

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

4,800

Open access books available

122,000

International authors and editors

135M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.

For more information visit www.intechopen.com



Roles of ncRNAs in Ovarian Dysfunction of Polycystic Ovary Syndrome

Junyong Han, Zhen Yu, Gang Chen and Fan Wang

Abstract

Polycystic ovary syndrome (PCOS) is a common endocrine disease in women of childbearing age. Many heterogeneous clinical manifestations of PCOS, including hyperandrogenism, obesity, insulin resistance, hirsutism, acne, chronic anovulation and infertility, seriously affected the quality of life of women worldwide and made it difficult to clearly demonstrate the specific pathophysiology. In recent years, large-scale studies have shown that non-coding RNAs (ncRNAs) play an important role in the regulation of ovarian functions, which did not have the ability to encode proteins and could regulate hormone synthesis and germ cell development, differentiation, and apoptosis by silencing transposable elements and regulating coding genes. A number of researches by whole transcriptome sequencing of polycystic ovaries (PCO) from PCOS patients or PCOS model animals found that the abnormal expressions of many ncRNAs were involved in the regulation of ovarian dysfunctions of PCOS, including the development of oocytes, the microenvironment of follicular fluid, and the proliferation, differentiation, and apoptosis of granulosa cells. The present review focused on the roles of ncRNAs in the PCO of PCOS, in order to provide a theoretical basis for further understanding of the molecular mechanisms of PCO formation in PCOS.

Keywords: ncRNAs, granule cell, oocyte, follicle fluid, polycystic ovary syndrome

1. Introduction

Polycystic ovary syndrome (PCOS) is an endocrine and reproductive disease in women that often occurs during the childbearing years [1]. PCOS is closely related to metabolic syndrome, and has become an increasingly serious public health problem worldwide, these patients have an increased risk of endometrial cancer, type 2 diabetes, and cardiovascular disease. The clinical characteristics of PCOS women show heterogeneity, include excessive androgen, infertility, obesity, anovulation, irregular menstruation, polycystic ovaries (PCO), hairy and recurrent miscarriage, insulin resistance and abnormal blood lipids [2].

Anovulatory infertility of PCOS patients accounts for more than 75%, and the spontaneous abortion rate of them in early pregnancy is 30–50% [3, 4]. Ovulation failure (such as ovulation or anovulation) is the main clinical features of PCOS. There are a large number of small follicles in the bilateral ovaries of PCOS patients, which can not grow into dominant follicle, suggesting the occurrence of follicular dysplasia [5]. The processes of ovarian follicular development and atresia

are a complex and delicate. However, ovulatory dysfunction in PCOS women is related to various factors such as abnormalities in proliferation and apoptosis of granule cells (GCs) and in follicular development and atresia. As is well-known that GCs are involved in the normal development and maturation of follicles. Some studies have found that the proliferation rate of GCs in PCOS patients is significantly higher, and the apoptotic rate is significantly lower [6, 7] suggesting that abnormalities in the proliferation and apoptosis of GCs may be a pathogenic mechanism of ovarian dysfunction of PCOS.

Due to the complexity of the etiology of PCOS, its specific pathophysiology has not yet been fully elucidated. More and more researches are currently being conducted to analyze the gene expression profiling of PCOS at the mRNA and protein levels, in order to explore its molecular mechanisms. PCOS is caused by the imbalance of multiple gene pathways [8]. The candidate genes are involved in sex hormone synthesis, insulin synthesis, chronic inflammatory factors, lipid metabolism, cell proliferation and apoptosis [8]. With the advancement of RNA sequencing technologies, many researchers have discovered that non-coding RNA (ncRNA, which refers to a class of RNA without open reading frame and can not translate into proteins) is involved in the regulation of numerous cell signaling pathways and biological processes for life activities, such as cell development, differentiation, apoptosis and hormone synthesis, and the occurrence and development of diseases [9].

Most of the human genome is transcribed into various ncRNAs, MicroRNA (miRNA) and long-chain non-coding RNAs (lncRNA) are hotspots for studying novel biomarkers and therapeutic targets in related diseases in recent years, such as cancer [10], diabetes [11] and PCOS [9]. Recent studies have shown that both miRNA and lncRNA play an important role in the pathogenesis of PCOS [9, 12]. miRNA and lncRNA can directly or indirectly affect the normal physiological functions of the ovaries, include the growth and development of follicles and oocytes [9, 12]. Moreover, ovarian dysfunction in PCOS seriously affects the reproductive capacity in reproductive women, but its specific molecular mechanism is not yet known. This chapter focuses on the roles of several ncRNAs in PCOS ovaries in recent years, which will help to further elucidate its pathogenesis and to discover new therapeutic targets.

2. ncRNA

ncRNAs as enzyme, regulatory signal, molecular sink, ligand, organizer of cellular structures, potential hormone and scaffold of molecular interactions, can play an important role in cell physiology and abnormal biological processes, such as nuclear transport, transcriptional regulation of genes, protein degradation, genomic imprinting and X chromosome silencing [13].

2.1 miRNAs

miRNAs are 21–23 nucleotides in length and widely found in eukaryotes, which can predict post-transcriptional regulation of at least half of the human transcriptome [14–16]. Mature miRNA is formed by removing and processing a longer primary transcript through a series of nucleases [17–19]. The main function of miRNAs is silencing or degrading the expressions of target genes at the post-transcriptional level by forming an RNA-induced silencing complex [20]. Abnormal expressions of miRNAs are associated with insulin resistance [21, 22], diabetes [11, 22, 23], inflammation [20, 23] and various cancer formations [10]. The expression profiles of some

miRNAs in ovarian various cells and follicular fluids of PCOS patients exist significant differences, which had been involved in the occurrence and development of PCOS by affecting the post-transcriptional regulation of the target genes [9, 12, 24].

2.2 lncRNAs

lncRNAs are a class of non-coding RNAs transcripts longer than 200 nucleotides in length, which produced by 4–9% of the sequence in mammalian genome [25, 26]. lncRNAs play critical roles in various human biological processes, such as chromatin modification, cell differentiation, proliferation and apoptosis, translational and post-translational regulation. Moreover, the abnormally expressed lncRNAs are involved in the occurrence and development of a variety of human diseases [27]. In recent years, more and more evidence has shown that lncRNAs are abnormally expressed in ovarian cumulus cells and/or GCs of PCOS patients [9, 28, 29]. These lncRNAs may be involved in ovarian steroid production, steroid receptor activity and IR, and future affect ovarian cell development, proliferation and apoptosis, which in turn leads to the occurrence of ovarian dysfunction of PCOS [28, 29].

3. PCO

PCO is one of the diagnostic indicators and symptoms for PCOS, which imaged the ovaries with ultrasound showing ≥ 12 follicles (each follicle is 2–9 mm in diameter) on one or both sides, and/or an ovarian volume ≥ 10 ml [5]. During the development and maturation of ovarian follicles, the oocyte interconnected and interdependent with surrounding GCs. The development of an oocyte requires GCs to provide nutrients, corresponding hormones and growth regulators [30, 31]. Abnormal interactions between GCs and oocytes are a possible cause of ovarian follicular dysplasia in PCOS [32]. Some vitro fertilization studies found that PCOS patients have a lower rate of implantation than normal women [33–35]. Microarray analysis indicated that abnormal endocrine and metabolism affect gene expression of oocytes in PCOS ovaries [36].

Moreover, ovarian follicular fluid is a fluid that fills the follicular cavity and surrounds cumulus cells. Follicular fluid is rich in substances such as hormones, growth factors, anti-apoptotic antibodies, various proteins, peptides, amino acids and nucleotides. Studies have confirmed that these substances are closely related to female reproductive system diseases, embryo quality and in vitro fertilization outcomes [37]. Therefore, the homeostasis of the microenvironment of follicular fluid directly affects follicular development and oocytes quality.

It is well known that insulin resistance and hyperinsulinemia play an important role in the pathophysiology of PCOS. Studies have found that most of PCOS patients with varying degrees of IR and compensatory hyperinsulinemia, including ovarian IR [38]. Ovarian GCs in PCOS patients are impaired by insulin-dependent glucose metabolism [38, 39]. Damaged glucose metabolism reduces the energy supply to GCs and oocytes, and thus hinders the proliferation of GCs and the development of oocytes [38, 39]. Therefore, GCs dysfunction is contributed to abnormal ovarian function in PCOS, including anovulation. Moreover, excessive apoptosis of cumulus GCs directly causes follicular dysplasia in PCOS, which are correlated with changes in ncRNAs expression profiles in follicular fluid. The abnormal expressions of ncRNAs from the ovaries of PCOS may contribute to its occurrence and development [9, 12, 40].

4. Effects of ncRNAs on ovarian dysfunction of PCOS

With the development of RNA sequencing technologies, many researchers have carried out miRNA or lncRNA sequencing studies on the ovaries of PCOS, in order to detect the molecular mechanisms of ovarian dysfunction. Studies have shown that compared with control rats, there are 129 miRNAs (49 miRNAs are up-regulated, 80 miRNAs are down-regulated) and 158 lncRNAs (114 lncRNAs are up-regulated, 44 lncRNAs are down-regulated) by deep-sequencing of ovaries tissue from letrozole-induced PCOS rats [41, 42]. Gene ontology (GO) and Kyoto encyclopedia of genes and genomes (KEG) Genes pathway analyzed and predicted that the differentially expressed ncRNAs in PCOS ovaries may be associated with abnormal ovarian GCs proliferation, apoptosis and steroidogenesis and ovarian insulin resistance [41, 42].

4.1 Roles of ncRNAs in the proliferation and apoptosis of GCs

4.1.1 *miR-141-3p*

In the ovaries of PCOS rat model, the expression of miR-141-3p is significantly decreased, which can target to death-associated protein kinase 1 (DAPK1) and mitogen-activated protein kinase 1 (MAPK1) signaling pathway to inhibit apoptosis of GCs, or regulate mitochondria-mediated apoptosis through phosphatidylinositol 3 kinase/protein kinase B (PI3K/Akt) and extracellular protein kinase (ERK) signaling pathways, and thus suppressing cell growth, promoting cell apoptosis, and further leading to the development of PCOS [41].

4.1.2 *miR-483-5p*

miR-483-5p is the most abundant known miRNA in human ovarian follicular fluid [43]. miR-483-5p is highly expressed in cumulus granulosa cells and follicular fluid of PCOS patients [44]. In PCOS cumulus cells, high concentrations of miR-483-5p hinders the expressions of Notch3 and MAPK3 protein by binding to the 3'UTR terminus of their mRNA, thereby blocking the Notch signaling pathway and the MAPK signaling pathway (the two pathways play an important role in cell proliferation, differentiation and apoptosis), which inhibits the proliferation and differentiation of cumulus GCs and promoted the apoptosis of GCs [44].

4.1.3 *LncRNA CD36-005*

lncRNA CD36-005 is a transcript encoding the fatty acid transporter CD36 gene, which may be involved in cell growth, development, transport, and metabolism by bioinformatics analysis [42, 45]. lncRNA CD36-005 is expressed in rat ovaries, and its expression level is related to the estrous cycle, which indicates that it plays a role in animal reproductive activities [45]. Moreover, the study found that lncRNA CD36-005 and CD36 were highly expressed in the ovaries of PCOS rats by high-throughput sequencing and qRT-PCR verification. lncRNA CD36-005 significantly inhibits the proliferation of GCs by reducing the viability and the S phase of the cell cycle, which may be involved in the pathogenesis of PCOS [42].

4.2 Roles of ncRNAs in ovarian steroidogenesis

4.2.1 *miR-320*

miR-320 is closely related to PCOS, especially in the process of follicular development [46, 47]. miR-320 can regulate the translation of its target genes by

post-transcriptional regulation, thereby improving the levels of steroid hormones. miR-320 was down-regulated in cumulus GCs from PCOS ovaries [48], and its expression in follicular fluid was controversial [49]. miR-320 deficiency not only impairs the expression of the steroid synthase CYP11A1 and CYP19A1, but also enhances the steroidogenesis in CGs by directly regulating the RUNX2/CYP11A1 cascade in the 3'UTR of Runx2, indicating that the cascade are a possible mechanism for the lack of estrogen synthesis in GCs of PCOS patients. In addition, miR-320 can target transcriptional factors E2F1 and steroidogenic factor-1 (SF-1) to inhibit the proliferation of GCs, and promote the synthesis of testosterone and progesterone [47].

4.2.2 *LncRNA HCG26*

lncRNA HCG26 is mainly distributed in the nucleus of ovarian GCs, which is highly expressed in PCOS patients [29]. HCG26 knockout can promote aromatase gene expression and estradiol synthesis, but can not affect the levels of androstenedione and follicle stimulating hormone, suggesting lncRNA HCG26 is involved in abnormal ovarian steroidogenic synthesis [50]. Moreover, the study also found that lncRNA HCG26 deficiency inhibits the proliferation of GCs and its expression correlated with the number of PCOS ovarian follicles, suggesting that lncRNA HCG26 may affect the proliferation of GCs and contributes to the formation of polycystic ovary morphology [29].

4.3 Roles of ncRNAs in ovarian insulin resistance

4.3.1 *miR-92*

Down-regulation of miR-92a/b expression in PCOS patients is associated with insulin resistance, hyperinsulinemia and hyperandrogenism [24], which can increase the expression of its target gene IRS-2, which activates the insulin signaling pathway in the ovaries of PCOS patients, thereby leading to be involved in the process of insulin resistance and hyperinsulinemia [24, 29].

4.3.2 *miR-145*

miR-145 overexpression in GCs of PCOS patients inhibits the target gene IRS1 and MAPK/ERK signaling pathways, and promotes the activity of PI3K/Akt signaling pathway through negative feedback, thereby improving IR in PCOS patients [7, 24, 50].

5. Conclusions

PCOS is a chronic disease that affects the patients throughout their lives. However, there is currently no gene that is recognized as the determinant of the pathogenesis of PCOS. Although the differential expression of ncRNAs in the ovaries from PCOS are involving in ovarian steroidogenesis, insulin resistance, cell proliferation and apoptosis and affect follicular development by regulating the expression of corresponding target genes at the post-transcriptional level, there are still many ncRNAs and the specific mechanism of their action in ovarian function of PCOS have not been discovered. Moreover, the roles and mechanisms of lncRNAs in PCOS have just been carried out, and the focus is on the study of lncRNA in peripheral blood of the patients or animal models, that in ovarian tissue is limited. The interactions between ncRNAs in PCOS have not been reported, such as miRNA-miRNA, lncRNA-lncRNA, and miRNA-lncRNA. Solving these problems will be contribution to further understand the etiology and pathogenesis of PCOS.

Acknowledgements

This study was supported by Young and Middle-aged Talents Training Project of Fujian Provincial Health and Family Planning (2018-ZQN-20).

Conflict of interest

The authors declare no conflict of interest.

A. Appendices

PCOS	polycystic ovary syndrome
ncRNAs	non-coding RNA
PCO	polycystic ovary
GCs	granule cells
lncRNA	long-chain non-coding RNAs
GO	gene ontology
KEGG	Kyoto encyclopedia of genes and genomes
DAPK1	death-associated protein kinase 1
MAP K1	mitogen-activated protein kinase 1
PI3K/Akt	phosphatidylinositol 3 kinase/protein kinase B
ERK	extracellular protein kinase
SF-1	steroidogenic factor-1

Author details

Junyong Han¹, Zhen Yu¹, Gang Chen¹ and Fan Wang^{1,2,3*}

¹ Fujian Provincial Key Laboratory of Medical Analysis, Fujian Academy of Medical Sciences, Fuzhou, China

² State Key Laboratory for Evaluation of Exercise Physiological Functions from General Administration of Sport of China, School of Physical Education and Sport Sciences, Fujian Normal University, Fuzhou, China

³ Laboratory of Sport Physiology and Biomedicine, School of Physical Education and Sport Sciences, Fujian Normal University, Fuzhou, China

*Address all correspondence to: 517239819@qq.com

IntechOpen

© 2019 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. 

References

- [1] Bazarganipour F, Taghavi SA, Montazeri A, Ahmadi F, Chaman R, Khosravi A. The impact of polycystic ovary syndrome on the health-related quality of life: A systematic review and meta-analysis. *International Journal of Reproductive Biomedicine*. 2015;**13**(2):61-70
- [2] Fan W, Shaobing W, Zhenghong Z, Qingqiang L, Yiping L, Yijun X, et al. Defective insulin signaling and the protective effects of dimethyldiguanide during follicular development in the ovaries of polycystic ovary syndrome. *Molecular Medicine Reports*. 2017;**16**(6):8164-8170. DOI: 10.3892/mmr.2017.7678
- [3] Caldwell A, Edwards M, Desai R, Jimenez M, Gilchrist RB, Handelsman DJ, et al. Neuroendocrine androgen action is a key extraovarian mediator in the development of polycystic ovary syndrome. *Proceedings of the National Academy of Sciences of the United States of America*. 2017;**114**(16):E3334-E3343. DOI: 10.1073/pnas.1616467114
- [4] Xuezhou Y, Xiaozhen Q, Yanli L, Qipeng W, Jinhai Y, Xiaofang Y, et al. Serum chemerin level in women with PCOS and its relation with the risk of spontaneous abortion. *Gynecological Endocrinology*. 2018;**34**(10):864-867. DOI: 10.1080/09513590.2018.1462316
- [5] Giampaolino P, Della Corte L, De Rosa N, Mercurio A, Bruzzese D, Bifulco G. Ovarian volume and PCOS: A controversial issue. *Gynecological Endocrinology*. 2018;**34**(3):229-232. DOI: 10.1080/09513590.2017.1391205
- [6] Song WJ¹, Shi X, Zhang J, Chen L, Fu SX, Ding YL. Akt-mTOR signaling mediates abnormalities in the proliferation and apoptosis of ovarian granulosa cells in patients with polycystic ovary syndrome. *Gynecologic and Obstetric Investigation*. 2018;**83**(2):124-132. DOI: 10.1159/000464351
- [7] Cai G, Ma X, Chen B, Huang Y, Liu S, Yang H, et al. MicroRNA-145 negatively regulates cell proliferation through targeting IRS1 in isolated ovarian granulosa cells from patients with polycystic ovary syndrome. *Reproductive Sciences*. 2016;**24**(6):902-910. DOI: 10.1177/1933719116673197
- [8] Li L, Baek KH. Molecular genetics of polycystic ovary syndrome: An update. *Current Molecular Medicine*. 2015;**15**(4):331-342. DOI: 10.2174/1566524015666150505160140
- [9] Qian Y, Chengliang Z, Jiexue P, Huanghe F. Research advances in the roles of ncRNAs in polycystic ovary syndrome. *Journal of Shanghai Jiaotong University*. 2016;**36**(6):921-925. DOI: 10.3969/j.issn.1674-8115.2016.06.027
- [10] Trang P, Weidhaas JB, Slack FJ. MicroRNAs and cancer. In: Coleman W, Tsongalis G, editors. *The Molecular Basis of Human Cancer*. New York, NY: Humana Press; 2017. https://doi.org/10.1007/978-1-59745-458-2_17
- [11] Tian Y, Xu J, Du X, Fu X. The interplay between noncoding RNAs and insulin in diabetes. *Cancer Letters*. 2018;**419**:53-63. DOI: 10.1016/j.canlet.2018.01.038
- [12] Huang X, Liu C, Hao C, Tang Q, Liu R, Lin S, et al. Identification of altered microRNAs and mRNAs in the cumulus cells of PCOS patients. *Reproduction*. 2016;**151**(6):643-655. DOI: 10.1530/REP-16-0071
- [13] Fu XD. Non-coding RNA: A new frontier in regulatory biology. *National Science Review*. 2014;**1**(2):190-204. DOI: 10.1093/nsr/nwu008

- [14] Yanli W, Meiling Y, En-Bo M, Jianzhen Z. Identification and analysis of the miRNAs targeting key genes involved in cuticle metabolism in *Locusta migratoria* (orthoptera:Acrididae). *Acta Entomologica Sinica*. 2017;**60**(3):309-317. DOI: 10.16380/j.kcxb.2017.03.008
- [15] Davis-Dusenbery BN, Hata A. Mechanisms of control of microRNA biogenesis. *Journal of Biochemistry*. 2010;**148**(4):381-392. DOI: 10.1093/jb/mvq096
- [16] Jinbiao M, Ying H. Post-transcriptional regulation of miRNA biogenesis and functions. *Frontiers in Biology*. 2010;**5**(1):32-40. DOI: 10.1007/s11515-010-0004-y
- [17] Büssing I, Yang JS, Lai EC, Grosshans H. The nuclear export receptor XPO-1 supports primary miRNA processing in *C. elegans* and *drosophila*. *EMBO Journal*. 2014;**29**(11):1830-1839. DOI: 10.1038/emboj.2010.82
- [18] Kawahara H, Imai T, Okano H. MicroRNAs in neural stem cells and neurogenesis. *Frontiers in Neuroscience*. 2012;**6**:30. DOI: 10.3389/fnins.2012.00030
- [19] Wu K, He J, Pu W, Yong P. The role of exportin-5 in MicroRNA biogenesis and cancer. *Genomics, Proteomics & Bioinformatics*. 2018;**65**(5):42-46. DOI: 10.1016/j.gpb.2017.09.004
- [20] Singh RP, Massachi I, Manickavel S, Singh S, Rao NP, Hasan S, et al. The role of miRNA in inflammation and autoimmunity. *Autoimmunity Reviews*. 2013;**12**(12):1160-1165. DOI: 10.1016/j.autrev.2013.07.003
- [21] Williams MD, Mitchell GM. MicroRNAs in insulin resistance and obesity. *Experimental Diabetes Research*. 2015;**2012**(3):484696. DOI: 10.1155/2012/484696
- [22] Honardoost M, Sarookhani MR, Arefian E, Soleimani M. Insulin resistance associated genes and miRNAs. *Applied Biochemistry and Biotechnology*. 2014;**174**(1):63-80. DOI: 10.1007/s12010-014-1014-z
- [23] Leti F, Taila M, Distefano JK. The role of ncRNA in diabetes[M]. In: *MicroRNAs and Other Non-Coding RNAs in Inflammation*. 2015
- [24] Saihua Z, Xuelian L, Gynecology DO. Expression and effect of MicroRNA in patients with polycystic ovary syndrome. *International Journal of Reproductive Health/Family Planning*. 2016;**35**(2):146-150
- [25] Qi P, Xiaoyan Z, Xiang D. Circulating long non-coding RNAs in cancer: Current status and future perspectives. *Molecular Cancer*. 2016;**15**(1):39. DOI: 10.1186/s12943-016-0524-4
- [26] Zhiping Z, Jian L, Yunchao W, Huan Q, Weiwu G, Chenhui W, et al. Effect of LncRNA LOC100506123 on proliferation and migration in pancreatic cancer cells. *Journal of Third Military Medical University*. 2017;**39**(9):840-845. DOI: 10.16016/j.1000-5404.201611074
- [27] Ping P, Wang L, Kuang L, Ye S, MFB I, Pei T. A novel method for LncRNA-disease association prediction based on an lncRNA-disease association network. *IEEE/ACM Transactions on Computational Biology and Bioinformatics*. 2019;**16**(2):688-693. DOI: 10.1109/TCBB.2018.2827373
- [28] Xin H, Cuifang H, Hongchu B, Meimei W, Huangguan D. Aberrant expression of long noncoding RNAs in cumulus cells isolated from PCOS patients. *Journal of Assisted Reproduction and Genetics*. 2016;**33**(1):111-121. DOI: 10.1007/s10815-015-0630-z

- [29] Yudong L, Ying L, Shuxian F, Desheng Y, Xin C, Xingyu Z, et al. Long noncoding RNAs: Potential regulators involved in the pathogenesis of polycystic ovary syndrome. *Endocrinology*. 2017;**158**(11):3890-3899. DOI: 10.1210/en.2017-00605
- [30] Munakata Y, Ichinose T, Ogawa K, tami N, Tasaki H, Shirasuna K, et al. Relationship between the number of cells surrounding oocytes and energy states of oocytes. *Theriogenology*. 2016;**86**(7):1789-1798.e1. DOI: 10.1016/j.theriogenology.2016.05.036
- [31] Zhengbin H, Guocheng L, Yanguang W, Dong H, Weiguo F, Junzuo W, et al. Interactive effects of granulosa cell apoptosis, follicle size, cumulus-oocyte complex morphology, and cumulus expansion on the developmental competence of goat oocytes: A study using the well-in-drop culture system. *Reproduction*. 2006;**132**(5):749-758. DOI: 10.1530/REP-06-0055
- [32] Kim E, Seok HH, Lee SY, Lee DR, Moon J, Yoon TK, et al. Correlation between expression of glucose transporters in granulosa cells and oocyte quality in women with polycystic ovary syndrome. *Endocrinology & Metabolism*. 2014;**29**(1):40-47. DOI: 10.3803/EnM.2014.29.1.40
- [33] Ashkenazi J, Farhi J, Orvieto R, Homburg R, Dekel A, Feldberg D, et al. Polycystic ovary syndrome patients as oocyte donors: The effect of ovarian stimulation protocol on the implantation rate of the recipient. *Fertility and Sterility*. 1995;**64**(3):564-567. DOI: 10.1016/s0015-0282(16)57793-0
- [34] Jabara S, Coutifaris C. In vitro fertilization in the PCOS patient: Clinical considerations. *Seminars in Reproductive Medicine*. 2003;**21**(03):317-324. DOI: 10.1055/s-2003-43310
- [35] Li R, Qiao J. The different endometrial receptivity between the normal women and PCOS patients in the implantation window. *Chinese Journal of Birth Health & Heredity*. 2007;**84**(3):S456-S456. DOI: 10.1016/j.fertnstert.2005.07.1193
- [36] Wood JR, Dumesic DA, Abbott DH, Strauss JF. Molecular abnormalities in oocytes from women with polycystic ovary syndrome revealed by microarray analysis. *The Journal of Clinical Endocrinology and Metabolism*. 2007;**92**(2):705-713. DOI: 10.1210/jc.2006-2123
- [37] Klein NA, Battaglia DE, Miller PB, Branigan EF, Giudice LC, Soules MR. Ovarian follicular development and the follicular fluid hormones and growth factors in normal women of advanced reproductive age. *The Journal of Clinical Endocrinology & Metabolism*. 1996;**81**(5):1946-1951. DOI: 10.1210/jcem.81.5.8626862
- [38] Ciaraldi TP. Molecular defects of insulin action in the polycystic ovary syndrome: Possible tissue specificity. *Journal of Pediatric Endocrinology & Metabolism*. 2000;**13**(Suppl 5):1291-1293. DOI: 10.1046/j.1440-1754.2000.00616.x
- [39] Diamanti-Kandaraki E, Argyrakopoulou G, Economou F, Kandaraki E, Koutsilieris M. Defects in insulin signaling pathways in ovarian steroidogenesis and other tissues in polycystic ovary syndrome (PCOS). *The Journal of Steroid Biochemistry and Molecular Biology*. 2008;**109**(3-5):242-246. DOI: 10.1016/j.jsbmb.2008.03.014
- [40] Xiaoke W, Risto E. Ovarian insulin resistance and insulin sensitizer effect on polycystic ovary syndrome. *Zhonghua Fu Chan Ke Za Zhi*. 2005;**39**(12):804-808. DOI: <http://dx.Doi.org/>
- [41] Li D, Xu D, Xu Y, Chen L, Li C, Dai X, et al. MicroRNA-141-3p

targets DAPK1 and inhibits apoptosis in rat ovarian granulosa cells. *Cell Biochemistry and Function*. 2017;**35**:197-201. DOI: 10.1002/cbf.3248

[42] Lulu F, Ying X, Dandan L, Xiaowei D, Xin X, Jingshun Z, et al. Expression profiles of mRNA and long noncoding RNA in the ovaries of letrozole-induced polycystic ovary syndrome rat model through deep sequencing. *Gene*. 2018;**657**:19-29. DOI: 10.1016/j.gene.2018.03.002

[43] Hua X, Xiaoyan Y, Boqun X. Involvement of MiRNA in pathogenesis of polycystic ovary syndrome. *International Journal of Reproductive Health/Family Planning*. 2017;**1**:66-69. DOI: 10.3969/j.issn.1674-1889.2017.01.018

[44] B Xu, YW Zhang, XH Tong, YS Liu. Characterization of microRNA profile in human cumulus granulosa cells: Identification of microRNAs that regulate notch signaling and are associated with PCOS. *Molecular and Cellular Endocrinology*. 2015;**404**:26-36. DOI: 10.1016/j.mce.2015.01.030

[45] Xueying Z, Ying X, Lulu F, Dandan L. Identification of mRNAs related to endometrium function regulated by lncRNA CD36-005 in rat endometrial stromal cells. *Reproductive Biology and Endocrinology*. 2018;**16**(1):96. DOI: 10.1186/s12958-018-0412-4

[46] Ruizhi F, Qing S, Yan Z, Wei F, Miao L, Yan X, et al. MiRNA-320 in the human follicular fluid is associated with embryo quality in vivo and affects mouse embryonic development in vitro. *Scientific Reports*. 2015;**5**:8689. DOI: 10.1038/srep08689

[47] Mianmian Y, Xiaorong W, Guidong Y, Mingrong L, Meng L, Yingpu S, et al. Transactivation of miR-320 by miR-383 regulates granulosa cell functions by targeting E2F1 and SF-1. *Journal of Biological Chemistry*.

2014;**289**(26):18239-18257. DOI: 10.1074/jbc.M113.546044

[48] Chenling Z, Hui W, Changyou Y, Xuefeng G, Xiaojuan L. Dereglulation of RUNX2 by miR-320a deficiency impairs steroidogenesis in cumulus granulosa cells from polycystic ovary syndrome (PCOS) patients. *Biochemical and Biophysical Research Communications*. 2017;**482**(4):1469-1476. DOI: 10.1016/j.bbrc.2016.12.059

[49] Sørensen Rensen A, Louise Wissing M, Salö S, Lis Mikkelsen Englund A. MicroRNAs related to polycystic ovary syndrome (PCOS). *Genes*. 2014;**5**(3):684-708. DOI: 10.3390/genes5030684

[50] Naji M, Nekoonam S, Aleyasin A, Arefian E, Mahdian R, Azizi E, et al. Expression of miR-15a, miR-145, and miR-182 in granulosa-lutein cells, follicular fluid, and serum of women with polycystic ovary syndrome (PCOS). *Archives of Gynecology and Obstetrics*. 2017;**297**(1):221-231. DOI: 10.1007/s00404-017-4570-y