

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

4,800

Open access books available

122,000

International authors and editors

135M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Toxoplasma gondii and Schizophrenia: A Relationship That Is Not Ruled Out

Antonio Sorlozano-Puerto and
Jose Gutierrez-Fernandez

Additional information is available at the end of the chapter

<http://dx.doi.org/10.5772/66018>

Abstract

Over recent years, it has been proposed that some diseases of unknown origin, such as schizophrenia, may be caused by persistent chronic infections coupled with a genetic component and may be perpetuated by the immune system. This hypothesis is supported by epidemiological and biological evidence on the exposure of schizophrenics to infectious diseases during prenatal or postnatal periods, including *Toxoplasma gondii*, chlamydia, human herpes virus, human endogenous retroviruses, parvovirus B19, mumps, and flu viruses. This growing list of microbes will undoubtedly continue to increase in the future. Linking infection to schizophrenia is a complex challenge that requires further experimental and epidemiological research. *T. gondii* is the infectious agent that has most frequently been related to neuropsychiatric disorders, including schizophrenia, and it is considered to represent a highly useful model to analyze the influence of a microorganism on human behavior and the development of psychiatric disease. It may also help to detect patient subpopulations susceptible to treatment with specific antimicrobials by improving definition of the differential phenotype of the disease, and it offers the possibility of a preventive approach.

Keywords: schizophrenia, *Toxoplasma gondii*, antibodies, behavior, cytokine, neurotransmitter, gene-infection interaction

1. Introduction

Over the past few years, it has been proposed by some authors that schizophrenia may be caused by central nervous system (CNS) disorders during neurodevelopment (i.e., congenital) or during the postnatal period, at least in some patient subgroups [1]. These disorders may be related to environmental exposure to toxic products, radiation, stress, fetal hypoxia,

nutritional problems, infections (especially when chronic and persistent), and/or, according to more recent data, gut microbiota [2, 3]. Any of these exposures could possibly affect cognitive functions and behavior patterns with important neuropsychiatric consequences, including irreversible neurological lesions leading to neuronal dysfunction, behavior problems, mental retardation, learning difficulties, or mood disorders [4–9]. Participation by microbial agents in the development of schizophrenia is suggested by medical evidence, with prenatal or perinatal infection being the most frequent cause of severe congenital malformations and mental impairment [10]. Their involvement is also supported by epidemiological evidence on the exposure of schizophrenic patients to *T. gondii*, chlamydia, human herpes virus, human endogenous retroviruses, parvovirus B19, and rubella, mumps, or influenza viruses, among other microorganisms [11].

According to current pathogenic models, microorganisms may produce various inflammatory and/or immunological disorders in the infected brain, giving rise to neurotransmitter synthesis disorders with important clinical repercussions [7]. Schizophrenia has been related to the production of inflammatory cytokines that alter the synthesis of dopamine and other neurotransmitters [12] and to fetal neuronal tissue damage due to the transplacental transfer of maternal antibodies, which might underlie development of the disease decades later [13]. This association with inflammatory and immunologic disorders has been observed in studies of animal models and human cells. Thus, maternal infection of mice and rats during pregnancy was associated with behavioral disorders in the offspring that were very similar to those reported in schizophrenic patients. Various studies in murine models revealed an association between prenatal infection and marked deficits in sensory information processing, in the expression of certain neurotransmitters (e.g., dopamine) and of cytokines, and in the immune function, all of which emerged in the offspring. Their onset is at an age equivalent to human adolescence and is earlier, with more severe effects, in male *versus* female rats, and these alterations can be reverted by the administration of antipsychotic drugs. In short, the fetus can be damaged by numerous infectious agents, whether or not they are primarily neurotropic, which may favor in a direct or immune-mediated manner the development of neurological damage, disorders in neurotransmitter expression, and modifications in sensory information processing [14].

There is intense and increasing research interest in the relationship between schizophrenia and infectious agents. Irreversible mild or severe neurophysiologic alterations may result from fetal infection, maternal infection with secondary fetal involvement *via* inflammatory and/or immunological mechanisms, or postnatal infection and may lead to the emergence of schizophrenia over the years. The full elucidation of these associations may allow specific antimicrobial treatments to be added to current symptomatic (or antipsychotic) treatments for these patients [5], potentially offering a preventive and curative approach to the disease, given that they would act on known and treatable etiologic factors.

T. gondii is the infectious agent that has most frequently been related to neuropsychiatric disorders, including schizophrenia, and it is considered to represent a useful model to analyze the influence of a microorganism on human behavior and the development of psychiatric disease [15]. It is an obligate intracellular protozoa belonging to the *Coccidia* subclass of the phylum *Apicomplexa* and causes toxoplasmosis. Its definitive hosts are cats and other felines, which

are the only animals in which the sexual stage of their life cycle takes place (in intestines), forming oocysts that are eliminated through the feces. Hot-blooded vertebrates such as birds and other mammals, including humans, are intermediate hosts. Humans can become infected by various pathways, such as: the intake of undercooked meat containing latent forms of the parasite (bradyzoites in tissue cysts), fresh food (e.g., fruit and vegetables), or water contaminated with oocysts from cat feces; blood transfusions; transplantation of solid organ or stem cells, or transplacental transmission. Upon reaching tissues, *T. gondii* rapidly replicates in the form of tachyzoites until tissue proliferation and expansion of the parasite are impeded by the immune response, after which its replication slows and it remains in tissue cysts in latent or bradyzoite form. Cysts are most frequently found in skeletal muscle, myocardium, CNS, and eyes and are responsible for persistent infection [16, 17].

Primary infection usually takes place during childhood, when only a small percentage of people show symptoms, which are mild and include general discomfort, lethargy, cervical lymphadenopathy, and/or eye disease, among others. Most parasitized individuals remain asymptomatic for a long time period, even throughout their life, and host the latent form of *T. gondii*. However, chronic infection can be reactivated in immunocompromised individuals (AIDS, transplanted, and oncology patients, etc.), giving rise to various symptoms and even, in death. This reactivation is often associated with nervous system symptoms, such as Guillain-Barré syndrome, diffuse encephalopathy, meningoencephalitis, or brain abscesses [17, 18]. Human parasitizations, although generally considered asymptomatic, may cause behavioral disorders and the development of a psychiatric disease such as schizophrenia due to damage resulting from the initial infection, from the host immune response to the parasite, or from the persistence of cysts in the CNS [19]. Accordingly, the concept of asymptomatic chronic parasitization is currently under debate [20].

T. gondii is a plausible candidate as an infectious origin of schizophrenia and has attracted considerable research attention for the following reasons: the possibility of its transplacental transmission; its marked neurotropism; its capacity for persistent infection, remaining in latent form but with the possibility of reactivation; its association with brain development disorders and anomalies; its relationship with behavior disorders in animal and human models; and *in vitro* evidence of the inhibition of its growth in cell culture by antipsychotic drugs.

2. Epidemiologic data on the association between toxoplasmosis and schizophrenia

One of the first approaches adopted to explore a possible relationship between *T. gondii* infection and schizophrenia was to analyze epidemiological data on the two diseases. Initial conclusions were as follows:

- Both have a familial incidence. This is explained in the case of schizophrenia by the possible participation of certain genes in its pathogeny [21], and in the case of toxoplasmosis by the possible common exposure of family members to the parasite, although a genetic influence has also been proposed [22].

- There is a relatively high frequency of stillborns among both schizophrenic [23] and parasitized [24] mothers.
- Both diseases typically show a decreased prevalence in geographic areas with small populations of felines [25, 26].
- Initial symptoms in both diseases commonly manifest between the second and third decade of life [27, 28].
- The prevalence of both diseases is higher among populations with lower socioeconomic level and living in overcrowded conditions [29, 30].

These and other published findings indicate that the two diseases have some similar features and may even be epidemiologically related. However, they are inadequate to establish etiological relationships, and a pathophysiological approach is required to explore causality.

3. Studies based on the detection of anti-*Toxoplasma gondii* antibodies

For more than six decades, the relationship between schizophrenia and toxoplasmosis has been explored by studying a specific immune response [31]. Various meta-analyses have demonstrated a significantly higher prevalence of anti-*T. gondii* antibodies in schizophrenic patients than in controls, with odds ratios ranging between 2.7 [11, 32–34] and 1.8 [35].

In the natural time course of toxoplasmosis, IgM antibodies against *T. gondii* are the first to be detected in serum, a few days after infection. These are usually negativized between weeks 4 and 12 but can remain detectable for months or even years in a large number of patients. IgG antibodies are detected at around 2 weeks later than IgM antibodies, reaching a maximum level in the 2nd to 3rd month and persisting throughout life in residual titers. The presence of IgM antibodies in the absence of IgG indicates recent infection, while the presence of IgG indicates chronic infection, especially in the absence of IgM. The reactivation of a persistent infection can be accompanied by increased IgG and/or IgM values, although these antibodies can be undetectable in immunocompromised patients [36].

Most studies have centered on the humoral immune response, comparing anti-*T. gondii* IgG and IgM antibodies between schizophrenic patients (in different clinical/therapeutic situations) and controls. This method is widely employed because of the ease with which samples (usually serum, occasionally cerebrospinal fluid) can be gathered and the high degree of reproducibility, specificity, and sensitivity obtained. Many of these studies reported higher levels of antibodies (IgG and, in some studies, IgM) against *T. gondii* in patients with schizophrenia than in other populations, including patients with a different psychiatric disorder [32, 37–51]. However, findings have been inconsistent [52, 53], and account should be taken of the publication bias against studies without significant results [11].

Clinical manifestations differ between seropositive and seronegative schizophrenic patients, with a predominance in the former of positive symptoms (delirium, hallucinations, disorganized thinking), cognitive disorders (abstract thinking difficulties, disorientation, attention

deficit), and agitation [50, 54]. Some researchers also observed that patients with schizophrenia and anti-*T. gondii* antibodies had a significantly higher risk of dying from natural causes [55] and were more likely to attempt suicide [56] in comparison with seronegative patients.

The above studies suggest a strong association between these diseases, with a significantly higher frequency of chronic parasitization in schizophrenic patients than in other population groups. However, if schizophrenia is a consequence of chronic CNS infection, which usually takes place at an early age, the question arises as to why it typically appears between the second and third decades of life. According to Yolken et al. [51], a significant increase in IgG titers observed in patients with a first schizophrenia episode may be compatible with a reactivation of the infection (previously in latent phase) that becomes clinically manifest in the onset of the disease *via* an immune-mediated mechanism. Some authors proposed that the immunoglobulins may cross the blood-brain barrier in this situation and react with brain tissue antigens due to their molecular mimicry with *T. gondii* antigens. This is similar to observations in such autoimmune diseases as systemic lupus erythematosus or in paraneoplastic syndromes [57]. Associations with the presence of anti-*Toxoplasma* IgM are less well documented [35], although Monroe et al. [58] reported in their meta-analysis a significant 1.7-fold greater likelihood of *T. gondii* IgM antibodies in patients with acute psychosis than in controls. It was concluded that *T. gondii* IgM antibodies may indicate either an acute/recent infection or a reinfection, possibly with a different genotype.

However, although a strong association has been described between schizophrenia and parasitization, these studies do not provide evidence to confirm the hypothesis on the infectious etiology of schizophrenia, and a causal relationship has not been demonstrated. Contact with *T. gondii* may be favored by the anomalous behavior, disorganized lifestyle, and/or weaker socioeconomic situation of schizophrenics, with infection being the consequence rather than the cause of their disease, which may explain the positive serological results [50].

4. Seroprevalence studies in mothers and newborns

The possible transplacental transmission of *T. gondii* has attracted considerable attention in seroprevalence studies. Maternal infection during the first or second trimester of pregnancy can lead to severe problems in the offspring, including intracranial calcifications, chorioretinitis, blindness, deafness, hydrocephaly, microcephaly, mental retardation, psychomotor retardation, pancytopenia, or epilepsy. The timing of the transmission is an influential factor: early maternal infection less frequently affects the fetus but is associated with a more severe congenital toxoplasmosis that may result in intrauterine death and miscarriages, while later maternal infection (third trimester) increases the risk of affecting the fetus but is associated with offspring who are asymptomatic [17]. Complications, possibly including schizophrenia, can appear decades later in patients with initially undetected infection due to its reactivation [59].

This type of study can be classified into two groups: those on the presence of antibodies in pregnant women and the development of schizophrenia in their offspring; those on the pres-

ence of antibodies in newborns and their later development of schizophrenia. Among the former, Brown et al. [60] and Blomström et al. [61] demonstrated associations between increased anti-*Toxoplasma* IgG levels in pregnant mothers and risk of schizophrenia in their offspring, although other researchers published discrepant results [62]. Xiao et al. [63] observed a significant association between the presence of maternal antibodies against type I *T. gondii* (but not against types II or III) and the onset of psychotic disorders in the offspring. Among the latter group of studies, Mortensen et al. [59] demonstrated that newborn levels of anti-*T. gondii* IgG levels (from the mother) were significantly higher in individuals who developed schizophrenia in adulthood.

Published data suggest that schizophrenia risk in offspring is associated with persistent maternal infection by *T. gondii* but is not directly related to acute maternal infection [64]. If this were the case, a significant association could be expected between the presence of IgM in the serum of mothers and/or newborns and the presence of the disease, which has not been demonstrated [60]. However, this relationship may be masked by the low frequency of anti-*Toxoplasma* IgM detection in pregnant women [24, 65].

As noted above, increased maternal IgG levels can cross the placenta (unlike IgM antibodies) and may damage fetal brain development by molecular mimicry [60, 64]. However, the presence of maternal IgG may indicate a reactivation of latent infection due to the impact of immune system disorders on protozoan replication control during pregnancy [66]; hence, brain development could also be impaired by transplacental transmission and/or the passage of inflammatory cytokines to the fetus [67, 68].

The majority of schizophrenic patients do not have anti-*Toxoplasma* antibodies, and the majority of seropositive patients are not schizophrenic. Therefore, *T. gondii* would only explain a minority of cases. Other factors under investigation that may explain why only some parasitized individuals develop schizophrenia include genetic susceptibility, the infective genotype of the parasite, the existence of different infection pathways, and the timing of toxoplasmosis onset [20, 33, 63].

5. Studies based on *Toxoplasma gondii* nucleic acid detection

Studies of animal brain biopsies have shown *T. gondii* to have high neurotropism, with the capacity to infect glial cells (especially microglia and astrocytes) and neurons, forming persistent cysts in brain tissue [69]. Although no tropism for specific brain regions has been observed, with cysts being detected in many areas, the most frequently parasitized regions are the hippocampus, thalamus, cerebral cortex, cerebellum, olfactory bulb, and, especially, the amygdala [70–73].

However, the presence of brain cysts can only be detected in *postmortem* biopsies, explaining the few studies of this type and the predominance of serological techniques for the detection of chronic infection by *T. gondii* in humans. The presence of glial anomalies, including a reduced amount of astrocytes, has been reported in the brains of schizophrenic patients [74], and it has been speculated that these may possibly result from infection by *T. gondii* [31].

Imaging techniques have revealed a lower density of gray matter in certain brain regions of schizophrenic patients [75], which may be directly related to the infection, given that non-parasitized schizophrenic patients were found to have the same brain morphology as healthy controls [76].

One of the few studies of *postmortem* brain biopsies found no parasite DNA in any subject (14 schizophrenic patients and 26 healthy controls) [77]. There appear to be three possible explanations: first, there was truly no association with the infection; second, the biopsies missed infected brain areas; and finally, the sensitivity of the nucleic acid detection technique might be inadequate. In addition, the detection of parasite DNA only demonstrates the presence of the parasite not its possible effect on the parasitized individual and would not establish an etiological relationship with schizophrenia. Thus, the detection of parasite DNA in brain tissues does not distinguish between asymptomatic patients with latent parasitization and those with encephalitis [17].

A study of blood samples detected parasite DNA in 33 out of 101 samples from schizophrenic patients *versus* 2 out of 55 samples from controls, a significant difference [46]. In contrast, Gutiérrez-Fernández et al. [32] detected parasite DNA in only 1 out of 128 blood samples from schizophrenic patients and in none of 143 samples from controls (nonsignificant difference). However, although the presence of parasite DNA in blood indicates acute infection [17], it does not necessarily signify infection of the brain, and no relationship was found between anti-*Toxoplasma* IgM and schizophrenia in the aforementioned study [46].

6. Studies on behavioral disorders in animals and humans

Research on this issue has included experimental animal studies, mainly in rats and mice. Parasitized animals have shown various behavioral changes, becoming more active, expressing less fear when examining new stimuli, reducing their natural aversion to cat odor or even becoming attracted to it, and demonstrating reduced learning ability and attention or memory deficits [78–83]. According to the “behavioral manipulation hypothesis,” these disorders in their intermediate hosts (rodents) represent an evolutionary adaptation of the parasite, facilitating their capture by their definitive host (felines) and completing their life cycle [84, 85]. Although the mechanism by which *T. gondii* induces these behavioral changes is poorly understood, various possibilities have been proposed. It may be due to a direct effect on tissue cysts in specific brain areas such as the amygdala or hippocampus, given that the host response to predator odors was changed by the parasite in male rats infected with *T. gondii* by inducing hypomethylation of the neuropeptide arginine vasopressin in the posterodorsal part of the medial amygdala, an important node of the extrahypothalamic vasopressin system that contains a large number of arginine vasopressin neurons. This epigenetic manipulation produced a greater activation of vasopressinergic neurons after exposure to cat odor, leading to the reversion of fear into attraction [86]. It may also result from the effect of a more diffuse and wider involvement of brain tissues, with no apparent changes, that nevertheless give rise to a series of neurophysiological disorders. Changes may also result from inflammation (encephalitis) caused by the immune activation induced by parasitization, which would increase

inflammatory cytokines in the rodent brain, such as tumor necrosis factor alpha (TNF- α), interleukin-1 β (IL-1 β), IL-10, interferon gamma (IFN γ), C-reactive protein, tissue inhibitor of metalloproteinases 1 (TIMP-1), or vascular cell adhesion molecule 1 (VCAM-1), similar to observations in *postmortem* biopsies of schizophrenic patients. Finally, the behavioral changes have also been related to neurochemical mechanisms, with an increase in dopamine and homovanillic acid and a decrease in norepinephrine levels [73, 84, 85, 87–90].

Any of the above mechanisms in rodents could also produce behavioral changes in the brains of other intermediate hosts, including humans. Thus, research in humans also suggests that toxoplasmosis may alter behavior, psychomotor abilities, or personality, with the corresponding clinical consequences [84]. These disorders would be more related to latent rather than acute toxoplasmosis, given that its emergence, frequently several years after primary infection and not during the acute phase, would indicate that it results from slow and possibly accumulative changes induced by parasite activity [91–93]. The study by Horacek et al. [76] demonstrates that, in seropositive schizophrenic patients, latent parasitization is associated with a significant reduction in gray matter volume in specific brain areas (cortical regions, hippocampus, and caudate nucleus), which is not observed in seronegative patients.

Reinforcing the relationship between the parasite and the psychiatric disease, it has been demonstrated that haloperidol, an antipsychotic drug that blocks D2 dopaminergic receptors in the mesolimbic system and often used in the symptomatic treatment of schizophrenia, inhibits the replication of tachyzoites in cell cultures *in vitro*. This effect may at least partly be due to the capacity of this drug to inhibit calcium transport, blocking cell ion channels [94]. The interaction between tachyzoites and host cells is calcium-dependent; hence, cell invasion capacity can be inhibited by the presence of drugs that block calcium channels, such as haloperidol [95]. Experimental studies with rodents have also demonstrated that some behavioral changes caused by the infection are reverted by using the antipsychotic, and that there are fewer parasitized neurons and glial cells after the treatment; this is observed using immunohistochemical techniques [96]. It is therefore possible that its therapeutic effect can be explained in patients with schizophrenia by various mechanisms, given that on the one hand, it blocks dopamine, whose levels are often elevated in schizophrenia patients parasitized with *T. gondii* [89, 97], and on the other hand, it can inhibit parasite replication in brain cells [96]. Other antipsychotic drugs such as fluphenazine, thioridazine, trifluoperazine, or zuclopenthixol, and mood stabilizers, e.g., valproic acid, were also found to inhibit *T. gondii* proliferation in cell cultures [94, 98, 99].

Antipsychotics are especially indicated in patients with a predominance of positive symptoms and agitation (as in the acute phase of schizophrenia), which are significantly more frequent in those parasitized with *T. gondii*, as noted above. The greater effectiveness of these drugs in these situations may be due not only to their dopamine blocking effect but also to their anti-*Toxoplasma* activity. Thus, these treatments were found to reduce anti-*Toxoplasma* antibody levels in seropositive schizophrenic patients, indicating their antiparasitic effect [44]. These findings suggest that these drugs may possibly have a beneficial effect on schizophrenic patients parasitized with *T. gondii*.

Studies to date on the possible effect in these patients of drugs with anti-*Toxoplasma* activity (e.g., pyrimethamine, sulfadiazine, azithromycin, or trimethoprim-sulfamethoxazole) have not demonstrated significant improvements in psychotic symptoms [100, 101]. In fact, drugs used to treat toxoplasmosis are largely active during the tachyzoite replication phase, and their effectiveness against bradyzoites in tissue cysts is drastically reduced once chronic infection by *T. gondii* is established [102].

The etiological relationship between parasitization and schizophrenia has not yet been established, despite the above data on behavioral changes in animals or humans and on the effects of antipsychotic drugs on symptoms. In addition, differences in behavioral disorders between humans and rodents may mean that results in animal models cannot be extrapolated to humans. It should also be borne in mind that the mild behavioral modifications associated with the infection cannot necessarily be considered symptoms of a psychotic disease.

7. The role of proinflammatory cytokines

The host response to the parasitization of glial cells and neurons involves the activation of immune system cells, including T lymphocytes (CD4+ and CD8+), B lymphocytes, NK cells, macrophages, and dendritic and glial cells. These produce a wide variety of inflammatory cytokines such as IFN γ , interleukins (IL-1, IL-1 β , IL-2, IL-4, IL-6, IL-10, IL-12, IL-15, IL-17, IL-18, IL-23), granulocyte macrophage colony-stimulating factor (GM-CSF), and/or TNF α [69, 103]. These cytokines halt protozoan proliferation and limit their replication, playing a key role in regulating the infection of host cells, thereby favoring the formation of tissue cysts and the development of the chronic latent form [20]. These and other inflammatory responses have also been reported in schizophrenia [104] and are therefore involved in brain disorders both in this disease and in *T. gondii* infection [105].

Thus, infection of brain tissue by *T. gondii* produces activation of the Jak/STAT pathway, which is recognized as an important regulatory mechanism in CNS development, function, and disease progression [106, 107]. This pathway comprises three elements: a ligand receptor, the majority are receptors of cytokines such as IFN γ ; Janus kinase (Jak) proteins associated with the receptor within the cell, which possess tyrosine-kinase activity; and signal transducer and activator of transcription (STAT) proteins, which act as transcription factors that move toward the cell nucleus after their phosphorylation, where they bind with regulatory sequences of genes designated gamma interferon activation sites (GAS) [108]. In mammals, the Jak/STAT pathway induces the transcription of genes that participate in multiple processes, including antimicrobial activity and the production of proinflammatory cytokines [109]. Among other effects, an increase is produced in the expression of NADPH oxidase enzyme (NOX2) and inducible nitric oxide synthase (iNOS). These enzymes are responsible for the production of reactive oxygen species (ROS) and reactive nitrogen species (RNS), which assist the destruction of foreign pathogens [110, 111] but have been linked to seizures, stroke, neurodegenerative diseases, and schizophrenia [111, 112] as a consequence of their toxic effect on neurons [113]. Scientific evidence points to ROS-mediated oxidative damage as a key pathogenic pathway involved in infection-mediated neuropathy. According to these findings, it can be

expected that a high degree of degenerated neuron degeneration and cognitive impairment is associated with the presence of *T. gondii* in the brain [111].

8. The importance of dopamine and other neurotransmitters

As already noted, some experimental animal and human studies concluded that behavioral changes may be explained by increased dopamine levels in the parasitized brain, and that these disorders could largely be resolved by administration of a dopaminergic receptor antagonist (e.g., haloperidol) or dopamine reuptake inhibitor (e.g., GBR-12909) [96, 114, 115]. It is therefore possible that dopamine represents the link between toxoplasmosis and schizophrenia [97]. This neurotransmitter is synthesized in the cytosol of neurons from L-tyrosine amino acid by the action of the tyrosine hydroxylase enzyme, which converts it to L-3,4-dihydroxyphenylalanine (L-DOPA). L-DOPA is in turn converted by the action of DOPA-decarboxylase (DDC) to dopamine, a precursor of norepinephrine (noradrenalin) and epinephrine (adrenalin) in the synthesis pathway of these catecholamines. It is subsequently packaged in vesicles and transported through the axon to the synapse, where it is released by exocytosis in response to an electrical stimulus. Dopamine is one of the main neurotransmitters in the prefrontal cortex and the mesolimbic system (mainly formed by the nucleus accumbens, amygdala, and hippocampus), where the presence of *T. gondii* tissue cysts is especially frequent [73].

The definitive mechanism by which *T. gondii* induces changes in the dopaminergic pathway has not been fully elucidated. However, an increase in dopamine with no modification of cellular tyrosine hydroxylase was demonstrated *in vitro* after parasitization of a rat pheochromocytoma cell line (PC12) and *in vivo* after the parasitization of mouse brains. This dopamine synthesis is attributable to the additional activity of the aromatic amino acid hydroxylase, which is encoded by two *T. gondii* genes [116] and has homologous activities to those of mammalian tyrosine hydroxylase, associated with the entry of cellular DDC enzymes into parasitophorous vacuoles (compartments formed by the parasite to invade the cell) and into tissue cysts (the protozoan encodes no enzyme with DDC activity) [114, 117]. Experiments in cell cultures have demonstrated that dopamine increases the replication of *T. gondii* tachyzoites [118]. This biochemical mechanism may play a role in the behavioral changes observed, which would result from the involvement of catecholaminergic neurons and consequent dopaminergic hyperactivity [19].

Parasitization in the fetal period may also impair the development of mesolimbic dopaminergic or prefrontal cortex neurons (inappropriate migration, altered position, reduced synapses, etc.) leading to neurodevelopmental disorders. Disease symptoms would not be induced immediately by these early anomalies but would rather manifest after a latency period of one to three decades. This is because the proliferation, migration, differentiation, and maturation of glial progenitor cells continue throughout childhood [119] and the volume of gray matter increases to a peak in puberty before beginning to diminish [120].

However, the hypothesis that increased dopamine levels or dopaminergic hyperactivity is the underlying cause of schizophrenia does not account for the negative symptoms in these

patients, which are more likely to result from dopaminergic hypoactivity. Therefore, neurotransmitters other than dopamine may play an important role in the development of this disease. Thus, it has been proposed that deficits in glutamatergic brain systems also participate in the physiopathology of schizophrenia based on findings of higher kynurenic acid levels in patients with psychotic symptoms than in healthy controls [121]. Kynurenic acid is a metabolite of tryptophan with important biological effects on the nervous system, related to its antagonism for the glutamate receptor in the human brain (it is a glutamatergic NMDA receptor antagonist). Increased kynurenic acid levels due to blockade of NMDA receptors in glutamatergic neurons have been related to changes in dopamine level in different brain areas. These modifications include cortical dopaminergic hypoactivity and mesolimbic dopaminergic hyperactivity, which would explain the negative and positive symptoms in patients. This relationship between glutamate pathway disorders and dopamine level changes may explain the presence of different symptoms in the psychosis [122].

Indoleamine-2,3-dioxygenase and therefore the metabolism of tryptophan, a precursor metabolite in serotonin (and melatonin) synthesis, are induced by the proinflammatory cytokines released in response to *T. gondii* infection, especially IFN γ [123]. Tryptophan is an essential amino acid for the parasite, and decreased levels inhibit its growth and replication capacity [124]. However, induction of this metabolite in turn increases kynurenic acid levels and therefore alters dopamine levels through the glutamatergic receptor antagonist effect of this acid [114]. Tryptophan degradation also reduces serotonin levels, which has been related to a higher incidence of depression and suicide [125, 126], as also observed in patients with high anti-*T. gondii* antibody levels [56].

Patients with schizophrenia also show anomalous levels of gamma-aminobutyric acid (GABA), another important neurotransmitter [20], which is synthesized from glutamate by the action of glutamic acid decarboxylase (GAD) [127]. GABA activates GABA_A receptors, which are ion channels, and GABA_B receptors, which are G-protein-coupled receptors [128]. It is the main neurotransmitter with inhibiting effect in the CNS, regulating dopaminergic activity and playing a key role in the reduction of neuronal excitability throughout the nervous system. Dopaminergic neurons in basal ganglia would be directly inhibited by GABAergic neurons, so that any GABAergic hypofunction would be accompanied by an increase in subcortical dopaminergic activity, as observed in schizophrenia.

More direct evidence of the involvement of this neurotransmitter in the etiology of schizophrenia derives from data on the reduction in neurons in the GABAergic system or in brain regions such as the hippocampus, temporal lobe, and prefrontal cortex of schizophrenic patients [129–131]. *Postmortem* molecular studies have demonstrated: a reduction in messenger RNA (mRNA) levels of isoform 67 of glutamic acid decarboxylase (GAD67) and of type 1 GABA transporter (GAT-1) in the prefrontal cortex of schizophrenics [132, 133]; an increase in subunit $\alpha 2$ of the GABA_A receptor in the initial segment of the axon of pyramidal neurons [134]; and a reduced expression of the receptor GABA_B, which regulates GABA release as a possible compensatory mechanism for GABAergic dysfunction [135]. As noted above, these findings may be the consequence of alterations during neurodevelopment in the differentiation and migration of these neurons toward their definitive localizations in the brain. This

would give rise to structural alterations and neurochemical dysregulation that would have a global effect on all of these neurotransmitters (dopamine, glutamate, serotonin, GABA) and would become manifest from adolescence onward, inducing the appearance of the disease. Once more, infection by *T. gondii* may play an important role in this process.

Outside the nervous system, GABAergic mechanisms have been observed in different tissues and peripheral organs, and GABA has also been found to exert a major role in the immune system, with important inter-regulatory functions between this and the CNS [136]. It has been reported that *T. gondii* infection is followed by an increase in the motility and migratory capacity of infected dendritic cells, permitting propagation of the parasite to different tissues, including the brain [128]. Although dendritic cells are considered guardians of the immune system, they can also, paradoxically, mediate in the spread of the parasite. This mechanism is produced by the induction in these cells of the GAD enzyme and therefore of GABA production and secretion, which in turn activate GABA receptors expressed by these same cells, stimulating their motility [137]. In experimental mouse models, inhibition of the GABAergic pathway by blockade of GABAA receptors or inhibition of the GAD enzyme markedly reduced the hypermotility and spread of *T. gondii*-infected dendritic cells and therefore of the parasite itself [137, 138]. Finally, it has also been reported that brain infection by *T. gondii* can interfere with the GABAergic system by inducing changes in the distribution of the GAD67 enzyme, although this event has been related more to possible neurological complications of toxoplasmic encephalitis, such as seizures [139], than to possible complications of latent toxoplasmosis, such as schizophrenia.

Accordingly, the inflammatory response of the host to parasitization, which aims to control parasite replication and alterations in differentiation and migration processes, can change levels of dopamine, tryptophan, kynurenic acid, serotonin, and GABA, leading to behavioral changes and giving rise to different psychotic symptoms.

In order to establish dopamine and other related neurotransmitters as a causal link between toxoplasmosis and schizophrenia development, it is necessary to confirm that this neurotransmitter is also involved in the disease genesis when there is infection by other pathogens [140], and this mechanism should also explain the possible contribution of *T. gondii* parasitization in other dopaminergic pathway diseases, e.g., Parkinson's disease [114].

9. The N-methyl-D-aspartate receptor hypofunction theory: anti-NMDAR antibodies

Encephalitis due to antibodies against the glutamatergic NMDA receptor (anti-NMDAR antibodies) is an autoimmune disease caused when antibodies produced by the host immune system identify NMDA receptors as foreign antigens. This receptor forms a heterotetramer between two GluN1 and two GluN2 subunits and participates in essential functions for reality perception, memory, and the control of unconscious activities. The disease is characterized by the hypofunction of NMDA receptors, which would account for the psychotic symptoms, personality changes, memory impairment, and psychomotor agitation

[141, 142]. It usually arises during the course of a paraneoplastic process and is frequently associated with the development of ovarian teratomas, explaining its higher incidence among females [143, 144]. Likewise, 14–75% of patients with systemic erythematosus lupus, another autoimmune disease, have been reported to manifest psychiatric symptoms related to the presence of the same antibodies [145, 146]. This involvement of anti-NMDAR antibodies (and other neurotransmission receptors) indicates an important link between immune abnormalities and altered neurotransmission in schizophrenia, major depression, or bipolar disorder [147, 148].

The presence of anti-NMDAR antibodies has been documented in schizophrenic patients in the absence of seizures, movement disorders, or other neurological signs or symptoms [149–151], although other researchers were unable to replicate these findings [152, 153]. For various reasons, the production of anti-NMDAR antibodies is a plausible mechanism to explain at least a percentage of schizophrenic cases [149]: several studies reported that 5–10% of cases are associated with the presence of these antibodies in serum and cerebrospinal fluid [150, 151, 154]; kynurenic acid is an antagonist of glutamate *via* blockade of NMDA receptors, as commented in the previous section, suggesting that it contributes to the pathogenesis of schizophrenia [122]; persistent blockade of NMDA receptors in experimental animals recreates clinical characteristics of schizophrenia [155]; selective elimination of subunit GluN1 of the NMDA receptor in neurons of the cortex and hippocampus in early postnatal development contributes to the pathophysiology of schizophrenia-related disorders in mice [156]; some of the genes associated with schizophrenia are related to the NMDA receptor [157]; NMDA receptors are reduced in medication-free schizophrenic patients [158]; blockade of the receptor with ketamine or phencyclidine produces psychotic symptoms [159, 160]; and *de novo* mutations (large chromosomal copy number changes) affect genes that encode one or more nucleotides among the glutamatergic postsynaptic proteins that form part of the receptor, providing insight into possible etiological mechanisms underlying schizophrenia [161].

Maternal infection during brain development or infection during childhood may produce anti-NMDAR antibodies, while other environmental or genetic factors may influence the age of disease onset [149]. Certain pathogens have been associated with elevated anti-NMDAR antibodies [162, 163]. Thus, a *T. gondii*-infected mouse model showed a significantly higher increase in serum GluN2A autoantibodies among juvenile- *versus* adult-infected mice. Adolescence is a critical window in neurodevelopment, and the authors hypothesized that early infection would have greater effects on behavior and the brain in comparison with adult infection. It is possible that chronic infection with *T. gondii* affects pre- or postnatal brain development by altering synaptic maturation. An increase in NMDAR autoantibodies due to *T. gondii* exposure might underlie behavioral alterations in symptomatic individuals [164].

10. Studies on gene-infection interaction

Various studies have demonstrated the participation of numerous genes in schizophrenia, providing firm evidence on the involvement of genetics in the etiology of the disease

[165]. Some authors have described inheritability in >80% of cases, and schizophrenia has been associated with polymorphic variability in certain genes [21, 166–168]. However, the genetic hypothesis alone cannot explain the familial association of schizophrenia with other diseases, the seasonal peaks of schizophrenia births, the different prevalences among residents of urban and rural areas, discordant results between monozygotic and dizygotic twins or between dizygotic twins and full siblings, or correlations in adopted children, which are, however, consistent with an infectious etiology [1]. Schizophrenia is likely a genetically complex disease that does not follow a Mendelian transmission pattern but rather involves multiple genes, each with a small effect, which act in combination with epigenetic and environmental factors [169]. Accordingly, epidemiological findings suggest that a combination of intrinsic (genetic) and extrinsic or environmental factors, including infections, may participate in the origin of this disease, operating during the development of the individual at some time between conception and adolescence [7]. Tomonaga [170] proposed that persistent chronic infections or the expression of microbial proteins may directly and/or indirectly affect CNS functions in infected individuals, changing the expressions of genes related to schizophrenia and increasing the risk of suffering this disease or at least some of its varied clinical phenotypes.

Genes whose variants or polymorphisms have been associated with the risk of schizophrenia include some that encode proteins with important functions in neurodevelopment or neurodegeneration and in neuronal neurotransmission circuits. This is the case of the gene that encodes neuregulin 1 (NRG1), a key molecule in maintaining brain synaptic plasticity in adults, which has been related to schizophrenia etiology [171, 172], and the genes that encode catechol-O-methyltransferase (COMT) [173], proline dehydrogenase (PRODH) [174], dysbindin protein (DTNBP1) [175], a regulator of G4 protein (RGS4) [176], a regulator of potassium calcium channels (KCNN3) [177], and D-amino-oxidase complex (G72, DAAO) [178], among others [179]. The genes that encode these proteins are located in chromosomal regions that have been described as relevant for the study of schizophrenia, and many of these proteins participate in glutamatergic, dopaminergic, or serotonergic neurotransmission circuits.

Genetic polymorphisms that increase susceptibility to schizophrenia, including some of the above, have also been related to resistance or susceptibility to certain infections through their important role in the life cycle of some pathogens, including *T. gondii* [169, 179, 180]. Schizophrenia may possibly correspond to a model in which various genes may interact with microbial agents in a process that is probably mediated by the inflammatory and immune response of the individual, increasing the risk of developing psychiatric disease [169, 179–182]. It appears reasonable to assume that infections may interact, thereby changing the expressions of schizophrenia-related genes and increasing the risk of suffering this condition.

Various rodent [79, 183, 184] and human [185, 186] studies have supported the existence of genetic susceptibility to *T. gondii* parasitization, suggesting that if the parasite were one of the possible causes underlying schizophrenia development, this genetic susceptibility might also explain familial cases of schizophrenia [1]. As commented above, some *T. gondii* genes encode proteins with a similar activity to that of enzymes (e.g., tyrosine hydroxylase) in the cells of their intermediate hosts. Therefore, this parasite has genes that allow it to “manipulate” the

behavior of the host and facilitate its capture by the cat, its definitive host, thereby favoring parasite survival. The presence of these genes is consequently an evolutionary advantage of *T. gondii* [19].

Genetic studies (in animals and humans) currently center on the possible presence of genes or specific allelic variants that interact with the genes of microorganisms that can infect the patient (gene-infection interaction hypothesis), increasing the risk of schizophrenia [187–189]. Thus, it has been demonstrated that a critical role in human congenital *T. gondii* infection is played by the *ALOX12* gene, which encodes arachidonate 5-lipoxygenase enzyme, which is involved in fatty acid metabolism and has been related to schizophrenia, at least in a Korean population [190, 191]. HLA-related genes such as *SGK1* on chromosome 6, which plays a role in regulating different brain functions [192] and mediates the effects of cortisol on hippocampal neurogenesis [193], have a modulating effect on some infectious agents, including *T. gondii*, consistent with the proposition that parasitization may modify the risk of schizophrenia [187]. In a study of mice parasitized with *T. gondii*, heterozygous deletion of the *Nurr1* gene (*Nurr1* ± genotype), an orphan nuclear receptor essential for the development of mesencephalic dopamine neurons [194], predisposed the animals to behavioral disorders that involve dopamine neurotransmission associated with schizophrenia symptoms [195].

A further example in support of this hypothesis is the Akt cell signaling system. The *Akt* gene encodes a serine-threonine kinase with three isoforms (*Akt1*, 2, and 3), whose activation mediates cell survival processes and whose inhibition favors apoptosis. As commented above, the innate immune system induces a range of processes after infection of brain cells by *T. gondii*, including antimicrobial activity and the generation of ROS to assist in the destruction of foreign pathogens. However, increases in ROS concentrations activate the Akt system, which guarantees cell survival and allows the pathogen to persist and replicate within the infected cell. Akt is above all activated in pathophysiological situations in which ROS increase as the result of ischemia-reperfusion, playing an important role in the protection of the different cells and tissues involved, including nerve tissue [196]. On the other hand, Akt is known to affect dopaminergic signaling, and polymorphisms of the *Akt1* gene have been found to increase the risk of developing schizophrenia through its relationship with dopaminergic pathways of the prefrontal cortex [197].

Other researchers reported similar associations between schizophrenia risk and other human pathogens, supporting the gene-infection interaction hypothesis [198–201]. This research line on the effects of interaction between genes or genetic variants on the risk of schizophrenia related to *T. gondii* parasitization is highly likely to establish the true causes of the disease, at least in some types of patient.

11. Is there an etiological association between *Toxoplasma gondii* infection and schizophrenia development?

Numerous studies have contributed evidence on the involvement of toxoplasmosis in the pathogenesis of numerous CNS diseases, including bipolar disorder, depression, Alzheimer's disease, Parkinson's disease, and epilepsy [49, 202–204]. However, the main advances over

the past few years have been achieved by research on deciphering the molecular mechanisms underlying the pathophysiology of schizophrenia.

This chapter analyzes data from *in vitro* and animal and human *in vivo* studies in order elucidate points of connection between *T. gondii* and schizophrenia. It can be concluded that infection by *T. gondii* is highly likely to be a cause of the disease for the following reasons: it is a neurotropic microorganism that persistently invades glial cells and neurons; it generates brain development anomalies; it reduces brain gray matter density; it elicits an inflammatory and immune response that alters neurotransmission systems; it affects cognitive function and behavior; and its replication is inhibited by some antipsychotics. All disorders reported for the parasite are associated with the development of psychotic symptoms. Furthermore, specific genetic polymorphisms linked to an increased risk of schizophrenia have also been associated with a higher likelihood of infection by this parasite. Nevertheless, despite all of the above evidence on this possible pathogenic association, one important

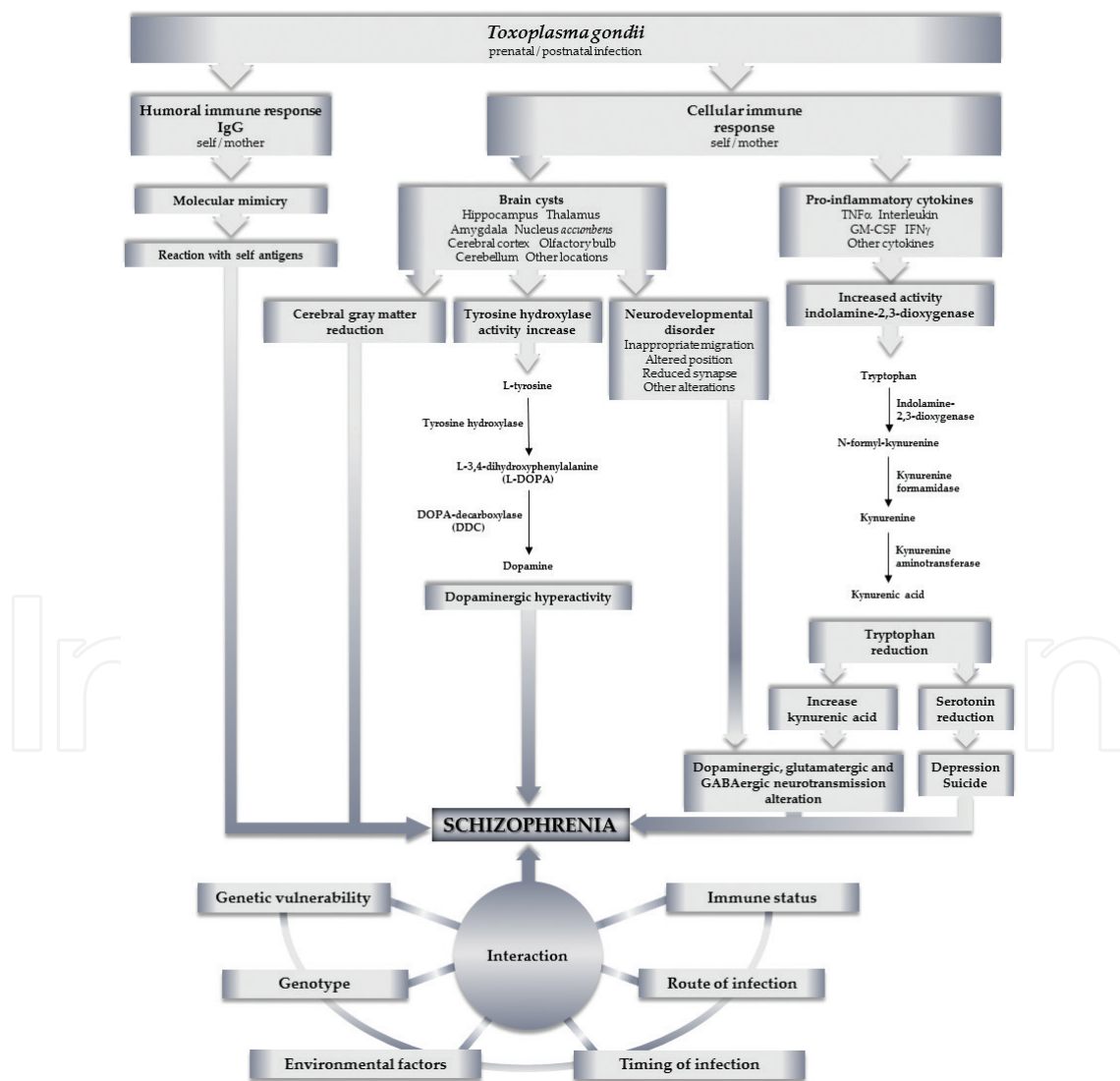


Figure 1. Likely involvement of infection by *Toxoplasma gondii* in the development of schizophrenia.

question remains to be resolved, which is why most individuals with signs of infection by *T. gondii* are asymptomatic and only a few develop psychiatric disorders.

Schizophrenia is a complex disease with innumerable symptoms, and its presentation and severity vary among patients. According to the infectious hypothesis of this disease (**Figure 1**), differences among patients would be influenced by their genetic predisposition or vulnerability, their immune status, the timing of parasitization (congenital, neonatal, or adult), the time interval since their first contact, and/or the particular brain area(s) affected. Characteristics of the infection also play a role, including its source (oocysts or tissue cysts), possible interactions with other infectious agents, and the genotype; thus, genotypes II and III more frequently establish chronic infections and show a greater expression of tyrosine hydroxylase genes in comparison with genotype I, and they may be more strongly related to behavioral changes [205].

Finally, the biology of schizophrenia must be fully elucidated to support the appropriate design of disease-modifying therapies or novel antipsychotic drugs. There appears to be sufficient evidence to suggest that schizophrenic patients with *T. gondii* infection could clinically benefit from a combined therapeutic approach based on the prescription of current or future antipsychotic drugs with antitoxoplasmic activity. However, published results have not been conclusive [206], and randomized controlled prospective trials are required in wider samples, stratifying schizophrenic patients into subgroups (e.g., by clinical phenotype, pathophysiological mechanism, or response to treatment) and in relation to specific types of *T. gondii* parasitization. Translational research must play a key role, with the involvement of psychiatric, neurologic, immunologic, biochemical, genetic, pharmacological, and microbiological investigators, among others, offering the possibility of using new and more effective methodologies. It appears highly likely that different causal agents are responsible for schizophrenia and that the pathogenic action of a particular microorganism such as *T. gondii* would only be relevant in certain patient subgroups, endorsing the need for personalized medicine.

Author details

Antonio Sorlozano-Puerto¹ and Jose Gutierrez-Fernandez^{1,2*}

*Address all correspondence to: josegf@ugr.es

1 Department of Microbiology, School of Medicine, University of Granada and Biosanitary Research Institute ibs, Granada, Spain

2 Area of Microbiology, Granada University Hospital Complex, Spain

References

- [1] Ledgerwood LG, Ewald PW, Cochran GM. Genes, germs, and schizophrenia: an evolutionary perspective. *Perspect Biol Med.* 2003;**46**(3):317–348. doi:10.1353/pbm.2003.0038

- [2] Tsuang M. Schizophrenia: genes and environment. *Biol Psychiatry*. 2000;**47**(3):210–220. doi:10.1016/S0006-3223(99)00289-9
- [3] Dinan TG, Borre YE, Cryan JF. Genomics of schizophrenia: time to consider the gut microbiome? *Mol Psychiatry*. 2014;**19**(12):1252–1257. doi:10.1038/mp.2014.93
- [4] Arseneault L, Cannon M, Wittton J, Murray RM. Causal association between cannabis and psychosis: examination of the evidence. *Br J Psychiatry*. 2004;**184**:110–117. doi:10.1192/bjp.184.2.110
- [5] Brown AS. The risk for schizophrenia from childhood and adult infections. *Am J Psychiatry*. 2008;**165**(1):7–10. doi:10.1176/appi.ajp.2007.07101637
- [6] Fatemi SH, Folsom TD. The neurodevelopmental hypothesis of schizophrenia, revisited. *Schizophr Bull*. 2009;**35**(3):528–548. doi:10.1093/schbul/sbn187
- [7] Matheson SL, Shepherd AM, Carr VJ. How much do we know about schizophrenia and how well do we know it? Evidence from the Schizophrenia Library. *Psychol Med*. 2014;**44**(16):3387–3405. doi:10.1017/S0033291714000166
- [8] Moore TH, Zammit S, Lingford-Hughes A, Barnes TR, Jones PB, Burke M, Lewis G. Cannabis use and risk of psychotic or affective mental health outcomes: a systematic review. *Lancet*. 2007;**370**(9584):319–328. doi:10.1016/S0140-6736(07)61162-3
- [9] Sullivan PF, Kendler KS, Neale MC. Schizophrenia as a complex trait: evidence from a meta-analysis of twin studies. *Arch Gen Psychiatry*. 2003;**60**(12):1187–1192. doi:10.1001/archpsyc.60.12.1187
- [10] Rawlins S. Nonviral sexually transmitted infections. *J Obstet Gynecol Neonatal Nurs*. 2001;**30**(3):324–331. doi:10.1111/j.1552-6909.2001.tb01551.x
- [11] Arias I, Sorlozano A, Villegas E, de Dios Luna J, McKenney K, Cervilla J, Gutierrez B, Gutierrez J. Infectious agents associated with schizophrenia: a meta-analysis. *Schizophr Res*. 2012;**136**(1–3):128–136. doi:10.1016/j.schres.2011.10.026
- [12] Müller N, Riedel M, Ackenheil M, Schwarz MJ. The role of immune function in schizophrenia: an overview. *Eur Arch Psychiatry Clin Neurosci*. 1999;**249**(Suppl 4):62–68. doi:10.1007/PL00014187
- [13] Nahmias AJ, Nahmias SB, Danielsson D. The possible role of transplacentally acquired antibodies to infectious agents, with molecular mimicry to nervous system sialic acid epitopes, as causes of neuromental disorders: prevention and vaccine implications. *Clin Dev Immunol*. 2006;**13**(2–4):167–183. doi:10.1080/17402520600801745
- [14] Gomez-Sintes R, Bortolozzi A, Artigas F, Lucas JJ. Reduced striatal dopamine DAD2 receptor function in dominant-negative GSK-3 transgenic mice. *Eur Neuropsychopharmacol*. 2014;**24**(9):1524–1533. doi:10.1016/j.euroneuro.2014.07.004
- [15] Flegr J. Influence of latent *Toxoplasma* infection on human personality, physiology and morphology: pros and cons of the *Toxoplasma*-human model in studying the manipulation hypothesis. *J Exp Biol*. 2013;**216**(Pt 1):127–133. doi:10.1242/jeb.073635

- [16] Gagne SS. Toxoplasmosis. Prim Care Update Ob Gyns. 2001;8(3):122–126. doi:10.1016/S1068-607X(00)00083-4
- [17] Montoya JG, Liesenfeld O. Toxoplasmosis. Lancet. 2004;363(9425):1965–1976. doi:10.1016/S0140-6736(04)16412-X
- [18] Bossi P, Caumes E, Paris L, Dardé ML, Bricaire F. *Toxoplasma gondii*-associated Guillain-Barré syndrome in an immunocompetent patient. J Clin Microbiol. 1998;36(12):3724–3725.
- [19] McConkey GA, Martin HL, Bristow GC, Webster JP. *Toxoplasma gondii* infection and behaviour—location, location, location? J Exp Biol. 2013;216(Pt 1):113–119. doi:10.1242/jeb.074153.
- [20] Fabiani S, Pinto B, Bruschi F. Toxoplasmosis and neuropsychiatric diseases: can serological studies establish a clear relationship? Neurol Sci. 2013;34(4):417–425. doi:10.1007/s10072-012-1197-4
- [21] Jablensky AV, Kalaydjieva LV. Genetic epidemiology of schizophrenia: phenotypes, risk factors, and reproductive behavior. Am J Psychiatry. 2003;160(3):425–429. doi:10.1176/appi.ajp.160.3.425
- [22] Johnson J, Suzuki Y, Mack D, Mui E, Estes R, David C, Skamene E, Forman J, McLeod R. Genetic analysis of influences on survival following *Toxoplasma gondii* infection. Int J Parasitol. 2002;32(2):179–185. doi:10.1016/S0020-7519(01)00321-6
- [23] Bennedsen BE, Mortensen PB, Olesen AV, Henriksen TB. Congenital malformations, stillbirths, and infant deaths among children of women with schizophrenia. Arch Gen Psychiatry. 2001;58(7):674–679. doi:10.1001/archpsyc.58.7.674
- [24] Sever JL, Ellenberg JH, Ley AC, Madden DL, Fuccillo DA, Tzan NR, Edmonds DM. Toxoplasmosis: maternal and pediatric findings in 23,000 pregnancies. Pediatrics. 1988;82(2):181–192.
- [25] Torrey EF, Torrey BB, Burton-Bradley BG. The epidemiology of schizophrenia in Papua New Guinea. Am J Psychiatry. 1974;131(5):567–573.
- [26] Wallace GD, Zigas V, Gajdusek DC. Toxoplasmosis and cats in New Guinea. Am J Trop Med Hyg. 1974;23(1):8–14.
- [27] Ryan M, Hall SM, Barrett NJ, Balfour AH, Holliman RE, Joynson DH. Toxoplasmosis in England and Wales 1981 to 1992. Commun Dis Rep CDR Rev. 1995;5(2):R13–R21.
- [28] Watt DC, Szulecka TK. The effect of sex, marriage and age at first admission on the hospitalization of schizophrenics during 2 years following discharge. Psychol Med. 1979;9(3):529–539. doi:10.1017/S0033291700032098
- [29] Kruszon-Moran D, McQuillan GM. Seroprevalence of six infectious diseases among adults in the United States by race/ethnicity: data from the third national health and nutrition examination survey, 1988–94. Adv Data. 2005;352:1–9.
- [30] Regier DA, Farmer ME, Rae DS, Myers JK, Kramer M, Robins LN, George LK, Karno M, Locke BZ. One-month prevalence of mental disorders in the United States and sociode-

- mographic characteristics: the Epidemiologic Catchment Area study. *Acta Psychiatr Scand.* 1993;**88**(1):35–47. doi:10.1111/j.1600-0447.1993.tb03411.x
- [31] Torrey EF, Yolken RH. *Toxoplasma gondii* and schizophrenia. *Emerg Infect Dis.* 2003;**9**(11):1375–1380.
- [32] Gutiérrez-Fernández J, Luna Del Castillo J de D, Mañanes-González S, Carrillo-Ávila JA, Gutiérrez B, Cervilla JA, Sorlózano-Puerto A. Different presence of *Chlamydia pneumoniae*, herpes simplex virus type 1, human herpes virus 6, and *Toxoplasma gondii* in schizophrenia: meta-analysis and analytical study. *Neuropsychiatr Dis Treat.* 2015;**11**:843–852. doi:10.2147/NDT.S79285
- [33] Torrey EF, Bartko JJ, Lun ZR, Yolken RH. Antibodies to *Toxoplasma gondii* in patients with schizophrenia: a meta-analysis. *Schizophr Bull.* 2007;**33**(3):729–736. doi:10.1093/schbul/sbl050
- [34] Torrey EF, Bartko JJ, Yolken RH. *Toxoplasma gondii* and other risk factors for schizophrenia: an update. *Schizophr Bull.* 2012;**38**(3):642–647. doi:10.1093/schbul/sbs043
- [35] Sutterland AL, Fond G, Kuin A, Koeter MW, Lutter R, van Gool T, Yolken R, Szoke A, Leboyer M, de Haan L. Beyond the association. *Toxoplasma gondii* in schizophrenia, bipolar disorder, and addiction: systematic review and meta-analysis. *Acta Psychiatr Scand.* 2015;**132**(3):161–179. doi:10.1111/acps.12423
- [36] Dard C, Fricker-Hidalgo H, Brenier-Pinchart MP, Pelloux H. Relevance of and new developments in serology for toxoplasmosis. *Trends Parasitol.* 2016;**32**(6):492–506. doi:10.1016/j.pt.2016.04.001.
- [37] Ahmad D, Mehdi S, Sayed HH, Sayed AK, Shirzad G. Serological survey of *Toxoplasma gondii* in schizophrenia patients referred to Psychiatric Hospital, Sari City, Iran. *Trop Biomed.* 2010;**27**(3):476–482.
- [38] Alvarado-Esquivel C, Alanis-Quiñones OP, Arreola-Valenzuela MA, Rodríguez-Briones A, Piedra-Nevarez LJ, Duran-Morales E, Estrada-Martínez S, Martínez-García SA, Liesenfeld O. Seroepidemiology of *Toxoplasma gondii* infection in psychiatric inpatients in a northern Mexican city. *BMC Infect Dis.* 2006;**6**:178. doi:10.1186/1471-2334-6-178
- [39] Alvarado-Esquivel C, Urbina-Álvarez JD, Estrada-Martínez S, Torres-Castorena A, Molotla-de-León G, Liesenfeld O, Dubey JP. *Toxoplasma gondii* infection and schizophrenia: a case control study in a low *Toxoplasma* seroprevalence Mexican population. *Parasitol Int.* 2011;**60**(2):151–155. doi:10.1016/j.parint.2010.12.003
- [40] Amminger GP, McGorry PD, Berger GE, Wade D, Yung AR, Phillips LJ, Harrigan SM, Francey SM, Yolken RH. Antibodies to infectious agents in individuals at ultra-high risk for psychosis. *Biol Psychiatry.* 2007;**61**(10):1215–1217. doi:10.1016/j.biopsych.2006.09.034
- [41] Cetinkaya Z, Yazar S, Gecici O, Namli MN. Anti-*Toxoplasma gondii* antibodies in patients with schizophrenia—preliminary findings in a Turkish sample. *Schizophr Bull.* 2007;**33**(3):789–791. doi:10.1093/schbul/sbm021

- [42] El-Sahn AA, Shatat HZ, Ghitany EM. Seropositivity of toxoplasmosis in patients with schizophrenia. *J Egypt Public Health Assoc.* 2005;**80**(5–6):509–524.
- [43] Hamidinejat H, Ghorbanpoor M, Hosseini H, Alavi SM, Nabavi L, Jalali MH, Borojeni MP, Jafari H, Mohammadaligol S. *Toxoplasma gondii* infection in first-episode and inpatient individuals with schizophrenia. *Int J Infect Dis.* 2010;**14**(11):e978–e981. doi:10.1016/j.ijid.2010.05.018
- [44] Leweke FM, Gerth CW, Koethe D, Klosterkötter J, Ruslanova I, Krivogorsky B, Torrey EF, Yolken RH. Antibodies to infectious agents in individuals with recent onset schizophrenia. *Eur Arch Psychiatry Clin Neurosci.* 2004;**254**(1):4–8. doi:10.1007/s00406-004-0481-6
- [45] Niebuhr DW, Millikan AM, Cowan DN, Yolken R, Li Y, Weber NS. Selected infectious agents and risk of schizophrenia among US military personnel. *Am J Psychiatry.* 2008;**165**(1):99–106. doi:10.1176/appi.ajp.2007.06081254
- [46] Omar A, Bakar OC, Adam NF, Osman H, Osman A, Suleiman AH, Manaf MR, Selamat MI. Seropositivity and serointensity of *Toxoplasma gondii* antibodies and DNA among patients with schizophrenia. *Korean J Parasitol.* 2015;**53**(1):29–34. doi:10.3347/kjp.2015.53.1.29
- [47] Pedersen MG, Stevens H, Pedersen CB, Nørgaard-Pedersen B, Mortensen PB. *Toxoplasma* infection and later development of schizophrenia in mothers. *Am J Psychiatry.* 2011;**168**(8):814–821. doi:10.1176/appi.ajp.2011.10091351
- [48] Tamer GS, Dundar D, Yalug I, Caliskan S, Yazar S, Aker A. The schizophrenia and *Toxoplasma gondii* connection: infectious, immune or both? *Adv Ther.* 2008;**25**(7):703–709. doi:10.1007/s12325-008-0063-5
- [49] Tedla Y, Shibre T, Ali O, Tadele G, Woldeamanuel Y, Asrat D, Aseffa A, Mihret W, Abebe M, Alem A, Medhin G, Habte A. Serum antibodies to *Toxoplasma gondii* and *Herpesviridae* family viruses in individuals with schizophrenia and bipolar disorder: a case-control study. *Ethiop Med J.* 2011;**49**(3):211–220.
- [50] Wang HL, Wang GH, Li QY, Shu C, Jiang MS, Guo Y. Prevalence of *Toxoplasma* infection in first-episode schizophrenia and comparison between *Toxoplasma*-seropositive and *Toxoplasma*-seronegative schizophrenia. *Acta Psychiatr Scand.* 2006;**114**(1):40–48. doi:10.1111/j.1600-0447.2006.00780.x
- [51] Yolken RH, Bachmann S, Ruslanova I, Lillehoj E, Ford G, Torrey EF, Schroeder J. Antibodies to *Toxoplasma gondii* in individuals with first-episode schizophrenia. *Clin Infect Dis.* 2001;**32**(5):842–844. doi:10.1086/319221
- [52] Xiao Y, Yin J, Jiang N, Xiang M, Hao L, Lu H, Sang H, Liu X, Xu H, Ankarklev J, Lindh J, Chen Q. Seroepidemiology of human *Toxoplasma gondii* infection in China. *BMC Infect Dis.* 2010;**10**:4. doi:10.1186/1471-2334-10-4.

- [53] Nascimento FS, de Rosalmeida Dantas C, Netto MP, Mella LF, Suzuki LA, Banzato CE, Rossi CL. Prevalence of antibodies to *Toxoplasma gondii* in patients with schizophrenia and mood disorders. *Schizophr Res*. 2012;**142**(1–3):244–245. doi:10.1016/j.schres.2012.08.036
- [54] Holub D, Flegr J, Dragomirecká E, Rodriguez M, Preiss M, Novák T, Čermák J, Horáček J, Kodym P, Libiger J, Höschl C, Motlová LB. Differences in onset of disease and severity of psychopathology between toxoplasmosis-related and toxoplasmosis-unrelated schizophrenia. *Acta Psychiatr Scand*. 2013;**127**(3):227–238. doi:10.1111/acps.12031
- [55] Dickerson F, Boronow J, Stallings C, Origoni A, Yolken R. *Toxoplasma gondii* in individuals with schizophrenia: association with clinical and demographic factors and with mortality. *Schizophr Bull*. 2007;**33**(3):737–740. doi:10.1093/schbul/sbm005
- [56] Okusaga O, Langenberg P, Sleemi A, Vaswani D, Giegling I, Hartmann AM, Konte B, Friedl M, Groer MW, Yolken RH, Rujescu D, Postolache TT. *Toxoplasma gondii* antibody titers and history of suicide attempts in patients with schizophrenia. *Schizophr Res*. 2011;**133**(1–3):150–155. doi:10.1016/j.schres.2011.08.006
- [57] Rice JS, Kowal C, Volpe BT, DeGiorgio LA, Diamond B. Molecular mimicry: anti-DNA antibodies bind microbial and nonnucleic acid self-antigens. *Curr Top Microbiol Immunol*. 2005;**296**:137–151. doi:10.1007/3-540-30791-5_8
- [58] Monroe JM, Buckley PF, Miller BJ. Meta-Analysis of anti-*Toxoplasma gondii* IgM antibodies in acute psychosis. *Schizophr Bull*. 2015;**41**(4):989–998. doi:10.1093/schbul/sbu159
- [59] Mortensen PB, Nørgaard-Pedersen B, Waltoft BL, Sørensen TL, Hougaard D, Torrey EF, Yolken RH. *Toxoplasma gondii* as a risk factor for early-onset schizophrenia: analysis of filter paper blood samples obtained at birth. *Biol Psychiatry*. 2007;**61**(5):688–693. doi:10.1016/j.biopsych.2006.05.024
- [60] Brown AS, Schaefer CA, Quesenberry CP Jr, Liu L, Babulas VP, Susser ES. Maternal exposure to toxoplasmosis and risk of schizophrenia in adult offspring. *Am J Psychiatry*. 2005;**162**(4):767–773. doi:10.1176/appi.ajp.162.4.767
- [61] Blomström A, Karlsson H, Wicks S, Yang S, Yolken RH, Dalman C. Maternal antibodies to infectious agents and risk for non-affective psychoses in the offspring—a matched case-control study. *Schizophr Res*. 2012;**140**(1–3):25–30. doi:10.1016/j.schres.2012.06.035
- [62] Buka SL, Tsuang MT, Torrey EF, Klebanoff MA, Wagner RL, Yolken RH. Maternal cytokine levels during pregnancy and adult psychosis. *Brain Behav Immun*. 2001;**15**(4):411–420. doi:10.1006/brbi.2001.0644
- [63] Xiao J, Buka SL, Cannon TD, Suzuki Y, Viscidi RP, Torrey EF, Yolken RH. Serological pattern consistent with infection with type I *Toxoplasma gondii* in mothers and risk of psychosis among adult offspring. *Microbes Infect*. 2009;**11**(13):1011–1018. doi:10.1016/j.micinf.2009.07.007

- [64] Mortensen PB, Nørgaard-Pedersen B, Waltoft BL, Sørensen TL, Hougaard D, Yolken RH. Early infections of *Toxoplasma gondii* and the later development of schizophrenia. *Schizophr Bull.* 2007;**33**(3):741–744. doi:10.1093/schbul/sbm009
- [65] Lebech M, Andersen O, Christensen NC, Hertel J, Nielsen HE, Peitersen B, Rechnitzer C, Larsen SO, Nørgaard-Pedersen B, Petersen E. Feasibility of neonatal screening for toxoplasma infection in the absence of prenatal treatment. Danish Congenital Toxoplasmosis Study Group. *Lancet.* 1999;**353**(9167):1834–1837.
- [66] Pope RM. Immunoregulatory mechanisms present in the maternal circulation during pregnancy. *Baillieres Clin Rheumatol.* 1990;**4**(1):33–52. doi:10.1016/S0950-3579(05)80242-0
- [67] Brown AS, Hooton J, Schaefer CA, Zhang H, Petkova E, Babulas V, Perrin M, Gorman JM, Susser ES. Elevated maternal interleukin-8 levels and risk of schizophrenia in adult offspring. *Am J Psychiatry.* 2004;**161**(5):889–895. doi:10.1176/appi.ajp.161.5.889
- [68] Buka SL, Tsuang MT, Torrey EF, Klebanoff MA, Bernstein D, Yolken RH. Maternal infections and subsequent psychosis among offspring. *Arch Gen Psychiatry.* 2001;**58**(11):1032–1037. doi:10.1001/archpsyc.58.11.1032
- [69] Carruthers VB, Suzuki Y. Effects of *Toxoplasma gondii* infection on the brain. *Schizophr Bull.* 2007;**33**(3):745–751. doi:10.1093/schbul/sbm008
- [70] Berenreiterová M, Flegr J, Kuběna AA, Němec P. The distribution of *Toxoplasma gondii* cysts in the brain of a mouse with latent toxoplasmosis: implications for the behavioral manipulation hypothesis. *PLoS One.* 2011;**6**(12):e28925. doi:10.1371/journal.pone.0028925
- [71] Di Cristina M, Marocco D, Galizi R, Proietti C, Spaccapelo R, Crisanti A. Temporal and spatial distribution of *Toxoplasma gondii* differentiation into bradyzoites and tissue cyst formation *in vivo*. *Infect Immun.* 2008;**76**(8):3491–3501. doi:10.1128/IAI.00254-08
- [72] Hermes G, Ajioka JW, Kelly KA, Mui E, Roberts F, Kasza K, Mayr T, Kirisits MJ, Wollmann R, Ferguson DJ, Roberts CW, Hwang JH, Trendler T, Kennan RP, Suzuki Y, Reardon C, Hickey WF, Chen L, McLeod R. Neurological and behavioral abnormalities, ventricular dilatation, altered cellular functions, inflammation, and neuronal injury in brains of mice due to common, persistent, parasitic infection. *J Neuroinflamm.* 2008;**5**:48. doi:10.1186/1742-2094-5-48
- [73] Vyas A, Kim SK, Giacomini N, Boothroyd JC, Sapolsky RM. Behavioral changes induced by *Toxoplasma* infection of rodents are highly specific to aversion of cat odors. *Proc Natl Acad Sci U S A.* 2007;**104**(15):6442–6447. doi:10.1073/pnas.0608310104
- [74] Cotter DR, Pariante CM, Everall IP. Glial cell abnormalities in major psychiatric disorders: the evidence and implications. *Brain Res Bull.* 2001;**55**(5):585–595. doi:10.1016/S0361-9230(01)00527-5
- [75] Shenton ME, Dickey CC, Frumin M, McCarley RW. A review of MRI findings in schizophrenia. *Schizophr Res.* 2001;**49**(1–2):1–52. doi:10.1016/S0920-9964(01)00163-3

- [76] Horacek J, Flegr J, Tintera J, Verebova K, Spaniel F, Novak T, Brunovsky M, Bubenikova-Valesova V, Holub D, Palenicek T, Höschl C. Latent toxoplasmosis reduces gray matter density in schizophrenia but not in controls: voxel-based-morphometry (VBM) study. *World J Biol Psychiatry*. 2012;**13**(7):501–509. doi:10.3109/15622975.2011.573809
- [77] Conejero-Goldberg C, Torrey EF, Yolken RH. Herpesviruses and *Toxoplasma gondii* in orbital frontal cortex of psychiatric patients. *Schizophr Res*. 2003;**60**(1):65–69. doi:10.1016/S0920-9964(02)00160-3
- [78] Berdoy M, Webster JP, Macdonald DW. Parasite-altered behaviour: is the effect of *Toxoplasma gondii* on *Rattus norvegicus* specific? *Parasitology*. 1995;**111**(Pt 4):403–409. doi:10.1017/S0031182000065902
- [79] Berdoy M, Webster JP, Macdonald DW. Fatal attraction in rats infected with *Toxoplasma gondii*. *Proc Biol Sci*. 2000;**267**(1452):1591–1594. doi:10.1098/rspb.2000.1182
- [80] Hay J, Aitken PP, Graham DI. Toxoplasma infection and response to novelty in mice. *Z Parasitenkd*. 1984;**70**(5):575–588. doi:10.1007/BF00926588
- [81] Webster JP. The effect of *Toxoplasma gondii* and other parasites on activity levels in wild and hybrid *Rattus norvegicus*. *Parasitology*. 1994;**109**(Pt 5):583–589. doi:10.1017/S0031182000076460
- [82] Webster JP, Brunton CF, MacDonald DW. Effect of *Toxoplasma gondii* upon neophobic behaviour in wild brown rats, *Rattus norvegicus*. *Parasitology*. 1994;**109**(Pt 1):37–43. doi:10.1017/S003118200007774X
- [83] Witting PA. Learning capacity and memory of normal and *Toxoplasma*-infected laboratory rats and mice. *Z Parasitenkd*. 1979;**61**(1):29–51.
- [84] Webster JP. Rats, cats, people and parasites: the impact of latent toxoplasmosis on behaviour. *Microbes Infect*. 2001;**3**(12):1037–1045. doi:10.1016/S1286-4579(01)01459-9
- [85] Webster JP. The effect of *Toxoplasma gondii* on animal behavior: playing cat and mouse. *Schizophr Bull*. 2007;**33**(3):752–756. doi:10.1093/schbul/sbl073
- [86] Hari Dass SA, Vyas A. *Toxoplasma gondii* infection reduces predator aversion in rats through epigenetic modulation in the host medial amygdala. *Mol Ecol*. 2014;**23**(24):6114–6122. doi:10.1111/mec.12888
- [87] Daniels BP, Sestito SR, Rouse ST. An expanded task battery in the Morris water maze reveals effects of *Toxoplasma gondii* infection on learning and memory in rats. *Parasitol Int*. 2015;**64**(1):5–12. doi:10.1016/j.parint.2014.09.002
- [88] Holliman RE. Toxoplasmosis, behaviour and personality. *J Infect*. 1997;**35**(2):105–110. doi:10.1016/S0163-4453(97)91380-3
- [89] Stibbs HH. Changes in brain concentrations of catecholamines and indoleamines in *Toxoplasma gondii* infected mice. *Ann Trop Med Parasitol*. 1985;**79**(2):153–157.

- [90] Tomasik J, Schultz TL, Kluge W, Yolken RH, Bahn S, Carruthers VB. Shared immune and repair markers during experimental *Toxoplasma* chronic brain infection and schizophrenia. *Schizophr Bull.* 2016;**42**(2):386–395. doi:10.1093/schbul/sbv134
- [91] Flegr J, Zitková S, Kodym P, Frynta D. Induction of changes in human behaviour by the parasitic protozoan *Toxoplasma gondii*. *Parasitology.* 1996;**113**(Pt 1):49–54. doi:10.1017/S0031182000066269
- [92] Flegr J, Kodym P, Tolarová V. Correlation of duration of latent *Toxoplasma gondii* infection with personality changes in women. *Biol Psychol.* 2000;**53**(1):57–68. doi:10.1016/S0301-0511(00)00034-X
- [93] Havlíček J, Gasová ZG, Smith AP, Zvára K, Flegr J. Decrease of psychomotor performance in subjects with latent ‘asymptomatic’ toxoplasmosis. *Parasitology.* 2001;**122**(Pt 5):515–520. doi:10.1017/S0031182001007624
- [94] Jones-Brando L, Torrey EF, Yolken R. Drugs used in the treatment of schizophrenia and bipolar disorder inhibit the replication of *Toxoplasma gondii*. *Schizophr Res.* 2003;**62**(3):237–244. doi:10.1016/S0920-9964(02)00357-2
- [95] Pezzella N, Bouchot A, Bonhomme A, Pingret L, Klein C, Burlet H, Balossier G, Bonhomme P, Pinon JM. Involvement of calcium and calmodulin in *Toxoplasma gondii* tachyzoite invasion. *Eur J Cell Biol.* 1997;**74**(1):92–101.
- [96] Webster JP, Lamberton PH, Donnelly CA, Torrey EF. Parasites as causative agents of human affective disorders? The impact of anti-psychotic, mood-stabilizer and anti-parasite medication on *Toxoplasma gondii*'s ability to alter host behaviour. *Proc Biol Sci.* 2006;**273**(1589):1023–1030. doi:10.1098/rspb.2005.3413
- [97] Flegr J, Preiss M, Klose J, Havlíček J, Vitáková M, Kodym P. Decreased level of psychobiological factor novelty seeking and lower intelligence in men latently infected with the protozoan parasite *Toxoplasma gondii*. Dopamine, a missing link between schizophrenia and toxoplasmosis? *Biol Psychol.* 2003;**63**(3):253–268. doi:10.1016/S0301-0511(03)00075-9
- [98] Fond G, Macgregor A, Tamouza R, Hamdani N, Meary A, Leboyer M, Dubremetz JF. Comparative analysis of anti-toxoplasmic activity of antipsychotic drugs and valproate. *Eur Arch Psychiatry Clin Neurosci.* 2014;**264**(2):179–183. doi:10.1007/s00406-013-0413-4
- [99] Goodwin DG, Strobl JS, Lindsay DS. Evaluation of five antischizophrenic agents against *Toxoplasma gondii* in human cell cultures. *J Parasitol.* 2011;**97**(1):148–151. doi:10.1645/GE-2536.1
- [100] Dickerson FB, Stallings CR, Boronow JJ, Origoni AE, Yolken RH. A double-blind trial of adjunctive azithromycin in individuals with schizophrenia who are seropositive for *Toxoplasma gondii*. *Schizophr Res.* 2009;**112**(1–3):198–199. doi:10.1016/j.schres.2009.05.005

- [101] Shibre T, Alem A, Abdulahi A, Araya M, Beyero T, Medhin G, Deyassa N, Negash A, Nigatu A, Kebede D, Fekadu A. Trimethoprim as adjuvant treatment in schizophrenia: a double-blind, randomized, placebo-controlled clinical trial. *Schizophr Bull*. 2010;**36**(4):846–851. doi:10.1093/schbul/sbn191
- [102] Wei HX, Wei SS, Lindsay DS, Peng HJ. A systematic review and meta-analysis of the efficacy of anti-*Toxoplasma gondii* medicines in humans. *PLoS One*. 2015;**10**(9):e0138204. doi:10.1371/journal.pone.0138204
- [103] da Silva RC, Langoni H. *Toxoplasma gondii*: host-parasite interaction and behavior manipulation. *Parasitol Res*. 2009;**105**(4):893–898. doi:10.1007/s00436-009-1526-6
- [104] Khandaker GM, Cousins L, Deakin J, Lennox BR, Yolken R, Jones PB. Inflammation and immunity in schizophrenia: implications for pathophysiology and treatment. *Lancet Psychiatry*. 2015;**2**(3):258–270. doi:10.1016/S2215-0366(14)00122-9
- [105] Doorduyn J, de Vries EF, Willemsen AT, de Groot JC, Dierckx RA, Klein HC. Neuroinflammation in schizophrenia-related psychosis: a PET study. *J Nucl Med*. 2009;**50**(11):1801–1807. doi:10.2967/jnumed.109.066647
- [106] Lin G, Zhang H, Sun F, Lu Z, Reed-Maldonado A, Lee YC, Wang G, Banie L, Lue TF. Brain-derived neurotrophic factor promotes nerve regeneration by activating the JAK/STAT pathway in Schwann cells. *Transl Androl Urol*. 2016;**5**(2):167–175. doi:10.21037/tau.2016.02.03
- [107] Qin H, Buckley JA, Li X, Liu Y, Fox TH 3rd, Meares GP, Yu H, Yan Z, Harms AS, Li Y, Standaert DG, Benveniste EN. Inhibition of the JAK/STAT pathway protects against α -synuclein-induced neuroinflammation and dopaminergic neurodegeneration. *J Neurosci*. 2016;**36**(18):5144–5159. doi:10.1523/JNEUROSCI.4658-15.2016
- [108] Gavrilescu LC, Butcher BA, Del Rio L, Taylor GA, Denkers EY. STAT1 is essential for antimicrobial effector function but dispensable for gamma interferon production during *Toxoplasma gondii* infection. *Infect Immun*. 2004;**72**(3):1257–1264. doi:10.1128/IAI.72.3.1257-1264.2004
- [109] Kim SK, Fouts AE, Boothroyd JC. *Toxoplasma gondii* dysregulates IFN-gamma-inducible gene expression in human fibroblasts: insights from a genome-wide transcriptional profiling. *J Immunol*. 2007;**178**(8):5154–5165.
- [110] Sun GY, Horrocks LA, Farooqui AA. The roles of NADPH oxidase and phospholipases A2 in oxidative and inflammatory responses in neurodegenerative diseases. *J Neurochem*. 2007;**103**(1):1–16.
- [111] Elsheikha HM, Büsselberg D, Zhu XQ. The known and missing links between *Toxoplasma gondii* and schizophrenia. *Metab Brain Dis*. 2016;**31**(4):749–759. doi:10.1007/s11011-016-9822-1
- [112] Vasconcelos AR, Yshii LM, Viel TA, Buck HS, Mattson MP, Scavone C, Kawamoto EM. Intermittent fasting attenuates lipopolysaccharide-induced neuroinflammation and memory impairment. *J Neuroinflamm*. 2014;**11**:85. doi:10.1186/1742-2094-11-85

- [113] Zindler E, Zipp F. Neuronal injury in chronic CNS inflammation. *Best Pract Res Clin Anaesthesiol.* 2010;**24**(4):551–562. doi:10.1016/j.bpa.2010.11.001
- [114] Prandovszky E, Gaskell E, Martin H, Dubey JP, Webster JP, McConkey GA. The neurotropic parasite *Toxoplasma gondii* increases dopamine metabolism. *PLoS One.* 2011;**6**(9):e232866. doi:10.1371/journal.pone.0023866
- [115] Skallová A, Kodym P, Frynta D, Flegr J. The role of dopamine in *Toxoplasma*-induced behavioural alterations in mice: an ethological and ethopharmacological study. *Parasitology.* 2006;**133**(Pt 5):525–535. doi:10.1017/S0031182006000886
- [116] Gaskell EA, Smith JE, Pinney JW, Westhead DR, McConkey GA. A unique dual activity amino acid hydroxylase in *Toxoplasma gondii*. *PLoS One.* 2009;**4**(3):e4801. doi:10.1371/journal.pone.0004801
- [117] Martin HL, Alsaady I, Howell G, Prandovszky E, Peers C, Robinson P, McConkey GA. Effect of parasitic infection on dopamine biosynthesis in dopaminergic cells. *Neuroscience.* 2015;**306**:50–62. doi:10.1016/j.neuroscience.2015.08.005
- [118] Strobl JS, Goodwin DG, Rzigalinski BA, Lindsay DS. Dopamine stimulates propagation of *Toxoplasma gondii* tachyzoites in human fibroblast and primary neonatal rat astrocyte cell cultures. *J Parasitol.* 2012;**98**(6):1296–1299. doi:10.1645/GE-2760.1
- [119] Stiles J, Jernigan TL. The basics of brain development. *Neuropsychol Rev.* 2010;**20**(4):327–348. doi:10.1007/s11065-010-9148-4
- [120] Paus T. Mapping brain maturation and cognitive development during adolescence. *Trends Cogn Sci.* 2005;**9**(2):60–88. doi:10.1016/j.tics.2004.12.008
- [121] Nilsson LK, Linderholm KR, Engberg G, Paulson L, Blennow K, Lindström LH, Nordin C, Karanti A, Persson P, Erhardt S. Elevated levels of kynurenic acid in the cerebrospinal fluid of male patients with schizophrenia. *Schizophr Res.* 2005;**80**(2–3):315–322. doi:10.1016/j.schres.2005.07.013
- [122] Erhardt S, Schwieler L, Nilsson L, Linderholm K, Engberg G. The kynurenic acid hypothesis of schizophrenia. *Physiol Behav.* 2007;**92**(1–2):203–209. doi:10.1016/j.physbeh.2007.05.025
- [123] Silva NM, Rodrigues CV, Santoro MM, Reis LF, Alvarez-Leite JI, Gazzinelli RT. Expression of indoleamine 2,3-dioxygenase, tryptophan degradation, and kynurenine formation during *in vivo* infection with *Toxoplasma gondii*: induction by endogenous gamma interferon and requirement of interferon regulatory factor 1. *Infect Immun.* 2002;**70**(2):859–868. doi:10.1128/IAI.70.2.859-868.2002
- [124] Miller CM, Boulter NR, Ikin RJ, Smith NC. The immunobiology of the innate response to *Toxoplasma gondii*. *Int J Parasitol.* 2009;**39**(1):23–39. doi:10.1016/j.ijpara.2008.08.002
- [125] Lidberg L, Belfrage H, Bertilsson L, Evenden MM, Asberg M. Suicide attempts and impulse control disorder are related to low cerebrospinal fluid 5-HIAA in mentally disordered violent offenders. *Acta Psychiatr Scand.* 2000;**101**(5):395–402.

- [126] Widner B, Laich A, Sperner-Unterweger B, Ledochowski M, Fuchs D. Neopterin production, tryptophan degradation, and mental depression—what is the link? *Brain Behav Immun*. 2002;**16**(5):590–595. doi:10.1016/S0889-1591(02)00006-5
- [127] Buddhala C, Hsu CC, Wu JY. A novel mechanism for GABA synthesis and packaging into synaptic vesicles. *Neurochem Int*. 2009;**55**(1–3):9–12. doi:10.1016/j.neuint.2009.01.020
- [128] Barragan A, Weidner JM, Jin Z, Korpi ER, Birnir B. GABAergic signalling in the immune system. *Acta Physiol (Oxf)*. 2015;**213**(4):819–827. doi:10.1111/apha.12467
- [129] Simpson MD, Slater P, Deakin JF, Royston MC, Skan WJ. Reduced GABA uptake sites in the temporal lobe in schizophrenia. *Neurosci Lett*. 1989;**107**(1–3):211–215. doi:10.1016/0304-3940(89)90819-7
- [130] Reynolds GP, Czudek C, Andrews HB. Deficit and hemispheric asymmetry of GABA uptake sites in the hippocampus in schizophrenia. *Biol Psychiatry* 1990;**27**(9):1038–1044. doi:10.1016/0006-3223(90)90039-5
- [131] Lewis DA, Pierri JN, Volk DW, Melchitzky DS, Woo TU. Altered GABA neurotransmission and prefrontal cortical dysfunction in schizophrenia. *Biol Psychiatry*. 1999; **46**(5):616–626. doi:10.1016/S0006-3223(99)00061-X
- [132] Volk D, Austin M, Pierri J, Sampson A, Lewis D. GABA transporter-1 mRNA in the prefrontal cortex in schizophrenia: decreased expression in a subset of neurons. *Am J Psychiatry*. 2001;**158**(2):256–265. doi:10.1176/appi.ajp.158.2.256
- [133] Woo TU, Walsh JP, Benes FM. Density of glutamic acid decarboxylase 67 messenger RNA-containing neurons that express the *N*-methyl-D-aspartate receptor subunit NR2A in the anterior cingulate cortex in schizophrenia and bipolar disorder. *Arch Gen Psychiatry*. 2004;**61**(7):649–657.
- [134] Volk DW, Pierri JN, Fritschy JM, Auh S, Sampson AR, Lewis DA. Reciprocal alterations in pre- and postsynaptic inhibitory markers at chandelier cell inputs to pyramidal neurons in schizophrenia. *Cereb Cortex*. 2002;**12**(10):1063–1070. doi:10.1093/cercor/12.10.1063
- [135] Fatemi SH, Folsom TD, Thuras PD. Deficits in GABA(B) receptor system in schizophrenia and mood disorders: a postmortem study. *Schizophr Res*. 2011;**128**(1–3):37–43. doi:10.1016/j.schres.2010.12.025
- [136] Olofsson PS, Rosas-Ballina M, Levine YA, Tracey KJ. Rethinking inflammation: neural circuits in the regulation of immunity. *Immunol Rev*. 2012;**248**(1):188–204. doi:10.1111/j.1600-065X.2012.01138.x
- [137] Fuks JM, Arrighi RB, Weidner JM, Kumar Mendu S, Jin Z, Wallin RP, Rethi B, Birnir B, Barragan A. GABAergic signaling is linked to a hypermigratory phenotype in dendritic cells infected by *Toxoplasma gondii*. *PLoS Pathog*. 2012;**8**(12):e1003051. doi:10.1371/journal.ppat.1003051

- [138] Weidner JM, Kanatani S, Hernandez-Castaneda MA, Fuks JM, Rethi B, Wallin RP, Barragan A. Rapid cytoskeleton remodelling in dendritic cells following invasion by *Toxoplasma gondii* coincides with the onset of a hypermigratory phenotype. *Cell Microbiol.* 2013;**15**(10):1735–1752. doi:10.1111/cmi.12145
- [139] Brooks JM, Carrillo GL, Su J, Lindsay DS, Fox MA, Blader IJ. *Toxoplasma gondii* infections alter GABAergic synapses and signaling in the central nervous system. *MBio.* 2015;**6**(6):e01428–15. doi:10.1128/mBio.01428-15
- [140] Xiao J, Li Y, Prandovszky E, Karuppagounder SS, Talbot CC Jr, Dawson VL, Dawson TM, Yolken RH. MicroRNA-132 dysregulation in *Toxoplasma gondii* infection has implications for dopamine signaling pathway. *Neuroscience.* 2014;**268**:128–138. doi:10.1016/j.neuroscience.2014.03.015
- [141] Dalmau J, Gleichman AJ, Hughes EG, Rossi JE, Peng X, Lai M, Dessain SK, Rosenfeld MR, Balice-Gordon R, Lynch DR. Anti-NMDA-receptor encephalitis: case series and analysis of the effects of antibodies. *Lancet Neurol.* 2008;**7**(12):1091–1098. doi:10.1016/S1474-4422(08)70224-2
- [142] Irani SR, Bera K, Waters P, Zuliani L, Maxwell S, Zandi MS, Friese MA, Galea I, Kullmann DM, Beeson D, Lang B, Bien CG, Vincent A. N-methyl-D-aspartate antibody encephalitis: temporal progression of clinical and paraclinical observations in a predominantly non-paraneoplastic disorder of both sexes. *Brain.* 2010;**133**(Pt 6):1655–1667. doi:10.1093/brain/awq113
- [143] Dalmau J, Tüzün E, Wu HY, Masjuan J, Rossi JE, Voloschin A, Baehring JM, Shimazaki H, Koide R, King D, Mason W, Sansing LH, Dichter MA, Rosenfeld MR, Lynch DR. Paraneoplastic anti-N-methyl-D-aspartate receptor encephalitis associated with ovarian teratoma. *Ann Neurol.* 2007;**61**(1):25–36. doi:10.1002/ana.21050
- [144] Fawcett RG. Acute psychosis associated with anti-NMDA-receptor antibodies and bilateral ovarian teratomas: a case report. *J Clin Psychiatry.* 2010;**71**(4):504. doi:10.4088/JCP.09l05609yel
- [145] Benros ME, Mortensen PB, Eaton WW. Autoimmune diseases and infections as risk factors for schizophrenia. *Ann N Y Acad Sci.* 2012;**1262**:56–66. doi:10.1111/j.1749-6632.2012.06638.x
- [146] DeGiorgio LA, Konstantinov KN, Lee SC, Hardin JA, Volpe BT, Diamond B. A subset of lupus anti-DNA antibodies cross-reacts with the NR2 glutamate receptor in systemic lupus erythematosus. *Nat Med.* 2001;**7**(11):1189–1193.
- [147] Pearlman DM, Najjar S. Meta-analysis of the association between N-methyl-D-aspartate receptor antibodies and schizophrenia, schizoaffective disorder, bipolar disorder, and major depressive disorder. *Schizophr Res.* 2014;**157**(1–3):249–258. doi:10.1016/j.schres.2014.05.001
- [148] Smyth AM, Lawrie SM. The neuroimmunology of schizophrenia. *Clin Psychopharmacol Neurosci.* 2013;**11**(3):107–117. doi:10.9758/cpn.2013.11.3.107

- [149] Deakin J, Lennox BR, Zandi MS. Antibodies to the *N*-methyl-*D*-aspartate receptor and other synaptic proteins in psychosis. *Biol Psychiatry*. 2014;**75**(4):284–291. doi:10.1016/j.biopsych.2013.07.018
- [150] Steiner J, Walter M, Glanz W, Sarnyai Z, Bernstein HG, Vielhaber S, Kästner A, Skalej M, Jordan W, Schiltz K, Klingbeil C, Wandinger KP, Bogerts B, Stoecker W. Increased prevalence of diverse *N*-methyl-*D*-aspartate glutamate receptor antibodies in patients with an initial diagnosis of schizophrenia: specific relevance of IgG NR1a antibodies for distinction from *N*-methyl-*D*-aspartate glutamate receptor encephalitis. *JAMA Psychiatry*. 2013;**70**(3):271–278. doi:10.1001/2013.jamapsychiatry.86
- [151] Zandi MS, Irani SR, Lang B, Waters P, Jones PB, McKenna P, Coles AJ, Vincent A, Lennox BR. Disease-relevant autoantibodies in first episode schizophrenia. *J Neurol*. 2011;**258**(4):686–688. doi:10.1007/s00415-010-5788-9
- [152] Masdeu JC, González-Pinto A, Matute C, Ruiz De Azúa S, Palomino A, De Leon J, Berman KF, Dalmau J. Serum IgG antibodies against the NR1 subunit of the NMDA receptor not detected in schizophrenia. *Am J Psychiatry*. 2012;**169**(10):1120–1121. doi:10.1176/appi.ajp.2012.12050646
- [153] Rhoads J, Guirgis H, McKnight C, Duchemin AM. Lack of anti-NMDA receptor autoantibodies in the serum of subjects with schizophrenia. *Schizophr Res*. 2011;**129**(2–3):213–214. doi:10.1016/j.schres.2010.12.018
- [154] Tsutsui K, Kanbayashi T, Tanaka K, Boku S, Ito W, Tokunaga J, Mori A, Hishikawa Y, Shimizu T, Nishino S. Anti-NMDA-receptor antibody detected in encephalitis, schizophrenia, and narcolepsy with psychotic features. *BMC Psychiatry*. 2012;**12**:37. doi:10.1186/1471-244X-12-37
- [155] Coyle JT. NMDA receptor and schizophrenia: a brief history. *Schizophr Bull*. 2012;**38**(5):920–926. doi:10.1093/schbul/sbs076
- [156] Belforte JE, Zsiros V, Sklar ER, Jiang Z, Yu G, Li Y, Quinlan EM, Nakazawa K. Postnatal NMDA receptor ablation in corticolimbic interneurons confers schizophrenia-like phenotypes. *Nat Neurosci*. 2010;**13**(1):76–83. doi:10.1038/nn.2447
- [157] Timms AE, Dorschner MO, Wechsler J, Choi KY, Kirkwood R, Girirajan S, Baker C, Eichler EE, Korvatska O, Roche KW, Horwitz MS, Tsuang DW. Support for the *N*-methyl-*D*-aspartate receptor hypofunction hypothesis of schizophrenia from exome sequencing in multiplex families. *JAMA Psychiatry*. 2013;**70**(6):582–590. doi:10.1001/jamapsychiatry.2013.1195
- [158] Pilowsky LS, Bressan RA, Stone JM, Erlandsson K, Mulligan RS, Krystal JH, Ell PJ. First *in vivo* evidence of an NMDA receptor deficit in medication-free schizophrenic patients. *Mol Psychiatry*. 2006;**11**(2):118–119. doi:10.1038/sj.mp.4001751
- [159] Pomarol-Clotet E, Honey GD, Murray GK, Corlett PR, Absalom AR, Lee M, McKenna PJ, Bullmore ET, Fletcher PC. Psychological effects of ketamine in healthy volunteers. Phenomenological study. *Br J Psychiatry*. 2006;**189**:173–179. doi:10.1192/bjp.bp.105.015263

- [160] Javitt DC. Glutamate and schizophrenia: phencyclidine, *N*-methyl-*D*-aspartate receptors, and dopamine-glutamate interactions. *Int Rev Neurobiol*. 2007;**78**:69–108.
- [161] Fromer M, Pocklington AJ, Kavanagh DH, Williams HJ, Dwyer S, Gormley P, Georgieva L, Rees E, Palta P, Ruderfer DM, Carrera N, Humphreys I, Johnson JS, Roussos P, Barker DD, Banks E, Milanova V, Grant SG, Hannon E, Rose SA, Chambert K, Mahajan M, Scolnick EM, Moran JL, Kirov G, Palotie A, McCarroll SA, Holmans P, Sklar P, Owen MJ, Purcell SM, O'Donovan MC. *De novo* mutations in schizophrenia implicate synaptic networks. *Nature*. 2014;**506**(7487):179–184. doi:10.1038/nature12929
- [162] Prüss H, Finke C, Höltje M, Hofmann J, Klingbeil C, Probst C, Borowski K, Ahnert-Hilger G, Harms L, Schwab JM, Ploner CJ, Komorowski L, Stoecker W, Dalmau J, Wandinger KP. *N*-methyl-*D*-aspartate receptor antibodies in herpes simplex encephalitis. *Ann Neurol*. 2012;**72**(6):902–911. doi:10.1002/ana.23689
- [163] Hammer C, Stepniak B, Schneider A, Papiol S, Tantra M, Begemann M, Sirén AL, Pardo LA, Sperling S, Mohd Jofrry S, Gurvich A, Jensen N, Ostmeier K, Lühder F, Probst C, Martens H, Gillis M, Saher G, Assogna F, Spalletta G, Stöcker W, Schulz TF, Nave KA, Ehrenreich H. Neuropsychiatric disease relevance of circulating anti-NMDA receptor autoantibodies depends on blood-brain barrier integrity. *Mol Psychiatry*. 2014;**19**(10):1143–1149. doi:10.1038/mp.2013.110
- [164] Kannan G, Crawford JA, Yang C, Gressitt KL, Ihenatu C, Krasnova IN, Cadet JL, Yolken RH, Severance EG, Pletnikov MV. Anti-NMDA receptor autoantibodies and associated neurobehavioral pathology in mice are dependent on age of first exposure to *Toxoplasma gondii*. *Neurobiol Dis*. 2016;**91**:307–314. doi:10.1016/j.nbd.2016.03.005
- [165] Freedman R. Schizophrenia. *N Engl J Med*. 2003;**349**(18):1738–1749.
- [166] Cardno AG, Marshall EJ, Coid B, Macdonald AM, Ribchester TR, Davies NJ, Venturi P, Jones LA, Lewis SW, Sham PC, Gottesman II, Farmer AE, McGuffin P, Reveley AM, Murray RM. Heritability estimates for psychotic disorders: the Maudsley twin psychosis series. *Arch Gen Psychiatry*. 1999;**56**(2):162–168. doi:10.1001/archpsyc.56.2.162
- [167] Fatemi SH, Pearce DA, Brooks AI, Sidwell RW. Prenatal viral infection in mouse causes differential expression of genes in brains of mouse progeny: a potential animal model for schizophrenia and autism. *Synapse*. 2005;**57**(2):91–99. doi:10.1002/syn.20162
- [168] Harrison PJ, Owen MJ. Genes for schizophrenia? Recent findings and their pathophysiological implications. *Lancet*. 2003;**361**(9355):417–419. doi:10.1016/S0140-6736(03)12379-3
- [169] Carter CJ. Schizophrenia: a pathogenetic autoimmune disease caused by viruses and pathogens and dependent on genes. *J Pathog*. 2011;**2011**:128318. doi:10.4061/2011/128318
- [170] Tomonaga K. Virus-induced neurobehavioral disorders: mechanisms and implications. *Trends Mol Med*. 2004;**10**(2):71–77. doi:10.1016/j.molmed.2003.12.001
- [171] Harrison PJ, Law AJ. Neuregulin 1 and schizophrenia: Genetics, gene expression, and neurobiology. *Biol Psychiatry*. 2006;**60**(2):132–140. doi:10.1016/j.biopsych.2005.11.002

- [172] Rico B, Marín O. Neuregulin signaling, cortical circuitry development and schizophrenia. *Curr Opin Genet Dev.* 2011;**21**(3):262–270. doi:10.1016/j.gde.2010.12.010
- [173] Chen CY, Lu RB, Yeh YW, Shih MC, Huang SY. Association study of catechol-O-methyltransferase gene polymorphisms with schizophrenia and psychopathological symptoms in Han Chinese. *Genes Brain Behav.* 2011;**10**(3):316–324. doi:10.1111/j.1601-183X.2010.00670.x
- [174] Willis A, Bender HU, Steel G, Valle D. PRODH variants and risk for schizophrenia. *Amino Acids.* 2008;**35**(4):673–679. doi:10.1007/s00726-008-0111-0
- [175] Voisey J, Swagell CD, Hughes IP, Connor JP, Lawford BR, Young RM, Morris CP. A polymorphism in the dysbindin gene (DTNBP1) associated with multiple psychiatric disorders including schizophrenia. *Behav Brain Funct.* 2010;**6**(1):41. doi:10.1186/1744-9081-6-41
- [176] Ding L, Hegde AN. Expression of RGS4 splice variants in dorsolateral prefrontal cortex of schizophrenic and bipolar disorder patients. *Biol Psychiatry.* 2009;**65**(6):541–545. doi:10.1016/j.biopsych.2008.10.026
- [177] Ritsner M, Modai I, Ziv H, Amir S, Halperin T, Weizman A, Navon R. An association of CAG repeats at the KCNN3 locus with symptom dimensions of schizophrenia. *Biol Psychiatry.* 2002;**51**(10):788–794. doi:10.1016/S0006-3223(01)01348-8
- [178] Madeira C, Freitas ME, Vargas-Lopes C, Wolosker H, Panizzutti R. Increased brain D-amino acid oxidase (DAAO) activity in schizophrenia. *Schizophr Res.* 2008;**101**(1–3):76–83. doi:10.1016/j.schres.2008.02.002
- [179] Carter CJ. Schizophrenia susceptibility genes directly implicated in the life cycles of pathogens: cytomegalovirus, influenza, herpes simplex, rubella, and *Toxoplasma gondii*. *Schizophr Bull.* 2009;**35**(6):1163–1182. doi:10.1093/schbul/sbn054
- [180] Carter CJ. Toxoplasmosis and polygenic disease susceptibility genes: extensive *Toxoplasma gondii* host/pathogen interactome enrichment in nine psychiatric or neurological disorders. *J Pathog.* 2013;**2013**:965046. doi:10.1155/2013/965046
- [181] Pearce BD. Schizophrenia and viral infection during neurodevelopment: a focus on mechanisms. *Mol Psychiatry.* 2001;**6**(6):634–646. doi:10.1038/sj.mp.4000956
- [182] Tsuang MT, Stone WS, Faraone SV. Genes, environment and schizophrenia. *Br J Psychiatry Suppl.* 2001;**178**(40):s18–s24. doi:10.1192/bjp.178.40.s18
- [183] Khan IA, Murphy PM, Casciotti L, Schwartzman JD, Collins J, Gao JL, Yeaman GR. Mice lacking the chemokine receptor CCR1 show increased susceptibility to *Toxoplasma gondii* infection. *J Immunol.* 2001;**166**(3):1930–1937. doi:10.4049/jimmunol.166.3.1930
- [184] Yap GS, Ortmann R, Shevach E, Sher A. A heritable defect in IL-12 signaling in B10.Q/J mice. II. Effect on acute resistance to *Toxoplasma gondii* and rescue by IL-18 treatment. *J Immunol.* 2001;**166**(9):5720–5725. doi:10.4049/jimmunol.166.9.5720

- [185] Mack DG, Johnson JJ, Roberts F, Roberts CW, Estes RG, David C, Grumet FC, McLeod R. HLA-class II genes modify outcome of *Toxoplasma gondii* infection. *Int J Parasitol*. 1999;**29**(9):1351–1358. doi:10.1016/S0020-7519(99)00152-6
- [186] Suzuki Y, Wong SY, Grumet FC, Fessel J, Montoya JG, Zolopa AR, Portmore A, Schumacher-Perdreau F, Schrappe M, Köppen S, Ruf B, Brown BW, Remington JS. Evidence for genetic regulation of susceptibility to toxoplasmic encephalitis in AIDS patients. *J Infect Dis*. 1996;**173**(1):265–268. doi:10.1093/infdis/173.1.265
- [187] Avramopoulos D, Pearce BD, McGrath J, Wolyniec P, Wang R, Eckart N, Hatzimanolis A, Goes FS, Nestadt G, Mulle J, Coneely K, Hopkins M, Ruczinski I, Yolken R, Pulver AE. Infection and inflammation in schizophrenia and bipolar disorder: a genome wide study for interactions with genetic variation. *PLoS One*. 2015;**10**(3):e0116696. doi:10.1371/journal.pone.0116696
- [188] Børghlum AD, Demontis D, Grove J, Pallesen J, Hollegaard MV, Pedersen CB, Hedemand A, Mattheisen M, Uitterlinden A, Nyegaard M, Ørntoft T, Wiuf C, Didriksen M, Nordentoft M, Nöthen MM, Rietschel M, Ophoff RA, Cichon S, Yolken RH, Hougaard DM, Mortensen PB, Mors O. Genome-wide study of association and interaction with maternal cytomegalovirus infection suggests new schizophrenia loci. *Mol Psychiatry*. 2014;**19**(3):325–333. doi:10.1038/mp.2013.2
- [189] Kannan G, Sawa A, Pletnikov MV. Mouse models of gene-environment interactions in schizophrenia. *Neurobiol Dis*. 2013;**57**:5–11. doi:10.1016/j.nbd.2013.05.012
- [190] Kim T, Kim HJ, Park JK, Kim JW, Chung JH. Association between polymorphisms of arachidonate 12-lipoxygenase (ALOX12) and schizophrenia in a Korean population. *Behav Brain Funct*. 2010;**6**:44. doi:10.1186/1744-9081-6-44
- [191] Witola WH, Liu SR, Montpetit A, Welti R, Hypolite M, Roth M, Zhou Y, Mui E, Cesbron-Delauw MF, Fournie GJ, Cavailles P, Bisanz C, Boyer K, Withers S, Noble AG, Swisher CN, Heydemann PT, Rabiah P, Muench SP, McLeod R. ALOX12 in human toxoplasmosis. *Infect Immun*. 2014;**82**(7):2670–2679. doi:10.1128/IAI.01505-13
- [192] Lang F, Strutz-Seebohm N, Seebohm G, Lang UE. Significance of SGK1 in the regulation of neuronal function. *J Physiol*. 2010;**588**(Pt 18):3349–3354. doi:10.1113/jphysiol.2010.190926
- [193] Anacker C, Cattaneo A, Musaelyan K, Zunszain PA, Horowitz M, Molteni R, Luoni A, Calabrese F, Tansey K, Gennarelli M, Thuret S, Price J, Uher R, Riva MA, Pariante CM. Role for the kinase SGK1 in stress, depression, and glucocorticoid effects on hippocampal neurogenesis. *Proc Natl Acad Sci U S A*. 2013;**110**(21):8708–8713. doi:10.1073/pnas.1300886110
- [194] Kadkhodaei B, Ito T, Joodmardi E, Mattsson B, Rouillard C, Carta M, Muramatsu S, Sumi-Ichinose C, Nomura T, Metzger D, Chambon P, Lindqvist E, Larsson NG, Olson L, Björklund A, Ichinose H, Perlmann T. Nurr1 is required for maintenance of matur-

- ing and adult midbrain dopamine neurons. *J Neurosci.* 2009;**29**(50):15923–15932. doi:10.1523/JNEUROSCI.3910-09.2009
- [195] Eells JB, Varela-Stokes A, Guo-Ross SX, Kummari E, Smith HM, Cox E, Lindsay DS. Chronic *Toxoplasma gondii* in Nurr1-null heterozygous mice exacerbates elevated open field activity. *PLoS One.* 2015;**10**(4):e0119280. doi:10.1371/journal.pone.0119280
- [196] Mullonkal CJ, Toledo-Pereyra LH. Akt in ischemia and reperfusion. *J Invest Surg.* 2007;**20**(3):195–203. doi:10.1080/08941930701366471
- [197] Tan HY, Nicodemus KK, Chen Q, Li Z, Brooke JK, Honea R, Kolachana BS, Straub RE, Meyer-Lindenberg A, Sei Y, Mattay VS, Callicott JH, Weinberger DR. Genetic variation in AKT1 is linked to dopamine-associated prefrontal cortical structure and function in humans. *J Clin Invest.* 2008;**118**(6):2200–2208. doi:10.1172/JCI34725
- [198] Fellerhoff B, Laumbacher B, Mueller N, Gu S, Wank R. Associations between *Chlamydomphila* infections, schizophrenia and risk of HLA-A10. *Mol Psychiatry.* 2007;**12**(3):264–272. doi:10.1038/sj.mp.4001925
- [199] Kim JJ, Shirts BH, Dayal M, Bacanu SA, Wood J, Xie W, Zhang X, Chowdari KV, Yolken R, Devlin B, Nimgaonkar VL. Are exposure to cytomegalovirus and genetic variation on chromosome 6p joint risk factors for schizophrenia? *Ann Med.* 2007;**39**(2):145–153. doi:10.1080/07853890601083808
- [200] Prasad KM, Bamne MN, Shirts BH, Goradia D, Mannali V, Pancholi KM, Xue B, McClain L, Yolken RH, Keshavan MS, Nimgaonkar VL. Grey matter changes associated with host genetic variation and exposure to herpes simplex virus 1 (HSV1) in first episode schizophrenia. *Schizophr Res.* 2010;**118**(1–3):232–239. doi:10.1016/j.schres.2010.01.007
- [201] Shirts BH, Kim JJ, Reich S, Dickerson FB, Yolken RH, Devlin B, Nimgaonkar VL. Polymorphisms in MICB are associated with human herpes virus seropositivity and schizophrenia risk. *Schizophr Res.* 2007;**94**(1–3):342–353. doi:10.1016/j.schres.2007.04.021
- [202] Kusbeci OY, Miman O, Yaman M, Aktepe OC, Yazar S. Could *Toxoplasma gondii* have any role in Alzheimer disease? *Alzheimer Dis Assoc Disord.* 2011;**25**(1):1–3. doi:10.1097/WAD.0b013e3181f73bc2
- [203] Miman O, Kusbeci OY, Aktepe OC, Cetinkaya Z. The probable relation between *Toxoplasma gondii* and Parkinson's disease. *Neurosci Lett.* 2010;**475**(3):129–131. doi:10.1016/j.neulet.2010.03.057
- [204] Yazar S, Arman F, Yalçın S, Demirtas F, Yaman O, Sahin I. Investigation of probable relationship between *Toxoplasma gondii* and cryptogenic epilepsy. *Seizure.* 2003;**12**(2):107–109. doi:10.1016/S1059-1311(02)00256-X

- [205] Webster JP, Kaushik M, Bristow GC, McConkey GA. *Toxoplasma gondii* infection, from predation to schizophrenia: can animal behaviour help us understand human behaviour? *J Exp Biol.* 2013;**216**(Pt 1):99–112. doi:10.1242/jeb.074716.
- [206] Fond G, Boyer L, Gaman A, Laouamri H, Attiba D, Richard JR, Delavest M, Houenou J, Le Corvoisier P, Charron D, Krishnamoorthy R, Oliveira J, Tamouza R, Yolken R, Dickerson F, Leboyer M, Hamdani N. Treatment with anti-toxoplasmic activity (TATA) for toxoplasma positive patients with bipolar disorders or schizophrenia: a cross-sectional study. *J Psychiatr Res.* 2015;**63**:58–64. doi:10.1016/j.jpsychires.2015.02.011

IntechOpen

