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# The Application of Membrane Separation Technology in the Dairy Industry

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## Abstract

Dairy industry is considered as an important food industry that provides various kinds of nutritionally rich dairy products for all age groups. Beside these nutritive values, dairy industry is contemplated as a good source of raw materials for other industries. Most importantly, dairy industry employs environment-friendly and energy-saving technologies. Membrane separation technology being one of those also focused on a cost-effective and environment-friendly manner, which can be widely applied in dairy industry for many useful purposes. In this chapter, we first define and classify the membrane separation technology and then comprehensively describe its applications, for instance, component separation, filtration, removal of bacteria, and wastewater treatment in dairy industry.

**Keywords:** membrane separation, milk concentration and component separation, milk sterilization, dairy wastewater treatment

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## 1. Introduction

Milk is a complete food, and its products are a good source of essential nutrients for human health and raw materials for other industries. Dairy industry is a predominant part of food industry that has rapid growth and stability in emerging markets. With the increase of dairy industries, to control cost and make it sustainable, it is necessary to adopt new energy-efficient and eco-friendly technologies. In new technologies, membrane separation is an emerging technology with suitable properties for dairy products in which solution is passed through a membrane of microscopic pores and pressure applied to separate the components [1].

Membrane separation technique	Principle	Driving force (kPa)	Intercept component	Application
Microfiltration	Sieving	20–100	0.1–20 $\mu\text{m}$	Clarification, separation, removal of bacteria, and filtration [2, 3]
Ultrafiltration	Sieving	100–1000	5–100 nm	Concentration, grading, and purification of macromolecular solution [4]
Nanofiltration	Dissolving diffusion, Donnan effect	500–1500	>1 nm	Separation, purification, and enrichment process of food, medicine, and biochemical industries [5]
Reverse osmosis	Dissolving diffusion	1000–10,000	0.1–1 nm	Concentration of low-molecular-weight components and removal of dissolved salts in aqueous solutions [6]
Electrodialysis	Ion exchange	Electrochemical potential—penetration	Large ions and water	Removal of salt and deacidification of solutions containing neutral components [7]
Pervaporation	Dissolving diffusion	Concentration difference	Insoluble or nonvolatile components	It is mainly used for volatile organic pollutants in the product separation and enrichment [8]

**Table 1.** Classification and characteristics of membrane separation techniques.

Most commonly used membrane separation techniques are microfiltration (MF), ultrafiltration (UF), nanofiltration, reverse osmosis, and electrodialysis. Their characteristics and applications are shown in **Table 1**. As these techniques have a good economic performance and are eco-friendly and uncomplicated to use, they are widely used in dairy industry for removal of bacteria, concentration, component separation, and wastewater treatment.

## 2. Applications of membrane separation technology in the dairy industry

### 2.1. Milk concentration and component separation

The removal of water from milk is known as milk concentration that reduces the cost of the packaging, storage, and transportation of milk and its products. To concentrate the milk, on the principle of heat exchange, flash [9] and falling film evaporation [10] methods are developed. However, these methods may change the composition, rheological characteristics, and heat stability and are energy consuming. As a result, the properties of final products are influenced [11]. Membrane separations are not phase separation technologies. They have advantages

of having lower cost, being environment friendly, and having a simple operation [12]. Kelly [13] and Jevons [14] applied ultrafiltration and reverse osmosis in preconcentration of quarg, soft cheese, and yogurt, respectively. The results showed that output of cheese significantly improved.

Besides milk concentration, milk components such as casein, whey proteins, mineral substance, lactose, and saccharides can also be isolated by membrane separation techniques. Milk proteins are whey protein (average diameter < 20 nm) and casein micelles (average diameter of 200 nm) that can be isolated by using membranes of 0.05 to 0.2  $\mu$ m diameter [15]. Whey protein consists of lactoferrin,  $\beta$ -lactoglobulin,  $\alpha$ -lactalbumin and immunoglobulins that demonstrate a range of immune-enhancing properties [16]. These components can be separated by permeation and phage retention by using one filtration process at the same time that gives approximately 60%  $\alpha$ -lactalbumin and 40%  $\beta$ -lactoglobulin [17]. Al-akoum et al. [18] reported that slightly higher transmission rates, 65% for  $\alpha$ -lactalbumin and 25–30% for  $\beta$ -lactoglobulin, were obtained by using vibratory shear enhanced processing. Rotating ceramic membranes are more suitable because they offered a better compromise between flux and whey protein transmission [19].

Casein is a major protein found in mammalian milk (80% of cow's milk proteins and 20–45% of human milk proteins) [20] that provides amino acids, carbohydrates, calcium, and phosphorus. Membrane separation processes do not affect the micellar structure of casein as compared to traditional methods such as acidification and rennet coagulation [21]. The temperature in membrane filtration is 45 to 50°C that is beneficial for high flux and growth control of mesophilic bacteria [22].  $\beta$ -Casein exists in the serum phase that requires low temperature [23, 24]. Therefore, new separation techniques are developed such as polyethersulfone (PES) and polyvinylidene fluoride (PVDF) membrane that applied in  $\beta$ -casein enrichment at refrigeration temperatures (<20°C). The final casein has same composition, physicochemical properties, and protein profile. Moreover, PES membrane has a higher flux and a lower fouling [25]. Chai et al. [26] applied the transverse vibrating membrane filtration system of 0.04  $\mu$ m PVDF membrane at 10°C to separate and concentrate the milk protein, and the structure of obtained protein was preserved better.

It is notable that membrane fouling is a serious problem and becomes more severe when protein concentrated and viscosity increased during protein separation. Therefore, effective methods for fouling removal are developed. High cross-flow velocities can effectively increase the shear force that controls the membrane fouling and maintains the productivity [27]. Dynamic membrane systems such as vibratory shear enhancing process are also helpful to control the fouling problem. The sugar present in milk is lactose that is a functional ingredient used in food and pharmaceutical products. It is used in bakery goods to reduce sweetness and enhance browning and as a protective carrier for sensitive proteins and peptides. But high amounts of lactose content lead to undesirable grainy texture and cause dyspepsia [28]. The wastewater of dairy industry contains high amount of lactose that increases the level of chemical and biochemical oxygen and causes pollution. To control this pollution, it is necessary to adopt membrane separation techniques to remove lactose before draining the wastewater [29–31].

The combination of microfiltration and ultrafiltration was applied to produce protein-enriched yogurt from fractionated skim milk. Results showed that the lactose content of final product also decreased up to 50% [32]. Morr and Brandon [33] evaluated that when MF in combination with UF membrane was applied to fractionate lactose and sodium from skim milk, 90–95% of lactose and sodium fractionated without affecting the consumer acceptance, product appearance, and flavor. When the lactose is separated from goat's milk by ultrafiltration membrane, some particular components such as serum proteins, casein, and fat globules are retained. The optimization of parameters usually involve transmembrane pressure and cross-flow velocity [34].

Lactose recovery from wastewater with ultrafiltration, nanofiltration, and reverse osmosis was also reported in many previous studies [35–37]. In general, nanofiltration and reverse osmosis are more efficient in terms of lactose recovery, but they require a higher operating pressure as compared to ultrafiltration [38–41]. Chollangi and Hossain [42] found that membranes with molar weight cutoff 3, 5, and 10 kDa provided 70–80%, 90–95%, and 100% recovery rate of lactose in permeate, respectively. In addition, lactose hydrolysis was applied in a continuous stirred tank-ultrafiltration (CSTR-UF) with  $\beta$ -galactosidase enzyme to produce galactose and glucose [43].

Human milk oligosaccharides (HMOs) play an important role in the growth and development of infants [44]. Animal milk also contains oligosaccharides with similar structure and function as human milk oligosaccharides that can be a functional food ingredient [45]. Sialyllactose is N-acetylneuraminic acid (sialic acid) bound to  $\beta$ -lactose, and Luo et al. [46] showed that high permeation of 3'-sialyllactose is obtained by using an integrated UF/NF membrane system for the valorization of dairy by-products with engineered sialidase. Continuous production of sialyllactose, as a typical sialylsaccharide, was also examined with a membrane reactor by Masuda et al. [47].

## 2.2. Removal of bacteria in milk

Milk contains particles with different sizes such as somatic cells (15–6  $\mu\text{m}$ ), fat globules (15–0.2  $\mu\text{m}$ ), bacteria (6–0.2  $\mu\text{m}$ ), and casein micelles (0.3–0.03  $\mu\text{m}$ ) [48]. Microbial and somatic cells of milk affect quality, flavor, and shelf life of final dairy products. Milk is treated with heat to remove microbial cells [49]. However, the heat treatment change the nutritional and flavor profile of the products [50, 51]. Membrane separation techniques are operated at low temperature, which remove bacteria effectively without affecting the nutritional and flavor profile. It also reduces the processing and transportation cost; that's why microfiltration is widely used for the removal of bacteria [52–55]. Cross-flow microfiltration (CFMF) has emerged as an industrial separation technique in the dairy industry [56–58].

Sterilization with inorganic ceramic membrane not only keeps the flavor of milk but also prolongs the shelf life of product. This processing technology is combined with slight heat treatment and applied in cold pasteurization. The products are called as extended shelf-life (ESL) milk [59]. ESL milk has a shelf life of 3 weeks, longer than HTST-pasteurized milk (10 days, typically), and sensory profile analysis shows that ESL milks have no appreciable difference from the pasteurized milk during storage. It fills the gap between high-temperature short-time (HTST) pasteurized milk and ultrahigh temperature (UHT) milk [60].

### 2.3. Wastewater treatment

Dairy industry is the major source of wastewater in the food processing [61] that contains large amount of organic matter and nutrients [62]. Common treatments include primary treatment and secondary biological treatment. Membrane separation usually plays an important role in secondary biological treatment as it is simple and energy saving and has wastewater zero emissions [63, 64]. In this processing, protein and sugar are also recycled from the wastewater. Membrane with different molecular weight cut off plays different roles in wastewater treatment. **Table 2** shows the applications of different membrane separation technologies for wastewater treatment.

### 2.4. Application of electrodialysis and pervaporation in dairy industry

Apart from above mentioned membrane separation technologies, electrodialysis and pervaporation are also used in dairy industry. Electrodialysis is a unit operation applied for the separation or concentration of ions in a solution, based on their selective electromigration through semipermeable membranes under the influence of a potential gradient. Nowadays, this operation has been widely used for demineralization in the dairy industry [66] and has successfully applied electrodialysis for desalination of skimmed milk and showed that the technique is useful in demineralization of dairy products. Demineralization is helpful for better use of milk protein such as application in infant formula. Laurent et al. [67] used electrodialysis for demineralization of skim milk with rate up to 75%. This is much better than their previous study (30–40% demineralization rate) [68]. Chen et al. [69] also successfully applied electrodialysis to remove the lactate ions from acid whey in order to solve operational problems in downstream spray drying operations. Alternatively, electrodialysis has been successfully demonstrated to recover lactic acid from fermentation broths [70, 71], as well as to demineralize sweet whey prior to whey powder production [72, 73]. However, ED applications are still in their infancy, and its potentialities have not been completely exploited probably because of the high specific electromembrane costs or their short lifetime [74].

Pervaporation is a selective membrane separation process in which some feed components are concentrated to a greater degree than others with the selectivity controlled by the membrane type [75]. It can be used to concentrate certain compounds in a mixture. In hydrophobic

Membrane separation technique	MWCO	Application
Microfiltration	100–500 kDa	Remove almost all pathogenic bacterial species and mold as well as a certain amount of halogenated salt [48]
Ultrafiltration	2–150 kDa	Remove almost all of the protein, fat, and some insoluble compounds and minerals in dairy wastewater, and only lactose, soluble salts, and ash content will be allowed to pass [65]
Nanofiltration	0.2–2 kDa	Intercept the lactose in the dairy wastewater, and recover more than 90% of the acid and alkali wastewater from clean in place (CIP)
Reverse osmosis	<0.2 kDa	Intercept almost all pollutants in dairy wastewater.

**Table 2.** Application of different membrane separation technologies in dairy wastewater treatment.



pervaporation, volatile hydrophobic compounds such as flavors pass through the polymeric membrane more readily than water and are thereby concentrated in the permeate [76]. In previous reports, it was used to concentrate acids, esters, and ketones in model flavor mixtures, and the characteristics of the feed mixture (pH and presence of dairy ingredients) were found to alter the pervaporation behavior of the flavor compounds [77].

### 3. Conclusion

The applications of membrane separation technology in dairy industry are wide. These are used in milk concentration, component separation such as protein and lactose, filtration, and bacteria reduction as well as play an important role in dairy industry wastewater treatments. All these applications fully demonstrate the advantages of membrane separation: simple operation, environment-friendly, and energy saving. However, there are still some problems such as membrane fouling that limit its further application. Therefore, more attention should be paid on the mechanism and control methods.

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### References

- [1] Ortega-Rivas E. Membrane separations. In: Non-thermal Food Engineering Operations. Boston: Springer; 2012. pp. 199-214. DOI: 10.1007
- [2] Chhaya SC, Mondal S, Majumdar GC, De S. Clarification of Stevia extract by ultrafiltration: Selection criteria of the membrane and effects of operating conditions. Food and Bioproducts Processing. 2012;**90**(C3):525-532. DOI: 10.1016/j.fbp.2011.10.002

- [3] Pinto MS, Pires ACS, Sant'Ana HMP, Soares NFF, Carvalho AF. Influence of multilayer packaging and microfiltration process on milk shelf life. *Food Packaging and Shelf Life*. 2014;**1**(2):151-159. DOI: 10.1016/j.fpsl.2014.01.006
- [4] Loginov M, Boussetta N, Lebovka N, Vorobiev E. Separation of polyphenols and proteins from flaxseed hull extracts by coagulation and ultrafiltration. *Journal of Membrane Science*. 2013;**442**:177-186. DOI: 10.1016/j.memsci.2013.04.036
- [5] Wang KY, Xiao Y, Chung T-S. Chemically modified polybenzimidazole nanofiltration membrane for the separation of electrolytes and cephalexin. *Chemical Engineering Science*. 2006;**61**(17):5807-5817. DOI: 10.1016/j.ces.2006.04.031
- [6] Skelton R. Membrane filtration applications in the food industry. *Filtration & Separation*. 2000;**37**(3):28-30. DOI: 10.1016/S0015-1882(00)88494-3
- [7] Strathmann H. Electrodialysis, a mature technology with a multitude of new applications. *Desalination*. 2010;**264**(3):268-288. DOI: 10.1016/j.desal.2010.04.069
- [8] Buvet R, Idier L, Inventors; MORGANE, assignee. Process for the concentration by pervaporation of an aqueous fluid containing volatile or carrier water vapour-distillable organic compounds; 1990. EP0380424
- [9] Kiesner C, Eggers R. Concept of a sterile concentration process for milk by multistage flash evaporation. *Chemical Engineering and Technology*. 1994;**17**:374-381. DOI: 10.1002/ceat.270170603
- [10] Liu D. Design considerations of single-effect falling film evaporator in liquid milk. *Journal of Dairy Science & Technology*. 2010;**4**:167-168. DOI: 10.15922/j.cnki.jdst.2010.04.008
- [11] Liu D, Li J, Zhang J, Liu X, Wang M, Hemar Y, et al. Effect of partial acidification on the ultrafiltration and diafiltration of skim milk: Physico-chemical properties of the resulting milk protein concentrates. *Journal of Food Engineering*. 2017;**212**:55-64. DOI: 10.1016/j.jfoodeng.2017.05.019
- [12] Kofschoten RC, Janssen AEM, Boom RM. Mass diffusion-based separation of sugars in a microfluidic contactor with nanofiltration membranes. *Journal of Separation Science*. 2011;**34**(11):1338-1346. DOI: 10.1002/jssc.201100018
- [13] Kelly PM. Application of membranes to whole milk separations: A review. *Irish Journal of Food Science and Technology*. 1987;**11**(2):153-161. DOI: 10.2307/25558165
- [14] Jevons M. Separate and concentrate. *Dairy Industries International*. 1997;**8**(62):19-21, 62
- [15] Heino AT, Uusi-Rauva JO, Rantamaki PR, Tossavainen O. Functional properties of native and cheese whey protein concentrate powders. *International Journal of Dairy Technology*. 2007;**60**(4):277-285. DOI: 10.1111/j.1471-0307.2007.00350.x
- [16] Marshall K. Therapeutic applications of whey protein. *Alternative Medicine Review: A Journal of Clinical Therapeutic*. 2004;**9**(2):136-156
- [17] Samtlebe M, Wagner N, Neve H, Heller KJ, Hinrichs J, Atamer Z. Application of a membrane technology to remove bacteriophages from whey. *International Dairy Journal*. 2015;**48**:38-45. DOI: 10.1016/j.idairyj.2014.12.004



- [18] Al-Akoum O, Ding L, Chotard-Ghodsni R, Jaffrin MY, Gésan-Guiziou G. Casein micelles separation from skimmed milk using a VSEP dynamic filtration module. *Desalination*. 2002;**144**(1):325-330. DOI: 10.1016/S0011-9164(02)00337-5
- [19] Espina VS, Jaffrin MY, Ding LH. Comparison of rotating ceramic membranes and polymeric membranes in fractionation of milk proteins by microfiltration. *Desalination*. 2009;**245**(1):714-722. DOI: doi.org/10.1016/j.desal.2009.02.042
- [20] Kunz C, Lonnerdal B. Human-milk proteins: Analysis of casein and casein subunits by anion-exchange chromatography, gel electrophoresis, and specific staining methods. *The American Journal of Clinical Nutrition*. 1990;**51**(1):37-46. DOI: 10.1093/ajcn/51.1.37
- [21] Lawrence ND, Kentish SE, O'Connor AJ, Barber AR, Stevens GW. Microfiltration of skim milk using polymeric membranes for casein concentrate manufacture. *Separation and Purification Technology*. 2008;**60**(3):237-244. DOI: 10.1016/j.seppur.2007.08.016
- [22] Hurt E, Zulewska J, Newbold M, Barbano DM. Micellar casein concentrate production with a 3X, 3-stage, uniform transmembrane pressure ceramic membrane process at 50 degrees C. *Journal of Dairy Science*. 2010;**93**(12):5588-5600. DOI: 10.3168/jds.2010-3169
- [23] Payens TAJ, Van Markwijk BW. Some features of the association of  $\beta$ -casein. *Biochimica et Biophysica Acta*. 1963;**71**(Supplement C):517-530. DOI: 10.1016/0006-3002(63)91124-7
- [24] Rose D. Relation between micellar and serum casein in bovine milk. *Journal of Dairy Science*. 1968;**51**(12):1897-1902. DOI: 10.3168/jds.S0022-0302(68)87308-4
- [25] Crowley SV, Caldeo V, McCarthy NA, Fenelon MA, Kelly AL, O'Mahony JA. Processing and protein-fractionation characteristics of different polymeric membranes during filtration of skim milk at refrigeration temperatures. *International Dairy Journal*. 2015;**48**:23-30. DOI: 0.1016/j.idairyj.2015.01.005
- [26] Chai M, Ye Y, Chen V. Separation and concentration of milk proteins with a submerged membrane vibrational system. *Journal of Membrane Science*. 2017;**524**:305-314. DOI: 10.1016/j.memsci.2016.11.043
- [27] Punidadas P, Rizvi SSH. Separation of milk proteins into fractions rich in casein or whey proteins by cross flow filtration. *Food Research International*. 1998;**31**(4):265-272. DOI: 10.1016/S0963-9969(98)00088-X
- [28] Somkuti GA, Holsinger VH. Microbial technologies in the production of low-lactose dairy foods. *Food Science and Technology International*. 1997;**3**(3):163-169. DOI: 10.1177/108201329700300302
- [29] Cuartas-Uribe B, Alcaina-Miranda MI, Soriano-Costa E, Mendoza-Roca JA, Iborra-Clar MI, Lora-Garcia J. A study of the separation of lactose from whey ultrafiltration permeate using nanofiltration. *Desalination*. 2009;**241**(1):244-255. DOI: 10.1016/j.desal.2007.11.086
- [30] Marwaha SS, Kennedy JF. Review: Whey-pollution problem and potential utilization. *International Journal of Food Science and Technology*. 1988;**23**(4):323-336. DOI: 10.1111/j.1365-2621.1988.tb00586.x

- [31] Trägårdh G. Membrane applications in the food industry. *Polymer Journal*. 1991;**23**(5):521-529. DOI: 10.1295/polymj.23.521
- [32] Rinaldoni AN, Campderros M, Menendez CJ, Padilla AP. Fractionation of skim milk by an integrated membrane process for yoghurt elaboration and lactose recuperation. *International Journal of Food Engineering*. 2009;**5**(3). DOI: 10.2202/1556-3758.1531
- [33] Morr CV, Brandon SC. Membrane fractionation processes for removing 90 to 95% of the lactose and sodium from skim milk and for preparing lactose and sodium-reduced skim milk. *Journal of Food Science*. 2008;**73**(9):C639-CC47. DOI: 10.1111/j.1750-3841.2008.00961.x
- [34] Abidin NSZ, Hussain SA, Kamal SMM. Removal of lactose from highly Goat's milk concentration through ultrafiltration membrane. In: Ahmed I, editor. *Process and Advanced Materials Engineering, Applied Mechanics and Materials*. Vol. 6252014. pp. 596-599. DOI: 10.4028/www.scientific.net/AMM.625.596
- [35] Barba D, Beolchini F, Veglió F. Minimizing water use in Diafiltration of whey protein concentrates. *Separation Science and Technology*. 2000;**35**(7):951-965. DOI: 10.1081/SS-100100204
- [36] Barba D, Beolchini F, Cifoni D, Veglio F. Whey protein concentrate production in a pilot scale two-stage diafiltration process. *Separation Science and Technology*. 2001;**36**(4):587-603. DOI: 10.1081/SS-100102948
- [37] Pouliot Y, Wijers MC, Gauthier SF, Nadeau L. Fractionation of whey protein hydrolysates using charged UF NF membranes. *Journal of Membrane Science*. 1999;**1;158**(1-2):105-114. DOI: 10.1016/S0376-7388(99)00006-X
- [38] Cheang BL, Zydney AL. A two-stage ultrafiltration process for fractionation of whey protein isolate. *Journal of Membrane Science*. 2004;**231**(1-2):159-167. DOI: 10.1016/j.memsci.2003.11.014
- [39] Atra R, Vatai G, Bekassy-Molnar E, Balint A. Investigation of ultra- and nanofiltration for utilization of whey protein and lactose. *Journal of Food Engineering*. 2005;**67**(3):325-332. DOI: 10.1016/j.jfoodeng.2004.04.035
- [40] Barba D, Beolchini F, Veglio F. Water saving in a two stage diafiltration for the production of whey protein concentrates. *Desalination*. 1998;**119**(1):187-188. DOI: 10.1016/S0011-9164(98)00146-5
- [41] Barba D, Beolchini F, Cifoni D, Veglio F. Whey ultrafiltration in a tubular membrane: Effect of selected operating parameters. *Separation Science and Technology*. 2002;**37**(8):1771-1788. DOI: doi.org/10.1081/SS-120003043
- [42] Chollangi A, Hossain MM. Separation of proteins and lactose from dairy wastewater. *Chemical Engineering and Processing*. 2007;**46**(5):398-404. DOI: 10.1016/j.cep.2006.05.022
- [43] Namvar-Mahboub M, Pakizeh M. Experimental study of lactose hydrolysis and separation in CSTR-UF membrane reactor. *Brazilian Journal of Chemical Engineering*. 2012;**29**(3):613-618. DOI: 10.1590/S0104-66322012000300018

- [44] Hickey RM. The role of oligosaccharides from human milk and other sources in prevention of pathogen adhesion. *International Dairy Journal*. 2012;**22**(2):141-146. DOI: 10.1016/j.idairyj.2011.09.012
- [45] Oliveira DL, Wilbey RA, Grandison AS, Roseiro LB. Milk oligosaccharides: A review. *International Journal of Dairy Technology*. 2015;**68**(3):305-321. DOI: 10.1111/1471-0307.12209
- [46] Luo J, Nordvang RT, Morthensen ST, Zeuner B, Meyer AS, Mikkelsen JD, et al. An integrated membrane system for the biocatalytic production of 3'-sialyllactose from dairy by-products. *Bioresource Technology*. 2014;**166**:9-16. DOI: 10.1016/j.biortech.2014.05.003
- [47] Masuda M, Kawase Y, Kawase M. Continuous production of sialyllactose from colominic acid using a membrane reactor. *Journal of Bioscience and Bioengineering*. 2000;**89**(2):119-125. DOI: 10.1016/S1389-1723(00)88724-1
- [48] Saboya LV, Maubois JL. Current developments of microfiltration technology in the dairy industry. *Le Lait*. 2000;**80**(6):541-553. DOI: 10.1051/lait:2000144
- [49] Hinrichs J, Atamer Z. Heat Treatment of Milk | Sterilization of Milk and Other Products A2 - Fuquay. In: John W, editor. *Encyclopedia of Dairy Sciences*. 2nd ed. San Diego: Academic Press; 2011. pp. 714-724. DOI: 10.1016/B978-0-12-374407-4.00218-1
- [50] Kilshaw PJ, Heppell LM, Ford JE. Effects of heat treatment of cow's milk and whey on the nutritional quality and antigenic properties. *Archives of Disease in Childhood*. 1982;**57**(11):842-847. DOI: 10.1136/adc.57.11.842
- [51] Greenbank GR, Wright PA. The deaeration of raw whole milk before heat treatment as a factor in retarding the development of the tallowy flavor in its dried product. *Journal of Dairy Science*. 1951;**34**(8):815-818. DOI: 10.3168/jds.S0022-0302(51)91786-9
- [52] Solanki G, Rizvi SSH. Physico-chemical properties of skim milk retentates from microfiltration. *Journal of Dairy Science*. 2001;**84**(11):2381-2391. DOI: 10.3168/jds.S0022-0302(01)74687-5
- [53] Pafylas I, Cheryan M, Mehaia MA, Saglam N. Microfiltration of milk with ceramic membranes. *Food Research International*. 1996;**29**(2):141-146. DOI: 10.3168/jds.S0022-0302(01)74687-5
- [54] Fritsch J, Moraru CI. Development and optimization of a carbon dioxide-aided cold microfiltration process for the physical removal of microorganisms and somatic cells from skim milk. *Journal of Dairy Science*. 2008;**91**(10):3744-3760. DOI: 10.3168/jds.2007-0899
- [55] Madaeni SS, Tavakolian HR, Rahimpour F. Cleaning optimization of microfiltration membrane employed for milk sterilization. *Separation Science and Technology*. 2011;**46**(4):571-580. DOI: 10.1080/01496395.2010.534118
- [56] Tomasula PM, Mukhopadhyay S, Datta N, Porto-Fett A, Call JE, Luchansky JB, et al. Pilot-scale crossflow-microfiltration and pasteurization to remove spores of *Bacillus anthracis* (Sterne) from milk. *Journal of Dairy Science*. 2011;**94**(9):4277-4291. DOI: 10.3168/jds.2010-3879

- [57] Fauquant J, Beaucher E, Sinet C, Robert B, Lopez C. Combination of homogenization and cross-flow microfiltration to remove microorganisms from industrial buttermilks with an efficient permeation of proteins and lipids. *Innovative Food Science & Emerging Technologies*. 2014;**21**:131-141. DOI: 10.1016/j.ifset.2013.10.004
- [58] Schmidt VSJ, Kaufmann V, Kulozik U, Scherer S, Wenning M. Microbial biodiversity, quality and shelf life of microfiltered and pasteurized extended shelf life (ESL) milk from Germany, Austria and Switzerland. *International Journal of Food Microbiology*. 2012;**154**(1-2):1-9. DOI: 10.1016/j.ijfoodmicro.2011.12.002
- [59] Hoffmann W, Kiesner C, Clawin-RÄDecker I, Martin D, Einhoff K, Lorenzen PC, et al. Processing of extended shelf life milk using microfiltration. *International Journal of Dairy Technology*. 2006;**59**(4):229-235. DOI: 10.1111/j.1471-0307.2006.00275.x
- [60] Lorenzen PC, Clawin-Raedecker I, Einhoff K, Hammer P, Hartmann R, Hoffmann W, et al. A survey of the quality of extended shelf life (ESL) milk in relation to HTST and UHT milk. *International Journal of Dairy Technology*. 2011;**64**(2):166-178. DOI: 10.1111/j.1471-0307.2010.00656.x
- [61] Rahimi Z, Zinatizadeh AA, Zinadini S. Milk processing wastewater treatment in a bioreactor followed by an antifouling O-carboxymethyl chitosan modified Fe<sub>3</sub>O<sub>4</sub>/PVDF ultrafiltration membrane. *Journal of Industrial and Engineering Chemistry*. 2016;**38**:103-112. DOI: 10.1016/j.jiec.2016.04.011
- [62] Perle M, Kimchie S, Shelef G. Some biochemical aspects of the anaerobic degradation of dairy wastewater. *Water Research*. 1995;**29**(6):1549-1554. DOI: 10.1016/0043-1354(94)00248-6
- [63] Luo J, Ding L, Wan Y, Paullier P, Jaffrin MY. Application of NF-RDM (nanofiltration rotating disk membrane) module under extreme hydraulic conditions for the treatment of dairy wastewater. *Chemical Engineering Journal*. 2010;**163**(3):307-316. DOI: 10.1016/j.cej.2010.08.007
- [64] Andrade LH, Mendes FDS, Espindola JC, Amaral MCS. Nanofiltration as tertiary treatment for the reuse of dairy wastewater treated by membrane bioreactor. *Separation and Purification Technology*. 2014;**126**:21-29. DOI: 10.1016/j.seppur.2014.01.056
- [65] Luo J, Ding L, Qi B, Jaffrin MY, Wan Y. A two-stage ultrafiltration and nanofiltration process for recycling dairy wastewater. *Bioresource Technology*. 2011;**102**(16):7437-7442. DOI: 10.1016/j.biortech.2011.05.012
- [66] Andrés LJ, Riera FA, Alvarez R. Skimmed milk demineralization by electro dialysis: Conventional versus selective membranes. *Journal of Food Engineering*. 1995;**26**(1):57-66. DOI: 10.1016/0260-8774(94)00042-8
- [67] Bazinet L, Ippersiel D, Gendron C, René-Paradis J, Tétrault C, Beaudry J. Bipolar membrane electroacidification of demineralized skim milk. *Journal of Agricultural and Food Chemistry*. 2001;**49**(6):2812-2818. DOI: 10.1021/jf000982r



- [68] Bazinet L, Lamarche F, Ippersiel D, Amiot J. Bipolar membrane electroacidification to produce bovine milk casein isolate. *Journal of Agricultural and Food Chemistry*. 1999;**47**(12):5291-5296. DOI: 10.1021/jf990524m
- [69] Chen GQ, Eschbach FII, Weeks M, Gras SL, Kentish SE. Removal of lactic acid from acid whey using electrodialysis. *Separation & Purification Technology*. 2016;**158**:230-237. DOI: 10.1016/j.seppur.2015.12.016
- [70] Lee EG, Moon SH, Yong KC, Yoo IK, Chang HN. Lactic acid recovery using two-stage electrodialysis and its modelling. *Journal of Membrane Science*. 1998;**145**:53-66. DOI: 10.1016/S0376-7388(98)00065-9
- [71] Wee YJ, Yun JS, Lee YY, Zeng AP, Ryu HW. Recovery of lactic acid by repeated batch electrodialysis and lactic acid production using electrodialysis wastewater. *Journal of Bioscience and Bioengineering*. 2005;**99**:104-108. DOI: 10.1263/jbb.99.104
- [72] Pérez A, Andrés LJ, Álvarez R, Coca JCGH Jr. Electrodialysis of whey permeates and retentates obtained by ultrafiltration. *Journal of Food Process Engineering*. 1994;**17**:177-190. DOI: 10.1111/j.1745-4530.1994.tb00334.x
- [73] Greiter M, Novalin S, Wendland M, Kulbe KD, Fischer J. Desalination of whey by electrodialysis and ion exchange resins: analysis of both processes with regard to sustainability by calculating their cumulative energy demand. *Journal of Membrane Science*. 2002;**210**:91-102. DOI: 10.1016/S0376-7388(02)00378-2
- [74] Fidaleo M, Moresi M. Electrodialysis Applications in the Food Industry. *Advances in Food & Nutrition Research*. 2006;**51**(51):265-360. DOI: 10.1016/S1043-4526(06)51005-8
- [75] Karlsson HOE, Trägårdh G. Pervaporation of dilute organic-waters mixtures. A literature review on modelling studies and applications to aroma compound recovery. *Journal of Membrane Science*. 1993;**76**(2-3):121-146. DOI: 10.1016/0376-7388(93)85211-E
- [76] Lipnizki F, Hausmanns S, Ten PK, Field RW, Laufenberg G. Organophilic pervaporation: Prospects and performance. *Chemical Engineering Journal*. **73**:113-129. DOI: 10.1016/S1385-8947(99)00024-8
- [77] Overington AR, Wong M, Harrison JA. Effect of feed pH and non-volatile dairy components on flavour concentration by pervaporation. *Journal of Food Engineering*. 2011;**107**(1):60-70. DOI: 10.1016/j.jfoodeng.2011.05.045