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Different Modulators of Airways and Distal Lung Parenchyma Contractile Responses in the Physiopathology of Asthma

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1. Introduction

Asthma outcomes from an allergen-driven Th2 (T helper 2) response in which airway hyperresponsiveness (AHR) is associated with chronic airway inflammation and airway remodeling have crucial clinical importance (1-3).

Recent investigations have emphasized the importance of lung tissue alterations in the pathophysiology of this syndrome. Additionally, current investigations have shown that patients who died of asthma presented important alterations in the lung parenchyma (4-7) that could also be observed in animal models of chronic allergic inflammation (8-11). In this regard, the importance of the mechanical properties of the lung parenchyma has been characterized as one of the major determinants of physiological function (8, 12-15).

Asthma physiopathology is highly complex and involves a diverse immune response and the release of different types of mediators. The bronchial and tissue inflammation is caused by eosinophils, mast cells and T lymphocytes (16), and the persistence of inflammation induces changes in the structural components of the airway and alveolar walls (5, 8, 17).

The airway smooth muscle (ASM) has been considered the main effector of the AHR in asthma (17-19) and is also believed to contribute to airway remodeling and inflammation due to its increased sensitivity to different bronchoconstrictor stimuli.

The continuous bronchial inflammation process associated with the release of various mediators is thought to be responsible for asthma symptoms directly and indirectly by inducing the constriction of the ASM, enhancing airway responsiveness to different stimuli,



and inducing changes in the structural components of the airway wall, leading to airway remodeling.

Inhaled corticosteroids, which are the gold-standard treatment for asthmatic patients, are more involved in counteracting the airway inflammation than in acting in the ASM. Although some studies have shown the potential of corticosteroids in causing bronchodilation, their role in airway smooth muscle relaxation is controversial. In its formulation (hydrofluoroalkane-HFA), this inhaled corticosteroid is delivered to the distal airways more effectively (68.3%) than chlorofluorocarbon formulations (19.7%) (20, 21). Although eosinophilic infiltration could be adequately controlled in the distal airways, whether both distal lung parenchyma eosinophilic infiltration and extracellular matrix remodeling may be sufficiently modulated by this new treatment is not clear (8, 20).

We discuss in this chapter the role of different mediators and modulators in the contractile responses of the airways and lung distal parenchyma. These studies contribute to the understanding of the mechanisms involved in asthma physiopathology and in smooth muscle contraction and also open opportunities to develop new therapeutic tools to treat asthma. In this regard, we will address the importance of the modulation of iNOS, arginase and Rho kinase pathways, the impact of inducing oral tolerance and the effects of exercise. In addition, aspects of neuroimmunomodulation, including stress effects, will be discussed.

2. Airway and lung parenchyma hyperresponsiveness and smooth muscle alterations in asthma

AHR is the hallmark of asthma, and it is characterized by an increase in the airway response to bronchoconstrictor stimuli. There are two components of AHR. AHR has a variable component that mainly reflects the current airway inflammation (22, 23) and an irreversible component that probably reflects pulmonary remodeling (24).

As described above, the ASM is the major effector of the AHR in asthma (17-19). There are two phenotypes of ASM cells in asthmatics: the contractile, which is responsive to contractile agonists and has an increased expression of contractile proteins, and the syntheticproliferative, which lacks the responsiveness to contractile stimuli and has a reduced expression of contractile proteins (17). Both phenotypes can coexist or not in the airways of the same person (25-29). Depending on the triggers, it can also induce the proliferation of the synthetic-proliferative cells or induce the maturation of these cells into contractile cells (17, 19).

In patients with asthma, the ASM was thought to generate more force and consequently a greater extent of contraction in response to different stimuli (30). Cultures of ASM cells isolated from lung tissue (trachea, bronchi) were used to study the contractile responses and the mitogenic and synthetic responses, which revealed that these cells are active players in inflammation (25, 31, 32).

In addition, ASM can contribute to lung inflammation. Many studies showed that there was an increased number of mast cells in the asthmatic ASM layer (33-38). Brightling et al. (32) evaluated patients with asthma and eosinophilic bronchitis and observed that both groups showed an increase in eosinophils but that the patients with eosinophilic bronchitis were not hyperresponsive to bronchoconstrictor stimuli. The analysis of the ASM layers in these patients showed that only the asthmatics showed a higher number of mast cells and a worsening of respiratory function, suggesting that the mast cells present in the ASM of asthmatics are responsible for the enhancement of airway narrowing.

The ASM cells release chemotactic agents for mast cells, such as CCL11 (25), CXCL10 (34) and CX3CL1 (35). Because the mast cells are in the airways, they adhere to the ASM cells and produce, together with the eosinophils, contractile mediators, such as prostaglandins (PGF2 α , PGD2, and thromboxane TXA2) (39).

Clinically, the AHR symptoms are described as cough, tightness of the chest and wheezing after exercise or exposure to cold air or other environmental irritants (40). Some studies suggest that monitoring of the AHR in asthmatic patients can serve as a guide to asthma therapy (24).

In clinical and experimental studies, AHR is evaluated by the aerosol administration of bronchoconstrictor agonists, such as histamine, methacholine or carbachol. This methodology considers that the ASM in asthmatics exposed to exogenous bronchoconstrictor stimuli showed an increased tonus and a concomitant bronchoconstriction. The hyperresponsiveness occurs due to an increase in both the sensitivity and/or reactivity of the airways (Figure 1). The increase in sensitivity is a reduction in the minimal dose that is necessary to induce bronchoconstriction, whereas the increase in reactivity is described by an increase in the intensity of the bronchoconstriction.

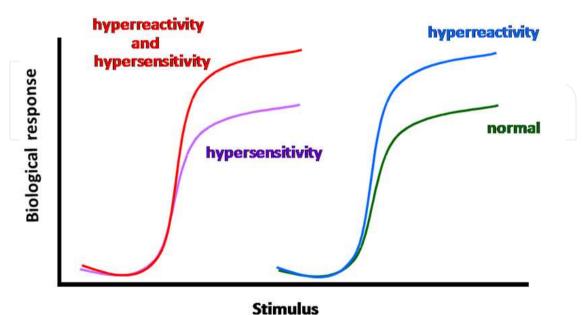


Figure 1. Airway hyperresponsiveness.

Considering that lung parenchyma strips have long been used to study the behavior of the peripheral lung, they are commonly used to evaluate the mechanics and pharmacological properties of the lung periphery (41). Dolhnikoff et al. (15) concluded that human lung tissue strips respond to an acetylcholine (ACh) challenge with changes in their dynamic mechanical behavior. In addition, Lanças et al. (10) have recently shown that the lung tissue is involved in the late asthmatic response in guinea pigs with chronic allergic lung inflammation, which is correlated to lung tissue eosinophilic recruitment and extracellular matrix remodeling.

Although the *in vivo* apparatus of oscillatory mechanics permits the evaluation of large and small airways, the oscillatory mechanics *in vitro* provide a tool for the specific evaluation of the lung periphery with minimal interference with the compartment represented by the small airways (10, 15). In addition, this *in vitro* methodology permits the specific analysis of the effects of several mediator/modulators in the lung periphery while avoiding other compensatory mechanisms that could be activated in *in vivo* studies. Lung parenchyma strips exclusively represent the distal units of the lung tissue and offer a better assessment of pure tissue properties. Thus, studies using this technique have been performed to evaluate the mechanical and pharmacological properties of the lung periphery (10, 42, 43).

Several authors have discussed the importance of these structures in the mechanical behavior of lung tissue, including the consequences of stiffening the extracellular matrix network and of elastin and collagen digestion in these responses (44, 45). In the subpleural region, there was a small number of bronchial and blood vessels (less than 30%). Romero et al. (46) concluded that pneumoconstriction significantly modifies the intrinsic mechanical properties of the connective matrix via a mechanism differing from that of passive stretching. In fact, the contractile cells could be accepted as being able to modulate the mechanical properties of the connective matrix.

3. Mediators involved in airways and distal lung parenchyma contractile responses

A large quantity of extracellular agonists (inflammatory mediators or neurotransmitters) released in an inflammatory milieu can stimulate the contraction of ASM in asthma. Mediators that are found in high concentrations in asthma, including leukotrienes (produced by inflammatory cells) (47), prostaglandins such as PGF2 α , PGD2, and thromboxane TXA2 (produced by mast cells and/or eosinophils) (39) and endothelin (produced by epithelial or endothelial cells) (48, 49), are direct contractile agonists of ASM. Neurotransmitters, such as ACh or neurokinins, are highly present in asthma and are also potent contractile messengers of ASM (50, 51).

To increase the release of the contractile mediators, there is also a lower release of relaxant mediators, such as vasoactive intestinal peptide (VIP), PGE 2, adrenaline and NO (35, 52). These mediators are involved in the mechanisms responsible for many of the structural and functional lung alterations observed in asthmatic patients and in animal models of chronic pulmonary allergic inflammation (53-55).

3.1. Excitatory non-adrenergic non-cholinergic mediators: Neurokinins and Substance P

Neurokinins and substance P are involved in the excitatory NANC responses and modulate several histopathological alterations observed in asthmatics, such as airway smooth muscle contraction, peribronchial edema formation and airway mucous secretion. In this regard, substance P (SP) and neurokinin A (NKA) play significant roles in priming and recruiting eosinophils and lymphocytes in models of allergic lung inflammation (56-58).

Asthmatic patients are hyperresponsive to the SP and NK1 expression that is augmented in their bronchi (59). Tibério et al. (60) showed that capsaicin infusion induced an increase in the respiratory system resistance that was attenuated mainly by a NK2 receptor antagonist. The NK receptors are also involved in eosinophil recruitment, which contributes to the hyperresponsiveness. Using a model of experimental asthma in guinea pigs, Tibério et al. (57) evaluated the airway inflammation induced by repeated exposure to ovalbumin and the effects of neurokinin depletion on these responses. These authors showed that neurokinin depletion reduced the peribronchial edema, CD4 lymphocytes and the hyperresponsiveness to the antigen challenge. In addition, Prado et al. (61) showed that the bronchodilation observed after 14 days of capsaicin infusion could be related to the increase in NO produced by nNOS, which counteracts the bronchoconstriction.

Emphasizing that SP has a preferential affinity for NK1 receptors and that neurokinin A has a preferential affinity for NK2 receptors is important (58). However, each neurokinin also exhibits activity at other NK receptors. In this regard, Regoli et al. (62) showed that NKA has 25% of the affinity of SP for the dog carotid artery, a preparation that contains only NK1 receptors. Tibério et al. (60) investigated the role of substance P (SP) and neurokinin A (NKA) and their receptor antagonists (RAs) SR140333 and SR48968 (respectively for the NK(1) and NK(2) receptors) in the pulmonary eosinophil influx induced by the stimulation of capsaicin (CAP)-sensitive nerve terminals. Both SP and NKA contribute to eosinophil lung recruitment in the distal airways and the alveolar wall, and these findings suggest that neurokinins may contribute to the development of eosinophilic inflammation in both allergic asthma and hypersensitive pneumonitis.

3.2. Cysteinyl leukotrienes

Cysteinyl leukotrienes (cysLTs) are synthesized *de novo* from arachidonic acid, and most of their actions are mediated by the CysLT1 receptor, a G protein-coupled receptor (63). Cys-LTs have many pulmonary actions, including human airway smooth muscle contraction, chemotaxis, mucous secretion, smooth muscle proliferation and increased vascular permeability (64-66).

The cysteinyl leukotrienes (LTC4, LTD4, LTE4) produced by inflammatory cells and endothelin, produced by epithelial or endothelial cells, are increased in asthma. They are also potent contractile agonists of ASM (48, 67). Leukotriene antagonists have been shown to reduce sputum and mucosal eosinophils in subjects with asthma (68, 69). However, recent long-duration trials have evaluated the impact of CysLT receptor antagonists compared with glucocorticoids and showed that spirometry, symptoms, β 2-agonist use and the quality of life were improved to a greater extent with glucocorticoids (70-72). Corroborating this idea, the blockade of leukotriene activity does not cause an improvement in airflow as intense as that obtained with glucocorticoids (70, 73).

Considering studies in animal models, Gardiner et al. (74) observed that the inhibition of leukotriene synthesis resulted in an attenuation of OVA-induced airway contraction in sensitized animals. Liu et al. (75) demonstrated that the CysLT1 receptor antagonists pranlukast and zafirlukast inhibited OVA-induced mucus secretion in the trachea of a sensitized guinea pig. Comparing the effects of montelukast and corticosteroid treatments in a guinea pig model, Leick-Maldonado et al. (76) showed that although montelukast, an antagonist of leukotriene, reduced some aspects of inflammation, this treatment was not able to attenuate the changes in lung mechanics.

3.3. Complex NOS-arginases

Nitric oxide derived either from constitutive isoforms (nNOS and eNOS) or from other NOadduct molecules (nitrosothiols) modulates bronchomotor and vascular tone. In addition, NO derived from inducible isoenzyme (iNOS) is mainly involved in the immunomodulation (77-80).

Prado et al. (81) tested the differences between chronic and acute nitric oxide inhibition by *N*-nitro-L-arginine methyl ester (L-NAME) treatment in lung mechanics, inflammation, and airway remodeling in an experimental asthma model in guinea pigs. Both acute and chronic L-NAME treatment reduced the exhaled nitric oxide in sensitized animals. Chronic L-NAME treatment increased the baseline and maximal responses after an antigen challenge (ovalbumin) of the respiratory system resistance and reduced peribronchial edema and airway infiltration by mononuclear cells. Acute administration of L-NAME increased the maximal values of respiratory system elastance and reduced the mononuclear cells and eosinophils in the airway wall, supporting the hypothesis that, in this model, nitric oxide acts as a bronchodilator in the airways.

iNOS enzyme activation has been found in many types of inflammatory cells, such as eosinophils, neutrophils and macrophages, as well as in respiratory epithelial cells. In fact, NO produced from this isoenzyme is related to the amplification of the inflammatory and remodeling responses (54, 78, 79, 82). Considering these aspects, a specific inhibition of iNOSderived NO has been considered to be a future therapeutic strategy for several diseases, such as asthma, sepsis and acute lung inflammation (82-85).

Considering the smooth muscle responses, NO mainly derived from cNOS relaxes the airway smooth muscle. Many studies have focused on the role of NO in the modulation of airway smooth muscle contraction in different models of experimental pulmonary allergic inflammation (78, 81, 85-87). NO that is mainly derived from the constitutive isoforms of NOS has been shown to attenuate the bronchoconstriction induced by allergens in sensitized experimental animals (54, 85, 88). In contrast, others have observed that nNOS-derived NO could contribute to airway constriction (61). We previously evaluated the effects of NO in respiratory system resistance using a guinea pig model of asthma and compared the cNOS and iNOS inhibition. We showed that chronic treatment with L-NAME, a false substrate that nonspecifically inhibits the production of NO, increased the respiratory system resistance in sensitized animals, whereas the iNOS-specific inhibition by 1400W reduced this response (54). Our results suggested a protective effect of NO derived from cNOS. In addition, we showed that iNOS contributes to the airway hyperresponsiveness in this model. Interestingly, in naïve animals, we observed that both L-NAME and 1400W treatments increased the resistance of the respiratory system. Because the role of iNOS is more pronounced in inflammatory situations, few studies have evaluated the effects of iNOS inhibition in physiologic situations. We have previously shown that there is a basal expression of iNOS in resident cells around the airways in guinea pigs not exposed to an inflammatory stimulus (54, 78). In addition, Guo and colleagues (89) showed that iNOS is continuously produced by the airway epithelium in normal humans. These data suggested that NO produced by iNOS under physiological conditions can also contribute to the control of the airway smooth muscle responses.

Analyzing the nitrergic nerve density, there appears to be a progressive reduction throughout the bronchial tree (90). In fact, Prado et al. (54) demonstrated that the inhibition of NO by chronic L-NAME treatment amplified the elastance responses. Considering that the respiratory system elastance responses are related to alterations in the distal airways and lung tissue, the authors suggested that NO could also be involved in the modulation of lung tissue constriction. Dupuy et al. (90) proposed that inhaled NO only affects the distal airways at high doses, suggesting that, although less intensive, NO can also modulate the responses of the distal airways and/or lung tissue.

Angeli et al. (11) evaluated the effects of chronic L-NAME treatment, a false substrate for all nitric oxide enzymes, on the modulation of lung tissue mechanics, eosinophilic inflammation and extracellular matrix tissue remodeling in guinea pigs with chronic lung inflammation. The authors suggested that nitric oxide plays an important role in lung tissue constriction and elastic fiber deposition within the alveolar septa in this animal model of chronic pulmonary inflammation. The activation of the pulmonary oxidative stress pathway, mainly via 8-iso-PGF2 α , may contribute to these responses.

Starling et al. (9) demonstrated that iNOS activation contributes to lung parenchyma inflammatory and remodeling alterations in guinea pigs with chronic pulmonary allergic inflammation. 1400W, an iNOS-specific inhibitor, diminished the lung tissue elastance and resistance as well as the eosinophilic infiltration, collagen and elastic fiber content and volume proportion of actin in lung tissue. To our knowledge, this study has provided the first evidence of the effects of iNOS inhibition on the distal lung parenchyma. In addition, the authors showed that specifically blocking iNOS reduced 8-isoprostane expression in the alveolar septa, which had previously been increased by repeated ovalbumin exposures (9). These findings suggest that the effects of iNOS-derived NO in the lung parenchyma depend, at least partially, on the activation of the oxidative stress pathway. The inhibition of NO production derived from iNOS activation also reduced the actin content (9). These results suggest an iNOS-derived effect on the myofibroblasts, which were believed to be the major cells responsible for the production of the extracellular matrix and the contraction of the parenchyma (91).

Another pathway to be discussed involves the arginases. These enzymes convert L-arginine into L-ornithine and urea and are the key enzymes of the urea cycle in the liver (arginase 1) but are also expressed in cells and tissues that lack a complete urea cycle, e.g., arginase 2 expression in the lung (88). Arginases are involved in cell growth and tissue repair via the increased production of L-ornithine, a precursor of polyamines and proline (88).

Que et al. (92) demonstrated the expression of arginase in the bronchial epithelium and in peribronchial connective tissue fibroblasts. In addition, Meurs et al. (87) showed that arginase appears to modulate the tone of the airway smooth muscle and potentiates methacholine-induced airway constriction. Arginase accomplishes these actions by forcing the common substrate L-arginine away from epithelial cNOS to diminish the agonist-induced production of NO. Arginases and NOS compete for the bioavailability of the same substrate, L-arginine, and are involved indirectly in the regulation of NO synthesis (53, 88). Corroborating this idea, Morris et al. (93) showed that there is a reduction in the levels of plasma arginine in asthmatic patients compared with patients without asthma but with increased serum arginase activity. Together, these results suggest that increased arginase activity in asthma may be a contributing factor to the decrease in the circulating levels of L-arginine and the consequent NO deficiency. Thus, blocking NO production could be a tool to study the indirect involvement of arginase in various pathophysiological processes (82, 87).

Several powerful drugs have been used to investigate the role of arginases in the pathophysiology of asthma, including nor-NOHA (N ω -hydroxy-nor-L-arginine), which is one of the most potent inhibitors of arginase (88). Meurs et al. (87), studying in vitro tracheal ring-sensitized guinea pigs, demonstrated that treatment with nor-NOHA reduced the hyperresponsiveness to methacholine, and this effect was reversed by treatment with L-NAME.

We demonstrated that chronic distal lung inflammation was associated with an increase in arginase content and iNOS-positive cells (data not published). These results were associated with constriction of the distal lung parenchyma. The increased iNOS expression leads to activation of the oxidative stress pathway and formation of PGF2 α , which had a procontractile effect. In addition, we showed that the mechanism involved in the activation of arginase and the iNOS pathways may be related to the modulation of NF-kB expression. Finally, we demonstrated that the association of iNOS and arginase 2 inhibitions potentiated the reduction of PGF2 α and NF-kB expression in the distal lung of guinea pigs with chronic pulmonary inflammation (data not published).

Airway inflammation is accompanied by a marked upregulation of iNOS expression, particularly in the airway epithelium (94), which has been associated with the activation of nuclear factor-kB (NF-kB), a transcription factor that is implicated in the induction of multiple genes expressed during the inflammatory response (95). Ckless et al. (96) showed that the activation of NF-kB may induce an increase in NOS and arginases. Furthermore, NF-κB activity can be affected by reactive oxygen species (ROS) and by reactive nitrogen species (RNS) (97).

Several mechanisms reported in the literature have tried to explain how NO could interfere with airway tone. The ability of NO to control airway tone could be related to both GMPc-dependent and GMPc-independent mechanisms (98-100).

Although the mechanisms involving the effects of NO in airway constriction have been extensively described, the exact mechanism involved in the effect of NOS inhibition on reducing lung parenchyma constriction is not completely understood. Another pathway discussed by some authors is related to the fact that the release of NO by NOS activation also contributes to oxidative stress, amplifying the deleterious and harmful effects on the lungs (9, 77).

The potent oxidant peroxynitrite is formed by the interaction of NO and superoxide by a rapid iso-stoichiometric reaction (77). Haddad et al. (101) suggested that peroxynitrite may contribute to the injury of pulmonary surfactant. Bhandari et al. (102) demonstrated that increased peroxynitrite formation was associated with a dose-dependent increase in the apoptotic cell death of type II pneumocytes. However, in strip preparations perfused with Krebs solution, the importance of reducing pulmonary surfactant was poorly associated with the pulmonary mechanical responses.

In contrast, peroxynitrite formation leads to lipid peroxidation and the generation of isoprostanes (8-iso-PGF_{2 α}). Jourdan et al. (103) showed that L-NAME treatment greatly inhibits 8-iso-PGF_{2 α}. Therefore, isoprostanes appear to induce airway and vascular smooth muscle contractions by acting through tyrosine kinase, Rho and Rho kinase, leading to the decreased activity of myosin light chain phosphatase. The net response is associated with an increased level of phosphorylated myosin light chain and contraction (104).

3.4. Rho kinase pathway

The protein Rho, a member of the Ras superfamily of small monomeric GTPases, controls a variety of downstream effector proteins, including Rho kinase. Rho exhibits GDP- and GTP-binding and GTPase activity and is able to alternate between a GDP-bound inactive state and a GTP-bound active state. This alternation allows Rho to function as a molecular switch to control downstream signal transduction, influencing the level of smooth muscle tone and changes in the actin cytoskeleton, which contributes to cell adhesion, motility, migration, and contraction (105). Effects on the airway smooth muscle responses may be one of the most important factors that need to be considered for the development of new therapies for asthmatics (106).

Current Basic and Pathological Approaches to 310 the Function of Muscle Cells and Tissues – From Molecules to Humans

The influence of Rho kinase on airway hyperresponsiveness is considered to be at least partly related to agonist-mediated Ca²⁺ sensitization. Ca²⁺ sensitization, which is also observed in the airways, is the increase in smooth muscle tension and/or phosphorylation of the 20-kDa regulatory light chain of myosin (MLC₂₀) at a constant Ca²⁺ concentration (107). In a variety of smooth muscles, this Ca²⁺ sensitization is mediated by a small G protein, RhoAp21, and its target protein, the Rho kinase (108), which is especially important during the sustained phase of contraction in smooth muscle (107).

Several studies have shown that the use of Rho kinase inhibitors might be beneficial for the treatment of airway diseases. Y-27632((+)-(R)-*trans*-4-(1-aminoethyl)-*N*-(4-pyrydil) cyclohexanecarboxamide, monohydrate) is one of the drugs that arose as a possible treatment for asthma. Y-27632 is a highly selective inhibitor of the Rho kinase pathway, capable of reversing G-protein sensitization and consequently relaxing the airway smooth muscle (108).

The effects of the acute inhibition of Rho kinase in sensitized animals have been analyzed by several authors. Schaafsma et al. (109) showed that the inhalation of Y-27632 at 30 min prevents the development of airway hyperresponsiveness both after the early and late airway reaction. Y-27632 reduces also reduces the cholinergic nerve-mediated contractions in the tracheal preparations of guinea pigs and mice in a dose-dependent manner (110). Witzenrath et al. (111) verified that the use of Y-27632 attenuated the methacholine-provoked airway response in the sensitized lungs.

Some studies suggested that the RhoA/ROCK system plays a role in eosinophil recruitment and Th-1 and Th-2 cytokine secretion (105, 112). In this regard, Henry et al. (112) demonstrated that pretreatment with Y-27632 reduced the number of eosinophils recovered from the bronchoalveolar lavage (BAL) fluid of OVA-sensitized mice.

Taki et al. (105) showed that another Rho kinase inhibitor, fasudil, reduced the presence of eosinophils in the BAL fluid, airways and blood vessels. In the BAL fluid, this Rho kinase inhibitor also diminished the augmented levels of IL-5, IL-13 and eotaxin. Aihara et al. (113) showed that Y-27632 suppressed the release of Th-1 cytokines and partially suppressed the release of Th-2 cytokines in healthy persons but reduced the release of IL-2 and IL-5 and weakly reduced the release of IL-4 and IFN-gamma in asthmatic patients.

Recently, we showed the chronic inhibition of Rho kinase reduced the airway and distal lung mechanical responses to an antigenic challenge with an associated reduction in NO_{EX}, eosinophilic infiltration, IL-2-, IL-4-, IL-5- and IL-13-positive cells, extracellular matrix remodeling and NF-κB-positive cells in the airways and distal lung. In addition, there was a significant reduction in the activation of the oxidative stress pathway, which was correlated with the attenuation of the maximal mechanical responses after antigen challenge (data not published).

These data suggest that treatment with an inhaled Rho kinase inhibitor contributes to the attenuation of the distal lung functional and structural changes induced by chronic allergic inflammation, both in the airways and distal lung. Taken together, this evidence suggests that Rho kinase inhibitors may be potential pharmacological tools to control distal lung asthmatic functional and histopathological alterations.

4. Modulators involved in airways and distal lung parenchyma contractile responses

4.1. Modulation of the lung contractile responses by physical exercises

The role of physical exercise in asthma is somewhat controversial. Exercise can induce bronchoconstriction in humans (114). Recently, however, various studies have shown that physical training, particularly at a moderate intensity, can improve lung function and is related to a reduction in asthma symptoms and AHR. Fanelli et al. (115) associated physical training improvements in the physiological variables at peak and submaximal exercise, and these authors also showed that trained patients have a reduction in the daily doses of inhaled steroids.

Studying adults, Mendes et al. (116) showed that 3 months after supervised training, patients presented a reduction in inflammation and asthma exacerbation and an increase in asthma symptom-free days. Although the authors did not directly measure the AHR, the reduction in symptoms and exacerbations indirectly reflects a reduction in the airway responsiveness. These authors clearly suggest that aerobic training might be useful as an adjuvant therapy in asthmatic patients under optimized medical care. In addition, physical training reduced the anxiety and depression levels with a significant correlation between improvements in the aerobic capacity and days without asthma symptoms (117).

Considering the experimental studies, Silva et al. (118) showed that aerobic training in mice with allergic chronic inflammation reduced both tissue elastance and resistance. These effects of aerobic training on lung mechanics could be at least partly mediated by the epithelium (119).

Based on these data, although AHR was frequently found among competitive athletes (120, 121), physical training may be beneficial to asthmatics, particularly when performed with supervision and at a moderate intensity.

4.2. Modulation of the lung contractile responses by stress

The stress response, which can be defined as the psychological reaction of the body to a variety of emotional or physical stimuli that threaten homeostasis (122), results in the activation of the hypothalamic-pituitary-adrenal (HPA) axis and the sympathetic and adrenomedullary systems. Although acute stress was shown to have anti-inflammatory effects, some studies have demonstrated that stressful situations and emotional states are triggers of asthmatic symptoms (123-125) and can influence the course and treatment of atopic diseases (126, 127). Chronic stress may induce a down-regulation of the expression and/or function of glucocorticoid receptors, leading to glucocorticoid resistance and contributing to the worsening of lung inflammation and pulmonary hyperreactivity.

Capelozzi et al. (128) showed that swimming-induced stress amplified mononuclear cell recruitment to the lungs in guinea pigs that performed 31 days of the stress protocol. These authors also showed that the amount of these cells was reduced when the animals were

treated with fluoxetine. Recently, Leick et al. (129), studying the effects of stress induced by forced swimming in bronchoconstriction, observed that stress amplified the airway response to ovalbumin in guinea pigs. In addition, Marques et al. (130) showed that the malefic effects of stress in asthma are related not only to the airways but to the lung distal parenchyma. In sensitized animals, they showed that repeated stress increased the distal lung constriction associated with an augmentation of actin content, which is indirect evidence of the alveolar smooth muscle content. The authors also showed that iNOS inhibition attenuated the effects of stress in the lung parenchyma response in this animal model.

Considering humans, Ritz and Steptoe (125) observed a negative association between mood states and a reduction in the forced expiratory volume in the first second in asthmatic patients. Höglund et al. (131) studied 41 undergraduate students 22 with allergies, 16 asthmatics and 19 controls in a low-stress period and in a period associated with a large exam. The values of the forced expiratory volume in the first second of the control group differed significantly from that of the group of asthmatics only during the exam stress phase. These results collectively reinforced the idea that stress is an important modulator of the AHR present in asthma.

Collectively, these studies showed that chronic stress is harmful to asthmatic individuals and is involved in the AHR.

4.3. Oral tolerance

Immunotherapy has been considered a possible therapeutic strategy for asthma. Oral tolerance has been recognized as an alternative treatment to autoimmune and allergic diseases (132-134). Oral tolerance has classically been defined as the specific suppression of the cellular and/or humoral immune response to an antigen by the prior administration of the antigen by the oral route (135). There are two primary effector mechanisms of oral tolerance: the induction of regulatory T cells that mediate the active suppression and the induction of clonal anergy or deletion (135-137). In atopic patients, the oral, sublingual, or inhaled administration of antigens leads to a reduction in symptoms and local inflammation as well as a reduction in dyspnea and airway hyperresponsiveness. Some meta-analyses found that sublingual immunotherapy is beneficial for asthma treatment, although the magnitude of the effect is not very large (138-140).

Some authors (141-143) have previously evaluated the effects of oral tolerance in experimental models of airway disease. In an animal model, oral tolerance induced an attenuation of airway eosinophilic recruitment, bronchial hyperresponsiveness, and mucous secretion (143, 144). Russo et al. (141, 142) observed that animals submitted to an oral antigen administration protocol presented low levels of Th2 cytokines in the bronchoalveolar lavage fluid and a reduction in the production of ovalbumin-specific antibodies. The tolerance process is known to attenuate B-cell responses. Hasegawa et al. (145) demonstrated that B-cells have been implicated in myofibroblast activation mainly by secreting IL-6, IL-9, and fibroblast growth factor. Thus, considering that myofibroblasts are one of the contractile elements that modulate lung parenchyma responses is important (146, 147).

Different Modulators of Airways and Distal Lung Parenchyma Contractile Responses in the Physiopathology of Asthma 313

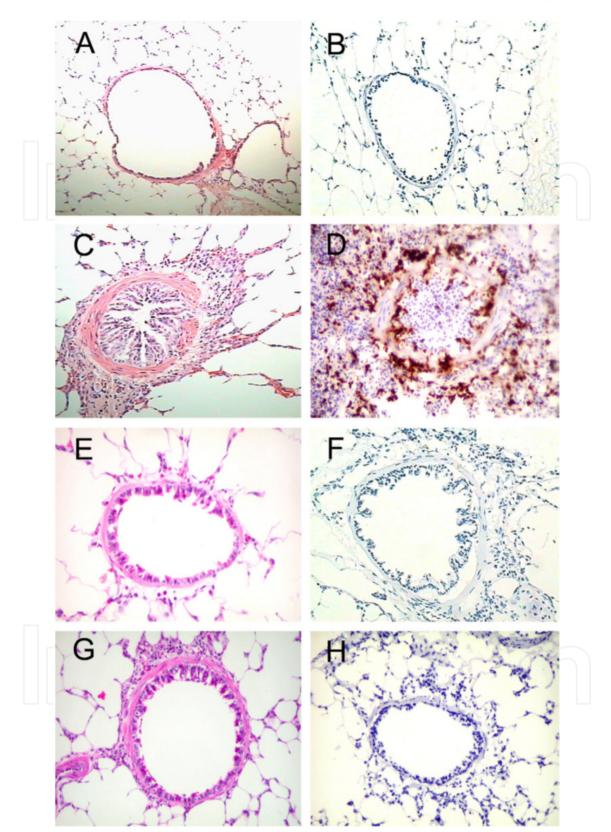


Figure 2. Photomicrographs of distal airways from the guinea pig (×200), stained with haematoxylineosin (left panels) and EPO+ eosinophils (right panels). Panels A and B: NS group. Panels C and D: OVA group. Panels E and F: OT1 group. Panels G and H: OT2 group. Reproduced with permission. *Published in Ruiz Schtüz et al.* (143).

Our group evaluated the airway responses in two different models of oral tolerance (ovalbumin-exposed and treat with oral tolerance beginning together with the 1st inhalation (OT1 group) and ovalbumin-exposed and treated with oral tolerance beginning after the 4th inhalation (OT2 group), and showed that both models counteract the bronchoconstriction induced by a specific antigen (ovalbumin) and by a nonspecific challenge using methacholine (143) (Figure 2). These data suggested that oral tolerance is an effective treatment to induce the relaxation of airway smooth muscle in asthma.

Although previous investigations showed that oral tolerance attenuated the airway responses, few studies have provided evidence of the effects of oral tolerance in lung periphery responses in an experimental model of chronic lung inflammation. In this regard, Nakashima et al. (43) showed that inducing oral tolerance attenuates peripheral lung tissue responsiveness, eosinophilic inflammation and extracellular matrix remodeling in an experimental model of chronic allergic pulmonary inflammation (Figure 3), suggesting that this approach could attenuate or prevent the distal lung functional and structural changes induced by chronic allergic inflammation.

5. Contribution of the airway and distal parenchyma structural changes to the pulmonary contractile responses.

The underlying persistent component of AHR, by contrast, is likely related to the structural (and/or physiological) airway changes often collectively referred to as airway remodeling. Structural changes in the airways and in the distal lung parenchyma, which were recently addressed, are involved in the remodeling process and include the epithelium basal membrane thickness, subepithelial fibrosis, mucous gland and goblet cell hypertrophy and hyperplasia, neoangiogenesis, increased ASM mass (hypertrophy of the smooth muscle cell and wall thickening), increased amount of actin and changes in the extracellular matrix (ECM), such as the deposition of fibronectin, laminin, and collagen fiber, alterations in the airway elastic fibers, and the increased expression of several metalloproteinases (MMP-1, MMP-2 and MMP-9) (45, 54). Such airway structural alterations or airway remodeling is associated with airway hyperresponsiveness to diverse triggers and with a decrease in the lung function of asthmatic patients.

In addition, an important structural change of the airways is related to the smooth muscle. One of the pathological consequences of remodeling is airway hyperresponsiveness. Myocyte hypertrophy and hyperplasia and myofibroblast hyperplasia are known to contribute to this hyperresponsiveness and the worsening of lung function in these patients (148). Throughout breathing, airway stiffening is a feasible contributor to airway hyperresponsiveness through the attenuation of the transmission of a potently bronchodilating cyclical stress to the ASM (37). ASM hyperplasia is characterized by a proliferation of cells, a reduction in the apoptosis of the ASM cells and migration of myofibroblasts within the ASM layer (19). Hence, alterations in the smooth muscle, either in the airways or in regions that are associated with perturbed alveolar attachments, may be factors that affect airwayparenchyma uncoupling and alterations in the mechanical properties of the distal lung that lead to constriction.

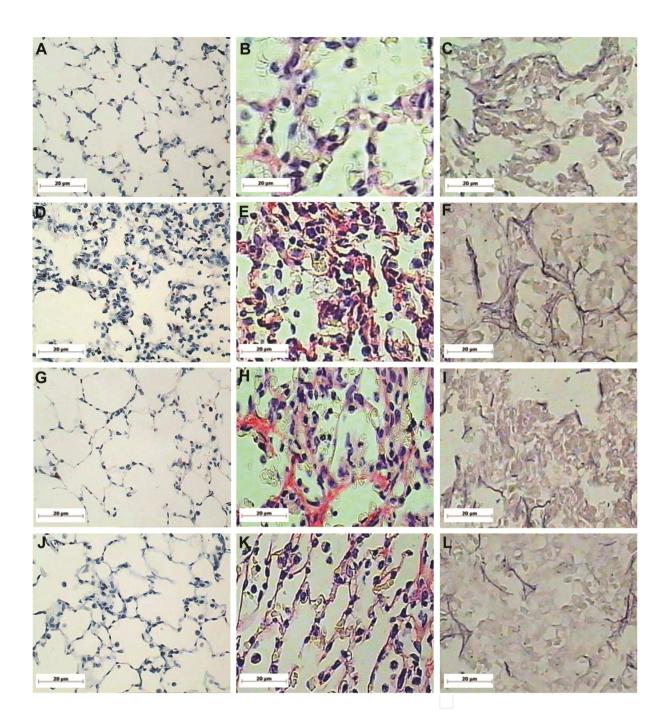


Figure 3. Photomicrographs of lung parenchymal strips eosinophilic infiltration (A, D, G, and J – x400), collagen density (B, E, H, and K – x1000) and elastic fibers (C, F, I, and L – x1000) in saline-exposed (NS group - panels A to C), ovalbumin-exposed (OVA group – panels D to F), ovalbumin-exposed and treat with oral tolerance beginning together with the 1st inhalation (OT1 group – panels G to I) and ovalbumin-exposed and treated with oral tolerance beginning after the 4th inhalation (OT2 group – panels J to L). Ovalbumin-exposed animals showed a significant increase in eosinophilic infiltration as well collagen and elastic density compared to saline-exposed ones. Both oral-induced tolerance protocols attenuated all these responses in ovalbumin-exposed animals. Reproduced with permission. *Published in Nakashima et al.* (43).

5.1. Mechanisms involved in lung remodeling

A chronic inflammatory process is almost invariably related to tissue damage and healing. The consequences of healing are repair and the replacement of injured cells by viable cells. Repair comprises regeneration (the replacement of damaged cells by cells of the same type) and replacement (by connective tissue). Chronic inflammatory processes have a wide varie-ty of consequences leading from the complete or partial restoration of the affected structure to fibrotic processes. The mechanisms underlying remodeling move from the highly dynamic process of cell migration, differentiation, and maturation to changes in the connective tissue deposition and to the altered restitution of the structures (149).

The airway epithelium constitutes a continuous physical barrier, crucial to maintaining tissue homeostasis, which lines the airway lumen and separates the underlying tissue from environmental antigens (150, 151). Currently, the airway epithelium is acknowledged to also sense and react to antigens by regulating innate (through pattern-recognition receptors, including Toll-like receptors [(TLRs]) and adaptive immune mechanisms, driving both allergic sensitization and airway remodeling through the release of inflammatory cytokines and chemokines. In addition, direct physical interactions with immune cells protect the internal milieu of the lung (152) and therefore contribute to airway narrowing. Furthermore, the increased loss of epithelial barrier integrity is known to correlate with more severe airway hyperresponsiveness, which may lead to the augmented exposure of the ASM to inhaled contractile agonists (153). Therefore, epithelial cells participate in a wide range of repair mechanisms, including the epithelization of the nude luminal surface, the production of chemotactic factors, and the expression of some surface markers and a broad range of molecules that participate in the tissue repair, such as fibronectin, growth factors, cytokines and chemokines (149).

One of the mechanisms that may account for ASM hyperplasia is the migration of myofibroblasts within the ASM layer, which differentiate into ASM-like cells (154). Fibroblasts differentiate into the highly synthetic and contractile myofibroblast phenotype when exposed to substrates with an elastic modulus corresponding to pathologically stiff fibrotic tissue. Myofibroblasts, which are cells that display features intermediate between fibroblasts and smooth muscle cells, are involved in this process and are able to synthesize several extracellular matrix substances and contract the lung parenchyma (155).

Although the hypertrophy in ASM has been described in studies with tissue specimens from intermittent, mild, severe (156) and fatal (45) asthma, which have been characterized as having an increase in the ASM cell size, there are conflicting findings (157) that suggest that the ASM cell hypertrophy could be a hallmark of severe asthma because it can be used to differentiate between patients with severe asthma and patients with milder disease (156). In asthmatics, ASM cell proliferation occurs faster than in nonasthmatics (27), and it can be explained by alterations in the calcium homeostasis in these cells and a subsequent increase in mitochondrial biogenesis (158).

The main characteristics of myofibroblasts are the secretion of extracellular matrix components, the development of adhesion structures with the substrate by the incorporation

of de novo expressed α -smooth muscle actin (α -SMA), and the formation of contractile bundles composed of actin and myosin, which help the myofibroblasts to develop a high contractile activity. These cytoskeletal features enable the myofibroblast to not only remodel and contract the extracellular matrix but also adapt its activity to changes in the mechanical microenvironment. In addition, immunohistochemistry and electron microscopy studies demonstrated that airway myofibroblasts and the smooth muscle bundles lie in close physical proximity in asthma (159, 160). The myofibroblasts have an intermediate phenotype between that of a fibroblast and that of a smooth muscle cell, which raises the possibility that these cells contribute to the increased smooth muscle mass because of their plasticity.

The arrangement and modification of the ECM involve dynamic processes of the production and degradation of matrix proteins, which are related to the ASM and parenchyma remodeling that are present and enhanced in asthma (161). The deposition of ECM proteins is increased by airway resident cells, such as epithelial cells, fibroblasts, myofibroblasts, and ASM cells. Some authors studying asthmatic bronchial samples demonstrated an increased deposition of ECM proteins in the bronchial wall, such as collagens I, III, and V, fibronectin, tenascin, hyaluronan, versican, laminin, lumican, and biglycan (162, 163), and a decreased deposition of collagen IV and elastin (164). Enhancing the ECM may be due to a reduced production of matrix metalloproteinases (MMPs), which degrade ECM proteins, and/or the enhanced production of tissue inhibitors of MMPs (TIMPs). Moreover, fibronectin and collagens III and V have been shown to enhance ASM migration (165) in the ASM cell contact with membranes coated with ECM components.

Notably, the epithelium in asthmatic children (aged 5-15 years) is stressed or injured without significant submucosal eosinophilic inflammation. This observation emphasizes the concept that the early pathological changes in asthma are linked to changes in the local tissue microenvironment related to epithelial stress and injury. The lamina reticularis from asthmatic biopsy sections was thicker than normal, with an increased deposition of collagen III. This alteration in the epithelial phenotype is associated with an enhanced collagen deposition in the lamina reticularis, suggesting that the epithelial mesenchymal trophic unit is active early in the natural history of asthma and may contribute to the pathogenesis of asthma (166).

ASM cells and the lung parenchyma have a crucial importance in the pathophysiology of asthma, leading to pulmonary remodeling, which remains unresponsive to conventional treatments, such as bronchodilators and anti-inflammatory drugs (167). Therefore, the development of new therapeutic tools targeting pulmonary remodeling is desirable.

6. Conclusions

ASM cells have a critical role in AHR in asthma, considering that these cells are part of the inflammatory process, have altered contractile, proliferative and secretory functions and contribute to airway remodeling.

Considering that many patients with AHR respond fairly well to conventional therapies, such as anti-inflammatory and bronchodilator drugs, and that ASM remodeling is insensitive to these treatments, further studies are necessary to evaluate ways to prevent or reverse ASM remodeling.

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7. References

- Brown RH, Pearse DB., Pyrgos G, Liu MC, Togias A, Permutt S. The Structural Basis of Airways Hyperresponsiveness in Asthma. Journal of Applied Physiology 2006;101 30-39.
- [2] Yamauchi K. Airway Remodeling in Asthma and its Influence on Clinical Pathophysiology. Tohoku Journal Experimental Medicine 2006;209 75-87.
- [3] Southam DS, Ellis R, Wattie J, Inman MD. Components of Airway Hyperresponsiveness and Their Associations with Inflammation and Remodeling In Mice. Journal of Allergy and Clinical Immunology 2007;119 848-854.
- [4] Kraft M, Djukanovic R, Wilson S, Holgate ST, Martin RJ. Alveolar Tissue Inflammation in Asthma. American Journal of Respiratory and Critical Care Medicine 1996;154 1505.
- [5] Kraft M. Part III: Location of Asthma Inflammation and the Distal Airways: Clinical Implications. Current Medical Research and Opinion 2007;3 S21-S27.
- [6] Martin RJ. Therapeutic Significance of Distal Airway Inflammation in Asthma. Journal of Allergy and Clinical Immunology 2002;109(2 Suppl) S447-S460.
- [7] Martin RJ. Exploring the Distal Lung: New Direction in Asthma. Israel Medical Association Journal 2008;10(12) 846-849.
- [8] Xisto DG, Farias LL, Ferreira HC, Pincanc, o MR, Amitrano D, Lapa e Silva JR, Negri EM, Carnielli D, F Silva LF, Capelozzi VL, Faffe DS, Zin WA, Rocco PM. Lung Parenchyma Remodeling in a Murine Model of Chronic Allergic Inflammation. American Journal of Respiratory and Critical Care Medicine 2005;171 829-837.
- [9] Starling CM, Prado CM, Leick-Maldonado EA, Lanças T, Reis FG, Aristóteles LR, Dolhnikoff M, Martins MA, Tibério IF: Inducible Nitric Oxide Synthase Inhibition Attenuates Lung Tissue Responsiveness and Remodeling in a Model of Chronic Pulmonary Inflammation in Guinea Pigs. Respiratory Physiology and Neurobiology 2009:165 185-194.

- [10] Lanças T, Kasahara DI, Prado CM, Tibério FLCI, Martins MA and Dolhnikoff M: Comparison of Early and Late Responses to Antigen of Sensitized Guinea Pig Parenchymal Lung Strips. Journal of Applied Physiology 2006;100(5) 1610-1616.
- [11] Angeli P, Prado CM, Xisto DG, Silva PL, Passaro CP, Nakazato HD, Leick-Maldonado EA, Martins MA, Rocco PR, Tiberio IF: Effects of Chronic L-NAME Treatment Lung Tissue Mechanics, Eosinophilic and Extracellular Matrix Responses Induced By Chronic Pulmonary Inflammation. American Journal Physiology and Lung Cell Molecular Physiology 2008;294 L1197-L1205.
- [12] Fredberg JJ, Stamenovic D. On the Imperfect Elasticity of Lung Tissue. Journal of Applied Physiology 1989;67 2408-2419.
- [13] Lauzon AM, Bates HT: Estimation of Time-Varying Respiratory Mechanical Parameters by Recursive Least Squares. Journal of Applied Physiology 1991;71 1159-1165.
- [14] Dolhnikoff M, Mauad T, Ludwig MS: Extracellular Matrix and Oscillatory Mechanics of Rat Lung Parenchyma in Bleomycin- Induced Fibrosis. American Journal of Respiratory and Critical Care Medicine 1999;160 1750-1757.
- [15] Dolhnikoff M, Morin J, Ludwig MS. Human Lung Parenchyma Responds to Contractile Stimulation. American Journal of Respiratory and Critical Care Medicine 1998;158 1607-1612.
- [16] Busse WW, Lemanske RF Jr. Asthma. New England Journal of Medicine. 2001;344(5) 350-362.
- [17] Zuyderduyn S, Sukkar MB, Fust A, Dhaliwal S, Burgess JK. Treating Asthma Means Treating Airway Smooth Muscle Cells. European Respiratory Journal 2008;32(2) 265-74.
- [18] Black JL, Roth M. Intrinsic Asthma: Is it Intrinsic to the Smooth Muscle? Clinical and Experimental Allergy, 2009;9(7) 962-965.
- [19] Ozier A, Allard B, Bara I, Girodet PO, Trian T, Marthan R, Berger P. The Pivotal Role of Airway Smooth Muscle in Asthma Pathophysiology. Journal of Allergy (Cairo) 2011;742710.
- [20] Bergeron C, Hauber HP, Gotfried M, Newman K, Dhanda R, Servi RJ, Ludwig M, Hamid Q. Evidence of Remodeling in Peripheral Airways of Patients with Mild to Moderate Asthma: Effect of Hydrofluoroalkane-Flunisolide. Journal of Allergy and Clinical Immunology 2005;116 983-989.
- [21] Micheletto C, Guerriero M, Tognella S, Dal Negro RW. Effects of HFA- and CFC-Beclomethasone Dipropionate on the Bronchial Response to Methacholine (Mch) in Mild Asthma. Respiratory Medicine 2005;99 850-855.
- [22] de Monchy JG, Kauffman HF, Venge P, et al. Bronchoalveolar Eosinophilia During Allergen-Induced Late Asthmatic Reactions. American Review and Respiratory Diseases 1985;131 373-376.
- [23] Hargreave FE. Late-Phase Asthmatic Responses and Airway Inflammation. Journal of Allergy and Clinical Immunology 1989;83 525-527.
- [24] Cockcroft DW, Davis BE. Mechanisms of Airway Hyperresponsiveness. Journal of Allergy and Clinical Immunology 2006;118 551-559.

Current Basic and Pathological Approaches to

320 the Function of Muscle Cells and Tissues – From Molecules to Humans

- [25] Chan V, Burgess JK, Ratoff JC, O'connor BJ, Greenough A, Lee TH, Hirst SJ. Extracellular Matrix Regulates Enhanced Eotaxin Expression in Asthmatic Airway Smooth Muscle Cells. American Journal and Respiratory Critical Care Medicine 2006;174(4) 379-385.
- [26] Johnson PR, Black JL, Carlin S, Ge Q, Underwood PA. The Production of Extracellular Matrix Proteins by Human Passively Sensitized Airway Smooth-Muscle Cells in Culture: The Effect of Beclomethasone. American Journal of Respiratory and Critical Care Medicine 2000;162: 2145-2151.
- [27] Johnson PR, Roth M, Tamm M, Hughes M, Ge Q, King G, Burgess JK, Black JL. Airway Smooth Muscle Cell Proliferation is Increased in Asthma. American Journal of Respiratory and Critical Care Medicine 2001;164(3) 474-477.
- [28] Burgess JK, Johnson PR, Ge Q, et al. Expression of Connective Tissue Growth Factor in Asthmatic Airway Smooth Muscle Cells. American Journal of Respiratory and Critical Care Medicine 2003;167 71-77.
- [29] Sukkar MB, Stanley AJ, Blake AE, et al. "Proliferative" and "Synthetic" Airway Smooth Muscle Cells Are Overlapping Populations. Immunology & Cell Biology 2004;82 471-478.
- [30] Jiang H, Rao K, Halayko AJ, Kepron W, Stephens NL. Bronchial Smooth Muscle Mechanics of a Canine Model of Allergic Airway Hyperresponsiveness. Journal of Applied Physiology 1992;72 39-45.
- [31] Ammit AJ, Bekir SS, Johnson PR, Hughes JM, Armour CL, Black JL. Mast Cell Numbers Are Increased in the Smooth Muscle of Human Sensitized Isolated Bronchi. American Journal of Respiratory and Critical Care Medicine 1997;155 1123-1129.
- [32] Brightling CE, Bradding P, Symon FA, Holgate ST, Wardlaw AJ, Pavord ID. Mast-Cell Infiltration of Airway Smooth Muscle in Asthma. New England Journal Medicine 2002;346 1699-1705.
- [33] Begueret H, Berger P, Vernejoux JM, Dubuisson L, Marthan R, Tunon-De-Lara JM. Inflammation of Bronchial Smooth Muscle in Allergic Asthma. Thorax 2007;62(1) 8-15.
- [34] Brightling CE, Ammit AJ, Kaur D et al. The CXCL10/ CXCR3 Axis Mediates Human Lung Mast Cell Migration to Asthmatic Airway Smooth Muscle. American Journal of Respiratory and Critical Care Medicine, 2005;171(10) 1103-1108.
- [35] El-Shazly A, Berger P, Girodet PO et al. Fraktalkine Produced By Airway Smooth Muscle Cells Contributes to Mast Cell Recruitment in Asthma. Journal of Immunology, 2006;176(3) 1860-1868.
- [36] Amin K, Janson C, Boman G, Venge P. The Extracellular Deposition of Mast Cell Products is Increased in Hypertrophic Airways Smooth Muscles in Allergic Asthma but not in Nonallergic Asthma. Allergy 2005;60(10) 1241-1247.
- [37] Siddiqui S, Martin JG. Structural Aspects of Airway Remodeling in Asthma. Current Allergy and Asthma Report 2008;8(6) 540-547.

- [38] Saha SK, Berry MA, Parker D et al. Increased Sputum and Bronchial Biopsy IL-13 Expression in Severe Asthma. Journal of Allergy and Clinical Immunology 2008;121(3) 685-691.
- [39] Liu MC, Bleecker ER, Lichtenstein LM, et al. Evidence for Elevated Levels of Histamine, Prostaglandin D2, and Other Bronchoconstriction Prostaglandins in the Airways of Subjects with Mild Asthma. *American Review of Respiratory Disease* 1990;142(1) 126-132.
- [40] Sterk PJ. The Place of Airway Hyperresponsiveness in the Asthma Phenotype. Clinical Experimental Allergy. 1995;25(2) 8-11; discussion 17-8.
- [41] Rocco PR, Facchinetti LD, Ferreira HC, Negri EM, Capelozzi VL, Faffe DS, Zin WA. Time Course of Respiratory Mechanics and Pulmonary Structural Remodelling in Acute Lung Injury. Respiratory Physiology Neurobiology 2004;143(1) 49-61.
- [42] Nagase T; Fukuchi Y; Dallaire MJ; Martin JG and Ludwig MS: In vitro Airway And Tissue Responses to Antigen in Sensitized Rats. American Journal of Respiratory and Critical Care Med 1995;153 81-86.
- [43] Nakashima AS, Prado CM, Lanças T, Ruiz VC, Kasahara DI, Leick-Maldonado EA, Dolhnikoff M, Martins MA, Tibério IF: Oral Tolerance Attenuates Changes in In Vitro Lung Tissue Mechanics and Extracellular Matrix Remodeling Induced by Chronic Allergic Inflammation in Guinea Pigs. Journal of Applied Physiology 2008;104(6) 1778-1785.
- [44] Romero PV, Rodriguez B, Lopez-Aguilar J, Manresa F. Parallel Airways Inhomogeneity and Lung Tissue Mechanics in Transition to Constricted State in Rabbits. Journal of Applied Physiology 1998;84 1040-1047.
- [45] Mauad T, Silva LF, Santos MA, Grinberg L, Bernardi FD, Martins MA, Saldiva PH, Dolhnikoff M. Abnormal Alveolar Attachments with Decreased Elastic Fiber Content in Distal Lung in Fatal Asthma. American Journal and Respiratory and Critical Care Medicine 2004;170 857-862.
- [46] Romero PV, Zin WA, Lopez-Aguilar J. Frequency Characteristics of Lung Tissue Strip during Passive Stretch and Induced Pneumoconstriction. Journal of Applied Physiology 2001;91 882-890.
- [47] Barnes NC, Piper PJ, Costello JF. Comparative Effects of Inhaled Leukotriene C4, Leukotriene D4, and Histamine in Normal Human Subjects. *Thorax* 1984;39(7) 500-504.
- [48] Perez-Zoghbi JF, Sanderson MJ. Endothelin-Induced Contraction of Bronchiole and Pulmonary Arteriole Smooth Muscle Cells is Regulated by Intracellular Ca2+ Oscillations and Ca2+ Sensitization. *American Journal of Physiology* 2007;293(4) L1000-L1011.
- [49] Howarth PH, Redington AE, Springall DR, et al. Epithelially Derived Endothelin and Nitric Oxide in Asthma. Interarch of Allergy and Immunology 1995;107(1-3) 228-230.
- [50] Gosens R, Zaagsma J, Grootte Bromhaar M, Nelemans A, Meurs H. Acetylcholine: A Novel Regulator of Airway Smooth Muscle Remodelling? European Journal of *Pharmacology* 2004;500(1-3) 193-201.

Current Basic and Pathological Approaches to

322 the Function of Muscle Cells and Tissues – From Molecules to Humans

- [51] Adcock IM, Peters M, Gelder C, Shirasaki H, Brown CR, Barnes PJ. Increased Tachykinin Receptor Gene Expression in Asthmatic Lung and its Modulation by Steroids. Journal Molecular and Endocrinology 1993;11(1) 1-7.
- [52] Chambers LS, Black JL, Ge Q et al. PAR-2 Activation, PGE2, and COX-2 in Human Asthmatic and Nonasthmatic Airway Smooth Muscle Cells. American Journal of Physiology 2003;285(3) L619-L627.
- [53] Meurs H, Hamer MAM, Pethe S, Goff SV, Boucher J, Zaagsma J: Modulation of Cholinergic Airway Reactivity and Nitric Oxide Production by Endogenous Arginase Activity. British Journal of Pharmacology 2000;130 1793-1798.
- [54] Prado CM; Leick- Maldonado EA; Yano L Leme AS; Capelozzi VL; Martins MA Tibério IF: Effects of Nitric Oxide Synthases in Chronic Allergic Airway Inflammation and Remodeling. American Journal of Physiology and Lung Cellular Molecular Biology 2006;35(4) 457-465.
- [55] Holgate S. Mechanisms of Allergy and Adult Asthma. Current Opinion in Allergy and Clinical Immunology 2001;1 47-50.
- [56] Numao T, Agrawal DK. Neuropeptides Modulate Human Eosinophil Chemotaxis. Journal of Immunology 1992;149 3309-3315.
- [57] Tibério IF, Turco GMG, Leick-Maldonado EA, Sakae RS, Paiva SO, do Patrocínio M, Warth TN, Lapa e Silva JR, Saldiva PH and Martins MA. Effects of Neurokinin Depletion on airway Inflammation Induced By Chronic Antigen Exposure. American Journal of Respiratory and Critical Care Medicine 1997;155 1739-1747.
- [58] Piedimonte G. Neural Mechanisms of Airway Syncytial Virus-Induced Inflammation and Prevention of Airway Syncytial Virus Sequelae. American Journal of Respiratory and Critical Care Medicine 2001;163 S18-S21.
- [59] O'Connor TM, O'Connell J, O'Brien DI, Goode T, Bredin CP, Shanahan F. The Role of Substance P in Inflammatory Disease. Journal of Cellular Physiology 2001, 167-180.
- [60] Tibério IF, Leick-Maldonado EA, Miyahara L, Kasahara DI, Spilborghs GM, Martins MA, Saldiva PH. Effects of Neurokinins on Airway and Alveolar Eosinophil Recruitment. Experimental Lung Research 2003;29(3) 165-177.
- [61] Prado CM, Leick-Maldonado EA, Miyamoto L, Yano LM, Kasahara DI, Martins MA, Tibério IF. Capsaicin-Sensitive Nerves and Neurokinins Modulate Non-Neuronal nNOS Expression in Lung. Respiratory Physiology and Neurobiology 2008;160(1) 37-44.
- [62] Regoli D., Drapeau G, Dion S, Couture R. New Selective Agonists for Neurokinin Receptors: Pharmacological Tools for Receptor Characterization. Trends Pharmacology Science 1988;9 290-295.
- [63] Holgate ST, Sampson AP. Antileukotriene Therapy: Future Directions. American Journal of Respiratory and Critical Care Medicine 2000;61 S147-S153.
- [64] O'Byrne PM. Leukotriene Bronchoconstriction Induced by Allergen and Exercises. American Journal of Respiratory and Critical Care Medicine 2000;161 S68-S72.
- [65] Drazen JM, Israel E, O'Byrne PM. Treatment of Asthma with Drugs Modifying the Leukotriene Pathway. New England Journal Medicine 1999;340 197-206.

- [66] Drazen JM. Leukotrienes as Mediators of Airway Obstruction. American Journal of Respiratory and Critical Care Medicine 1998;158 S193-S200.
- [67] Trakada G, Tsourapis S, Marangos M, Spiropoulos K. Arterial and Bronchoalveolar Lavage Fluid Endothelin-1 Concentration in Asthma. *Respiratory Medicine* 2000;94(10) 992-996.
- [68] Pizzichini E, Leff JA, Reiss TF et al. Montelukast Reduces Airway Eosinophilic Inflammation in Asthma: A Randomized, Controlled Trial. European Respiratory Journal 1999;14 12-18.
- [69] Nakamura Y, Hoshino M, Sim JJ, Ishii K, Hosaka K, Sakamoto T. Effect of the Leukotriene Receptor Antagonist Pranlukast on Cellular Infiltration in the Bronchial Mucosa of Patients with Asthma. Thorax 1998;53 835-841.
- [70] Smith LJ. Comparative Efficacy of Inhaled Corticosteroids and Antileukotriene Drugs in Asthma. Biodrugs 2001;15 239-249.
- [71] Malmstrom K, Rodriguez-Gomes G, Guerra J et al. Oral Montelukast, Inhaled Beclomethasone and Placebo for Chronic Asthma. Annals of Internal Medicine 1999;130 487-495.
- [72] Bleecker ER, Welch MJ, Weinstein SF et al. Low-Dose Inhaled Fluticasone Propionate Versus Oral Zafirlukast in the Treatment of Persistent Asthma. Journal of Allergy and Clinnical Immunology 2000;105 1123-1129.
- [73] Leff AR. Role of Leukotrienes in Bronchial Hyperresponsiveness and Cellular Responses In Airways. American Journal of Respiratory and Critical Care Medicine 2000;161 S125-S132.
- [74] Gardiner PJ, Cuthbert NJ, Francis HP et al. Inhibition of Antigeninduced Contraction of Guinea-Pig Airways by a Leukotriene Synthesis Inhibitor, BAY x1005. European Journal Pharmacology 1994;258 95-102.
- [75] Liu YC, Khawaja AM, Rogers DF. Effects of the Cysteinyl Leukotriene Receptor Antagonists Pranlukast and Zafirlukast on Tracheal Mucus Secretion in Ovalbumin-Sensitized Guinea-Pigs In Vitro. Brittish Journal of Pharmacology 1998;124 563-571.
- [76] Leick-Maldonado EA, Kay FU, Leonhardt MC, Kasahara DI, Prado CM, Fernandes FT, Martins MA and Tibério IFLC: Comparison of Glucocorticoid and Cysteinyl Leukotriene Receptor Antagonist Treatments in an Experimental Model of Chronic Airway Inflammation in Guinea-Pigs. Clinical Experimental Allergy 2004,34 145-152.
- [77] Ricciardolo FLM: Multiple Roles of Nitric Oxide in the Airways. Thorax 2003;58 175-182.
- [78] Prado CM, Leick-Maldonado EA, Arata V, Kasahara DI, Martins MA, Tibério IF. Neurokinins and Inflammatory Cell iNOS Expression in Guinea Pigs with Chronic Allergic Airway Inflammation. American Journal of Physiology and Lung Cell Molecular Physiology 2005;288(4) L741-L748.
- [79] Prado CM, Martins MA, Tiberio IF. Nitric Oxide in Asthma Physiopathology. International Scholarly Research Network. ISRN Allergy. 2011, 13 pages.

- [80] Prado CM, Yano L, Rocha G, Starling CM, Capelozzi VL, Leick-Maldonado EA, Martins M de A, Tibério IF. Effects of Inducible Nitric Oxide Synthase Inhibition in Bronchial Vascular Remodeling-Induced by Chronic Allergic Pulmonary Inflammation. Experimental Lung Research 2011;137(5) 259-268.
- [81] Prado CM, Leick-Maldonado EA, Kasahara DI, Capelozzi VL, Martins MA, Tiberio IF. Effects of Acute and Chronic Nitric Oxide Inhibition in an Experimental Model of Chronic Pulmonary Allergic Inflammation in Guinea Pigs. American Journal Physiology Lung Cellular Mollecular Biology 2005;289 L677-L683.
- [82] Ricciardollo FL, Sterk PJ, Gaston B, Folkers G. Nitric Oxide in Health and Disease of the Respiratory System. Physiology Review 2004;84(3) 731-765.
- [83] Garvey EP, Oplinger JA, Furfine ES, Kiff RJ, Laszlo F, Whittle BJR and Knowel RG: 1400W is a slow, Tight Binding and Highly Selective Inhibitor of Inducible Nitric Oxide Synthase *In Vitro* and *In Vivo*. Journal of Biochemistry 1997;272(8) 4959-63.
- [84] De Boer J, Meurs H, Coers W, Koopal M, Bottone AE, Visser AC, Timens W, Zaagsma J: Deficiency of Nitric Oxide in Allergen-Induced Airway Hyperreactivity to Contractile Agonists after the Early Asthmatic Reaction: An Ex Vivo Study. Brittish Journal of Pharmacol 1996;119 1109-1116.
- [85] Koarai A, Ichinose M, Sugiura H, Tomaki M, Watanabe M, Yamagata S, Komaki Y, Shirato K, Hattori T. iNOS Depletion Completely Diminishes Reactive Nitrogen-Species Formation after an Allergic Response. European Respiratory Journal 2002;20(3) 609-616.
- [86] Eynott PR, Groneberg DA, Caramori G, Adcock IM, Donnely LE, Kharitonov S, Barnes PL, Chung KF. Role of Nitric Oxide in Allergic Inflammation and Bronchial Hyperresponsiveness. European Journal of Pharmacology 2002;452 123-133.
- [87] Meurs H, McKay S, Maarsingh H, Hamer M , Macic L, Molendijk N, Zaagsma J: Increased Arginase Activity Underlies Allergen-Induced Deficiency of cNOS Derived Nitric Oxide and Airway Hyperresponsiveness. Brittish Journal of Pharmacology 2002;136 391-398.
- [88] Meurs H, Maarsingh H, Zaagsma J. Arginase and Asthma: Novel Insights into Nitric Oxide Homeostasis and Airway Hyperresponsiveness. Trends Pharmacology in Science 2003;24 450-455.
- [89] Guo FH, De Raeve HR, Rice TW, Stuehr DJ, Thunnissen FBJM, Erzurum SC. Continuous Nitric Oxide Synthesis by Inducible Nitric Oxide Synthase in Normal Human Airway Epithelium In Vivo. Proceedings of the National Academy of Science USA 1995;92 7809-7813.
- [90] Dupuy PM, Shore SA, Drazen JM, Frostell C, Hill Zapol WA. Bronchodilator Action of inhaled Nitric Oxide in Guinea Pigs. Journal of Clinical Investigation 1992;90 421-442.
- [91] Hsu YC, Wang LF, Chien YW. Nitric Oxide In The Pathogenesis of Diffuse Pulmonary Fibrosis. Free Radical Biology Medicine 2007;42 599-607.
- [92] Que LG, George SE, Gotoh T, Mori M, Huang YC. Effects of Arginase Isoforms on NO Production by nNOS. Nitric Oxide 2002;6(1) 1-8.

- [93] Morris CR, Poljakovic M, Lavrisha L, Machado L, Kuypers FA, Morris SM: Decreased Arginine Bioavailability and Increased Serum Arginase Activity in Asthma. American Journal and Respiratory and Critical Care Medicine 2004;170 148-53.
- [94] Hamid Q, Springall DR, Riveros-Moreno V, Chanez P, Howarth P, Redington A, Bousquet J, Godard P, Holgate S, Polak JM. Induction of Nitric Oxide Synthase in Asthma. Lancet 1993;342 1510-1513.
- [95] Ckless K, Lampert, Reiss J, Kasahara D, Poynter ME, Irvin CG, Lundblad LKA, Norton R, Vliet A, Janssen-Heininger YMW. Inhibition of Arginase Activity Enhances Inflammation in Mice with Allergic Airway Disease, in Association with Increases in Protein S-Nitrosylation and Tyrosine Nitration1. The Journal of Immunology 2008;181 4255-4264.
- [96] Ckless K, Van der Vliet A, Janssen-Heininger Y. Oxidative Nitrosative Stress and Post-Translational Protein Modifications: Implications to Lung Structure-Function Relations. Arginase Modulates NF-kappaB Activity Via a Nitric Oxide-Dependent Mechanism. American Journal and Respiratory Cellular Mollecular Biology 2007;36 645-53.
- [97] Poynter ME, Cloots R, Van Woerkom T, Butnor KJ, Vacek P, Taatjes DJ, Irvin CG, Janssen-Heininger YM: NF-kB: Activation in airways modulates allergic inflammation but not hyperresponsiveness. Journal of Immunology 2004;173 7003–7009.
- [98] Bannenberg G, Xue J, Engman L, Cotgreave I, Moldeus P, Ryrfeldt A. Characterization of Bronchodilator Effects and Fate of S-nitrosothiols in the Isolated Perfused and Ventilated Guinea Pig Lung. *Journal* of *Pharmacology* and Experimental Therapeutics 1995;272 1238-1245.
- [99] Perkins WJ, Pabelick C, Warner DO, Jones KA. cGMP-Independent Mechanism of Airway Smooth Muscle Relaxation Induced by S-Nitrosoglutathione. American Journal of Physiology 1998;275 C468-C474.
- [100] Janssen LJ, Premji M, Lu-Chao H, Cox G, Keshavjee S. NO but not NO Radical Relaxes Airway Smooth Muscle via cGMP-Independent Release of Internal Ca(2+). American Journal of Physiology Lung Cellular Molecular Physiology 2000;278 L899-L905.
- [101] Haddad IY, Ischiropoulos H, Holm BA, Beckman JS, Baker JR, Matalon S. Mechanisms of Peroxynitrite-Induced Injury to Pulmonary Surfactants. American Journal Physiology 1993;265 L555-L564.
- [102] Bhandari V, Johnson L, Smith-Kirwin S, Vigliotta G, Funanage V, Chander A. Hyperoxia and Nitric Oxide Reduce Surfactant Components (DSPC and Surfactant Proteins) and Increase Apoptosis in Adult and Fetal Rat Type II Pneumocytes. Lung 2002;180(6) 301-317.
- [103] Jourdan KB, Mitchell JA, Evans TW. Release of Isoprostanes by Human Pulmonary Artery in Organ Culture: A Cyclo-Oxygenase and Nitric Oxide Dependent Pathway. Biochemical and Biophysical Research Communications 1997;233(3) 668-672.
- [104] Janssen LJ. Isoprostanes: An Overview and Putative Roles in Pulmonary Pathophysiology. American Journal of Physiology and Lung Cellular Molecular Physiology 2001;280(6) L1067-L1082.

- [105] Taki F, Kume H, Kobayashi T, Ohta H, Aratake H, Shimokata K. Effects of Rho-Kinase Inactivation on Eosinophilia and Hyper-Reactivity In Murine Airways by Allergen Challenges. Clinical and Experimental Allergy 2007;37 599-607.
- [106] Gosens R, Schaafsma D, Grootte Bromhaar MM, Vrugt B, Zaagsma J, Meurs H, Nelemans SA. Growth Factor-Induced Contraction of Human Bronchial Smooth Muscle is Rho-Kinase-Dependent. European Journal of Pharmacology 2004;494 73-76.
- [107] Heasman SJ, Ridley AJ. Multiple Roles for RhoA during T Cell Transendothelial Migration. Small Gtpases 2010;1(3) 174-179.
- [108] Kume H. RhoA/Rho-kinase as a Therapeutic Target in Asthma. Current Medicinal Chemistry 2008;15(27) 2876-2885.
- [109] Schaafsma D, Bos ST, Zuidhof AB, Zaagsma J, Meurs H. The Inhaled Rho Kinase Inhibitor Y-27632 Protects Against Allergen-Induced Acute Bronchoconstriction, Airway Hyperresponsiveness, and Inflammation. American Journal of Physiology and Lung Cellular Molecular Physiology 2008;295 L214-L219.
- [110] Fernandes L, D'Aprile A, Self G, McGuire M, Sew T, Henry P, Goldie R A Rho-Kinase Inhibitor, Y-27632, Reduces Cholinergic Contraction but not Neurotransmitter Release. European Journal Pharmacology. 2006;550(1-3) 155-161.
- [111] Witzenrath M, Ahrens B, Schmeck B, Kube SM, Hippenstiel S, Rosseau S, Hamelmann E, Suttorp N, Schütte H. Rho-Kinase and Contractile Apparatus Proteins in Murine Airway Hyperresponsiveness. Toxicology Pathology 2008;60(1) 9-15.
- [112] Henry PJ, Mann TS, Goldie RG. A Rho Kinase Inhibitor, Y-27632 Inhibits Pulmonary Eosinophilia, Bronchoconstriction and Airways Hyperresponsiveness in Allergic Mice. Pulmonary Pharmacology Therapeutics 2005;18(1) 67-74.
- [113] Aihara M, Dobashi K, Iizuka K, Nakazawa T, Mori M. Effect of Y-27632 on Release of Cytokines from Peripheral T Cells in Asthmatic Patients and Normal Subjects. International Immunopharmacology 2004;4 557-561.
- [114] Spector S, Tan R. Exercise-Induced Bronchoconstriction Update: Therapeutic Management. Allergy Asthma Proceedings 2012;33(1) 7-12.
- [115] Fanelli A, Cabral AL, Neder JA, Martins MA, Carvalho CR. Exercise Training on Disease Control and Quality of Life in Asthmatic Children. Medicine and Science Sports Exercise. 2007;39(9) 1474-1480.
- [116] Mendes FA, Almeida FM, Cukier A, Stelmach R, Jacob-Filho W, Martins MA, Carvalho CR. Effects of Aerobic Training on Airway Inflammation in Asthmatic Patients. Medicine and Science Sports Exercise 2011;43(2) 197-203.
- [117] Mendes FA, Gonçalves RC, Nunes MP, Saraiva-Romanholo BM, Cukier A, Stelmach R, Jacob-Filho W, Martins MA, Carvalho CR. Effects of Aerobic Training on Psychosocial Morbidity and Symptoms in Patients with Asthma: A Randomized Clinical Trial. Chest 2010;138(2) 331-337.
- [118] Silva RA, Vieira RP, Duarte AC, Lopes FD, Perini A, Mauad T, Martins MA, Carvalho CR. Aerobic Training Reverses Airway Inflammation and Remodelling in an Asthma Murine Model. European Respiratory Journal 2010;35(5) 994-1002.

- [119] Vieira RP, Toledo AC, Ferreira SC, Santos AB, Medeiros MC, Hage M, Mauad T, Martins Mde A, Dolhnikoff M, Carvalho CR. Airway Epithelium Mediates the Anti-Inflammatory Effects of Exercise on Asthma. Respiratory Physiology and Neurobiology 2011;175(3) 383-9.
- [120] Stadelmann K, Stensrud T, Carlsen KH. Respiratory Symptoms and Bronchial Responsiveness in Competitive Swimmers. Medicine and Science Sports Exercise 2011;43(3) 375-381.
- [121] Lund TK. Asthma in Elite Athletes: How do we Manage Asthma-Like Symptoms and Asthma in Elite Athletes? Clinnical Respiratory Journal 2009;3(2) 123.
- [122] Forsythe P, Ebeling C, Gordon JR, Befus AD, Vliagoftis H: Opposing Effects of Shortand Asthma. Allergy Asthma Proceedings 2000;21 241-246.
- [123] Webster EL, Torpy DJ, Elenkov IJ, Chrousos GP: Corticotropin-Releasing Hormone and Inflammation. Annual NY Academic Science 1998;840 21-32.
- [124] Ritz T, Steptoe A, DeWilde S, Costa M: Emotions and Stress Increase Respiratory Resistance in Asthma. Psychosomatic Medicine 2000;62 401-412.
- [125] Ritz T, Steptoe A: Emotion and Pulmonary Function in Asthma: Reactivity in the Field and Relationship with Laboratory Induction of Emotion. Psychosomatic Medicine 2000;62 808-815.
- [126] Marshall GD Jr, Agarwal SK: Stress, Immune Regulation, and Immunity: Applications for Long-Term Stress on Airway Inflammation. American Journal and Respiratoory Critical Care Med 2004;169 220-226.
- [127] Miller GE, Cohen S, Ritchey AK: Chronic Psychological Stress and the Regulation of Pro-Inflammatory Cytokines: A Glucocorticoid-Resistance Model. Health Psychology 2002;21 531-541.
- [128] Capelozzi MA, Leick-Maldonado EA, Parra ER, Martins MA, Tibério IF, Capelozzi VL: Morphological and Functional Determinants of Fluoxetine (Prozac)-Induced Pulmonary Disease in an Experimental Model. Respiratory Physiology and Neurobiology 2007;156 171-178.
- [129] Leick EA, Reis FG, Honorio-Neves FA, Almeida-Reis R, Prado CM, Martins MA, Tibério IF. Effects of Repeated Stress on Distal Airway Inflammation, Remodeling and Mechanics in an Animal Model of Chronic Airway Inflammation. Neuroimmunomodulation 2012;19(1) 1-9.
- [130] Marques RH, Reis FG, Starling CM, Cabido C, de Almeida-Reis R, Dohlnikoff M, Prado CM, Leick EA, Martins MA, Tibério IF. Inducible Nitric Oxide Synthase Inhibition Attenuates Physical Stress-Induced Lung Hyper-Responsiveness and Oxidative Stress in Animals with Lung Inflammation. Neuroimmunomodulation 2012;19(3) 158-170.
- [131] Höglund CO, Axen J, Kemi C, Jernelov S, Grunewald J, Muller-Suur C, Smith Y, Gronneberg R, Eklund A, Stierna P, Lekander M: Changes in Immune Regulation in Response to Examination Stress in Atopic and Healthy Individuals. Clinical Experimental Allergy 2006;36 982-992.

- [132] Karlsson MR, Kahu H, Hanson LA, Telemo E, Dahlgren UI. Tolerance and Bystander Suppression, with Involvement of CD25-Positive Cells, is Induced in Rats Receiving Serum from Ovalbumin-Fed Donors. Immunology 2000;100 326-333.
- [133] Smith KM, Eaton AD, Finlayson LM, Garside P. Oral Tolerance. American Journal and Respiratoory Critical Care Medicine 2000;162 175-178.
- [134] Weiner HL. Oral Tolerance: Immune Mechanisms and Treatment of Autoimmune Diseases. Immunology Today 1997;18 335-343.
- [135] Faria AM, Weiner HL. Oral Tolerance. Immunology Review 2005;206 232-259.
- [136] Garside P, Millington O, Smith KM. The Anatomy of Mucosal Immune Responses. Annual NY Academic Science 2004;1029 9-15.
- [137] Dubois B, Goubier A, Joubert G, Kaiserlian D. Oral Tolerance and Regulation of Mucosal Immunity. Cellular Molecular Life Science 2005;62 1322-1332.
- [138] Abramson MJ, Puy RM, Weiner JM. Allergen Immunotherapy for Asthma. American Journal and Respiratory and Critical Care Medicine 1999;160 1750-1757.
- [139] Calamita Z, Saconato H, Pela AB, Atallah AN. Efficacy of Sublingual Immunotherapy in Asthma: Systematic Review of Randomized-Clinical Trials using the Cochrane Collaboration Method. Allergy 2006;61 1162-1172.
- [140] Sopo SM, Macchiaiolo M, Zorzi G, Tripodi S. Sublingual Immunotherapy in Asthma and Rhinoconjunctivitis; Systematic Review of Paediatric Literature. Archives of Diseaes in Childhood 2004;89 620-624.
- [141] Russo M, Jancar S, Siqueira ALP, Mengel J, Gomes E, Ficker SM, Faria AMC. Prevention of Lung Eosinophilic Inflammation by Oral Tolerance. Immunology Letters 1998;61 15-23.
- [142] Russo M, Nahori MA, Lefort J, Gomes E, Keller AC, Rodriguez D, Ribeiro OG, Adriouch S, Gallois V, de Faria AM, Vargaftig BB. Suppression of Asthma-Like Responses in Different Mouse Strains by Oral Tolerance. American Journal and Respiratory and Cellular Molecular Biology 2001;24 518-526.
- [143] Ruiz-Schütz VC, Drewiacki T, Nakashima AS, Arantes-Costa FM, Prado CM, Kasahara DI, EA, Martins M de A, Tibério IF. Oral Tolerance Attenuates Airway Inflammation and Remodeling in a Model of Chronic Pulmonary Allergic Inflammation. Respiratory Physiology and Neurobiology 2009;165 13-21.
- [144] Chung Y, Cho J, Chang YS, Cho SH, Kang CY. Preventive and Therapeutic Effects of Oral Tolerance in a Murine Model of Asthma. Immunobiology 2002;206 408-423.
- [145] Hasegawa M, Fujimoto M, Takehara K, Sato S. Pathogenesis of Systemic Sclerosis: Altered B Cell Function is the Key Linking Systemic Autoimmunity and Tissue Fibrosis. Journal of Dermatological Science 2005;39 1-7.
- [146] Ludwig MS, Dallaire MJ. Structural Composition of Lung Parenchymal Strip and Mechanical Behavior during Sinusoidal Oscillation. Journal of Applied Physiology 1994;77 2029-2035.
- [147] Wynn TA. Cellular and Molecular Mechanisms of Fibrosis. Journal of Pathology 2008;214 199-210.

- [148] Mehrotra AK, Henderson WR Jr. The Role of Leukotrienes in Airway Remodeling. Currents in Molecular Medicine 2009:9(3) 383-391.
- [149] Bergeron C, Al-Ramli W, Hamid Q. Remodeling in Asthma. Proceedings of American Thoracic Society 2009;6(3) 301-305.
- [150] Swindle EJ, Collins JE, Davies DE. Breakdown in Epithelial Barrier Function in Patients with Asthma: Identification of Novel Therapeutic Approaches. Journal of Allergy Clinical Immunology 2009;124 23-34.
- [151] Hackett TL. Epithelial-Mesenchymal Transition in the Pathophysiology of Airway Remodelling in Asthma. Currents Opinion Allergy Clinical Immunology 2012;12(1) 53-59.
- [152] Nawijn MC, Hackett TL, Postma DS, et al. E-Cadherin: Gatekeeper of Airway Mucosa and Allergic Sensitization. Trends Immunology 2011;32 248-255.
- [153] Holgate ST, Roberts G, Arshad HS, Howarth PH, Davies DE. The Role of the Airway Epithelium and its Interaction with Environmental Factors in Asthma Pathogenesis. *Proceedings of the American Thoracic Society* 2009;6(8) 655-659.
- [154] Gerthoffer WT. Migration of Airway Smooth Muscle Cells. Proceedings of the American Thoracic Society 2008;5(1) 97-105.
- [155] Zhang HY, Phan SH. Inhibition of Myofibroblast Apoptosis by Transforming Growth Factor Beta-1. American Journal of Respiratory and Cellular and Molecular Biology 1999;21 658-665.
- [156] Benayoun L, Druilhe A, Dombret MC, Aubier M, and Pretolani M. Airway Structural Alterations Selectively Associated with Severe Asthma. American Journal of Respiratory and Critical Care Medicine 2003;167(10) 1360-1368.
- [157] Woodruff PG, Dolganov GM, Ferrando RE et al. Hyperplasia of Smooth Muscle in Mild to Moderate Asthma without Changes in Cell Size or Gene Expression. American Journal of Respiratory and Critical Care Medicine 2004;169(9) 1001-1006.
- [158] Trian T, Benard G, Begueret H et al. Bronchial Smooth Muscle Remodeling Involves Calcium-Dependent Enhanced Mitochondrial Biogenesis in Asthma. Journal of Experimental Medicine, 2007;204(13) 3173-3181.
- [159] Jeffery P. Structural Alterations and Inflammation of Bronchi in Asthma. Int Journal Clin Pract Suppl 1998;96 5-14.
- [160] Richter A, Puddicombe SM, Lordan JL, Bucchieri F, Wilson SJ, Djukanovic R. The Contribution of Interleukin IL-4 and IL-13 to the Epithelial-Mesenchymal Trophic Unit in Asthma. Am J Respir Cell Mol Biol 2001;25 385-391.
- [161] James A. Remodelling of Airway Smooth Muscle in Asthma: What Sort do You Have? Clinical Experimental Allergy 2005;35(6) 703-707. Review.
- [162] Laitinen LA, Laitinen A, Altraja A, et al. Bronchial Biopsy Findings in Intermittent or "Early" Asthma. Journal of Allergy and Clinical Immunology 1996;98(5 part 2) S33-S40.
- [163] Roberts CR. Remodelling of the Extracellular Matrix in Asthma: Proteoglycan Synthesis and Degradation. Canadian Respiratory Journal 1998;5(1) 48-50.

Current Basic and Pathological Approaches to 330 the Function of Muscle Cells and Tissues – From Molecules to Humans

- [164] Bousquet J, Chanez P, Lacoste JY et al. Asthma: A Disease Remodeling the Airways. Allergy 1992;47(1) 3-11.
- [165] Parameswaran K, Radford K, Zuo J, Janssen LJ, O' Byrne PM, Cox PG. Extracellular Matrix Regulates Human Airway Smooth Muscle Cell Migration. European Respiratory Journal 2004;24(4) 545-551.
- [166] Fedorov IA, Wilson SJ, Davies DE, Holgate ST. Epithelial Stress and Structural Remodelling in Childhood Asthma. Thorax 2005;60 389-394.
- [167] Girodet PO, Ozier A, Bara I, Tunon De Lara JM, Marthan R, Berger P. Airway Remodeling in Asthma: New Mechanisms and Potential for Pharmacological Intervention. Pharmacology Therapeutics 2011;130(3) 325-337
- Adcock IM, Peters M, Gelder C, Shirasaki H, Brown CR, Barnes PJ. Increased Tachykinin Receptor Gene Expression in Asthmatic Lung and its Modulation by Steroids. *Journal of Molecular Endocrinology* 1993;11(1) 1-7.

