## we are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists



122,000

135M



Our authors are among the

TOP 1%





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

## Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



### Mantle Cell Lymphoma: Decision Making for Transplant

Yener Koc and Taner Demirer Medical Park Hospital, Antalya and Ankara University Medical School, Ankara Turkey

#### 1. Introduction

#### 1.1 Definition and clinical characteristics

Mantle Cell Lymphoma (MCL) is a relatively rare type of mature B-cell lymphoma that comprises 5% of Non-Hodgkin's Lymphomas(NHL)<sup>1-3</sup>. MCL was added to the Revised European–American Lymphoma classification in 1994. Having both indolent and incurable features associated with aggressive clinical course, MCL is most frequently seen in 6<sup>th</sup> decade of life, with male dominance 3 to 4:1<sup>2</sup>. Malignant origin of MCL cells appear to derive from an antigen-naive pregerminal center cell<sup>4, 5</sup>.

#### 1. B-cell neoplasms

- 1. Precursor B-cell
- 2. Mature B-cell
  - Chronic lymphocytic leukemia/small lymphocytic lymphoma
  - Lymphoplasmacytic lymphoma
  - Splenic marginal zone lymphoma
  - Extranodal marginal zone B-cell lymphoma of MALT
  - Nodal marginal zone B-cell lymphoma
  - Follicular lymphoma
  - Mantle cell lymphoma
    - Diffuse large B-cell lymphoma
    - Mediastinal (thymic) large B-cell lymphoma
    - Intravascular large B-cell lymphoma
    - Primary effusion lymphoma
    - Burkitt's lymphoma/leukemia
- 3. B-cell proliferations of uncertain malignant potential

#### 2. T-Cell and NK-Cell neoplasms

Table 1. World Health Organization Classification of Lymphomas<sup>1</sup>.

At the time of diagnosis, patients tend to have more extranodal disease and low serum albumin<sup>6</sup>. Although MCL has differential diagnosis with Chronic Lymphocytic Leukemia (CLL) and low-grade NHL, mimicking malignancies with indolent behavior, it follows an aggressive course with 10% to 15% long-term survivors<sup>7</sup> despite administration of standard chemotherapy courses commonly used in NHLs<sup>8</sup>, corresponding to a median survival of 3 to 5 years.

#### 2. Diagnosis

Diagnosis of MCL can be made by lymph node or bone marrow biopsy, or analysis of malignant cells obtained from peripheral blood, if the disease is in the leukemic phase<sup>9, 10</sup>. Differential diagnosis with CLL is important since both MCL and CLL cell have co-expression of CD5 and CD19/20<sup>11</sup>. Malignant cells are negative for CD10, CD23 and BCL6. Although absence of CD23 antigen expression on malignant cell population strongly favor a diagnosis of MCL<sup>12, 13</sup>, presence of cyclinD1 expression by immunohistochemical staining<sup>14</sup> or determination of t(11;14) translocation by molecular analysis<sup>15-18</sup> is required to confirm the diagnosis<sup>19</sup>. Cyclin D1 is overexpressed in MCL as a result of the landmark t(11;14)(q13;q32) translocation<sup>20</sup>. Cyclin D1 complex with cyclin dependent kinases 4 and 6 (Cdk4 and Cdk6) and cyclin E-Cdk2, leading to phosphorylation of retinoblastoma protein (Rb), irreversibly inducing progression of the cell from G1 to S phase, which is not the only biologic dysregulation on the way to malignant transformation<sup>21-24</sup>.

Disease	CD5	CD10	CD23	CD43	Cyclin D1	Ig class
FL	-	+	+/-	-	-	IgM, IgG
MCL	+	-	-	+	+	IgM/IgD
CLL/SLL	+	-	+	+	-	IgM/IgD
LPL	-	-	-	+/-	-	IgM (c)
MALT	-	-	-	+/-	-	IgM (c, s)
SMZL	-	-	Π - /		-	IgM/IgD
HCL	1974			}	-/+	IgG

FL, follicular lymphoma; MCL, mantle cell lymphoma; CLL/SLL, chronic lymphocytic leukemia/small lymphocytic lymphoma; LPL, Lymphoplasmacytic lymphoma; MALT, marginal zone lymphoma of MALT type; SMZL, splenic marginal zone lymphoma; HCL, hairy cell leukemia; Ig class, most commonly expressed heavy chain classes; c, cytoplasmic Ig; s, surface Ig.

Table 2. Differential Diagnosis of "Small" B-Cell Lymphomas<sup>25</sup>.

#### 3. Prognostic parameters

Prognosis can be estimated by using MIPI (mantle cell lymphoma international prognostic index, Figure 1)<sup>26</sup> which seems to be more efficient than international prognostic index (IPI)<sup>27</sup> or follicular lymphoma international prognostic index (FLIPI)<sup>28, 29</sup>, which includes leukemic phase<sup>10, 30, 31</sup>, besides other clinical parameters used in the IPI.

320

Points	Age, y	ECOG	LDHULN	WBC, 10 <sup>9</sup> /L
0	<50	0-1	<0.67	< 6.700
1	50-59		0.67-0.99	6.700-9.999
2	60-69	2-4	1.000 -1.49	1.000-14.999
3	≥70	1 <u>11111</u>	≥1.5000	≥15000

#### Table 3. Simplified MIPI<sup>26</sup>.

For each prognostic factor, 0 to 3 points were given to each patient and points were summed up to a maximum of 11. Patients with 0 to 3 points in summary were classified as low risk, patients with 4 to 5 points as intermediate risk, and patients with 6 to 11 points as high risk.

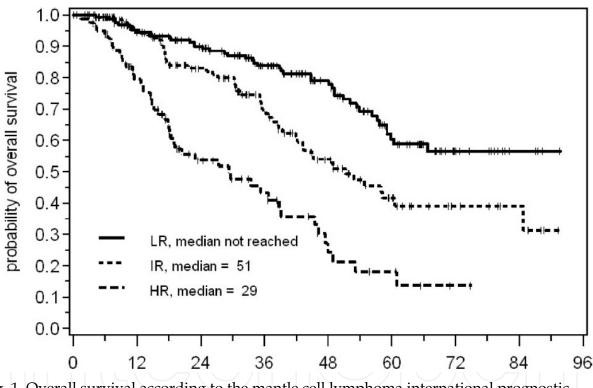


Fig. 1. Overall survival according to the mantle cell lymphoma international prognostic index (MIPI).

Although Ki-67 has previously been shown to predict prognosis (Figure 2)<sup>32-34</sup>, analysis of Ki-67 did not substantially change the regression coefficient of the MIPI score and served as an important biologic marker with strong additional prognostic relevance<sup>26</sup>.

Recently, proliferation gene expression signature has been reported to be the best molecular predictor of survival in patients with MCL<sup>21</sup>, leading to a prognostic model defined as an optimized survival predictor composed of five genes: RAN, MYC, TNFRSF10B, POLE2, and SLC29A2<sup>35</sup>. Furthermore, this model was validated for application in formalin-fixed paraffin-embedded tissue samples and appeared superior to the immunohistochemical marker Ki-67<sup>35</sup>.

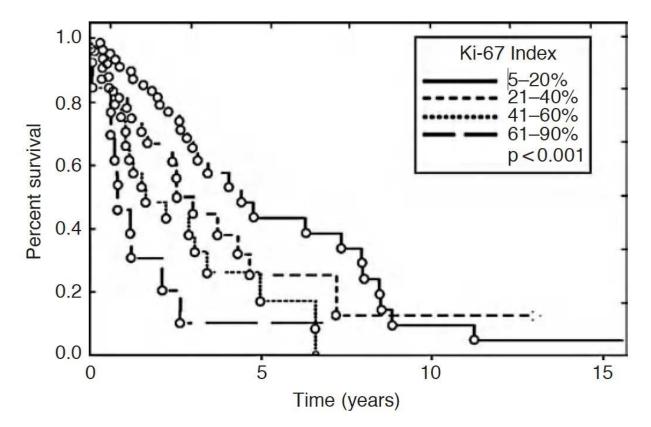


Fig. 2. Overall survival of patients with MCL according to Ki-67 proliferation index<sup>33</sup>.

#### 4. Role of chemotherapy

When it comes to assessment of long term overall survival, MCL has the worst prognosis among all lymphoma types<sup>36</sup>, a watch-and-wait strategy should be avoided. Retrospective analyses of administration of standard chemotherapy have not shown any improvement in overall survival in patients with MCL<sup>6, 37</sup>. Most regiments induce around 80% response rates with complete remissions up to 30% of cases<sup>6, 8</sup>. While more aggressive regimens were used<sup>38</sup>, reliable cure rates with conventional treatment has not been reported until now<sup>36</sup>. Compared to historical controls, median survival with conventional chemotherapy CVP and CHOP have not been improved<sup>37</sup>. In a randomized trial, these two regimens were associated with similar response rates (84% and 89%) and median overall survivals (32 and 37 months)<sup>39</sup>. Only one third of patients with untreated<sup>40</sup> or relapsed<sup>41</sup> MCL will respond to rituximab as single agent. Rituximab may also be used for purging tumor cells either with standard chemotherapy<sup>42</sup> or prior to high dose therapy regimens with stem cell support<sup>43</sup>. But in a randomized trial, addition of rituximab to CHOP regimen failed to improve overall survival (Figure 3)<sup>44</sup>. Maintenance interferon-alfa therapy following induction regimen has not been proven to improve survival either<sup>45, 46</sup>.

Fludarabine can also be used in patients with lymphoma with or without rituximab combination<sup>47</sup>. Fludarabine alone in previously treated MCL can induce temporary responses in a third of patients lasting between 4 to 8 months<sup>48</sup>. In newly diagnosed patients, fludarabine appears to be more active, inducing responses in 60%, half of which is complete response<sup>49</sup>. Combining fludarabine with idarubicin or cyclophosphamide may improve induction of complete responses<sup>50, 51</sup>. Fludarabine and cyclophosphamide

combination(FC) can induce a higher response rate of 63% in patients recurrent MCL<sup>51</sup>. Previously untreated patients have response rate up to 100% with a complete response rate of 70% and 28 months of progression free survival<sup>51</sup>.

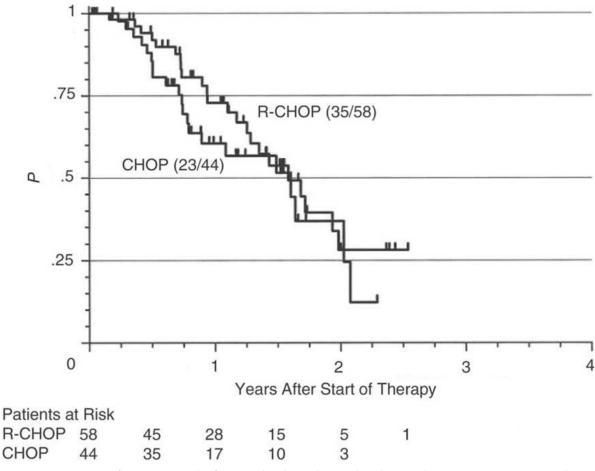


Fig. 3. Progression-free survival after cyclophosphamide, doxorubicin, vincristine, and prednisone (CHOP) and rituximab and CHOP (R-CHOP)<sup>44</sup>.

In a randomized study testing addition of rituximab to fludarabine, cyclophosphamide and mitoxantrone (FCM), patients in the rituximab arm were found to have significant improvement in disease free and overall survival<sup>52-54</sup>. Fludarabine combined with cyclophosphamide and rituximab is a highly effective regimen in patients relapsing from previously received CHOP regimen<sup>55</sup>. Addition of rituximab to the chemotherapy regimens does not appear to increase toxicity<sup>56</sup>.

Although tolerating patients treated with the hyper-CVAD regimen had excellent response and survival rates of greater than 90% at 3 years and 4.6 years of time to treatment failure (TTF at 8yr , 16% if age>65, 46% if age<65) following 10 year observation period<sup>7, 38, 57, 58</sup>, patients treated with conventional therapies have also reported to have similar 3 year survival rates<sup>59, 60</sup>. The hyper-CVAD regimen without stem cell transplantation was not associated with a plateau in the survival curves (Figure 4)<sup>57</sup>. In this study, beta-2microglobulin levels and IPI/MIPI scores were found to predict survival<sup>57</sup>. Hyper-CVAD regimen data may suggest high efficacy of the high-dose Ara-C (HIDAC) regimen, which needs to be tested in further clinical trials.

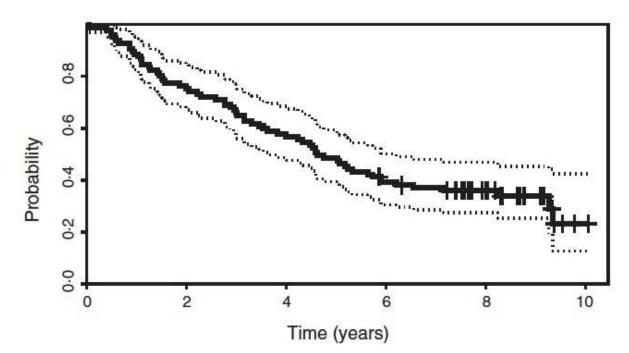


Fig. 4. Overall survival and time to failure in 97 patients treated with R hyper-CVAD alternating with  $R M/A^{57}$ .

Due to advanced age at the time of presentation of MCL, there is a concern about intensive therapy regimens being associated with higher toxicity rates requiring patient selection and patients tolerating intensive therapies are being selected for high-dose therapies with stem cell support.

#### 5. New agents

Recently, bortezomib<sup>61-63</sup>, lenalidomide<sup>64-66</sup>, bendamustine<sup>67-69</sup>, pixantrone<sup>70</sup>, azaepothilone ixabepilone<sup>71</sup> and mTOR inhibitor temsirolimus<sup>72-74</sup> has been shown to demonstrate activity alone and in combination with rituximab and mitoxantrone, and may induce complete responses in relapsed or refractory MCL<sup>61, 64, 69, 75, 76</sup>. Along with rituximab, these biotherapy agents can be used frontline or may also be introduced into post transplant maintenance to prevent or treat relapses<sup>61, 68, 77-81</sup>. There may be an advantage of combining rituximab with bendamustine compared to combining with the conventional CHOP regimen in terms of disease free survival<sup>82</sup>. Recently, it was reported that frontline use of cladribine and rituximab can induce an overall response rate of 87%, with 61% of patients achieving complete remission, suggesting use of this combination for the initial treatment of MCL<sup>83</sup>. Radioimmunotherapy with <sup>90</sup>Y-labeled anti-CD20 monoclonal antibody (ibritumomab tiuxetan) or <sup>131</sup>I-rituximab is now being tested in phase I-II trials, including stem cell transplant setting<sup>84-87</sup>.

#### 6. High dose chemotherapy with autologous stem cell support (ASCT)

Since conventional and dose intense chemotherapy regimens have failed to induce cure or a plateau phase in time to treatment failure curves, high dose therapy with autologous stem cell support have been studied in the relapse setting<sup>88</sup> as well as consolidating complete responses following frontline chemotherapy regimens<sup>88-94</sup>.

In a randomized trial testing ASCT versus IFN-alfa consolidation following response to CHOP or CHOP like regimens with or without rituximab, consolidation with BEAM/ASCT improved progression free survival (PFS) but has not been shown to improve overall survival (OS)(Figure 5)<sup>93</sup>. When patients responding to R-CHOP regimen received HIDAC therapy followed with autologous tranplantation with the BEAM regimen, also a plateau was not observed in time to treatment failure (TTF) and OS curves<sup>94</sup>.

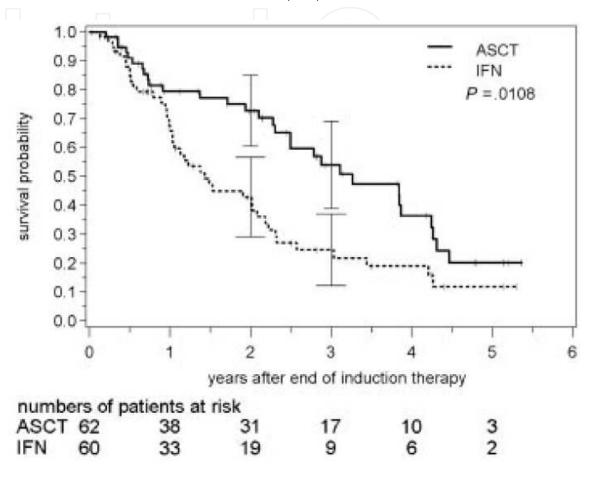
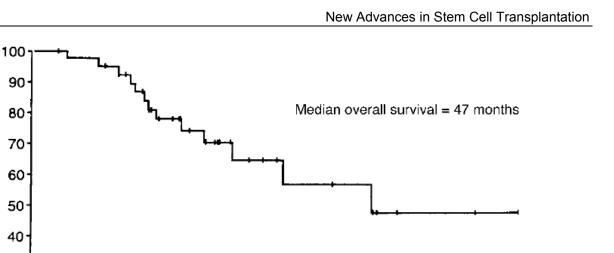


Fig. 5. Progression-free survival after high-dose radiochemotherapy followed by ASCT and IFN-alfa maintenance in MCL<sup>93</sup>.

Autologous transplantation with conventional conditioning regimens without in vivo purging induced a median survival of 47 months<sup>88</sup>. Analysis of this patient group did not reveal a plateau in the survival curve<sup>88</sup>.

Addition of rituximab to ASCT regimen leads to better event free survival (EFS) curves in patients with first remission, not affecting progression in patients with relapsed/refractory disease<sup>91</sup>.

The R-HDS regimen consisting of high-dose sequential chemotherapy (including intravenous administration of high-dose cyclophosphamide, high-dose cytarabine, high-dose melphalan, and high-dose mitoxantrone plus melphalan) and in vivo purging with rituximab resulted in OS and EFS rates at 54 months were 89% and 79%, respectively<sup>96</sup>. These results compare favorably with the 42% OS rate and the 18% EFS rate observed in 35 age-matched historic controls treated with standard-dose chemotherapy at the participating centers (Figure 6)<sup>96</sup>.



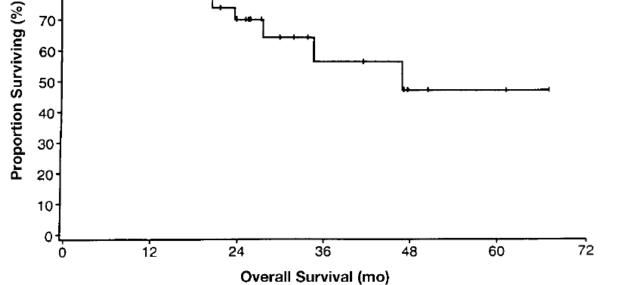


Fig. 6. Overall survival for mantle cell lymphoma patients after autologous transplantation without in vivo purging<sup>88</sup>.

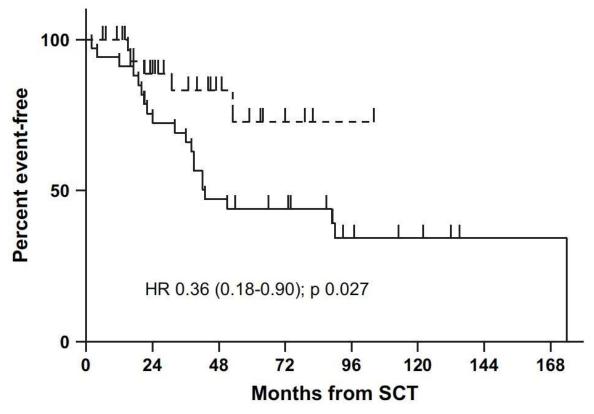
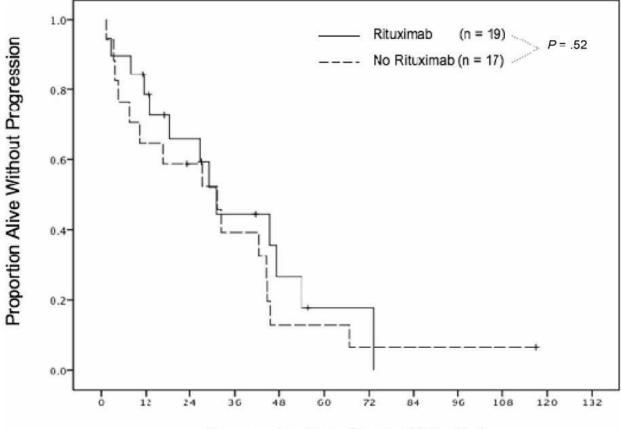


Fig. 7. EFS following ASCT after myeloablative therapy with (broken line) or without peritransplant rituximab (solid line)<sup>95</sup>.



Progression Free Survival (Months)

Fig. 8. PFS for patients receiving ASCT for relapsed/refractory disease<sup>91</sup>.

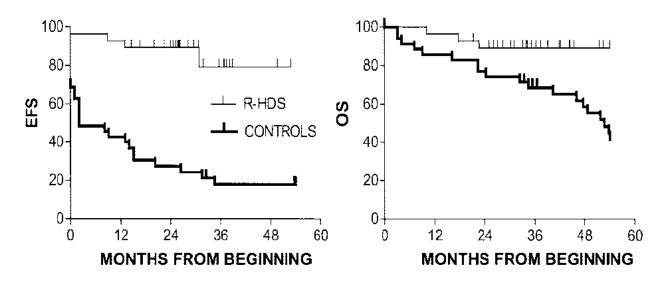


Fig. 9. Overall survival and event-free survival of patients treated with R-HDS versus conventional chemotherapy<sup>96</sup>.

Heavily treated patients with multiple recurrences may undergo an effective salvage with high-dose radioimmunotherapy (RIT) coupled with autologous stem cell support<sup>97</sup>.

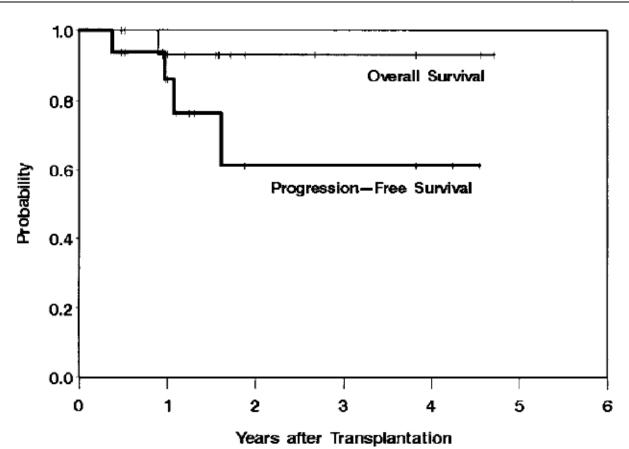


Fig. 10. OS and PFS obtained in patients with relapsed MCL treated with <sup>131</sup>I-tositumomab, etoposide, cyclophosphamide, and ASCT<sup>97</sup>.

#### 7. Allogeneic transplantation for achieving cure

Myeloablative or non-myeloablative allo-HSCT is generally performed in patients with lymphoma relapsing following auto-HSCT, since patients need tumor-free grafts that can induce a graft-versus-lymphoma (GVL) effect.

Physicians should consider the need for an allogeneic transplant when there is a need for a GVL effect (high risk of relapse) which can be predicted by the presence of;

- A high MIPI score
- Aggressive clinical behavior characterized by not achieving a satisfactory response to chemotherapy regimens
- remaining PET positive following ASCT<sup>98</sup>
- Multiple relapses
- and;
- If the patient is young (<55 years) or,
- Autologous stem cells cannot be mobilized
- in patients considered to be eligible following pretransplant screening tests.

Evidence for presence of graft versus lymphoma (GVL) effect has been reported in patients with MCL who underwent allo-HSCT<sup>99</sup>. CVL effect may be observed as conversion to pcr negativity for t(11;14) or achieving CR in the presence of GVHD<sup>99</sup>, or observing lower relapse rates in allo-HSCT recipients compared to patients undergoing autologous

transplantation<sup>100-102</sup>. A healthy comparison of autologous and allogeneic transplants cannot be made at present, due to lack of randomized trials and the different prognostic groups undergoing each transplant type. By using myeloablative regimens for relapsed patients, allogeneic transplants can induce three year event free survivals (EFS)around 50%<sup>99, 101-104</sup>. However, if performed in first CR or PR, EFS induced by allo transplants at three years can be as high as 70%<sup>103</sup>.

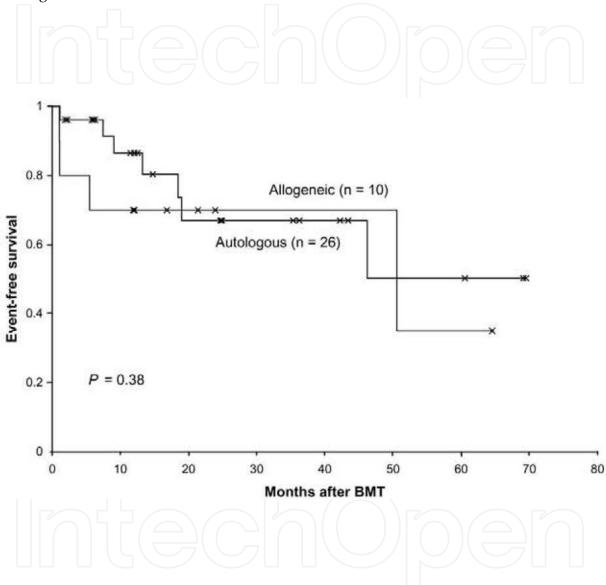


Fig. 11. Event-free survival according to type of BMT<sup>103</sup>.

#### 8. Reduced intensity allogeneic stem cell transplantation

In the allogeneic transplant setting, benefit of cure over transplant related mortality can be positively affected by using reduced intensity (RIC) or non-myeloablative (NMA) conditioning regimens<sup>105-116</sup>. Using donor lymphocyte infusions for GVL effect in patients undergoing allogeneic transplantation with RIC or NMA regimens, low transplant mortality rates (<10%) and higher OS (73 to 85%) and EFS rates (73 to 82%) can be achieved<sup>105, 117, 118</sup>.

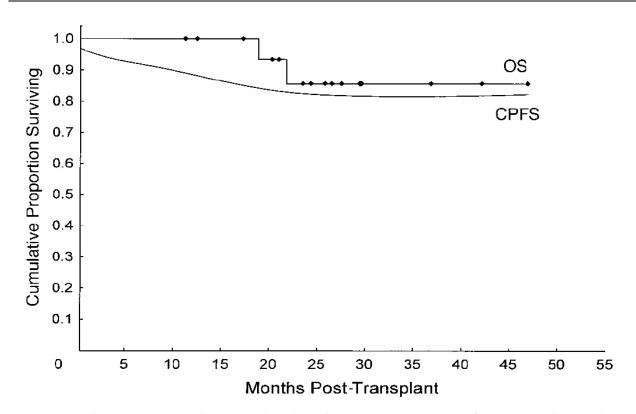


Fig. 12. Kaplan-Meier overall survival (OS) and current progression-free survival (CPFS) accounting for salvage post-DLI in with relapsed MCL who received NMA allogeneic transplantation<sup>105</sup>.

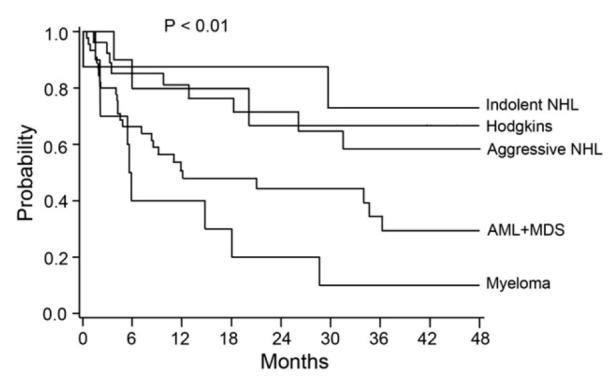


Fig. 13. Favorable overall survival following reduced intensity allogeneneic transplants by disease group<sup>117</sup>.

#### 9. Decision making for transplant options

While standard chemotherapy regimens do not offer long term progression free survival, intensive therapy followed by ASCT should be considered for each newly diagnosed patient<sup>92</sup>, especially if the MIPI score is translating into poor prognosis.

Multiply relapsed patients will not do well after autologous transplants<sup>91, 103</sup>. Best time to perform an autologous transplant is following CR<sup>91, 92, 119, 120</sup> obtained after high-dose therapy with hyperCVAD or HDS regimen, coupled with rituximab<sup>91</sup>, including in vivo purging prior to stem cell collection<sup>81, 92, 96</sup>. With this approach, patients eligible for these sequential intensive regimens may enjoy long term disease free survival, although cure cannot be achieved<sup>121</sup>.

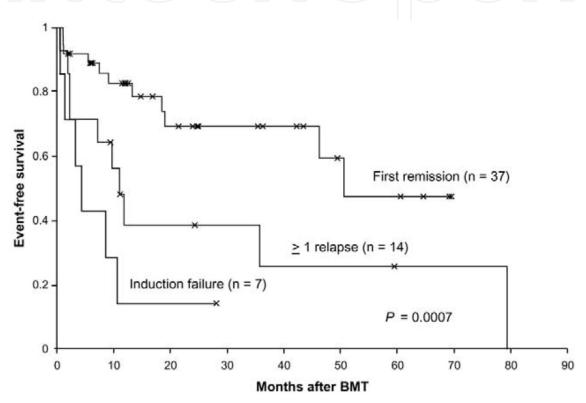


Fig. 14. Event-free survival according to remission status at BMT<sup>103</sup>.

Newly diagnosed and relatively younger patients less than 60 years of age who are not eligible for ASCT or relapsing after ASCT, or patients with recurrence following chemotherapy, can still be cured by allogeneic SCT following an effective salvage regimen inducing CR or near CR<sup>121</sup>.

As the outcome of transplantation is most promising in the newly diagnosed patients without chemorefractory disease, graft contamination and the lack of a survival plateau following autologous transplantation, allogeneic transplantation deserve investigation as the upfront therapy in the management of patients with MCL<sup>92</sup>.

Patients over age 65, and individuals who are not eligible for ASCT with poor prognostic MIPI scores remain as a major challenge for the hematologist. In this group of patients, promising new drugs such as bendamustine, cladribine and bortezomib in combination with synergizing agents such as rituximab, may be offered to patients in the context of clinical trials. One should remember that ASCT with intermediate dose melphalan at doses

between 100 to 140 mg/m<sup>2</sup> can still be utilized in patients over age 65, as well as nonmyeloablative allogeneic stem cell transplants, immediately after achieving a clinical complete response in experienced transplant centers. These approaches have to be tested in clinical trials in the near future.

#### 10. Rational approaches in managing patients with MCL

Due to heterogeneity of disease, paucity of randomized trials, availability of a wide range of chemotherapy regimens including new agents, and altering prognostic factors among patients, an individualized treatment approach should be adapted for each patient diagnosed with MCL. As in therapy of patients with myeloma, long term survival may be achieved following sequential and risk-adapted use of available therapies such as intensive or nucleoside analogue based chemotherapy, followed by autologous and/or reduced intensity allogeneic transplantation, including radioimmunotherapy.

Rational strategy at present can be outlined as;

- 1. After assessing the MIPI score, low risk patients can be followed without transplantation strategy, because of late age of onset, which is over 60 in most patients. This group of patients will do well with standard chemotherapy regimens utilized in NHLs, such as R-CHOP or new generation combination regimens including a purine analogue and rituximab, such as R-FCM or FCR.
- 2. Due to its poor prognosis and failure to achieve improved survival curves with conventional chemotherapy regimens, consideration should be given to upfront autologous transplantation in patients with high MIPI scores. To induce remission FCR, R-hyperCVAD and R-CHOP-14 regimens seem to be effective approaches. Inducing high quality complete responses -molecular remission for t(11;14) translocation- prior to autologous transplantation may further improve disease free and overall survival curves. Success of this strategy has been shown in comparison to historical controls, and also has to be proven in randomized clinical trials.
- 3. Young patients with intermediate MIPI scores may choose to continue with the same approach with the ones with high scores as described above. Elderly patients may receive R-CHOP, R-FCM or FCR as the initial therapy.
- 4. Patients having recurrence following conventional chemotherapy or an autologous transplant and who are determined to be fit for an allogeneic transplant following a careful evaluation and screening tests should be given a chance to have this treatment option to achieve cure.
- 5. Regardless of the MIPI score, patients who are not eligible for high dose therapy and autologous stem cell support and relapsing after conventional chemotherapy regimens and not a candidate for an allogeneic transplant may be treated with bendamustine and cladribine containing regimens in the context of clinical trials.

Transplant strategy can be summarized as;

Patients with predicted poor survival by MIPI score should undergo intensive induction regimens if eligible, to achieve CR, followed by ASCT preferably in first remission. If cure is targeted, especially in young patients, allogeneic SCT may be performed following an excellent cytoreduction with ASCT or intensive chemotherapy regimen including high dose Ara-C and rituximab, such as R-HDS or hyper-CVAD regimens.

#### 11. Emerging new drugs to improve results in the near future

New agents which may have potential to improve outcome of relapsed or refractory patients with MCL include chemicals targeting cyclin D1 and the cell cycle regulatory proteins (cdk), inhibitors of mammalian target of rapamycin, the proteasome, and proapoptotic family members<sup>122</sup>.

Both histone deacetylase (HDAC) and mTOR inhibitors may downregulate cyclin D1 reducing cell cycle drive, which is upregulated in patients with MCL. Cyclin D1 is the key factor for upregulating cellular proliferation rate<sup>22</sup>. HDAC inhibitor sueroylanilide hydroxamic acid (vorinostat) has ability to reduce intracellular cyclin D1 levels in MCL cells, leading to inhibition of progression to S-phase in the cell cycle, and may induce response in MCL patients, currently being tested in phase I clinical trials<sup>123</sup>. Cyclin D1 protein itself can be used as a target for sensitized T-cells, generating a potential treatment option by cellular therapy<sup>124</sup>.

Other than overexpression of cyclin D1, activation of mammalian target of rapamycin (mTOR) also plays a major role in growth and proliferation in MCL cells<sup>125</sup>. Inhibitiors of mTOR (rapamycin, temsirolimus, everolimus) may increase apoptosis and decrease proliferation rate by causing G1 arrest in the cell cycle<sup>126</sup>. Single agent temsirolimus has been shown to have clinical activity in phase II trials, and a randomized phase III trial<sup>127-129</sup>. Alternative mTOR inhibitor everolimus is currently in phase II trials<sup>130, 131</sup>.

Deficiencies of Noxa and Bim proteins and aberrant expression of Bcl-2 may reduce apoptosis and lead to increased resistance to chemotherapeutic agents<sup>132</sup>. Proteasome inhibitors such as bortezomib and flavopiridol may function as accumulating cdk inhibitors (p21/p27), causing cell cycle arrest in malignant cells<sup>132-134</sup>. Accumulation of proapoptotic Noxa protein has been consistently shown in MCL cells treated with bortezomib<sup>135</sup>.

There is a rationale combining bortezomib with HDAC inhibitor vorinostat. Vorinostat may be able to turn off cyclin D1, whereas bortezomib turns on cdk inhibitors p27 and p21, targeting two complementary genetic lesions involved in the cell cycle aberration in MCL cells which may lead to induction of apoptosis<sup>136</sup>. Bortezomib may also be combined with cyproheptadine, recently identified inhibitor of cyclin D1, synergistic in inducing apoptosis in vitro<sup>137</sup>.

Development of new drugs for MCL will further improve the recent achievements in the management options published in the last decade, in a disease currently not curable by medical therapy except allogeneic transplantation.

#### 12. References

- [1] Jaffe ESHNL, Stein H., et al.: Tumours of haematopoietic and lymphoid tissues In: Jaffe ES, et al ed. World Health Organization Classification of Tumours, Lyon, France: IARC Press 2001: 1-352.
- [2] Lenz G, Dreyling M, Hiddemann W. Mantle Cell Lymphoma. *In: Canellos G.P. e al ed. The Lymphomas. 2nd Edition Philadelphia, USA: Elsevier Inc.* 2006: 397-405.
- [3] Velders GA, Kluin-Nelemans JC, De Boer CJ, Hermans J, Noordijk EM, Schuuring E *et al.* Mantle-cell lymphoma: a population-based clinical study. *J Clin Oncol* 1996; 14(4): 1269-74.

- [4] Walsh SH, Rosenquist R. Immunoglobulin gene analysis of mature B-cell malignancies: reconsideration of cellular origin and potential antigen involvement in pathogenesis. *Med Oncol* 2005; 22(4): 327-41.
- [5] Welzel N, Le T, Marculescu R, Mitterbauer G, Chott A, Pott C *et al.* Templated nucleotide addition and immunoglobulin JH-gene utilization in t(11;14) junctions: implications for the mechanism of translocation and the origin of mantle cell lymphoma. *Cancer Res* 2001; 61(4): 1629-36.
- [6] Vandenberghe E, De Wolf-Peeters C, Vaughan Hudson G, Vaughan Hudson B, Pittaluga S, Anderson L *et al.* The clinical outcome of 65 cases of mantle cell lymphoma initially treated with non-intensive therapy by the British National Lymphoma Investigation Group. *Br J Haematol* 1997; 99(4): 842-7.
- [7] Dreyling M. Hyper-CVAD in mantle-cell lymphoma: really "hyper" or just hype? *Leuk Lymphoma* 2008; 49(6): 1017-8.
- [8] Zucca E, Roggero E, Pinotti G, Pedrinis E, Cappella C, Venco A *et al.* Patterns of survival in mantle cell lymphoma. *Ann Oncol* 1995; 6(3): 257-62.
- [9] Singleton TP, Anderson MM, Ross CW, Schnitzer B. Leukemic phase of mantle cell lymphoma, blastoid variant. *Am J Clin Pathol* 1999; 111(4): 495-500.
- [10] Schlette E, Lai R, Onciu M, Doherty D, Bueso-Ramos C, Medeiros LJ. Leukemic mantle cell lymphoma: clinical and pathologic spectrum of twenty-three cases. *Mod Pathol* 2001; 14(11): 1133-40.
- [11] Bentz M, Plesch A, Bullinger L, Stilgenbauer S, Ott G, Muller-Hermelink HK et al. t(11;14)-positive mantle cell lymphomas exhibit complex karyotypes and share similarities with B-cell chronic lymphocytic leukemia. *Genes Chromosomes Cancer* 2000; 27(3): 285-94.
- [12] Zukerberg LR, Medeiros LJ, Ferry JA, Harris NL. Diffuse low-grade B-cell lymphomas. Four clinically distinct subtypes defined by a combination of morphologic and immunophenotypic features. *Am J Clin Pathol* 1993; 100(4): 373-85.
- [13] Pittaluga S, Wlodarska I, Stul MS, Thomas J, Verhoef G, Cassiman JJ *et al.* Mantle cell lymphoma: a clinicopathological study of 55 cases. *Histopathology* 1995; 26(1): 17-24.
- [14] de Boer CJ, Schuuring E, Dreef E, Peters G, Bartek J, Kluin PM *et al.* Cyclin D1 protein analysis in the diagnosis of mantle cell lymphoma. *Blood* 1995; 86(7): 2715-23.
- [15] Fan H, Gulley ML, Gascoyne RD, Horsman DE, Adomat SA, Cho CG. Molecular methods for detecting t(11;14) translocations in mantle-cell lymphomas. *Diagn Mol Pathol* 1998; 7(4): 209-14.
- [16] Samaha H, Dumontet C, Ketterer N, Moullet I, Thieblemont C, Bouafia F *et al.* Mantle cell lymphoma: a retrospective study of 121 cases. *Leukemia* 1998; 12(8): 1281-7.
- [17] Espinet B, Sole F, Woessner S, Bosch F, Florensa L, Campo E et al. Translocation (11;14)(q13;q32) and preferential involvement of chromosomes 1, 2, 9, 13, and 17 in mantle cell lymphoma. *Cancer Genet Cytogenet* 1999; 111(1): 92-8.
- [18] Li JY, Gaillard F, Moreau A, Harousseau JL, Laboisse C, Milpied N *et al.* Detection of translocation t(11;14)(q13;q32) in mantle cell lymphoma by fluorescence in situ hybridization. *Am J Pathol* 1999; 154(5): 1449-52.
- [19] Lai R, Medeiros LJ. Pathologic diagnosis of mantle cell lymphoma. *Clin Lymphoma* 2000; 1(3): 197-206; discussion 207-8.

- [20] de Boer CJ, van Krieken JH, Kluin-Nelemans HC, Kluin PM, Schuuring E. Cyclin D1 messenger RNA overexpression as a marker for mantle cell lymphoma. *Oncogene* 1995; 10(9): 1833-40.
- [21] Rosenwald A, Wright G, Wiestner A, Chan WC, Connors JM, Campo E *et al.* The proliferation gene expression signature is a quantitative integrator of oncogenic events that predicts survival in mantle cell lymphoma. *Cancer Cell* 2003; 3(2): 185-97.
- [22] Wiestner A, Tehrani M, Chiorazzi M, Wright G, Gibellini F, Nakayama K et al. Point mutations and genomic deletions in CCND1 create stable truncated cyclin D1 mRNAs that are associated with increased proliferation rate and shorter survival. Blood 2007; 109(11): 4599-606.
- [23] Garcia-Conde J, Cabanillas F. Mantle cell lymphoma: a Tymphoproliferative disorder associated with aberrant function of the cell cycle. *Leukemia* 1996; 10 Suppl 2: s78-83.
- [24] Hartwell LH, Kastan MB. Cell cycle control and cancer. Science 1994; 266(5192): 1821-8.
- [25] Feldman AL, Pittaluga S, Jaffe ES. Classification and Histopathology of the Lymphomas. In: Canellos G.P. et al. The Lymphomas. 2nd Edition Philadelphia, USA: Elsevier Inc. 2006: 2-38.
- [26] Hoster E, Dreyling M, Klapper W, Gisselbrecht C, van Hoof A, Kluin-Nelemans HC et al. A new prognostic index (MIPI) for patients with advanced-stage mantle cell lymphoma. Blood 2008; 111(2): 558-65.
- [27] Shipp MA. Prognostic factors in aggressive non-Hodgkin's lymphoma: who has "high-risk" disease? *Blood* 1994; 83(5): 1165-73.
- [28] Moller MB, Pedersen NT, Christensen BE. Mantle cell lymphoma: prognostic capacity of the Follicular Lymphoma International Prognostic Index. *Br J Haematol* 2006; 133(1): 43-9.
- [29] Solal-Celigny P, Roy P, Colombat P, White J, Armitage JO, Arranz-Saez R *et al.* Follicular lymphoma international prognostic index. *Blood* 2004; 104(5): 1258-65.
- [30] Tiemann M, Schrader C, Klapper W, Dreyling MH, Campo E, Norton A *et al.* Histopathology, cell proliferation indices and clinical outcome in 304 patients with mantle cell lymphoma (MCL): a clinicopathological study from the European MCL Network. *Br J Haematol* 2005; 131(1): 29-38.
- [31] Geisler CH, Kolstad A, Laurell A, Raty R, Jerkeman M, Eriksson M *et al.* The Mantle Cell Lymphoma International Prognostic Index (MIPI) is superior to the International Prognostic Index (IPI) in predicting survival following intensive firstline immunochemotherapy and autologous stem cell transplantation (ASCT). *Blood* 2010; 115(8): 1530-3.
- [32] Determann O, Hoster E, Ott G, Wolfram Bernd H, Loddenkemper C, Leo Hansmann M et al. Ki-67 predicts outcome in advanced-stage mantle cell lymphoma patients treated with anti-CD20 immunochemotherapy: results from randomized trials of the European MCL Network and the German Low Grade Lymphoma Study Group. Blood 2008; 111(4): 2385-7.

- [33] Katzenberger T, Petzoldt C, Holler S, Mader U, Kalla J, Adam P *et al.* The Ki67 proliferation index is a quantitative indicator of clinical risk in mantle cell lymphoma. *Blood* 2006; 107(8): 3407.
- [34] Klapper W, Hoster E, Determann O, Oschlies I, van der Laak J, Berger F *et al.* Ki-67 as a prognostic marker in mantle cell lymphoma-consensus guidelines of the pathology panel of the European MCL Network. *J Hematop* 2009.
- [35] Hartmann E, Fernandez V, Moreno V, Valls J, Hernandez L, Bosch F *et al.* Five-gene model to predict survival in mantle-cell lymphoma using frozen or formalin-fixed, paraffin-embedded tissue. *J Clin Oncol* 2008; 26(30): 4966-72.
- [36] Wilson WH, Armitage JO. Non Hodgkin's Lymphoma. *In: Abeloff, M.D. et al ed. Abeloff's Clinical Oncology* 2008: 2371-2404.
- [37] Zucca E, Fontana S, Roggero E, Pedrinis E, Pampallona S, Cavalli F. Treatment and prognosis of centrocytic (mantle cell) lymphoma: a retrospective analysis of twenty-six patients treated in one institution. *Leuk Lymphoma* 1994; 13(1-2): 105-10.
- [38] Khouri IF, Romaguera J, Kantarjian H, Palmer JL, Pugh WC, Korbling M *et al.* Hyper-CVAD and high-dose methotrexate/cytarabine followed by stem-cell transplantation: an active regimen for aggressive mantle-cell lymphoma. *J Clin* Oncol 1998; 16(12): 3803-9.
- [39] Meusers P, Engelhard M, Bartels H, Binder T, Fulle HH, Gorg K *et al.* Multicentre randomized therapeutic trial for advanced centrocytic lymphoma: anthracycline does not improve the prognosis. *Hematol Oncol* 1989; 7(5): 365-80.
- [40] Foran JM, Rohatiner AZ, Cunningham D, Popescu RA, Solal-Celigny P, Ghielmini M et al. European phase II study of rituximab (chimeric anti-CD20 monoclonal antibody) for patients with newly diagnosed mantle-cell lymphoma and previously treated mantle-cell lymphoma, immunocytoma, and small B-cell lymphocytic lymphoma. J Clin Oncol 2000; 18(2): 317-24.
- [41] Coiffier B, Haioun C, Ketterer N, Engert A, Tilly H, Ma D et al. Rituximab (anti-CD20 monoclonal antibody) for the treatment of patients with relapsing or refractory aggressive lymphoma: a multicenter phase II study. Blood 1998; 92(6): 1927-32.
- [42] Howard OM, Gribben JG, Neuberg DS, Grossbard M, Poor C, Janicek MJ et al. Rituximab and CHOP induction therapy for newly diagnosed mantle-cell lymphoma: molecular complete responses are not predictive of progression-free survival. J Clin Oncol 2002; 20(5): 1288-94.
- [43] Gianni AM, Cortelazzo S, Magni M, Martelli M. Rituximab: enhancing stem cell transplantation in mantle cell lymphoma. *Bone Marrow Transplant* 2002; 29 Suppl 1: S10-3.
- [44] Lenz G, Dreyling M, Hoster E, Wormann B, Duhrsen U, Metzner B et al. Immunochemotherapy with rituximab and cyclophosphamide, doxorubicin, vincristine, and prednisone significantly improves response and time to treatment failure, but not long-term outcome in patients with previously untreated mantle cell lymphoma: results of a prospective randomized trial of the German Low Grade Lymphoma Study Group (GLSG). J Clin Oncol 2005; 23(9): 1984-92.
- [45] Hiddemann W, Unterhalt M, Herrmann R, Woltjen HH, Kreuser ED, Trumper L *et al.* Mantle-cell lymphomas have more widespread disease and a slower response to

chemotherapy compared with follicle-center lymphomas: results of a prospective comparative analysis of the German Low-Grade Lymphoma Study Group. *J Clin Oncol* 1998; 16(5): 1922-30.

- [46] Teodorovic I, Pittaluga S, Kluin-Nelemans JC, Meerwaldt JH, Hagenbeek A, van Glabbeke M *et al.* Efficacy of four different regimens in 64 mantle-cell lymphoma cases: clinicopathologic comparison with 498 other non-Hodgkin's lymphoma subtypes. European Organization for the Research and Treatment of Cancer Lymphoma Cooperative Group. J Clin Oncol 1995; 13(11): 2819-26.
- [47] Lenz G, Hiddemann W, Dreyling M. The role of fludarabine in the treatment of follicular and mantle cell lymphoma. *Cancer* 2004; 101(5): 883-93.
- [48] Decaudin D, Bosq J, Tertian G, Nedellec G, Bennaceur A, Venuat AM *et al.* Phase II trial of fludarabine monophosphate in patients with mantle-cell lymphomas. J Clin Oncol 1998; 16(2): 579-83.
- [49] Zinzani PL, Magagnoli M, Moretti L, Battista R, Ronconi F, De Renzo A *et al.* Fludarabine-based chemotherapy in untreated mantle cell lymphomas: an encouraging experience in 29 patients. *Haematologica* 1999; 84(11): 1002-6.
- [50] Zinzani PL, Magagnoli M, Moretti L, De Renzo A, Battista R, Zaccaria A *et al.* Randomized trial of fludarabine versus fludarabine and idarubicin as frontline treatment in patients with indolent or mantle-cell lymphoma. *J Clin Oncol* 2000; 18(4): 773-9.
- [51] Cohen BJ, Moskowitz C, Straus D, Noy A, Hedrick E, Zelenetz A. Cyclophosphamide/fludarabine (CF) is active in the treatment of mantle cell lymphoma. *Leuk Lymphoma* 2001; 42(5): 1015-22.
- [52] Forstpointner R, Dreyling M, Repp R, Hermann S, Hanel A, Metzner B *et al.* The addition of rituximab to a combination of fludarabine, cyclophosphamide, mitoxantrone (FCM) significantly increases the response rate and prolongs survival as compared with FCM alone in patients with relapsed and refractory follicular and mantle cell lymphomas: results of a prospective randomized study of the German Low-Grade Lymphoma Study Group. *Blood* 2004; 104(10): 3064-71.
- [53] Forstpointner R, Unterhalt M, Dreyling M, Bock HP, Repp R, Wandt H *et al.* Maintenance therapy with rituximab leads to a significant prolongation of response duration after salvage therapy with a combination of rituximab, fludarabine, cyclophosphamide, and mitoxantrone (R-FCM) in patients with recurring and refractory follicular and mantle cell lymphomas: Results of a prospective randomized study of the German Low Grade Lymphoma Study Group (GLSG). *Blood* 2006; 108(13): 4003-8.
- [54] Hiddemann W, Dreyling M, Unterhalt M. Rituximab plus chemotherapy in follicular and mantle cell lymphomas. *Semin Oncol* 2003; 30(1 Suppl 2): 16-20.
- [55] Thomas DW, Owen RG, Johnson SA, Hillmen P, Seymour JF, Wolf MM *et al.* Superior quality and duration of responses among patients with mantle-cell lymphoma treated with fludarabine and cyclophosphamide with or without rituximab compared with prior responses to CHOP. *Leuk Lymphoma* 2005; 46(4): 549-52.
- [56] Eve HE, Linch D, Qian W, Ross M, Seymour JF, Smith P *et al.* Toxicity of fludarabine and cyclophosphamide with or without rituximab as initial therapy for patients

with previously untreated mantle cell lymphoma: results of a randomised phase II study. *Leuk Lymphoma* 2009; 50(2): 211-5.

- [57] Romaguera JE, Fayad LE, Feng L, Hartig K, Weaver P, Rodriguez MA *et al.* Ten-year follow-up after intense chemoimmunotherapy with Rituximab-HyperCVAD alternating with Rituximab-high dose methotrexate/cytarabine (R-MA) and without stem cell transplantation in patients with untreated aggressive mantle cell lymphoma. *Br J Haematol* 2010; 150(2): 200-8.
- [58] Romaguera JE, Khouri IF, Kantarjian HM, Hagemeister FB, Rodriguez MA, McLaughlin P *et al.* Untreated aggressive mantle cell lymphoma: results with intensive chemotherapy without stem cell transplant in elderly patients. *Leuk Lymphoma* 2000; 39(1-2): 77-85.
- [59] Dreyling MH, Hoster E, Hermine O, Kluin-Nelemans JC, Walewski J, Trneny M et al. European MCL Network: An Update on Current First Line Trials. Blood 2007; 110: 388 Abstract.
- [60] Martin P, Chadburn A, Christos P, Furman R, Ruan J, Joyce MA et al. Intensive treatment strategies may not provide superior outcomes in mantle cell lymphoma: overall survival exceeding 7 years with standard therapies. Ann Oncol 2008; 19(7): 1327-30.
- [61] Friedberg JW, Vose JM, Kelly JL, Young F, Bernstein SH, Peterson D *et al.* The combination of bendamustine, bortezomib, and rituximab for patients with relapsed/refractory indolent and mantle cell non-Hodgkin lymphoma. *Blood* 2011; 117(10): 2807-12.
- [62] Ruan J, Martin P, Furman RR, Lee SM, Cheung K, Vose JM *et al.* Bortezomib plus CHOP-rituximab for previously untreated diffuse large B-cell lymphoma and mantle cell lymphoma. *J Clin Oncol* 2011; 29(6): 690-7.
- [63] Lamm W, Kaufmann H, Raderer M, Hoffmann M, Chott A, Zielinski C et al. Bortezomib combined with rituximab and dexamethasone is an active regimen for patients with relapsed and chemotherapy-refractory mantle cell lymphoma. *Haematologica* 2011; 96(7): 1008-14.
- [64] Habermann TM, Lossos IS, Justice G, Vose JM, Wiernik PH, McBride K *et al.* Lenalidomide oral monotherapy produces a high response rate in patients with relapsed or refractory mantle cell lymphoma. *Br J Haematol* 2009; 145(3): 344-9.
- [65] Richardson SJ, Eve HE, Copplestone JA, Dyer MJ, Rule SA. Activity of thalidomide and lenalidomide in mantle cell lymphoma. *Acta Haematol* 2010; 123(1): 21-9.
- [66] Qian Z, Zhang L, Cai Z, Sun L, Wang H, Yi Q *et al.* Lenalidomide synergizes with dexamethasone to induce growth arrest and apoptosis of mantle cell lymphoma cells in vitro and in vivo. *Leuk Res* 2011; 35(3): 380-6.
- [67] Weide R, Hess G, Koppler H, Heymanns J, Thomalla J, Aldaoud A *et al.* High antilymphoma activity of bendamustine/mitoxantrone/rituximab in rituximab pretreated relapsed or refractory indolent lymphomas and mantle cell lymphomas. A multicenter phase II study of the German Low Grade Lymphoma Study Group (GLSG). *Leuk Lymphoma* 2007; 48(7): 1299-306.
- [68] Rummel MJ, Al-Batran SE, Kim SZ, Welslau M, Hecker R, Kofahl-Krause D *et al.* Bendamustine plus rituximab is effective and has a favorable toxicity profile in the

treatment of mantle cell and low-grade non-Hodgkin's lymphoma. *J Clin Oncol* 2005; 23(15): 3383-9.

- [69] Rummel MJ, Gregory SA. Bendamustine's emerging role in the management of lymphoid malignancies. *Semin Hematol* 2011; 48 Suppl 1: S24-36.
- [70] Abbott BL. Diagnosis and management of lymphoma. *Clin Lymphoma Myeloma* 2006; 7(1): 30-2.
- [71] O'Connor OA, Portlock C, Moskowitz C, Straus D, Hamlin P, Stubblefield M *et al.* A multicentre phase II clinical experience with the novel aza-epothilone Ixabepilone (BMS247550) in patients with relapsed or refractory indolent non-Hodgkin lymphoma and mantle cell lymphoma. *Br J Haematol* 2008; 143(2): 201-9.
- [72] Ansell SM, Tang H, Kurtin PJ, Koenig PA, Inwards DJ, Shah K *et al.* Temsirolimus and rituximab in patients with relapsed or refractory mantle cell lymphoma: a phase 2 study. *Lancet Oncol* 2011; 12(4): 361-8.
- [73] Galimberti S, Petrini M. Temsirolimus in the treatment of relapsed and/or refractory mantle cell lymphoma. *Cancer Manag Res* 2010; 2: 181-9.
- [74] Hoy SM, McKeage K. Temsirolimus: In relapsed and/or refractory mantle cell lymphoma. *Drugs* 2010; 70(14): 1819-29.
- [75] Garnock-Jones KP. Bendamustine: a review of its use in the management of indolent non-Hodgkin's lymphoma and mantle cell lymphoma. *Drugs* 2010; 70(13): 1703-18.
- [76] Zhang L, Qian Z, Cai Z, Sun L, Wang H, Bartlett JB *et al.* Synergistic antitumor effects of lenalidomide and rituximab on mantle cell lymphoma in vitro and in vivo. *Am J Hematol* 2009; 84(9): 553-9.
- [77] Ohmachi K, Ando K, Ogura M, Uchida T, Itoh K, Kubota N *et al.* Multicenter phase II study of bendamustine for relapsed or refractory indolent B-cell non-Hodgkin lymphoma and mantle cell lymphoma. *Cancer Sci* 2010; 101(9): 2059-64.
- [78] Ogura M, Ando K, Taniwaki M, Watanabe T, Uchida T, Ohmachi K *et al.* A Feasibility and Pharmacokinetic Study of Bendamustine Hydrochloride in Combination With Rituximab in Relapsed or Refractory Aggressive B-Cell Non-Hodgkin Lymphoma. *Cancer Sci* 2011.
- [79] Ogura M. Current treatment strategy and new agents in mantle cell lymphoma. *Int J Hematol* 2010; 92(1): 25-32.
- [80] Cortelazzo S, Ponzoni M, Ferreri AJ, Dreyling M. Mantle cell lymphoma. *Crit Rev Oncol Hematol* 2011.
- [81] Oyan B, Koc Y, Kansu E. Successful salvage with high-dose sequential chemotherapy coupled with in vivo purging and autologous stem cell transplantation in 2 patients with primary refractory mantle cell lymphoma presenting in the leukemic phase. *Int J Hematol* 2005; 81(2): 155-8.
- [82] Keating GM. Rituximab: a review of its use in chronic lymphocytic leukaemia, lowgrade or follicular lymphoma and diffuse large B-cell lymphoma. *Drugs* 2010; 70(11): 1445-76.
- [83] Spurgeon SE, Pindyck T, Okada C, Chen Y, Chen Z, Mater E *et al.* Cladribine plus rituximab is an effective therapy for newly diagnosed mantle cell lymphoma. *Leuk Lymphoma* 2011.

- [84] Weigert O, Illidge T, Hiddemann W, Dreyling M. Recommendations for the use of yttrium-90 ibritumomab tiuxetan in malignant lymphoma. *Cancer* 2006; 107(4): 686-95.
- [85] Bethge WA, Lange T, Meisner C, von Harsdorf S, Bornhaeuser M, Federmann B *et al.* Radioimmunotherapy with yttrium-90-ibritumomab tiuxetan as part of a reducedintensity conditioning regimen for allogeneic hematopoietic cell transplantation in patients with advanced non-Hodgkin lymphoma: results of a phase 2 study. *Blood* 2010; 116(10): 1795-802.
- [86] Leahy MF, Turner JH. Radioimmunotherapy of relapsed indolent non-Hodgkin lymphoma with 131I-rituximab in routine clinical practice: 10-year singleinstitution experience of 142 consecutive patients. *Blood* 2011; 117(1): 45-52.
- [87] Krishnan A, Nademanee A, Fung HC, Raubitschek AA, Molina A, Yamauchi D et al. Phase II trial of a transplantation regimen of yttrium-90 ibritumomab tiuxetan and high-dose chemotherapy in patients with non-Hodgkin's lymphoma. J Clin Oncol 2008; 26(1): 90-5.
- [88] Vose JM, Bierman PJ, Weisenburger DD, Lynch JC, Bociek Y, Chan WC et al. Autologous hematopoietic stem cell transplantation for mantle cell lymphoma. Biol Blood Marrow Transplant 2000; 6(6): 640-5.
- [89] Jacobsen E, Freedman A. An update on the role of high-dose therapy with autologous or allogeneic stem cell transplantation in mantle cell lymphoma. *Curr Opin Oncol* 2004; 16(2): 106-13.
- [90] Murali S, Winton E, Waller EK, Heffner LT, Lonial S, Flowers C *et al.* Long-term progression-free survival after early autologous transplantation for mantle-cell lymphoma. *Bone Marrow Transplant* 2008; 42(8): 529-34.
- [91] Tam CS, Bassett R, Ledesma C, Korbling M, Alousi A, Hosing C *et al.* Mature results of the M. D. Anderson Cancer Center risk-adapted transplantation strategy in mantle cell lymphoma. *Blood* 2009; 113(18): 4144-52.
- [92] Kasamon YL. Blood or marrow transplantation for mantle cell lymphoma. *Curr Opin* Oncol 2007; 19(2): 128-35.
- [93] Dreyling M, Lenz G, Hoster E, Van Hoof A, Gisselbrecht C, Schmits R *et al.* Early consolidation by myeloablative radiochemotherapy followed by autologous stem cell transplantation in first remission significantly prolongs progression-free survival in mantle-cell lymphoma: results of a prospective randomized trial of the European MCL Network. *Blood* 2005; 105(7): 2677-84.
- [94] van 't Veer MB, de Jong D, MacKenzie M, Kluin-Nelemans HC, van Oers MH, Zijlstra J *et al.* High-dose Ara-C and beam with autograft rescue in R-CHOP responsive mantle cell lymphoma patients. *Br J Haematol* 2009; 144(4): 524-30.
- [95] Dreger P, Laport GG. Controversies in lymphoma: the role of hematopoietic cell transplantation for mantle cell lymphoma and peripheral T cell lymphoma. *Biol Blood Marrow Transplant* 2008; 14(1 Suppl 1): 100-7.
- [96] Gianni AM, Magni M, Martelli M, Di Nicola M, Carlo-Stella C, Pilotti S et al. Long-term remission in mantle cell lymphoma following high-dose sequential chemotherapy and in vivo rituximab-purged stem cell autografting (R-HDS regimen). Blood 2003; 102(2): 749-55.

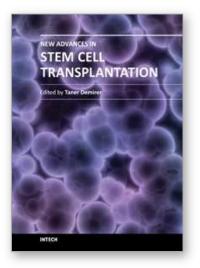
- [97] Gopal AK, Rajendran JG, Petersdorf SH, Maloney DG, Eary JF, Wood BL *et al.* Highdose chemo-radioimmunotherapy with autologous stem cell support for relapsed mantle cell lymphoma. *Blood* 2002; 99(9): 3158-62.
- [98] Svoboda J, Andreadis C, Elstrom R, Chong EA, Downs LH, Berkowitz A *et al.* Prognostic value of FDG-PET scan imaging in lymphoma patients undergoing autologous stem cell transplantation. *Bone Marrow Transplant* 2006; 38(3): 211-6.
- [99] Khouri IF, Lee MS, Romaguera J, Mirza N, Kantarjian H, Korbling M *et al.* Allogeneic hematopoietic transplantation for mantle-cell lymphoma: molecular remissions and evidence of graft-versus-malignancy. *Ann Oncol* 1999; 10(11): 1293-9.
- [100] Nademanee A. Transplantation for non-Hodgkin lymphoma. *Expert Rev Hematol* 2009; 2(4): 425-42.
- [101] Popplewell L, Nademanee A, Carter N, al. e. Autologous vs allogeneic cell transplantation for mantle cell lymphoma (MCL): outcomes over a 10-year period at City of Hope. *Blood* 2004; 104: abstract 894.
- [102] Ganti AK, Bierman PJ, Lynch JC, Bociek RG, Vose JM, Armitage JO. Hematopoietic stem cell transplantation in mantle cell lymphoma. *Ann Oncol* 2005; 16(4): 618-24.
- [103] Kasamon YL, Jones RJ, Diehl LF, Nayer H, Borowitz MJ, Garrett-Mayer E *et al.* Outcomes of autologous and allogeneic blood or marrow transplantation for mantle cell lymphoma. *Biol Blood Marrow Transplant* 2005; 11(1): 39-46.
- [104] Laudi N, Arora M, Burns L, McGlave P, Miller J, Bohac G *et al.* Efficacy of high-dose therapy and hematopoietic stem cell transplantation for mantle cell lymphoma. *Am J Hematol* 2006; 81(7): 519-24.
- [105] Khouri IF, Lee MS, Saliba RM, Jun G, Fayad L, Younes A *et al.* Nonablative allogeneic stem-cell transplantation for advanced/recurrent mantle-cell lymphoma. J Clin Oncol 2003; 21(23): 4407-12.
- [106] Corradini P, Tarella C, Olivieri A, Gianni AM, Voena C, Zallio F *et al.* Reducedintensity conditioning followed by allografting of hematopoietic cells can produce clinical and molecular remissions in patients with poor-risk hematologic malignancies. *Blood* 2002; 99(1): 75-82.
- [107] Maris MB, Sandmaier BM, Storer BE, Chauncey T, Stuart MJ, Maziarz RT et al. Allogeneic hematopoietic cell transplantation after fludarabine and 2 Gy total body irradiation for relapsed and refractory mantle cell lymphoma. Blood 2004; 104(12): 3535-42.
- [108] Morris E, Thomson K, Craddock C, Mahendra P, Milligan D, Cook G et al. Outcomes after alemtuzumab-containing reduced-intensity allogeneic transplantation regimen for relapsed and refractory non-Hodgkin lymphoma. Blood 2004; 104(13): 3865-71.
- [109] Robinson SP, Goldstone AH, Mackinnon S, Carella A, Russell N, de Elvira CR et al. Chemoresistant or aggressive lymphoma predicts for a poor outcome following reduced-intensity allogeneic progenitor cell transplantation: an analysis from the Lymphoma Working Party of the European Group for Blood and Bone Marrow Transplantation. Blood 2002; 100(13): 4310-6.

- [110] Le Gouill S, Mohty M, Guillaume T, Gastinne T, Moreau P. Allogeneic stem cell transplantation in mantle cell lymphoma: where are we now and which way should we go? *Semin Hematol* 2011; 48(3): 227-39.
- [111] Sorror ML. Comorbidities and hematopoietic cell transplantation outcomes. *Hematology Am Soc Hematol Educ Program* 2010; 2010: 237-47.
- [112] Baron F, Storb R, Storer BE, Maris MB, Niederwieser D, Shizuru JA *et al.* Factors associated with outcomes in allogeneic hematopoietic cell transplantation with nonmyeloablative conditioning after failed myeloablative hematopoietic cell transplantation. *J Clin Oncol* 2006; 24(25): 4150-7.
- [113] Sorror ML, Maris MB, Storer B, Sandmaier BM, Diaconescu R, Flowers C *et al.* Comparing morbidity and mortality of HLA-matched unrelated donor hematopoietic cell transplantation after nonmyeloablative and myeloablative conditioning: influence of pretransplantation comorbidities. *Blood* 2004; 104(4): 961-8.
- [114] Hill BT, Bolwell BJ, Rybicki L, Dean R, Kalaycio M, Pohlman B et al. Nonmyeloablative second transplants are associated with lower nonrelapse mortality and superior survival than myeloablative second transplants. *Biol Blood Marrow Transplant* 2010; 16(12): 1738-46.
- [115] Tomblyn M, Brunstein C, Burns LJ, Miller JS, MacMillan M, DeFor TE et al. Similar and promising outcomes in lymphoma patients treated with myeloablative or nonmyeloablative conditioning and allogeneic hematopoietic cell transplantation. Biol Blood Marrow Transplant 2008; 14(5): 538-45.
- [116] Dreger P, Brand R, Milligan D, Corradini P, Finke J, Lambertenghi Deliliers G et al. Reduced-intensity conditioning lowers treatment-related mortality of allogeneic stem cell transplantation for chronic lymphocytic leukemia: a population-matched analysis. *Leukemia* 2005; 19(6): 1029-33.
- [117] Warlick ED, Tomblyn M, Cao Q, Defor T, Blazar BR, Macmillan M et al. Reducedintensity conditioning followed by related allografts in hematologic malignancies: long-term outcomes most successful in indolent and aggressive non-Hodgkin lymphomas. Biol Blood Marrow Transplant 2011; 17(7): 1025-32.
- [118] Shea T, Johnson J, Westervelt P, Farag S, McCarty J, Bashey A et al. Reduced-Intensity Allogeneic Transplantation Provides High Event-Free and Overall Survival in Patients with Advanced Indolent B Cell Malignancies: CALGB 109901. Biol Blood Marrow Transplant 2011.
- [119] Till BG, Gooley TA, Crawford N, Gopal AK, Maloney DG, Petersdorf SH *et al.* Effect of remission status and induction chemotherapy regimen on outcome of autologous stem cell transplantation for mantle cell lymphoma. *Leuk Lymphoma* 2008; 49(6): 1062-73.
- [120] Evens AM, Winter JN, Hou N, Nelson BP, Rademaker A, Patton D et al. A phase II clinical trial of intensive chemotherapy followed by consolidative stem cell transplant: long-term follow-up in newly diagnosed mantle cell lymphoma. Br J Haematol 2008; 140(4): 385-93.
- [121] Tam CS, Khouri IF. Autologous and allogeneic stem cell transplantation: rising therapeutic promise for mantle cell lymphoma. *Leuk Lymphoma* 2009; 50(8): 1239-48.

- [122] Diefenbach CS, O'Connor OA. Mantle cell lymphoma in relapse: the role of emerging new drugs. *Curr Opin Oncol* 2010; 22(5): 419-23.
- [123] Watanabe T, Kato H, Kobayashi Y, Yamasaki S, Morita-Hoshi Y, Yokoyama H et al. Potential efficacy of the oral histone deacetylase inhibitor vorinostat in a phase I trial in follicular and mantle cell lymphoma. *Cancer Sci* 2010; 101(1): 196-200.
- [124] Wang M, Sun L, Qian J, Han X, Zhang L, Lin P *et al.* Cyclin D1 as a universally expressed mantle cell lymphoma-associated tumor antigen for immunotherapy. *Leukemia* 2009; 23(7): 1320-8.
- [125] Rudelius M, Pittaluga S, Nishizuka S, Pham TH, Fend F, Jaffe ES *et al.* Constitutive activation of Akt contributes to the pathogenesis and survival of mantle cell lymphoma. *Blood* 2006; 108(5): 1668-76.
- [126] Peponi E, Drakos E, Reyes G, Leventaki V, Rassidakis GZ, Medeiros LJ. Activation of mammalian target of rapamycin signaling promotes cell cycle progression and protects cells from apoptosis in mantle cell lymphoma. Am J Pathol 2006; 169(6): 2171-80.
- [127] Ansell SM, Inwards DJ, Rowland KM, Jr., Flynn PJ, Morton RF, Moore DF, Jr. *et al.* Low-dose, single-agent temsirolimus for relapsed mantle cell lymphoma: a phase 2 trial in the North Central Cancer Treatment Group. *Cancer* 2008; 113(3): 508-14.
- [128] Witzig TE, Geyer SM, Ghobrial I, Inwards DJ, Fonseca R, Kurtin P et al. Phase II trial of single-agent temsirolimus (CCI-779) for relapsed mantle cell lymphoma. J Clin Oncol 2005; 23(23): 5347-56.
- [129] Hess G, Herbrecht R, Romaguera J, Verhoef G, Crump M, Gisselbrecht C *et al.* Phase III study to evaluate temsirolimus compared with investigator's choice therapy for the treatment of relapsed or refractory mantle cell lymphoma. *J Clin Oncol* 2009; 27(23): 3822-9.
- [130] Witzig TE, Reeder CB, LaPlant BR, Gupta M, Johnston PB, Micallef IN *et al.* A phase II trial of the oral mTOR inhibitor everolimus in relapsed aggressive lymphoma. *Leukemia* 2011; 25(2): 341-7.
- [131] Yee KW, Zeng Z, Konopleva M, Verstovsek S, Ravandi F, Ferrajoli A et al. Phase I/II study of the mammalian target of rapamycin inhibitor everolimus (RAD001) in patients with relapsed or refractory hematologic malignancies. Clin Cancer Res 2006; 12(17): 5165-73.
- [132] Marshansky V, Wang X, Bertrand R, Luo H, Duguid W, Chinnadurai G et al. Proteasomes modulate balance among proapoptotic and antiapoptotic Bcl-2 family members and compromise functioning of the electron transport chain in leukemic cells. J Immunol 2001; 166(5): 3130-42.
- [133] Perez-Galan P, Roue G, Villamor N, Campo E, Colomer D. The BH3-mimetic GX15-070 synergizes with bortezomib in mantle cell lymphoma by enhancing Noxa-mediated activation of Bak. *Blood* 2007; 109(10): 4441-9.
- [134] Goy A, Kahl B. Mantle cell lymphoma: The promise of new treatment options. *Crit Rev* Oncol Hematol 2010.
- [135] Rummel MJ, de Vos S, Hoelzer D, Koeffler HP, Hofmann WK. Altered apoptosis pathways in mantle cell lymphoma. *Leuk Lymphoma* 2004; 45(1): 49-54.

- [136] Paoluzzi L, Gonen M, Gardner JR, Mastrella J, Yang D, Holmlund J et al. Targeting Bcl-2 family members with the BH3 mimetic AT-101 markedly enhances the therapeutic effects of chemotherapeutic agents in in vitro and in vivo models of Bcell lymphoma. *Blood* 2008; 111(11): 5350-8.
- [137] Paoluzzi L, Scotto L, Marchi E, Seshan VE, O'Connor OA. The anti-histaminic cyproheptadine synergizes the antineoplastic activity of bortezomib in mantle cell lymphoma through its effects as a histone deacetylase inhibitor. *Br J Haematol* 2009; 146(6): 656-9.





### New Advances in Stem Cell Transplantation

Edited by Prof. Taner Demirer

ISBN 978-953-51-0013-3 Hard cover, 582 pages **Publisher** InTech **Published online** 24, February, 2012 **Published in print edition** February, 2012

This book documents the increased number of stem cell-related research, clinical applications, and views for the future. The book covers a wide range of issues in cell-based therapy and regenerative medicine, and includes clinical and preclinical chapters from the respected authors involved with stem cell studies and research from around the world. It complements and extends the basics of stem cell physiology, hematopoietic stem cells, issues related to clinical problems, tissue typing, cryopreservation, dendritic cells, mesenchymal cells, neuroscience, endovascular cells and other tissues. In addition, tissue engineering that employs novel methods with stem cells is explored. Clearly, the continued use of biomedical engineering will depend heavily on stem cells, and this book is well positioned to provide comprehensive coverage of these developments.

#### How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Yener Koc and Taner Demirer (2012). Mantle Cell Lymphoma: Decision Making for Transplant, New Advances in Stem Cell Transplantation, Prof. Taner Demirer (Ed.), ISBN: 978-953-51-0013-3, InTech, Available from: http://www.intechopen.com/books/new-advances-in-stem-cell-transplantation/mantle-cell-lymphoma-decision-making-for-transplant

# Open science | open minds

#### InTech Europe

University Campus STeP Ri Slavka Krautzeka 83/A 51000 Rijeka, Croatia Phone: +385 (51) 770 447 Fax: +385 (51) 686 166 www.intechopen.com

#### InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai No.65, Yan An Road (West), Shanghai, 200040, China 中国上海市延安西路65号上海国际贵都大饭店办公楼405单元 Phone: +86-21-62489820 Fax: +86-21-62489821 © 2012 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the <u>Creative Commons Attribution 3.0</u> <u>License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

# IntechOpen

# IntechOpen