

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

4,800

Open access books available

122,000

International authors and editors

135M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.

For more information visit www.intechopen.com



Astrocytes Role in Parkinson: A Double-Edged Sword

Ricardo Cabezas, Marco Fidel Avila, Daniel Torrente,
Ramon Santos El-Bachá, Ludis Morales,
Janneth Gonzalez and George E. Barreto

Additional information is available at the end of the chapter

<http://dx.doi.org/10.5772/54305>

1. Introduction

Parkinson Disease (PD) is the second most chronic neurodegenerative disorder in the world, after Alzheimer's Disease (AD), and is estimated to affect about 1% of the population over 60 years of age. PD is caused by the disruption of dopaminergic neurotransmission in the basal ganglia, which causes a reduction in the numbers of dopaminergic neurons in the substantia nigra and formation of cytoplasmic inclusions called Lewy bodies [1].

Both in normal and pathological circumstances, astrocytes are critical supporters of neuronal function in processes such as antioxidant protection, glutamate clearance, the development and/or maintenance of blood brain barrier characteristics, the release of gliotransmitters and cytokines [2-4]. In recent years, much research on PD has focused on the astrocytic-neuronal crosstalk, suggesting that this interaction is important for future therapies against neurodegenerative processes. During brain damage events, astrocytes become transiently or permanently impaired, and the subsequent impact on neuronal cells may lead to pathological conditions such as PD [5-7].

In the present chapter, we provide a brief overview of the astrocytic functions and the pathophysiological events elicited during PD. Additionally, we explore the beneficial and damaging consequences of reactive astrogliosis in dopaminergic neurons during PD, particularly on oxidative damage, which is a main component of numerous neuropathological conditions, and that may have a damaging effect in astrocytic functions. We also highlight some of the cellular and animal models currently used in Parkinson research, such rotenone, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) and paraquat as inducers, which have many similar features with this disease. Finally, a brief overview of the future perspectives in astrocytic protection during Parkinson development is discussed.

2. Parkinson's disease

PD is a progressive neurodegenerative disorder caused by the neuronal death in the substantia nigra (SN), degeneration of dopaminergic neurotransmission, and the presence of α -synuclein and protein inclusions in neuronal cell bodies (Lewy bodies) [4-5,7]. Main symptoms of Parkinson are asymmetrical bradikinesia, rigidity, resting tremor and postural instability. Other non-motor symptoms that generate serious disability problems have also been noted, including fatigue, pain, Lewy Body dementia, psychosis, depression, and apathy [1]. Although there is not a cure for the disease, the most used and cheaper treatment for PD continues to be Levodopa [1,8]. However, about 40% of patients developed motor fluctuations and dyskinesias after 4 to 6 years of treatment [1], demonstrating that further pharmacological research is needed in order to counterbalance side effects. In this aspect, treatments using long-acting dopaminergic agents or a continuous dopaminergic effect in the striatum have been associated with less severe motor complications, given alone or in combination with L-dopa [9]. Some pharmacological agents that have shown promising applications, include dopamine agonist like apomorphine and ropinirole, and catechol-O-methyltransferase (COMT; EC 2.1.1.6) inhibitors [9].

Numerous reviews and articles agree that the exact cause of PD remains unknown [1,9-10]. Mutations in various proteins such as leucine-rich repeat kinase 2 (LRRK2; EC 2.7.11.1), Parkinson protein 2 (PARK2), probable cation-transporting ATPase type 13A2 (ATP13A2; EC 3.6.3.-), phosphatase and tensin homolog (PTEN)-induced putative kinase 1 (PINK1; EC 2.7.11.1), and Parkinson disease (autosomal recessive, early onset) 7 protein (DJ-1) have been observed in familiar cases of Parkinson, which only accounts for 10-15% of diagnosed cases [6,11-12]. Interestingly, LRRK2, PINK1, and DJ-1, which are present in mitochondrial membranes, have been suggested to play a role in reactive oxygen species (ROS) production by a defective maintenance of the mitochondrial membrane potential [12-13].

A number of environmental factors have been found to induce PD-like symptoms, and are currently used in animal and cellular models of the disease. Environmental factors include vascular insults to the brain, oxidative stress, neuroleptic drugs and repeated head trauma. [6,14]. Additionally, the exposure to pesticides like rotenone or 1-methyl-4-phenylpyridinium (MPP⁺) and heavy metals (manganese) increases the risk of PD development [6, 10, 14-15]. In this aspect, numerous epidemiologic and toxicologic studies have examined pesticides as a risk factor for PD and parkinsonism and the possible mechanisms by which pesticides may act [14-17].

Initiation and progression of PD is dependent upon cellular events, including failures in the protein degradation machinery, oxidative stress, mitochondrial dysfunction, defects in mitochondrial autophagy (mitophagy) and the continuous accumulation of α -synuclein, driven through cell to cell interactions between glial cells and neurons that ultimately lead to apoptosis [7,10,18]. Previous studies pointed that astrocytic α -synuclein deposition initiates the recruitment of phagocyte microglia that attacks and kills neurons in restricted brain regions [7,19], correlating this α -synuclein accumulation with nigral neuronal cell death [20], and suggest the importance of astrocytes in the initiation of the disease. Conversely, astrocytes

also have beneficial roles during PD progression [21-22]. For example, astrocytes express different antioxidant molecules such as glutathione peroxidase (EC 1.11.1.9), which have been inversely correlated with the severity of dopaminergic cell loss in the respective cell groups in patients with PD [4].

3. Astrocytes in PD

3.1. Astrocytic functions

Astrocytes are the most common cell type in the mammalian brain, conforming the glia with oligodendrocytes and microglia [23]. They are characterized by the expression of the intermediate filaments glial fibrillary acidic protein (GFAP) and vimentin (Vim). Astrocytes are essential for the metabolism of the brain, transporting multiple nutrients and metabolic precursors to the neurons by the malate-aspartate shuttle and other transporters [24]. There are two main types of astrocytes in the SNC: Protoplasmic astrocytes, which envelope neuronal bodies and synapses and fibrous astrocytes which interact with the nodes of Ranvier and oligodendroglia [7]. Current research has shown that only protoplasmic astrocytes have an increase in the accumulation of α -synuclein, whereas fibrous astrocytes do not [7,19].

Current knowledge indicates that astrocytes are critical for some cellular processes, such as the development and/or maintenance of blood-brain barrier characteristics, the promotion of neurovascular coupling, the attraction of cells through the release of chemokines, K^+ buffering, release of gliotransmitters, release of glutamate by calcium signaling, maintenance of general metabolism, control of the brain pH, metabolization of dopamine and other substrates by monoamine oxidases (MAOs; EC 1.4.3.4), uptake of glutamate and γ -aminobutyric acid (GABA) by specific transporters and production of antioxidants [2-3,25-27] (Figure 1). Recent evidence has shown that astrocytes are arranged in non-overlapping domains forming a syncytial network that may contact approximately 160.000 synapses, thus integrating neural activity with the vascular network [4,28]. In this aspect, astrocytic terminal processes, known as endfeet, contact the brain vasculature and enwrap the neuronal synapses, enabling the modulation of both neuronal activity and cerebral blood flow, following an elevation in intracellular Ca^{2+} levels in the endfeet [24,29].

During brain damage (including diseases, brain injury and oxidative stress), these astrocytic functions become transiently or permanently impaired, and the subsequent impact on neuronal cells may lead to pathological conditions and neurodegenerative diseases [3,26]. Neurons are more susceptible to injury than astrocytes, as they have limited antioxidant capacity, and rely heavily on their metabolic coupling with astrocytes to combat oxidative stress [3]. However, severe brain damage also results in astrocyte dysfunction, leading to increased neuronal death [30].

As previously stated, astrocytes exert both neuroprotective and neurodegenerative roles, depending on the molecules released by them, and the pathological or normal circumstances of their microenvironment [6]. For example, astrocytes release antioxidant molecules like

- ROS formation of substantia nigra dopaminergic neurons. *Biochimica et Biophysica Acta* 2011; 1812(6) 674-684.
- [69] Greenamyre JT, Betarbet R, Sherer TB. The rotenone model of Parkinson's disease: genes, environment and mitochondria. *Parkinsonism & Related Disorders* 2003; 9 (Suppl 2) S59-S64.
- [70] Diaz-Corrales FJ, Asanuma M, Miyazaki I, Miyoshi K, Ogawa N. Rotenone induces aggregation of gamma-tubulin protein and subsequent disorganization of the centrosome: relevance to formation of inclusion bodies and neurodegeneration. *Neuroscience* 2005; 133(1) 117-135.
- [71] Kawasaki A, Hayashi T, Nakachi K, Trosko JE, Sugihara K, Kotake Y, et al. Modulation of connexin 43 in rotenone-induced model of Parkinson's disease. *Neuroscience* 2009; 160(1) 61-68.
- [72] Zhang S, Liang R, Zhou F, Huang X, Ding JH, Hu, G. Reversal of rotenone-induced dysfunction of astrocytic connexin43 by opening mitochondrial ATP-sensitive potassium channels. *Cellular and Molecular Neurobiology* 2010; 31(1) 111-117.
- [73] Gao XF, Wang W, Yu Q, Burnstock G, Xiang ZH, He C. Astroglial P2X7 receptor current density increased following long-term exposure to rotenone. *Purinergic Signaling* 2011; 7(1) 65-72.
- [74] Sarafian TA, Montes C, Imura T, Qi J, Coppola G, Geschwind DH, et al. Disruption of astrocyte STAT3 signaling decreases mitochondrial function and increases oxidative stress in vitro. *PloS One* 2010; 5(3) e9532.
- [75] Ahmadi FA, Grammatopoulos TN, Poczobutt AM, Jones SM, Snell LD, Das M, et al. 2008. Dopamine selectively sensitizes dopaminergic neurons to rotenone-induced apoptosis. *Neurochemical Research* 2008; 33(5) 886-901.
- [76] Norazit A, Meedeniya AC, Nguyen MN, Mackay-Sim A. Progressive loss of dopaminergic neurons induced by unilateral rotenone infusion into the medial forebrain bundle. *Brain Research* 2010; 1360, 119-129.
- [77] Radad K, Gille G, Rausch WD. Dopaminergic neurons are preferentially sensitive to long-term rotenone toxicity in primary cell culture. *Toxicology In Vitro* 2008; 22(1) 68-74.
- [78] Verkhratsky A, Rodriguez JJ, Parpura V. Calcium signalling in astroglia. *Molecular and Cellular Endocrinology* 2012; 353(1-2) 45-56.
- [79] McGeer PL, McGeer EG. Glial reactions in Parkinson's disease. *Movement Disorders* 2008; 23(4) 474-83.
- [80] Sonsalla PK, Zeevalk GD, German DC. Chronic intraventricular administration of 1-methyl-4-phenylpyridinium as a progressive model of Parkinson's disease. *Parkinsonism & Related Disorders* 2008; 14(Suppl 2): S116-S118.

- [81] Di Monte, DA, Tokar I, Langston JW, Impaired glutamate clearance as a consequence of energy failure caused by MPP(+) in astrocytic cultures. *Toxicology and Applied Pharmacology* 1999; 158(3) 296-302.
- [82] Javitch JA, D'Amato RJ, Strittmatter SM, Snyder SH. Parkinsonism-inducing neurotoxin, *N*-methyl-4-phenyl-1,2,3,6-tetrahydropyridine: uptake of the metabolite *N*-methyl-4-phenylpyridine by dopamine neurons explain selective toxicity. *Proceedings of the National Academy of Sciences of the United States of America* 1985; 82(7) 2173–2177.
- [83] Xie HR, Hu LS, Li GY. SH-SY5Y human neuroblastoma cell line: in vitro cell model of dopaminergic neurons in Parkinson's disease. *Chinese Medical Journal* 2010; 123(8) 1086-1092.
- [84] Cui M, Aras R, Christian WV, Rappold PM, Hatwar M, Panza J, et al. The organic cation transporter-3 is a pivotal modulator of neurodegeneration in the nigrostriatal dopaminergic pathway. *Proceedings of the National Academy of Sciences of the United States of America* 2009; 106(19) 8043– 8048.
- [85] Di Monte DA, Wu EY, Delaney LE, Irwin I, Langston JW. Toxicity of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine in primary cultures of mouse astrocytes. *The Journal of Pharmacology and Experimental Therapeutics* 1992; 261(1) 44-49.
- [86] Bi J, Wang XB, Chen L, Hao S, An LJ, Jiang B, et al. Catalpol protects mesencephalic neurons against MPTP induced neurotoxicity via attenuation of mitochondrial dysfunction and MAO-B activity. *Toxicology in Vitro* 2008; 22(8) 1883-1889.
- [87] Berry C, La Vecchia C, Nicotera P. Paraquat and Parkinson's disease. *Cell Death and Differentiation* 2010; 17(7) 1115-1125.
- [88] Richardson JR, Quan Y, Sherer TB, Greenamyre JT, Miller GW. Paraquat neurotoxicity is distinct from that of MPTP and rotenone. *Toxicological Sciences* 2005; 88(1) 193–201.
- [89] Liou HH, Tsai MC, Chen CJ, Jeng JS, Chang YC, Chen SY, et al. Environmental risk factors and Parkinson's disease: a case-control study in Taiwan. *Neurology* 1997; 48(6) 1583–1588.
- [90] Olesen BT, Clausen J, Vang O. Characterization of the transcriptional profile in primary astrocytes after oxidative stress induced by Paraquat. *Neurotoxicology* 2008; 29(1) 13-21.
- [91] Rathinam ML, Watts LT, Narasimhan M, Riar AK, Mahimainathan L, Henderson GI. Astrocyte mediated protection of fetal cerebral cortical neurons from rotenone and paraquat. *Environmental toxicology and pharmacology* 2012; 33(2) 353-60.
- [92] Bové J, Prou D, Perier C, Przedborski S Toxin-induced models of Parkinson's disease. *NeuroRX: the journal of the American society for experimental Neurotherapeutics* 2005; 2(3) 484–494.

- [93] Wachter B, Schurger S, Rolinger J, von Ameln-Mayerhofer A, Berg D, Wagner HJ, et al. Effect of 6-hydroxydopamine (6-OHDA) on proliferation of glial cells in the rat cortex and striatum: evidence for de-differentiation of resident astrocytes. *Cell and Tissue Research* 2010; 342(2) 147-60.
- [94] Bambrick L, Kristian T, Fiskum G. Astrocyte mitochondrial mechanisms of ischemic brain injury and neuroprotection. *Neurochemical Research* 2004; 29(3) 601-608.
- [95] Dringen R. Metabolism and functions of glutathione in brain. *Progress in Neurobiology* 2000; 62(6) 649-671.
- [96] Safi R, Gardaneh M, Panahi Y, Maghsoudi N, Zaefizadeh M, Gharib E. Optimized quantities of GDNF overexpressed by engineered astrocytes are critical for protection of neuroblastoma cells against 6-OHDA toxicity. *The Journal of Molecular Neuroscience* 2011; 46(3) 654-665.
- [97] Zheng L, Ishii Y, Tokunaga A, Hamashima T, Shen J, Zhao QL, et al. Neuroprotective effects of PDGF against oxidative stress and the signaling pathway involved. *The Journal of Neuroscience Research* 2010; 88(6) 1273-1284.
- [98] Ouyang YB, Xu LJ, Emery JF, Lee, AS, Giffard RG. Overexpressing GRP78 influences Ca²⁺ handling and function of mitochondria in astrocytes after ischemia-like stress. *Mitochondrion* 2011; 11(2) 279-286.
- [99] Anderson CM, Swanson RA. Astrocyte glutamate transport: review of properties, regulation, and physiological functions. *Glia* 2000; 32(1) 1-14.
- [100] Sims NR, Nilsson M, Muyderman H. Mitochondrial glutathione: a modulator of brain cell death. *Journal of Bioenergetics and Biomembranes* 2004; 36(4) 329-333.
- [101] Lindenau J, Noack H, Possel H, Asayama K, Wolf G. Cellular distribution of superoxide dismutases in the rat CNS. *Glia* 2000; 29(1) 25-34.
- [102] Hirrlinger J, Dringen R The cytosolic redox state of astrocytes: Maintenance, regulation and functional implications for metabolite trafficking. *Brain Research Review* 2010; 63(1-2) 177-188.
- [103] Duncan AJ, Heales SJ. Nitric oxide and neurological disorders. *Molecular Aspects of Medicine* 2005; 26(1-2) 67-96.
- [104] Giordano G, Kavanagh TJ, Costa LG. Mouse cerebellar astrocytes protect cerebellar granule neurons against toxicity of the polybrominated diphenyl ether (PBDE) mixture DE-71. *Neurotoxicology* 2009; 30(2) 326-329.
- [105] Maier CM, Chan PH. Role of superoxide dismutases in oxidative damage and neurodegenerative disorders. *Neuroscientist* 2002; 8(4) 323-334.
- [106] Slemmer JE, Shacka JJ, Sweeney MI, Weber JT. Antioxidants and free radical scavengers for the treatment of stroke, traumatic brain injury and aging. *Current Medicinal Chemistry* 2008; 15(4) 404-414.

- [107] Damier P, Hirsch EC, Zhang P, Agid Y, Javoy-Agid F. Glutathione peroxidase, glial cells and Parkinson's disease. *Neuroscience* 1993; 52(1) 1-6.
- [108] Johnson JA, Johnson DA, Kraft AD, Calkins MJ, Jakel RJ, Vargas MR, et al. The Nrf2-ARE pathway: an indicator and modulator of oxidative stress in neurodegeneration. *Annals of the New York Academy of Sciences* 2008; 1147 61-69.
- [109] Martin HL, Teismann P. Glutathione—a review on its role and significance in Parkinson's disease. *The FASEB Journal* 2009; 23(10) 3263-3272
- [110] Hauser RA, Lyons KE, McClain T, Carter S, Perlmutter D. Randomized, double-blind, pilot evaluation of intravenous glutathione in Parkinson's disease. *Movement Disorders* 2009; 24(7) 979-83
- [111] Chan PH. Role of oxidants in ischemic brain damage. *Stroke* 1996; 27(6) 1124-1129.
- [112] Botella JA, Bayersdorfer F, Schneuwly S. Superoxide dismutase overexpression protects dopaminergic neurons in a *Drosophila* model of Parkinson's disease. *Neurobiology of Disease* 2008; 30(1) 65-73.
- [113] Radunović A, Porto WG, Zeman S, Leigh PN. Increased mitochondrial superoxide dismutase activity in Parkinson's disease but not amyotrophic lateral sclerosis motor cortex. *Neuroscience Letters* 1997; 239(2-3) 105-108.
- [114] Witt SN. Hsp70 molecular chaperones and Parkinson's disease. *Biopolymers* 2010; 93(3) 218-28.
- [115] Kakizuka, A. Protein precipitation: a common etiology in neurodegenerative disorders? *Trends in Genetics* 1998; 14(10) 396-402.
- [116] Giffard RG, Xu L, Zhao H, Carrico W, Ouyang Y, Qiao Y, et al. Chaperones, protein aggregation, and brain protection from hypoxic/ischemic injury. *Journal of Experimental Biology* 2004; 207(Pt 18) 3213-3220.
- [117] Slodzinski H, Moran LB, Michael GJ, Wang B, Novoselov S, Cheetham ME, et al. Homocysteine-induced endoplasmic reticulum protein (herp) is up-regulated in parkinsonian substantia nigra and present in the core of Lewy bodies. *Clinical Neuropathology* 2009; 28(5) 333-43.
- [118] Kalia SK, Kalia LV, McLean PJ. Molecular chaperones as rational drug targets for Parkinson's disease therapeutics. *CNS & Neurological Disorders-Drug Targets* 2010; 9(6) 741-53.
- [119] Bandopadhyay R, de Belleruche J. Pathogenesis of Parkinson's disease: emerging role of molecular chaperones. *Trends in Molecular Medicine* 2010; 16(1) 27-36.
- [120] Kahle PJ, Waak J, Gasser T. DJ1 and prevention of oxidative stress in Parkinson's disease and other age related disorders. *Free Radical Biology and Medicine* 2009; 47(10) 1354-61.

- [121] Shendelman S, Jonason A, Martinat C, Leete T, Abeliovich A. DJ-1 is a redox-dependent molecular chaperone that inhibits alpha-synuclein aggregate formation. *PLoS Biol* 2004; 2(11) e362.
- [122] Klucken J, Shin Y, Masliah E, Hyman BT, McLean PJ. Hsp70 Reduces alpha-synuclein aggregation and Toxicity. *The Journal of Biological Chemistry* 2004; 279(24) 25497-25502.
- [123] Lee J, Giordano S, Zhang J. Autophagy, mitochondria and oxidative stress: cross-talk and redox signalling. *The Biochemical Journal* 2012; 441(2) 523-540.
- [124] Mattson MP. Glutamate and neurotrophic factors in neuronal plasticity and disease. *Annals of the New York Academy of Sciences* 2008; 1144 97-112.
- [125] Nutt JG, Burchiel KJ, Comella CL, Jankovic J, Lang AE, Laws ER, Jr., et al. Randomized, double-blind trial of glial cell line-derived neurotrophic factor (GDNF) in PD. *Neurology* 2003; 60(1) 69-73.
- [126] Gill SS, Patel NK, Hotton GR, O'Sullivan K, McCarter R, Bunnage M, et al. Direct brain infusion of glial cell line-derived neurotrophic factor in Parkinson disease. *Nature Medicine* 2003; 9(5) 589-95.
- [127] Patel NK, Bunnage M, Plaha P, Svendsen CN, Heywood P, Gill SS. Intraputamenal infusion of glial cell line-derived neurotrophic factor in PD: a two-year outcome study. *Annals of Neurology* 2005; 57(2) 298-302.
- [128] Aberg ND, Brywe KG, Isgaard J. Aspects of growth hormone and insulin-like growth factor-I related to neuroprotection, regeneration, and functional plasticity in the adult brain. *Scientific World Journal* 2006; 6, 53-80.
- [129] Pang, Y., Zheng, B., Campbell, L.R., Fan, L.W., Cai, Z., Rhodes, P.G. IGF-1 can either protect against or increase LPS-induced damage in the developing rat brain. *Pediatric Research* 2010; 67(6) 579-584.
- [130] Kao SY. Rescue of alpha-synuclein cytotoxicity by insulin-like growth factors. *Biochemical and Biophysical Research Communications* 2009; 385(3) 434-438.
- [131] Yasuhara T, Shingo T, Kobayashi K, Takeuchi A, Yano A, Muraoka K, et al. Neuroprotective effects of vascular endothelial growth factor (VEGF) upon dopaminergic neurons in a rat model of Parkinson's disease. *European Journal of Neuroscience* 2004; 19(6) 1494-1504.
- [132] Yasuhara T, Shingo T, Muraoka K, wen Ji Y, Kameda M, Takeuchi A, et al. The differences between high and low-dose administration of VEGF to dopaminergic neurons of *in vitro* and *in vivo* Parkinson's disease model. *Brain Research* 2005; 1038(1) 1-10.
- [133] Tian YY, Tang CJ, Wang JN, Feng Y, Chen XW, Wang L, et al. Favorable effects of VEGF gene transfer on a rat model of Parkinson disease using adeno-associated viral vectors. *Neuroscience Letters* 2007; 421(3) 239-244.

- [134] Timmer M, Muller-Ostermeyer F, Kloth V, Winkler C, Grothe C, Nikkhah G. Enhanced survival, reinnervation, and functional recovery of intrastriatal dopamine grafts co-transplanted with Schwann cells overexpressing high molecular weight FGF-2 isoforms. *Experimental Neurology* 2004; 187(1) 118-136.
- [135] Tang Z, Arjunan P, Lee C, Li Y, Kumar A, Hou X, et al. Survival effect of PDGF-CC rescues neurons from apoptosis in both brain and retina by regulating GSK3beta phosphorylation. *Journal of Experimental Medicine* 2010; 207(4) 867-880.
- [136] Sullivan AM, Toulouse A. Neurotrophic factors for the treatment of Parkinson's disease. *Cytokine & Growth Factor Reviews* 2011; 22(3) 157-65.

