmentation can effectively resolve the issue. Although, this comes with economic and environmental problems. Supplementing amino acids adds costs to farmers. For example, the average cost for amino acids supplementation for dairy farmers is 20 cents per head per day [21]. Maximizing crude protein for a growth limiting factor also negatively impacts livestock nitrogen-use-efficiency and environmental nitrogen outputs [8, 22]. Synthetic amino acid production can produce hazardous environmental waste and synthetic methionine, the most limiting soybean amino acid for poultry, has also been banned for organic poultry production [20, 23]. Movement towards sustainable agriculture will pressure the feed industry to alter how soybean meal is enhanced for essential amino acid livestock maximization. Furthermore, the increasing popularity of meat-less diets in humans will create new markets for soybean's well-balanced amino acid profile.

3. Nonessential amino acids

Nonessential amino acids should not be misconstrued as unimportant amino acids. Of the 20 proteinogenic amino acids, those considered nonessential are still necessary for living organisms. Healthy organisms are able to biosynthesize them and are not required from food and feed consumption. The 20 standard nonessential amino acids for humans found in soybean are: arginine, alanine, asparagine, aspartic acid, cysteine, glutamine, glutamic acid, glycine, proline, serine, and tyrosine [24-26]. As previously mentioned, the necessity of amino acids such as arginine and glycine can differ amongst species. Some nonessential amino acids are also affected by the presence and amounts of essential amino acids.

Cysteine not provided from food consumption is directly biosynthesized from methionine via trans-sulfuration [16, 27–30]. Consequently, if cysteine is not provided in the diet, then enough methionine must be provided to compensate for both amino acid needs. For that reason, feed research for poultry occasionally measures methionine and cysteine jointly [10, 31, 32]. Tyrosine is also directly formed from phenylalanine via hydroxylation [16, 29, 33]. Other amino acids like arginine, glycine, and proline can be required from the diet when animals are young, old, sick, or otherwise deficient in body protein regulation. As human and livestock diets become more sustainably plant-based, it will become more important to evaluate nonessential amino acids, specifically the ones immediately affected by essential amino acids.

4. Proteinogenic sulfur-containing amino acids

The two proteinogenic sulfur-containing amino acids, methionine and cysteine, are critical to evaluating soybean meal as food and feed. While present in soybean, methionine and cysteine levels are both inadequate for consumer needs [8, 20, 24]. Similar to research determinations, the nutritional requirements for methionine and cysteine intake are often grouped together as overall sulfur consumption from protein. Adult humans are recommended to intake 910–1120 mg methionine and cysteine (based on body weight) per day [13]. For livestock, sulfur amino acids recommendations can vary based on species, age, end-use, and diet formulation. The importance of dietary sulfur amino acids for livestock is greatly emphasized in literature, especially with soybean meal as base feed [8, 20, 34, 35].

Methionine and cysteine are vital to biological functions because of the sulfur contained in their R-groups and versatility in macromolecule synthesis. Methionine is well-known for being the typical initiating amino acid for protein synthesis and has hydrophobic properties when incorporated into proteins [36]. These hydrophobic properties usually result in methionine incorporation within the core of proteins. However, certain proteins have surface-exposed methionine susceptible to oxidation that is associated with age-related disease [30, 37, 38]. S-adenosylmethionine, a methionine metabolism intermediate, is widely-used with functions in methylation as well as amine, methylene, and sulfur atom donation [30, 39, 40]. Methionine is especially important for poultry production as birds have exceedingly high sulfur amino acid requirements, and low methionine levels can negatively affect growth rate, carcass yield, fat content, and disease immunity [41–43]. Cysteine's ability to form disulfide bonds makes it incredibly important to tertiary protein structure and can occur with and without enzyme interactions [30, 44]. Cysteine is involved with keratin and feather production in poultry and deficiencies have been correlated to poor breast muscle development [45, 46]. Swine also need higher amounts of cysteine as they age to compensate for body maintenance [29, 47].

5. Breeding efforts

Soybean has an extensive cultivated history dating back thousands of years to its country of origin, China. *Glycine max*, the contemporary species of soybean, was domesticated from the wild species *Glycine soja* and has been continually been improved through selective and molecular breeding [48–50]. Once harvesting traits such as seed shatter and lodging were improved in the late 1930s to make soybean a competitive row crop, other cultivar improvements became a valuable research goal [49]. Current breeding programs tend to focus on traits such as yield, disease resistance, abiotic stress tolerance, and seed composition. Seed composition improvements include protein and oil content, fatty acid levels, anti-nutritional factors, isoflavones, and amino acids profiles. Before 1972, there had been zero reported research for improvement of soybean amino acid profiles, rather with emphasis on overall protein content [51]. Modern breeders are also inclined to concentrate efforts on protein content and consider amino acid levels an afterthought [52]. TN04-5321 is the only released germplasm in the United States that maintains yield and protein content while improving amino acid balance by increasing methionine and cysteine to levels recommended livestock needs [53].

The major soybean storage proteins are 11S (glycinin) and 7S (conglycinin) and provide the bulk of amino acids with limited non-proteinogenic amino acids in seed [54–56]. By increasing 11S and 7S quantity, more protein can ultimately be present in food and feed. Overemphasis on crude protein content can have negative ramifications on overall protein quality, specifically

deficient amino acids. While an increase in protein content would theoretically entail an increase in amino acids including methionine and cysteine [57], the opposite effect has been more notable [58, 59]. Molecular breeding techniques have recently improved the understanding of amino acid genomic regulation. Multiple quantitative trait loci (QTL) studies have been performed to identify genomic regions that control amino acids levels in soybean seed [60-64]. A myriad of QTL's were found to create amino acid phenotypic variation. Individual amino acids had reoccurring or proximal QTL's discovered such as Satt 518 [60], ss107913002 [61], and BARC-048619 [62] for glycine and threonine. QTL's for methionine and cysteine were also discovered which could lead to valuable improvements for soybean livestock feed [60–64]. Other genomic studies such as genome-wide association studies (GWAS) and genetic diversity analyses would further improve genetic understanding.

6. Genetic engineering

Genetic engineering experiments such as genetically modified organisms (GMOs) and gene editing are also promising avenues for improving amino acid profiles of soybeans. Compared to conventionally bred varieties, transgenic soybeans face additional adversity from registration requirements and public opinion. Transgenic efforts generally have one of three targets: magnifying biosynthesis genes, adjusting biosynthesis regulation, and modifying storage proteins. The earliest example would be a Brazil nut gene transfer in 1992. This successfully increased protein content and methionine biosynthesis, however a major food allergen was also transferred making commercialization impossible [65]. Expressing zein proteins from corn has also been shown to increase sulfur-containing amino acids levels in soybean [66, 67]. Altering biosynthesis feedback regulation amplified both non-proteinogenic and proteogenic lysine by circumventing normal enzymatic pathways [68]. Tryptophan in soybean also exhibited increased non-proteinogenic levels when a feedback-insensitive enzyme was transferred [25]. While soybean is not deficient in lysine or tryptophan, corn is deficient in both. By increasing lysine and tryptophan concentrations, soybean becomes an even more useful feed additive to corn rations. Even with limited research on modifying overall amino acids profiles in soybeans, modifying 11S and 7S storage proteins ratios [69] or silencing their expression entirely [70] has displayed increased amino acids levels. Similarly, a study using irradiated mutant soybeans lacking storage proteins as breeding parents demonstrated increased non-proteinogenic amino acids contents [54]. In addition, further research should be conducted to determine the bioavailability and digestibility of increased non-proteinogenic amino acids in soybean.

7. Agronomic relations

Amino acids concentrations in soybean are not only affected by their genetic potential. Agronomic properties greatly impact the final levels of amino acids. Agronomy encompasses all aspects of crop production including environmental effects, climatic variables, and abiotic factors. Perhaps the most considered agronomic factor is soil nutrient availability. Insufficient soil nutrient levels of nitrogen, potassium, phosphorous, sulfur, calcium, and magnesium create poor amino acid profiles in soybean plants [71]. Increased phosphorous rates have been shown to increase the percentage of methionine and tryptophan in seed but had no effect on protein content percentage [72]. Applications of sulfur, phosphorous, and nitrogen (individual and combined) produced a variety of different methionine and cysteine seed concentrations [73]. Sulfur deficiencies were also shown to inhibit the production of 11S proteins while almost eliminating methionine and cysteine in 7S proteins [74]. It is becoming more popular to also apply biological substances such as amino acids to plants through foliar and seed application. Amino acid uptake by soybean and wheat have been proven, and improved soybean growth rates and antioxidant effects have also occurred [75, 76]. Further research should be conducted to determine if biofortification solutions are possible through amino acid application.

Amino acid variation has also been shown to occur across environments [24, 59]. Specific correlations have emerged in response to temperature, solar radiation, and rainfall. One study shows that increased temperature leads to increased concentrations of all proteinogenic amino acids [77], while another concludes that only methionine and cysteine increase alongside temperature [78]. Increased solar radiation and greater available water appeared to have a negative relationship with amino acid content [77]. These favorable conditions would increase yield which has been shown to have a negative correlation with overall protein content [57]. The multitude of agronomic factors that affect amino acid profiles in soybean make it exceedingly important to compensate for variables when researching.

8. Processing impacts

The diversity in food, feed, and industrial used for soybean require the whole seed or seed components to be processed. Processing can affect the nutritional value of soybean protein and presence of amino acids in food and feed. Processing procedures can either separate seed components for different purposes or convert the entire seed into a product (usually human food). Some human soy foods such as edamame and soybean sprouts need little to no processing. Others including soymilk, tofu, natto, and soy sauce involve more processing. Soymilk and tofu processing are interconnected. Soymilk is a water-extract of whole or crushed soybeans that is coagulated and pressed into tofu [48]. While not all seed proteins convert into protein in tofu, 11S/7S storage protein ratios have been shown to be both positively and negatively correlated with tofu hardness [79, 80]. Natto is a soy food created by fermenting whole soybeans with *Bacillus subtilis*. Fermentation time affects final amino acid concentrations, and proper fermentation length could potentially increase nutritional values [81]. Soy sauce is produced by traditional and commercial methods, but both are based around whole seed or meal fermentation with *Aspergillus* sp. However, commercial methods have a lower amino acid to nitrogen ratio [48].

Soybean meal processing also impacts the level of amino acids in livestock feed. The first step in soybean meal processing is essentially separating protein from oil. A variety of methods

exist including solvent extraction, screw pressing, and extruding [48, 82, 83]. All three processes have three final products: oil, meal (usually toasted to lessen anti-nutritional factors), and hulls. Over processing of solvent extracted soymeal has been shown to decrease lysine, cysteine, and arginine levels [84, 85]. Protein solubility and dispersibility measurements may be a useful indicator of over processing [86, 87]. Soybean hulls are sometimes added to livestock feed for additional fiber, however an increase in hull/meal ratios decrease the digestibility of amino acids [88]. While soybean is renowned for its protein and amino acid content, actual nutritional values can be decreased through certain processing methods.

9. Evaluation methods

All previously mentioned aspects of soybean production in regard to amino acid levels and human and animal nutrition depend on a single common denominator: amino acid quantification. Amino acids must be reliably, effectively, and accurately identified, measured and evaluated. A Google Scholar search of "amino acid analysis" will display over 1 million results. Several reviews have been published regarding the development of amino acid analysis [89–91]. In general, contemporary analysis of amino acids from any source will be performed by chromatography or near-infrared reflectance spectroscopy. Chromatography is the common method with specific techniques including ion exchange chromatography (IEC), high-performance liquid chromatography (HPLC), and gas chromatography (GC). HPLC is the more validated method for soybean amino acid analysis. It is more efficient than IEC, and it does not require the transformation into volatiles like GC [91–94]. Near-infrared reflectance spectroscopy (NIRS) is a more recent addition to amino acid analysis, and it has the potential to drastically improve the efficiency in soybean feed evaluation [19]. The inability to actually measure amino acid levels is main hindrance for NIRS amino acid analysis. NIRS methods must be developed from a calibration set of raw data (often from HPLC) [95-97]. Nonetheless, efficiency improvements should persuade researchers to continually explore future NIRS amino acid analysis applications.

10. Conclusion

Soybean is a valuable source of protein and amino acids for humans and livestock. Soybean's well-balanced amino acid profile provides all essential amino acids as well as most nonessential. However, there is much room for nutritional improvement. Proteinogenic sulfurcontaining amino acids, methionine and cysteine, are deficient in soybean and are especially needed in livestock rations. Increased levels of these amino acids would augment soybean meal value and lessen the need for synthetic amino acid supplements. Breeding efforts have made little progress in adjusting amino acid profiles thus far, however significant developments in understanding genomic control regions promise future success. Genetic engineering efforts have shown promising amino acid improvements, but regulations and public opinion made commercialization difficult. New gene-editing technology could be the key to unlocking true nutritional improvement.

Agronomic properties and processing methods both impact the final quantities of amino acids available to humans and livestock. Understanding these impacts are essential to improve the nutritional quality of soybeans. Amino acid evaluation through HPLC provides reliable and efficient quantification, yet even quicker measurements are possible through NIRS. As the world's population continues to grow, soybeans will be essential to both human and livestock for amino acid requirements. Wholesome approaches that understand the complexities of amino acids in soybean will be required to maximize overall success and feed the world with balance soy proteins.

Acknowledgements

The authors would like to acknowledge Virginia Polytechnic Institute and State University's Open Access Subvention Fund for funding the publishing fee.

Conflict of interest

The authors declare there is no conflict of interest.

Author details

William Monte Singer^{1*}, Bo Zhang¹, M.A. Rouf Mian² and Haibo Huang³

- *Address all correspondence to: wilmsing@vt.edu
- 1 School of Plant and Environmental Sciences, Virginia Polytechnic and State Institution, Blacksburg, Virginia, USA
- 2 United States Department of Agriculture, Agricultural Research Service, Raleigh, North Carolina
- 3 Department of Food Science and Technology, Virginia Polytechnic Institute and State University, Blacksburg, Virginia, USA

References

- [1] International: World Oilseed Production. 2019. Available from: http://soystats.com/international-world-oilseed-production/
- [2] International: World Soybean Production. 2019. Available from: http://soystats.com/international-world-soybean-production/

- [3] U.S. Exports: Soy Exports by Customer. 2019. Available from: http://soystats.com/u-sexports-soy-exports-by-customersda/
- [4] Soybean Meal. United Soybean Board. 2019. Available from: https://unitedsoybean.org/ topics/soybean-meal/
- [5] Osborne TB, Mendel LB. Amino-acids in nutrition and growth. 1914. Journal of the American College of Nutrition. 1993;12(5):484-485
- [6] Herrmann K, Somerville RL. Amino Acids: Biosynthesis and Genetic Regulation. Reading, Massachusetts: Addison-Wesley Publishing Company, Inc; 1983
- [7] Wade C. Carlson Wade's Amino Acids Book. New Canaan, Connecticut: Keats Publishing Inc.; 1985
- [8] Berry TH, Becker DE, Rasmussen OG, Jensen AH, Norton HW. The limiting amino acids in soybean protein. Journal of Animal Science. 1962;21(3):558-561
- [9] Boisen S, Hvelplund T, Weisbjerg MR. Ideal amino acid profiles as a basis for feed protein evaluation. Livestock Production Science. 2000;64(2):239-251
- [10] Han Y, Suzuki H, Parsons CM, Baker DH. Amino acid fortification of a low-protein corn and soybean meal diet for chicks. Poultry Science. 1992;71(7):1168-1178
- [11] Waldroup PW, Mitchell RJ, Payne JR, Hazen KR. Performance of chicks fed diets formulated to minimize excess levels of essential amino acids. Poultry Science. 1976;55(1):243-253
- [12] D'Mello JPF, editor. Amino Acids in Human Nutrition and Health. Wallingford, United Kingdom: CAB International; 2012
- [13] Jez JM, Fukagawa NK. Plant sulphur compounds and human health. In: Sulfur: A Missing Link between Soils, Crops, and Nutrition. American Society of Agronomy, Crop Science Society of America, Soil Science Society of America; 2008. pp. 281-291
- [14] Weaver R. Molecular Biology. 4th ed. New York, NY: McGraw-Hill; 2008
- [15] Wagner I, Musso H. New naturally occurring amino acids. Angewandte Chemie (International Ed. in English). 1983;22(11):816-828
- [16] Buttery PJ, D'Mello JPF. Amino acid metabolism in farm animals: An overview. In: Amino Acids in Farm Animal Nutrition. Wallingford, United Kingdom: CAB International; 1994. pp. 1-10
- [17] Kuiken KA, Lyman M. Essential amino acid composition of soy bean meals prepared from twenty strains of soy beans. Journal of Biological Chemistry. 1949;177:29-36
- [18] Pfarr MD, Kazula MJ, Miller-Garvin JE, Naeve SL. Amino acid balance is affected by protein concentration in soybean. Crop Science. 2018;58(5):2050-2062
- [19] Measuring Beyond the Bushel. United Soybean Board. 2019. Available from: https://unitedsoybean.org/article/measuring-beyond-the-bushel-1/

- [20] Fernandez SR, Aoyagi S, Han Y, Parsons CM, Baker DH. Limiting order of amino acids in corn and soybean meal for growth of the chick. Poultry Science. 1994;73(12):1887-1896
- [21] Control Feed Costs with Amino Acids. Drovers. 2019. Available from: https://www.drovers.com/article/control-feed-costs-amino-acids
- [22] Managing Nutrients and Pathogens from Animal Agriculture. Natural Resource, Agriculture, and Engineering Service. Ithaca, NY; 2000
- [23] Willke T. Methionine production—A critical review. Applied Microbiology and Biotechnology. 2014;98(24):9893-9914
- [24] Goldflus F, Ceccantini M, Santos W. Amino acid content of soybean samples collected in different Brazilian states: Harvest 2003/2004. Brazilian Journal of Poultry Science. 2006;8(2):105-111
- [25] Kita Y, Nakamoto Y, Takahashi M, Kitamura K, Wakasa K, Ishimoto M. Manipulation of amino acid composition in soybean seeds by the combination of deregulated tryptophan biosynthesis and storage protein deficiency. Plant Cell Reports. 2010;**29**(1):87-95
- [26] Rackis JJ, Anderson RL, Sasame HA, Smith AK, VanEtten CH. Soybean amino acids, amino acids in soybean hulls and oil meal fractions. Journal of Agricultural and Food Chemistry. 1961;9(5):409-412
- [27] Mato JM, Martinez-Chantar ML, Lu SC. Methionine metabolism. In: Amino Acids in Human Nutrition and Health. Wallingford, United Kingdom: CAB International; 2012. pp. 173-188
- [28] Kredich NM. Regulation of cysteine biosynthesis in *Escherichia coli* and *Salmonella typhimurium*. In: Amino Acids: Biosynthesis and Genetic Regulation. Reading, Massachusetts: Addison-Wesley Publishing Company, Inc; 1983. pp. 115-132
- [29] Fuller MF. Amino acid requirements for maintenance, body protein accretion and reproduction in pigs. In: Amino Acids in Farm Animal Nutrition. Wallingford, United Kingdom: CAB International; 1994. pp. 155-184
- [30] Brosnan JT, Brosnan ME. The sulfur-containing amino acids: An overview. The Journal of Nutrition. 2006;**136**(6):1636S-1640S
- [31] Jankowski J, Kubińska M, Zduńczyk Z. Nutritional and immunomodulatory function of methionine in poultry diets—A review. Annals of Animal Science. 2014;14(1):17-32
- [32] D'Mello JPF. Responses of growing poultry to amino acids. In: Amino Acids in Farm Animal Nutrition. Wallingford, United Kingdom: CAB International; 1994. pp. 205-244
- [33] Herrmann K. The common aromatic biosynthetic pathway. In: Amino Acids: Biosynthesis and Genetic Regulation. Reading, Massachusetts: Addison-Wesley Publishing Company, Inc; 1983
- [34] Allee GL, Kerley MS. Modifications in soybean seed composition to enhance animal feed use and value: Moving from a dietary ingredient to a functional dietary component.

- Journal of Agrobiotechnology Management & Economics. 2003. Vol 6. Number 1 & 2. Article 5. Available from: http://agbioforum.org/v6n12/v6n12a05-kerley.htm
- [35] Wu G. Dietary requirements of synthesizable amino acids by animals: A paradigm shift in protein nutrition. Journal of Animal Science and Biotechnology. 2014;5(1):34
- [36] Ingenbleek Y, Kimura H. Nutritional essentiality of sulfur in health and disease. Nutrition Reviews. 2013;71(7):413-432
- [37] Levine RL, Mosoni L, Berlett BS, Stadtman ER. Methionine residues as endogenous antioxidants in proteins. Proceedings of the National Academy of Sciences of the United States of America. 1996;93(26):15036-15040
- [38] Moskovitz J. Methionine sulfoxide reductases: Ubiquitous enzymes involved in antioxidant defense, protein regulation, and prevention of aging-associated diseases. Biochimica et Biophysica Acta. 2005;1703(2):213-219
- [39] Catoni GL. S-Adenosylmethionine—A new intermediate formed enzymatically from L-methionineandadenosinetriphosphate. The Journal of Biological Chemistry. 1953;204(1): 403-416
- [40] Fontecave M, Atta M, Mulliez E. S-adenosylmethionine: Nothing goes to waste. Trends in Biochemical Sciences. 2004;29(5):243-249
- [41] Bunchasak C. Role of dietary methionine in poultry production. The Journal of Poultry Science. 2009;46(3):169-179
- [42] Conde-Aguilera JA, Cobo-Ortega C, Tesseraud S, Lessire M, Mercier Y, van Milgen J. Changes in body composition in broilers by a sulfur amino acid deficiency during growth. Poultry Science. 2013;92(5):1266-1275
- [43] Wu B, Cui H, Peng X, Fang J, Cui W, Liu X. Effect of methionine deficiency on the thymus and the subsets and proliferation of peripheral blood T-cell, and serum IL-2 contents in broilers. Journal of Integrative Agriculture. 2012;11(6):1009-1019
- [44] Jessop CE, Chakravarthi S, Watkins RH, Bulleid NJ. Oxidative protein folding in the mammalian endoplasmic reticulum. Biochemical Society Transactions. 2004;32(Pt 5:655-658
- [45] Wylie LM, Robertson GW, Macleod MG, Hocking PM. Effects of ambient temperature and restricted feeding on the growth of feathers in growing turkeys. British Poultry Science. 2001;42(4):449-455
- [46] Bonato MA, Sakomura NK, Siqueira JC, Fernandes JBK, Gous RM. Maintenance requirements for methionine and cysteine, and threonine for poultry. South African Journal of Animal Science. 2011;41(3):209-222-222
- [47] Lewis AJ. Methionine-cysteine relationships in pig nutrition. In: Absorption and Utilization of Amino Acids. Wallingford, United Kingdom: CAB International; 2003. pp. 143-156

- [48] Liu K. Soybeans: Chemistry, Technology, and Utilization. New York, NY: Chapman & Hall; 1997
- [49] Sleper DA, Poehlman JM. Breeding Field Crops. 5th ed. Ames, Iowa: Blackwell Publishing Professional; 2006
- [50] Hermann FJ. A Revision of the Genus Glycine and its Immediate Allies. Washington, D.C: U.S. Department of Agriculture; 1962. 84 p
- [51] Howell RW, Brim CA, Rinne RW. The plant geneticist's contribution toward changing lipid and amino acid composition of soybeans. Journal of the American Oil Chemists' Society. 1972;49(1):30-32
- [52] Mahmoud AA, Natarajan SS, Bennett JO, Mawhinney TP, Wiebold WJ, Krishnan HB. Effect of six decades of selective breeding on soybean protein composition and quality: A biochemical and molecular analysis. Journal of Agricultural and Food Chemistry. 2006;54(11):3916-3922
- [53] Panthee D, Pantalone VR. Registration of soybean germplasm lines TN03-350 and TN04-5321 with improved protein concentration and quality. Crop Science. 2006;46:2328-2329
- [54] Takahashi M, Uematsu Y, Kashiwaba K, Yagasaki K, Hajika M, Matsunaga R, et al. Accumulation of high levels of free amino acids in soybean seeds through integration of mutations conferring seed protein deficiency. Planta. 2003;217(4):577-586
- [55] Meinke DW, Chen J, Beachy RN. Expression of storage-protein genes during soybean seed development. Planta. 1981;153:130-139
- [56] Wolf WJ. Soybean protein nomenclature: A progress report. Cereal Science Today. Number 3. 1969;14:75-129. Available from: https://pubag.nal.usda.gov/catalog/31396
- [57] Wilcox JR, Shibles RM. Interrelationships among seed quality attributes in soybean. Crop Science. 2001;41(1):11-14
- [58] Paek NC, Imsande J, Shoemaker RC, Shibles R. Nutritional control of soybean seed storage protein. Crop Science. 1997;37(2):498-503
- [59] Thakur M, Hurburgh CR. Quality of US soybean meal compared to the quality of soybean meal from other origins. Journal of the American Oil Chemists' Society. 2007;84(9):835-843
- [60] Panthee DR, Pantalone VR, Saxton AM, West DR, Sams CE. Genomic regions associated with amino acid composition in soybean. Molecular Breeding. 2006;**17**:79-89
- [61] Fallen B, Hatcher C, Allen F, Kopsell D, Saxton A, Chen P, et al. Soybean seed amino acid content QTL detected using the universal soy linkage panel 1.0 with 1,536 SNPs. Journal of Plant Genome Sciences. 2013;1(3):68-79. Agronomy and Horticulture—Faculty Publications. Available from: https://digitalcommons.unl.edu/agronomyfacpub/807

- [62] Warrington CV, Abdel-Haleem H, Hyten DL, Cregan PB, Orf JH, Killam AS, et al. QTL for seed protein and amino acids in the Benning × Danbaekkong soybean population. Theoretical and Applied Genetics. 2015;128(5):839-850
- [63] Li X, Tian R, Kamala S, Du H, Li W, Kong Y, et al. Identification and verification of pleiotropic QTL controlling multiple amino acid contents in soybean seed. Euphytica. 2018;214(6):93
- [64] Panthee DR, Pantalone VR, Sams CE, Saxton AM, West DR, Orf JH, et al. Quantitative trait loci controlling sulfur containing amino acids, methionine and cysteine, in soybean seeds. Theoretical and Applied Genetics. 2006;112(3):546-553
- [65] Townsend JA, Thomas LA, Kulisek ES, Daywalt MJ, Winter KRK, Altenbach AB. Improving the quality of seed proteins in soybean. In: Proceedings of the 4th Biennial Conference on Molecular and Cellular Biology of Soybean. Ames, Iowa: Iowa State University; 1992. p. 4
- [66] Kerr P. Utilization and Quality of Identity Preserved Oilseed Co-Products. Presentation, Institute of Food Technologists; Symposium. New Orleans, LA; 1996
- [67] Kim W-S, Krishnan HB. Impact of co-expression of maize 11 and 18 kDa δ-zeins and 27 kDa γ-zein in transgenic soybeans on protein body structure and sulfur amino acid content. Plant Science. 2019;280:340-347
- [68] Falco SC, Guida T, Locke M, Mauvais J, Sanders C, Ward RT, et al. Transgenic canola and soybean seeds with increased lysine. Bio/Technology. 1995;13(6):577
- [69] El-Shemy H, Khalafalla M, Fujita K, Ishimoto M. Improvement of protein quality in transgenic soybean plants. Biologia Plantarum. 2007;51:277-284
- [70] Schmidt MA, Barbazuk WB, Sandford M, May G, Song Z, Zhou W, et al. Silencing of soybean seed storage proteins results in a rebalanced protein composition preserving seed protein content without major collateral changes in the metabolome and transcriptome. Plant Physiology. 2011;**156**(1):330-345
- [71] Haghiri F. Influence of macronutrient elements on the amino aicd composition of Soybean plants. Agronomy Journal. 1966;58:609-612
- [72] Kapoora AC, Gupta YP. Changes in proteins and amino acids in developing soybean seed and effect of phosphorus nutrition. Journal of the Science of Food and Agriculture. 1977;**28**(2):113-120
- [73] Arora SK, Luthra YP. Relationship between sulphur content of leaf with methionine, cysteine and cysteine contents in the seeds of Phaseolus aureus L. As affected by S, P, and N application. Plant and Soil. 1971;34(1):91-96
- [74] Gayler KR, Sykes GE. Effects of nutritional stress on the storage proteins of soybeans. Plant Physiology. 1985;78(3):582-585

- [75] Gioseffi E, de Neergaard A, Schjoerring JK. Interactions between uptake of amino acids and inorganic nitrogen in wheat plants. Biogeosciences. 2012;9(4):1509-1518
- [76] Teixeira WF, Fagan EB, Soares LH, Umburanas RC, Reichardt K, Neto DD. Foliar and seed application of amino acids affects the antioxidant metabolism of the Soybean crop. Frontiers in Plant Science. 2017;8. Available from: https://www.frontiersin.org/articles/10.3389/fpls.2017.00327/full
- [77] Carrera CS, Reynoso CM, Funes GJ, Martínez MJ, Dardanelli J, Resnik SL. Amino acid composition of soybean seeds as affected by climatic variables. Brazilian Agricultural Research. 2011;46(12):1579-1587
- [78] Wolf RB, Cavins JF, Kleiman R, Black LT. Effect of temperature on soybean seed constituents: Oil, protein, moisture, fatty acids, amino acids and sugars. Journal of the American Oil Chemists' Society. 1982;59(5):230-232
- [79] Cai T, Chang K-C. Processing effect on soybean storage proteins and their relationship with tofu quality. Journal of Agricultural and Food Chemistry. 1999;47(2):720-727
- [80] Mujoo R, Trinh DT, Ng PKW. Characterization of storage proteins in different soybean varieties and their relationship to tofu yield and texture. Food Chemistry. 2003;82(2):265-273
- [81] Weng TM, Chen MT. Changes of protein in Natto (a fermented soybean food) affected by fermenting time. Food Science and Technology Research. 2010;**16**(6):537-542
- [82] Lusas EW. Soybean processing and utilization. In: Soybeans: Improvement, Production, and Uses. 3rd ed. Madison, WI: ASA, CSSA, SSSA; 2004. pp. 949-1036
- [83] Johnson L, Smith K. Soybean Processing. United Soybean Board. Soybean Meal INFO Center. Fact Sheet; 2019. 7 pages. Available from: https://www.soymeal.org/wp-content/uploads/2018/04/soybean_processing.pdf
- [84] Parsons CM, Hashimoto K, Wedekind KJ, Han Y, Baker DH. Effect of overprocessing on availability of amino acids and energy in soybean meal. Poultry Science. 1992;71(1):133-140
- [85] Taira H. Studies on amino acid contents of processed soybean: Part X. the influence of added sugars on the heat destruction of the basic and sulphur containing amino acids in soybean products. Agricultural and Biological Chemistry. 1966;30(9):847-855
- [86] Araba M, Dale NM. Evaluation of protein solubility as an indicator of overprocessing soybean meal. Poultry Science. 1990;69(1):76-83
- [87] Batal AB, Douglas MW, Engram AE, Parsons CM. Protein dispersibility index as an indicator of adequately processed soybean meal. Poultry Science. 2000;79(11):1592-1596
- [88] Dilger RN, Sands JS, Ragland D, Adeola O. Digestibility of nitrogen and amino acids in soybean meal with added soyhulls. Journal of Animal Science. 2004;82(3):715-724

- [89] Husek P, Simek P. Advances in amino acid analysis. Analytical and Bioanalytical Chemistry. 2001;**19**(9):986-999
- [90] Tristram GR, Rattenbury JM. The development of amino acid analysis. In: Amino Acid Analysis. Chichester, UK: Ellis Horwood; 1981. pp. 16-36
- [91] Williams AP. Recent developments in amino acid analysis. In: Amino Acids in Farm Animal Nutrition. Wallingford, United Kingdom: CAB International; 1994. pp. 11-36
- [92] Jajić I, Krstović S, Glamočić D, Jakšić SM, Abramović BF. Validation of an HPLC method for the determination of amino acids in feed. Journal of the Serbian Chemical Society. 2013;78(6):839-850
- [93] Oomah BD, Voldeng H, Fregeau-Reid JA. Characterization of soybean proteins by HPLC. Plant Foods for Human Nutrition. 1994;45:251-263
- [94] Malmer MF, Schroeder LA. Amino acid analysis by high-performance liquid chromatography with methanesulfonic acid hydrolysis and 9-fluorenylmethylchloroformate derivatization. Journal of Chromatography. A. 1990;514:227-239
- [95] Baianu I, Prisecaru V. Novel NIR spectroscopy correlation approach to amino acid analysis of soybean proteins for composition improvements. Nature Precedings. University of Illinois at Urbana. 2011. Available from: http://dx.doi.org/10.1038/npre.2011.6231.1
- [96] Kovalenko IV, Rippke GR, Hurburgh CR. Determination of amino acid composition of soybeans (Glycine max) by near-infrared spectroscopy. Journal of Agricultural and Food Chemistry. 2006;54(10):3485-3491
- [97] Pazdernik DL, Killam AS, Orf JH. Analysis of amino and fatty acid composition in soybean seed, using near infrared refelctance spectroscopy. Agronomy Journal;89:679-685



IntechOpen

IntechOpen