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Leflunomide an Immunosuppressive Drug for Antiviral Purpose in Treatment for BK Virus-Associated Nephropathy After Kidney Transplantation

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1. Introduction

1.1 BK Virus

BK virus is a polyomavirus belonging to the *papovaviridae* branch. In addition to BK, the human polyomavirus family includes John Cunningham virus (JCV), Washington University virus (WUV), Karolinska Institute virus (KIV) and Merkel cell viruses (Boothpur et al. 2010). BK virus is a virus without a shell and it has a double-stranded circular non-enveloped DNA. It was first discovered and isolated in 1971 just like JC virus, responsible for Progressive Multifocal Leukoencephalopathy (PML). Contamination usually occurs during early childhood through the airway without clinical symptoms. BK virus seroprevalence in general population is around 60%. The main latency areas are the kidney and the urothelium. Asymptomatic BK virus infection is often acquired in childhood and the virus persists in a dormant state in urothelium and kidneys of healthy and immunocompetent individuals, where it can be reactivated under immunosuppression (Nickeleit et al. 2000a; Brocker et al. 2011).

1.2 Prevalence and incidence

Urinary viral prevalence for BK virus is between 0.3% and 6% in general population, and increases in functions of immunosuppression degree; between 10% and 45% in patients after renal transplant, 30% in patients after bone marrow graft and 25% in patients with Human immunodeficiency virus. In patients with renal graft, the annual incidence of the nephropathy is between 3% and 5% (Randhawa et al. 2000; Pavlakis et al. 2006).

1.3 Risk factors

BK virus-associated nephropathy seems to be promoted by the concurrent presence of several risk factors. The immunosuppressive regimen strength, with high level blood concentrations, is the first factor involved. Most patients affected by BK virus-associated nephropathy previously had an intensification of immunosuppressive regimen due to a rejection event or a treatment including tacrolimus and/or mycophenolate mofetil

combined with monoclonal or polyclonal antibodies (De Luca et al. 2000; Nickeleit et al. 2000b; Randhawa et al. 2000; Hirsch et al. 2001). Conversely, no cases have been reported in patients treated with cyclosporine and corticosteroids (Binet et al. 1999; Mengel et al. 2003).

The other risk factors identified comprise donor characteristics, such as female gender, deceased donation, ischemia-reperfusion injury, high BK virus specific antibody titres, HLA mismatch and African-American ethnicity. The recipient characteristics in cause are older age, male gender, white race, diabetes, obesity, retransplantation, lack of HLA C-7, low or absent BK virus specific T-cell activity. Lastly, in addition to high immunosuppressive drug levels and tacrolimus based combinations, other post-transplant factors can be mentioned as acute rejection and antirejection treatment, cumulative steroid exposure and lymphocyte depleting antibodies (Gupta & Gupta, 2011).

Although immunosuppression increases the probability of latent BK virus reactivation, clinical manifestation of disease is rare. When symptoms occur, on the clinical point of view, a progressive decline of the renal functions can be observed up to 45 % of patients, usually 9 to 12 months after the renal transplant (Nickeleit et al. 2000a; Randhawa et al. 2000). The most serious form of the infection turns out to be the interstitial nephritis; although the BK virus was discovered in the 70's, this serious complication has first been seen in 1995. This fact can probably be explained by the commercialization of two drugs in 1995 and 1996, tacrolimus and mycophenolate mofetil.

Interestingly, BK virus-associated nephropathy happens hardly only in patients with renal graft. Some explanations could be found, such as the role of vesico-urethral reflux, quite usual in renal transplantation, with the systemic pathway of collecting tubes in peritubular capillary and the tubular localization of the infection. Some authors evoked easiness in the viral antigens presentation in a context of allograft, cold ischemia, tubular necrosis and graft rejection. For that matter BK virus-associated nephropathy is generally related to rejection, both events being linked in time; most cases of nephropathy are falsely tagged and treated just like a rejection. This confusion suggests that rejection is a risk factor on its own. Viral antigens could probably lead to rejection and conversely a rejection event could reactivate viral replication. In mice, Atencio et al proved an inductive effect of tubular damage upon BK virus linked interstitial nephritis (Atencio et al. 1993).

1.4 Clinical aspects

BK virus infection may lead to encephalitis, retinitis, pneumonitis, damage of the kidneys, bleeding of the bladder, and blockage of urine passageways. Minor infections are most of the time asymptomatic and can lead to urethral stenosis. This infection occurs 1 to 45 months (average 12.5 months) after the graft. It is linked to the conjunction of multiple factors, including an intense immunosuppressive regimen, viral reactivation, existence of an immune-allogenic conditions, and a suffering tubular due to ischemia or rejection (Nickeleit et al. 2003).

1.5 Genotypes

BK virus comes in the form of 4 different genotypes, type I being the most common seen. The coding regions for non structural proteins T and t antigens (pathogenic viral power), viral capsid proteins (cellular tropism) and a regulatory non coding zone have a vital

importance. Some authors have brought to light emerging mutations which could explain the renal physiopathologic effects of these viruses (Chen et al. 2001). Virus selection in patients with renal graft results in rearrangements in the T antigen region, mutations in the non coding regulatory zone, and above all variations in VP1 protein (Smith et al. 1998; Baksh et al. 2001; Randhawa et al. 2002). Heterogeneity and genetic instability in a same patient seem to favor renal damage and the risk of escaping immunologic surveillance (Chen et al. 2001; Randhawa et al. 2002).

1.6 Histology

BK virus is usually associated with changes in the kidney and sometimes haemorrhagic cystitis and urethral stenosis. The virus affects tubular epithelial cells that show characteristic intranuclear inclusion bodies. Diagnosis relies upon urinary cytology, detection of viral DNA in fluids and renal biopsy. The nephropathy diagnosis can only be made histologically in a graft biopsy. Intranuclear viral inclusions are exclusively seen in epithelial cells and tubular cells reveal focal necrosis. Four different variants of intranuclear inclusion bodies can be seen throughout the entire nephron. Type 1 is the most frequently observed; it is an amorphous basophilic ground-glass variant. Type 2 is an eosinophilic granular type, halo surrounded. Type 3 is a finely granular form lacking a halo. And finally type 4 is a vesicular variant presenting markedly enlarged nuclei and irregular chromatin. Infected cells which are rounded-up and extruded from the epithelial cell layer into tubular lumens are frequently observed. Viral replication often causes tubular epithelial cell necrosis with denudation of basement membranes. Although cytopathic signs can be seen along the entire nephron, they are mostly abundant in distal tubular parts and collecting ducts (Nickeleit et al. 2000a).

1.7 Interstitial inflammation

Interstitial inflammation in BK virus-associated nephropathy still remains controversial and needs to be fully explained. The major outcome is to distinguish between virally induced interstitial nephritis and cellular rejection. As lowering immunosuppression is the first option which can be chosen in the treatment, this choice requires two conditions, first the absence of rejection and second the BK virus should not trigger rejection. BK virus is frequently accompanied by an heterogeneous inflammatory reaction (Drachenberg et al. 1999). This inflammation can be minimal or absent in up to 17% of biopsies (Nickeleit et al. 2000a). When inflammation is encountered, the inflammatory cell infiltrate is composed of lymphocytes, macrophages and occasional plasma cells. Polymorphonuclear leukocytes can be seen in response to markedly damaged tubules with urinary leakage (Drachenberg et al. 1999). About 50% of biopsies performed during persistent BK virus-associated nephropathy show evidence of cellular rejection as conventionally defined with abundant tubulitis and transplant endarteritis in about 25%. Typically, mononuclear cell infiltrates and tubulitis are pronounced in areas without viral inclusions making virally induced interstitial nephritis highly unlikely (Nickeleit et al. 2000a).

The upregulation of MCH-class II (HLA-DR) and ICAM-1 on tubular epithelial cells is a typical finding in graft biopsies with cellular rejection and can serve as an adjunct diagnostic tool (Seron et al. 1989; Nickeleit et al. 1998). HLA-DR expression can stimulate an allogenic

lymphocytic reaction and also enhance T cell mediated lysis (Rosenberg et al. 1992). Consequently, BK virus could probably trigger rejection episodes by inducing HLA-DR upregulation as previously proposed for CMV (von Willebrand et al. 1986). However, no association could be found between BK virus infection and tubular HLA-DR expression based on immunofluorescence double labeling staining techniques. It is only in biopsies showing characteristic morphological evidence of rejection with marked tubulitis that typical upregulation of HLA-DR and ICAM-1 could be observed (Nickeleit et al. 2000a). Therefore, BK virus does not stimulate HLA-DR expression. Consequently no significant difference can be found between the prevalence of rejection in tissue samples taken during persistent BK virus-associated nephropathy and time matched controls without BK virus nephropathy. Thus, BK virus does not seem to provoke a constant and pronounced interstitial inflammatory reaction and should probably not be considered as associated with an increased prevalence of rejection episodes (Nickeleit et al. 2000a).

1.8 PCR

BK-virus DNA in the plasma and the urine, which can be detected by PCR (Polymerase Chain Reaction), is closely associated with nephropathy. Quantitative PCR can be used to follow the disease evolution and the treatment efficiency (Randhawa et al. 2004).

As for BK virus infection, this technique has proven a 100% sensitivity, a 88% specificity and above all a negative predictive test of 100%. Hirsch et al. have even shown a correlation between viral load and nephropathy and proposed a cut-off above which the risk of nephropathy is significant: all patients with more than 7700 copies/mL in plasma had typical BK virus-associated nephropathy lesions on the biopsy (Hirsch et al. 2002).

The nephropathy evolution is very poor with a cytopathogenic effect persistent in up to 70% of patients, a graft loss in 45% of cases; and major sequel fibrosis in 75% of cases, even if viremia can be controlled (Nickeleit et al. 2000a; Randhawa et al. 2000; Mylonakis et al. 2001; Mengel et al. 2003).

2. Classical treatments for BKV nephropathy

Therapeutic alternatives are quite few in number. Despite the absence of randomized clinical trials, the current approach generally includes reduction of immunosuppression (Brennan et al. 2005; Hardinger et al. 2010). The rationale is to allow host immune function to combat the virus, with the risk to increase acute and subclinical rejection. Lowering immunosuppression with smaller dosage and/or less drugs is partially efficient and seems to be the first thing to do. Except from lowering immunosuppression, to date no treatment seem to be efficient enough to be recommended to all patients, and new research have to be performed because of the poor evidence in small series of patients (Johnston et al. 2010).

2.1 Lowering immunosuppression

Reduction of immunosuppression is to date the only consensus regarding the treatment of BK virus-associated nephropathy. Lowering tacrolimus dosage of 41% and mycophenolate mofetil dosage of 44% allowed to eradicate 24 patients' viremia in 6 months (Saad et al. 2008).

In a previous study, mycophenolate mofetil was stopped the day leflunomide treatment was initiated; tacrolimus and everolimus were respectively reduced of 50% and 12.5%. Therapeutic drug monitoring target for tacrolimus was lowered to 4 - 6 ng/mL on immunoenzymatic techniques on whole blood. Corticosteroids were kept with average dosage of 5 to 10 mg per day (Bazin et al. 2009). Other authors recommend even lower targets with 3 ng/mL for tacrolimus and 100 ng/mL for cyclosporine (Gupta & Gupta, 2011).

Besides, lowering immunosuppressive regimens together with a specific treatment for BK virus-associated nephropathy recently turns out to be effective to prolong graft survival, and moreover a safe treatment with acute rejection rates not increased significantly after lowering immunosuppression (Dheir et al. 2011).

Two different therapeutic strategies have been evaluated: the immunosuppression withdrawal (3-drug to 2-drug immunosuppression) within the first month versus reduction of immunosuppression. The regimen modifications and results are presented in table 1 and figure 1. The Withdrawal cohort had significantly better graft survival at 1 year compared with the Reduction cohort (1-year graft survival 87.8% versus 56.2%, $P = 0.03$) (Weiss et al. 2008).

	Withdrawal cohort (n = 17)	Reduction cohort (n = 18)	p
CNI, sirolimus, prednisone at diagnosis	12	11	0.56
CNI, MMF, prednisone at diagnosis	5	7	0.56
Median serum creatinine at diagnosis (mg/dl)	2.5	2.2	0.30
Agent withdrawal within 1 mo of diagnosis			
CNI withdrawal	14	-	-
AP withdrawal	3	-	-
Dose reduction within 1 mo of diagnosis			
CNI reduction, AP reduction < 50%	-	8	-
CNI reduction, AP reduction ≥ 50%	-	7	-
Tac to CsA switch, AP reduction < 50%	-	3	-
Ancillary therapy			
Cidofovir	2	5	0.40
Intravenous Ig	8	8	0.88
Leflunomide	4	5	1.0
Acute rejection after diagnosis	1	1	1.0

Table 1. Immunosuppression modifications comparing immunosuppression withdrawal *versus* immunosuppression reduction after diagnosis of BK virus-associated nephropathy. CNI, calcineurin inhibitor; MMF, mycophenolate; AP, antiproliferative; Tac, tacrolimus; CsA, cyclosporine A (Weiss et al. 2008).

2.2 Cidofovir

Cidofovir (Vistide®) is an injectable antiviral drug. It belongs to nucleoside analogues. It is used in infections due to human Cytomegalovirus (CMV) in adults suffering of AIDS (Acquired immune deficiency syndrome) without renal insufficiency, and it should only be used when other treatments are considered as inappropriate. Cidofovir counters CMV replication thanks to a selective inhibition of viral DNA polymerase in *herpesviridae* viruses

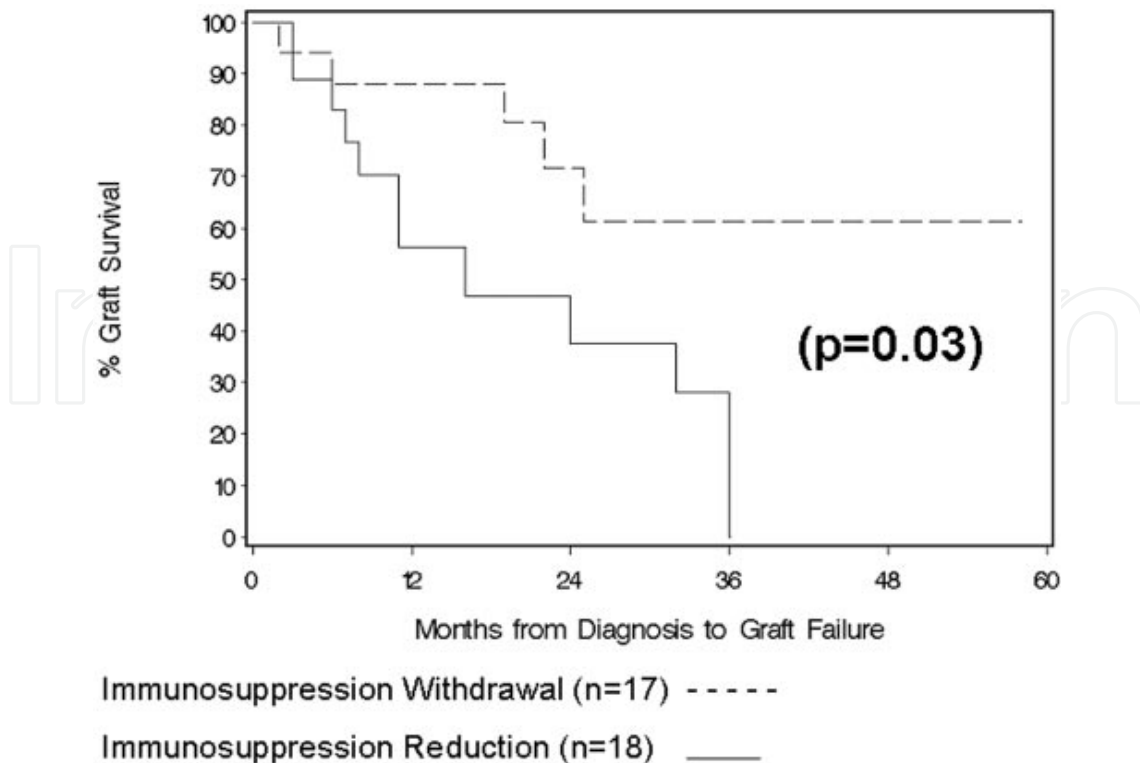


Fig. 1. Immunosuppression withdrawal preserves graft function compared with reduction (Weiss et al. 2008).

(Gilead 2010). Cidofovir has also demonstrated *in vitro* activity against murine and simian polyomavirus strains and appears to have activity against JC virus *in vivo* (De Luca et al. 2000). Pharmacokinetic studies have demonstrated that cidofovir is highly concentrated in urine and renal tissue which are the primary sites of BK virus infection (Kadambi et al. 2003). This fact highlights the possibility that low doses might be sufficient for treating an infectious process, such as BK virus-associated nephropathy, that appears to be largely localized to the kidney and genitourinary tract.

The treatment consists in a low-dose treatment, 0.25 mg/kg/day intravenous during 2 weeks, associated to a prior hydration of 1 litre of saline solution. Cidofovir seems to be efficient in BK virus as well, but it tends to concentrate itself inside the kidney and can be responsible of a nephrotoxicity mostly for tubular cells leading to renal insufficiency.

Few cases have been described in literature and no conclusion can be given on the real efficacy of cidofovir. Indeed, despite viremia control, viruria remains detectable and the treatment is not able to avoid the evolution towards fibrosis and renal insufficiency (Kadambi et al. 2003; Kuypers et al. 2005).

In some cases, cidofovir may also become deleterious (Pallet et al. 2010; Talmon et al. 2010).

3. Leflunomide

3.1 Drug generalities

Leflunomide (Arava®) is a disease-modifying antirheumatic drug (DMARD) used in adult patients with methotrexate intolerance, failure or loss of efficiency; it is also used in a second

line to treat severe and active forms of psoriatic arthritis (Maddison et al. 2005; Sanofi-Aventis 2009).

3.2 Pharmacodynamics

Its immunosuppressive action lies in the dihydroorotate dehydrogenase (DHOH) inhibition, an enzyme necessary for de novo synthesis of pyrimidic bases in lymphocytes. It also has an anti-proliferative action (Williamson et al. 1995; Fox et al. 1999).

Besides, leflunomide has proven abilities to reduce the viral proliferation for Human Cytomegalovirus (CMV), Herpes Simplex Viruses (HSV) *in vitro* (Knight et al. 2001) and respiratory syncytial virus (RSV) *in vitro* and *in vivo* (Dunn et al. 2011).

3.3 Pharmacokinetics

After per os administration, leflunomide is promptly and almost fully metabolized into its active form, terflunomide or A77 1726. This metabolism happens during first pass and consists in a carbon cycle opening in the intestinal wall and the liver. 95% of leflunomide is turned into A77 1726 this way, the remains into minor metabolites. Terflunomide is the drug responsible for the activity and side effects of leflunomide.

Leflunomide bioavailability is about 82% in healthy volunteers (Sanofi-Aventis 2009). Elimination plasma half-life of A77 1726 is quite considerable, with some 15 days in average. Patients are so compelled to take a 100 mg charging dose for 3 days before a 10 to 20 mg maintenance dose per day.

After a unique charging dose, A77 1726 T_{max} is comprised between 6 and 12 hours, with a high inter-individual variability in patients with rheumatoid arthritis (Rozman 2002).

The volume of distribution (V_d) is quite low, with about 12.7 L (6 to 30.8 L), which is logical with its high affinity and linkage to albumin (99.4% in healthy volunteers) (Rozman 2002). Elimination of A77 1726 is slow, it is characterized by an apparent clearance of 0.051 L/h (Rozman 2002). This elimination is mostly renal (43%) and biliary (48%), as a consequence renal insufficiency alone does not significantly impair A77 1726 plasma concentrations (Beaman et al. 2002). Furthermore haemodialysis does not modify concentrations or clearance of A77 1726, which allows the patients to be on a dialysis without any dose adjustment. *In vitro* studies showed that cytochroms P450, in particular cytochroms 1A2, 2C19, 3A4 and 3A5 were involved in leflunomide metabolism (Kalgutkar et al. 2003). A pharmacogenetic study also showed the link between a polymorphism of cytochrom 1A2 and a risk of toxicity for patients with rheumatoid arthritis (Bohanec Grabar et al. 2008).

3.4 Predictive efficiency

In rheumatoid arthritis, plasma concentrations above 13 µg/mL seem to be efficient. These concentrations are usually reached with 20 mg per day dosage (van Roon et al. 2005). Some authors tried to establish a relation between plasma concentrations and efficiency in patients with BK virus nephropathy, showing a tendency but with no absolute proof. Finally to date, no link between plasma concentrations and side effects has been shown (Bazin et al. 2009). Yet *in vitro* studies seem to show a predictive correlation between concentrations and the

viral inhibitory effect: 10 $\mu\text{g}/\text{mL}$ reduced the extracellular BKV load by 90% (IC_{90}) but with significant host cytostatic effects (see figure 2) (Bernhoff et al. 2010).

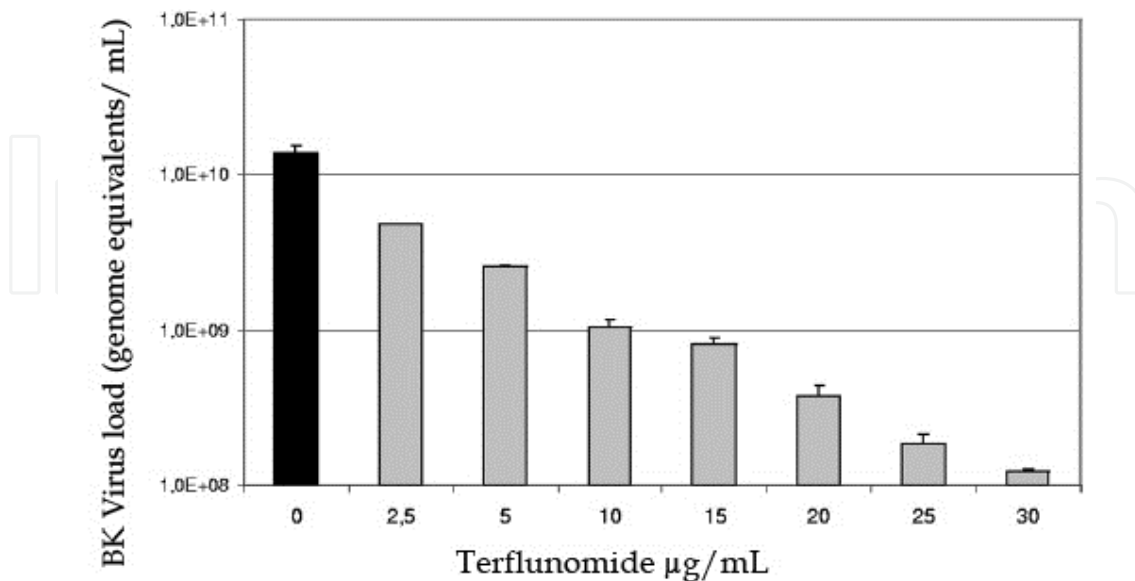


Fig. 2. Effect of Terflunomide on BK Virus load *in vitro* (Bernhoff et al. 2010)

3.5 Mechanism of action

Researches about the mechanism of leflunomide have recently been brightened. Leflunomide has two mechanisms of action: inhibition of dihydroorotate dehydrogenase, a key enzyme in the pyrimidine synthesis pathway, and tyrosine kinase inhibition. Dihydroorotate dehydrogenase inhibition is the primary mechanism involved in rheumatoid arthritis treatment. Interactions between the BK virus and the cellular protein kinase Akt / mammalian target of rapamycin (mTOR) pathway have been discovered (Liacini et al. 2010). These interactions are described in figure 3.

Akt (protein kinase B) is a serine/threonine kinase activated by growth factors, cytokines and mitogens (Fayard et al. 2010). The mTOR pathway which controls protein synthesis is located downstream of Akt. Akt indirectly activates mTOR. Two mTOR complexes have yet been described, mTOR complex 1 (mTORC1) which controls translation initiation, and mTOR complex 2 (mTORC2) which controls cytoskeletal changes and is also a 3'-phosphoinositide-dependent kinase-2 (PDK2), phosphorylating Akt, which may alter its substrate specificity (Bhaskar et al. 2007). Liacini et al showed that BK virus infecting renal tubular epithelial cells was able to activate the Akt/mTOR pathway; that leflunomide active metabolite, A77 1726 could inhibit PDK1 and Akt phosphorylation in a dose-dependent manner and in this way to reduce BK large T antigen expression and DNA replication. The combination of serine/threonine kinase inhibition of mTOR and tyrosine kinase inhibition significantly reduce the ability of the virus to survive and to produce new virions. More interesting though seems to be the combination of leflunomide and sirolimus targeting the Akt/mTOR pathway on different sites. Because both leflunomide and sirolimus possess immunosuppressive activity, this combination may allow treatment of BK virus-associated nephropathy without reduction of immunosuppression (Liacini et al. 2010).

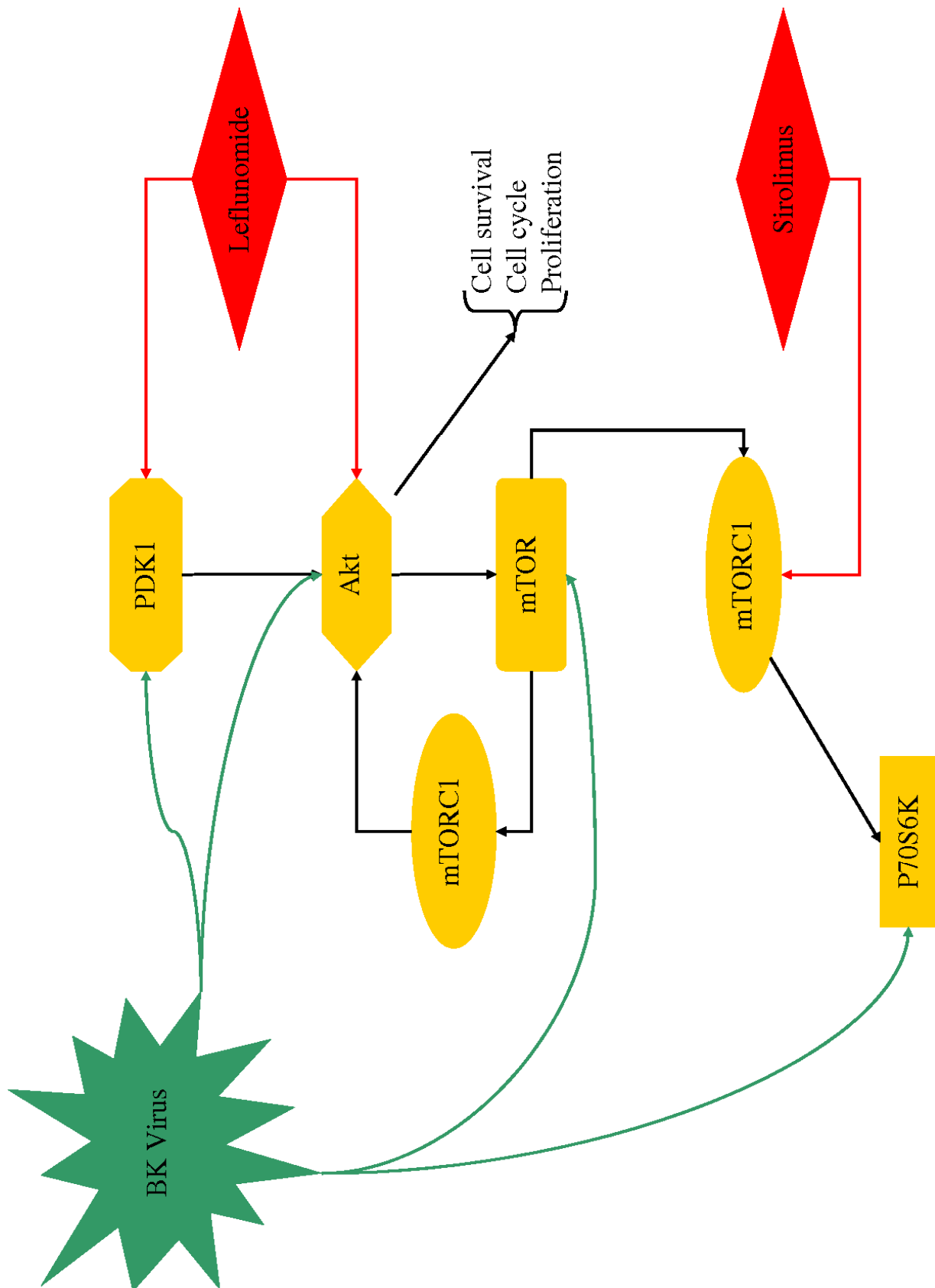


Fig. 3. Interactions between BK virus and inhibitors, sirolimus and leflunomide (Liacini et al. 2010)

Results in terms of biological evolution in patients speak for itself. In a mean monitoring time of 16 months (12-24 months), viral load with leflunomide can be reduced about up to 50%, and even be brought to undetectable, in blood like in urine. But more important is the lowering of renal failure and graft rejection thanks to this treatment. Creatinine clearance (Cockcroft-Gault) can be stabilized and even improved (Bazin et al. 2009).

3.6 Therapeutic drug monitoring

Initial dosage for leflunomide is 20 mg once a day, and can be raised to 30 or 40 mg for patients with viral loads remaining important. Plasma concentrations in therapeutic drug monitoring fluctuate between 15 and 135 $\mu\text{g}/\text{mL}$. These concentrations set out low intra-individual but high inter-individual variability, and moreover without apparent correlation with prescribed dosage. It is interesting to notice that these concentrations were outside usual targets used in most studies - 50 to 100 $\mu\text{g}/\text{mL}$ - which are supposed to offer the best efficiency and to limit the hepatotoxicity risk which can be lethal. Besides, the patient with the highest concentration - 135 $\mu\text{g}/\text{mL}$ - had its viremia turned undetectable after only a two months treatment and showed no side effect of any kind. This result suggests that higher concentrations lead to higher efficacy and vice versa (Bazin et al. 2009).

3.7 Tolerance

Concerning tolerance, very few patients suffer from serious side effects. Loss of taste or lethargy can be observed but without any correlation with plasma concentrations. These side effects can prompt the treatment to be stopped, but in most cases viremia tends to increase strongly (Bazin et al. 2009).

4. Discussion

The main risk factor for BK virus-associated nephropathy is undeniably the immunosuppressive regimen intensity, in particular an intensification due to an acute rejection event (Binet et al. 1999; Nicleleit et al. 2000a; Barri et al. 2001; Nicleleit et al. 2003). Drugs in cause for these events seem to be the combination of tacrolimus, mycophenolate mofetil and monoclonal or polyclonal antibodies (Binet et al. 1999; Nicleleit et al. 1999; Nicleleit et al. 2000a; Nicleleit et al. 2003; Benavides et al. 2007). The organ and the graft type also play a role. For instance, Benavides et al. showed that the incidence of BK virus-associated nephropathy is higher in patients with kidney and pancreas rather than kidney alone; and that an alive donor would had a protective effect, probably explained by a lighter immunosuppressive regimen (Benavides et al. 2007).

Other risk factors have been evoked, like age and sex: nephropathy incidence seem to be greater for aged men (Ramos et al. 2002).

Furthermore many patients improve their symptoms at a distance of the surgery with the lowering of immunosuppression. We already have at our disposal a few experimental studies testing leflunomide on chronic or acute graft rejection (Williams et al. 1994; Xiao et al. 1995; Shen et al. 1998). More recently the inhibitory effects of leflunomide upon HSV, CMV and BK virus have been proved *in vitro* like *in vivo* (Waldman et al. 1999; Waldman et al. 1999; Knight et al. 2001; Farasati et al. 2005). Indeed a study suggests leflunomide is at

least as efficient as ganciclovir in CMV infections and does not seem to be affected by resistant viruses (John et al. 2004). Leflunomide has even been successfully used in a patient with bone marrow graft and infected by a resistant virus to ganciclovir, foscarnet and cidofovir (Avery et al. 2004).

The studies where leflunomide is used as an immunosuppressive drug in renal and hepatic graft are more and more, because leflunomide allows to reduce anti-calcineurin drugs which have the major inconvenient of nephrotoxicity, and potentially protects aside from CMV, HSV and BK virus infections (Hardinger et al. 2002; Williams et al. 2002). Moreover leflunomide seems to be an interesting alternative in BK virus-associated nephropathy in renal transplant by eradicating detectable viremia in some patients. Leflunomide also allows avoiding rejection in most cases in spite of classical immunosuppressive drugs dosage reduction. Besides one of leflunomide's main asset is its absence of renal toxicity, contrary to cidofovir (Williams et al. 2005; Josephson et al. 2006; Teschner et al. 2006; Faguer et al. 2007).

Thanks to the encountered success in renal transplant, leflunomide is now used to treat hemorrhagic cystitis linked to BK virus in bone marrow transplant (Dropulic et al. 2008). However, due to the absence of randomized clinical trials with a sufficient number of patients, some authors consider its use in a first-line drug not recommended (Chon et al. 2011).

Leflunomide pharmacokinetics is characterized by a great inter-individual variability with terflunomide concentrations from 15 to 130 $\mu\text{g}/\text{mL}$ obtained with the same dosage (Bazin et al. 2009). In BK virus infection, terflunomide concentrations between 15-30 $\mu\text{g}/\text{mL}$ and 35-100 $\mu\text{g}/\text{mL}$ are sufficient to suppress respectively 50% and 90% of the replication for CMV and BK virus *in vitro*. That is why a therapeutic margin between 50 and 100 $\mu\text{g}/\text{mL}$ has been proposed in this indication (Josephson et al. 2006). However, current strategic therapy so as to limit BK virus incidence tends to manage an early reduction of immunosuppressive regimen to avoid the apparition of a nephropathy. A prospective study with a significant number of patients would be probably necessary to definitely conclude about this relation between plasma concentrations and efficacy or in terms of rapidity of viral load eradication.

5. Conclusion

Leflunomide appears to be an alternative treatment in nephropathy due to BK virus in addition to lower immunosuppression regimen. In case of leflunomide use, a major standard seems to be high plasma terflunomide concentrations so as to obtain rapid virus eradication. Concentrations comprised between 15 and 60 $\mu\text{g}/\text{mL}$ appear to be pertinent; these concentrations are usually reached with 20-40 mg per day. In patients with insufficient concentrations, further studies should be carried out to determine whether exists a benefit to use higher dosage up to 60 or 80 mg a day. Even if tolerance is quite satisfying, it will probably be the most important parameter in such high-dose treatments.

Despite the small number of studies and the weak number of patients in each of them, a correlation seems to exist between plasma terflunomide concentrations and the treatment efficacy. This relation has not yet been proved with tolerance.

Due to its great inter-individual variability and alongside classical virological and clinical follow-up therapeutic drug monitoring appears to be an important step to take into care patients with BK virus related nephropathy.

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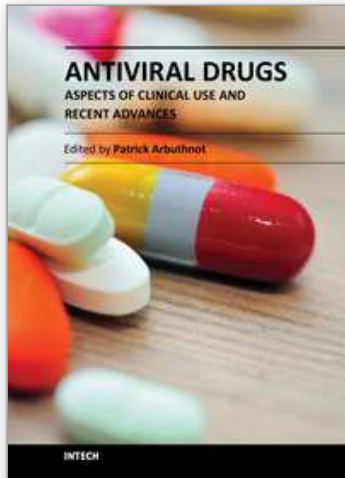
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The articles that appear in Antiviral Drugs - Aspects of Clinical Use and Recent Advances cover several topics that reflect the varied mechanisms of viral disease pathogenesis and treatment. Clinical management and new developments in the treatment of virus-related diseases are the two main sections of the book. The first part reviews the treatment of hepatitis C virus infection, the management of virus-related acute retinal necrosis, the use of leflunomide therapy in renal transplant patients, and mathematical modeling of HIV-1 treatment responses. Basic research topics are dealt with in the second half of the book. New developments in the treatment of the influenza virus, the use of animal models for HIV-1 drug development, the use of single chain camelid antibodies against negative strand RNA viruses, countering norovirus infection, and the use of plant extracts to treat herpes simplex virus infection are described. The content of the book is not intended to be comprehensive, but aims to provide the reader with insights into selected aspects of established and new viral therapies.

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