the world's leading publisher of Open Access books Built by scientists, for scientists

4,800

Open access books available

122,000

International authors and editors

135M

Downloads

154

TOD 10/

Our authors are among the

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.

For more information visit www.intechopen.com



The Metamorphosis of Acute Renal Failure to Acute Kidney Injury

John W Pickering¹ and Zoltán H Endre^{1,2}

¹Christchurch Kidney Research Group, University of Otago Christchurch

²Department of Nephrology, Prince of Wales Hospital,

University of New South Wales, Sydney

¹New Zealand

²Australia

1. Introduction

Successive generations of scientists and nephrologists have failed to prevent or cure Acute Kidney Injury (AKI) and thousands every year die because of this. Recent innovations in proteomics and genomics have brought hope and renewed interest in preventing this blight. Motivated by high incidence and lack of effective treatments, researchers have focussed on how to detect AKI early in the disease process so as to provide the maximum opportunity for early intervention and positive outcomes. AKI Incidence is greatest in the intensive care, at about 11-52% in larger studies (n>500) (Ahlstrom et al. (2006); Bagshaw et al. (2008); Cruz et al. (2007)). Cardiac surgery and procedures involving radiocontrast pose smaller, but significant, risk of AKI with an incidence of 3-15% depending on cohort (Harjai et al. (2008); Lassnigg et al. (2008)). From 13 studies the mortality with AKI was 31.2% and was associated with an increase in relative risk of death from 2.40 to 6.15 depending on AKI severity (Ricci et al. (2008)). Stimulating much recent research has been the discovery of new kidney injury biomarkers, some of which appear to have sufficient sensitivity and specificity to be clinically useful.

This chapter will outline the history of the development of the concepts of clearance, acute renal failure, and acute kidney injury. This history provides the context for the current clearance based AKI diagnostic paradigm. The discovery of novel kidney injury biomarkers is challenging that paradigm. We will discuss the nature of that challenge and the opportunity it provides for development of early intervention treatments. All epidemiology, biomarker studies and clinical trials rely on tools to quantify AKI and assess efficacy of diagnostic or treatment efficacy. In section 3 we will discuss those tools before moving on to considering how they may best be applied in practice (section 4).

2. From ARF to AKI

2.1 Clearance and the rise and fall of creatinine

While suppression of urine flow, ischuria renalis, was recognised as a fundamental manifestation of renal disease from the 17th Century, clear metabolic manifestations of AKI

were not documented until World War I in the German and World War II in the English literature (see Eknoyan (2002); McGrath (1852)). The term "Acute Renal Failure" first appeared in the literature in 1946 (Frank et al. (1946)), although it has been attributed to Homer Smith (Eknoyan (2002)). In keeping with a recent change in nomenclature we shall use the term "Acute Kidney Injury" (AKI) unless we are specifically referring to an historic use of ARF.

The historical development of clearance techniques and the relationship to glomerular filtration rate (GFR) are discussed by Berliner in his tribute to the great renal physiologist Homer Smith (Berliner (1995)). The term clearance was introduce in 1928 with reference to urea and to clearance of a defined volume of plasma in unit time (Moller et al. (1928)). The idea of creatinine clearance as a measure of glomerular filtration rate (GFR) was beautifully first demonstrated by Rehberg in experiments on himself (Rehberg (1926)). These also highlighted how variations in serum creatinine could be induced by diet: Rehberg ingested different quantities of creatinine to vary his serum creatinine. The utility of clearance as a technique was further established in the laboratory of Homer Smith, especially with para-amminohippuric acid clearance, a measure of secretion and renal blood flow (Smith et al. (1945)).

Creatinine is formed non-enzymatically from creatine in muscle, has a molecular weight of 113.12 Da, is freely filtered at the glomerulus and completely cleared by renal excretion when renal function is normal. The proximal tubules secrete creatinine, which accounts for 10 to 20% of the excreted load, and results in overestimation of GFR when measured by creatinine clearance (Perrone et al. (1992); Shemesh et al. (1985)). The contribution of tubular creatinine secretion to clearance, is increased when GFR is reduced and may reach 50%, but is highly variable amongst individuals (Perrone et al. (1992)). In contrast, the tubules reabsorb creatinine in some clinical settings such as decompensated heart failure and uncontrolled diabetes (Levinsky & Berliner (1959); Perrone et al. (1992)).

With creatinine clearance firmly established as a reasonable approximation to GFR, the next step was to estimate creatinine clearance based on the reciprocal relationship with plasma creatinine. This was popularized by the Cockroft-Gault equation which was derived by, firstly, a regression to estimate creatinine excretion/kg body weight according to age in hospitalised male patients; then clearance was calculated by multiplying by weight and dividing by the serum creatinine (Cockcroft & Gault (1976)). An untested 15% reduction for female gender was included, based on the observation that, on average, females had less fat and muscle mass than males (Cockcroft & Gault (1976)). This formula has been widely replaced by other estimates of GFR (eGFR), most notably by the Modification of Diet in Renal Disease (MDRD) equation originally developed in a population of CKD patients (Levey et al. (1999)).

Many alternative algorithms for creatinine-based eGFR have been developed, including those regularly in use for children; these equations are more accurate and precise than estimates from measurement of creatinine alone (*KDOQI Clinical Practice Guidelines for Chronic Kidney Disease: Evaluation, Classification, and Stratification* (2002)). The various iterations of the MDRD equation rely on plasma creatinine plus several other variables, including gender and race, and originally albumin and urea (as blood urea nitrogen) concentrations, but excluding mass although they do incorporate body surface area via the units; the latest version is more accurate than the MDRD equation for patients with GFR>60 ml/min (Levey et al. (2009)) but caveats remain (Rule (2010)) and drug dosing tends to be based on Cockroft-Gault (Ryzner (2010)).

Classical measurement of clearance with timed urine collection is cumbersome, so logically, the surrogate of clearance, the serum creatinine (actually measured in plasma), is usually the sole measure used to define renal status including development of AKI. However, serum creatinine increases slowly in response to a single step alteration in GFR. Serum creatinine has a half-life of approximately 4 hours when GFR is normal and 77 hours when GFR is reduced to 5% (Chiou & Hsu (1975); see section 3.1). As 3 to 5 half-lives are required after any change in GFR to obtain a new steady-state estimate, a reliable GFR based on the serum creatinine will require at least 12 hrs after even a minimal change in GFR. The current consensus definition (RIFLE: Risk, Injury, Failure, Loss, End-stage) of AKI requires at least a 33% decline in GFR, but a 25% reduction is traditionally accepted for diagnosis of contrast-induced AKI) (Bagshaw et al. (2008); Endre et al. (1989); Pickering & Endre (2009a)). Thus a new steady-state creatinine-based estimate of GFR will take from 24 to 72 hours. Obviously, the extent of increase in creatinine, which is determined by the extent of initial decrease in GFR and creatinine production, may allow diagnosis of AKI prior to the time needed to reach steady-state. However, the limited precision of creatinine measurement and the extent of intra-patient variation mean that a minimum 10% change in creatinine has traditionally been required by clinicians to demonstrate measurably significant change. An increase in serum creatinine has also been used as a major trigger for intervention (renal replacement therapy, RRT) in both AKI and chronic kidney disease (Gibney et al. (2008)).

The recognised imprecision in determining change in GFR led to removal of GFR from the new Acute Kidney Injury Network (AKIN) consensus classification of AKI so that an absolute or percentage increase in creatinine alone or in combination with oliguria has become the new consensus definition of AKI (Mehta et al. (2007)). Thus, despite many limitations, such as dependence on muscle mass, diet etc (Perrone et al. (1992)), serum creatinine became the accepted shorthand for estimating GFR and significant change in creatinine became the definition of AKI and marker of AKI severity.

Since there is an inverse relationship between serum creatinine and GFR, it is easy to forget that a small increase in serum creatinine represents a substantial decline in GFR. Consequently, prior to the now widespread reporting of eGFR estimated from creatinine, clinicians often ignored small increases in creatinine, and characterised these as "mild" or "moderate" increases. Combined with the uncertainty associated with the precision of measurement, years of ignoring such small increases in creatinine have impaired insight into how AKI is triggered. Even if an increase is observed, the delay required for diagnosis creates uncertainty regarding the timing of the renal insult leading to AKI. This represents a lost opportunity to investigate and identify the underlying pathophysiology of AKI in humans. Even mild grades of severity of AKI and transient (less than 24 hour increases in creatinine) are associated with increased hospital mortality (Chertow et al. (2005); Uchino et al. (2010; 2006)).

Inevitably, much of our interpretation of the pathophysiology of AKI is based on an animal model; the classical model utilises temporary cessation of renal blood flow to induce injury, usually through bilateral renal artery clamping. While this model may parallel human AKI after aortic surgery or renal transplantation, it greatly exaggerates the degree of tubular injury compared with that in the limited number of available human renal biopsies and is probably less relevant to AKI that does not follow hypoperfusion (Heyman et al. (2010)). Even in this model there is limited understanding of how major pathophysiological events in AKI are integrated, for example, the mechanism and timing of the switching "off" and "on" of

glomerular filtration. With few exceptions (Alejandro et al. (1995); Myers et al. (1984)), most of our experimental interventions have been validated only in this and nephrotoxic animal models (Vaidya et al. (2010)).

While there is a great deal of information about a large number of cellular (tubular and endothelial) events and the autonomic, inflammatory and renal vascular responses in experimental ischemia-reperfusion injury (Devarajan (2006); Heemskerk et al. (2009)), there is little corroborative and time-relevant clinical pathophysiological data. Usually, the clinical diagnosis of ischemic AKI is a diagnosis of exclusion. Thus, the potential delay imposed by reliance on creatinine is further delayed by investigations (eg exclusion of urinary outflow obstruction) or interventions (fluid loading to treat underlying "pre-renal" AKI) designed to exclude rather than confirm the diagnosis of ischemic AKI. Other investigations such as measurement of global or parenchymal renal blood flow, or renal biopsy, which might provide insight into human AKI, are delayed, difficult to interpret in the absence of baseline data, and usually not performed.

Since diagnosis is delayed it is not surprising, that there has been failure of pharmacological intervention in clinical trials, which are largely based on experimental interventions to prevent rather than treat AKI (Jo et al. (2007)). However, pharmacologic prevention of AKI has also largely failed, even when the apparent aetiology is known, eg parenteral administration of iodinated radiocontrast (Fishbane (2008); Nigwekar et al. (2009); Zoungas et al. (2009)). The failure to translate apparently effective pharmacologic preventive measures in animal models into clinical practice, suggests that the serum creatinine-inspired delay in diagnosis, merely complements a lack of fundamental understanding of pathophysiology in human AKI.

2.2 The rise and rise of injury

The change in nomenclature from acute renal failure to acute kidney injury recognised that an acute decline in renal function is usually secondary to injury (*American Society of Nephrology Renal Research Report* (2005); Mehta et al. (2007)). The need for a renal-specific biomarkers of injury akin to a troponin was presaged in the late 1990s (Star (1998)) and the discovery of such proteins was later accorded highest priority by The American Society of Nephrology (*American Society of Nephrology Renal Research Report* (2005)) with the expectation that such biomarkers would: "Diagnose AKI before the rise in serum creatinine; Stratify patients with respect to severity of injury and; Provide prognostic indicators." The term "secondary prevention" was introduced to highlight that biomarker detection would lead to early intervention ideally prior to loss of GFR (Pickering & Endre (2009c)).

Some urinary proteins, notably α_1 -microglobulin, β_1 -microglobulin, and N-acetyl- β -D-glucosaminidase (NAG), were already known to be associated with acute tubular injury (Yu et al. (1983)), whilst others awaited discovery. In 1998 Kidney Injury Molecule-1 (KIM-1) was identified as being upregulated in the proximal tubule cells after ischemic/reperfusion (I/R) injury (Ichimura et al. (1998)) and soon discovered in the urine of patients with I/R injury (Han et al. (2002)). Around the same time a small study of ICU patients identified urinary tubular injury makers, α and α -glutathone S-transferase (α and α -GST), and the bursh border enzymes, α -glutamyl-transpeptidase (GGT) and alkaline phosphatase (AP), as diagnostic of AKI (Westhuyzen et al. (2003)). A transcriptome wide interrogation study to identify genes induced early after I/R injury identified neutrophil gelatinase-associated lipocalin (NGAL) to be upregulated in a mouse model (Mishra et al. (2003)). Plasma and urinary NGAL quickly showed spectacular success as diagnostic

markers. The first, a trial of 71 children undergoing cardiopulmonary bypass of whom 20 developed AKI showed urinary and plasma NGAL to increase 10-fold in AKI 2-h post surgery (Mishra et al. (2005)). Similarly, interluekin-18, IL-18, first identified as being released into the urine following I/R injury in mice (Melnikov et al. (2001)) was found to appear in large quantities in the urine of patients with acute tubular necrosis (Parikh et al. (2004)). Proteomic and genomic approaches continue to be a rich source of new urinary proteins which may predict AKI (Bennett & Devarajan (2011); Devarajan (2008)).

AKI Biomarker discovery has followed the well trodden path of "early promise" with some highly sensitive and specific biomarkers in demographically homogeneous populations (eg paediatric cardiopulmonary bypass surgery), followed by a more tempered response as studies evaluated candidate biomarkers in demographically heterogeneous populations with multiple causes of AKI and co-morbidities. There is no one biomarker which will successfully diagnose or predict a decline in renal function in all situations. Three factors have emerged which are likely to become determinant factors in the choice of biomarker for a particular clinical context, namely: (i) likely aetiology of AKI, (ii) pre-existing renal function, (iii) time from renal insult. IL-18 is an example of a biomarker which has been shown to be elevated following ischaemic/repurfusion injury (Hall et al. (2010); Parikh et al. (2006)), but possibly not following radiocontrast induced nephrotoxic injury (Bulent Gul et al. (2008)). Mcilroy et al demonstrated that in a cohort of 426 adult cardiac surgery patients urinary NGAL concentrations post-operatively did not differ between those who developed and those who did not develop AKI when their estimated baseline GFR (eGFR) was less than 60 ml/min, yet for those with a normal baseline eGFR (90-120 ml/min) NGAL was significantly elevated in the AKI cohort (Mcilroy et al. (2010)). In our own head to head comparison of 6 urinary biomarkers (GGT, AP, NGAL, Cystatin C, IL-18, and KIM-1) in 529 adult patients on entry to an intensive care unit, we demonstrated that the performance of biomarkers is critically dependent on both baseline renal function and time from renal insult (Endre et al. (2011)). Peak diagnostic performance at a level that may be considered clinically useful was limited to patients with eGFR 90-120 ml/min within 12-h of insult for GGT, 6-h for NGAL and from 6 to 12-h for Cystatin C, IL-18 and KIM-1, and to patients with eGFR < 60 ml/min from 12 to 36-h for GGT, Cystatin C, NGAL and IL-18.

There are many excellent recent reviews of biomarkers of AKI. The pathophysiology of AKI in relation to potential biomarkers has been reviewed and discussed specifically in relation to AKI following cardiopulmonary bypass (Haase et al. (2010)), AKI with varying aetiology (Vaidya et al. (2008)), and AKI involving biomarker mediators of inflammation (Akcay et al. (2009)). Reviews of biomarker performance specific to nephrotoxic injury (Bonventre et al. (2010); Ferguson et al. (2008)), septic AKI (Bagshaw et al. (2007)), ischemic injury following cardiopulmonary bypass (Haase et al. (2010)), acute allograft rejection and ischemic injury (Alachkar et al. (2010)) as well as broader reviews across aetiologies (Coca et al. (2008); Edelstein & Faubel (2010); Endre & Westhuyzen (2008); Malyszko (2010)) have been published within the last 4 years. We have published a more specialist review considering biomarkers in the early phase of injury (Pickering & Endre (2009c)) and there has been one meta-analysis of the performance of NGAL (Haase et al. (2009)).

2.3 Paradigm lost and paradigm found

The relationship between clearance (the function paradigm) and injury has, to date, been studied in humans primarily in sample populations of about 20 to 600 in which a biomarker's

ability to diagnose a subsequent rise in a surrogate marker of function (normally creatinine) is assessed. However, we need to explore the early stages of, and prior to, loss of renal function in order to understand the relevant pathophysiology of clinical AKI. We can conceptualise this as an evaluation of the evolution of injury phase which leads to loss of GFR and the early loss of function phase immediately following a GFR decrease (Figure 1). Detection and characterisation of this very early phase in man appears essential for progress. This highlights the need for quantifying the time course of injury biomarkers in relation to change in GFR and for real-time assessment of renal function. Two promising techniques are under development. The ambulatory renal monitor (ARM) is a shielded detector which monitors extracellular excretion of 99m TcDTPA over up to 24 hours following injection and relates this to GFR (Rabito et al. (2010)). The ratiometric fluorescence approach monitors the plasma disappearance of both a rapidly filtered and a poorly filtered fluorescent marker introduced by a single bolus infusion, changes in the ratio of which enable a calculation of GFR (Wang et al. (2010)). Both techniques allow a rapid (5 to 15 min) measure of GFR. Ideally these or similar techniques would allow monitoring of kidney function from prior to the time of insult through the evolution time until change of GFR and beyond.

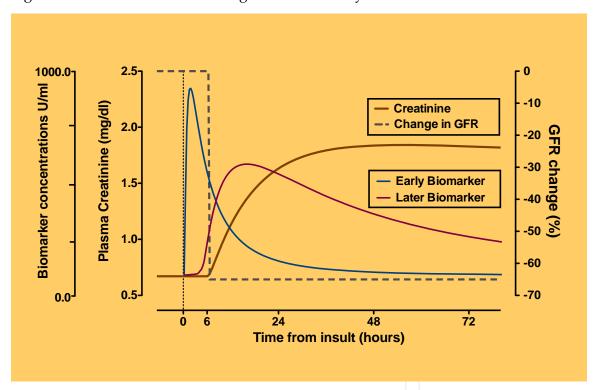


Fig. 1. The paradigm of function compared with the paradigm of injury. Following injury the early injury biomarker is elevated prior to change in function (here, a 65% loss of GFR, approximately 6 hours post-injury). The later biomarker takes takes longer to be elevated. Creatinine is only elevated following loss of GFR (following equation 6)

Much of the literature describes a biomarker as "predicting" AKI when it precedes an increase in plasma creatinine. However, a biomarker is only truly predictive if it precedes a decrease in GFR. If it is elevated shortly after such a decrease, then it should be described as *diagnostic* rather than predictive of AKI even though it may predict the latter increase in plasma creatinine.

3. Defining AKIs

In its broadest sense renal failure is simply the rapid loss of renal filtration (ie decrease in GFR). As discussed, current clinical diagnosis of AKI and evaluation of novel biomarkers of kidney injury are largely dependent on observation of changes in creatinine as a surrogate for a change in GFR. An understanding of the kinetics of the relationship between creatinine and GFR provides a basis for understanding the definitions of AKI and their limitations. Plasma cystatin C, an alternative to creatinine (Nejat et al. (2010); Westhuyzen (2006)), follows similar kinetics.

3.1 Creatinine and cystatin C kinetics

Creatinine and cystatin C are generated in tissue outside of the plasma compartment, diffuse into the plasma compartment from where they are lost by renal and non-renal excretion. Because both creatinine and cystatin C are not bound to plasma proteins the exchange between the extravascular and plasma compartments is rapid (compared with rates of production and elimination) allowing us to conflate the two compartments into one compartment with volume of distribution, *V* (Figure 2).

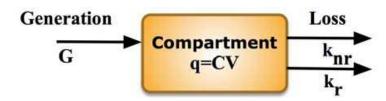


Fig. 2. One compartment pharmacokinetics model

The change in total mass (q) of creatinine/cystatin C depends on the rate at which it is entering and the rate at which it is leaving the compartment:

$$\frac{dq}{dt} = \text{gain from generation} - (\text{renal loss} + \text{non-renal loss})$$
 (1)

Under normal circumstances non-renal losses are much less than renal losses and may be ignored. The renal loss is the product of the renal elimination rate constant, k_r , and the total mass, q. As the total mass is the product of the concentration C and volume of distribution V, equation 1 becomes:

$$C\frac{dV}{dt} + \frac{dC}{dt}V = G - k_r CV \tag{2}$$

where G is the generation rate of creatinine/cystatin C (see Box 3.1). The volume of distribution of creatinine is equal to the total body water (TBW) and that of cystatin C to the extracellular fluid (about one third of TBW)(see Box 3.2). If this is assumed not to change then equation 2 becomes:

$$\frac{dC}{dt} = \frac{G}{V} - k_r C \tag{3}$$

At equilibrium (dC/dt = 0 at t = 0), prior to any change in GFR the renal elimination rate constant may be determined from equation 3:

$$k_{r0} = \frac{G}{C_h V} \tag{4}$$

where C_b is the baseline concentration (at time t=0). k_{r0} may also be estimated from the renal clearance (Cl) which is simply the product k_{r0} and V. For creatinine, this may be measured in critically ill paitients using a short duration creatinine clearance and an estimate of an individual's volume of distribution (see below). From the EARLYARF study (Endre et al. (2010)) where 4-h creatinine clearances were measured on entry to ICU in 484 patients we were able to calculate a median (interquartile range) for k_{r0} of 0.11 (0.07-0.17) h⁻¹. Following a loss in GFR (Δg %), the renal elimination rate constant becomes:

$$k_r = (1 - \frac{\Delta g}{100})k_{r0} \tag{5}$$

and equation 3 may be solved numerically, or, if we assume of G, V, or k_r are not varying, may be integrated to give the concentration as a function of time (Chiou & Hsu (1975); Chow (1985)):

$$C(t) = \frac{G}{k_r V} (1 - e^{-k_r t}) + C_b e^{-k_r t}$$
(6)

As $t \to \infty$ the concentration asymptotically approaches a new steady state, C_{ss} which from equation 6 is:

$$C_{ss} = \frac{G}{k_r V} \tag{7}$$

Mathematically the concentration is within 5% of the new steady state in 4.4 half lives (Half life: $t_{1/2} = ln(2)/k_r$). In practice this is well within the uncertainty in creatinine measurements. The new steady state may also be determined by substituting equations 5 and 4 into equation 7:

$$C_{ss} = \frac{C_b}{\left(1 - \frac{\Delta g}{100}\right)} \tag{8}$$

From this equation we may determine the equivalences between a decline in GFR and a rise in creatinine used in the RIFLE definition of AKI (Table 1).

Box 3.1. Creatinine and Cystatin C production rates

Creatinine production (Bjornsson (1979)):

Male (
$$r^2 = 0.919$$
): $G_0 = (27 - 0.173 \times age) \times weight/24$ (mg/h) Female ($r^2 = 0.966$): $G_0 = (25 - 0.175 \times age) \times weight/24$ (mg/h)

Creatinine production may decrease during critical illness (Griffiths (1996)). If the reduction is at constant rate m%, (eg 2% per day as suggested by Griffiths) then:

$$G(t) = G_0 e^{-mt}$$
 and $\frac{dG}{dt} = -mG$

Cystatin C production

Only one study has measured the rate constant of Cystatin C (Sjostrom et al. (2005)). There was no significant difference with age, sex or lean body mass. The rate constant per $1.73m^2$ of body surface area was:

$$G_{CysC} = 7.44 \quad (mg/h/1.73m^2)$$

Box 3.2. Estimating the volume of distribution

The volume of distribution of creatinine is equal to the total body water (TBW) and of cystatin C to the volume of the extracellular fluid which is about $1/3^{rd}$ of the TBW (Hansen (2002)). TBW is often estimated from the body weight as:

$$TBW = 0.6 \times Body \ weight \ (L)$$

More accurate formulae have been derived from population studies (Watson et al. (1980)):

```
Male (r^2 = 0.704): TBW = 2.447 - 0.09516 \times age + 0.1074 \times height + 0.3662 \times weight (L) Female (r^2 = 0.736): TBW = 2.097 + 0.1069 \times height + 0.2466 \times weight (L)
```

where *height* is measured in cm, *weight* in kg, and *age* in years.

Where height is not available, the slightly less robust equations may be used:

```
Male (r^2 = 0.689): TBW = 20.03 - 0.1183 \times age + 0.3626 \times weight (L) Female (r^2 = 0.717): TBW = 14.46 + 0.2549 \times weight (L)
```

3.2 Categorical consensus

In 2004 the Acute Dialysis Quality Initiative Group (ADQI) developed a consensus definition for AKI and severity staging (RIFLE: Risk, Injury, Failure, Loss, End Stage) (Bellomo et al. (2004)). The scheme involved both a creatinine based classification and a urine output based classification (Table 1). A decrease in GFR of more than 25% or increase in serum creatinine of 50% was deemed sufficient to diagnose AKI. Unfortunately, there was an error in calculating this relationship which was not identified until 2009 (Pickering & Endre (2009a)). A 50% increase in creatinine is equivalent to a one third decrease in GFR (see equation 8) not a 25% decrease. Similarly, for RIFLE stage F a 200% increase in creatinine is equivalent to a two thirds decrease in GFR not a 75% decrease. Whilst plasma creatinine rather than a GFR measure, or an estimation with creatinine clearance, is the analyte of choice in AKI studies, creatinine is but a surrogate for GFR and GFR should remain the principal diagnostic parameter of AKI (Pickering & Endre (2009b)). The ADQI group recommended AKI be defined as "sustained" (lasting at least 24hrs) and "abrupt" (1-7 days) (http://www.ccm.upmc.edu/adqi/ADQI2/ADQI2g1.pdf). Whilst duration did not appear in the seminal RIFLE publication it was included in later publications (Hoste et al. (2006); Lameire et al. (2006)).

On the back of new evidence that even minor changes in serum creatinine are associated with poor outcomes (eg Chertow et al. (2005); Lassnigg et al. (2004)), the Acute Kidney Injury Network modified the RIFLE definition to include a small absolute rise in creatinine (0.3 mg/dl or 26.4 μ mol/l). Further modifications included requiring the change to occur within 48 hours for the definition of AKI, and removing RIFLE stages L and E in preference to all patients requiring renal replacement therapy to be assigned to severity stage III (Table 1). Both definitions have received broad support and there is considerable evidence for an association of increased mortality with increased RIFLE or AKIN stages (Bagshaw et al. (2008); Ricci et al. (2008)). More recently, duration of AKI (using the AKIN criteria) has been shown to be independently associated with long-term mortality (Brown et al. (2010); Coca et al. (2010); Goldberg et al. (2009)). KDIGO (Kidney Disease for Improving Global Outcomes:

www.kdigo.org) has recently reviewed the use of the AKIN and RIFLE criteria and is shortly
to release a new consensus definition which combines the two definitions.

RIFLE			AKIN		RIFLE and AKIN	
Stage	Creatinine increase	GFR decrease	Stage	Creatinine increase	Urine output	
Risk (R)	≥ 50%	> 33.3%*	I	≥ 0.3 mg/dl or ≥ 50%	< 0.5 mg/kg/h for 6 h	
Injury (I)	≥ 100%	> 50%	II	≥ 100%	< 0.5 mg/kg/h for 12 h	
Failure (F)	\geqslant 200% or \geqslant 0.5 mg/dl and above 4.0 mg/dl	> 66.7%**	III	\geqslant 200% or \geqslant 0.5 mg/dl and above 4.0 mg/dl	< 0.3 mg/kg/h for 24 h or anuria for 12 h	

^{*,**} corrected from 25% and 75% see Pickering & Endre (2009a)

Table 1. Severity Staging: Consensus definitions

3.3 A continuum needing continuous variables

One of the intentions of ADQI in setting up RIFLE was that it would provide common outcomes in clinical trials. During the four year period (2005-08) only 36% of published AKI (non Contrast Induced Nephropathy, CIN) trials used RIFLE or AKIN as an outcome variable, and the use was not consistent in terms of timing and duration of injury (Endre & Pickering (2010)). Amongst CIN intervention trials only 13% used RIFLE or AKIN, whereas most continued to use an increase in creatinine of \geq 25% and/or \geq 0.5 mg/dl (44.2 μ mol/l) to diagnose CIN. This later definition is slightly anomalous as only those with pre-existing kidney disease (creatinine > 2.0 mg/dl) can have a greater than 0.5 mg/dl elevation in creatinine that is less than 25%. We have recommended that all CIN studies adopt the AKIN or RIFLE definition of AKI (Endre & Pickering (2010)).

We investigated whether a continuous variable measure of kidney function, the Relative Average Creatinine (RAVC), performed better than the RIFLE and AKIN categorical definitions as an outcome variable in AKI prevention or intervention trials. The RAVC is the integral of the area under the plasma creatinine curve above baseline creatine divided by the total time and baseline creatinine:

$$RAVC(\%) = \frac{100}{C_h t} \int_0^t (C(t) - C_b) dt$$
 (9)

which in practice is calculated using the trapezoidal rule (Figure 3):

$$RAVC(\%) = \frac{100}{C_b(t_N - t_1)} \sum_{0 \le n \le N - 1} \frac{(C_{n+1} - C_b) + (C_n - C_b)}{2} (t_{n+1} - t_n)$$
(10)

where N is the number of creatinine measurements.

We created a population of 10,000 Virtual-In-Patients (VIPs) whose baseline creatinine and changes in GFR were based on real ICU populations. Placebo controlled trials were simulated by randomly assigning half the VIPs to treatments which ameliorated loss of renal

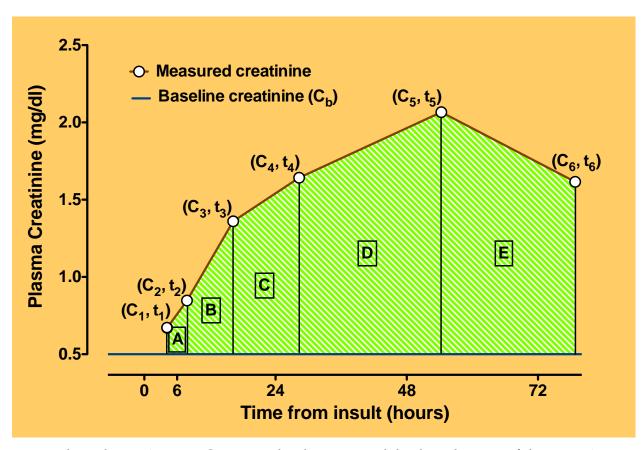


Fig. 3. The Relative Average Creatinine by the trapezoidal rule is the sum of the areas A to E divided by $(t_6 - t_1)$ (following equation 10).

function (ie reduced the reduction in GFR). The more efficacious the treatment, the less the decrease in GFR. Creatinine profiles were calculated using equation 6 (Pickering et al. (2009)). AKIN, RIFLE and RAVC as outcome variables were compared. At low treatment efficacy, the categorical outcomes underestimated and at high treatment efficacy overestimated the effect of treatment. These effects were exaggerated when the population contained a high proportion of patients with more severe AKI. The RAVC, on the other hand, responded in an almost linear fashion across treatment efficacies. Importantly, when the efficacy was low it was best able to distinguish between placebo and treatment arms. The advantage of the RAVC over the categorial metrics is two fold, first it includes the effect of treatment on those patients who had mild kidney injury which in normal circumstances would not exceed the diagnostic threshold for AKI according to a categorical definition, but which may result in a small increase (eg 20-30%) in creatinine and, second, it measures function over a (pre-determined) time period. As discussed previously, the length of time creatinine is elevated is independently of the maximum elevation associated with mortality. The RAVC captures both severity and duration of injury and was used as the primary outcome in the EARLYARF trial, which was the first randomised control trial to use an injury biomarker to triage patients to placebo or high-dose erythropoietin (Endre et al. (2010)).

3.4 The baseline issue

All creatinine based definitions of AKI and the RAVC depend on knowing the normal, or "baseline", plasma creatinine for each individual. For patients undergoing elective surgery

this is easily obtainable prior to surgery. However, about half of the patients entering the ICU have no previous record of plasma creatinine to serve as a baseline. For trials this requires a retrospective determination of renal function. The ADQI recommend assuming a normal (eg 75 ml/min) GFR for all these patients and "back-calculating" a plasma creatinine using the MDRD equation. Unfortunately, this has proved erroneous. We showed that in our VIP population and in an ICU population that this approach seriously overestimates the proportion of patients with AKI using either the AKIN or RIFLE definitions (Pickering & Endre (2010); Pickering et al. (2009)). Randomly assigning a baseline creatinine produced just as accurate results as back-calculation. In a population already with AKI the presence of patients with CKD was seen to be driving the overestimation (Bagshaw et al. (2009)). Using the emergency department creatinine as an alternative baseline underestimated AKI (AKIN) and lowered the sensitivity (Siew et al. (2010)).

In the EARLYARF trial we overcame this problem by using an adjudicated hierarchical approach to choose in each patient the measured plasma creatinine that best represented normal renal function. Outpatient plasma creatinine prior to admission were considered the most likely to represent true baseline function. Whilst CKD can be diagnosed over three months, up to twelve months appears to be a reasonable time period prior to admission in which to ascertain baseline creatinine as it reduces misclassification of AKI (Lafrance & Miller (2010)). Amongst patients with no pre-admission creatinine measurement, a post-discharge measurement is the next best option if it is stable. Increasing creatinine may be indicative of developing CKD. Hence, it is preferable that a post-discharge creatinine is within three months of the insult. As a last resort the lower of the first hospital or final hospital (when there is recovery) may be used. We have presented our recommended hierarchical approach in table 2. This differs from the earlier approach in that we now consider that for cardiac arrest and trauma patients where there is no baseline prior to admission available the first hospital sample is likely to be the best estimate of baseline function if it is measured close to the time of renal insult. In our experience this is less than 2-h for most cardiac arrest and trauma patients. For the creatinine to have increased considerably in this time frame the loss of GFR would have had to have been substantial.

3.5 Quantifying function in a dilute environment

Fluid resuscitation dilutes plasma creatinine concentrations, which in turn may lead to delayed diagnosis or severity underestimation. This may explain why the only successful AKI intervention in the ICU has been early consultation with a nephrologist (Mehta et al. (2002)). The effect of fluids may be estimated by adjusting plasma creatinine for fluid balance (Macedo et al. (2010)):

$$C_{adjusted} = C_{measured} \times \frac{\text{admission weight (kg)} \times 0.6 + \sum \text{daily cumulative fluid balance (L)}}{\text{admission weight (kg)} \times 0.6}$$
(11)

3.6 Injury meets function

Current evaluation of novel biomarkers of renal injury is largely confined to evaluation of their performance to predict increases in plasma creatinine which lead to a diagnosis of AKI according to RIFLE or AKIN. This runs the risk of missing significant injury because creatinine did not increase beyond the diagnostic threshold of 50% or 0.3 mg/dl. In a large series

Situation	Timing of baseline creatinine sample				
1 Elective surgery	Prior to surgery				
2 All	Pre-hospital outpatient or prior admission				
	Preferably this is between 7 and 90 days prior to admission. 7 days avoids a period which may reflect changing renal function. Less than 90 days is preferable because 90 days is the period usually used to diagnose CKD. Up to 365 days may be used if there is little likelihood of CKD having developed during that period.				
3 Cardiac Arrest	First hospital				
	If a pre-hospital value is not available and the time between insult and the first measurement is short (< 2h). Within a short time frame the creatinine will not (yet) have become elevated.				
4 Trauma	First hospital				
	If a pre-hospital value is not available and the time between insult and the first measurement is short (< 2h) and crush-injuries are not involved.				
5	Post ICU discharge				
	Within 90 days of admission to avoid capturing the development of CKD.				
6	Lowest of first hospital or final ICU				

Table 2. Hierarchical determination of baseline creatinine

of experimental studies of nephrotoxic AKI biomarkers, serum creatinine and blood urea nitrogen (BUN) were the poorest predictors of histologically determined injury compared with numerous urinary biomarkers (Dieterle et al. (2010); Ozer et al. (2010); Vaidya et al. (2010); Yu et al. (2010)). In what may turn out to be a seminal publication Haase et al. (2011) demonstrated that across 10 studies patients with a positive NGAL, yet negative plasma creatinine, for AKI had worse outcomes (mortality, need for dialysis, and ICU length of stay) than those with both negative NGAL and negative creatinine.

Biomarker clinical utility is most often quantified by the area under the receiver operator characteristic curve (AUC or c-statistic, see Box 3.3). The AUC is crude estimation of the ability of the biomarker to distinguish between those with and without AKI. However, a high AUC does not necessarily imply clinical utility. Clinical utility depends on what alternative biomarkers there are, what treatments are available, and the risks and costs involved with false negatives or false positives. The calculation of the sensitivity, specificity and particularly the negative and positive predictive values at either an established cut-off from the literature, or a cut-off chosen for clinical reasons, or derived mathematically from the ROC. The latter is usually either the cut-off closest to a Sensitivity and Specificity of 1 or the Youden index.

Comparing AUCs between different studies is problematical. AUCs for AKI are highly dependent on the AKI definition. Typically a definition requiring greater injury (eg RIFLE sustained for 24-h, compared with an increase of 0.3 mg/dl within 48-h) will result in higher AUCs. Within a study when more than one biomarker is being measured they should be compared using the method of DeLong (DeLong et al. (1988)). One biomarker should not be described as "better" than another unless the difference between the two is statistically significant at p < 0.05.

Box 3.3. The Area Under the Curve (AUC)

The receiver operator characteristic curve (ROC) is a plot of sensitivity verse 1-specificity. An AUC of 1 means the biomarker always discriminates between patients with and without the disease (no false negatives and no false positives). An AUC of 0.5 is equivalent to a coin toss. Whilst an AUC of less than 0.5 means the reciprocal of the biomarker is diagnostic. Some statistics packages, however, will always express the AUC as greater than 0.5 by inverting the biomarker concentration where necessary. The AUC from a study is strictly speaking an estimate of the AUC of the population, therefore it should always be presented with appropriate confidence intervals (usually 95%). The 95% confidence interval is ± 1.96 times the standard error of a proportion:

$$\widehat{AUC} - 1.96\sqrt{\widehat{AUC}(1 - \widehat{AUC})/n}$$
 to $\widehat{AUC} + 1.96\sqrt{\widehat{AUC}(1 - \widehat{AUC})/n}$

where n is the sample size and \widehat{AUC} the estimated AUC. This is equation is adequate for sample sizes > 30, but for smaller sample sizes a bootstrapping should be used. Only an AUC for which the lower limit of its 95% confidence interval is greater than 0.5 may be described as diagnostic (or prognistic).

Figure 4 is an example of a ROC from the EARLYARF trial (Endre et al. (2011)). It shows the ability of NGAL to diagnose AKI when the sample was taken between 12 and 36 hours following renal insult. The AUC lower limit of the 95% confidence interval, 0.62, is greater than 0.5, therefore it is diagnostic.

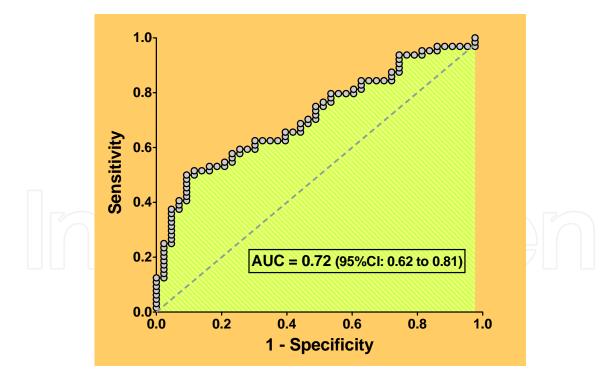


Fig. 4. An example of a receiver operator characteristic curve

It is anticipated that panels of biomarkers will be needed to diagnose AKI in heterogeneous populations with multiple AKI aetiologies. Attempts to assess a panel of biomarkers usually involve logistic regression models with two or more biomarkers measured at the same time point. The EARLYARF trial has demonstrated that the efficacy of any one particular biomarker is very much time dependent, and that each biomarker has its own "window of opportunity" during which it has have diagnostic utility (Endre et al. (2011)). Logistic regression models are not a suitable approach for assessment of biomarker panels because of this. Until we know the time courses of biomarkers much better, an "either and/or" approach is likely to yield greater results. For example, in an ICU population either an elevated GGT and/or an elevated NGAL may be considered diagnostic.

There is a need to move away from assessing injury biomarkers only in relation to function. All trials should report mortality data, even if the incidence is too low for statistical analysis. This will facilitate later meta-analysis of the relationship between a biomarker and mortality. The EARLYARF trial paved the way for future trials of early intervention based on an elevated biomarker. Since the inception of that trial new biomarkers with rapid assay turn-around (necessary in an early intervention trial) have become available. Plasma and urinary NGAL, and KIM-1 head the list along with urinary Cystatin C and GGT which are already routinely available in many hospital laboratories. It is anticipated that the next early intervention trial will use one or more of these biomarkers.

3.7 Quantifying injury in a dilute environment

Changes in GFR and water handling will change urinary biomarker concentrations independent of injury. Normalising biomarkers to urinary creatinine has been proposed to account for these effects. This process also amplifies the signal soon after a decline in function (Waikar et al. (2010)). This may be advantageous in an early intervention trial if the threshold for intervention is set high enough, but it may distort the analysis of biomarker performance in a biomarker performance study. Whilst there is no consensus on whether biomarkers should or should not be normalised to urinary creatinine we recommend reporting both normalised and non-normalised results.

4. Practical considerations

Table 3 presents a summary of practical measures to take into account when planning AKI epidemiology, biomarker efficacy studies or prevention or intervention trials.

4.1 Epidemiology

Most epidemiological studies are retrospective and face the difficulty of missing data, particularly baseline creatinine data. This was discussed in section 3.4. It is important to quantify the severity of AKI, as more severe AKI is likely to have greater long term impact on health resources. Where possible, data on the duration of AKI as well as severity should be captured as this is an independent predictor of outcome. Three areas of epidemiology lack data. First, there are comparatively few good epidemiology studies in countries other than in Europe, North America or Australasia (Cerda et al. (2008)). The incidence of AKI in countries with large populations such as China, India, Indonesia, Nigeria has significance beyond their own borders. Some countries have numbers of particular AKI aetiologies which, if well studied, could provide useful data world-wide. For example, with anti-retroviral therapy

reducing mortality in HIV patients, the incidence of AKI in this population may be increasing (Lopes et al. (2011)). In South-East Asia, AKI is a prominent complication of paraquat (a contact herbicide) self-poisoning (Roberts et al. (2011)). In both these cases epidemiology in countries with relevant populations would help identify the extent of risk in other countries where incidence of HIV or paraquat poisoning is relatively low. Second, there are few studies that have investigated AKI on CKD (Ali et al. (2007)). The growing world wide epidemic of CKD demands epidemiology that quantifies the additional risk CKD has for AKI and the effects of AKI on the progression of CKD. Third, there are few studies on CKD induced by AKI that have comparable control groups of hospitalised non-AKI patients (Coca et al. (2009)).

4.2 Biomarker studies

It is important to report the timing of sampling for biomarker measurements in relation to putative onset of injury. As we have seen, different biomarkers are likely to have different time profiles. A weakness of many studies is that samples are taken only at one or two time points, and even then they may be outside the temporal "window of opportunity" of the biomarker, possibly rendering the study results misleading. Temporal profiles are available for many biomarkers following the time of injury which can be used to plan sampling (eg Endre et al. (2011)) . For novel biomarkers, frequent sampling will be necessary to establish their time course. The first 12 hours following injury are crucial as this is the window for early intervention. There is also a need to discover and assess injury biomarkers with slightly longer (~24 to 36-h) time courses as samples may not be able to be taken earlier in many critically ill patients. Very few studies have measured biomarker profiles beyond a two or three days, yet there is tantilising evidence that some biomarkers (eg KIM-1) may become elevated during a repair phase which in some patients may not begin until several days following injury.

The choice of outcome variable is particularly important. Despite the caveats discussed with respect to using creatinine changes as a surrogate for change in GFR and the issue of comparing injury to function, we are still in a period where a functional definition of AKI is likely to be the outcome of choice for biomarker studies. Use of non-standard AKI definitions are unhelpful and should be avoided in preference to the AKIN, RIFLE and, potentially, new KDIGO AKI definitions. The duration as well as severity of creatinine increase is also of importance. The AKIN definition requires only one time point at which plasma creatinine may be elevated. This allows for mild and transient changes in function to equally be recorded as AKI. We have shown that transient increases in creatinine are associated with injury biomarkers (Nejat et al (2011b)), although not to the extent of more sustained increases. We recommend evaluating biomarkers in relation to duration of increase of creatinine as well as the increase itself.

Most biomarker studies are underpowered. The number of participants are rarely calculated *a priori*. Typically we want to ascertain if a biomarker is clinically useful. At a minimum we would want an AUC of 0.7, more likely 0.85 or greater. Given an expected incidence (proportion), *I*, in the population, how many participants (*N*) will we need in the study? Hanley and McNeil provide a general equation which we may adapt as we are interested in the difference between the true AUC and 0.5 (Hanley & McNeil (1982)):

$$N = \frac{1}{I} \left[\frac{0.5773Z_{\alpha} + Z_{\beta} \sqrt{0.1667 + \frac{AUC}{2 - AUC} + \frac{2AUC^2}{1 + AUC} - 2AUC^2}}{AUC - 0.5} \right]^2$$
(12)

where Z_{α} is the Z coefficient for a Type I error and Z_{β} for a Type II error. In order to detect a true AUC of 0.7 with p<0.05 ($Z_{\alpha}=1.96$ (two sided)) and at 80% power ($Z_{\beta}=0.84$) in a population with a 30% incidence 212 participants would be needed. A population with a much lower incidence, say 5%, would need a much greater sample (N=1269). If we were comparing a known biomarker with an AUC of 0.7 and wanting to know if another biomarker was better (AUC of 0.8 or more), then at p<0.05, 80% power and 30% incidence we would need 676 participants. This highlights the importance of knowing *a priori* the AKI incidence, what a clinically relevant AUC would be, and choosing a sample size appropriately.

In larger studies it is possible to assess the biomarker's ability to predict premature death or need for renal replacement therapy. In both cases, it is important to assess the risk relative to known risk factors. A useful technique is to use logistic regression models and compare a model of known risk factors with one with the same risk factors plus the biomarker. The integrated discrimination improvement (IDI) is a useful technique which, along with the AUC, allows for analysis of biomarker performance across the whole range of biomarker concentrations (Pencina et al. (2008)).

Studies in heterogeneous populations with multiple aetiologies and timing of injury are particularly difficult. Given the recent findings with respect to biomarker dependency on pre-existing renal function it is important that study size is sufficient to allow for cohort analysis, in particular cohorts of CKD and sepsis. In studies with CKD patients it is important to measure urinary albumin. Albumin is a possible AKI biomarker in its own right, but also competes for reabsorption in the proximal tubules with cystatin C and NGAL, IL-18 and possibly other markers (Nejat et al (2011a)).

The quality of reporting of biomarker studies varies widely. The Standards of Reporting of Diagnostic Accuracy (STARD) statement provides minimum standards and is worth referring to in the planning of the study (Bossuyt et al. (2003)). Biomarker concentrations should almost always be presented as median with inter-quartile range as they are usually non-normal distributions. With the jury still out on "normalising" urinary biomarkers to urinary creatinine we have recommended that both normalised and non-normalised results be presented.

4.3 Clinical trials

Clinical trial design follows one of three paradigms: (i) Prevention, namely treatment prior to a procedure which may cause injury; (ii) Early intervention following following a raised biomarker, but prior to observation of functional change (eg Endre et al. (2010)); (iii) Late intervention following an elevation of creatinine (Pickering & Endre (2009c); Pickering et al. (2011)). Whilst it may be anticipated that if successful prevention or early intervention treatments are developed there will be less need for late intervention treatments, late intervention treatments will still be needed either where prevention or early intervention fails, or where it is not possible to administer it. Injury biomarkers will play a role in all three paradigms. Some are being identified as risk factors, prior to a procedure (Bennett et al. (2008)), whilst others will be elevated early enough after injury to allow early intervention, finally others will serve as outcome variables. As with plasma creatinine as an outcome variable, analogous to the RAVC, it is likely that a continuous measurement of the biomarker over a pre-specified duration will serve best as an outcome variable.

		Epidemiology	Biomarker Studies	Clinical Trials
1	Use a continuous outcome variable (eg RAVC)	n/a	possibly	✓
2	Use RIFLE, AKIN or KDIGO definition as outcome variables (including for CIN trials)	~	•	✓
3	Use a hierarchical-adjudicated approach to determine baseline creatinine		V	
4	Include hard outcomes (RRT, Death)			~
5	Include plasma cystatin C as an outcome	n/a		/
6	For subcohort analysis use KDIGO guidelines for CKD staging (Levey et al. (2003))	,	✓	V
7	Measure 2 to 4-h creatinine clearance	n/a	✓	✓
8	Use corrected ΔGFR (%) for RIFLE classes R or F (Pickering & Endre (2009a))	n/a	•	✓
8	Apply treatment within the time window following insult determined by experimental &/or pilot data	n/a	n/a	•
9	Measure biomarkers within the time window following insult determined by experimental &/or pilot data	n/a	V	n/a
10	Report urinary biomarker concentrations & concentrations normalised to urinary creatinine	n/a	✓	n/a
11	Report median and inter-quartile range for biomarker concentrations	n/a	•	n/a
12	Report times and duration for which plasma outcomes were determined	~	✓	✓

Table 3. Practical considerations

5. Conclusion

The discovery of many early biomarkers of kidney injury has begun to shift the paradigm from assessment of change in filtration function to measurement of direct injury. Lest we throw the baby out with the bathwater we must recognise the complementary role of assessing both renal injury and renal function. Our techniques for assessing injury are still in their infancy, but show much promise. Our techniques for assessing function have a long history, yet, as we have shown, have room for improvement. In particular, we are learning to use appropriate surrogates of function, categorical or continuous, depending on the type of study we are conducting. We await the development of rapid, near "real-time" measures of function which are the missing link in enabling us to understand the temporal profile of injury in relation to functional change. We also look forward to an era of clinical trials which utilise the injury biomarkers discovered to date so as to properly test of drugs found to be effective soon after injury in experimental models.

6. Acknowledgements

Dr Pickering is supported by an Australia and New Zealand Society of Nephrologists enabling and infrastructure grant and the Marsden Fund Council from government funding, administered by the Royal Society of New Zealand.

7. References

- Ahlstrom, A., Kuitunen, A., Peltonen, S., Hynninen, M., Tallgren, M., Aaltonen, J. & Pettila, V. (2006). Comparison of 2 acute renal failure severity scores to general scoring systems in the critically ill, *Am J Kid Dis* 48(2): 262–268.
- Akcay, A., Nguyen, Q. & Edelstein, C. L. (2009). Mediators of inflammation in acute kidney injury, *Mediat Inflamm* 2009: 137072.
- Alachkar, N., Rabb, H. & Jaar, B. G. (2010). Urinary Biomarkers in Acute Kidney Transplant Dysfunction, *Nephron Clin Pract* 118(2): c173–c181.
- Alejandro, V., Scandling, J., Sibley, R., Dafoe, D., Alfrey, E., Deen, W. & Myers, B. D. (1995). Mechanisms of filtration failure during postischemic injury of the human kidney. A study of the reperfused renal allograft, *J Clin Invest* 95(2): 820–831.
- Ali, T., Khan, I., Simpson, W., Prescott, G., Townend, J., Smith, W. & Macleod, A. (2007). Incidence and outcomes in acute kidney injury: a comprehensive population-based study, *J Am Soc Nephrol* 18(4): 1292–1298.
- American Society of Nephrology Renal Research Report (2005). 16(7): 1886–1903.
- Bagshaw, S. M., George, C., Dinu, I. & Bellomo, R. (2008). A multi-centre evaluation of the RIFLE criteria for early acute kidney injury in critically ill patients., *Nephrol Dial Transpl* 23(4): 1203–1210.
- Bagshaw, S. M., Langenberg, C., Haase, M., Wan, L., May, C. N. & Bellomo, R. (2007). Urinary biomarkers in septic acute kidney injury, *Intens Care Med* 33(7): 1285–1296.
- Bagshaw, S. M., Uchino, S., Cruz, D., Bellomo, R., Morimatsu, H., Morgera, S., Schetz, M., Tan, I., Bouman, C., Macedo, E., Gibney, N., Tolwani, A., Oudemans-van Straaten, H. M., Ronco, C., Kellum, J. A. & Beginning and Ending Supportive Therapy for the Kidney (BEST Kidney) Investigators (2009). A comparison of observed versus estimated baseline creatinine for determination of RIFLE class in patients with acute kidney injury., *Nephrol Dial Transpl* 24(9): 2739–2744.
- Bellomo, R., Ronco, C., Kellum, J. A., Mehta, R. L., Palevsky, P. M. & workgroup, A. D. Q. I. (2004). Acute renal failure definition, outcome measures, animal models, fluid therapy and information technology needs: the Second International Consensus Conference of the Acute Dialysis Quality Initiative (ADQI) Group, *Crit Care* 8(4): R204–12.
- Bennett, M. R. & Devarajan, P. (2011). Proteomic analysis of acute kidney injury: Biomarkers to mechanisms, *Proteom Clin Appl* 5(1-2): 67–77.
- Bennett, M. R., Ravipati, N., Ross, G., Nguyen, M. T., Hirsch, R., Beekman, R. H., Rovner, L. & Devarajan, P. (2008). Using proteomics to identify preprocedural risk factors for contrast induced nephropathy, *Proteom Clin Appl* 2(7-8): 1058–1064.
- Berliner, R. (1995). Homer Smith: His contribution to physiology, *J Am Soc Nephrol* 5(12): 1988–1992.
- Bjornsson, T. (1979). Use of serum creatinine concentrations to determine renal-function, *Clin Pharmacokinet* 4(3): 200–222.

- Bonventre, J. V., Vaidya, V. S., Schmouder, R., Feig, P. & Dieterle, F. (2010). Next-generation biomarkers for detecting kidney toxicity, *Nat Biotechnol* 28(5): 436–440.
- Bossuyt, P., Reitsma, J., Bruns, D., Gatsonis, C., Glasziou, P., Irwig, L., Moher, D., Rennie, D., de Vet, H. & Lijmer, J. (2003). The STARD statement for reporting studies of diagnostic accuracy: Explanation and elaboration, *Clin Chem* 49(1): 7–18.
- Brown, J. R., Kramer, R. S., Coca, S. G. & Parikh, C. R. (2010). Duration of Acute Kidney Injury Impacts Long-Term Survival After Cardiac Surgery, *Ann Thorac Surg* 90(4): 1142–1149.
- Bulent Gul, C., Gullulu, M., Oral, B., Aydinlar, A., Oz, O., Budak, F., Yilmaz, Y. & Yurtkuran, M. (2008). Urinary IL-18: a marker of contrast-induced nephropathy following percutaneous coronary intervention?, *Clin Biochem* 41(7-8): 544–547.
- Cerda, J., Lameire, N., Eggers, P., Pannu, N., Uchino, S., Wang, H., Bagga, A. & Levin, A. (2008). Epidemiology of acute kidney injury, *Clin J Am Soc Nephro* 3(3): 881–886.
- Chertow, G. M., Burdick, E., Honour, M., Bonventre, J. V. & Bates, D. (2005). Acute kidney injury, mortality, length of stay, and costs in hospitalized patients, *J Am Soc Nephrol* 16: 3365–3370.
- Chiou, W. L. & Hsu, F. H. (1975). Pharmacokinetics of creatinine in man and its implications in the monitoring of renal function and in dosage regimen modifications in patients with renal insufficiency, *J Clin Pharmacol* 15(5-6): 427–434.
- Chow, M. S. (1985). A method for determining the pharmacokinetics of endogenous creatinine without exogenous creatinine administration, *Biopharm Drug Dispos* 6(2): 201–208.
- Coca, S. G., King, J. T., Rosenthal, R. A., Perkal, M. F. & Parikh, C. R. (2010). The duration of postoperative acute kidney injury is an additional parameter predicting long-term survival in diabetic veterans, *Kidney Int* 78(9): 926–933.
- Coca, S. G., Yalavarthy, R., Concato, J. & Parikh, C. R. (2008). Biomarkers for the diagnosis and risk stratification of acute kidney injury: A systematic review, *Kidney Int* 73: 1001–1016.
- Coca, S. G., Yusuf, B., Shlipak, M. G., Garg, A. X. & Parikh, C. R. (2009). Long-term Risk of Mortality and Other Adverse Outcomes After Acute Kidney Injury: A Systematic Review and Meta-analysis, *Am J Kid Dis* 53(6): 961–973.
- Cockcroft, D. & Gault, M. (1976). Prediction of creatinine clearance from serum creatinine, *Nephron* 16(1): 31–41.
- Cruz, D. N., Bolgan, I., Perazella, M. A., Bonello, M., de Cal, M., Corradi, V., Polanco, N., Ocampo, C., Nalesso, F., Piccinni, P., Ronco, C. & on Acute Kidney Injury NEiPHROS-AKI Investigators, N. E. I. P. H. R. O. S. (2007). North East Italian Prospective Hospital Renal Outcome Survey on Acute Kidney Injury (NEiPHROS-AKI): targeting the problem with the RIFLE Criteria, *Clin J Am Soc Nephro* 2(3): 418–425.
- DeLong, E., DeLong, D. & Clarke-Pearson, D. (1988). Comparing the areas under 2 or more correlated receiver operating characteristic curves a nonparametric approach, *Biometrics* 44(3): 837–845.
- Devarajan, P. (2006). Update on mechanisms of ischemic acute kidney injury, *J Am Soc Nephrol* 17(6): 1503–1520.
- Devarajan, P. (2008). Proteomics for the investigation of acute kidney injury, *Contrib Nephrol* 160: 1–16.

- Dieterle, F., Perentes, E., Cordier, A., Roth, D. R., Verdes, P., Grenet, O., Pantano, S., Moulin, P., Wahl, D., Mahl, A., End, P., Staedtler, F., Legay, F., Carl, K., Laurie, D., Chibout, S.-D., Vonderscher, J. & Maurer, G. (2010). Urinary clusterin, cystatin C, beta2-microglobulin and total protein as markers to detect drug-induced kidney injury, *Nat Biotechnol* 28(5): 463–469.
- Edelstein, C. L. & Faubel, S. (2010). Chapter 5, Biomarkers in Acute Kidney Injury, Elsevier Inc., pp. 177–232.
- Eknoyan, G. (2002). Emergence of the concept of acute renal failure, *Am J Nephrol* 22(2-3): 225–230.
- Endre, Z. H., Allis, J. & Radda, G. (1989). Toxicity of dysprosium shift-reagents in the isolated perfused rat kidney, *Magnet Reson Med* 11: 267–274.
- Endre, Z. H. & Pickering, J. W. (2010). Outcome definitions in non-dialysis intervention and prevention trials in acute kidney injury (AKI)., *Nephrol Dial Transpl* 25(1): 107–118.
- Endre, Z. H., Pickering, J. W., Walker, R. J., Devarajan, P., Edelstein, C. L., Bonventre, J. V., Frampton, C. M., Bennett, M. R., Ma, Q., Sabbisetti, V. S., Vaidya, V. S., Walcher, A. M., Shaw, G. M., Henderson, S. J., Nejat, M., Schollum, J. B. W. & George, P. M. (2011). Improved performance of urinary biomarkers of acute kidney injury in the critically ill by stratification for injury duration and baseline renal function., *Kidney Int* 79(10): 1119–1130.
- Endre, Z. H., Walker, R. J., Pickering, J. W., Shaw, G. M., Frampton, C. M., Henderson, S. J., Hutchison, R., Mehrtens, J. E., Robinson, J. M., Schollum, J. B. W., Westhuyzen, J., Celi, L. A., McGinley, R. J., Campbell, I. J. & George, P. M. (2010). Early intervention with erythropoietin does not affect the outcome of acute kidney injury (the EARLYARF trial)., *Kidney Int* 77(11): 1020–1030.
- Endre, Z. H. & Westhuyzen, J. (2008). Early detection of acute kidney injury: Emerging new biomarkers (Review Article), *Nephrology* 13(2): 91–98.
- Ferguson, M. A., Vaidya, V. S. & Bonventre, J. V. (2008). Biomarkers of nephrotoxic acute kidney injury, *Toxicology* 245(3): 182–193.
- Fishbane, S. (2008). N-acetylcysteine in the prevention of contrast-induced nephropathy, *Clin J Am Soc Nephro* 3(1): 281–287.
- Frank, H. A., Seligman, A. M. & Fine, J. (1946). Treatment of uremia after acute renal failure by peritoneal irrigation., *J Am Med Assoc* 130: 703–705.
- Gibney, N., Hoste, E., Burdmann, E. A., Bunchman, T., Kher, V., Viswanathan, R., Mehta, R. L. & Ronco, C. (2008). Timing of initiation and discontinuation of renal replacement therapy in AKI: Unanswered key questions, *Clin J Am Soc Nephro* 3(3): 876–880.
- Goldberg, A., Kogan, E., Hammerman, H., Markiewicz, W. & Aronson, D. (2009). The impact of transient and persistent acute kidney injury on long-term outcomes after acute myocardial infarction, *Kidney Int* 76(8): 900–906.
- Griffiths, R. (1996). Muscle mass, survival, and the elderly ICU patient, *Nutrition* 12(6): 456–458.
- Haase, M., Bellomo, R., Devarajan, P., Schlattmann, P., Haase-Fielitz, A. & NGAL Meta-analysis Investigator Group (2009). Accuracy of neutrophil gelatinase-associated lipocalin (NGAL) in diagnosis and prognosis in acute kidney injury: a systematic review and meta-analysis., *Am J Kid Dis* 54(6): 1012–1024.

- Haase, M., Bellomo, R. & Haase-Fielitz, A. (2010). Novel biomarkers, oxidative stress, and the role of labile iron toxicity in cardiopulmonary bypass-associated acute kidney injury, *J Am Coll Cardiol* 55(19): 2024–2033.
- Haase, M., Devarajan, P., Haase-Fielitz, A., Bellomo, R., Cruz, D. N., Wagener, G., Krawczeski, C. D., Koyner, J. L., Murray, P., Zappitelli, M., Goldstein, S. L., Makris, K., Ronco, C., Martensson, J., Martling, C.-R., Venge, P., Siew, E., Ware, L. B., Ikizler, T. A. & Mertens, P. R. (2011). The outcome of neutrophil gelatinase-associated lipocalin-positive subclinical acute kidney injury a multicenter pooled analysis of prospective studies., *J Am Coll Cardiol* 57(17): 1752–1761.
- Hall, I., Yarlagadda, S., Coca, S. G., Wang, Z., Doshi, M., Devarajan, P., Han, W., Marcus, R. & Parikh, C. R. (2010). IL-18 and Urinary NGAL Predict Dialysis and Graft Recovery after Kidney Transplantation, *J Am Soc Nephrol* 21(1): 189–197.
- Han, W. K., Bailly, V., Abichandani, R., Thadhani, R. & Bonventre, J. V. (2002). Kidney Injury Molecule-1 (KIM-1): a novel biomarker for human renal proximal tubule injury., *Kidney Int* 62(1): 237–244.
- Hanley, J. A. & McNeil, B. J. (1982). The meaning and use of the area under a receiver operating characteristic (ROC) curve, *Radiology* 143(1): 29–36.
- Hansen, J. T. (2002). *Netter's Atlas of Human Physiology*, Icon Learning Systems, Teterboro, NJ, USA.
- Harjai, K. J., Ralzada, A., Shenoy, C., Sattur, S., Orshaw, P., Yaeger, K., Boura, J., Aboufares, A., Sporn, D. & Stapleton, D. (2008). A comparison of contemporary definitions of contrast nephropathy in patients undergoing percutaneous coronary intervention and a proposal for a novel nephropathy grading system, *Am J Cardiol* 101(6): 812–819.
- Heemskerk, S., Masereeuw, R., Russel, F. G. M. & Pickkers, P. (2009). Selective iNOS inhibition for the treatment of sepsis-induced acute kidney injury, *Nat Rev Nephrol* 5(11): 629–640.
- Heyman, S. N., Rosenberger, C. & Rosen, S. (2010). Experimental ischemia-reperfusion: biases and myths-the proximal vs. distal hypoxic tubular injury debate revisited, *Kidney Int* 77(1): 9–16.
- Hoste, E. A. J., Clermont, G., Kersten, A., Venkataraman, R., Angus, D. C., de Bacquer, D. & Kellum, J. A. (2006). RIFLE criteria for acute kidney injury are associated with hospital mortality in critically ill patients: a cohort analysis, *Crit Care* 10(3): R73.
- Ichimura, T., Bonventre, J. V., Bailly, V., Wei, H., Hession, C., Cate, R. & Sanicola, M. (1998). Kidney injury molecule-1 (KIM-1), a putative epithelial cell adhesion molecule containing a novel immunoglobulin domain, is up-regulated in renal cells after injury, *J Biol Chem* 273(7): 4135–4142.
- Jo, S. K., Rosner, M. H. & Okusa, M. D. (2007). Pharmacologic treatment of acute kidney injury: why drugs haven't worked and what is on the horizon, *Clin J Am Soc Nephro* 2(2): 356–365.
- KDOQI Clinical Practice Guidelines for Chronic Kidney Disease: Evaluation, Classification, and Stratification (2002).
- Lafrance, J.-P. & Miller, D. R. (2010). Defining acute kidney injury in database studies: the effects of varying the baseline kidney function assessment period and considering CKD status., *Am J Kid Dis* 56(4): 651–660.
- Lameire, N. H., van Biesen, W. & Vanholder, R. (2006). The changing epidemiology of acute renal failure, *Nat Clin Pract Nephrol* 2(7): 364–377.

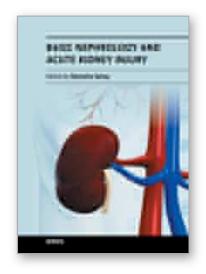
- Lassnigg, A., Schmid, E. R., Hiesmayr, M., Falk, C., Druml, W., Bauer, P. & Schmidlin, D. (2008). Impact of minimal increases in serum creatinine on outcome in patients after cardiothoracic surgery: do we have to revise current definitions of acute renal failure?, *Crit Care Med* 36(4): 1129–1137.
- Lassnigg, A., Schmidlin, D., Mouhieddine, M., Bachmann, L., Druml, W., Bauer, P. & Hiesmayr, M. (2004). Minimal changes of serum creatinine predict prognosis in patients after cardiothoracic surgery: A prospective cohort study, *J Am Soc Nephrol* 15(6): 1597–1605.
- Levey, A., Bosch, J., Lewis, J., Greene, T., Rogers, N. & Roth, D. (1999). A more accurate method to estimate glomerular filtration rate from serum creatinine: A new prediction equation, *Ann Intern Med* 130(6): 461–470.
- Levey, A., Coresh, J., Balk, E., Kausz, A., Levin, A., Steffes, M., Hogg, R., Perrone, R., Lau, J. & Eknoyan, G. (2003). National kidney foundation practice guidelines for chronic kidney disease: Evaluation, classification, and stratification, *Ann Intern Med* 139(2): 137–147.
- Levey, A. S., Stevens, L. A., Schmid, C. H., Zhang, Y., Castro, A. F., Feldman, H. I., Kusek, J. W., Eggers, P., van Lente, F., Greene, T. & Coresh, J. (2009). A New Equation to Estimate Glomerular Filtration Rate, *Ann Intern Med* 150(9): 604–613.
- Levinsky, N. & Berliner, R. (1959). Changes in composition of the urine in ureter and bladder at low urine flow, *Am J Physiol* 196(3): 549–553.
- Lopes, J. A., Melo, M. J., Viegas, A., Raimundo, M., Camara, I., Antunes, F. & Gomes da Costa, A. (2011). Acute kidney injury in hospitalized HIV-infected patients: a cohort analysis, *Nephrol Dial Transpl*.
- Macedo, E., Bouchard, J., Soroko, S. H., Chertow, G. M., Himmelfarb, J., Ikizler, T. A., Paganini, E. P., Mehta, R. L. & Program to Improve Care in Acute Renal Disease Study (2010). Fluid accumulation, recognition and staging of acute kidney injury in critically-ill patients., *Crit Care* 14(3): R82.
- Malyszko, J. (2010). Biomarkers of acute kidney injury in different clinical settings: a time to change the paradigm?, *Kidney Blood Press Res* 33(5): 368–382.
- McGrath, J. (1852). Case of Ischuria Renalsis, Am J Med Sci 48(October): 390–392.
- Mcilroy, D. R., Wagener, G. & Lee, H. T. (2010). Neutrophil gelatinase-associated lipocalin and acute kidney injury after cardiac surgery: the effect of baseline renal function on diagnostic performance., *Clin J Am Soc Nephro* 5(2): 211–219.
- Mehta, R. L., Kellum, J., Shah, S., Molitoris, B. A., Ronco, C., Warnock, D., Levin, A. & the Acute Kidney Injury Network (2007). Acute Kidney Injury Network: report of an initiative to improve outcomes in acute kidney injury, *Crit Care* 11(2): R31.
- Mehta, R., McDonald, B., Gabbai, F., Pahl, M., Farkas, A., Pascual, M., Zhuang, S., Kaplan, R. & Chertow, G. (2002). Nephrology consultation in acute renal failure: Does timing matter?, *Am J Med* 113(6): 456–461.
- Melnikov, V. Y., Ecder, T., Fantuzzi, G., Siegmund, B., Lucia, M. S., Dinarello, C. A., Schrier, R. W. & Edelstein, C. L. (2001). Impaired IL-18 processing protects caspase-1-deficient mice from ischemic acute renal failure., *J Clin Invest* 107(9): 1145–1152.
- Mishra, J., Dent, C. L., Tarabishi, R., Mitsnefes, M. M., Ma, Q., Kelly, C., Ruff, S. M., Zahedi, K., Shao, M., Bean, J., Mori, K., Barasch, J. M. & Devarajan, P. (2005). Neutrophil gelatinase-associated lipocalin (NGAL) as a biomarker for acute renal injury after cardiac surgery, *Lancet* 365(9466): 1231–1238.

- Mishra, J., Ma, Q., Prada, A., Mitsnefes, M., Zahedi, K., Yang, J., Barasch, J. M. & Devarajan, P. (2003). Identification of neutrophil gelatinase-associated lipocalin as a novel early urinary biomarker for ischemic renal injury, *J Am Soc Nephrol* 14: 2534–2543.
- Moller, E., McIntosh, J. & van Slyke, D. (1928). Studies of urea excretion II Relationship between urine volume and the rate of urea excretion by normal adults, *J Clin Invest* 6(3): 427–465.
- Myers, B. D., Miller, D., Mehigan, J., Olcott, C., Golbetz, H., Robertson, C., Derby, G., Spencer, R. & Friedman, S. (1984). Nature of the renal injury following total renal ischemia in man, *J Clin Invest* 73(2): 329–341.
- Nejat, M., Hill, J. V., Pickering, J. W., Edelstein, C. L., Devarajan, P. & Endre, Z. H. (2011). Albuminuria increases cystatin C excretion: implications for urinary biomarkers, *Nephrol Dial Transpl*.
- Nejat, M., Pickering, J. W., Devarajan, P., Bonventre, J. V., Edelstein, C. L. & Endre, Z. H. (2011). Acute Kidney Injury (AKI) biomarkers are increased in prerenal AKI, *World Congress of Nephrology*.
- Nejat, M., Pickering, J. W., Walker, R. J. & Endre, Z. H. (2010). Rapid detection of acute kidney injury by plasma cystatin C in the intensive care unit., *Nephrol Dial Transpl* 25(10): 3283–3289.
- Nigwekar, S., Navaneethan, S., Parikh, C. R. & Hix, J. (2009). Atrial natriuretic peptide for preventing and treating acute kidney injury, *Cochrane Db Syst Rev* (4): Art. No.: CD006028.
- Ozer, J. S., Dieterle, F., Troth, S., Perentes, E., Cordier, A., Verdes, P., Staedtler, F., Mahl, A., Grenet, O., Roth, D. R., Wahl, D., Legay, F., Holder, D., Erdos, Z., Vlasakova, K., Jin, H., Yu, Y., Muniappa, N., Forest, T., Clouse, H. K., Reynolds, S., Bailey, W. J., Thudium, D. T., Topper, M. J., Skopek, T. R., Sina, J. F., Glaab, W. E., Vonderscher, J., Maurer, G., Chibout, S.-D., Sistare, F. D. & Gerhold, D. L. (2010). A panel of urinary biomarkers to monitor reversibility of renal injury and a serum marker with improved potential to assess renal function, *Nat Biotechnol* 28(5): 486–494.
- Parikh, C. R., Jani, A., Melnikov, V., Faubel, S. & Edelstein, C. L. (2004). Urinary interleukin-18 is a marker of human acute tubular necrosis, *Am J Kid Dis* 43: 405–414.
- Parikh, C. R., Mishra, J., Thiessen-Philbrook, H., Dursun, B., Ma, Q., Kelly, C., Dent, C. L., Devarajan, P. & Edelstein, C. L. (2006). Urinary IL-18 is an early predictive biomarker of acute kidney injury after cardiac surgery, *Kidney Int* 70: 199–203.
- Pencina, M. J., D'Agostino, R. B. & Vasan, R. S. (2008). Evaluating the added predictive ability of a new marker: From area under the ROC curve to reclassification and beyond, *Stat Med* 27(2): 157–172.
- Perrone, R., Madias, N. & Levey, A. (1992). Serum creatinine as an index of renal-function new insights into old concepts, *Clin Chem* 38(10): 1933–1953.
- Pickering, J. W. & Endre, Z. H. (2009a). GFR shot by RIFLE: errors in staging acute kidney injury, *Lancet* 373(9672): 1318–1319.
- Pickering, J. W. & Endre, Z. H. (2009b). RIFLE and AKIN–maintain the momentum and the GFR!, *Crit Care* 13(5): 416.
- Pickering, J. W. & Endre, Z. H. (2009c). Secondary prevention of acute kidney injury., *Current opinion in critical care* 15(6): 488–497.

- Pickering, J. W. & Endre, Z. H. (2010). Back-calculating baseline creatinine with MDRD misclassifies acute kidney injury in the intensive care unit., *Clin J Am Soc Nephro* 5(7): 1165–1173.
- Pickering, J. W., Frampton, C. M. & Endre, Z. H. (2009). Evaluation of Trial Outcomes in Acute Kidney Injury by Creatinine Modeling, *Clin J Am Soc Nephro* 4(11): 1705–1715.
- Pickering, J. W., Md Ralib, A., Nejat, M. & Endre, Z. H. (2011). New considerations in the design of clinical trials of acute kidney injury, *Clin Invest* 1(5): 637–650.
- Rabito, C., Halpern, E. F., Scott, J. & Tolkoff-Rubin, N. (2010). Accurate, fast, and convenient measurement of glomerular filtration rate in potential renal transplant donors, *Transplantation* 90(5): 510–517.
- Rehberg, P. B. (1926). Studies on Kidney Function: The Rate of Filtration and Reabsorption in the Human Kidney., *Biochem J* 20(3): 447–460.
- Ricci, Z., Cruz, D. & Ronco, C. (2008). The RIFLE criteria and mortality in acute kidney injury: A systematic review, *Kidney Int* 73(5): 538–546.
- Roberts, D. M., Wilks, M. F., Roberts, M. S., Swaminathan, R., Mohamed, F., Dawson, A. H. & Buckley, N. A. (2011). Changes in the concentrations of creatinine, cystatin C and NGAL in patients with acute paraquat self-poisoning., *Toxicol. Lett.* 202(1): 69–74.
- Rule, A. D. (2010). The CKD-EPI equation for estimating GFR from serum creatinine: real improvement or more of the same?, *Clin J Am Soc Nephro* 5(6): 951–953.
- Ryzner, K. L. (2010). Evaluation of Aminoglycoside Clearance Using the Modification of Diet in Renal Disease Equation Versus the Cockcroft-Gault Equation as a Marker of Glomerular Filtration Rate, *Ann Pharmacother* 44(6): 1030–1037.
- Shemesh, O., Golbetz, H., Kriss, J. P. & Myers, B. D. (1985). Limitations of creatinine as a filtration marker in glomerulopathic patients., *Kidney Int* 28(5): 830–838.
- Siew, E. D., Matheny, M. E., Ikizler, T. A., Lewis, J. B., Miller, R. A., Waitman, L. R., Go, A. S., Parikh, C. R. & Peterson, J. F. (2010). Commonly used surrogates for baseline renal function affect the classification and prognosis of acute kidney injury, *Kidney Int* 77(6): 536–542.
- Sjostrom, P., Tidman, M. & Jones, I. (2005). Determination of the production rate and non-renal clearance of cystatin C and estimation of the glomerular filtration rate from the serum concentration of cystatin C in humans, *Scand J Clin Lab Inv* 65(2): 111–124.
- Smith, H., Finkelstein, N., Alimnosa, L., Crawford, B. & Graber, M. (1945). The renal clearances of substituted hippuric acid derivatives and other aromatic acids in dog and man, *J Clin Invest* 24(3): 388–404.
- Star, R. A. (1998). Treatment of acute renal failure, Kidney Int 54: 1817–1831.
- Uchino, S., Bellomo, R., Bagshaw, S. M. & Goldsmith, D. (2010). Transient azotaemia is associated with a high risk of death in hospitalized patients, *Nephrol Dial Transpl* 25(6): 1833–1839.
- Uchino, S., Bellomo, R., Goldsmith, D., Bates, S. & Ronco, C. (2006). An assessment of the RIFLE criteria for acute renal failure in hospitalized patients, *Crit Care Med* 34: 1913–1917.
- Vaidya, V. S., Ferguson, M. A. & Bonventre, J. V. (2008). Biomarkers of acute kidney injury, *Annu Rev Pharmacol* 48: 463–493.
- Vaidya, V. S., Ozer, J. S., Dieterle, F., Collings, F. B., Ramirez, V., Troth, S., Muniappa, N., Thudium, D., Gerhold, D., Holder, D. J., Bobadilla, N. A., Marrer, E., Perentes, E., Cordier, A., Vonderscher, J., Maurer, G., Goering, P. L., Sistare, F. D. & Bonventre,

- J. V. (2010). Kidney injury molecule-1 outperforms traditional biomarkers of kidney injury in preclinical biomarker qualification studies, *Nat Biotechnol* 28(5): 478–485.
- Waikar, S. S., Sabbisetti, V. S. & Bonventre, J. V. (2010). Normalization of urinary biomarkers to creatinine during changes in glomerular filtration rate, *Kidney Int* 78(5): 486–494.
- Wang, E., Sandoval, R. M., Campos, S. B. & Molitoris, B. A. (2010). Rapid diagnosis and quantification of acute kidney injury using fluorescent ratio-metric determination of glomerular filtration rate in the rat, *Am J Physiol-Renal* 299(5): F1048–55.
- Watson, P., Watson, I. & Batt, R. (1980). Total-body water volumes for adult males and females estimated from simple anthropometric measurements, *Am J Clin Nutr* 33(1): 27–39.
- Westhuyzen, J. (2006). Cystatin C: a promising marker and predictor of impaired renal function, *Ann Clin Lab Sci* 36(4): 387–394.
- Westhuyzen, J., Endre, Z. H., Reece, G., Reith, D., Saltissi, D. & Morgan, T. (2003). Measurement of tubular enzymuria facilitates early detection of acute renal impairment in the intensive care unit, *Nephrol Dial Transpl* 18: 543–551.
- Yu, H., Yanagisawa, Y., Forbes, M. A., Cooper, E. H., Crockson, R. A. & MacLennan, I. C. (1983). Alpha-1-microglobulin: an indicator protein for renal tubular function., *J Clin Pathol* 36(3): 253–259.
- Yu, Y., Jin, H., Holder, D., Ozer, J. S., Villarreal, S., Shughrue, P., Shi, S., Figueroa, D. J., Clouse, H., Su, M., Muniappa, N., Troth, S. P., Bailey, W., Seng, J., Aslamkhan, A. G., Thudium, D., Sistare, F. D. & Gerhold, D. L. (2010). Urinary biomarkers trefoil factor 3 and albumin enable early detection of kidney tubular injury, *Nat Biotechnol* 28(5): 470–477.
- Zoungas, S., Ninomiya, T., Huxley, R., Cass, A., Jardine, M., Gallagher, M., Patel, A., Vasheghani-Farahani, A., Sadigh, G. & Perkovic, V. (2009). Systematic Review: Sodium Bicarbonate Treatment Regimens for the Prevention of Contrast-Induced Nephropathy, *Ann Intern Med* 151(9): 631–638.





Basic Nephrology and Acute Kidney Injury

Edited by Prof. Manisha Sahay

ISBN 978-953-51-0139-0 Hard cover, 226 pages Publisher InTech Published online 02, March, 2012 Published in print edition March, 2012

The first section of the book covers the basics of nephrology and second section focuses on acute kidney injury. This easy to reference text examines the physiological and biochemical aspects of renal diseases - all in one convenient resource. Experts in the field discuss topics of increasing concern in nephrology including newer methods of assessing renal function. The field of acute kidney injury in nephrology is a rapidly evolving one with research translating into clinical guidelines and standards. This text brings together experts to provide an authoritative reference for management of AKI in various clinical settings. Pregnancy related AKI is an important entity which has also been discussed in detail. The recent advances in the field of critical care AKI have been incorporated as well and help the reader to update their knowledge.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

John W Pickering and Zoltán H Endre (2012). The Metamorphosis of Acute Renal Failure to Acute Kidney Injury, Basic Nephrology and Acute Kidney Injury, Prof. Manisha Sahay (Ed.), ISBN: 978-953-51-0139-0, InTech, Available from: http://www.intechopen.com/books/basic-nephrology-and-acute-kidney-injury/the-metamorphosis-of-acute-renal-failure-to-acute-kidney-injury



InTech Europe

University Campus STeP Ri Slavka Krautzeka 83/A 51000 Rijeka, Croatia Phone: +385 (51) 770 447

Fax: +385 (51) 686 166 www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai No.65, Yan An Road (West), Shanghai, 200040, China 中国上海市延安西路65号上海国际贵都大饭店办公楼405单元

Phone: +86-21-62489820 Fax: +86-21-62489821 © 2012 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the <u>Creative Commons Attribution 3.0</u> <u>License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



