we are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists



122,000

135M



Our authors are among the

TOP 1%





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



Chapter

Antioxidant Activity of Faba Bean Extracts

Wojciech Rybiński, Magdalena Karamać, Katarzyna Sulewska and Ryszard Amarowicz

Abstract

Phenolic compounds were extracted from seeds of 22 cultivars of faba bean (Vicia faba L.) by using 80% (v/v) aqueous acetone. The total phenolic compound and condensed tannins contents of the extracts and their antioxidant activity were determined using the Folin-Ciocalteu's phenol reagent, vanillin/HCl method, and ABTS and FRAP assays, respectively. The content of total phenolic compounds ranged from 40.7 to 66.1 mg/g extract and from 326 to 574 mg/100 g seeds. Contents of condensed tannins ranged from 2.40 to 49.9 mg/g extract and from 22.2 (FAB) to 365 mg/100 seeds. The extracts and seeds were characterized by Trolox equivalent antioxidant capacity (TEAC) values ranging from 0.550 (FAB 443) to 1.030 mmol Trolox/g extract (FAB 187) and from 4.85 (FAB 318) to 9.81 mmol Trolox/100 g seeds (FAB 187). Ferric-reducing antioxidant power (FRAP) values varied from 0.595 (FAB 443) to 0.908 mmol Fe^{2+}/g extract (FAB 5023) and from 4.61 (FAB 297) to 7.90 mmol $Fe^{2+}/100$ g seeds (FAB 187). The total phenolic content of faba bean extract was correlated with the results of the ABTS (r = 0.864) and FRAP (r = 0.862) assays. The coefficients of correlations between the contents of condensed tannins and ABTS and FRAP results were 0.543 and 0.862. We also noted a correlation between results of ABTS and FRAP assays (r = 0.795).

Keywords: faba bean, *Vicia faba* L., phenolic compounds, antioxidant activity, ABTS assay, DPPH assay

1. Introduction

Phenolic compounds of plant origin can inhibit or delay the oxidation of nutrients present in food products. In the human organism, natural antioxidants can protect lipids, proteins, and DNA against reactive oxygen and nitrogen species (ROS, RNS) [1, 2]. Results of numerous research groups demonstrate the protective effect of consuming phenolic-rich grains, legumes, oilseeds, fruits, berries, and nuts against several chronic diseases [3–7].

In human nutrition, legumes are an important source of proteins, starch, oligosaccharides (prebiotics), dietary fibers, vitamins, and minerals [8, 9]. As a rich source of natural antioxidants, legumes can play an important role in cardio and cancer protection [5, 10–15].

Faba bean (*Vicia faba*) is a species of Fabaceae family. It is native to South America, North Africa, and southwest and south Asia and is extensively cultivated elsewhere. Similar to other legumes, faba bean seeds contain phenolic compounds

[16] including condensed tannins [17–19]. The antioxidant potential of faba bean was determined using DPPH, FRAP, and ORAC assays [17, 20, 21]. Very high antioxidant capacity of *Vicia faba* sprouts was confirmed by Okumura et al. [22].

2. Experimental

2.1 Material

Plant material consisted of a collection of 22 faba bean accessions derived from Syria, Morocco, Tunisia, Sudan, Egypt, Yemen, Israel, Georgia, Azerbaijan, Tajikistan, Mongolia, Afghanistan, India, Australia, and Poland (**Table 1**). Information about seed coat color and 100 seed weight was provided on the basis of results obtained in a field experiment conducted in Cerekwica (Poland, 51°55′N, 17°21′E).

Description of faba bean seeds is reported in Table 1.

2.2 Chemicals

Ferrous chloride, sodium persulfate, ferrous chloride, 2,4,6-tri(2-pyridyl)*s*-triazine (TPTZ), 2,2'-azinobis-(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS),

No.	Accession code	Country of origin	Seeds coat color	Weight of 100 seeds (g)
1	FAB 337	Syria	Dark	47.2
2	FAB 443	Syria	Bright	76.6
3	FAB 5023	Morocco	Bright	45.8
4	FAB 5019	Morocco	Bright	52.1
5	FAB 6440	Tunisia	Bright	68.0
6	FAB 6441	Tunisia	Bright	65.0
7	FAB 225	Sudan	Bright	64.7
8	FAB 297	Sudan	Bright	45.0
9	FAB 6474	Egypt	Bright	27.4
10	FAB 219	Yemen	Bright	57.0
11	FAB 6318	Israel	Bright	81.5
12	FAB 344	Israel	Dark	46.8
13	FAB 604	Georgia	Bright	66.3
14	FAB 294	Azerbaijan	Bright	43.4
15	FAB 354	Tajikistan	Bright	40.0
16	FAB 202	Mongolia	Bright	68.4
17	FAB 144	Afghanistan	Dark	65.5
18	FAB 187	Afghanistan	Bright	30.9
19	FAB 250	India	Bright	38.6
20	FAB 446	India	Bright	41.1
21	FAB 7077	Australia	Bright	72.5
22	Martin	Poland	Bright	55.8

Table 1.Description of faba bean seeds.

6-hydroxy-2,5,7,8-tetramethyl-chroman-2-carboxylic acid (Trolox), Folin– Ciocalteu's phenol reagent, and (+)-catechin were purchased from Sigma (Poznań, Poland). Methanol, acetone, and hexane were obtained from P.O.Ch. Company (Gliwice, Poland).

2.3 Extraction

Phenolic compounds were extracted from grounded and defatted with hexane faba bean seed by using 80% acetone (v/v) at a solid to solvent ratio of 1:10 (w/v), for 15 min at 50°C [23]. Extraction was carried out in flasks placed in a shaking water bath (Elpan 357, Wrocław, Poland). Acetone from the combined extract was evaporated using a Büchi rotary evaporator. The sample was then freeze-dried.

2.4 Determination of total phenolic compound contents

The content of total phenolic compounds in the extracts was determined using the Folin–Ciocalteu's phenol reagent [24]. The results were expressed as (+)-cat-echin equivalents per g of the extract and per 100 g of seeds.

2.5 Determination of condensed tannins content

The content of condensed tannins was determined using a vanillin/HCl colorimetric method [25]. The results obtained were reported as mg catechin equivalent per g extract and 100 g of seeds.

2.6 ABTS assay

The Trolox equivalent antioxidant capacity (TEAC) was determined using the method of Re et al. [26]. The results obtained were reported as mmol Trolox equivalents per g extract and 100 g of seeds.

2.7 FRAP assay

The ferric-reducing antioxidant power (FRAP) assay was performed as previously described by Benzie and Strain [27]. The results obtained were reported as mmol Fe²⁺ equivalents per g extract and 100 g of seeds.

2.8 Statistical analysis

The results are reported as a mean value of three determinations ± standard deviation. The Pearson correlation was used to determine the relation between results of total phenolics, condensed tannins, TEAC, and FRAP values. Moreover the principal component analysis (PCA) and hierarchical cluster analysis (HCA) with Ward's method using Euclidean distances were conducted. Statistical and chemometric analyses of data were performed using the Statistica software (Windows software package 8.0).

3. Results and discussion

3.1 Content of total phenolics and condensed tannins

The content of total phenolics in the extracts was determined using the Folin– Ciocalteu's phenol reagent. The results were expressed as (+)-catechin equivalents per g of the extract or 100 g seeds. The results are reported in **Tables 2** and **3**. The content of total phenolics ranged from 40.7 (FAB 219) to 66.1 mg/g extract (FAB 187) and from 326 (FAB 219) to 574 mg/100 g seeds (FAB 5019).

The content of condensed tannins in the extracts was determined using the vanillin/HCl colorimetric method. The results ranged from 2.40 (FAB 297) to 49.9 mg/g extract (FAB 225) and from 22.2 (FAB 297) to 441 mg/100 seeds (FAB 5019) (**Tables 1** and **3**).

The results obtained in this research confirm the fact that faba bean is a rich source of phenolic compounds as well as condensed tannins. A lower content of total phenolic compounds was previously reported for extracts of pea [28], white bean [29], broad bean [23, 30] lupin [31], and grass pea [32]. A similarly high content of total phenolic compounds was previously reported for red lentil [33], green lentil [34], red bean [35], and adzuki bean [36].

The presence of condensed tannins in faba bean seeds determined with the vanillin method was reported by Amarowicz et al. [37], Baginsky et al. [38], Amarowicz and Shahidi [39], and Zduńczyk et al. [17]. In a research conducted by

No	Total phenolics ^a (mg/g extract)	Condensed tannins ^b (mg/g extract)	TEAC (mmol TE/g extract)	FRAP (mmol Fe ²⁺ /g extract)
1	51.5 ± 0.4	26.4 ± 0.7	0.656 ± 0.014	0.673 ± 0.014
2	42.1 ± 1.0	10.9 ± 0.9	0.550 ± 0.016	0.535 ± 0.010
3	61.2 ± 0.5	25.9 ± 1.4	0.860 ± 0.011	0.908 ± 0.012
4	62.8 ± 2.4	48.2 ± 1.3	0.854 ± 0.022	0.753 ± 0.008
5	60.5 ± 1.6	43.2 ± 1.8	0.775 ± 0.023	0.742 ± 0.011
6	47.6 ± 0.2	21.5 ± 1.1	0.563 ± 0.008	0.634 ± 014
7	60.2 ± 0.7	49.9 ± 0.7	0.886 ± 0.017	0.715 ± 0.015
8	46.9 ± 0.9	2.40 ± 0.72	0.710 ± 0.018	0.572 ± 0.013
9	41.8 ± 0.4	8.77 ± 1.06	0.669 ± 0.014	0.636 ± 0.009
10	40.7 ± 1.4	4.32 ± 0.78	0.661 ± 0.012	0.603 ± 0.016
11	45.5 ± 0.3	4.15 ± 0.69	0.592 ± 0.011	0.651 ± 0.014
12	55.0 ± 1.6	22.1 ± 1.6	0.826 ± 0.022	0.734 ± 0.012
13	59.1 ± 1.1	21.3 ± 1.5	0.806 ± 0.020	0.807 ± 0.011
14	46.8 ± 1.0	35.3 ± 2.1	0.691 ± 0.004	0.671 ± 0.011
15	61.1 ± 1.4	49.1 ± 1.4	0.844 ± 0.017	0.824 ± 0.014
16	51.7 ± 1.5	28.2 ± 1.3	0.841 ± 0.024	0.703 ± 0.012
17	46.3 ± 1.5	23.8 ± 1.4	0.721 ± 0.014	0.588 ± 0.008
18	66.1 ± 1.9	32.5 ± 1.3	1.035 ± 0.014	0.833 ± 0.009
19	47.9 ± 1.0	14.4 ± 0.8	0.631 ± 0.011	0.718 ± 0.013
20	48.8 ± 1.7	47.8 ± 2.3	0.694 ± 0.009	0.675 ± 0.010
21	48.7 ± 0.6	34.2 ± 1.5	0.631 ± 0.010	0.694 ± 0.015
22	65.7 ± 1.9	39.9 ± 1.1	0.894 ± 0.015	0.807 ± 0.007

Table 2.

Contents of total phenolic compounds and condensed tannins and antioxidant activity of faba bean extracts. Results are reported per g of extract.

No	Total phenolics ^a (mg/100 g seeds)	Condensed tannins ^b (mg/100 g seeds)	TEAC (mmol TE/100 g seeds)	FRAP (mmol Fe ²⁺ /100 g seeds
1	484 ± 4	247 ± 6	6.16 ± 0.13	6.32 ± 0.13
2	421 ± 10	109 ± 19	5.50 ± 0.16	5.35 ± 0.10
3	524 ± 5	222 ± 12	7.37 ± 0.09	7.78 ± 0.10
4	574 ± 22	441 ± 12	7.80 ± 0.20	6.88 ± 0.08
5	509 ± 13	362 ± 15	6.51 ± 0.20	6.23 ± 0.09
6	425 ± 2	192 ± 9	5.82 ± 0.07	5.66 ± 0.13
7	539 ± 6	447 ± 6	7.94 ± 0.15	6.40 ± 0.13
8	378 ± 7	22.2 ± 1.0	5.72 ± 0.15	4.61 ± 0.10
9	395 ± 4	82.8 ± 9.9	6.31 ± 0.13	6.01 ± 0.08
10	326 ± 11	34.5 ± 7.5	5.28 ± 0.10	4.82 ± 0.13
11	373 ± 2	34.0 ± 4.9	4.85 ± 0.09	5.34 ± 0.11
12	475 ± 13	191 ± 14	7.13 ± 0.19	6.34 ± 0.10
13	559 ± 9	201 ± 14	7.63 ± 0.19	7.64 ± 0.10
14	410 ± 9	309 ± 18	6.04 ± 0.03	5.85 ± 0.12
15	561 ± 13	451 ± 13	7.75 ± 0.16	7.57 ± 0.11
16	486 ± 14	265 ± 12	7.91 ± 0.22	6.61 ± 0.08
17	421 ± 14	217 ± 12	6.56 ± 0.13	5.34 ± 0.08
18	626 ± 18	308 ± 13	9.81 ± 0.14	7.90 ± 0.12
19	418 ± 9	126 ± 7	5.52 ± 0.09	6.28 ± 0.09
20	482 ± 17	471 ± 23	6.85 ± 0.09	6.66 ± 0.15
21	464 ± 6	326 ± 14	6.01 ± 0.10	6.60 ± 0.07
22	602 ± 17	365 ± 10	8.20 ± 0.14	7.40 ± 0.10

^bCatechin equivalents.

Table 3.

Contents of total phenolic compounds and condensed tannins and antioxidant activity of faba bean seeds. Results are reported per 100 g of seeds.

Luo et al. [40], the content of condensed tannins in faba bean extracts ranged from 0.9 to 1.9 g of gallic acid equivalents/100 g extract. In this research, the authors used the Folin-Ciocalteu's phenol reagent to determine the content of tannins precipitated from the extract by using polypyrrolidone (PVPP). Amarowicz and Shahidi [39] identified gallate procyanidin dimer and three gallate procyanidins in faba bean extract by using HPLC-DAD-MS. The content of the abovementioned compounds was 689, 89.8, 28.6, and 18.3 μ g/g extract. Five procyanidin dimmers and three procyanidin trimers were determined in faba bean extract using an UHPLC-ESI-QTOF-MS method [41]. The presence of procyanidin B1, B2, B3, B4, C1, and C2 has been reported by De Pascual-Teresa et al. [42].

3.2 Antioxidant activity

Antioxidant properties of the extracts were investigated using ABTS and FRAP assays. The extracts and seeds were characterized by the Trolox equivalent antioxidant capacity (TEAC) values ranging from 0.550 (FAB 443) to 1.030 mmol Trolox/g extract (FAB 187) and from 4.85 (FAB 318) to 9.81 mmol Trolox/100 g

seeds (FAB 187). Ferric-reducing antioxidant power (FRAP) values varied from 0.595 (FAB 443) to 0.908 mmol Fe^{2+}/g extract (FAB 5023) and from 4.61 (FAB 297) to 7.90 mmol $Fe^{2+}/100$ g seeds (FAB 187).

The results of ABTS assay obtained in this study for faba bean extracts were much higher than those reported before for extracts of grass pea (0.017-0.037 mmol Trolox/g) [32], cow pea (0.285-0.665 mmol Trolox/g) [43], white bean (0.0270-0.043 mmol Trolox/g) [29], mung bean (0.021-0.031 mmol Trolox/g) [35], and lupin (0.260-0.620 mmol Trolox/g) [31]. The results of FRAP assay were also much higher than those reported for extracts of grass pea $(0.045-0.120 \text{ mmol} \text{Fe}^{2+}/\text{g})$, [32], cow pea $(0.487-1.566 \text{ mmol} \text{Fe}^{2+}/\text{g})$ [43], white bean $(0.066-0.089 \text{ mmol} \text{Fe}^{2+}/\text{g})$ [29], and lupin 0.046-0.064 [31].

3.3 Statistical analysis

In our study, for the first time, a correlation was calculated between the content of phenolic compounds in the faba bean extracts and their antioxidant activity. The

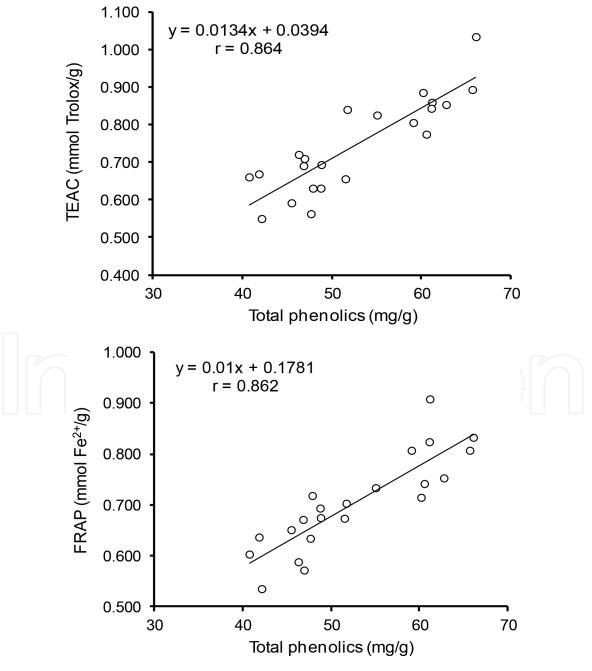


Figure 1. Correlation between the total phenolic contents and the results of ABTS and FRAP assays.

coefficients of correlation between the total phenolic content and the results of the ABTS and FRAP assays were 0.864 and 0.862, respectively (**Figure 1**). The correlations between the content of condensed tannins and results of ABTS and FRAP assays were weaker and characterized by r = 0.543 and r = 0.528 (**Figure 2**). A correlation was also observed between the results of both assays (r = 0.795) (**Figure 3**).

In our previous study, we noted a correlation between the total phenolic content and ABTS and FRAP results determined for red bean (r = 0.997 and 0.997, respectively) [35], white (r = 0.480 and 0.850, respectively) [29], and grass pea (r = 0.881and 0.781, respectively) [32]. Statistically significant correlations between the content of total phenolics and TEAC as well as between the content of condensed tannins and TEAC (r = 0.857 and 0.787, respectively) were reported for extracts obtained from seeds of faba bean, broad bean, adzuki bean, red bean, pea, red lentil, and green lentil [44].

In the principal component analysis (PCA) (**Figure 4**), the two first components accounted for 93.6% of the total variability between the faba bean cultivars. The analysis includes the content of total phenolic compounds and condensed tannins

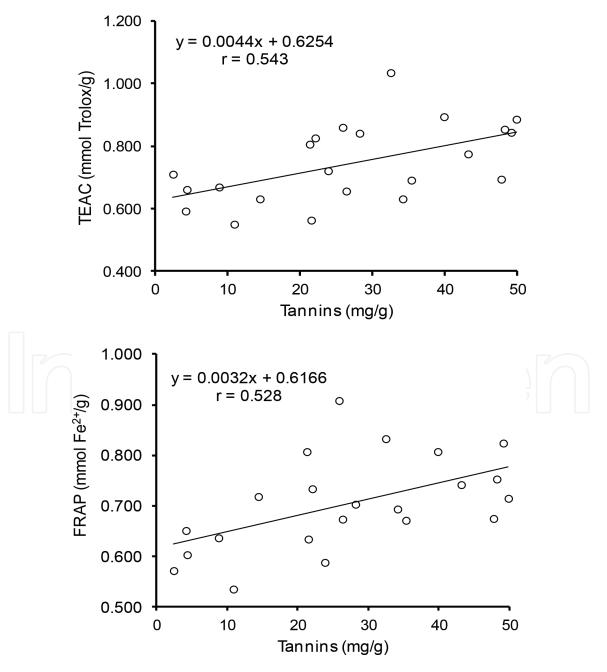


Figure 2. Correlation between the content of tannins and the results of ABTS and FRAP assays.

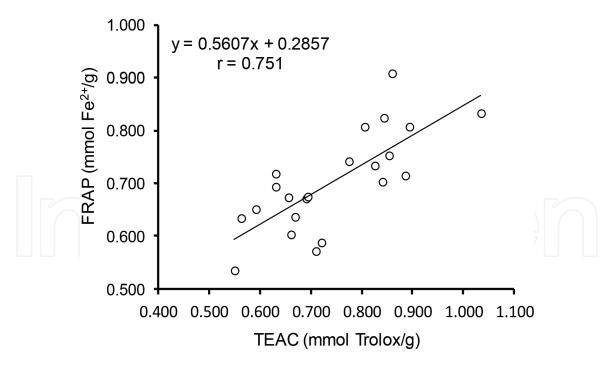
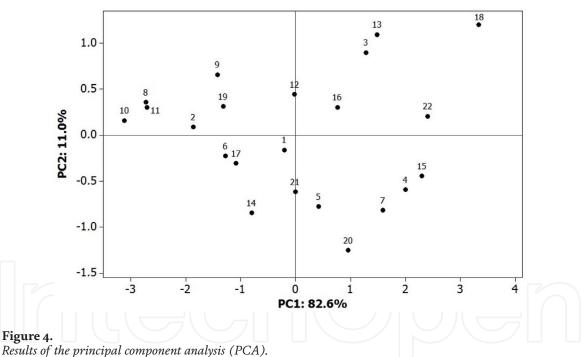


Figure 3. *Correlation between the results of FRAP and ABTS assays.*



in seeds as well as their antioxidant potential determined using ABTS and FRAP methods. A considerable variability in terms of the analyzed traits expressed jointly with the grates Mahalanobis distance was recorded for sample 18 from Afghanistan (FAB 187) and samples 8, 10, and 11 from Sudan (FAB 297), Yemen (FAB 219), and Israel (FAB 6318). The close clustering was observed for seeds from Sudan, Yemen, and Afghanistan and for seeds from Morocco, Sudan, and Tajikistan (sample 4, 7, and 15; FAB 5019, FAB 225, and FAB 354).

The hierarchical cluster analysis (**Figure 5**) showed several pairs of faba bean accessions (e.g., FAB 337 and FAB 202, FAB 5023 and FAB 604). Two main clusters were observed. The first contained 16 accessions, whereas the second contained only 6. The presence of similar pairs of faba bean accessions from different countries confirms the limitation of the hierarchical cluster analysis in the discrimination of the geographical origin of samples.

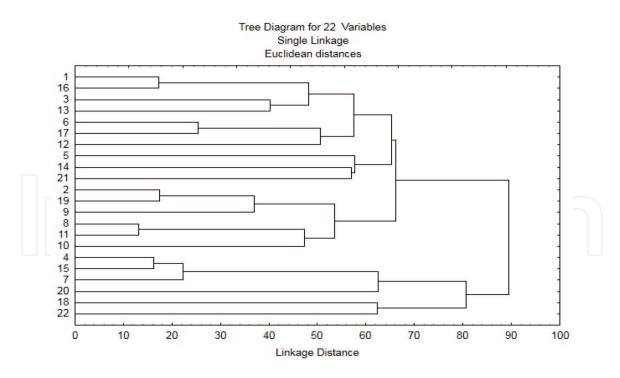


Figure 5. *The hierarchical cluster analysis.*

4. Conclusions

The extracts obtained from faba bean were characterized by a high content of phenolic compounds and condensed tannins. Their antioxidant potential was higher than that reported previously for the extracts of other legume seeds. The content of total phenolics and condensed tannins in the faba bean extracts strongly influenced the antioxidant activity of extracts determined using ABTS and FRAP assays.

Acknowledgements

The technical assistance of Kamila Penkacik is acknowledged.

Conflict of interest

The authors declare no conflict of interest.

Intechopen

Author details

Wojciech Rybiński¹, Magdalena Karamać², Katarzyna Sulewska² and Ryszard Amarowicz^{2*}

1 Institute of Plant Genetics, Polish Academy of Sciences, Poznań, Poland

2 Institute of Animal Reproduction and Food Research, Polish Academy of Sciences, Olsztyn, Poland

*Address all correspondence to: r.amarowicz@pan.olsztyn.pl

IntechOpen

© 2019 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

References

[1] Halliwell B, Gutteridge JC, Cross CE. Free radicals, antioxidants, and human disease: Where are we now? Journal of Laboratory and Clinical Medicine. 1992;**119**:598-620

[2] Willett WC. Diet and health: What should we eat? Science. 1994;**264**: 532-537. DOI: 10.1126/science.8160011

[3] Zhang B, Peng H, Deng Z, Tsao R. Phytochemicals of lentil (*Lens culinaris*) and their antioxidant and antiinflammatory effects. Journal of Food Bioactives. 2018, 2018;1:93-103. DOI: 10.31665/JFB.2018.1128

[4] Chang SK, Alasalvar C,
Shahidi F. Review of dried fruits:
Phytochemicals, antioxidant efficacies,
and health benefits. Journal of
Functional Foods. 2016, 2016;21:
113-132. DOI: 10.1016/j.jff.2015.11.034

[5] Shahidi F, Ambigaipalan P. Phenolics and polyphenolics in foods, beverages and spices: Antioxidant activity and health effects–A review. Journal of Functional Foods. 2015;**18**:820-897. DOI: 10.1016/j. jff.2015.06.018

[6] Alshikh N, de Camargo AC, Shahidi F. Phenolics of selected lentil cultivars: Antioxidant activities and inhibition of low-density lipoprotein and DNA damage. Journal of Functional Foods. 2016;**18**:1022-1038. DOI: 10.1016/j. jff.2015.06.018

[7] Shahidi F, Chandrasekara A. Millet grain phenolics and their role in disease risk reduction and health promotion: A review. Journal of Functional Foods. 2013;5:570-581. DOI: 10.1016/j. jff.2013.02.004

[8] Flight I, Clifton P. Cereal grains and legumes in the prevention of coronary heart disease and stroke: A review of the literature. European Journal of Human Nutrition. 2006;**60**:1145-1159. DOI: 10.1038/sj.ejcn.1602435

[9] Vaz Patto MC, Amarowicz R, Aryee ANA, Boye JI, Chung H-J, Martín-Cabrejas MA, et al. Achievements and challenges in improving the nutritional quality of food legumes. Critical Reviews in Plant Sciences. 2015;**34**:105-143. DOI: 10.1080/07352689.2014.897907

[10] Bouchenak M, Lamri-Senhadji M. Nutritional quality of legumes, and their role in cardiometabolic risk prevention: A review. Journal of Meducinal Foods. 2013;**16**:185-198. DOI: 10.1089/jmf.2011.0238

[11] Madhujith T, Shahidi F. Antioxidant potential of pea beans (*Phaseolus vulgaris* L.). Journal of Food Science.
2005;**70**:S85-S95. DOI: 10.1111/j.1365-2621.2005.tb09071.x

[12] Aparicio-Fernandez X, Reynoso-Camacho R, Castano-Tostado E, Garcia-Gasca T, Mejia SH, Guzman-Maldonado SH. Antiradical capacity and induction of apoptosis on HeLa cells by a *Phaseolus vulgaris* extract. Plant Foods for Human Nutrition. 2008;**63**:35-40. DOI: 10.1007/ s11130-007-0066-4

[13] Peeters PHM, Boker LK, van der Schouw YT, Grobbee DE. Phytoestrogens and breast cancer risk. Review of the epidemiologic evidence. Breast Cancer Research and Treatment. 2013;77:171-183. DOI: 10.1186/bcr628

[14] Kolonel LN, Hankin JH, Whittemore AS, Wu AH, Gallagher RP, Wilkens LR, et al. Vegetables, fruits, legumes and prostate cancer: A multicenter case-control study. Cancer Cancer Epidemiol Biomarkers 2000;9: 795-804 [15] Kushi LH, Meyer KA, Jacobs DR
Jr. Cereals, legumes, and chronic disease risk reduction: Evidence from epidemiological studies. The American Journal of Clinical Nutrition.
1999;70:451S-458S. DOI: 10.1093/ ajcn/70.3.451s

[16] Turco I, Ferretti G, Bacchetti T.
Review of the health benefits of faba bean (*Vicia faba* L.) polyphenols.
Journal of Food and Nutrition Research.
2016;55:283-293

[17] Zduńczyk Z, Mikulski D, Jankowski J, Przybylska-Górnowicz B, Sosnowska E, Juśkiewicz J, et al. Effects of dietary inclusion of high- and lowtannin faba bean (*Vicia faba* L.) seeds on microbiota, histology and fermentation processes of the gastrointestinal tract in finisher turkeys. Animal Nutrition and Feed Science and Technology. 2018;**240**:184-196. DOI: 10.1016/j.anifeedsci.2018.04.006

[18] Makkar HPS, Becker K, Abel HJ, Pawlicki E. Nutrient contents, rumen protein degradability and antinutritional factors in some colour- and white-flowering cultivars of *Vicia faba* beans. Journal of the Science of Food and Agriculture. 1997;75:511-520. DOI: 10.1002/ (SICI)1097-0010(199712)75:4<511::AID-JSFA907>3.0.CO;2-M

[19] Khalil AH, Mansour EH. The effect of cooking, autoclaving and germination on the nutritional quality of faba beans. Food Chemistry. 1995;**54**:177-182. DOI: 10.1016/0308-8146(95)00024-D

[20] Khan MA, Ammar MH, Migdadi HM, El-Harty EH, Osman MA, Farooq M, et al. Comparative nutritional profiles of various faba bean and chickpea genotypes. International Journal of Agriculture and Biology. 2015;**17**: 449-457. DOI: 10.17957/IJAB/17.3.14.990

[21] Siah S, Konczak I, Wood JA, Agboola S, Blanchard CL. Effect on phenolic composition and in vitro antioxidant capacity of Australian grown faba beans (*Vicia faba* L.). Plant Foods for Human Nutrition. 2014;**69**:85-91

[22] Okumura K, Hosoya T,
Kawarazaki K, Izawa N, Kumazawa S.
Antioxidant activity of phenolic compounds from fava bean sprouts. Journal of Food Science.
2016;81:C1394-C1398. DOI: 10.1111/1750-3841.13330

[23] Amarowicz R, Shahidi F. Antioxidant activity of broad bean seed extract and its phenolic composition. Journal of Functional Foods. 2017, 2017;**38**:656-662. DOI: 10.1016/j. jff.2017.04.002

[24] Naczk M, Shahidi F. The effect of methanol-ammonia-water treatment on the content of phenolic acids of canola. Food Chemistry. 1989;**31**:159-164. DOI: 10.1016/0308-8146(89)90026-5

[25] Price ML, van Scoyoc S, Butler LG. A critical evaluation of the vanillin reaction as an assay for tannin in sorghum grain. Journal of Agricultural and Food Chemistry. 1978;**26**:1214-1218. DOI: 10.1021/jf60219a031

[26] Re R, Pellegrini N, Proteggente A, Pannala A, Yang M, Rice-Evans C. Antioxidant activity applying an improved ABTS radical cation decolorization assay. Free Radical Biology & Medicine. 1999;**26**:1231-1237. DOI: 10.1016/S0891-5849(98)00315-3

[27] Benzie IFF, Strain JJ. The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": The FRAP assay. Analytical Biochemistry. 1996;**239**:70-76. DOI: 10.1006/ abio.1996.0292

[28] Amarowicz R, Troszyńska A. Antioxidant activity of extract of pea and its fractions of low molecular phenolics and tannins. Polish Journal

of Food and Nutrition Sciences. 2003;**53**:10-15

[29] Orak HH, Karamać M, Orak A, Amarowicz R. Antioxidant potential and phenolic compounds of some widely consumed Turkish white bean (*Phaseolus vulgaris* L.) varieties. Polish Journal of Food and Nutrition Sciences. 2016;**66**:253-260. DOI: 10.1515/ pjfns-2016-0022

[30] Boudjou S, Oomah BD, Zaidi F, Hosseinian F. Phenolics content and antioxidant and anti-inflammatory activities of legume fractions. Food Chemistry. 2013;**138**:1543-1550. DOI: 10.1016/j.foodchem.2012.11.108

[31] Karamać M, Orak HH, Amarowicz R, Orak A, Piekoszewski W. Phenolic contents and antioxidant capacities of wild and cultivated white lupin (*Lupinus albus* L.) seeds. Food Chemistry. 2018;**258**:1-7. DOI: 10.1016/j. foodchem.2018.03.041

[32] Rybiński W, Karamać M, Sulewska K, Börner A, Amarowicz R. Antioxidant potential of grass pea seeds from European countries. Food. 2018, 2018;7(9):142. DOI: 10.3390/ foods7090142

[33] Amarowicz R, Estrella I, Hernández T, Dueñas M, Troszyńska A, Kosińska A, et al. Antioxidant activity of a red lentil extract and its fractions. International Journal of Molecular Sciences. 2009;**10**:5513-5527. DOI: 10.3390/ijms10125513

[34] Amarowicz R, Estrella I, Hernández T, Robredo S, Troszyńska A, Kosińska A, et al. Free-radical scavenging capacity, antioxidant activity, and phenolic composition of green lentil (*Lens culinaris*). Food Chemistry. 2010;**121**:705-711. DOI: 10.1016/j.foodchem.2010.01.009

[35] Amarowicz R, Karamać M, Dueñas M, Pegg RB. Antioxidant activity and phenolic composition of a red bean (*Phasoelus vulgaris*) extract and its fractions. Natural Product Communications. 2017;**12**:541-544

[36] Amarowicz R, Estrella I, Hernández T, Troszyńska A. Antioxidant activity of extract of adzuki bean and its fractions. Journal of Food Lipids. 2008;**15**:119-136. DOI: 10.1111/j.1745-4522.2007.00106.x

[37] Amarowicz R, Naczk M, Zadernowski R, Shahidi F. Antioxidant activity of condensed tannins of beach pea, canola hulls, evening primrose, and faba bean. Journal of Food Lipids. 2000, 2000;7:195-205. DOI: 10.1111/j.1745-4522.2000.tb00171.x

[38] Baginsky C, Peña-Neira Á, Cáceres A, Hernández T, Estrella I, Morales H, et al. Phenolic compound composition in immature seeds of fava bean (*Vicia faba* L.) varieties cultivated in Chile. Journal of Food Composition and Analysis. 2013;**31**:1-6. DOI: 10.1016/j.jfca.2013.02.003

[39] Amarowicz R, Shahidi F. Antioxidant activity of faba bean extract and fractions thereof. Journal of Food Bioactives. 2018;**2**:112-118. DOI: 10.31665/JFB.2018.2146

[40] Luo Y-W, Wang Q, Li J, Wang Y, Xiao-Xiao J, Zheng-Ping H. The impact of processing on antioxidant activity of faba bean (*Vicia faba* L.). Advance Journal of Food Science and Technology. 2015;7:361-367

[41] Abu-Reidah IM, del Mar CM, Arráez-Román D, Fernández-Gutiéraz A, Segura-Carretero A. UHPLC-ESI-QTOF-MS-based metabolic profiling of *Vicia faba* L. (Fabaceae) seeds as a key strategy for characterization in foodomics. Electrophoresis. 2014;**35**:1571-1581. DOI: 10.1002/elps.201300646

[42] De Pascual-Teresa S, Santos-Buelga C, Rivas-Gonzalo JC. Quantitative analysis of flavan-3-ols in Spanish foodstuffs and beverages. Journal of Agricultural and Food Chemistry. 2000;**48**:5331-5337. DOI: 10.1021/jf000549h

[43] Siddhuraju P, Becker K. The antioxidant and free radical scavenging activities of processed cowpea (*Vigna unuguiculata* (L.) Walp.) seed extracts. Food Chemistry. 2007;**101**:10-19. DOI: 10.1016/j.foodchem.2006.01.004

[44] Amarowicz R, Troszyńska A, Baryłko-Pikielna N, Shahidi F. Polyphenolics extracts from legume seeds: Correlations between total antioxidant activity, total phenolics content, tannins content and astringency. Journal of Food Lipids. 2004;**11**:278-286. DOI: 10.1111/j.1745-4522.2004.01143.x

