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Molecular Pathogenesis of Gastric Adenocarcinoma

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Abstract

The incidence and mortality of gastric cancer (GC) rank top five and top three, respectively, among cancers around the world. It is an intricate malignancy caused by the reciprocity of intrinsically genetic, environmental, and host-related elements. The silent property, advanced clinical characterization, and potential heterogeneity have made GC a thorny disease with a high death rate. The increasing knowledge of the abundant genetic abnormalities regarding GC will definitely elongate the patients' survival. Scientists have been working hard to discover the myths beneath gastric tumorigenesis: novel biomarkers have been established, and cell transduction cascades have been well described. The study grouping GC into four molecular subtypes by The Cancer Genome Atlas (TCGA) broadens our horizon of GC etiologies. Knowledge regarding to the sophisticated networks in tumor microenvironment also bring new insights into the mechanisms assist GC development. In the future, people will strive for translating more research achievements into clinical utility. Successful translational medicine will lead to new methods for early GC diagnosis and precise medical strategies for individuals.

Keywords: gastric adenocarcinoma, molecular classification, pathogenesis

1. Introduction

Gastric cancer (GC) is the fifth most common malignancy worldwide and third leading cancer-related death. Because of poor diagnosis, 5-year survival rate of GC is rather low, ranging from 15 to 52% [1, 2]. The incident rates are higher in men than in women. GC contributes over 20% of morbidity and mortality to cancer all over the world annually, following lung and liver cancer, which account for 23 and 28%, respectively. In 2012, approximate 723,000 deaths and more than 950,000 new GC cases were reported worldwide. Despite the incidence of GC has been declining in the North American and most of the west European countries, GC remains a type of prevailing cancer with increasing risk in regions including

Asia, Eastern Europe, and certain areas of Latin America [3]. Particularly, adenocarcinomas, which developed from the glands of the most superficial layer or the mucosa, take up 90% of GC. The rest types of GC include mucosa-associated lymphoid tissue (MALT) lymphomas, which originate from the muscles surrounding mucosa areas of the stomach [4].

The risk factors include environmental factors, such as *Helicobacter pylori* infection, cigarettes smoking and high-salt diet, and host genetic alterations. Although the incidence of GC has shown a dramatic decrease in recent years due to *H. pylori* eradication, the overall survival is still quite poor due to its silent nature, late clinical presentation, and genetic heterogeneity. Thus, comprehensive understanding of the detailed molecular mechanisms and accurate pathogenesis of GC will improve patient outcome. Recently, several kinds of molecular classification of GC have been provided to reveal the genomic landscape of GC and decipher the crucial molecular changes. Among them, The Cancer Genome Atlas (TCGA) classification is a milestone for the molecular characterization of GC. Clinical translation of these molecular findings will provide novel strategies for early GC detection and promote precision therapies for GC patients.

2. The risk factors of GC

GC is defined as a tumor with multifactorial etiologies. Environmental alteration and genetic factors play major roles in GC, particularly, virus infection, dietary habits, and lifestyles are recognized to be critical factors.

H. pylori is a common human pathogen, contributing to both malignant and benign diseases. Among all the risk factors of GC, *H. pylori* is considered to be the most predominant factor, up to 80% of GC cases are led by *H. pylori* infection. At least 660,000 new diagnoses are making annually [5, 6]. Data in western countries indicated that *H. pylori* is a major risk factor for only non-cardia GC instead of cardia GC [5]. It has been noted that the death number of GC is decreasing in many developed regions, partially related to eradication therapy and improved living condition in certain populations and regions [7]. Given the worldwide aging problem and the migration trend of people from high to low prevalence regions, the mortality is very likely to increase in the future [8]. *H. pylori* infection alone is not sufficient to trigger GC. Chiba et al. suggested *H. pylori* infection contributed to GC development via two potential mechanisms, either caused chronic inflammation or directly acted on epithelial cells [9]. In general, *H. pylori* colonizes the gastric mucosal epithelium and induces chronic inflammation at the beginning of infection, while persistent inflammation resulted in GC [10, 11]. Cytokines and chemokines produced by the tumor microenvironment facilitate cell proliferation and migration, and keep cells from apoptosis and immune detection [12]. Cytokines released by different cell sources have been described as suggestive indicators in the progression of GC. The serum level of Tumor necrosis factor- α (TNF- α) in GC was found greatly reduced, however, levels of TNF- α in stage III or IV GC patients showed a significant elevation when compared to levels in earlier stage patients [13]. Besides, the secretion of Interleukin-1 (IL-1) contributes to tumor cell proliferation and progression. In addition, IL-6, IL-10, and Transforming

growth factor- β (TGF- β) improve survival of GC cells by promoting cell invasion and suppression of antitumor immunity. IL-11, IL-17, IL-18, IL-22, and chemokines secreted by a specific cell type also improve the progression of GC [14]. Major types of virulence factors of *H. pylori* include cytotoxin VacA, the type IV secretion system (T4SS), and the CagA effector protein. These factors are associated with multiple cellular responses, such as induction of oxidative stress and epithelial barrier disruption, in various model systems [15].

Epstein-Barr virus (EBV), also termed as human herpesvirus 4 (HHV-4), is one of the most prevalent viruses in humans. EBV was the first virus identified in human carcinoma in 1964 [16]. Approximately 95% of adults in the world are infected by EBV due to the positive detection of serological EBV markers. EBV infection may not cause severe symptoms and disease. After primary infection, EBV establishes a carrier state called latent stage. However, latent EBV infection could subsequently become high-risk oncogenic factors associated with human malignancies. EBV has been known as another significant pathogen exists in GC cells. According to worldwide data, EBV-associated GC (EBVaGC) accounts for 10%, in average, of total GC cases. Most of the EBVaGC cases were epithelial tumors, while lymphoepithelioma-like carcinomas take up 90% of the rest rare EBVaGC cases. EBVaGC presents distinct clinic properties, such as predominance among male and younger individuals and predominant proximal stomach location [17]. EBV infection can be achieved by two different entry mechanisms, either via B cell entry or fusion with epithelial cells directly [18, 19]. EBVaGC occurs in the upper and middle stomach in the majority. Fukayama et al. indicated the tumor distribution in the stomach, with the proportion that 58% in the cardia, 33% in the body, and 9% in the antrum [20, 21]. They also depicted the appearance of EBVaGC as ulcerated or saucer-like, with obvious thickened gastric wall. Moreover, the lymph node was less frequently involved during early stage within the submucosa. These characteristics are proposed to be favorable prognosis indicators. Histological studies provide evidence that immune cell infiltration was a feature of EBVaGC. For instance, infiltrating lymphocytes, containing EBV-specific cytotoxic T cells, communicate with carcinoma cells directly, in the opposite, cytokine IL-1 β was upregulated to recruit noninfiltrating lymphocytes against this cell-cell contact [22, 23]. Therefore, in EBVaGC, immune responses in tumor microenvironment also accompany with the progression of EBVaGC [24].

Dietary factors: dietary risks, including salty and spicy food intake, cigarettes smoking, frequent coffee, and high-temperature drinking habits are positively associated with GC. Intriguingly, excess salt intake showed susceptibility to EBVaGC and *H. pylori*-induced GC. Habitual excess salt intake was suggested to progressively increase the risk across consumption levels of GC via a meta-analysis [25]. The association between salty food intake and *H. pylori* infection was also evaluated in a cross-sectional study of 634-middle age male cohort. The result supported habitual salt-rich Japanese food intake was prevalent in *H. pylori*-induced GC cases [26]. It is probably due to the increase of *H. pylori* colonization and persistent infection [27]. Besides salty food intake, Camargo et al. indicated that smoking was strongly associated with EBVaGC by a case-case comparison between EBV-positive tumors and EBV-negative tumors [28]. Although alcohol drinking was a suggestive risk factor of GC, heavy alcohol drinking, rather than moderate alcoholic drinks, was significantly correlated with GC development [29, 30].

3. Molecular classification and pathogenies of GC

Up to 90% of stomach malignancies are adenocarcinomas. Non-Hodgkin's lymphomas and gastrointestinal stromal tumor (GIST) make up most of the remaining 10% [31]. Even though infrequently, adenosquamous, squamous, and undifferentiated carcinomas also occur. In regard to clinical diagnosis, several pathological characterization varied from time to time. Several histological classification systems for gastric adenocarcinoma have been described, but the most frequently used are those of the World Health Organization (WHO) and Lauren [32]. In the World Health Organization (WHO) classification, there are 10 histological types [33]. The Lauren classification is commonly applied and it makes the distinction between intestinal and diffuse types. The intestinal GC consists of cohesive neoplastic cells forming gland-like structures while the diffuse type has lost cell cohesion and resulting in diffuse discohesive cellular infiltration [31]. Men and elderly are more likely to suffer intestinal type, whereas diffuse type carcinomas are relatively more common among the younger population with an equal male-to-female ratio [32]. Recently, a project named The Cancer Genome Atlas (TCGA) has proposed a brand new classification, in which GC is grouped by four subtypes: EBV-positive (EBV), microsatellite instability (MSI), genomically stable [34], and chromosomal instability [35, 36].

According to previous studies, about 9% of GC cases are infected by EBV [37]. All the EBV-positive GCs harbor the property of CpG island methylator phenotype (CIMP) [36, 38, 39]. EBV-positive tumors exhibited a higher incidence of whole-genomic DNA hypermethylation than any molecular subtypes. The genes with promoter hypermethylation showed most differentially silenced expression in EBV-associated GC [36]. Moreover, PI(3)-kinase inhibition was also strongly detected in EBV-positive GC, which offered a new method for the evaluation of this subtype [36]. The most highly transcribed EBV viral, message RNAs (mRNAs) and microRNAs (miRNAs), fell within the BamH1A region of the viral genome and showed similar expression patterns across tumors [36]. The mutation rate of PIK3CA is exclusively high in EBV-positive gastric cancer compared with other molecular subtypes. The mutation rate of PIK3CA in this subtype is about 80 and 68% of the mutations belongs to recurrent mutation in this dataset. In contrast, in other molecular subtypes, the mutation rates of PIK3CA are from 3 to 42%. So, this result provides a hint that using PI3K inhibitor might have the clinical therapeutic potential for this kind of molecular subtype.

The next subtypes of GC are abundant in MSI, which display increased mutation rates (in major histocompatibility complex class I genes, including B2M and HLA-B) and hypermethylation (containing hypermethylation at the *MLH1* promoter). The most obvious difference between EBV-CIMP (CpG island methylator phenotype) and MSI-associated gastric-CIMP methylation profiles is that all EBV-positive gastric tumors show promoter hypermethylation of *CDKN2A* (*p16^{INK4A}*), but the *MLH1* hypermethylation was only detected in MSI-associated CIMP [38].

In genomically stable subtype, *RHOA* mutation was detected [36]. When binding with Guanosine-5'-triphosphate (GTP), *RHOA* behaves through a great number of downstream effectors, such as ROCK1, mDia, and protein kinase N. This will lead to actin-myosin-dependent cell contractility and cellular motility [40, 41] and activation of STAT3 to promote carcinogenesis [42, 43]. Except from activating mDia or ROCK1, the *RHOA* mutation Y42C has been confirmed to attenuate the

activation of protein kinase N. Because RHOA is strongly associated with cell motility, the *RHOA* mutations might contribute to the invasive growth patterns. In diffuse type GC or genomically stable GC, the lack of cellular cohesion is a hallmark for this diffuse phenotype. Apart from *RHOA* mutation, an inter-chromosomal translocation called *CLDN18-ARHGAP26* fusion gene was identified. *ARHGAP26* is a GTPase-activating protein that converts GTP-RHO to GDP-RHO and it is been reported to facilitate cellular motility. *CLDN18* is a tight junction component that involves in cell adhesion. This fusion gene thus was thought to correlate with cell metastasis in this kind of molecular subtype.

With the somatic copy-number aberrations (SCNAs), the last group of GC was clustered as CIN subtype. In this subtype, a bunch of genes shows dysregulated, such as *TP53* mutations (in 71% of tumors) as well as *CDH1* somatic mutations (enriched in the genomically stable subtype, about 37% of cases).

4. The dysregulated miRNAs involved in GC

MicroRNAs (miRNAs) are one predominant category of small (roughly 20–30 nucleotides) non-coding RNAs that participate in gene expression and control [44]. Their effects are mostly lead to the degradation of message RNAs (mRNAs) or inhibitory of the translations, and subsequently affect a series of biological behaviors of cells, such as inflammation, cell proliferation, apoptosis and differentiation. In the nucleus, together with its cofactor Pasha (DGCR8), the RNase III enzyme Droscha cut out primary miRNA transcripts into a fragment of approximately 60 nucleotides named precursor miRNAs (pre-miRNAs), and initiate the biogenesis of miRNAs [45]. A cytoplasmic RNase III called Dicer will be responsible for the further processing of the pre-miRNAs and makes them mature after they are transported to the cytoplasm [46, 47]. A mature miRNA, with the length of about 18–24 nucleotides, is single-stranded, which can sometimes aim at multiple targets. These mature miRNAs always bind to the complementary sequences of targeted mRNAs directly to make mRNAs degrade or bind directly to 3'-untranslated regions (3'-UTR) of mRNAs to decrease their translation, so that miRNAs can exert their effects on regulating certain gene expression [44, 48]. Accordingly, miRNAs regulate at least 30% of genes of human as it is estimated [49].

In other words, miRNAs are capable of acting as a switch to control genes related to cell proliferation and apoptosis under pathogenic circumstances, consequently, they may have a chance to be involved in both cancer initiation and progression. It seems that no matter how clear the mechanism of malignancy behaviors or an effective therapy that might prevent tumorigenesis from the beginning, an increasing knowledge of these miRNAs is crucial. During the physiological periods, miRNAs present or absent in proper time of different stage of lives. However, they are produced abnormally in tumors, that the levels of some miRNAs are highly detected while some are lower or even none. Hence, those which are upregulated called onco-miRs, whereas the downregulated ones called tumor-suppressive miRNAs. As the names suggested, genes controlled by onco-miRs are oncogenes whose products may promote tumor cells in many aspects, whereas the opposite site of genes is tumor suppressive, which plays a role of inhibitor among the initiation or development of tumors (e.g., miR-15a

and miR-16-1, which target a member of Hippo pathway called YAP1, are downregulated in GC) [50]. Thus, identifying the target genes of these miRNAs is crucial and it may lead us to a better understanding of the miRNAs themselves.

For example, miR-21 was the first miRNA which was influenced by *H. pylori*. In tissues of both GC and *H. Pylori* infection, it was highly detected [51, 52]. Several data have reported that this miRNA can be used as a biomarker in GC diagnosis in the clinic [53–55]. And most recently, a research conducted in China suggested this miR-21 to be a GC biomarker in both diagnosis and prognosis, for the reason that, besides the high levels found in tumor tissues compared with the normal ones, miR-21 was revealed to be associated with poor survivals in clinical patients [56]. Behind the phenomena, the molecular mechanism is still unclear in GC. In other types of cancer, however, such as colorectal cancer, miR-21 decreased the tumor suppressor protein programmed cell death 4 (PDCD4) and exhibited an oncogenic function [57]. Additionally, in nonsmall cell lung carcinoma, miR-21 was found to deregulate PTEN, a tumor suppressor, to promote carcinogenesis [58].

5. The tumor microenvironment and gastric carcinogenesis

More and more evidence supported the idea that not only malignant GC cells, but also those nonmalignant cells involved in the tumor microenvironment play indispensable roles throughout GC pathogenesis. Generally, nonmalignant cells participate in various mechanisms related to GC development, such as stromal interactions, angiogenesis, and some immune responses.

Stromal components, including fibroblasts and extracellular matrix (ECM) adjacent to cancer cells, create a suitable environment for GC development. Stromal fibroblasts are known to play a central role in tumor microenvironment by interacting with cancer cells [59, 60]. However, other than ordinary fibroblasts, cancer-associated fibroblasts (CAFs) undergo a phenotypic change into myofibroblasts. Also, CAFs exhibit distinct gene expression patterns that pertain to aberrant cell growth, focal adhesion, and ECM remodeling [61]. The remodeling ECM promotes the survival ability and distant colonization cancer cells by synthesizing components, such as type I and type III collagens, fibronectins, tenascin, and versican [62–64], as well as proteases like urokinase, plasminogen activator, fibroblast activation protein (FAP), and matrix metalloproteinases (MMPs) [65–67]. These characteristics are usually associated with poor prognostics. Moreover, tumor-related stromal fibroblasts secrete small molecules, such as IL-6 to stimulate cell growth, as well as factors associated with TGF- β signaling, triggering epithelial-mesenchymal transition (EMT) [68–70].

Angiogenesis is also a pivotal process contributes to tumor progression. Cell population in GC tumor microenvironment regulates the density and architecture of new blood vessels by stimulating the proliferation and differentiation of myofibroblasts and vascular endothelial cells [71, 72]. The growing number of new vessels was reported to facilitate GC metastasis [73, 74]. To promote angiogenesis, GC cells provide numerous angiogenic factors, including VEGF, FGF-2, CXCL1, and Ang-2, to microenvironment [75–78]. It has been noted that high level of VEGF-A contributed to endothelium-dependent angiogenesis. VEGF-A signaling increased both the new blood vessel number and permeability in GC [79]. VEGF-A strongly

promotes the angiogenesis and aggressive phenotype of human intestinal-type GC [80]. Bevacizumab is a specific antibody against VEGF-A, and dominantly regulates normal and pathological angiogenesis. Clinical trials of bevacizumab in phase II advanced GC suggested a satisfied curative effect [81]. Unfortunately, in the randomized phase III Avastin in Gastric Cancer (AVAGAST) study, the combination of chemotherapy and bevacizumab did not show a better overall survival extension in the first-line treatment when compared to advanced GC patients, who only subjected chemotherapy [82]. Intriguingly, REGARD and RAINBOW trials using VEGFR2 targeting antibody ramucirumab have also shown a significant increase in the overall survival of patients with advanced GC [83]. It can be partially explained by the geographical differences of GC patients.

Immune reactivity in GC development is based on various types of immune cells in different stages [84]. First, in eliminating stage, macrophages and dendritic cells recruit to secrete chemokines and cytokines, such as IL-12 and IFN- γ , to phagocytize and remove apoptotic cancer cells [85]. Cancer cells survived from elimination subsequently under equilibrium stage and adapted to immune-suppression. The number of tumor-infiltrating lymphocytes (TILs) was noted as independent predictors to evaluate lymph node metastasis and GC patient survival in this stage. Interestingly, TILs exert either oncogenic or tumor-suppressive influence in GC cells, attributing to their unique functions. In a recent report, tumor-associated macrophages with high levels of CD163 expression exhibited aggressive characteristics and expression of cancer stem cell markers in recurrent GC cases. CD163⁺ macrophages might, therefore, related to independent worse prognosis [86]. T-lymphocyte subsets are also significant cell types associated with both early onset of tumor and tumor progression. It has been reported that the high ratio of CD8⁺ cytotoxic TILs/FOXP3⁺ regulatory T cells (Treg) correlated with high overall survival rate and good prognosis, especially in MSI gastric cancer [87, 88]. Eventually, cancer cells escape from immune responses.

Immunogenicity could be blunted by releasing cytokines including TGF- β , TNF- α , and IL-10. Therefore, cancer cells can evade the detection by immune effectors in the tumor microenvironment [89]. PD-1 is one of the most important receptors expressed by T cells and monocytes. PD-1 negatively regulates the effectors by binding to its ligands PD-L1 and PD-L2. Blocking PD-1/PD-L1 has been suggested as a promising immune-therapeutic option [90, 91]. The relationship between PD-L1 level and prognosis remains under debate, however, a recent meta-analysis pooled all present data and suggested PD-L1 to be a valuable prognostic indicator of GC patients by dividing patients into different groups [92].

Collectively, tumor microenvironment composed of multiple subjects, such as tumor stromal fibroblasts, lymphocytes, and angiogenic factors. All these components cooperate together, configuring a context for GC development.

6. Conclusion

There is still a large realm of the unknown area about GC pathogenies even though the related research studies have been going deeper and our horizon has been broadened in the past few years. Epigenetically, we are seeking to unravel the mechanisms of gastric tumorigenesis

affected by chromatin remodeling. Besides, there are miRNAs, lncRNAs, and some other classic molecules that are involved in GC development, which requires a more detailed investigation regarding these aspects. The most crucially and urgently, we are struggling to explore specific and precise therapies for patients who are suffering or likely to suffer from GC, especially genomically stable GC (a type of GC with a high mortality).

When it comes to the novel small entities, there are even more issues waiting to be addressed, such as how lncRNAs interact with chromatin alterations during gastric carcinogenesis. Moreover, in addition to *H. pylori* and EBV, scientists have identified a wild variety of bacteria in the stomach through DNA sequencing. These uncultivable bacteria have formed a complicated community and there are few clues about how they interact with *H. pylori* and host immunity during GC initiation and progression. In the early detection field, molecular findings may facilitate new approaches for diagnosing GC early, by identifying high-risk potential sufferers via the molecular features of precancerous lesions. Finally, therapeutic strategies need to be designed precisely for each GC subtypes, according to their somatic driver change of genome or tumor-associated cell compartments, such as stromal cells and tumor-infiltrating immune cells.

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References

- [1] Marchet A, Mocellin S, Ambrosi A, Morgagni P, Vitimberga G, Roviello F, Marrelli D, de Manzoni G, Minicozzi A, Coniglio A, Tiberio G, Pacelli F, Rosa F, Nitti D. Validation of the new AJCC TNM staging system for gastric cancer in a large cohort of patients (n = 2,155): Focus on the T category. *European Journal of Surgical Oncology*. 2011;**37**:779-785
- [2] Nashimoto A, Akazawa K, Isobe Y, Miyashiro I, Katai H, Kodera Y, Tsujitani S, Seto Y, Furukawa H, Oda I, Ono H, Tanabe S, Kaminishi M. Gastric cancer treated in 2002 in Japan: 2009 annual report of the JGCA nationwide registry. *Gastric Cancer*. 2013;**16**:1-27

- [3] International Agency for Research on Cancer. GLOBOCAN Database. IARC; 2008
- [4] Karimi P, Islami F, Anandasabapathy S, Freedman ND, Kamangar F. Gastric cancer: Descriptive epidemiology, risk factors, screening, and prevention. *Cancer Epidemiology Biomarkers & Prevention*. 2014;**23**:700-713
- [5] Helicobacter and Cancer Collaborative Group. Gastric cancer and *Helicobacter pylori*: a combined analysis of 12 case control studies nested within prospective cohorts. *Gut*. 2001;**49**:347-353
- [6] de Martel C, Ferlay J, Franceschi S, Vignat J, Bray F, Forman D, Plummer M. Global burden of cancers attributable to infections in 2008: A review and synthetic analysis. *Lancet Oncology*. 2012;**13**:607-615
- [7] O'Connor A, O'Morain CA, Ford AC. Population screening and treatment of *Helicobacter pylori* infection. *Nature Reviews Gastroenterology & Hepatology*. 2017;**14**:230-240
- [8] Yeh JM, Goldie SJ, Kuntz KM, Ezzati M. Effects of *Helicobacter pylori* infection and smoking on gastric cancer incidence in China: A population-level analysis of trends and projections. *Cancer Causes Control*. 2009;**20**:2021-2029
- [9] Chiba T, Marusawa H, Seno H, Watanabe N. Mechanism for gastric cancer development by *Helicobacter pylori* infection. *Journal of Gastroenterology and Hepatology*. 2008;**23**:1175-1181
- [10] Correa P. Human gastric carcinogenesis: A multistep and multifactorial process—First American Cancer Society Award Lecture on Cancer Epidemiology and Prevention. *Cancer Research*. 1992;**52**:6735-6740
- [11] Sheibani S, Mahmoudian RA, Abbaszadegan MR, Chamani J, Memar B, Gholamin M. Expression analysis of matrix metalloproteinase-13 in human gastric cancer in the presence of *Helicobacter Pylori* infection. *Cancer Biomarkers*. 2017;**18**(4):349-356
- [12] Hanahan D, Weinberg RA. Hallmarks of cancer: The next generation. *Cell*. 2011;**144**:646-674
- [13] Wu CW, Chi CW, Hsieh MC, Chao MF, Lui WY, P'Eng FK. Serum tumor necrosis factor in patients with gastric cancer. *Anticancer Research*. 1998;**18**:1597-1599
- [14] Bagheri V, Memar B, Momtazi AA, Sahebkar A, Gholamin M, Abbaszadegan MR. Cytokine networks and their association with *Helicobacter pylori* infection in gastric carcinoma. *Journal of Cellular Physiology*. 2017;**9999**:1-13
- [15] Naumann M, Sokolova O, Tegtmeyer N, Backert S. *Helicobacter pylori*: A paradigm pathogen for subverting host cell signal transmission. *Trends in Microbiology*. 2017;**25**:316-328
- [16] Epstein MA, Achong BG, Barr YM. Virus particles in cultured lymphoblasts from Burkitt's lymphoma. *Lancet*. 1964;**1**:702-703
- [17] Fukayama M, Hino R, Uozaki H. Epstein-Barr virus and gastric carcinoma: Virus-host interactions leading to carcinoma. *Cancer Science*. 2008;**99**:1726-1733

- [18] Tsang CM, Deng W, Yip YL, Zeng MS, Lo KW, Tsao SW. Epstein-Barr virus infection and persistence in nasopharyngeal epithelial cells. *Chinese Journal of Cancer*. 2014;**33**:549-555
- [19] Tsai MH, Raykova A, Klinke O, Bernhardt K, Gartner K, Leung CS, Geletneky K, Sertel S, Munz C, Feederle R, Delecluse HJ. Spontaneous lytic replication and epitheliotropism define an Epstein-Barr virus strain found in carcinomas. *Cell Reports*. 2013;**5**:458-470
- [20] Abe H, Maeda D, Hino R, Otake Y, Isogai M, Ushiku AS, Matsusaka K, Kunita A, Ushiku T, Uozaki H, Tateishi Y, Hishima T, Iwasaki Y, Ishikawa S, Fukayama M. ARID1A expression loss in gastric cancer: Pathway-dependent roles with and without Epstein-Barr virus infection and microsatellite instability. *Virchows Archiv*. 2012;**461**:367-377
- [21] Abe H, Kaneda A, Fukayama M. Epstein-Barr Virus-Associated gastric carcinoma: Use of host cell machineries and somatic gene mutations. *Pathobiology*. 2015;**82**:212-223
- [22] Chong JM, Sakuma K, Sudo M, Osawa T, Ohara E, Uozaki H, Shibahara J, Kuroiwa K, Tominaga S, Hippo Y, Aburatani H, Funata N, Fukayama M. Interleukin-1beta expression in human gastric carcinoma with Epstein-Barr virus infection. *Journal of Virology*. 2002;**76**:6825-6831
- [23] Fukayama M. Epstein-Barr virus and gastric carcinoma. *Pathology International*. 2010;**60**:337-350
- [24] Cho J, Kang MS, Kim KM. Epstein-Barr Virus-Associated gastric carcinoma and specific features of the accompanying immune response. *Journal of Gastric Cancer*. 2016;**16**:1-7
- [25] D'Elia L, Rossi G, Ippolito R, Cappuccio FP, Strazzullo P. Habitual salt intake and risk of gastric cancer: A meta-analysis of prospective studies. *Clinical Nutrition*. 2012;**31**:489-498
- [26] Tsugane S, Tei Y, Takahashi T, Watanabe S, Sugano K. Salty food intake and risk of *Helicobacter pylori* infection. *Japanese Journal of Cancer Research*. 1994;**85**:474-478
- [27] Fox JG, Dangler CA, Taylor NS, King A, Koh TJ, Wang TC. High-salt diet induces gastric epithelial hyperplasia and parietal cell loss, and enhances *Helicobacter pylori* colonization in C57BL/6 mice. *Cancer Research*. 1999;**59**:4823-4828
- [28] Camargo MC, Koriyama C, Matsuo K, Kim WH, Herrera-Goepfert R, Liao LM, Yu J, Carrasquilla G, Sung JJ, Alvarado-Cabrero I, Lissowska J, Meneses-Gonzalez F, Yatabe Y, Ding T, Hu N, Taylor PR, Morgan DR, Gulley ML, Torres J, Akiba S, Rabkin CS. Case-case comparison of smoking and alcohol risk associations with Epstein-Barr virus-positive gastric cancer. *International Journal of Cancer*. 2014;**134**:948-953
- [29] Tramacere I, Negri E, Pelucchi C, Bagnardi V, Rota M, Scotti L, Islami F, Corrao G, La Vecchia C, Boffetta P. A meta-analysis on alcohol drinking and gastric cancer risk. *Annals of Oncology*. 2012;**23**:28-36
- [30] Everatt R, Tamosiunas A, Kuzmickiene I, Virviciute D, Radisauskas R, Reklaitiene R, Milinaviciene E. Alcohol consumption and risk of gastric cancer: A cohort study of men in Kaunas, Lithuania, with up to 30 years follow-up. *BMC Cancer*. 2012;**12**:475

- [31] Kelley JR, Duggan JM. Gastric cancer epidemiology and risk factors. *Journal of Clinical Epidemiology*. 2003;**56**:1-9
- [32] Lauren P. The two histological main types of gastric carcinoma: Diffuse and So-Called Intestinal-Type carcinoma. An attempt at a Histo-Clinical classification. *Acta Pathologica et Microbiologica Scandinavica*. 1965;**64**:31-49
- [33] Brunicki FC, Schwartz SI. *Schwartz's Principles of Surgery*. 8th ed. New York: McGraw-Hill, Medical Pub. Division; 2005
- [34] Nguyen LH, Robinton DA, Seligson MT, Wu L, Li L, Rakheja D, Comerford SA, Ramezani S, Sun X, Parikh MS, Yang EH, Powers JT, Shinoda G, Shah SP, Hammer RE, Daley GQ, Zhu H. Lin28b is sufficient to drive liver cancer and necessary for its maintenance in murine models. *Cancer Cell*. 2014;**26**:248-261
- [35] Finkelstein GP, Forcinito P, Lui JC, Barnes KM, Marino R, Makaroun S, Nguyen V, Lazarus JE, Nilsson O, Baron J. An extensive genetic program occurring during postnatal growth in multiple tissues. *Endocrinology*. 2009;**150**:1791-1800
- [36] The Cancer Genome Atlas Research Network. Comprehensive molecular characterization of gastric adenocarcinoma. *Nature*. 2014;**513**:202-209
- [37] Murphy G, Pfeiffer R, Camargo MC, Rabkin CS. Meta-analysis shows that prevalence of Epstein-Barr virus-positive gastric cancer differs based on sex and anatomic location. *Gastroenterology*. 2009;**137**:824-833
- [38] Matsusaka K, Kaneda A, Nagae G, Ushiku T, Kikuchi Y, Hino R, Uozaki H, Seto Y, Takada K, Aburatani H, Fukayama M. Classification of Epstein-Barr virus-positive gastric cancers by definition of DNA methylation epigenotypes. *Cancer Research*. 2011;**71**:7187-7197
- [39] Toyota M, Ahuja N, Suzuki H, Itoh F, Ohe-Toyota M, Imai K, Baylin SB, Issa JP. Aberrant methylation in gastric cancer associated with the CpG island methylator phenotype. *Cancer Research*. 1999;**59**:5438-5442
- [40] Ridley AJ, Schwartz MA, Burridge K, Firtel RA, Ginsberg MH, Borisy G, Parsons JT, Horwitz AR. Cell migration: Integrating signals from front to back. *Science*. 2003;**302**:1704-1709
- [41] Thumkeo D, Watanabe S, Narumiya S. Physiological roles of Rho and Rho effectors in mammals. *European Journal of Cell Biology*. 2013;**92**:303-315
- [42] Aznar S, Valeron PF, del Rincon SV, Perez LF, Perona R, Lacal JC. Simultaneous tyrosine and serine phosphorylation of STAT3 transcription factor is involved in Rho A GTPase oncogenic transformation. *Molecular Biology of the Cell*. 2001;**12**:3282-3294
- [43] Yu H, Jove R. The STATs of cancer—New molecular targets come of age. *Nature Reviews Cancer*. 2004;**4**:97-105

- [44] Carthew RW, Sontheimer EJ. Origins and mechanisms of miRNAs and siRNAs. *Cell*. 2009;**136**:642-655
- [45] Lee Y, Han J, Yeom KH, Jin H, Kim VN. Drosha in primary microRNA processing. *Cold Spring Harbor Symposia on Quantitative Biology*. 2006;**71**:51-57
- [46] Han J, Lee Y, Yeom KH, Kim YK, Jin H, Kim VN. The Drosha-DGCR8 complex in primary microRNA processing. *Genes & Development*. 2004;**18**:3016-3027
- [47] Lund E, Guttinger S, Calado A, Dahlberg JE, Kutay U. Nuclear export of microRNA precursors. *Science*. 2004;**303**:95-98
- [48] Bartel DP. MicroRNAs: Genomics, biogenesis, mechanism, and function. *Cell*. 2004;**116**:281-297
- [49] Miranda KC, Huynh T, Tay Y, Ang YS, Tam WL, Thomson AM, Lim B, Rigoutsos I. A pattern-based method for the identification of MicroRNA binding sites and their corresponding heteroduplexes. *Cell*. 2006;**126**:1203-1217
- [50] Kang W, Tong JH, Lung RW, Dong Y, Zhao J, Liang Q, Zhang L, Pan Y, Yang W, Pang JC, Cheng AS, Yu J, To KF. Targeting of YAP1 by microRNA-15a and microRNA-16-1 exerts tumor suppressor function in gastric adenocarcinoma. *Molecular Cancer*. 2015;**14**:52
- [51] Volinia S, Calin GA, Liu CG, Ambs S, Cimmino A, Petrocca F, Visone R, Iorio M, Roldo C, Ferracin M, Prueitt RL, Yanaihara N, Lanza G, Scarpa A, Vecchione A, Negrini M, Harris CC, Croce CM. A microRNA expression signature of human solid tumors defines cancer gene targets. *Proceedings of the National Academy of Sciences of the United States of America*. 2006;**103**:2257-2261
- [52] Zhang Z, Li Z, Gao C, Chen P, Chen J, Liu W, Xiao S, Lu H. miR-21 plays a pivotal role in gastric cancer pathogenesis and progression. *Laboratory Investigation; A Journal of Technical Methods and Pathology*. 2008;**88**:1358-1366
- [53] Shiotani A, Murao T, Kimura Y, Matsumoto H, Kamada T, Kusunoki H, Inoue K, Uedo N, Iishi H, Haruma K. Identification of serum miRNAs as novel non-invasive biomarkers for detection of high risk for early gastric cancer. *British Journal of Cancer*. 2013;**109**:2323-2330
- [54] Li BS, Zhao YL, Guo G, Li W, Zhu ED, Luo X, Mao XH, Zou QM, Yu PW, Zuo QF, Li N, Tang B, Liu KY, Xiao B. Plasma microRNAs, miR-223, miR-21 and miR-218, as novel potential biomarkers for gastric cancer detection. *Plos One*. 2012;**7**:e41629
- [55] Cui L, Zhang X, Ye G, Zheng T, Song H, Deng H, Xiao B, Xia T, Yu X, Le Y, Guo J. Gastric juice MicroRNAs as potential biomarkers for the screening of gastric cancer. *Cancer*. 2013;**119**:1618-1626
- [56] Wang D, Fan Z, Liu F, Zuo J. Hsa-miR-21 and Hsa-miR-29 in tissue as potential diagnostic and prognostic biomarkers for gastric cancer. *Cellular Physiology and Biochemistry: International Journal of Experimental Cellular Physiology, Biochemistry, and Pharmacology*. 2015;**37**:1454-1462

- [57] Asangani IA, Rasheed SA, Nikolova DA, Leupold JH, Colburn NH, Post S, Allgayer H. MicroRNA-21 (miR-21) post-transcriptionally downregulates tumor suppressor Pcd4 and stimulates invasion, intravasation and metastasis in colorectal cancer. *Oncogene*. 2008;**27**:2128-2136
- [58] Zhang JG, Wang JJ, Zhao F, Liu Q, Jiang K, Yang GH. MicroRNA-21 (miR-21) represses tumor suppressor PTEN and promotes growth and invasion in non-small cell lung cancer (NSCLC). *Clinica Chimica Acta; International Journal of Clinical Chemistry*. 2010;**411**:846-852
- [59] Orimo A, Gupta PB, Sgroi DC, Arenzana-Seisdedos F, Delaunay T, Naeem R, Carey VJ, Richardson AL, Weinberg RA. Stromal fibroblasts present in invasive human breast carcinomas promote tumor growth and angiogenesis through elevated SDF-1/CXCL12 secretion. *Cell*. 2005;**121**:335-348
- [60] Pietras K, Ostman A. Hallmarks of cancer: Interactions with the tumor stroma. *Experimental Cell Research*. 2010;**316**:1324-1331
- [61] Bhowmick NA, Neilson EG, Moses HL. Stromal fibroblasts in cancer initiation and progression. *Nature*. 2004;**432**:332-337
- [62] Lagace R, Grimaud JA, Schurch W, Seemayer TA. Myofibroblastic stromal reaction in carcinoma of the breast: Variations of collagenous matrix and structural glycoproteins. *Virchows Archiv A Pathological Anatomy and Histology*. 1985;**408**:49-59
- [63] Brown LF, Guidi AJ, Schnitt SJ, Van De Water L, Iruela-Arispe ML, Yeo TK, Tognazzi K, Dvorak HF. Vascular stroma formation in carcinoma in situ, invasive carcinoma, and metastatic carcinoma of the breast. *Clinical Cancer Research*. 1999;**5**:1041-1056
- [64] Hanamura N, Yoshida T, Matsumoto E, Kawarada Y, Sakakura T. Expression of fibronectin and tenascin-C mRNA by myofibroblasts, vascular cells and epithelial cells in human colon adenomas and carcinomas. *International Journal of Cancer*. 1997;**73**:10-15
- [65] Park JE, Lenter MC, Zimmermann RN, Garin-Chesa P, Old LJ, Rettig WJ. Fibroblast activation protein, a dual specificity serine protease expressed in reactive human tumor stromal fibroblasts. *Journal of Biological Chemistry*. 1999;**274**:36505-36512
- [66] DeClerck YA. Interactions between tumour cells and stromal cells and proteolytic modification of the extracellular matrix by metalloproteinases in cancer. *European Journal of Cancer*. 2000;**36**:1258-1268
- [67] Huang WB, Zhou XJ, Chen JY, Zhang LH, Meng K, Ma HH, Lu ZF. CD10-positive stromal cells in gastric carcinoma: Correlation with invasion and metastasis. *Japanese Journal of Clinical Oncology*. 2005;**35**:245-250
- [68] Wu X, Tao P, Zhou Q, Li J, Yu Z, Wang X, Li C, Yan M, Zhu Z, Liu B, Su L. IL-6 secreted by cancer-associated fibroblasts promotes epithelial-mesenchymal transition and metastasis of gastric cancer via JAK2/STAT3 signaling pathway. *Oncotarget*. 2017;**8**:20741-20750

- [69] Landskron G, De la Fuente M, Thuwajit P, Thuwajit C, Hermoso MA. Chronic inflammation and cytokines in the tumor microenvironment. *Journal of Immunology Research*. 2014;**2014**:149185
- [70] Huang L, Wu RL, Xu AM. Epithelial-mesenchymal transition in gastric cancer. *American Journal of Translational Research*. 2015;**7**:2141-2158
- [71] Direkze NC, Hodivala-Dilke K, Jeffery R, Hunt T, Poulsom R, Oukrif D, Alison MR, Wright NA. Bone marrow contribution to tumor-associated myofibroblasts and fibroblasts. *Cancer Research*. 2004;**64**:8492-8495
- [72] Matsumoto T, Kuroda R, Mifune Y, Kawamoto A, Shoji T, Miwa M, Asahara T, Kurosaka M. Circulating endothelial/skeletal progenitor cells for bone regeneration and healing. *Bone*. 2008;**43**:434-439
- [73] Takahashi Y, Cleary KR, Mai M, Kitadai Y, Bucana CD, Ellis LM. Significance of vessel count and vascular endothelial growth factor and its receptor (KDR) in intestinal-type gastric cancer. *Clinical Cancer Research*. 1996;**2**:1679-1684
- [74] Chung HW, Lim JB. Role of the tumor microenvironment in the pathogenesis of gastric carcinoma. *World Journal of Gastroenterology*. 2014;**20**:1667-1680
- [75] Maeda K, Chung YS, Ogawa Y, Takatsuka S, Kang SM, Ogawa M, Sawada T, Sowa M. Prognostic value of vascular endothelial growth factor expression in gastric carcinoma. *Cancer*. 1996;**77**:858-863
- [76] Tanimoto H, Yoshida K, Yokozaki H, Yasui W, Nakayama H, Ito H, Ohama K, Tahara E. Expression of basic fibroblast growth factor in human gastric carcinomas. *Virchows Archiv. B, Cell Pathology Including Molecular Pathology*. 1991;**61**:263-267
- [77] Kasashima H, Yashiro M, Nakamae H, Kitayama K, Masuda G, Kinoshita H, Fukuoka T, Hasegawa T, Nakane T, Hino M, Hirakawa K, Ohira M. CXCL1-Chemokine (C-X-C Motif) receptor 2 signaling stimulates the recruitment of bone Marrow-Derived mesenchymal cells into Diffuse-Type gastric cancer stroma. *American Journal of Pathology*. 2016;**186**:3028-3039
- [78] Huang MM, Guo AB, Sun JF, Chen XL, Yin ZY. Angiotensin II promotes the progression of human gastric cancer. *Molecular Medicine Reports*. 2014;**9**:1056-1060
- [79] Wang L, Zhou R, Zhao Y, Dong S, Zhang J, Luo Y, Huang N, Shi M, Bin J, Liao Y, Liao W. MACC-1 promotes Endothelium-Dependent angiogenesis in gastric cancer by activating TWIST1/VEGF-A signal pathway. *PLoS One*. 2016;**11**:e0157137
- [80] Yancopoulos GD, Davis S, Gale NW, Rudge JS, Wiegand SJ, Holash J. Vascular-specific growth factors and blood vessel formation. *Nature*. 2000;**407**:242-248
- [81] Shah MA, Ramanathan RK, Ilson DH, Levnor A, D'Adamo D, O'Reilly E, Tse A, Trocola R, Schwartz L, Capanu M, Schwartz GK, Kelsen DP. Multicenter phase II study of irinotecan, cisplatin, and bevacizumab in patients with metastatic gastric or gastroesophageal junction adenocarcinoma. *Journal of Clinical Oncology*. 2006;**24**:5201-5206

- [82] Ohtsu A, Shah MA, Van Cutsem E, Rha SY, Sawaki A, Park SR, Lim HY, Yamada Y, Wu J, Langer B, Starnawski M, Kang YK. Bevacizumab in combination with chemotherapy as first-line therapy in advanced gastric cancer: A randomized, double-blind, placebo-controlled phase III study. *Journal of Clinical Oncology*. 2011;**29**:3968-3976
- [83] Shan F, Miao R, Xue K, Li Z, Bu Z, Wu A, Zhang L, Wu X, Zong X, Wang X, Li S, Ji X, Jia Z, Ji J. Controlling angiogenesis in gastric cancer: A systematic review of anti-angiogenic trials. *Cancer Letters*. 2016;**380**:598-607
- [84] Blankenstein T, Coulie PG, Gilboa E, Jaffee EM. The determinants of tumour immunogenicity. *Nature Reviews Cancer*. 2012;**12**:307-313
- [85] Subhash VV, Yeo MS, Tan WL, Yong WP. Strategies and advancements in harnessing the immune system for gastric cancer immunotherapy. *Journal of Immunology Research*. 2015;**2015**:308574
- [86] Zhang WJ, Zhou ZH, Guo M, Yang LQ, Xu YY, Pang TH, Gao ST, Xu XY, Sun Q, Feng M, Wang H, Lu CL, Wu GZ, Guan WX, Xu GF. High infiltration of polarized CD163+ tumor-associated macrophages correlates with aberrant expressions of CSCs markers, and predicts prognosis in patients with recurrent gastric cancer. *Journal of Cancer*. 2017;**8**:363-370
- [87] Kim KJ, Lee KS, Cho HJ, Kim YH, Yang HK, Kim WH, Kang GH. Prognostic implications of tumor-infiltrating FoxP3+ regulatory T cells and CD8+ cytotoxic T cells in microsatellite-unstable gastric cancers. *Human Pathology*. 2014;**45**:285-293
- [88] Liu K, Yang K, Wu B, Chen H, Chen X, Jiang L, Ye F, He D, Lu Z, Xue L, Zhang W, Li Q, Zhou Z, Mo X, Hu J. Tumor-Infiltrating immune cells are associated with prognosis of gastric cancer. *Medicine (Baltimore)*. 2015;**94**:e1631
- [89] Mittal D, Gubin MM, Schreiber RD, Smyth MJ. New insights into cancer immunoediting and its three component phases—Elimination, equilibrium and escape. *Current Opinion in Immunology*. 2014;**27**:16-25
- [90] Tumeh PC, Harview CL, Yearley JH, Shintaku IP, Taylor EJ, Robert L, Chmielowski B, Spasic M, Henry G, Ciobanu V, West AN, Carmona M, Kivork C, Seja E, Cherry G, Gutierrez AJ, Grogan TR, Mateus C, Tomasic G, Glaspy JA, Emerson RO, Robins H, Pierce RH, Elashoff DA, Robert C, Ribas A. PD-1 blockade induces responses by inhibiting adaptive immune resistance. *Nature*. 2014;**515**:568-571
- [91] Herbst RS, Soria JC, Kowanzet M, Fine GD, Hamid O, Gordon MS, Sosman JA, McDermott DF, Powderly JD, Gettinger SN, Kohrt HE, Horn L, Lawrence DP, Rost S, Leabman M, Xiao Y, Mokatrin A, Koeppen H, Hegde PS, Mellman I, Chen DS, Hodi FS. Predictive correlates of response to the anti-PD-L1 antibody MPDL3280A in cancer patients. *Nature*. 2014;**515**:563-567
- [92] Zhang M, Dong Y, Liu H, Wang Y, Zhao S, Xuan Q, Zhang Q. The clinicopathological and prognostic significance of PD-L1 expression in gastric cancer: A meta-analysis of 10 studies with 1,901 patients. *Scientific Reports*. 2016;**6**:37933

