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Physiology and Pathology of Autoimmune Diseases: Role of CD4+ T cells in Rheumatoid Arthritis

Patricia Castro-Sánchez and Pedro Roda-Navarro

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Abstract

Rheumatoid arthritis (RA) is a chronic autoimmune disease characterised by synovial inflammation leading to bone erosion and to systemic manifestations in patients with long RA duration. Although the aetiology is unknown, several observations make currently clear that CD4 T cells play a key role in the pathogenesis: (1) RA associates with certain polymorphisms of HLA class II molecules, and (2) the repertoire and aging of CD4 T cells as well as the intracellular signalling mediating CD4 T cell activation are altered in RA patients. We describe herein the alterations found in CD4 T cells and the role of these cells in the development and progression of RA.

Keywords: autoimmunity, lymphocytes, synovitis, T cell signalling, T cell aging

1. Introduction

Rheumatoid arthritis (RA) is a chronic autoimmune disease, which affects 0.33 to 2.65% of the population, showing differences between countries and studies [1–7]. It is more frequent in North America than Northern Europe, with Southern Europe having the lowest rate of incidence [8]. As other autoimmune diseases, RA is more prevalent in women than in men, suggesting that hormonal [9] and gender-related genetic factors [10] contribute to the development of the disease. RA is also more frequent in the elderly, consistent with a key role of immune system aging in this disease [11, 12].

RA physiopathology is characterised by persistent synovial inflammation that leads to joint deformity, stiffness and bone erosion. Consequently, patients suffer pain and progressive disability. Although the most evident feature of RA is synovitis, extra-articular manifestations of



RA (ExRA) such as cardiovascular disease can be present in long-duration disease, raising the risk of early death [13, 14].

RA is associated to certain alleles of the major histocompatibility complex class II (MHC-II), and CD4 T cells of RA patients show abnormalities in intracellular signalling, repertoire and aging. It is then conceivable that CD4 T cells could be essential mediators in the development of the chronic inflammation occurring in RA. These cells are key regulators of the immune response secreting pro-inflammatory cytokines and cooperating with B cells for secreting antibodies. In fact, certain RA patients develop autoantibodies such as anti-citrullinated protein antibodies (ACPA) or rheumatoid factor (RF, which recognises the Fc portion of IgG), while other patients do not, indicating that RA comprises at least two different pathologies, seropositive and seronegative [15].

The study of CD4 T cell population has changed our understanding of RA: from the traditional paradigm, which considered that a small set of joint antigens causes the selective expansion of few antigen-specific cells, to a new model in which RA would be a systemic disease caused by alterations in T cell homeostasis and aging. In this chapter, we will describe the role of CD4 T cells in the development of RA and the abnormalities that these lymphocytes show in diseased individuals.

2. Aetiology of rheumatoid arthritis

Although the aetiology of RA remains elusive, genetic and environmental risk factors have been described [16, 17]. MHC-II genes, particularly HLA (human leukocyte antigen) -DRB1 alleles (the so-called shared epitope [18, 19]), constitute the strongest genetic risk factor, accounting for 50% of the genetic contribution to RA [20]. Association with HLA-DRB1 has been established in different populations across the world [21–25], especially in ACPA-positive pathology, and different haplotypes of HLA-DRB1 associate with distinct RA severity and treatment response [26]. Single-nucleotide polymorphisms (SNPs) in other genes have also been linked to RA [16], including genes coding for molecules that regulate T cell activation, which will be discussed below. These genetic associations strongly indicate a decisive role of helper T lymphocytes in the pathology.

The major environmental risk factor is smoking habit, which seems to alter citrullination of mucosal proteins [27]. Genetic and environmental risk factors work together in promoting the disease. For example, smoking habit alters methylation of the HLA-DRB1 region, increasing the chance of developing ACPA-positive RA [28, 29].

Some infectious agents might also be risk factors of RA. For example, there is a positive association between the prevalence of periodontitis and RA [30]. *Porphyromonas gingivalis*, the major causative agent of periodontitis, produces an enzyme that induces aberrant citrullination of host proteins [31]. This generates neoantigens that can then be recognised by the immune system of the host, triggering ACPA production. In addition, it has been shown that ACPA from RA patients cross-react with various autoantigens and microbial and plant-citrullinated proteins [32]. This suggests that environmental factors such as infections and diet may trigger

the production of ACPA in individuals with genetic predisposition. ACPA can then cross-react with self-proteins through molecular mimicry, inducing RA.

3. Pathophysiology of rheumatoid arthritis

A healthy joint (**Figure 1A**, left side) is composed of two adjacent bony ends covered with a layer of cartilage. The space between ends is called articular cavity, which is delimited by the synovial membrane on both sides and contains synovial fluid. The synovial membrane is a thin layer of cells, formed by two types of synoviocytes: type A or macrophage-like synovial cells

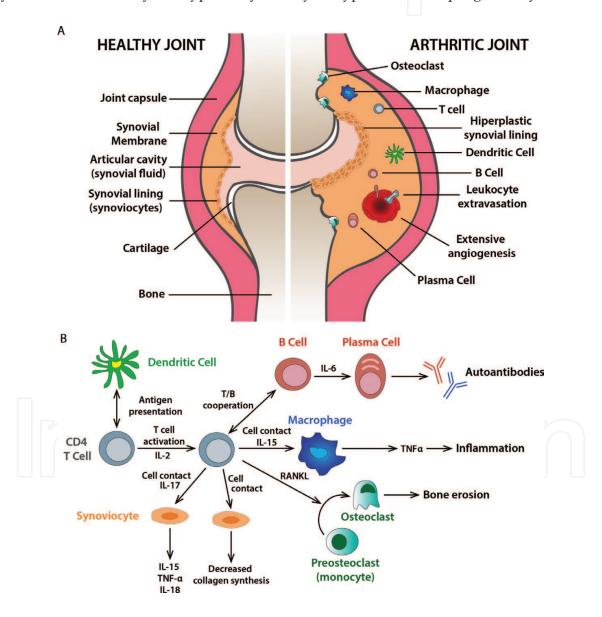


Figure 1. Role of CD4 T cells in rheumatoid synovitis. (**A**) In a healthy synovial joint (left), a thin layer of synoviocytes delimits the joint capsule. By contrast, in RA (right), synoviocytes form an invasive synovial lining and leukocytes infiltrate the synovial membrane. (**B**) Activated CD4 T cells play a central role in inflammatory responses in the synovial membrane, including autoantibody production by plasma cells, secretion of inflammatory cytokines by macrophages and synoviocytes, bone erosion by osteoclasts and inhibition of collagen secretion by synoviocytes.

and type B or fibroblast-like synoviocytes (FLSs). The synovial membrane produces synovial fluid and due to its porous organisation allows diffusion of the nutrients in serum to the avascular cartilage.

The confluence of genetic susceptibility and environmental factors determines the development of an autoimmune response that precedes clinical arthritis. For reasons poorly understood, this autoimmune response exacerbates in the synovium, where leukocytes infiltrate causing synovial membrane inflammation (rheumatoid synovitis) (**Figure 1A**, right side). Synovial infiltrate includes both innate and adaptive immune cells [33, 34] and creates a microenvironment where FLSs acquire an invasive and inflammatory phenotype, leading to hyperplasia of the synovial lining [35, 36]. FLSs secrete matrix metalloproteinases (MMPs) and collagenase, promoting cartilage destruction [37]. Leukocyte infiltration and secretion of pro-inflammatory cytokines favour maturation of pre-osteoclasts to osteoclasts, which leads to bone erosion [38–40]. Cytokines and growth factors released by infiltrated cells, together with the hypoxia resulting from synovial hyperplasia, trigger angiogenesis [41–43], establishing a feedback loop that favours continuous leukocyte infiltration and chronic inflammation.

Inflammation initiated in the synovium gives way to systemic inflammation that alters the function of distant tissues and organs, such as vascular endothelium, adipose tissue, liver and lungs. As a result, ExRA is present in RA patients, such as cardiovascular disease (CVD), anaemia or rheumatoid lung, among others [44].

Although different immune cells infiltrate the inflamed joint, we will focus on CD4 T cells, which, as mentioned above, seem to be central in the pathophysiology of RA by secreting cytokines and by cooperating with synovial cells.

4. Pathogenic role of CD4 T cells in rheumatoid arthritis

4.1. CD4 T cell activation and function in synovitis

CD4 T cells are the most abundant lymphocyte in the synovial infiltrate [45], where they regulate other cell types in the synovium and play a central role in the pathological immune response leading to the joint damage (**Figure 1B**).

4.1.1. CD4 T cell activation by DCs

Dendritic cells (DCs) are key initiators of adaptive immune responses, since they are professional antigen-presenting cells (APCs), able to present to T cell antigenic peptides in the context of the MHC-II. Initially, infiltrated CD4 T cells interact with synovial DCs, resulting in T cell stimulation (**Figure 1B**). Activation of CD4 T cells requires the engagement of the T cell receptor (TCR) by antigen-MHC-II complexes on the surface of the APC. In addition, full T cell activation requires interaction between the molecule CD28 on the T cell and its ligands CD80 and CD86 expressed by APCs, which provides costimulatory signals. Activated CD4 T cells upregulate the expression of the inhibitory molecule cytotoxic T lymphocyte antigen-4 (CTLA-4), which binds CD80 and CD86 with higher affinity

than CD28 [46]. During consecutive contacts with APCs, CTLA-4 will compete with CD28 for CD80/CD86, and binding of CTLA-4 to these ligands will result in inhibition of T cell activation [47]. The importance of APC-mediated T cell costimulation for the progression of RA has been proved by therapy with the CTLA-4-immunoglobulin fusion protein abatacept. This molecule binds to CD80/CD86 on the APC, impeding binding of CD28 and, therefore, blocking T cell costimulation [48]. Treatment with abatacept reduces disease activity and radiographic progression of RA [49, 50].

4.1.2. Cooperation between CD4 T cells and B cells

B cells play a fundamental role in seropositive RA, in which patients develop autoantibodies contributing to inflammation and tissue damage. Autoantibodies are synthesised by plasma cells, which differentiate from B cells after cooperation with CD4 T cells. Upon activation, T cells upregulate the surface expression of CD40 ligand (CD40L or CD154), which interacts with CD40 expressed by B cells. During T/B cooperation, stimulation through CD40 together with IL-6 signalling favours isotype switching, differentiation of B cells into plasma cells and synthesis of antibodies such as ACPA (Figure 1B) [51]. CD4 T cells, B cells and DCs found in joints of RA patients range from diffuse infiltrates to follicular structures, forming ectopic germinal centres (EGCs) in some patients [52]. Formation of EGCs favours the formation of high affinity autoantibodies, increasing the severity of the disease [53]. EGCs and B cells seem to be critical for T cell activation in the synovium [54].

4.1.3. Regulation of FLSs by CD4 T cells

As mentioned before, FLSs are an important component of joint architecture. In a healthy joint (Figure 1A, left side), FLSs form the synovial lining and produce synovial fluid. FLSs acquire an invasive phenotype in RA, causing hyperplasia of the synovial lining (Figure 1A, right side). This hyperplasia originates a hypoxic environment where angiogenesis is activated, favouring perpetuation of inflammation. In addition, RA FLSs secrete high amounts of proteases, which trigger cartilage destruction, and pro-inflammatory cytokines.

Antigen-experienced CD4 T cells affect the function of FLSs by direct cell-cell interaction. For example, CD4 T cells induce the production of the pro-inflammatory cytokines IL-15, TNF- α and IL-18 by FLSs (Figure 1B). This is dependent on CD40L-CD40 engagement as demonstrated by a blocking agent [55]. Collagen synthesis by FLSs is also decreased by CD4 T cells, a process mediated, at least in part, by T cell membrane-associated IFN- γ , TNF- α and IL- 1α [56].

4.1.4. Regulation of macrophages/monocytes by CD4 T cells

Macrophages infiltrate the RA joint, where they interact with synovial cells and produce the pro-inflammatory cytokine TNF- α . CD4 T cells regulate macrophages in the synovium, as shown by the finding that freshly isolated synovial T cells can induce the expression of the pro-inflammatory cytokine TNF- α by macrophages in an IL-15-dependent manner (Figure 1B) [57]. Resembling the behaviour of T cells in RA patients, T cells of healthy donors stimulated with an inflammatory cytokine cocktail can induce the production of TNF- α by resting monocytes [58]. It should be noted that TNF- α production by myeloid cells is also induced by IL-15-stimulated NK cells [59]. Due to the central role of TNF- α in the progression of RA, as demonstrated by the succeeded neutralising therapy [60], it will be needed to further investigate this complex regulation of immune cells in the inflamed joint.

Monocytes are the progenitors of osteoclasts, which constitute the only cell type that is able to degrade bone. In health, bone resorption by osteoclasts and bone generation by osteoblasts are tightly regulated to maintain skeletal integrity and homeostasis. In RA, osteoclast activity in the joint is increased, resulting in an unbalanced bone erosion. Synovial CD4 T cells from RA patients, as well as activated peripheral blood T cells from healthy donors, express receptor activator of nuclear factor κB ligand (RANKL), which engages RANK expressed on monocytes, inducing their differentiation to osteoclasts [61, 62] and, consequently, triggering bone erosion (**Figure 1B**).

4.1.5. Role of IL-17 secretion by T cells

Synovial CD4 T cells produce pro-inflammatory cytokines themselves (**Table 1**). Among these, IL-17 expression is increased in the synovial tissue of RA patients [63], its levels correlate with disease activity [64] and it has a predominant role in rheumatoid pathology [65]. This cytokine is produced by Th17 cells that are critical drivers of synovitis [66]. In the synovium, IL-17 stimulates the production of pro-inflammatory cytokines by rheumatoid synovial cells [67, 68], triggers osteoclastogenesis [69] and impairs cartilage repair [70]. Methotrexate, a first-line conventional therapeutic agent in RA, attenuates IL-17 production by peripheral blood mononuclear cells in vitro [71], supporting the pathogenic role of this cytokine.

Interestingly, the balance between Th17 and regulatory T cells (Treg), which exert anti-inflammatory functions, is shifted towards the Th17 subset in RA [72]. The first hypothesis explaining the excessive Th17 response in RA is that it might be an enhanced Th17 differentiation due to the inflammatory environment. Th17 cells differentiate in the presence of IL-1β, IL-6 and IL-23 [73], which are secreted by activated macrophages and dendritic cells in inflammatory conditions [74]. Supporting this hypothesis, both IL-23 and IL-6 levels are increased in patients with RA [75, 76]. IL-23 levels correlate with the activity of early arthritis [77]. A second hypothesis would be that intrinsic alterations in naïve CD4 T cells might prone Th17 rather than Treg differentiation. Supporting this hypothesis, naïve RA T cells overexpress glucose-6-phosphate dehydrogenase (G6PD), which causes insufficient activation of ataxia telangiectasia mutated (ATM), leading to biased differentiation of CD4 T cells towards Th17 and Th1 subsets (**Table 2**) [78].

4.2. Abnormalities in CD4 T cell activation and signalling

As mentioned in the previous sections, CD4 T cell activation in the synovium is a key event in RA pathology. CD4 T cell activation is initiated by interaction of the TCR with the antigen-MHC-II expressed on the surface of an APC. Engagement of TCR/MHC-II-antigen complex triggers the activation of intracellular signalling networks in which phosphorylation plays a decisive role. The kinases Lck and ZAP70 are rapidly activated after TCR stimulation and activate downstream effectors such as extracellular signal-regulated kinase (ERK) to induce

Cytokine	Pathogenic role			
TNF-α	Activates leukocytes, synovial fibroblasts, endothelial cells and osteoclasts			
	Induces production of inflammatory cytokines			
	Enhances metalloproteinase expression			
	Suppresses Treg cells			
IFN-γ	 Increases antigen presentation Activates macrophages Increases chemokine secretion 			
IL-1	Activates leukocytes, synovial fibroblasts, endothelial cells and osteoclasts			
	Induces production of matrix proteinases			
IL-6	Activates leukocytes and osteoclasts			
	Stimulates antibody production			
IL-17	Induces production of inflammatory cytokines			
	Activates innate immune cells			
	Increases osteoclastogenesis			
	Stimulates neutrophil recruitment			
IL-21	Activates Th17 and B cells			

Table 1. Pathogenic role of cytokines secreted by CD4 T cells in the RA synovium.

gene expression and cell proliferation. In physiologic conditions, signalling downstream the TCR is tightly regulated by proteins such as phosphatases. In T cell-mediated autoimmune pathologies, such as RA, intracellular signalling is deregulated, leading to alterations in T cell responses.

Another physiological mechanism regulating T cell responses and preventing autoimmunity is the elimination of self-reactive T cells. This mechanism is called tolerance and occurs both on immature T cells in the thymus (central tolerance) and on mature circulating T cells (peripheral tolerance). In RA, activation of CD4 T cells by self-antigens seems to be permitted by losing peripheral or central tolerance and promoted by enhanced sensitivity to self-antigens due to alterations in signalling networks integrating extracellular stimuli.

Several observations indicate that peripheral blood, and not only synovial-infiltrating T cells, show hyper-activation in RA patients [79, 80]. An aberrant function or expression of signalling molecules, some of them regulating T cell responses, has been found in CD4 T cells of RA patients (Table 2) and will be discussed below.

Protein	Alteration	Consequence in CD4 T cells	Reference(s)
G6PD	Overexpression	Insufficient ATM activation	[78]
		Hyperproliferation	
		• Increased Th1/Th17 differentiation	
LYP (rs2476601 SNP)	Gain of function mutation	• T cell hyporesponsiveness	[88–92]
TC-PTP (rs1893217(C) SNP)) Reduced expression	Decreased STAT5 phosphorylation	[95, 96]
		Decreased FOXP3 expression upon activation	
CDC25B	Reduced expression	Not reported	[99]
DUSP7	Reduced expression	Not reported	[99]
B-RAF K-RAS	Overexpression	 Increased ERK phosphory- lation and signalling 	[101]
		Autoreactive response to citrullinated peptides	
PD-1	Reduced expression	Not reported	[91–95]
Telomerase	Insufficient induction	Susceptibility to apoptosis	[12]
MRE11A	Reduced expression	Telomeric damage	[11]
		• Senescence	

G6PD, glucose-6-phosphate dehydrogenase; ATM, ataxia telangiectasia mutated; LYP, lymphoid-specific tyrosine phosphatase; TC-PTP, T cell protein tyrosine phosphatase; STAT5, signal transducer and activator of transcription 5; FOXP3, forkhead box P3; CDC25B, cell division cycle 25 B; DUSP7, dual-specificity phosphatase 7; ERK, extracellular signal-regulated kinase; PD-1, programmed death 1; MRE11A, meiotic recombination 11 homolog A

Table 2. Alterations in gen/protein expression or activity found in CD4 T cells from RA patients and their phenotype.

4.2.1. PD-1

Programmed death-1 (PD-1) receptor is inducibly expressed on CD4 T cells upon activation through the TCR [81]. Upon binding to its ligands during TCR stimulation, PD-1 delivers inhibitory signals that suppress T cell activation and proliferation and impair T cell survival [82]. A set of SNPs in the gene coding for PD-1 are linked to RA [83–85], and PD-1 expression is decreased in T cells from RA patients [86]. This reduced expression would lead to a defect in peripheral tolerance, favouring autoimmunity.

4.2.2. LYP

The lymphoid-specific tyrosine phosphatase (LYP) is encoded by the gene *PTPN22*. This protein is exclusively expressed in cells of the immune system and in T cells negatively regulates TCR signalling by inactivating the kinases Lck and ZAP70 [87]. Therefore, LYP is an important inhibitor of signalling downstream the TCR. The SNP rs2476601 in *PTPN22* is associated with RA [88, 89]. The pathological function of this SNP, which results in the LYP mutant R620W, remains controversial. Various reports show that the LYP R620W variant is more effective in

downregulating TCR signalling than the LYP WT [90, 91]. In this situation, LYP R620W would trigger autoimmunity because it would suppress TCR signalling of autoreactive T cells during negative selection in the thymus, promoting their survival and compromising central tolerance [92]. Molecular mechanisms leading to autoimmunity in the presence of this polymorphism should be further studied.

4.2.3. TC-PTP

The T cell-phosphotyrosine phosphatase (TC-PTP) is encoded by the gene PTPN2. This tyrosine phosphatase negatively regulates TCR and JAK-STAT signalling, being an inhibitor of T cell activation [93, 94]. The SNP rs1893217(C) in PTPN2 is associated with juvenile idiopathic arthritis and results in decreased gene expression [95]. Strikingly, decreased phosphorylation of STAT5 and reduced FOXP3 expression are found in cells carrying this mutation [96]. Because FOXP3 is the master regulator of Treg differentiation [97], this SNP might cause abnormalities in Treg functions, resulting in increased inflammation. The mechanism for this phenotype should be investigated.

4.2.4. CDC25B

The dual-specificity phosphatase cell division cycle 25 B (CDC25B) positively regulates cell proliferation by promoting G2/M transition [98]. Recently, our group has found a reduced expression of this phosphatase in CD4 T cells of patients diagnosed with early arthritis [99]. Importantly, altered CDC25B levels associate to the activity of the disease. Whether this alteration causes or is a consequence of the inflammatory environment characteristic of RA, and its effect in T cell responses will need further investigation.

4.2.5. Regulators of ERK signalling

As mentioned before, ERK is a key effector molecule downstream TCR activation. Hence, defective regulation of ERK phosphorylation levels could lead to aberrant T cell responses. The expression of some ERK regulator is altered in T cells of RA patients.

The dual-specificity phosphatase 7 (DUSP7) negatively regulates ERK phosphorylation and activity [100]. Although its role in T cells has not been addressed, it is conceivable that DUSP7 could be a negative regulator of MAPK signalling in T cells being activated. CD4 T cells of patients with seropositive early arthritis have reduced expression of DUSP7 [99]. The fact that defective expression is restricted to seropositive patients could indicate a role of this phosphatase in T/B cooperation. Further investigation is needed to determine the functional significance of DUSP7 in T cells.

The GTPase K-RAS and the kinase B-RAF are positive regulators of ERK signalling upon TCR stimulation. A higher TCR-induced ERK phosphorylation results in a lower T cell activation threshold, contributing to autoimmunity. K-RAS and B-RAF are overexpressed in T cells of RA patients [101]. Interestingly, overexpression of B-RAF and K-RAS increases the activation of CD4 T cells of healthy donors by a citrullinated vimentin peptide. This finding provides support to the notion that higher CD4 sensitivity could cause loss of peripheral tolerance in RA patients.

4.3. Abnormalities in CD4 T cell repertoire and aging

The ability of the adaptive immune system to respond to the large diversity of pathogens found throughout life depends on the generation of a wide TCR repertoire. This repertoire is generated in the thymus, where the V, D and J segments of the TCR rearrange randomly. Newly generated naive T cells migrate from the thymus to the periphery to exert their functions. The thymic output, however, declines throughout life. In the elderly the thymus no longer functions as a source of new naïve T cells, which have to be produced by replication of mature peripheral T cells, a process called homeostatic proliferation [102]. The expansion of peripheral T cell clones generates a contraction in T cell repertoire and induces a phenotype of replicative stress that is characteristic of aged people [103]. Clone expansion of peripheral cells might favour an increased presence of autoreactive clones. Consistent with this idea, autoimmune signs such as autoantibody production are higher in elderly individuals [104].

Repertoire contraction and clonally expanded populations in the CD4 compartment have been reported in RA [105]. Clonal expansion was initially interpreted as a consequence of specific responses to synovial self-antigens, but this hypothesis is unlikely. Contraction in CD4 T cell diversity is not limited to the memory compartment, but involves also naïve T cells [106]. This seems to be due to an accelerated aging of the immune system in RA patients, in which the thymus function is lost earlier than in healthy people [107].

A hallmark of immune aging is the accumulation of end-differentiated effector CD4 T cells that lack expression of the costimulatory receptor CD28 [108]. Indeed, the frequency of CD4+ CD28– lymphocytes is higher in RA patients [109, 110]. These cells are producers of IFN- γ , display cytotoxic functions and are autoreactive [109, 111, 112]. Such phenotype could be mediated, at least in part, by increased expression of the NK cell-activating receptor NKG2D. Ligands of NKG2D are highly expressed in inflamed synovium [113].

Another hallmark of cellular aging is telomere shortening [114], and lymphocytes from RA patients show premature telomeric loss [115]. In naïve CD4 T cells, this is due to insufficient upregulation of telomerase activity (**Table 2**), which in addition promotes apoptosis in these cells [12]. Excessive loss of naïve T cells will further stimulate homeostatic proliferation of effector T cells, providing a positive feedback loop of replicative stress.

Recently, another alteration in DNA repair machinery was found in CD4 T cells from RA patients [11]. The expression of repair nuclease MRE11A is decreased in these cells, leading to telomeric damage and upregulated senescence markers (**Table 2**).

4.4. CD4 T cells in extra-articular disease

Although the main site of inflammation in RA is the synovium, pro-inflammatory cytokines and activated cells are released to the bloodstream, leading to systemic inflammation. This inflammatory state has multiple ExRA on distant organs, such as skin, lungs, heart, blood or bone [116]. Smoking habit and autoantibodies predispose to severe ExRA [117]. Several systemic

pathologies are frequent in RA patients, such as systemic vasculitis, interstitial lung disease and pericarditis, which is the most common cardiac complication [116]. We focus here on CVD.

Chronic inflammation generates a pro-atherogenic environment in RA. Indeed, RA patients have increased risk of cardiovascular death [118] and higher incidence of atherosclerotic heart disease [119]. Atherosclerosis is an inflammatory process in which the plaque, constituted by lipid accumulation on arterial walls, causes endothelial injury and activation. This promotes the recruitment of leukocytes, which culminates in the disruption of the plaque and thrombosis. Vascular inflammation in atherosclerosis and synovial inflammation in RA share features of immune activation, including accumulation of inflammatory macrophages and T cells, production of inflammatory cytokines and degradation of the extracellular matrix. High levels of soluble factors such as C-reactive protein, TNF- α and IL-6 are associated with coronary artery disease [120–122]. These cytokines are also elevated in chronic inflammation, which renders lipoproteins more atherogenic, reduces the repair of injured endothelium and upregulates the expression of endothelial adhesion molecules, which enhance leukocyte recruitment [123]. Consistent with a role of systemic inflammation in atherosclerosis, RA therapies based on methotrexate and TNF- α antagonists decrease CVD rates [124, 125].

As mentioned before, the CD4+CD28– T cell subset is expanded in RA [109, 110]. This T cell subset is also expanded in patients with unstable angina (UA) [126], a pathology in which the atheroma plaque is disrupted causing thrombosis. The percentage of CD4+CD28– cells correlates with recurrence of UA, pointing to a direct role of these cells in the progression of the pathology [127]. In addition, expanded CD4+ CD28– found in the atherosclerotic lesion includes large monoclonal populations, suggesting that these cells can recognise antigens in the atheroma plaque [128]. Consistently, RA patients with expansion of circulating CD4+CD28– cells show preclinical atherosclerotic changes, including endothelial dysfunction [129]. The implication of CD4+CD28– cells in atherosclerosis is further supported by anti-TNF therapy, which normalises CD28 expression [130] and decreases CVD rates [125].

5. Conclusion

RA is a chronic inflammatory disease characterised by synovitis and systemic features, such as exacerbated atherosclerosis. CD4 T cells are key mediators of tissue damage, both in the joint and in extra-articular lesions, through a variety of mechanisms. Certain alleles of the MHC-II as well as different alterations of signalling molecules and checkpoints for activation seem to favour self-antigen recognition, activation and break of tolerance. Besides, abnormalities found in CD4 T cell repertoire and phenotype in patients with RA strongly suggest that in these patients there is an accelerated aging of the immune system that leads to oligoclonality and senescence of T cells, making these lymphocytes autoreactive. Understanding the mechanisms underlying these systemic alterations will be essential for the development of more effective therapies for RA treatment.

Author details

Patricia Castro-Sánchez and Pedro Roda-Navarro*

*Address all correspondence to: proda@ucm.es

Department of Microbiology I (Immunology), School of Medicine, Universidad Complutense de Madrid, 12 de Octubre Health Research Institute (imas12), Madrid, Spain

References

- [1] Jacobs P, Bissonnette R, Guenther LC. Socioeconomic burden of immune-mediated inflammatory diseases—Focusing on work productivity and disability. The Journal of Rheumatology Supplement. 2011;88:55-61. DOI: 10.3899/jrheum.110901
- [2] Alamanos Y, Voulgari PV, Drosos AA. Incidence and prevalence of rheumatoid arthritis, based on the 1987 American College of Rheumatology criteria: A systematic review. Seminars in Arthritis and Rheumatism. 2006;36(3):182-188. DOI: 10.1016/j. semarthrit.2006.08.006
- [3] Rossini M, Rossi E, Bernardi D, Viapiana O, Gatti D, Idolazzi L, et al. Prevalence and incidence of rheumatoid arthritis in Italy. Rheumatology International. 2014;34(5):659-664. DOI: 10.1007/s00296-014-2974-6
- [4] Andrianakos A, Trontzas P, Christoyannis F, Kaskani E, Nikolia Z, Tavaniotou E, et al. Prevalence and management of rheumatoid arthritis in the general population of Greece—The ESORDIG study. Rheumatology (Oxford). 2006;45(12):1549-1554. DOI: 10.1093/rheumatology/kel140
- [5] Carmona L, Villaverde V, Hernández-García C, Ballina J, Gabriel R, Laffon A; EPISER Study Group. The prevalence of rheumatoid arthritis in the general population of Spain. Rheumatology (Oxford). 2002;41(1):88-95
- [6] Neovius M, Simard JF, Askling J; ARTIS study group. Nationwide prevalence of rheumatoid arthritis and penetration of disease-modifying drugs in Sweden. Annals of the Rheumatic Diseases. 2011;70(4):624-629. DOI: 10.1136/ard.2010.133371
- [7] Langley PC, Mu R, Wu M, Dong P, Tang B. The impact of rheumatoid arthritis on the burden of disease in urban China. Journal of Medical Economics. 2011;14(6):709-719. DOI: 10.3111/13696998.2011.611201
- [8] Silman AJ, Pearson JE. Epidemiology and genetics of rheumatoid arthritis. Arthritis Research. 2002;4(Suppl 3):S265-S272. DOI: 10.1186/ar578
- [9] Capellino S, Cosentino M, Wolff C, Schmidt M, Grifka J, Straub RH. Catecholamine-producing cells in the synovial tissue during arthritis: Modulation of sympathetic neurotransmitters as new therapeutic target. Annals of the Rheumatic Diseases. 2010;69(10):1853-1860. DOI: 10.1136/ard.2009.119701

- [10] Martin G, Kanaan S, Azzouz D, Balandraud N, Picard C, Auger I, et al. A6.40 Copy number increase of TLR7 and TLR8 genes in men with rheumatoid arthritis. Annals of the Rheumatic Diseases. 2015;74:A72
- [11] Li Y, Shen Y, Hohensinner P, Ju J, Wen Z, Goodman SB, et al. Deficient Activity of the Nuclease MRE11A Induces T Cell Aging and Promotes Arthritogenic Effector Functions in Patients with Rheumatoid Arthritis. Immunity. 2016;45(4):903-916. DOI: 10.1016/j. immuni.2016.09.013
- [12] Fujii H, Shao L, Colmegna I, Goronzy JJ, Weyand CM. Telomerase insufficiency in rheumatoid arthritis. Proceedings of the National Academy of Sciences of the United States of America. 2009;106(11):4360-4365. DOI: 10.1073/pnas.0811332106
- [13] Watson DJ, Rhodes T, Guess HA. All-cause mortality and vascular events among patients with rheumatoid arthritis, osteoarthritis, or no arthritis in the UK General Practice Research Database. The Journal of Rheumatology. 2003;30(6):1196-1202
- [14] Gonzalez A, Maradit-Kremers H, Crowson CS, Nicola PJ, Davis JM 3rd, Therneau TM, et al. The widening mortality gap between rheumatoid arthritis patients and the general population. Arthritis and Rheumatism. 2007;56(11):3583-3587. DOI: 10.1002/art.22979
- [15] Daha NA, Toes RE. Rheumatoid arthritis: Are ACPA-positive and ACPA-negative RA the same disease?. Nature Reviews. Rheumatology. 2011;7(4):202-203. DOI: 10.1038/nrrheum.2011.28
- [16] Okada Y, Wu D, Trynka G, Raj T, Terao C, Ikari K, et al. Genetics of rheumatoid arthritis contributes to biology and drug discovery. Nature. 2014;506(7488):376-381. DOI: 10.1038/nature12873
- [17] Tobón GJ, Youinou P, Saraux A. The environment, geo-epidemiology, and autoimmune disease: Rheumatoid arthritis. Autoimmunity Reviews. 2010;9(5):A288-A292. DOI: 10.1016/j.autrev.2009.11.019
- [18] Gregersen PK, Silver J, Winchester RJ. The shared epitope hypothesis. An approach to understanding the molecular genetics of susceptibility to rheumatoid arthritis. Arthritis and Rheumatism. 1987;30(11):1205-1213
- [19] du Montcel ST, Michou L, Petit-Teixeira E, Osorio J, Lemaire I, Lasbleiz S, et al. New classification of HLA-DRB1 alleles supports the shared epitope hypothesis of rheumatoid arthritis susceptibility. Arthritis and Rheumatism. 2005;52(4):1063-1068. DOI: 10.1002/art.20989
- [20] MacGregor AJ, Snieder H, Rigby AS, Koskenvuo M, Kaprio J, Aho K, et al. Characterizing the quantitative genetic contribution to rheumatoid arthritis using data from twins. Arthritis and Rheumatism. 2000;43(1):30-37. DOI: 10.1002/1529-0131(200001)43:1<30::AID-ANR5>3.0.CO;2-B
- [21] Liu X, Guo J, Jia Y, Zhao Y, Liu X, Cheng F, et al. HLA-DRB1 shared epitope-dependent DR-DQ haplotypes are associated with both anti-CCP-positive and -negative rheumatoid arthritis in Chinese Han. PLoS One. 2013;8(8):e71373. DOI: 10.1371/journal.pone.0071373

- [22] Mohan VK, Ganesan N, Gopalakrishnan R, Venkatesan V. HLA-DRB1 shared epitope alleles in patients with rheumatoid arthritis: Relation to autoantibodies and disease severity in a south Indian population. International Journal of Rheumatic Diseases. 2016. DOI: 10.1111/1756-185X.12948. [Epub ahead of print]
- [23] Lagha A, Messadi A, Boussaidi S, Kochbati S, Tazeghdenti A, Ghazouani E, et al. HLA DRB1/DQB1 alleles and DRB1-DQB1 haplotypes and the risk of rheumatoid arthritis in Tunisians: A population-based case-control study. HLA. 2016;88(3):100-109. DOI: 10.1111/tan.12855
- [24] Louthrenoo W, Kasitanon N, Wangkaew S, Kuwata S, Takeuchi F. Distribution of HLA-DR alleles among Thai patients with rheumatoid arthritis. Human Immunology. 2015;76(2-3):113-117. DOI: 10.1016/j.humimm.2015.01.018
- [25] Balsa A, Minaur NJ, Pascual-Salcedo D, McCabe C, Balas A, Fiddament B, et al. Class II MHC antigens in early rheumatoid arthritis in Bath (UK) and Madrid (Spain). Rheumatology (Oxford). 2000;39(8):844-849
- [26] Viatte S, Plant D, Han B, Fu B, Yarwood A, Thomson W, et al. Association of HLA-DRB1 haplotypes with rheumatoid arthritis severity, mortality, and treatment response. JAMA. 2015;**313**(16):1645-1656. DOI: 10.1001/jama.2015.3435
- [27] Damgaard D, Friberg Bruun Nielsen M, Quisgaard Gaunsbaek M, Palarasah Y, Svane-Knudsen V, Nielsen CH. Smoking is associated with increased levels of extracellular peptidylarginine deiminase 2 (PAD2) in the lungs. Clinical and Experimental Rheumatology. 2015;33(3):405-408
- [28] Meng W, Zhu Z, Jiang X, Too CL, Uebe S, Jagodic M, et al. DNA methylation mediates genotype and smoking interaction in the development of anti-citrullinated peptide antibody-positive rheumatoid arthritis. Arthritis Research and Therapy. 2017;**19**(1):71. DOI: 10.1186/s13075-017-1276-2
- [29] Klareskog L, Stolt P, Lundberg K, Källberg H, Bengtsson C, Grunewald J, et al. A new model for an etiology of rheumatoid arthritis: Smoking may trigger HLA-DR (shared epitope)-restricted immune reactions to autoantigens modified by citrullination. Arthritis and Rheumatism. 2006;54(1):38-46. DOI: 10.1002/art.21575
- [30] de Pablo P, Chapple IL, Buckley CD, Dietrich T. Periodontitis in systemic rheumatic diseases. Nature Reviews. Rheumatology. 2009;5(4):218-224. DOI: 10.1038/nrrheum.2009.28
- [31] Wegner N, Wait R, Sroka A, Eick S, Nguyen K, Lundberg K, et al. Peptidylarginine deiminase from *Porphyromonas gingivalis* citrullinates human fibrinogen and α-enolase: Implications for autoimmunity in rheumatoid arthritis. Arthritis and Rheumatism. 2010;62(9):2662-2672. DOI: 10.1002/art.27552
- [32] Tsuda R, Ozawa T, Kobayashi E, Hamana H, Taki H, Tobe K, et al. Monoclonal antibody against citrullinated peptides obtained from rheumatoid arthritis patients reacts with numerous citrullinated microbial and food proteins. Arthritis & Rheumatology (Hoboken, N.J.). 2015;67(8):2020-2031. DOI: 10.1002/art.39161

- [33] Kraan MC, Reece RJ, Smeets TJ, Veale DJ, Emery P, Tak PP. Comparison of synovial tissues from the knee joints and the small joints of rheumatoid arthritis patients: Implications for pathogenesis and evaluation of treatment. Arthritis and Rheumatism. 2002;46(8):2034-2038. DOI: 10.1002/art.10556
- [34] Tak PP, Smeets TJ, Daha MR, Kluin PM, Meijers KA, Brand R, et al. Analysis of the synovial cell infiltrate in early rheumatoid synovial tissue in relation to local disease activity. Arthritis and Rheumatism. 1997;40(2):217-225
- [35] Alvaro-Gracia JM, Zvaifler NJ, Firestein GS. Cytokines in chronic inflammatory arthritis. V. Mutual antagonism between interferon-gamma and tumor necrosis factor-alpha on HLA-DR expression, proliferation, collagenase production, and granulocyte macrophage colony-stimulating factor production by rheumatoid arthritis synoviocytes. The Journal of Clinical Investigation. 1990;86(6):1790-1798
- [36] Bottini N, Firestein GS. Duality of fibroblast-like synoviocytes in RA: Passive responders and imprinted aggressors. Nature Reviews. Rheumatology. 2013;9(1):24-33. DOI: 10.1038/nrrheum.2012.190
- [37] Konttinen YT, Ceponis A, Takagi M, Ainola M, Sorsa T, Sutinen M, et al. New collagenolytic enzymes/cascade identified at the pannus-hard tissue junction in rheumatoid arthritis: Destruction from above. Matrix Biology: Journal of the International Society for Matrix Biology. 1998;17(8-9):585-601
- [38] Gravallese EM, Manning C, Tsay A, Naito A, Pan C, Amento E, et al. Synovial tissue in rheumatoid arthritis is a source of osteoclast differentiation factor. Arthritis and Rheumatism. 2000;43(2):250-258. DOI: 10.1002/1529-0131(200002)43:2<250::AID-ANR3>3.0.CO;2-P
- [39] Lam J, Takeshita S, Barker JE, Kanagawa O, Ross FP, Teitelbaum SL. TNF-alpha induces osteoclastogenesis by direct stimulation of macrophages exposed to permissive levels of RANK ligand. The Journal of Clinical Investigation. 2000;106(12):1481-1488. DOI: 10.1172/JCI11176
- [40] Axmann R, Böhm C, Krönke G, Zwerina J, Smolen J, Schett G. Inhibition of interleukin-6 receptor directly blocks osteoclast formation in vitro and in vivo. Arthritis and Rheumatism. 2009;60(9):2747-2756. DOI: 10.1002/art.24781
- [41] Honorati MC, Neri S, Cattini L, Facchini A. Interleukin-17, a regulator of angiogenic factor release by synovial fibroblasts. Osteoarthritis and Cartilague. 2006;14(4):345-352. DOI: 10.1016/j.joca.2005.10.004
- [42] Koch AE, Harlow LA, Haines GK, Amento EP, Unemori EN, Wong WL, et al. Vascular endothelial growth factor. A cytokine modulating endothelial function in rheumatoid arthritis. Journal of Immunology (Baltimore). 1994;152(8):4149-4156
- [43] Giatromanolaki A, Sivridis E, Maltezos E, Athanassou N, Papazoglou D, Gatter KC, et al. Upregulated hypoxia inducible factor- 1α and -2α pathway in rheumatoid arthritis and osteoarthritis. Arthritis Research and Therapy. 2003;5(4):R193-R201. DOI: 10.1186/ar756

- [44] Cojocaru M, Cojocaru IM, Silosi I, Vrabie CD, Tanasescu R. Extra-articular manifestations in rheumatoid arthritis. Maedica. 2010;5(4):286-291
- [45] Duke O, Panayi GS, Janossy G, Poulter LW. An immunohistological analysis of lymphocyte subpopulations and their microenvironment in the synovial membranes of patients with rheumatoid arthritis using monoclonal antibodies. Clinical and Experimental Immunology. 1982;49(1):22-30
- [46] Krummel MF, Allison JP. CD28 and CTLA-4 have opposing effects on the response of T cells to stimulation. The Journal of Experimental Medicine. 1995;**182**(2):459-465
- [47] Parry RV, Chemnitz JM, Frauwirth KA, Lanfranco AR, Braunstein I, Kobayashi SV, et al. CTLA-4 and PD-1 receptors inhibit T-cell activation by distinct mechanisms. Molecular and Cellular Biology. 2005;25(21):9543-9553. DOI: 10.1128/MCB.25.21.9543-9553.2005
- [48] Herrero-Beaumont G, Martínez Calatrava MJ, Castañeda S. Abatacept mechanism of action: Concordance with its clinical profile. Reumatologia Clinica. 2012;8(2):78-83. DOI: 10.1016/j.reuma.2011.08.002
- [49] Emery P, Burmester GR, Bykerk VP, Combe BG, Furst DE, Barré E, et al. Evaluating drugfree remission with abatacept in early rheumatoid arthritis: Results from the phase 3b, multicentre, randomised, active-controlled AVERT study of 24 months, with a 12-month, double-blind treatment period. Annals of the Rheumatic Diseases. 2015;74(1):19-26. DOI: 10.1136/annrheumdis-2014-206106
- [50] Kubo S, Nakano K, Nakayamada S, Hirata S, Fukuyo S, Sawamukai N, et al. Clinical, radiographic and functional efficacy of abatacept in routine care for rheumatoid arthritis patients: Abatacept Leading Trial for RA on Imaging Remission (ALTAIR) study. Clinical and Experimental Rheumatology. 2016;34(5):834-841
- [51] Humby F, Bombardieri M, Manzo A, Kelly S, Blades MC, Kirkham B, et al. Ectopic lymphoid structures support ongoing production of class-switched autoantibodies in rheumatoid synovium. PLoS Medicine. 2009;6(1):e1. DOI: 10.1371/journal.pmed.0060001
- [52] Weyand CM, Goronzy JJ. Ectopic germinal center formation in rheumatoid synovitis. Annals of the New York Academy of Sciences. 2003;987:140-149
- [53] Weyand CM, Kurtin PJ, Goronzy JJ. Ectopic lymphoid organogenesis: A fast track for autoimmunity. The American Journal of Pathology. 2001;159(3):787-793. DOI: 10.1016/ S0002-9440(10)61751-8
- [54] Takemura S, Klimiuk PA, Braun A, Goronzy JJ, Weyand CM. T cell activation in rheumatoid synovium is B cell dependent. Journal of Immunology (Baltimore). 2001;167(8):4710-4718
- [55] Cho ML, Yoon CH, Hwang SY, Park MK, Min SY, Lee SH, et al. Effector function of type II collagen-stimulated T cells from rheumatoid arthritis patients: Cross-talk between T cells and synovial fibroblasts. Arthritis and Rheumatism. 2004;50(3):776-784. DOI: 10.1002/art.20106

- [56] Rezzonico R, Burger D, Dayer JM. Direct contact between T lymphocytes and human dermal fibroblasts or synoviocytes down-regulates types I and III collagen production via cell-associated cytokines. The Journal of Biological Chemistry. 1998;273(30):18720-18728
- [57] McInnes IB, Leung BP, Sturrock RD, Field M, Liew FY. Interleukin-15 mediates T celldependent regulation of tumor necrosis factor-alpha production in rheumatoid arthritis. Nature Medicine. 1997;3(2):189-195
- [58] Brennan FM, Hayes AL, Ciesielski CJ, Green P, Foxwell BM, Feldmann M. Evidence that rheumatoid arthritis synovial T cells are similar to cytokine-activated T cells: Involvement of phosphatidylinositol 3-kinase and nuclear factor kappaB pathways in tumor necrosis factor alpha production in rheumatoid arthritis. Arthritis and Rheumatism. 2002;46(1):31-41. DOI: 10.1002/1529-0131(200201)46:1<31::AID-ART10029>3.0.CO;2-5
- [59] González-Alvaro I, Domínguez-Jiménez C, Ortiz AM, Núñez-González V, Roda-Navarro P, Fernández-Ruiz E, et al. Interleukin-15 and interferon-gamma participate in the crosstalk between natural killer and monocytic cells required for tumour necrosis factor production. Arthritis Research & Therapy. 2006;8(4):R88. DOI: 10.1186/ar1955
- [60] Feldmann M. Development of anti-TNF therapy for rheumatoid arthritis. Nature Reviews. Immunology. 2002;2(5):364-371. DOI: 10.1038/nri802
- [61] Kotake S, Udagawa N, Hakoda M, Mogi M, Yano K, Tsuda E, et al. Activated human T cells directly induce osteoclastogenesis from human monocytes: Possible role of T cells in bone destruction in rheumatoid arthritis patients. Arthritis and Rheumatism. 2001;44(5):1003-10012. DOI: 10.1002/1529-0131(200105)44:5<1003::AID-ANR179>3.0.CO;2-#
- [62] Kim HR, Kim KW, Kim BM, Jung HG, Cho ML, Lee SH. Reciprocal activation of CD4+ T cells and synovial fibroblasts by stromal cell-derived factor 1 promotes RANKL expression and osteoclastogenesis in rheumatoid arthritis. Arthritis & Rheumatology. 2014;66(3):538-548. DOI: 10.1002/art.38286
- [63] Li N, Wang JC, Liang TH, Zhu MH, Wang JY, Fu XL, et al. Pathologic finding of increased expression of interleukin-17 in the synovial tissue of rheumatoid arthritis patients. International Journal of Clinical and Experimental Pathology. 2013;6(7):1375-1379
- [64] Metawi SA, Abbas D, Kamal MM, Ibrahim MK. Serum and synovial fluid levels of interleukin-17 in correlation with disease activity in patients with RA. Clinical Rheumatology. 2011;**30**(9):1201-1207. DOI: 10.1007/s10067-011-1737-y
- [65] Benedetti G, Miossec P. Interleukin 17 contributes to the chronicity of inflammatory diseases such as rheumatoid arthritis. European Journal of Immunology. 2014;44(2):339-347. DOI: 10.1002/eji.201344184
- [66] van Hamburg JP, Asmawidjaja PS, Davelaar N, Mus AM, Colin EM, Hazes JM, et al. Th17 cells, but not Th1 cells, from patients with early rheumatoid arthritis are potent inducers of matrix metalloproteinases and proinflammatory cytokines upon synovial fibroblast interaction, including autocrine interleukin-17A production. Arthritis and Rheumatism. 2011;63(1):73-83. DOI: 10.1002/art.30093

- [67] Fossiez F, Djossou O, Chomarat P, Flores-Romo L, Ait-Yahia S, Maat C, et al. T cell interleukin-17 induces stromal cells to produce proinflammatory and hematopoietic cytokines. The Journal of Experimental Medicine. 1996;183(6):2593-2603
- [68] Jovanovic DV, Di Battista JA, Martel-Pelletier J, Jolicoeur FC, He Y, Zhang M, et al. IL-17 stimulates the production and expression of proinflammatory cytokines, IL-beta and TNF-alpha, by human macrophages. Journal of Immunology. 1998;160(7):3513-3521
- [69] Sato K, Suematsu A, Okamoto K, Yamaguchi A, Morishita Y, Kadono Y, et al. Th17 functions as an osteoclastogenic helper T cell subset that links T cell activation and bone destruction. The Journal of Experimental Medicine. 2006;**203**(12):2673-2682. DOI: 10.1084/jem.20061775
- [70] Schminke B, Trautmann S, Mai B, Miosge N, Blaschke S. Interleukin 17 inhibits progenitor cells in rheumatoid arthritis cartilage. European Journal of Immunology. 2016;46(2):440-445. DOI: 10.1002/eji.201545910
- [71] Li Y, Jiang L, Zhang S, Yin L, Ma L, He D, et al. Methotrexate attenuates the Th17/IL-17 levels in peripheral blood mononuclear cells from healthy individuals and RA patients. Rheumatology International. 2012;32(8):2415-2422. DOI: 10.1007/s00296-011-1867-1
- [72] Niu Q, Cai B, Huang ZC, Shi YY, Wang LL. Disturbed Th17/Treg balance in patients with rheumatoid arthritis. Rheumatology International. 2012;**32**(9):2731-2736. DOI: 10.1007/s00296-011-1984-x
- [73] Acosta-Rodriguez EV, Napolitani G, Lanzavecchia A, Sallusto F. Interleukins 1beta and 6 but not transforming growth factor-beta are essential for the differentiation of interleukin 17-producing human T helper cells. Nature Immunology. 2007;8(9):942-949. DOI: 10.1038/ni1496
- [74] Segura E, Touzot M, Bohineust A, Cappuccio A, Chiocchia G, Hosmalin A, et al. Human inflammatory dendritic cells induce Th17 cell differentiation. Immunity. 2013;38(2):336-348. DOI: 10.1016/j.immuni.2012.10.018
- [75] Kim HR, Cho ML, Kim KW, Juhn JY, Hwang SY, Yoon CH, et al. Up-regulation of IL-23p19 expression in rheumatoid arthritis synovial fibroblasts by IL-17 through PI3-kinase-, NF-kappaB- and p38 MAPK-dependent signalling pathways. Rheumatology (Oxford, England). 2007;46(1):57-64. DOI: 10.1093/rheumatology/kel159
- [76] Hirano T, Matsuda T, Turner M, Miyasaka N, Buchan G, Tang B, et al. Excessive production of interleukin 6/B cell stimulatory factor-2 in rheumatoid arthritis. European Journal of Immunology. 1988;18(11):1797-1801
- [77] Rasmussen TK, Andersen T, Hvid M, Hetland ML, Hørslev-Petersen K, Stengaard-Pedersen K, et al. Increased interleukin 21 (IL-21) and IL-23 are associated with increased disease activity and with radiographic status in patients with early rheumatoid arthritis. The Journal of Rheumatology. 2010;37(10):2014-2020. DOI: 10.3899/jrheum.100259
- [78] Yang Z, Shen Y, Oishi H, Matteson EL, Tian L, Goronzy JJ, et al. Restoring oxidant signaling suppresses proarthritogenic T cell effector functions in rheumatoid arthritis. Science Translational Medicine. 2016;8(331):331ra38. DOI: 10.1126/scitranslmed.aad7151

- [79] López-Santalla M, Salvador-Bernáldez M, González-Alvaro I, Castañeda S, Ortiz AM, García-García MI, et al. Tyr³²³-dependent p38 activation is associated with rheumatoid arthritis and correlates with disease activity. Arthritis and Rheumatism. 2011;63(7):1833-1842. DOI: 10.1002/art.30375
- [80] Singh K, Deshpande P, Pryshchep S, Colmegna I, Liarski V, Weyand CM, et al. ERKdependent T-cell receptor threshold calibration in rheumatoid arthritis. Journal of Immunology. 2009;183(12):8258-8267. DOI: 10.4049/jimmunol.0901784
- [81] Keir ME, Butte MJ, Freeman GJ, Sharpe AH. PD-1 and its ligands in tolerance and immunity. Annual Review of Immunology. 2008;26:677-704. DOI: 10.1146/annurev. immunol.26.021607.090331
- [82] Riley JL. PD-1 signaling in primary T cells. Immunological Reviews. 2009;229(1):114-125. DOI: 10.1111/j.1600-065X.2009.00767.x
- [83] Lin SC, Yen JH, Tsai JJ, Tsai WC, Ou TT, Liu HW, et al. Association of a programmed death 1 gene polymorphism with the development of rheumatoid arthritis, but not systemic lupus erythematosus. Arthritis and Rheumatism. 2004;50(3):770-775
- [84] Prokunina L, Padyukov L, Bennet A, de Faire U, Wiman B, Prince J. Association of the PD-1.3A allele of the PDCD1 gene in patients with rheumatoid arthritis negative for rheumatoid factor and the shared epitope. Arthritis and Rheumatism. 2004;50(6):1770-1773. DOI: 10.1002/art.20280
- [85] Kong EK, Prokunina-Olsson L, Wong WH, Lau CS, Chan TM, Alarcón-Riquelme M, et al. A new haplotype of PDCD1 is associated with rheumatoid arthritis in Hong Kong Chinese. Arthritis and Rheumatism. 2005;52(4):1058-1062. DOI: 10.1002/art.20966
- [86] Li S, Liao W, Chen M, Shan S, Song Y, Zhang S, et al. Expression of programmed death-1 (PD-1) on CD4+ and CD8+ T cells in rheumatoid arthritis. Inflammation. 2014;37(1):116-121. DOI: 10.1007/s10753-013-9718-8
- [87] Cloutier JF, Veillette A. Cooperative inhibition of T-cell antigen receptor signaling by a complex between a kinase and a phosphatase. The Journal of Experimental Medicine. 1999;189(1):111-121
- [88] The Wellcome Trust Case Control Consortium. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. Nature. 2007;447(7145):661-678. DOI: 10.1038/nature05911
- [89] Lee AT, Li W, Liew A, Bombardier C, Weisman M, Massarotti EM, et al. The PTPN22 R620W polymorphism associates with RF positive rheumatoid arthritis in a dose-dependent manner but not with HLA-SE status. Genes and Immunity. 2005;6(2):129-133. DOI: 10.1038/sj.gene.6364159
- [90] Vang T, Congia M, Macis MD, Musumeci L, Orrú V, Zavattari P, et al. Autoimmuneassociated lymphoid tyrosine phosphatase is a gain-of-function variant. Nature Genetics. 2005;**37**(12):1317-1319. DOI: 10.1038/ng1673

- [91] Rieck M, Arechiga A, Onengut-Gumuscu S, Greenbaum C, Concannon P, Buckner JH. Genetic variation in PTPN22 corresponds to altered function of T and B lymphocytes. Journal of Immunology (Baltimore). 2007;179(7):4704-4710
- [92] Bottini N, Vang T, Cucca F, Mustelin T. Role of PTPN22 in type 1 diabetes and other autoimmune diseases. Seminars in Immunology. 2008;**18**(4):207-213. DOI: 10.1016/j. smim.2006.03.008
- [93] Wiede F, Shields BJ, Chew SH, Kyparissoudis K, van Vliet C, Galic S, et al. T cell protein tyrosine phosphatase attenuates T cell signaling to maintain tolerance in mice. Journal of Clinical Investigation. 2011;**121**(12):4758-4774. DOI: 10.1172/JCI59492
- [94] Pike KA, Tremblay ML. TC-PTP and PTP1B: Regulating JAK-STAT signaling, controlling lymphoid malignancies. Cytokine. 2016;82:52-57. DOI: 10.1016/j.cyto.2015.12.025
- [95] Thompson SD, Sudman M, Ramos PS, Marion MC, Ryan M, Tsoras M, et al. The susceptibility loci juvenile idiopathic arthritis shares with other autoimmune diseases extend to PTPN2, COG6, and ANGPT1. Arthritis and Rheumatism. 2010;62(11):3265-3276. DOI: 10.1002/art.27688
- [96] Long SA, Cerosaletti K, Wan JY, Ho JC, Tatum M, Wei S, et al. An autoimmune-associated variant in PTPN2 reveals an impairment of IL-2R signaling in CD4(+) T cells. Genes and Immunity. 2011;12(2):116-125. DOI: 10.1038/gene.2010.54
- [97] Fontenot JD, Gavin MA, Rudensky AY. Foxp3 programs the development and function of CD4+CD25+ regulatory T cells. Nature Immunology. 2003;4:330-336. DOI: 10.1038/ni904
- [98] Sur S, Agrawal DK. Phosphatases and kinases regulating CDC25 activity in the cell cycle: Clinical implications of CDC25 overexpression and potential treatment strategies. Molecular and Cellular Biochemistry. 2016;416(1-2):33-46. DOI: 10.1007/s11010-016-2693-2
- [99] Castro-Sánchez P, Ramirez-Munoz R, Lamana A, Ortiz A, González-Álvaro I, Roda-Navarro P. mRNA profilin identifies low levels of phosphatases dual-specific phosphatase-7 (DUSP7) and cell division cycle-25B (CDC25B) in patients with early arthritis. Clinical and Experimental Immunology. 2017;189(1):113-119. DOI: 10.1111/cei.12953
- [100] Caunt CJ, Keyse SM. Dual-specificity MAP kinase phosphatases (MKPs): Shaping the outcome of MAP kinase signalling. The FEBS Journal. 2013;**280**(2):489-504. DOI: 10.1111/j.1742-4658.2012.08716.x
- [101] Singh K, Deshpande P, Li G, Yu M, Pryshchep S, Cavanagh M. K-RAS GTPaseand B-RAF kinase-mediated T-cell tolerance defects in rheumatoid arthritis. Proceedings of the National Academy of Sciences of the United States of America. 2012;109(25):E1629-E1637. DOI: 10.1073/pnas.1117640109
- [102] Ernst B, Lee DS, Chang JM, Sprent J, Surh CD. The peptide ligands mediating positive selection in the thymus control T cell survival and homeostatic proliferation in the periphery. Immunity. 1999;11(2):173-181

- [103] Naylor K, Li G, Vallejo AN, Lee WW, Koetz K, Bryl E, et al. The influence of age on T cell generation and TCR diversity. Journal of Immunology (Baltimore). 2005;**174**(11):7446-7452
- [104] Manoussakis MN, Tzioufas AG, Silis MP, Pange PJ, Goudevenos J, Moutsopoulos HM. High prevalence of anti-cardiolipin and other autoantibodies in a healthy elderly population. Clinical and Experimental Immunology. 1987;69(3):557-565
- [105] Waase I, Kayser C, Carlson PJ, Goronzy JJ, Weyand CM. Oligoclonal T cell proliferation in patients with rheumatoid arthritis and their unaffected siblings. Arthritis and Rheumatism. 1996;39(6):904-913
- [106] Wagner UG, Koetz K, Weyand CM, Goronzy JJ. Perturbation of the T cell repertoire in rheumatoid arthritis. Proceedings of the National Academy of Sciences of the United States of America. 1998;95(24):14447-14452
- [107] Koetz K, Bryl E, Spickschen K, O'Fallon WM, Goronzy JJ, Weyand CM. T cell homeostasis in patients with rheumatoid arthritis. Proceedings of the National Academy of Sciences of the United States of America. 2000;97(16):9203-9208
- [108] Vallejo AN, Nestel AR, Schirmer M, Weyand CM, Goronzy JJ. Aging-related deficiency of CD28 expression in CD4+ T cells is associated with the loss of gene-specific nuclear factor binding activity. The Journal of Biological Chemistry. 1998;273(14):8119-8129
- [109] Schmidt D, Goronzy JJ, Weyand CM. CD4+ CD7- CD28- T cells are expanded in rheumatoid arthritis and are characterized by autoreactivity. Journal of Clinical Investigation. 1996;97(9):2027-2037
- [110] Pawlik A, Ostanek L, Brzosko I, Brzosko M, Masiuk M, Machalinski B, et al. The expansion of CD4+CD28- T cells in patients with rheumatoid arthritis. Arthritis Research & Therapy. 2003;5(4):R210-R213. DOI: 10.1186/ar766
- [111] Park W, Weyand CM, Schmidt D, Goronzy JJ. Co-stimulatory pathways controlling activation and peripheral tolerance of human CD4+CD28- T cells. European Journal of Immunology. 1997;**27**(5):1082-1090
- [112] Weyand CM, Brandes JC, Schmidt D, Fulbright JW, Goronzy JJ. Functional properties of CD4+ CD28- T cells in the aging immune system. Mechanisms of Ageing and Development. 1998;102(2-3):131-147
- [113] Groh V, Brühl A, El-Gabalawy H, Nelson JL, Spies T. Stimulation of T cell autoreactivity by anomalous expression of NKG2D and its MIC ligands in rheumatoid arthritis. Proceedings of the National Academy of Sciences of the United States of America. 2003;100(16):9452-9457. DOI: 10.1073/pnas.1632807100
- [114] López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. Cell. 2013;**153**(6):1194-1217. DOI: 10.1016/j.cell.2013.05.039
- [115] Schönland SO, Lopez C, Widmann T, Zimmer J, Bryl E, Goronzy JJ, Weyand CM. Premature telomeric loss in rheumatoid arthritis is genetically determined and involves both myeloid and lymphoid cell lineages. Proceedings of the National Academy of Sciences of the United States of America. 2003;100(23):13471-13476. DOI: 10.1073/pnas.2233561100

- [116] Young A, Koduri G. Extra-articular manifestations and complications of rheumatoid arthritis. Best practice & research. Clinical Rheumatology. 2007;21(5):907-927. DOI: 10.1016/j.berh.2007.05.007
- [117] Prete M, Racanelli V, Digiglio L, Vacca A, Dammacco F, Perosa F. Extra-articular manifestations of rheumatoid arthritis: An update. Autoimmunity Reviews. 2011;11(2):123-131. DOI: 10.1016/j.autrev.2011.09.001
- [118] Aviña-Zubieta JA, Choi HK, Sadatsafavi M, Etminan M, Esdaile JM, Lacaille D. Risk of cardiovascular mortality in patients with rheumatoid arthritis: A meta-analysis of observational studies. Arthritis and Rheumatism. 2008;59(12):1690-1697. DOI: 10.1002/ art.24092
- [119] Manzi S, Wasko MC, Manzi S. Inflammation-mediated rheumatic diseases and atherosclerosis. Annals of the Rheumatic Diseases. 2000;59(5):321-325. DOI: 10.1136/ ard.59.5.321
- [120] Goodson NJ, Symmons DP, Scott DG, Bunn D, Lunt M, Silman AJ. Baseline levels of C-reactive protein and prediction of death from cardiovascular disease in patients with inflammatory polyarthritis: A ten-year followup study of a primary care-based inception cohort. Arthritis and Rheumatism. 2005;52(8):2293-2299. DOI: 10.1002/art.21204
- [121] Ridker PM, Rifai N, Pfeffer M, Sacks F, Lepage S, Braunwald E. Elevation of tumor necrosis factor-alpha and increased risk of recurrent coronary events after myocardial infarction. Circulation. 2000;101(18):2149-2153
- [122] Ridker PM, Rifai N, Stampfer MJ, Hennekens CH. Plasma concentration of interleukin-6 and the risk of future myocardial infarction among apparently healthy men. Circulation. 2000;**101**(15):1767-1772
- [123] Ku IA, Imboden JB, Hsue PY, Ganz P. Rheumatoid arthritis: Model of systemic inflammation driving atherosclerosis. Circulation Journal: Official Journal of the Japanese Circulation Society. 2009;73(6):977-985
- [124] Choi HK, Hernán MA, Seeger JD, Robins JM, Wolfe F. Methotrexate and mortality in patients with rheumatoid arthritis: A prospective study. Lancet. 2002;359(9313):1173-1177. DOI: 10.1016/S0140-6736(02)08213-2
- [125] Dixon WG, Watson KD, Lunt M, Hyrich KL; British Society for Rheumatology Biologics Register Control Centre Consortium, Silman AJ, et al. Reduction in the incidence of myocardial infarction in patients with rheumatoid arthritis who respond to anti-tumor necrosis factor alpha therapy: Results from the British Society for Rheumatology Biologics Register. Arthritis and Rheumatism. 2007;56(9):2905-2912. DOI: 10.1002/ art.22809
- [126] Liuzzo G, Kopecky SL, Frye RL, O'Fallon WM, Maseri A, Goronzy JJ, et al. Perturbation of the T-cell repertoire in patients with unstable angina. Circulation. 1999;100(21): 2135-2139

- [127] Liuzzo G, Biasucci LM, Trotta G, Brugaletta S, Pinnelli M, Digianuario G, et al. Unusual CD4+CD28null T lymphocytes and recurrence of acute coronary events. Journal of the American College of Cardiology. 2007;50(15):1450-1458. DOI: 10.1016/j.jacc.2007.06.040
- [128] Liuzzo G, Goronzy JJ, Yang H, Kopecky SL, Holmes DR, Frye RL. Monoclonal T-cell proliferation and plaque instability in acute coronary syndromes. Circulation. 2000;101(25):2883-2888
- [129] Gerli R, Schillaci G, Giordano A, Bocci EB, Bistoni O, Vaudo G. CD4+CD28- T lymphocytes contribute to early atherosclerotic damage in rheumatoid arthritis patients. Circulation. 2004;109(22):2744-2748. DOI: 10.1161/01.CIR.0000131450.66017.B3
- [130] Bryl E, Vallejo AN, Matteson EL, Witkowski JM, Weyand CM, Goronzy JJ. Modulation of CD28 expression with anti-tumor necrosis factor alpha therapy in rheumatoid arthritis. Arthritis and Rheumatism. 2005;**52**(10):2996-3003. DOI: 10.1002/art.21353



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