We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists



122,000

135M



Our authors are among the

TOP 1%





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



Biomarkers in Lung Cancer: Integration with Radiogenomics Data

Elena Aréchaga-Ocampo, Nicolas Villegas-Sepulveda, Eduardo Lopez-Urrutia, Mayra Ramos-Suzarte, César López-Camarillo, Carlos Perez-Plasencia, Claudia H. Gonzalez-de la Rosa, Cesar Cortes-Gonzalez and Luis A. Herrera

Additional information is available at the end of the chapter

http://dx.doi.org/10.5772/53426

1. Introduction

Lung cancer remains as one of the most aggressive cancer types with nearly 1.6 million new cases worldwide each year. There are an estimated 222,520 new cases and 157,300 deaths from lung cancer in the United States in 2010 [1]. Non-small cell lung cancer (NSCLC) is the most common subtype of lung cancer, comprising three major histological subtypes: adenocarcinoma, squamous cell carcinoma, and large cell carcinoma. Chronic exposure to carcinogens drives genetic and epigenetic damage that can result in lung epithelial cells progressively acquiring growth and/or survival advantages, giving as a result the generation of tumor cells. Studies have shown that some specific molecules contribute to sporadic tumors of lung cancer; even now, they are useful as predictive biomarkers. Mutations in at least one of the established lung cancer driver genes including egfr, kras, braf, her2, akt1, nras, pik3ca, mek1, eml4-alk and met amplification are found in approximately 60% of tumor specimens, and greater than 90% were "exclusive": only one mutation was found in a particular tumor [2]. Epidermal growth factor receptor (EGFR) exhibits overexpression or aberrant activation by mutations in 50 to 90% of NSCLC. Much effort has been focused on the development of targeted molecular inhibitors for this molecule, but it has become clear that molecular-targeted cancer therapies can only reach their full potential through appropriate patient selection. Conventional therapies as chemo- and radiotherapy continue being the first option of treatment for lung cancer patients, even their mutation status of NSCLC driver genes. Radiotherapy, alone or in combination with surgery, chemotherapy or biological therapies, play a critical role in the management of lung cancer. Currently, there are several clinical studies in radiation response of NSCLC tumors, which exhibit a wide spectrum of response to this modality treatment. Thus, a successful radiation sensitivity assay to calculate individual tumor radioresponse is central for the development of personalized strategies in radio-



© 2013 Arechaga-Ocampo et al., licensee InTech. This is an open access chapter distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Oncogenomics and Cancer Proteomics – 50 Novel Approaches in Biomarkers Discovery and Therapeutic Targets in Cancer

oncology. Some research groups have done effort in radiogenomics and proteomics in lung cancer with the purpose of finding specific molecules to predict resistance or sensibility to radiotherapy. NSCLC tumors with mutations in well-known molecular markers as EGFR and KRAS represent two molecularly distinct tumor entities, with different clinical behaviors. In this chapter we focus on the biomarkers used as biological therapy targets in lung cancer and their impact on resistance to therapeutic interventions. Moreover, we highlight genomic and proteomic data in radiation response to lung cancer.

2. Lung cancer

Lung cancer remains as one of the most aggressive cancer types with nearly 1.6 million new cases worldwide each year. In 2010, in the United States were estimated 222,520 new cases and 157,300 deaths from lung cancer [1]. Non-small cell lung cancer (NSCLC) subtype represents 85% of all cases of lung cancer, while small cell lung cancer (SCLC) subtype comprises 15%. Histologically, NSCLC is classified as adenocarcinoma, squamous cell carcinoma, and large cell carcinoma. This classification has important implications for the clinical management and prognosis of the disease [3]. Yet early detection methods are not extensively used in the wider population, malignancy is most commonly diagnosed at a late stage resulting in poor patient survival. Overall 5-year survival rates for lung cancer vary globally but are consistently low (7.5-16%) [1]. Approximately 40% of patients with advanced unresectable disease at the time of diagnosis have a poor prognosis. At present, no single chemo-radiation therapy regimen can be considered standard; despite the treatment choice for unresectable stage III NSCLC, a platinum-based chemotherapy regimen and thoracic radiation are concurrently administered. Chemotherapy concurrently with chest radiation therapy significantly improves the survival of patients with unresectable stage IIIA and IIIB disease. Decades of research have increased understanding the lung cancer as a multistep process involving genetic and epigenetic alterations, through which, resulting DNA damage transforms normal epithelial cells that progressively acquire growth and/or survival advantages until cancer arises [2,4-7]. Malignant transformation of lung epithelial cells is characterized by genetic instability, which can exist at the chromosomal level (with large-scale loss or gain of genomic material, translocations, and microsatellite instability) or at the nucleotide level (with single or several nucleotide base changes). Moreover, lung cancer is also related to genomic and epigenomic changes at the transcriptome (with altered gene and microRNA expression) and proteome [8-11] level. As many kinds of tumors, molecular abnormalities in lung cancer cells are typically targeted to proto-oncogenes, tumor suppressor genes, DNA repair genes, and other genes that can promote outgrowth and immortality of affected cells [12,13]. It is accepted that the successful discovery, validation and implementation of specific molecular markers for early diagnosis, clinical surveillance and determination of tumor response to therapeutic intervention could improve survival rates for patients, but only few biomarkers turned out to be useful in the clinic. *egfr* and *kras* gene mutations are prognosis markers in NSCLC [2,12,14]. Because of the importance of EGFR as a prognostic factor in NSCLC, mutated EGFR has been the target for development of biological therapies; at present, these therapies are being used in treatment of a certain group of patients [15]. In this context, current research focuses on identifying other potential molecular targets for the development of new agents and the assessment of better combinations of established therapies. Intensive research has originated numerous potential lung carcinoma molecular biomarkers related to therapy response in order to establish an appropriate molecular selection of patients, with focus on personalized medicine.

3. Genome biomarkers: The opening to personalized medicine in lung cancer

Nowadays, molecular and genetic studies have shown that some specific molecules contribute to sporadic tumors of lung cancer; they are useful as therapeutic targets and predictive biomarkers [16]. Recently, the National Cancer Institute's lung cancer mutation consortium (NCI's LCMC) performed such a study on more than 800 lung adenocarcinoma tumor specimens, examining mutations in established lung cancer driver genes (*egfr, kras, braf, her2, akt1, pik3ca, mek1, eml4-alk, met* amplification) [2]. Mutations in at least one of these genes were found in approximately 60% of tumor specimens, and greater than 90% were "exclusive", namely, only one mutation was found in a particular tumor. EGFR regulates important tumorigenic processes, including proliferation, apoptosis, angiogenesis, and invasion. EGFR, along with its ligands, is frequently overexpressed during the development and progression of NSCLC. *egfr* gene are amplified and over-expressed in 6% of NSCLC. However, activating mutations in exons 18 to 21 comprised in the kinase domain of EGFR (Figure 1) occur early in the development of adenocarcinomas with clinic characteristics like never-smoking, female sex and Asian ethnicity [7,15].

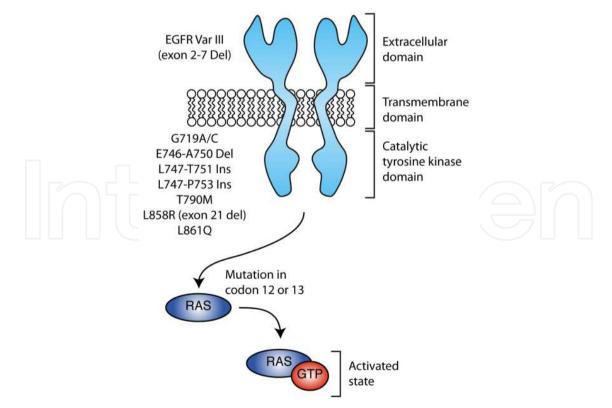


Figure 1. EGFR and KRAS mutations in NSCLC. Mutations in extracellular domain of EGFR have been implicated in resistance to treatment with mAb against EGFR. Mutations in TK domain are most

52 Novel Approaches in Biomarkers Discovery and Therapeutic Targets in Cancer

common in NSCLC, including L858R and E746-A750 deletion in exon 19. These mutations are target for small molecules inhibitors of tyrosine kinases domain (TKI). T790M is a mutation related to resistance to TKI treatment. Mutations in codon 12 or 13 of *kras* gene can lead to constantly union of GTP to KRAS protein, this represent the activate state of KRAS. GTP/KRAS induces activation of signaling depending to KRAS, permitting uncontrolled cell proliferation.

Mutated EGFR are present in 10-15% of NSCLC tumors [2,17]. Mutant EGFRs (either by exon 19 deletion or punctual mutation in exon 21 known as L858R) show an increased amount and duration of EGFR activation compared with wild-type receptors [18]. Mutated EGFR can activate RAS/RAF/MEK/MAPK and phosphoinositide 3-kinase (PI3K)/AKT and STAT3/STAT5 pathways [19-21]. Beside the importance of EGFR on lung carcinogenesis, some other molecules have been described as molecular markers for prognosis and therapeutic targets. Gene amplification and mutations in the kinase domain of C-erbB2 (HER-2/neu), a member of EGFR family, have been identified in patients with lung adenocarcinomas with a frequency of less than 5% and 5 to 10% respectively, and its overexpression are involved in ~25% of NSCLC cases [22]. EGFR and HER-2 kinase domain mutations have similar associations with female sex, non-smoking status and Asian background in patients with adenocarcinoma [15,22]. RAS/RAF/MEK/MAPK pathway is involved in signaling downstream from EGFR leading the growth and tumor progression in NSCLC. Activating kras gene mutation occurs in ~30% of cases of NSCLC, mostly adenocarcinomas. KRAS mutations are localized in exon 12 (in 90% of patients) or exon 13, and they are smoking-related G-T transversion and nonsmoking-related G-A transition [23]. KRAS mutations appear to be an early event in smoking-related lung adenocarcinoma, representing a poor prognosis in these patients. Another promising predictive markers in NSCLC are BRAF [24] and the oncogenic fusion gene of EML4-ALK [25]. BRAF, an effector molecule of RAS pathway, is mutated in about 2% of adenocarcinomas that does not show kras gene mutations. While eml4-alk is present in 2% to 7% of NSCLC cases; essentially, this fusion gene is present in young patients with adenocarcinoma and no exposure to smoking [26] (Figure 2).

Some other molecules have been identified based on expression and genomic data such as MYC and Cyclin D1 which are amplified and over-expressed in 2.5–10% and 5% of NSCLC respectively, while BCL-2 over-expression is involved in ~25% of cases of NSCLC [8,16]. Recent data have shown that methylation of the promoter regions of genes is a common event in NSCLC, which contributes to oncogenes over-expression or tumor genes suppressors silenced. These epigenetic changes may be an early event in NSCLC, since that promoter region of p16 gene is frequently methylated in smokers and premalignant lesion of lung cancer [27]. PI3K-AKT-mTOR pathway is altered in NSCLC. AKT overexpression has been described in a subgroup of NSCLC tumors jointly with mutations or amplification of PIK3CA gene. These genomic modifications are related with enhanced activity of PI3-K pathway mainly in squamous cell carcinoma tumors [28]. On the other hand, tissues of smoker patients show higher levels of angiogenic factors such as VEGF. VEGF expression increases in relationship with tumoral grade, which in turn, correlates with increased microvessel density, development and poor prognosis of lung cancer. Tumoral angiogenesis and angiogenic factors are regulated by hypoxic inductor factor (HIF) 1 α and 2 α or through

oncogenes as *egfr, kras* and *p53* [29]. Genomics and proteomics tools have permitted the identification of molecules associated with a specific phenotype in cancer. Gene, microRNA and protein-expression signatures in lung cancer have allowed for the identification of molecules that show promise as biomarkers or therapeutic target for diagnosis, prognosis and therapeutic treatments [review 11,30]. The research focused on improving anti-tumor treatments in lung cancer has focused on genomic and proteomic study of tumors with specific genetic background, such as tumors with mutations in EGFR and KRAS. This molecular classification has had an influence on the response to biological therapies based on monoclonal antibodies (mAb) and tyrosine kinase inhibitors (TKIs) in lung cancer patients [15, 31-32], but now, we also know that the genetic background of lung tumors has an impact on the response to chemotherapy [33] and radiotherapy [34,35].

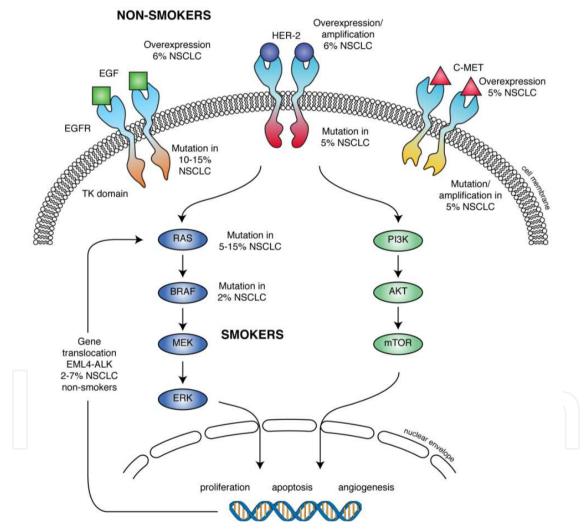


Figure 2. EGFR pathway in NSCLC. Mutations, amplification or overexpression of growth factors receptors such as EGFR, HER-2 and C-MET are most frequent in NSCLC tumors from non-smokers patients. All these genetic alterations have been observed commonly in adenocarcinomas, women and Asiatic ethnicity. EML4/ALK fusion gene is associated to NSCLC from young and non-smokers patients. KRAS mutations and signaling pathway depending to KRAS are most frequent in smoker patients. PI3K signaling pathway modifications are most frequently observed in squamous cell carcinomas.

4. Molecular and radiology therapies in lung cancer

4.1. Molecular therapy: Response and biological resistance

EGFR exhibits overexpression or aberrant activation in 50 to 90% of NSCLC. Mutations in EGFR allow sustained activation of EGF signaling for tumor cell survival, therefore, has been development targeted inhibitors for this molecule [16]. mAbs target the extracellular domain of EGFR and small molecules that inhibit intracellular EGFR tyrosine kinase domain function. In 2004, a significant advancement in the treatment of NSCLC was made following the observation that somatic mutations in the kinase domain of EGFR strongly correlated with sensitivity to EGFR TKIs [31, 32]. EGFR mutations are particularly prevalent in a patient subgroups with specific characteristics as adenocarcinoma histology, women, never smokers, and East Asian ethnicity [36]. This subgroup shows an exquisite sensitivity and marked tumor response to TKIs treatment. Despites the results obtained with biological therapies, there is a group of patients who do not respond to molecular therapy. Moreover, there is another group of patients with EGFR mutant lung cancer who initially respond to TKI treatment, but subsequently develop disease progression after a median of 10 to 14 months on treatment with biological therapy [37,38]. Hence, no optimal therapy thereafter has yet been established. Presumably, tumors do not respond because their molecular lesions are downstream of the therapeutic target [39]. Resistance to biologic therapy in NSCLC has been associated with EGFR exon-20 insertions [40] or a secondary T790M mutation [41], KRAS mutation [42], or amplification of the MET proto-oncogene [43,44], where MET is a transmembrane receptor with a tyrosine kinase domain, which activates signaling survival depending to PI3K and MAPK pathways. Of importance, Some reports showed that inhibition of MET signaling can restore sensitivity to TKIs [45]. HER-2 kinase domain mutations are associated with resistance to EGFR TKIs, but also with sensitivity to HER-2-targeted therapy [46].

Genomics data have provided information for developing targeted therapies in lung cancer patients based upon identification of cancer-specific vulnerabilities and set the stage for molecular biomarkers that provide information on clinical outcome and response to treatment. It has become clear that molecular-targeted cancer therapies can only reach their full potential through appropriate patient selection. In addition, there are now large clinical studies of lung cancer showing distinct chemotherapy and radiation responses. The majority of patients with lung cancer display advanced disease, these patients have obtained modest improvements in overall survival and quality of life through the use of systemic chemotherapy; however, the survival is still low, getting a median survival of 8 to 10 months [1]. Once recurred or metastasized, the disease is essentially incurable with survival rates at 5 years of less than 5%, and this has improved only marginally during the past 25 years [1]. The substantial genetic heterogeneity inherent to human cancers as an indicator of distinct phenotypes makes the identification of patients most likely to benefit from a given anticancer agent challenging. The description of molecules associated with resistance or sensitivity to cytotoxic treatments will improve personalized therapy for lung cancer. Radiotherapy, alone or in combination with surgery or chemotherapy, plays a critical role in the management of lung cancer. More than 60% of lung cancer patients receive radiotherapy at least once during the course of their disease [47].

4.2. Role of EGFR pathways in resistance and sensibility to radiotherapy

NSCLC tumors exhibit a wide response spectrum to radiation therapy but the molecular basis for this responsiveness is unknown. Some patients with NSCLC have a good response to radiation therapy with long-term local control while others relapse even with high dose treatment [48]. Many factors are involved in biological process of lung damage induced by radiation. At the molecular level, it is established that ionizing radiation causes various types of cellular damage; the creation of DNA breaks represents the principal damage induced by direct action of ionizing radiation or indirect action provoked by reactive species oxygen (ROS). Inadequately repaired DNA breaks leads to loss of cell clonogenicity via the generation of lethal chromosomal aberrations or the direct induction of apoptosis [49]. In addition to DNA breaks, ROS rapidly triggers the production of cytokines, growth factors, and more ROS, ultimately leading to chronic oxidative stress, hypoxia and the nonhealing tissue response in the lung [50,51]. Tumor radioresistance, including intrinsic resistance before treatments and acquired resistance during radiotherapy, is one of the main obstacles for radiotherapy efficiency for NSCLC. Some of the most important mechanisms associated with radioresistance in cancer including checkpoint pathway, mismatch repair process, and DNA damage repair [52-54]. Accumulating evidence suggests that radioresistance is often correlated with some genes, such as p53 [55] and EGFR [56]. In this regard, targeting EGFR pathway activation radiosensitizes human cancer cells [57-59], suggesting that the presence of overexpressed or activated oncogenes such as EGFR or RAS may be a mechanism for increased cellular resistance to radiation. In some models, it has been demonstrated that EGFR/Ras/Raf/MEK/ERK signaling may be activated in response to radiation, promoting cancer cell survival and proliferation [52-54,60] (Figure 3).

Variations in NSCLC responses to radiotherapy alone or in combination with chemotherapy or biological therapy are most likely due in the majority of cases to the genetic and epigenetic constitution of tumors [61,62]. In NSCLC, EGFR and KRAS oncogenes play an important role as prognostic factors; therefore, their role in radioresistance has been documented [63]. NSCLC cell lines harboring EGFR with mutations in tyrosine kinase domain were many folds more sensitive to radiation compared to cell lines with wild type EGFR. Radiosensitivity of NSCLC cell lines with mutant EGFR and human bronchial epithelial cells stably expressing mutant forms of EGFR was attributed to delayed DNA repair kinetics, defective radiation-induced arrest during DNA synthesis or mitosis, and pronounced increases in apoptosis or the occurrence of micronuclei [63]. Apparently, mutant EGFR is unable to translocate into the nucleus, which hinders its interaction with DNA-dependent protein kinase (DNA-PK), which is a fundamental enzyme for repair radiation-induced double strand breaks [63]. Besides of the promising role of mutant EGFR in radiosensitivity, the effort by blocking EGFR pathway to induce better response to radiotherapy has been limited. Inhibition of the EGFR by TKI or mAb, has been shown to

56 Novel Approaches in Biomarkers Discovery and Therapeutic Targets in Cancer

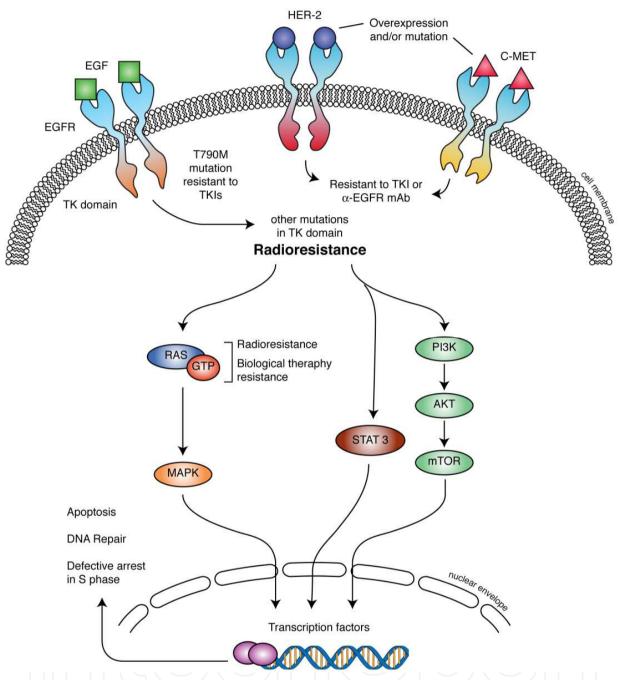


Figure 3. Role of EGFR pathway in radioresistance and radiosensibility in NSCLC. Aberrantly activation of EGFR pathways, including receptor mutations, KRAS activation, PI3K/AKT/mTOR pathway activation allows expression of specific genes for to regulate apoptosis, DNA repair, cell cycle and cell proliferation in order to get resistance to radiation.

radiosensitize a limited number of NSCLC cell lines *in vitro* and *in vivo* [34,35,63-65]. In NSCLC cell lines with wild-type or mutant p53, cell proliferation and clonogenic survival could be disturbed by senescence induced by EGFR inhibition and double strand breaks (DSB) produced by radiation. Apparently, radiosensitization by EGFR inhibitors is due to an increase in the levels of non-repairable DSB and disturbance of the MEK-ERK pathway [66]. Although a variety of signaling pathways downstream of EGFR have been implicated in

radioresistance, including PI3K-AKT, MEK-ERK, and PLC-PKC [67-69], no evidence of a common molecular pathway of radiosensitization, and cellular mechanisms by which EGFR TKI and mAb may cause radiosensitization remained largely elusive. Activating KRAS mutations is a marker for worse prognosis in NSCLC [23, 26]. Sun et al. evaluated whether the presence of mutation could be a potential factor for radioresistance. The results showed a reduced level of apoptosis in response to radiation in lung cancer cell line HCC2429 transfected with mutant KRAS 12V (mutation in codon 12). The authors suggested that phosphorylation of ERK could contribute to the low levels of apoptosis induced by radiotherapy in mutated KRAS lung cancer cells. This work suggests that KRAS mutation status is one potential factor associated with increased resistance to radiation-induced apoptosis in lung cancer cells [70]. The same group has recently shown that the specific inhibition of JAK2 by the novel molecule TG101209, induces radiosensibility through inhibition of phosphorylation of STAT3 and reduced expression of survivin in HCC2429 lung cancer cells. Moreover, the inhibition of survivin by treatment with TG101209 in experiments in vivo, was related to increased apoptosis, reducing tumor proliferation and vascular density [70]. Lu et al. demonstrated that overexpression of survivin leads to radioresistance in H460 lung cancer cells by inhibiting apoptosis and promoting cell survival, however, when survivin is inhibited by antisense oligonucleotides the cytotoxic effect of radiation is enhanced [71]. These results suggested that survivin might be a molecular marker for prognostic response to radiotherapy in NSCLC. While inhibition of survivin expression in HCC2429 and H460 cells were related to radiosensibility, both cell lines showed different apoptosis levels which were related to radioresistance depending on KRAS mutation status.

5. Lung cancer radiogenomics

Radiotherapy has played a key role in the control of tumor growth in many cancer patients, including lung cancer. Studies that originated more than 40 years ago [72,73] have indicated that tumors respond to radiotherapy by initiating a process called accelerated repopulation. In this process, the few surviving cells that escaped death after exposure to radiotherapy or chemotherapy can rapidly repopulate the badly damaged tumor by proliferating at a markedly faster pace. This phenomenon suggested that tumoral heterogeneity permits a cell population in the tumor to have advantages to avoid cell death induced by radiation. Cellular senescence, DNA repair and cell cycle checkpoint are cellular mechanisms that influence the resistance to radiotherapy. However, the molecular mechanisms that regulate the radioresistance phenotype have not been clear in cancer. For this reason, some research groups have focused in the study of biological models to obtain genomic and proteomic signatures in order to find genes and proteins that could predict radiosensitivity or radioresistance in lung tumors (Table 1). Although such researches have contributed to a partial understanding of the mechanisms underlying cellular radioresistance, the comprehensive functional mechanisms remain largely elusive. This may be quite reasonable since the mechanisms of radioresistance are a complex multigene interaction. In this sense, Torres-Roca et al. [74] in 2005, hypothesized that a radiation sensitivity classifier or predictor Oncogenomics and Cancer Proteomics – 58 Novel Approaches in Biomarkers Discovery and Therapeutic Targets in Cancer

could be developed based on gene expression profiles derived from DNA microarrays. This hypothesis was based in the fact of three main biological mechanisms partially correlated with clinical failure to radiotherapy, which are: hypoxia, intrinsic radiosensitivity and proliferation. These mechanisms, in turn, are handled by changes in gene expression.

Radiosensibility	
c-Jun* HDAC-1 RELA (p65 subunit NFkB PKC-beta	[75]
Sumo1	
c-Ab1	
STAT1	
AR	
CDK1	
IRF1	
Innate Radioresistance	
Up-regulated in basal condition	[76]
XRCC5	
ERCC5	
ERCC1	
RAD9A	
ERCC4	
Up-regulated after radiation	[76]
MDM2*	
BCL-2	
PKC-2	
PIM2	
Acquired Radioresistance	
Up-regulated	[77]
DDB2	
LOX	
CDH2	
CR4AB	
Livin α*	[79]
Down-regulated	[77]
GBP-1	
CD83	
TNNC1	[70]
TP53I3*	[78]

* Validated genes

Table 1. Genes associated to radiation response in NSCLC from genomics data

The authors developed a radiation classifier to calculate the radiosensitivity of tumor cell lines based on basal gene expression profiles obtained from the literature. They predicted the survival fraction to 2 Gy (SF2) value in 22 of 35 cell lines from the National Cancer Institute, a result significantly different from chance (P = 0.0002). In their approach, radiation sensitivity as a continuous variable, significance analysis of microarrays is used for gene selection, and a multivariate linear regression model is used for radiosensitivity prediction. In gene selection, they identified three novel genes: RbAp48, RGS19, and R5PIA, whose expression values correlated with radiation sensitivity. Exogenous overexpression of RbAp48 into three cancer cell lines (HS-578T, MALME-3M, and MDA-MB-231) induced radiosensitization (1.5- to 2-fold), moreover, higher proportion of transfected cells with RbAp48 were in G2-M phase of the cell cycle (27% versus 5%). Finally, RbAp48 overexpression is correlated with dephosphorylation of Akt, suggesting that RbAp48 might be exerting its effect radiosensitized by antagonizing the Ras pathway, but it could also do so through PI3K. The authors establish that radiation sensitivity can be predicted based on gene expression profiles and they introduce a genomic approach to the identification of novel molecular markers of radiation sensitivity. Despite of results in different tumor cell lines, this work included only four NSCLC cell lines and they were able to predict correct SF2 values for only two of them [74]. So, the study should be performed on a broader panel of NSCLC cell lines. In lung cancer, multiple studies have identified a wide array of genetic and epigenetic alterations, including mutations in DNA sequence, DNA copy number changes, aberrant DNA promoter methylation, changes in mRNA, microRNAs and protein expression [8], revealing many potential determinants and signaling pathways governing lung tumorigenesis and progression. Gene expression profiling analysis allows for an increase in the understanding of the molecular mechanisms and pathways that involve radioresistance. Thus, the strategy followed by Torres-Roca and collaborators can be applied to gene expression data reported in lung cancer, in order to identify new molecular targets for radiotherapy response. In this sense, we know that the response of tumor cells to radiation is accompanied by complex changes in the gene expression pattern. Based on mRNA expression profiles and systems-biology approach, Eschrich et al. [75] applied a linear regression algorithm that integrates gene expression with biological variables, including RAS and p53 status (mut/wt), and tissue of origin, with the aim of understanding radiosensitivity and identifying radiation specific markers. The modeling of radiosensitivity represented for the survival fraction at 2 Gy of 48 human cancer cell lines reported a direct correlation between gene expression and radiosensitivity of the lung cancer cell lines. The authors developed a model that classified four different clusters of genes that were markers for radiosensitivity. They identified 10 gene networks comprised by c-Jun, HDAC1, RELA (p65 subunit of NFKB), PKC-beta, SUMO-1, c-Abl, STAT1, AR, CDK1 and IRF1. Interestingly, RAS was a dominant variable in the analysis, as was the tissue of origin (lung), and their interaction with gene expression but not with p53. Moreover, when they knockeddown c-Jun in eight different cancer cell lines (lung, colon and breast cancer) there was an overall trend toward radioresistance, predominantly in lung cancers, but not in breast or colon cancers, implying that the origin of the tissue was important [75].

60 Novel Approaches in Biomarkers Discovery and Therapeutic Targets in Cancer

A problem in radiogenomics research is the difficulty to determine what fraction of the tumor cell population is radioresistant after a course of radiotherapy. For understand the radiation-mediated changes in gene expression that might result in different responses to radiation, Guo W et al. in 2005 [76] designed an oligonucleotide microarray to analyze the expression of 143 genes in lung cancer cell lines that differed in radiosensitivity. In the radioresistant A549 cells, 8 genes were significantly up-regulated and 10 genes were downregulated compared to radiosensitivity NCI-H446 cells. When the lung cancer cell lines were irradiated with 5Gy of γ rays, they identified genes showing altered expression and potential candidate genes that might confer radioresistance. In A549 cells, 19 up-regulated and 3 down-regulated genes, and 8 up-regulated and 18 down-regulated genes were found 6 and 24 h after irradiation, respectively. In NCI-H446 cells, the expression of 9 up-regulated and 8 down-regulated genes, and 8 up-regulated and 12 down-regulated genes was altered 6 and 24 h after irradiation, respectively. They found that MDM2, BCL2, PKCZ and PIM2 expression levels were increased in A549 cells and decreased in NCI-H446 cells after irradiation. Whereas, XRCC5, ERCC5, ERCC1, RAD9A, ERCC4 and the gene encoding DNA-PK were found to be increased to a higher level in A549 cells than in NCI-H446 cells. Inhibition of MDM2 by an antisense oligonucleotide in A549 cells resulted in increased radiosensitivity. The authors demonstrate the possibility that a group of genes involved in DNA repair, regulation of the cell cycle, cell proliferation and apoptosis are responsible for the different endogenous radioresistance between these two lung cancer cell lines [76]. To continue searching for new molecular evidences for radioresistance, Qing-Yong et al. in 2008 identified gene expression profiles in lung adenocarcinoma cell line Anip973 and obtained radioresistant phenotype cells (Anip973R). Expression profiles were obtained by oligonucleotide microarrays consisting of 21,522 human genes, while radioresistant cells Anip973R were obtained by fractionated ionizing radiation treatment of 4 Gy until a total dose of 60 Gy. In Anip973R cells, the authors reported 59 up-regulated genes associated with DNA damage repair (DDB2), extracellular matrix (LOX), cell adhesion (CDH2), and apoptosis (CRYAB); and 43 down-regulated genes associated with angiogenesis (GBP-1), immune response (CD83), and calcium signaling pathway (TNNC1). Validation of the selected eleven genes, including CD24, DDB2, IGFBP3, LOX, CDH2, CRYAB, PROCR, ANXA1 DCN, GBP-1 and CD83 by Q-RT-PCR was consistent with microarray analysis [77]. In 2010, Lee et al.. analyzed expression profiles of H460 NSCLC radiosensitive cell lines and their radioresistant counterpart (H460R) cells established by fractionated irradiation. By utilizing a cDNA microarray, they identified 1,463 genes altered more than 1.5-fold in H460R compared with parental H460. Tumor protein p53-inducible protein 3 (TP53I3) gene was significantly down-regulated in radioresistant H460R cells predicting a link to p53dependent cell death signaling. Interestingly, mRNA expression of TP53I3 differed in X-ray– irradiated H460 and H460R cells, and overexpression of TP53I3 significantly affected the cellular radiosensitivity of H460R cells [78]. These works showed that fractionated ionizing radiation can lead to the development of acquired radiation resistance across altered gene profiles. Genomic profile using in vivo models of radioresistance may provide new insights into mechanisms underlying the promotion of clinical resistance for NSCLC. Some other researches have been focused in describing specific molecules that revert the radioresistant phenotype. It is well known that there is a large amount of cell death during cytotoxic cancer therapy such as radiotherapy; therefore, radioresistance is associated with deregulation of apoptosis proteins. Sun et al. in 2011 reported the role of livin in radioresistance of lung adenocarcinomas cell lines A549 and SPC-A1. Livin is a IAPs family member whose expression is related with apoptosis inhibition, in some studies, it has been suggested that livin may be of clinical significance [79]. This work showed that A549 lung adenocarcinoma cells do not express livin in basal condition, but it is expressed after cells were irradiated. Moreover, gene silencing of livin by siRNA in SPC-A1 lung cell line induced a remarkable sensibility to radiation. Additionally, the authors showed that the isoform livin α had more impact on radioresistance that livin β had. These results suggested that livin expression in lung adenocarcinoma cells could be a radioresistance mechanism through downregulation of apoptosis. The cytotoxicity of oncological therapies is highly dependent on the cell cycle phase. G2/M phase is the one most sensitive to ionizing radiation. A work published in 2010 determined that arresting time on G2/M cell cycle phase is different between NSCLC cell lines sensitive and resistance to ionizing radiation. Radiosensitive H460 NSCLC cell line showed a significant G2/M arrest after 12 h of irradiation with 5 Gy of γ rays, while radioresistant A549 cell lines showed a significant G2/M arrest after 12 h of radiation. Interestingly, the arrest in A549 completely disappeared after 24 h of radiation. The arrest on G2/M correlated with higher methylated CpG sites of PTEN gene and consequently, reducing expression of the protein. PTEN negatively regulate pAKT which regulate negatively to p53. Therefore, radioresistance of A549 may depend to over activation of p53 signaling pathways. Epigenetic gene modification may be a way for regulating genes that participate in radiation response [80]. Signal transduction pathways depending to STAT have been explored. In A549 and SK-MES-1 cells, the exogenous over-expression of STAT3 was evaluated for its role in radioresponse. STAT3 over-expression enhanced the sensitivity to ionizing radiation in vitro and in vivo. Apparently, the radiosensibility may induce through STAT3-dependent inhibition of growth and induction of apoptosis [81]. These works showed that the regulation of signaling molecules that control apoptosis, cell growth and cell cycle has an important role in positive or negative radiation response.

6. Proteomics of radiation response in lung cancer

Despite proteomics being useful to find molecular markers associated to lung cancer cells [82], in radiation resistance research there are very few studies focused on applying proteomics to find new markers associated to radiotherapy response in lung cancer. Recently, Wei R *et al.* [83] in 2012 evaluated the multidrug resistance (MDR) effect on the radioresistance (RDR) in human lung adenocarcinoma cell lines and tissues. In this work, the authors screen MDR- and RDR-related proteins after irradiation of A549 and A549/DDP (resistant to cisplatin) human lung adenocarcinoma cells. The cell lines were analyzed by colony-forming assay and flow cytometry. Two-dimensional electrophoresis (2-DE) and matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-

TOF–MS) were utilized to identify differentially expressed proteins between irradiated A549 and A549/DDP. The SF2 value increased and the mean percentage of G2 phase and apoptosis rate decreased significantly in A549/DDP cells compared with A549 cells. Forty spots were found, and among them, 27 were identified through proteomics. Four up-regulated proteins (HSPB1, Vimentin, Cofilin-1, and Annexin A4) were confirmed by Western blot in MDR cells as compared with non-MDR cells. Immunohistochemistry showed that they were also over-expressed in MDR tissues compared with non-MDR counterparts of human lung adenocarcinomas. These results proved that the MDR in lung adenocarcinoma cells and tissues increased the radioresistance. HSPB1, Vimentin, Cofilin-1, and Annexin A4 are potential biomarkers for predicting lung adenocarcinomas response to chemo- and radiotherapy, as well as novel targets for treatment of lung adenocarcinomas [83].

7. Conclusion

One of the most important problems in lung oncology is lack of suitable biomarkers as therapeutic targets or the absence of predictors of therapy response. The genetic heterogeneity of the lung tumors influences the initial molecular resistance to therapies, but also in the development of resistance during treatment. The molecular mechanisms that influence the resistance to biological or radiological treatments, referring to the resistance mechanisms occurring naturally because of the carcinogenic process, or those developing as a result of evolutionary pressure that tumor cells undergoing during the treatment administration, is a barrier that has not been fully elucidated. With current genomics and proteomics studies in lung cancer focused on solving the mystery of therapeutic resistance, it has been possible to identify molecules that may serve as prognostic markers of response to radiological and molecular therapy resistance. Genes and proteins that regulate cell proliferation and survival, including signaling molecules and transcription factors such as KRAS, BRAF, PI3K, MAPK, mTOR, JAK2, STAT, survivin and others have demonstrated to be part of the molecular machinery that regulates therapeutic resistance. Moreover, gene and protein expression profiling of lung cancer has focused specifically on searching predictive markers to radiotherapy. Some studies have generated data on molecules involved in radioresistance or radiosensitivity either natural or acquired. Using therapeutic doses of radiation in in vitro models, it have described proteins implicated in DNA repair, cell cycle checkpoint and cell death. Mutations in EGFR pathway have played an important role as therapeutic targets for development of new therapies, moreover, mutations in this pathway represent a mechanism of radioresistance, suggesting that aberrant activation of EGFR pathway, including activated mutations in EGFR and KRAS might be an innate radioresistance mechanism in NSCLC. Despite advances in proteomics and radiogenomics in lung cancer, an enormous need to implement in vivo and clinical models for identification of effective biomarkers predictive in radio-oncology has also became evident. This is currently a promising field of cancer research in which genomics, tumor molecular biology and clinical experience interact to

achieve more effective combination therapies adjusted to the patient profile. Understanding the mechanisms of radioresistance of cells from solid tumors is of prime importance for further improvement of radiotherapy.

Author details

Elena Aréchaga-Ocampo^{*} Oncogenomics Lab, National Institute of Cancerology, Mexico

Nicolas Villegas-Sepulveda

Department of Molecular Biomedicine. Center for Research and Advanced Studies of the National Polytechnic Institute, Mexico

Eduardo Lopez-Urrutia Molecular Biochemistry Lab, UBIPRO, FES-I, National Autonomous University of Mexico, Mexico

Mayra Ramos-Suzarte Center of Molecular Immunology, Atabey, Havana, Cuba

César López-Camarillo Genomics Science Program, Oncogenomics and Cancer Proteomics Lab, Autonomous University of Mexico City, Mexico

Carlos Perez-Plasencia

Oncogenomics Lab, National Institute of Cancerology, Mexico Massive Sequencing Unite, National Institute of Cancerology-Genomics Lab, FES-I, UBIMED, National Autonomous University of Mexico, Mexico

Claudia H. Gonzalez-de la Rosa Department of Natural Science, Metropolitan Autonomous University-C, Mexico

Cesar Cortes-Gonzalez and Luis A. Herrera Cancer Biomedical Research Unit, National Institute of Cancerology-Biomedical Research Institute National Autonomous University of Mexico, Mexico

Acknowledgement

Authors gratefully acknowledge the financial support from the National Council of Science and Technology (CONACyT), Mexico (grants 115552 and 115591), and The Institute of Science and Technology (ICyT-DF), Mexico (grant PIUTE147).

8. References

[1] Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. CA Cancer Journal for Clinician 2011; 61(2) 69-90.

^{*} Corresponding Author

Oncogenomics and Cancer Proteomics – 64 Novel Approaches in Biomarkers Discovery and Therapeutic Targets in Cancer

- [2] Kris MG, Johnson BE, Kwiatkowski DJ, Iafrate AJ, Wistuba II, Aronson SL, Engelman JA, Shyr Y, Khuri FR, Rudin CM, Garon EB, Pao W, Schiller JH, Haura EB, Shirai K, Giaccone G, Berry LD, Kugler K, Minna JD, Bunn PA. Identification of driver mutations in tumor specimens from 1000 patients with lung adenocarcinoma: the NCI's lung cancer mutation consortium (LCMC) [abstract CRA7506]. Journal of Clinical Oncology 2011; 29(Suppl).
- [3] Wistuba II, Gazdar AF. Lung cancer preneoplasia. Annual Review of Pathology 2006; 1 331-348.
- [4] Sekido Y, Fong KM, Minna JD. Progress in understanding the molecular pathogenesis of human lung cancer. Biochemical and Biophysical Acta 1998; 1378(1) F21-59.
- [5] Silvestri GA, Alberg AJ, Ravenel J. The changing epidemiology of lung cancer with a focus on screening. British Medical Journal 2009; 339 451-454.
- [6] Mao L, Lee JS, Kurie JM, Fan YH, Lippman SM, Lee JJ, Ro JY, Broxson A, Yu R, Morice RC, Kemp BL, Khuri FR, Walsh GL, Hittelman WN, Hong WK. Clonal genetic alterations in the lungs of current and former smokers. Journal of National Cancer Institute 1997; 89(12) 857-862.
- [7] Sun S, Schiller JH, Gazdar AF. Lung cancer in never smokers-a different disease. Nature Reviews Cancer 2007; 7(10) 778-790.
- [8] Nikliński J, Niklińska W, Laudanski J, Chyczewska E, Chyczewski L. Prognostic molecular markers in non-small cell lung cancer. Lung Cancer 2001; 4-Suppl 2 S53-58.
- [9] Enfield KS, Pikor LA, Martinez VD, Lam WL. Mechanistic roles of noncoding RNAs in lung cancer biology and their clinical implications. Genetic Research International 2012; 2012 ID:737416.
- [10] Belinsky SA. Gene-Promoter Hypermethylation as a biomarker in lung cancer. Nature Reviews Cancer 2004; 4(9) 707-717.
- [11] Cho JY, Sung HJ. Proteomic approaches in lung cancer biomarker development. Expert Review of Proteomics 2009; 6(1) 27-42.
- [12] Sato M, Shames DS, Gazdar AF, Minna JD. A translational view of the molecular pathogenesis of lung cancer. Journal of Thoracic Oncology 2007; 2(4) 327-343.
- [13] Lee W, Jiang Z, Liu J, Haverty PM, Guan Y, Stinson J, Yue P, Zhang Y, Pant KP, Bhatt D, Ha C, Johnson S, Kennemer MI, Mohan S, Nazarenko I, Watanabe C, Sparks AB, Shames DS, Gentleman R, de Sauvage FJ, Stern H, Pandita A, Ballinger DG, Drmanac R, Modrusan Z, Seshagiri S, Zhang Z. The mutation spectrum revealed by paired genome sequences from a lung cancer patient. Nature 2010; 465(7297) 473-477.
- [14] Aviel-Ronen S, Blackhall FH, Shepherd FA, Tsao MS. K-ras mutations in non-small-cell lung carcinoma: a review. Clinical Lung Cancer 2006; 8(1) 30-38.
- [15] Pao W, Miller V, Zakowski M, Doherty J, Politi K, Sarkaria I, Singh B, Heelan R, Rusch V, Fulton L, Mardis E, Kupfer D, Wilson R, Kris M, Varmus H. EGF receptor gene mutations are common in lung cancers from "never smokers" and are associated with sensitivity of tumors to gefitinib and erlotinib. Proceedings of the National Academy of Science USA 2004; 101(36) 13306-13311.
- [16] Choong NW, Salgia R, Vokes EE. Key signaling pathways and targets in lung cancer therapy. Clinical Lung Cancer 2007; 8 Suppl 2 S52-S60.

- [17] Tang X, Shigematsu H, Bekele BN, Roth JA, Minna JD, Hong WK, Gazdar AF, Wistuba II. EGFR tyrosine kinase domain mutations are detected in histologically normal respiratory epithelium in lung cancer patients. Cancer Research 2005; 65(17) 7568-7572.
- [18] Lynch TJ, Bell DW, Sordella R, Gurubhagavatula S, Okimoto RA, Brannigan BW, Harris PL, Haserlat SM, Supko JG, Haluska FG, Louis DN, Christiani DC, Settleman J, Haber DA. Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. New England Journal of Medicine 2004; 350(21) 2129-2139.
- [19] Sordella R, Bell DW, Haber DA, Settleman J. Gefitinib sensitizing EGFR mutations in lung cancer activate anti-apoptotic pathways. Science 2004; 305(5687) 1163-1167.
- [20] de Mello RA, Marques DS, Medeiros R, Araujo AM. Epidermal growth factor receptor and K-Ras in non-small cell lung cancer-molecular pathways involved and targeted therapies. World Journal of Clinical Oncology 2011; 2(11) 367-376.
- [21] Li H, Schmid-Bindert G, Wang D, Zhao Y, Yang X, Su B, Zhou C. Blocking the PI3K/AKT and MEK/ERK signaling pathways can overcome gefitinib-resistance in nonsmall cell lung cancer cell lines. Advances in Medical Science 2011; 56(2) 275-284.
- [22] Shigematsu H, Takahashi T, Nomura M, Majmudar K, Suzuki M, Lee H, Wistuba II, Fong KM, Toyooka S, Shimizu N, Fujisawa T, Minna JD, Gazdar AF. Somatic mutations of the HER2 kinase domain in lung adenocarcinomas. Cancer Research 2005; 65(5) 1642-1646.
- [23] Riely GJ, Kris MG, Rosenbaum D, Marks J, Li A, Chitale DA, Nafa K, Riedel ER, Hsu M, Pao W, Miller VA, Ladanyi M. Frequency and distinctive spectrum of KRAS mutations in never smokers with lung adenocarcinoma. Clinical Cancer Research 2008; 14(18) 5731-5734.
- [24] Kobayashi M, Sonobe M, Takahashi T, Yoshizawa A, Ishikawa M, Kikuchi R, Okubo K, Huang CL, Date H. Clinical significance of BRAF gene mutations in patients with nonsmall cell lung cancer. Anticancer Research 2011; 31(12) 4619-4623.
- [25] Soda M, Choi YL, Enomoto M, Takada S, Yamashita Y, Ishikawa S, Fujiwara S, Watanabe H, Kurashina K, Hatanaka H, Bando M, Ohno S, Ishikawa Y, Aburatani H, Niki T, Sohara Y, Sugiyama Y, Mano H. Identification of the transforming EML4-ALK fusion gene in non-small cell lung cancer. Nature 2007; 448(7153) 561-566.
- [26] Brose MS, Volpe P, Feldman M, Kumar M, Rishi I, Gerrero R, Einhorn E, Herlyn M, Minna J, Nicholson A, Roth JA, Albelda SM, Davies H, Cox C, Brignell G, Stephens P, Futreal PA, Wooster R, Stratton MR, Weber BL. BRAF and RAS mutations in human lung cancer and melanoma. Cancer Research 2002; 62(23) 6997-7000.
- [27] Belinsky SA, Liechty KC, Gentry FD, Wolf HJ, Rogers J, Vu K, Haney J, Kennedy TC, Hirsch FR, Miller Y, Franklin WA, Herman JG, Baylin SB, Bunn PA, Byers T. Promoter hypermethylation of multiple genes in sputum precedes lung cancer incidence in a high-risk cohort. Cancer Research 2006; 66(6) 3338-3344.
- [28] Rekhtman N, Paik PK, Arcila ME, Tafe LJ, Oxnard GR, Moreira AL, Travis WD, Zakowski MF, Kris MG, Ladanyi M. Clarifying the spectrum of driver oncogene mutations in biomarker-verified squamous carcinoma of lung: lack of EGFR/KRAS and presence of PIK3CA/AKT1 mutations. Clinical Cancer Research 2012; 18(4) 1167-1176.

66 Novel Approaches in Biomarkers Discovery and Therapeutic Targets in Cancer

- [29] Giatromanolaki A. Prognostic role of angiogenesis in non-small cell lung cancer. Anticancer Research 2001; 6B 4373-4382.
- [30] Vrabec-Branica B, Gajovic S. Molecular biomarkers of lung carcinoma. Front Bioscience. (Elite Ed). 2012; 4 865-875.
- [31] Lynch TJ, Bell DW, Sordella R, Gurubhagavatula S, Okimoto RA, Brannigan BW, Harris PL, Haserlat SM, Supko JG, Haluska FG, Louis DN, Christiani DC, Settleman J, Haber DA. Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. New England of Journal Medicine 2004; 350(21) 2129-2139.
- [32] Paez JG, Jänne PA, Lee JC, Tracy S, Greulich H, Gabriel S, Herman P, Kaye FJ, Lindeman N, Boggon TJ, Naoki K, Sasaki H, Fujii Y, Eck MJ, Sellers WR, Johnson BE, Meyerson M. EGFR mutations in lung cancer: correlation with clinical response to gefitinib therapy. Science 2004; 304(5676) 1497-500.
- [33] Eberhard DA, Johnson BE, Amler LC, Goddard AD, Heldens SL, Herbst RS, Ince WL, Jänne PA, Januario T, Johnson DH, Klein P, Miller VA, Ostland MA, Ramies DA, Sebisanovic D, Stinson JA, Zhang YR, Seshagiri S, Hillan KJ. Mutations in the epidermal growth factor receptor and in KRAS are predictive and prognostic indicators in patients with non-small-cell lung cancer treated with chemotherapy alone and in combination with erlotinib. Journal of Clinical Oncology 2005; 23(25) 5900-5909.
- [34] Das AK, Sato M, Story MD. Non-small-cell lung cancers with kinase domain mutations in the epidermal growth factor receptor are sensitive to ionizing radiation. Cancer Research 2006; 66(19) 9601-9608.
- [35] Raben D, Helfrich B, Bunn PA Jr. Targeted therapies for non-small-cell lung cancer: biology, rationale, and preclinical results from a radiation oncology perspective. International Journal of Radiation Oncology Biology Physic 2004; 59(2 Suppl) 27-38.
- [36] Fujino S, Enokibori T, Tezuka N, Asada Y, Inoue S, Kato H, Mori A. A comparison of epidermal growth factor receptor levels and other prognostic parameters in non-small cell lung cancer. European Journal of Cancer 1996; 32A(12) 2070-2074.
- [37] Rosell R, Moran T, Queralt C, Porta R, Cardenal F, Camps C, Majem M, Lopez-Vivanco G, Isla D, Provencio M, Insa A, Massuti B, Gonzalez-Larriba JL, Paz-Ares L, Bover I, Garcia Campelo R, Moreno MA, Catot S, Rolfo C, Reguart N, Palmero R, Sánchez JM, Bastus R, Mayo C, Bertran-Alamillo J, Molina MA, Sanchez JJ, Taron M; Spanish Lung Cancer Group. Screening for epidermal growth factor receptor mutations in lung cancer. New England of Journal Medicine 2009; 361(10) 958-967.
- [38] Mok TS, Wu YL, Thongprasert S, Yang CH, Chu DT, Saijo N, Sunpaweravong P, Han B, Margono B, Ichinose Y, Nishiwaki Y, Ohe Y, Yang JJ, Chewaskulyong B, Jiang H, Duffield EL, Watkins CL, Armour AA, Fukuoka M. Gefitinib or carboplatin-paclitaxel in pulmonary adenocarcinoma. New England Journal of Medicine 2009; 361(10) 947-957.
- [39] Gazdar AF. Activating and resistance mutations of EGFR in non-small-cell lung cancer: role in clinical response to EGFR tyrosine kinase inhibitors. Oncogene 2009; 28(Suppl 1) S24-31.

- [40] Yasuda H, Kobayashi S, Costa DB. EGFR exon 20 insertion mutations in non-small-cell lung cancer: preclinical data and clinical implications. Lancet Oncology 2012; 13(1) e23-31.
- [41] Pao W, Miller VA, Politi KA, Riely GJ, Somwar R, Zakowski MF, Kris MG, Varmus H. Acquired resistance of lung adenocarcinomas to gefitinib or erlotinib is associated with a second mutation in the EGFR kinase domain. PLoS Medicine 2005; 2(3) e73.
- [42] Pao W, Wang TY, Riely GJ, Miller VA, Pan Q, Ladanyi M, Zakowski MF, Heelan RT, Kris MG, Varmus HE. KRAS mutations and primary resistance of lung adenocarcinomas to gefitinib or erlotinib. PLoS Medicine 2005; 2(1) e17.
- [43] Bean J, Brennan C, Shih JY, Riely G, Viale A, Wang L, Chitale D, Motoi N, Szoke J, Broderick S, Balak M, Chang WC, Yu CJ, Gazdar A, Pass H, Rusch V, Gerald W, Huang SF, Yang PC, Miller V, Ladanyi M, Yang CH, Pao W. MET amplification occurs with or without T790M mutations in EGFR mutant lung tumors with acquired resistance to gefitinib or erlotinib. Proceedings of the National Academy of Science USA 2007; 104(52) 20932-20937.
- [44] Engelman JA, Zejnullahu K, Mitsudomi T, Song Y, Hyland C, Park JO, Lindeman N, Gale CM, Zhao X, Christensen J, Kosaka T, Holmes AJ, Rogers AM, Cappuzzo F, Mok T, Lee C, Johnson BE, Cantley LC, Jänne PA. MET amplification leads to gefitinib resistance in lung cancer by activating ERBB3 signaling. Science 2007; 316(5827) 1039-1043.
- [45] Xu L, Kikuchi E, Xu C, Ebi H, Ercan D, Cheng KA, Padera R, Engelman JA, Jänne PA, Shapiro GI, Shimamura T, Wong KK. Combined EGFR/MET or EGFR/HSP90 inhibition is effective in the treatment of lung cancers codriven by mutant EGFR containing T790M and MET. Cancer Research 2012; 72(13) 3302-3311.
- [46] Wang SE, Narasanna A, Perez-Torres M, Xiang B, Wu FY, Yang S, Carpenter G, Gazdar AF, Muthuswamy SK, Arteaga CL. HER2 kinase domain mutation results in constitutive phosphorylation and activation of HER2 and EGFR and resistance to EGFR tyrosine kinase inhibitors. Cancer Cell 2006; 10 25-38.
- [47] Kong FM, Ten Haken RK, Schipper MJ, Sullivan MA, Chen M, Lopez C, Kalemkerian GP, Hayman JA. High-dose radiation improved local tumor control and overall survival in patients with inoperable/unresectable non-small-cell lung cancer: long-term results of a radiation dose escalation study. International Journal of Radiation Oncology Biology Physics 2005; 63(2) 324-333.
- [48] Bradley JD, Paulus R, Graham MV, Ettinger DS, Johnstone DW, Pilepich MV, Machtay M, Komaki R, Atkins J, Curran WJ; Radiation Therapy Oncology Group. Phase II trial of postoperative adjuvant paclitaxel/carboplatin and thoracic radiotherapy in resected stage II and IIIA non-small-cell lung cancer: promising long-term results of the Radiation Therapy Oncology Group-RTOG 9705. Journal of Clinical Oncology 2005; 23(15) 3480-3487.
- [49] Willers H, Dahm-Daphi J, Powell SN. Repair of radiation damage to DNA. British Journal of Cancer 2004; 90 1297-1301.
- [50] Marks LB, Yu X, Vujaskovic Z, Small W Jr, Folz R, Anscher MS. Radiation-induced lung injury. Seminars in Radiation Oncology 2003; 13(3) 333-345.

68 Novel Approaches in Biomarkers Discovery and Therapeutic Targets in Cancer

- [51] Fleckenstein K, Gauter-Fleckenstein B, Jackson IL, Rabbani Z, Anscher M, Vujaskovic Z. Using biological markers to predict risk of radiation injury. Seminars Radiation Oncology 2007; 17(2) 89-98.
- [52] Contessa JN, Hampton J, Lammering G, Mikkelsen RB, Dent P, Valerie K, Schmidt-Ullrich RK. Ionizing radiation activates Erb-B receptor dependent Akt and p70 S6 kinase signaling in carcinoma cells. Oncogene 2002; 21(25) 4032-4041.
- [53] Bernhard EJ, Stanbridge EJ, Gupta S, Gupta AK, Soto D, Bakanauskas VJ, Cerniglia GJ, Muschel RJ, McKenna WG. Direct evidence for the contribution of activated N-ras and K-ras oncogenes to increased intrinsic radiation resistance in human tumor cell lines. Cancer Research 2000; 60(23) 6597-6600.
- [54] Rodemann HP, Dittmann K, Toulany M. Radiation-induced EGFR signaling and control of DNA-damage repair. International Journal of Radiation Biology 2007; 83 781-791.
- [55] Biard DS, Martin M, Rhun YL, Duthu A, Lefaix JL, May E, May P. Concomitant p53 gene mutation and increased radiosensitivity in rat lung embryo epithelial cells during neoplastic development. Cancer Research 1994; 54(13) 3361-3364.
- [56] Wang H, Yu JM, Yang GR, Song XR, Sun XR, Zhao SQ, Wang XW, Zhao W. Further characterization of the epidermal growth factor receptor ligand 11C-PD153035. Chinese Medical Journal (Engl) 2007; 120(11) 960-964.
- [57] Camp ER, Summy J, Bauer TW, Liu W, Gallick GE, Ellis LM. Molecular mechanisms of resistance to therapies targeting the epidermal growth factor receptor. Clinical Cancer Research 2005; 11 397-405.
- [58] Chinnaiyan P, Huang S, Vallabhaneni G, Armstrong E, Varambally S, Tomlins SA, Chinnaiyan AM, Harari PM. Mechanisms of enhanced radiation response following epidermal growth factor receptor signaling inhibition by erlotinib (Tarceva). Cancer Research. 2005; 65(8) 3328-3335.
- [59] Schmidt-Ullrich RK, Mikkelsen RB, Dent P, Todd DG, Valerie K, Kavanagh BD, Contessa JN, Rorrer WK, Chen PB. Radiation-induced proliferation of the human A431 squamous carcinoma cells is dependent on EGFR tyrosine phosphorylation. Oncogene 1997; 15(10) 1191-1197.
- [60] Yacoub A, McKinstry R, Hinman D, Chung T, Dent P, Hagan MP. Epidermal growth factor and ionizing radiation up-regulate the DNA repair genes XRCC1 and ERCC1 in DU145 and LNCaP prostate carcinoma through MAPK signaling. Radiation Research 2003;159 439-452.
- [61] Pao W, Kris MG, Iafrate AJ. Integration of molecular profiling into the lung cancer clinic. Clinical Cancer Research 2009; 15(17) 5317-5322.
- [62] Minna JD, Girard L, Xie Y. Tumor mRNA expression profiles predict responses to chemotherapy. Journal of Clinical Oncology 2007; 25(28) 4329-4336.
- [63] Das AK, Chen BP, Story MD. Somatic mutations in the tyrosine kinase domain of epidermal growth factor receptor (EGFR) abrogate EGFR-mediated radioprotection in non-small cell lung carcinoma. Cancer Research 2007; 67(11) 5267-5274.
- [64] Chinnaiyan P, Huang S, Vallabhaneni G. Mechanisms of enhanced radiation response following epidermal growth factor receptor signaling inhibition by erlotinib (Tarceva). Cancer Research 2005; 65(8) 3328-3335.

- [65] Shibuya K, Komaki R, Shintani T. Targeted therapy against VEGFR and EGFR with ZD6474 enhances the therapeutic efficacy of irradiation in an orthotopic model of human non-small-cell lung cancer. International Journal of Radiation Oncology Biology Physics 2007; 69(5) 1534-1543.
- [66] Wang M, Morsbach F, Sander D, Gheorghiu L, Nanda A, Benes C, Kriegs M, Krause M, Dikomey E, Baumann M, Dahm-Daphi J, Settleman J, Willers H. EGF receptor inhibition radiosensitizes NSCLC cells by inducing senescence in cells sustaining DNA double-strand breaks. Cancer Research 2011; 71(19) 6261-6269.
- [67] Kriegs M, Kasten-Pisula U, Rieckmann T, Holst K, Saker J, Dahm-Daphi J, Dikomey E. The epidermal growth factor receptor modulates DNA double-strand break repair by regulating non-homologous end-joining. DNA Repair (Amst). 2010; 9(8) 889-897.
- [68] Sturla LM, Amorino G, Alexander MS, Mikkelsen RB, Valerie K, Schmidt-Ullrichr RK. Requirement of Tyr-992 and Tyr-1173 in phosphorylation of the epidermal growth factor receptor by ionizing radiation and modulation by SHP2. Journal of Biological Chemistry 2005; 280 14597-14604.
- [69] Toulany M, Dittmann K, Kruger M, Baumann M, Rodemann HP. Radioresistance of K-Ras mutated human tumor cells is mediated through EGFR-dependent activation of PI3K-AKT pathway. Radiotherapy Oncology 2005; 76 143-150.
- [70] Sun Y, Moretti L, Giacalone NJ, Schleicher S, Speirs CK, Carbone DP, Lu B. Inhibition of JAK2 signaling by TG101209 enhances radiotherapy in lung cancer models. Journal of Thoracic Oncology 2011; 6(4) 699-706.
- [71] Lu B, Mu Y, Cao C, Zeng F, Schneider S, Tan J, Price J, Chen J, Freeman M, Hallahan DE. Survivin as a therapeutic target for radiation sensitization in lung cancer. Cancer Research. 2004; 64 2840-2845.
- [72] Hermens AF, Barendsen GW. Changes of cell proliferation characteristics in a rat rhabdomyosarcoma before and after x-irradiation. European Journal of Cancer 1969; 5 173-189.
- [73] Stephens TC, Currie GA, Peacock JH. Repopulation of gamma-irradiated Lewis lung carcinoma by malignant cells and host macrophage progenitors. British Journal of Cancer 1978; 38(5) 573-582.
- [74] Torres-Roca JF, Eschrich S, Zhao H, Bloom G, Sung J, McCarthy S, Cantor AB, Scuto A, Li C, Zhang S, Jove R, Yeatman T. Prediction of radiation sensitivity using a gene expression classifier. Cancer Research 2005; 65(16) 7169-7176.
- [75] Eschrich S, Zhang H, Zhao H. Systems biology modeling of the radiation sensitivity network: a biomarker discovery platform. International Journal of Radiation Oncology Biology Physics 2009; 75(2) 497-505.
- [76] Guo WF, Lin RX, Huang J, Zhou Z, Yang J, Guo GZ, Wang SQ. Identification of differentially expressed genes contributing to radioresistance in lung cancer cells using microarray analysis. Radiation Research 2005; 164 27-35.
- [77] Qing-yong X, Yuan G, Yan L, Wei-zhi Y, Xiang-ying X. Identification of differential gene expression profiles of radioresistant lung cancer cell line established by fractionated ionizing radiation *in vitro*. Chinese Medicine Journal 2008; 121(18) 1830-1837.

- 70 Novel Approaches in Biomarkers Discovery and Therapeutic Targets in Cancer
 - [78] Lee YS, Oh JH, Yoon S, Kwon MS, Song CW, Kim KH, Cho MJ, Mollah ML, Je YJ, Kim YD, Kim CD, Lee JH. Differential gene expression profiles of radioresistant non-small-cell lung cancer cell lines established by fractionated irradiation: tumor protein p53-inducible protein 3 confers sensitivity to ionizing radiation. International Journal of Radiation Oncology Biology Physics 2010; 77(3) 858-866.
 - [79] Sun JG, Liao RX, Zhang SX, Duan YZ, Zhuo WL, Wang XX, Wang ZX, Li DZ, Chen ZT. Role of inhibitor of apoptosis protein Livin in radiation resistance in non small cell lung cancer. Cancer Biotherapy and Radiopharmacy 2011; 26(5) 585-592.
 - [80] Jung IL, Kang HJ, Kim KC, Kim IG. PTEN/pAkt/p53 signaling pathway correlates with the radioresponse of non-small cell lung cancer. International Journal of Molecular Medicine 2010; 25(4) 517-523.
 - [81] Yin ZJ, Jin FG, Liu TG, Fu EQ, Xie YH, Sun RL. Overexpression of STAT3 potentiates growth, survival, and radioresistance of non-small-cell lung cancer (NSCLC) cells. Journal of Surgical Research 2011; 171(2) 675-683.
 - [82] Indovina P, Marcelli E, Pentimalli F, Tanganelli P, Tarro G, Giordano A. Mass spectrometry-based proteomics: The road to lung cancer biomarker discovery. Mass Spectrometry Reviews. 2012; DOI: 10.1002/mas.21355.
 - [83] Wei R, Zhang Y, Shen L, Jiang W, Li C, Zhong M, Xie Y, Yang D, He L, Zhou Q. Comparative proteomic and radiobiological analyses in human lung adenocarcinoma cells. Molecular and Cellular Biochemistry 2012; 359(1-2) 151-159.

