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Studies on the Character of Hypothalamic GnRH Neurons and Kisspeptin Neurons Using Hypothalamic Cell Models

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Abstract

The hypothalamic-pituitary-gonadal (HPG) axis controls the hormonal network responsible for reproductive functions. In the past, hypothalamic gonadotropin-releasing hormone (GnRH) neurons have been positioned at the highest level in the HPG axis. After the discovery of the indispensable roles of hypothalamic kisspeptin in GnRH neurons, our understanding of the neuroendocrine regulation of the HPG axis was reconfirmed, and it is now recognized that hypothalamic kisspeptin neurons are positioned at the summit of the HPG axis. Accumulating evidence shows that kisspeptin neurons are responsible for the onset of puberty and sex steroid feedback mechanisms by modulating the activity of GnRH neurons. Furthermore, the identification of kisspeptin in the hypophyseal portal circulation suggests that this peptide has some direct roles in the pituitary gland. The detailed mechanisms underlying the regulation of GnRH by kisspeptin and the regulatory control of kisspeptin neurons are still largely unknown because of the limitations of the experimental models. The establishment of GnRH-expressing and kisspeptin-expressing cell models has enabled us to examine the character of these neuronal cells. In this chapter, we describe our in vivo studies examining the character of GnRH neurons and kisspeptin neurons in the hypothalamus using hypothalamic GnRH- and/or kisspeptin-expressing cell models.

Keywords: kisspeptin, GnRH, gonadotropin, HPG axis

1. Introduction

The hypothalamus maintains the homeostasis within the body and controls endocrine functions. The hypothalamic-pituitary-gonadal (HPG) axis is a hormonal network responsible



for female reproductive function. After the discovery of inactivating mutations in the gene encoding the kisspeptin receptor (Kiss1R) in patients with idiopathic hypogonadotropic hypogonadism [1, 2], a new concept of the HPG axis was established. Now, it is generally agreed that kisspeptin produced from kisspeptin neurons, which are located in different regions of hypothalamus, stimulates gonadotropin-releasing hormone (GnRH) synthesis and release through Kiss1R within the GnRH neurons.

2. Kisspeptin as a regulator of the HPG axis

Kisspeptin is positioned upstream of GnRH in the HPG axis. Kisspeptin, which is encoded by the Kiss1 gene, was first discovered as a peptide that has potency to suppress metastasis of malignant melanoma and was initially named metastin [3]. The Kiss1 gene product is translated into a 145-amino acid precursor protein and further cleaved into a 54-residue peptide (kisspeptin 54), which can be further cleaved into 14-, 13-, and 10-amino acid peptides [4]. Kiss1R was discovered 4 years after kisspeptin, and it was found that Kiss1R is a member of the G protein-coupled receptor superfamily and is structurally similar to the galanin receptor [4, 5]. Discovery of loss-of-function mutations in Kiss1R in the family of a hypogonadotropic hypogonadism patient [1, 2] clearly linked kisspeptin and reproduction, and these observations indicated that kisspeptin acts as a major stimulator of the HPG axis.

Two different populations of kisspeptin-expressing neurons have been identified in rodents. The most predominant population is located in the arcuate nucleus (ARC) of the hypothalamus, where kisspeptin-expressing neurons co-express dynorphin and neurokinin B (NKB). Another kisspeptin-expressing cell population is located in the anteroventral periventricular nucleus (AVPV) of the hypothalamus. In humans and primates, kisspeptin is expressed predominantly within the infundibular nucleus, which is equivalent to the ARC in rodent [6]. Kisspeptin neurons in both populations make direct synaptic contacts with GnRH neurons and their terminals in the median eminence [7].

3. GnRH release is influenced by kisspeptin

In rodents, GnRH is released into the portal circulation by GnRH neurons located in the preoptic area and eventually reaches the anterior pituitary. GnRH is released in a pulsatile manner, and the pulse frequency and amplitude of GnRH release vary physiologically during reproductive cycles in females. Secretion of luteinizing hormone (LH) and follicle-stimulating hormone (FSH) is maintained by pulsatile, not continuous, release of GnRH [8]. Moreover, the frequency of the GnRH pulse differentially regulates the production and release of FSH and LH from the anterior pituitary [9]. Administration of high-frequency GnRH pulses increases the secretion of LH, whereas a lower frequency of GnRH pulses shifts the gonadotropin secretion to more FSH dominant [10]. Taken together, these

observations show that the secretory mode of pituitary gonadotropins is controlled by the so-called GnRH pulse generator.

The neuronal mechanisms underlying the pulsatile release of GnRH are still not fully understood, but at present it is agreed that kisspeptin neurons located in the ARC of the hypothalamus may be involved. Previous studies recorded the electrical activity of neurons, measured as multiunit activity (MUA), within the ARC region of the hypothalamus and reported that MUA correlates with pulsatile secretion of LH in several animal models [11, 12]. The neuronal population of kisspeptin-expressing cells is called KNDy neurons because kisspeptin-expressing cells located in the ARC region of the hypothalamus co-express kisspeptin, neurokinin B, and dynorphin. KNDy neurons generate synchronized oscillatory patterns of neuronal activity by sharing excitatory and inhibitory input from NKB and dynorphin produced within themselves [13, 14]. Selective and synchronized activation of KNDy neurons induces pulsatile release of LH in a mouse model [15]. Furthermore, exogenous kisspeptin administration can increase LH pulse frequency and amplitude in healthy women [16]. Because kisspeptin antagonism suppresses both mean GnRH and GnRH pulses [17], it is natural to think that kisspeptin neurons in the ARC of the hypothalamus (KNDy neurons) comprise the GnRH pulse generator.

The pattern of GnRH release and subsequent LH release across the reproductive cycle is modulated by gonadal steroid feedback. In rodents, the estrogen-induced GnRH/LH surge is mediated by kisspeptin neurons in the AVPV region (positive feedback). However, KNDy neurons in the ARC region of the hypothalamus are sensitive to estradiol (E2) and reduce the GnRH/LH secretion (negative feedback). This concept is based on the observations that kisspeptin expression was increased in the AVPV region at the time of the estrogen- and progesterone-induced LH surge in ovariectomized rats, whereas kisspeptin expression levels in the ARC were lowest during this time [18]. Another experiment showing that Kiss1 gene expression in the kisspeptin neurons in the AVPV is upregulated by E2, whereas those in the ARC are inhibited by E2 [19], also supports the current understanding that kisspeptin neurons in the AVPV play a role in the GnRH/LH surge, and kisspeptin neurons in the ARC region maintain the pulsatile release of GnRH and also play a pivotal role in the negative feedback control by E2.

4. Hypothalamic cell models, GT1-7 and rHypoE8, for investigating neuroendocrine mechanisms of the HPG axis

The hypothalamus is the control center for the HPG axis; however, it has been difficult to study in detail the GnRH neurons as well as the kisspeptin neurons because of the inherent heterogeneity of this brain region. The hypothalamus is composed of a complex network of neurons, and there are different neuronal phenotypes that express a specific complement of neuropeptides, neurotransmitters, and receptors [20]. Immortalized, clonal cell lines represent a relatively homogeneous population of specific neuronal cells and can be used as

experimental models. For the study of the character or the functions of hypothalamic GnRH neurons, we are using GT1-7 cell lines, which have proven to be a valuable GnRH-expressing cell model for GnRH neurons. These cells were created from the hypothalamic tumor cells in a transgenic female mouse that expressed the SV40 T-antigen under the control of the GnRH promoter [21]. GT1-7 cells display neuronal morphology and secrete GnRH; therefore, these cells have become one of the most highly utilized neuronal cell models for studies related to GnRH neurons.

The embryonic rat hypothalamic cell line R8 (rHypoE8) consists of hypothalamic neurons from rat embryonic day 18 hypothalamic primary cultures immortalized by retroviral transfer of SV40 T-antigen. These cells express neuroendocrine markers such as kisspeptin, GnRH, and RF-amide-related peptide-3 (RFRP-3, the mammalian ortholog of the avian gonadotropin-inhibiting hormone, GnIH). Because the expression of kisspeptin or RFRP-3 is functionally altered by physiological neuropeptides, these cells serve as tools for the analysis of the cellular and molecular mechanisms involved in the hypothalamic control of a number of physiological processes [22].

5. Effect of kisspeptin on hypothalamic GT1-7 cells

It is generally agreed that hypothalamic kisspeptin regulates GnRH release from GnRH neurons by kisspeptin binding the Kiss1R that is expressed by GnRH neurons [23]. A previous study by Novaira et al. demonstrated a functional role for kisspeptin in GT1-7 cells, in which they showed that kisspeptin stimulates the expression and secretion of GnRH [24]. Similarly, Terasaka et al. demonstrated the stimulatory effect of kisspeptin on GnRH gene expression, and they also found that this stimulatory effect was antagonized in the presence of bone morphogenetic protein in these cells [25]. In our study using GT1-7 cells, we did not observe any effect of kisspeptin on GnRH expression [26]. Because GT1-7 cells express Kiss1R, we suspected that Kiss1R function was lost or diminished in our GT1-7 cells, probably because of a change in cell character due to cell immortalization or multiple passages. On the other hand, when GT1-7 cells overexpressed Kiss1R after transfection with a Kiss1R expression vector, exogenous Kiss1R was absolutely functional. Furthermore, both extracellular signal-regulated kinase (ERK) and cAMP/protein kinase A (PKA) pathways were activated by kisspeptin in Kiss1R-overexpressing GT1-7 cells. These observations suggested that overexpression of exogenous Kiss1R could lead to activation of the intracellular signaling pathways mediated by kisspeptin stimulation in these cells. It is also noteworthy that, even when GT1-7 cells overexpressed Kiss1R, GnRH expression was not stimulated by kisspeptin [26]. It is still unclear why kisspeptin did not increase GnRH expression in our GT1-7 cells even when Kiss1R was overexpressed; instead, we clearly observed that kisspeptin could stimulate the expression of the GnRH receptor (GnRHR) in GT1-7 cells overexpressing Kiss1R [26] (Figure 1). GnRH-producing cells have been reported to respond to GnRH and modify their GnRH expression levels [27]. Furthermore, GnRHRs within GnRH neurons were reported to be involved in the pulsatile secretion of GnRH by an autocrine or paracrine interaction between GnRH and GnRHR [28, 29]. These

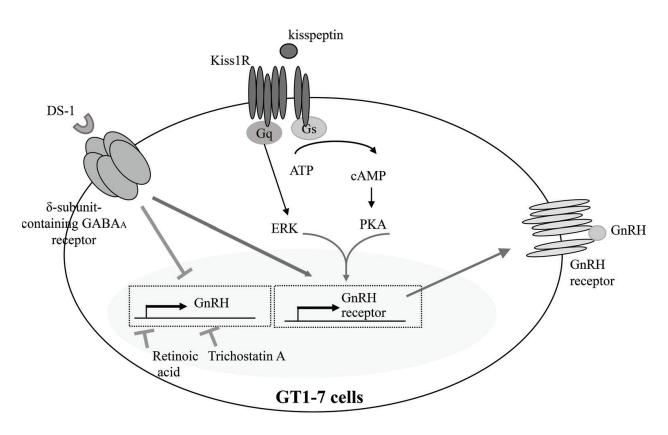


Figure 1. Schematic summary of the regulation of GnRH in GT1-7 cells. GT1-7 cells express Kiss1R, but endogenous Kiss1R does not respond to kisspeptin. Therefore, we used GT1-7 cells overexpressing Kiss1R. GT1-7 cells overexpressing Kiss1R did not show an increase in GnRH mRNA expression upon kisspeptin treatment. However, kisspeptin increased GnRH receptor expression in these cells. We also found that a γ -subunit-containing GABA_A receptor agonist, DS1, as well as histone deacetylase inhibitor trichostatin A, reduces GnRH mRNA expression. Retinoic acid also has an inhibitory effect on GnRH expression.

observations implied that kisspeptin could affect the function of GnRH neurons by changing their expression levels of GnRHR.

6. Effect of kisspeptin on primary cultures of fetal rat brain

Although GT1-7 cells endogenously express Kiss1R, kisspeptin does not affect these cells. When GT1-7 cells overexpress Kiss1R, kisspeptin stimulates intracellular signaling pathways and increases GnRHR, but not GnRH expression. To determine the character of kisspeptin neurons in their original, non-transformed state, we used primary cultures of fetal rat brain that contain both GnRH and kisspeptin neurons. GnRH neurons in these cells did not respond to E2, which failed to stimulate GnRH mRNA expression. This observation was consistent with a previous study that revealed a lack of estrogen receptor immunoreactivity in GnRH neurons, raising doubts about the role of E2 in GnRH neuronal function [30]. In contrast, kisspeptin neurons in these primary cultures responded to E2, and Kiss1 mRNA expression was upregulated by E2 [31], suggesting that kisspeptin neurons, but not GnRH neurons, could be a target of E2 in neuronal cells in the fetal brain. GnRH mRNA expression within these primary cultures of fetal rat brain containing GnRH-producing neurons was

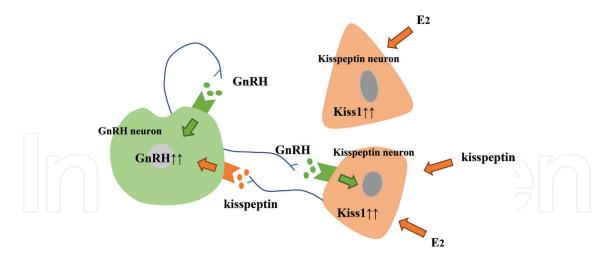


Figure 2. Schematic summary of the regulation of Kiss1 mRNA and GnRH mRNA expression in primary cultures of fetal rat brain and the proposed interaction between kisspeptin neurons and GnRH neurons. In experiments using primary cultures of fetal rat neuronal cells, Kiss1 mRNA, but not GnRH mRNA expression, was upregulated by estradiol (E2). GnRH mRNA expression was clearly increased by treatment with kisspeptin. GnRH stimulation increased the expression of both Kiss1 and GnRH mRNAs, and kisspeptin itself was found to increase the expression of the Kiss1 gene. We postulate that GnRH neurons reversibly interact with kisspeptin neurons and also form an autocrine interaction with kisspeptin neurons.

clearly increased by treatment with kisspeptin. Therefore, we could conclude that kisspeptin can stimulate GnRH synthesis in GnRH-expressing neurons in vivo. However, kisspeptin increased GnRH mRNA expression only up to about 1.5-fold [31]. In addition, we have found that GnRH stimulation increased the expression of the Kiss1 gene as well as that of the GnRH gene and also found that kisspeptin itself increased the expression of the Kiss1 gene. We postulate that GnRH neurons reversibly interact with kisspeptin neurons and also form an autocrine interaction with kisspeptin neurons (**Figure 2**).

7. Trichostatin A, a selective inhibitor of mammalian histone deacetylase, reduces GnRH expression in GT1-7 cells

Observations from the studies using GT1-7 cells and primary cultures of fetal rat brain imply that kisspeptin could affect GnRH neurons and increase GnRH expression. In addition, kisspeptin may change the GT1-7 cells' expression levels of Kiss1R. GnRH synthesis is not only regulated by kisspeptin, but several experimental reagents can modify the GnRH synthesis in GnRH-producing cells. Trichostatin A (TSA), a selective inhibitor of histone deacetylase, is an experimental reagent that modifies gene expression by opening chromatin structure through hyperacetylation of histones [32]. The structural change in chromatin allows transcription factors to bind DNA to modify gene expression. In GnRH-producing GT1-7 cells, TSA significantly reduced GnRH expression, with a concomitant increase in the gene encoding retinal-dehyde dehydrogenase, which catalyzes the oxidation of retinol to retinoic acid [33]. Because retinoic acid also reduces GnRH expression in these cells, epigenetic mechanisms modified through retinaldehyde dehydrogenase, and retinoic acid might have an inhibitory effect on GnRH production (Figure 1).

8. DS1, a δ -subunit-containing GABA_A receptor agonist, reduces GnRH mRNA expression and increases that of GnRHR in GT1-7 cells

It is well documented that the neurotransmitter γ-aminobutyric acid (GABA) can modulate the activity of GnRH neurons. GnRH neurons possess functional GABA, receptors [34], and GABAergic neurons establish synapses with GnRH neurons [35]. GABA neurons predominantly exert their inhibitory effect on GnRH neurons in rodents and sheep. GT1-7 cells also express functional GABA, receptors [36]. GABA, receptors are multimeric proteins that are composed of five subunits drawn from a repertoire of several homologous protein groups (α 1-6, β 1-3, γ 1-3, δ , ϵ , θ , and π); the majority of GABA receptors in the central nervous system are composed of α , β , and γ subunits, and less abundant populations of GABA receptor contain the δ subunit [37]. DS1, an $\alpha 4\beta 3\delta$ GABA, receptor agonist, reduces GnRH mRNA expression in GT1-7 cells, although DS1 can exert a stimulatory effect on signal transduction systems, such as ERK and cAMP/PKA [38]. The δ -subunit-containing $\alpha 4\beta 3\delta$ GABA receptor was found in extra-synaptic sites and is known to control neuronal excitability [39]. Interestingly, although GnRH mRNA expression was decreased, GnRHR expression within GT1-7 cells was significantly increased by DS1 stimulation [38] (Figure 1). At present, it is still unknown why δ-subunit-containing GABA_A receptor agonism decreases the production of GnRH in spite of increasing GnRHR expression. We currently speculate that GABA could modulate GnRH-producing neurons through δ-containing GABA, receptors and deplete their GnRH content by modulating gene expression and secretory function in association with the expression of their GnRHR within the cell (**Figure 1**).

9. Kisspeptin expression is induced by glucagon-like peptide-1 in rHypoE8 cells and GT1-7 cells

As described above, we used GT1-7 cells as a model for GnRH-producing neurons; however, GT1-7 cells also express the Kiss1 gene, which encodes kisspeptin [40]. rHypoE8 cells, another hypothalamic model that was developed from rat embryonic hypothalamic primary cultures, express the Kiss1 gene, and they also express the GnRH gene [22]. Because both rHypoE8 and GT1-7 are immortalized cell lines derived from heterogeneous hypothalamic cell populations, they express several types of neuropeptides. Using these hypothalamic cell models, we found that Kiss1 mRNA was regulated by several metabolic factors. Glucagon-like peptide-1 (GLP-1) is a gastrointestinal hormone produced by the small intestine and colon in response to food intake [41]. GLP-1 is also expressed in the central nervous system, and its expression in the brain is altered during fasting or feeding [42], suggesting that GLP-1 plays a role as a satiety factor. We found that GLP-1 increased the expression of Kiss1 mRNA in rHypoE8 cells as well as GT1-7 cells [43]. Moreover, leptin, which is an anorexigenic factor that is released from adipocytes, can also stimulate Kiss1 mRNA expression in these cells (Figure 3). These observations suggest that the levels of metabolic factors such as GLP-1 or leptin, which change during a state of starvation or negative energy balance, can critically influence the HPG axis by changing kisspeptin expression.

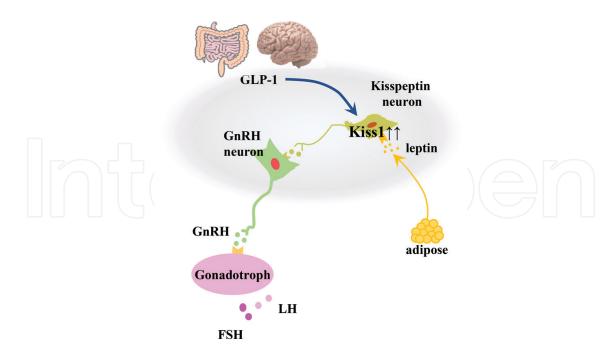


Figure 3. The metabolic factors glucagon-like peptide-1 (GLP-1) and leptin can affect the HPG axis via kisspeptin neurons. From experiments using the hypothalamic cell models rHypoE8 and GT1-7 cells, we found that metabolic factors such as GLP-1 and leptin had the ability to stimulate Kiss1 gene expression. These metabolic factors may affect the HPG axis by modulating the synthesis and release of kisspeptin.

10. Conclusion

Within the past decade, our understanding of the hypothalamic control of female reproductive function has matured considerably. The identification of hypothalamic kisspeptin, a regulator of GnRH, has provided us decisive insight into previously unanswerable questions. Kisspeptin neurons within the hypothalamus play a pivotal role in the control of the HPG axis, but it is still not entirely clear how kisspeptin release and expression are regulated in the brain during the reproductive cycle. Furthermore, the precise biology of kisspeptin and GnRH neurons remains unknown because of the difficulty of isolation of these neurons from heterogeneous neuronal populations of the hypothalamus.

In this review, we described our observations concerning the regulation of kisspeptin and GnRH neurons using hypothalamic cell models. Because we believe these cell models may reflect the original character of genuine kisspeptin and GnRH neurons, future studies using these cells are likely to contribute to our understanding of the mechanisms of regulation of the HPG axis.

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Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the review reported.

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