

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

4,800

Open access books available

122,000

International authors and editors

135M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.

For more information visit www.intechopen.com



Role of Astrocytes in Neurodegenerative Diseases

George E. Barreto¹, Janneth Gonzalez¹,
Francisco Capani² and Ludis Morales¹

¹*Departamento de Nutrición y Bioquímica, Facultad de Ciencias,
Pontificia Universidad Javeriana, Bogotá D.C.,*

²*Instituto de Investigaciones Cardiológicas, Prof. Dr. Alberto Taquini,
CONICET-UBA Buenos Aires,*

¹*Colombia*

²*Argentina*

1. Introduction

The past several decades have given rise to many important discoveries and novel insights into the role of astrocytes in normal brain function and disease, firmly establishing concepts that describe the dynamic and reciprocal signaling networks between astrocytes, neurons and other cell types.

Brain aging, overt any neurodegenerative state, leads to inflammation, oxidative stress and cell death. Neurons are more susceptible to injury than astrocytes, as they have fewer antioxidant mechanisms and are therefore prone to excitotoxicity (Swanson *et al.*, 2004). Both normally and with aging, astrocytes support neurons by providing antioxidant protection, substrates for neuronal metabolism via neurovascular coupling, and glutamate clearance. Although astrocytes are generally more resilient than neurons, severe damage also results in astrocyte dysfunction, leading to increased neuronal death (Nedergaard & Dirnagl, 2005). Therefore, many recent efforts have focused on the astrocyte-neuron interaction and how astrocyte function can be improved to enhance neuronal support and survival (Swanson *et al.*, 2004). A growing body of data demonstrates that astrocytes play a multifaceted and complex role in the response to neuropathologies, including neurodegenerative, as they have potential to both enhance neuronal survival and regeneration and contribute to further injury (Sofroniew, 2000, 2009; Sofroniew & Vinters, 2010). Because of the diverse nature and complex biology of these cells, and the limited number of studies to date, their role in neurodegeneration deserves further study.

It is likely that diminished astrocytes function throughout the neurodegenerative process is a prominent determinant of both neuronal survival as well as survival of the entire organism (Shibata & Kobayashi, 2008). In this chapter we provide a brief overview of the pathophysiological events underlying brain aging, and in neurodegenerative diseases, and discusses how these events affect astrocytes response to these chronic neuropathologies, such as Alzheimer's AD and Parkinson's PD diseases, Amyotrophic Lateral Syndrome (ALS), and Multiple Sclerosis (MS).

2. Astrocytes function in the brain

Astrocytes are the most common cell type in mammalian brain. Glial fibrillary acidic protein (GFAP) and vimentin (Vim) constitute intermediate filaments (known also as nanofilaments) as part of the cytoskeleton in astrocytes. Reactive gliosis is a response of astrocytes to a variety of brain insults that is characterized by hypertrophy of the cell bodies and processes, altered gene expression, increase in the expression of GFAP, Vim and the calcium binding protein S100 β (Ridet *et al.*, 1997), and proliferation that may likely occur in some neurodegenerative diseases (Sofroniew, 2009; Sofroniew & Vinters, 2010). In contrast, because reactive astrocytes are ubiquitous in aged central nervous system (CNS) tissue, they are often regarded as uniformly harmful, provoking inflammation, releasing cytotoxins and chemokines that serve no purpose but to inhibit axonal regeneration and increase damage. The wide range of activities that astrocytes can exhibit *in vitro* contributes to uncertainty over whether these cells exert beneficial or detrimental effects after CNS degeneration. For example, potential protective effects could be provided by glutamate uptake and neurotrophin release, while potential detrimental effects might be caused by the release of inflammatory cytokines and cytotoxic radicals. Little information has been available on the roles played by reactive astrocytes in the response to experimental models of neurodegenerative diseases *in vivo*. For instance, aged astrocytes exhibit an elevated content of GFAP and of S100 β (Barreto *et al.*, 2009; Nichols, 1999). Use of oligonucleotide arrays has yielded the first profile of gene expression from the aging brain of mice and evidence that aging seems to be associated with an inflammatory response and oxidative stress both in neocortex, hippocampus and in cerebellum (Lee *et al.*, 2000; Zeier *et al.*, 2011), with parallels to human neurodegenerative disorders. GFAP is also one of the genes that undergoes a twofold increase in expression. Thus, the GFAP increases of the aged astrocytes may be the result of a response to the inflammatory and oxidative state of the aging brain. Indeed, better comprehension of the features that distinguish a normal, “healthy” old brain from a brain that is at an early stage of a neurodegenerative disease is a key aspect in developing treatments.

It is interesting to note that one of the characteristics of astrocytes in the aging brain – the number of astrocytes – is increased by ~20% (Peinado *et al.*, 1998; Pilegaard & Ladefoged, 1996; Rozovsky *et al.*, 1998; Salminen *et al.*, 2011). This response has been compared with reactive gliosis in response to injured or damaged neurons during aging. However, an alternative explanation is that increased number of astrocytes in the aging brain is required to provide the same level of neuroprotection that is present in the brain of a young animal.

One hallmark of the cellular response to brain aging, and in neurodegenerative states, is a rapid, dramatic increase in damaging free radicals, including nitric oxide (NO), superoxide, and peroxynitrite (Shibata & Kobayashi, 2008). On the other hand, astrocytes produce the beneficial antioxidants glutathione, superoxide dismutases (SODs 1, 2 and 3), and ascorbate (Figure 1, Anderson & Swanson, 2000; Dringen, 2000; Dringen *et al.*, 2000; Lindenau *et al.*, 2000; Sims *et al.*, 2004). Interestingly, neurons cocultured with astrocyte exhibit higher levels of glutathione compared with neurons cultured alone (Giordano *et al.*, 2009), suggesting that astrocytes provide additional antioxidant defense to neurons (Slemmer *et al.*, 2008). Similarly, astrocytes upregulate HO-1 (heme-oxygenase 1, Figure 2), a 32 kDa stress protein that degrades heme to biliverdin, free iron and carbon monoxide. Although the upregulation of this enzyme has been previously reported to confer neuroprotection following various brain insults (Beschorner *et al.*, 2000; Chen *et al.*, 2000; Espada *et al.*, 2010;

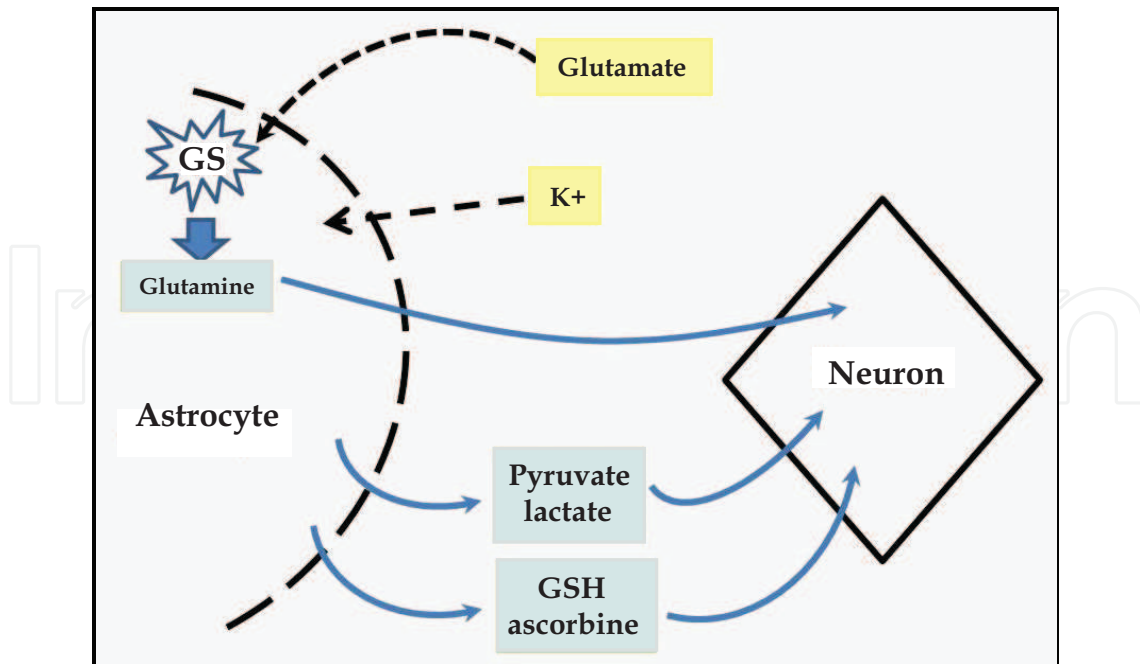


Fig. 1. Mechanisms of astrocyte support of neurons in the normal brain. Antioxidant defence includes release of glutathione and ascorbate. Regulation of extracellular levels of ions and neurotransmitters, especially K^+ and glutamate, strongly influence neuronal excitability. Elevated extracellular K^+ triggers astrocyte glycolysis and enhances lactate and pyruvate release which support neuronal metabolism. Sodium dependent glutamate uptake by astrocytes activates the Na^+/K^+ ATPase, stimulating glycolytic activity and production of lactate. Astrocytes and neurons are also coupled by the glutamate-glutamine cycle. Astrocytes take up glutamate, convert it to glutamine, release glutamine to the extracellular space where it is taken up by neurons and used to synthesize glutamate to replenish the neurotransmitter pool. Any deregulation of these mechanisms, as a common situation in some neurodegenerative diseases, will likely influence neuronal survival

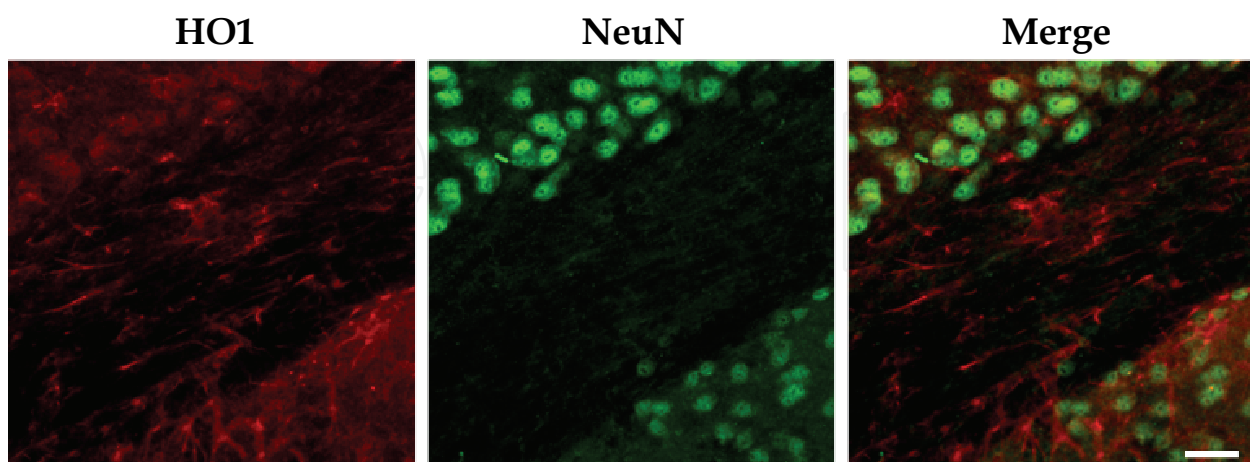


Fig. 2. Astrocytic HO-1 expression in corpus callosum. Immunostaining for HO-1 and NeuN (neuronal marker) was carried out in free floating brain sections of 3 months-old naïve male mice. Morphologically, HO-1 is expressed in shaped-like astrocytes, and does not seem to be expressed by neurons. Scale bar, 50 μm

Imuta *et al.*, 2007; Ku *et al.*, 2006; Le *et al.*, 1999; Takeda *et al.*, 2000), its overproduction in astrocytes may contribute to iron overload and mitochondrial insufficiency, characteristic of some neurodegenerative disorders (Fernandez-Checa *et al.*, 2010; Serviddio *et al.*, 2011). HO-1 is expressed by approximately 86% (Schipper *et al.*, 1995) and 77.1% (Schipper *et al.*, 1998) of GFAP-positive astrocytes in AD and PD, respectively, suggesting a possible role in the pathogenesis of these neurodegenerative diseases.

Control of energy metabolism is also controlled by astrocytes in the CNS. When astrocytes take up extracellular glutamate as a result of neuronal activity, the Na⁺/K⁺-ATPase and AMPA signaling trigger astrocyte uptake of glucose from the blood, as astrocytic endfeet contact capillaries (Caesar *et al.*, 2008; Magistretti, 2006). The glucose is then made into lactate, a substrate for neuronal energy, to further “fuel” active neurons (Magistretti & Pellerin, 1999; Figure 1). As mentioned above, astrocytes produce glutathione. In addition to its antioxidant properties, glutathione is the enzyme needed for the conversion of methylglyoxal, a toxic by-product of metabolism, into D-lactate by glyoxalase 1 (Cliffe & Waley, 1961). Although the role of astrocyte metabolism is relatively well-established in normal tissues, the role of astrocyte metabolism maintenance with aging and in neurodegenerative diseases is less clear (Bartnik-Olson *et al.*, 2010; Bentzer *et al.*, 2000; Floyd & Lyeth, 2007).

Astrocytes are also key players in the production and regulation of neurotransmitters, antioxidant production, potassium uptake, energy metabolism and neurovascular coupling in the CNS. Notably, astrocytes make glutamine, the precursor for the neurotransmitters glutamate and GABA, from glucose (Zou *et al.*, 2010). In addition to providing the precursors for neurotransmitters, one important role of astrocytes in the normal brain is to take up glutamate using the glutamate transporters GLAST and GLT-1 (Anderson & Swanson, 2000; Romera *et al.*, 2004; Schousboe & Waagepetersen, 2006), as excess glutamate leads to cell death via excitotoxicity (Tilleux & Hermans, 2007).

Astrocytes regulate neuronal activation by extracellular potassium uptake, and proper maintenance of ion gradients, such as potassium, as an important mechanism for regulating cell volume in both normal and pathological conditions (Jayakumar & Norenberg, 2010; Lambert & Oberwinkler, 2005; Lang *et al.*, 1998; Obara *et al.*, 2008). Indeed, astrocytes upregulate glucose transporters in order to provide energy to dying neuronal cells (Floyd & Lyeth, 2007; Scafidi *et al.*, 2009; Yi & Hazell, 2006,) suggesting that astrocytes are necessary for improvement in chronic neurodegenerative diseases energy metabolism. In summary, astrocytes are important producers of antioxidants in the normal CNS, and astrocytic production of these molecules after brain injury may enhance neuronal survival and protect astrocyte function.

Astrocytes are critical in the development and/or maintenance of blood-brain barrier characteristics (Gordon *et al.*, 2007; Koehler *et al.*, 2009). Astrocytes are arranged in non-overlapping spatial domains (Bushong *et al.*, 2002; Halassa *et al.*, 2007), but coupled to each other in a syncytial network (Haydon & Carmignoto, 2006). Since one astrocyte maintains contacts with approximately 160,000 synapses (Bushong *et al.*, 2002), this cell population is well positioned to integrate neuronal activity and link neuronal activity to the vascular network (Ransom *et al.*, 2003).

Astrocytes terminal processes are also known as “endfeet” cover 99% of the abluminal vascular surface of capillaries, intracerebral arterioles, and venules (Simard *et al.*, 2003). The extent of contact between endfeet and penetrating and pial arterioles remains unclear. Pial arterioles and arteries lying free in the subarachnoid space are not covered (Jones, 1970).

Nevertheless, much of the pial circulation is in contact with the glia-limitans, a de-facto extension of astrocytic processes (Kontos *et al.*, 1971; Xu *et al.*, 2004). This domain organization has been proposed as being the key linking element of the neuronal-(astrocyte)-vascular unit (Volterra & Meldolesi, 2005). For example, working with neocortical slices, Zonta *et al.* (Zonta *et al.*, 2003) demonstrated that electrical stimulation of neuronal processes raises intracellular Ca^{2+} levels in astrocytic endfeet and leads to a slowly developing dilatation of local intracerebral arterioles. Additionally, electrical stimulation of individual astrocytes had the same effect. Since this initial report, several investigators observed a vascular response in conjunction with an elevation of intracellular Ca^{2+} levels in astrocytic endfeet. However, these studies reported inconsistent vascular responses ranging from vasorelaxation to vasodilatation or the combination of both (Gordon *et al.*, 2007; Iadecola & Nedergaard, 2007). Mediators implicated in this mechanism are vasoactive metabolites of the cyclooxygenase or cytochrome P450 ω -hydroxylase pathways. All of these studies were performed in brain slices in which the vessels are lacking in intraluminal pressure. This might account for disparate results. In vivo analysis with two-photon laser scanning microscopy revealed that increases of astrocytic Ca^{2+} by photolysis of caged Ca^{2+} evoked a vasodilatation of cortical arterioles (Takano *et al.*, 2006). This interaction between the vessel and the endfeet appeared to be mediated by metabolites of the COX-1 pathway, because inhibitors of nitric oxide synthetase (NOS), COX-2, p450 epoxygenases, and adenosine receptor antagonists had no effect. These and other studies strongly implicate a role for astrocytes in cerebral blood flow regulation during neuronal activation (Haydon & Carmignoto, 2006).

It is important to point that some, if not all, of these astrocytic functions may likely be altered or reduced in neurodegenerative states (Rossi & Volterra, 2009). The role of astrocytes in various neurodegenerative diseases will briefly be discussed more thoroughly below, specifically looking at their involvement during the pathologic processes of Alzheimer's and Parkinson diseases, Amyotrophic Lateral Syndrome (ALS) and Multiple Sclerosis.

3. Astrocytes dysfunction in neurodegenerative diseases

Neurodegenerative diseases represent a heterogeneous group of disorders affecting the nervous system. In most instances, they affect adults, their causes are unknown, and progression is relentless. Some are genetic, but most are sporadic. They involve all parts of the nervous system, although the cerebral cortex and the basal ganglia are the most frequent loci of pathology. The historical classification of neurodegenerative diseases, based on clinical and pathological characteristics, is imperfect. New classifications are rather based on molecular determinants. Contrary to common belief, it is now recognized that neurodegenerative disorders are multisystemic, even if specific neuronal pathways are more affected than others. The death of astrocytes and specific types of neurons in neurodegenerative diseases is provoked, not by a single pathogenic factor, but rather by a cascade of multiple deleterious molecular and cellular events as described earlier.

3.1 Oxidative stress and neurodegeneration

Mitochondria are central neuronal organelles that play a vital role in neuronal life and death. Both mitochondrial dysfunction and proper function are essential components in neurodegeneration. Further elucidation of the mechanisms of interaction between

mitochondria and neuronal death will allow better description of the pathogenesis of neurodegenerative diseases and provide potential targets for therapeutic intervention.

One of the hallmarks of various neurodegenerative and neuroinflammatory disorders is oxidative stress-induced CNS damage. Similarly, the natural aging process per se is associated with increased oxidative stress (Figure 3). Such oxidative stress can damage lipids, proteins and nucleic acids of cells and power-house mitochondria causing cell death in assorted cell types including astrocytes and neurons. However, astrocytes having high levels of anti-oxidant enzymes (glutathione peroxidase, catalase, glutathione reductase, and superoxide dismutase) and antioxidants (glutathione and ascorbic acid) try to absorb reactive oxygen species ($O_2 =$, O_2^- , and $OH\cdot$) and reactive nitrogen species (NO , $ONOO^-$), maintain redox homeostasis and defend the insulted CNS (Chen & Swanson, 2003; Dringen & Hirrlinger, 2003; Wilson, 1997). In addition, astrocytes also scavenge detrimental

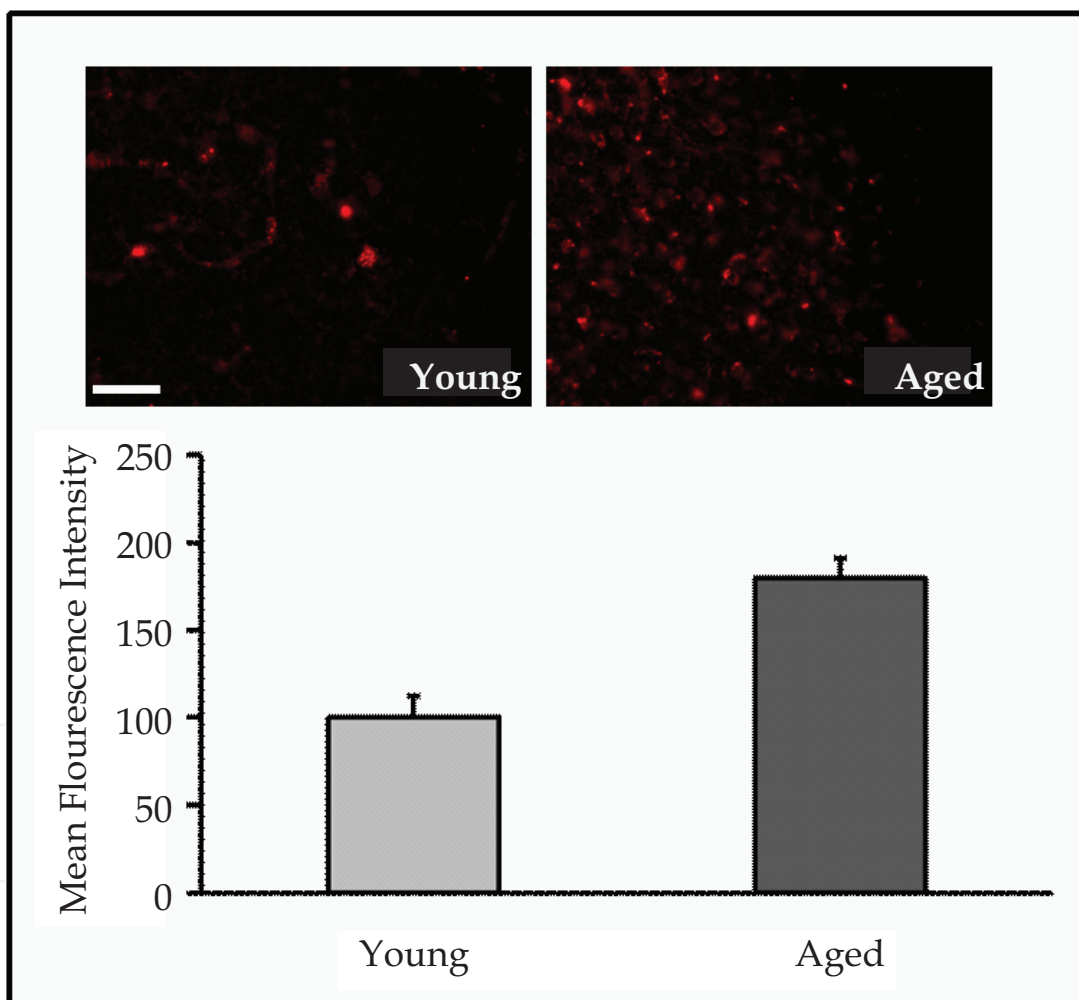


Fig. 3. Increased oxidative stress production in the normal aged brain. 3- or 18-months naïve old male mice were sacrificed and 40 μ m brain sections were stained for 4-Hydroxynonenal (4-HNE), a marker of lipid peroxidation. Mean Fluorescence intensity assessed by ImageJ program showed that aged animals, overt any pathological condition, had an increased expression of 4-HNE in cortical layers II-III, compared to young mice. 3 sections/animal for 4 animals were analyzed in each condition. Data are represented as Means \pm SEM. Scale bar, 100 μ m

molecules such as glutamate, produced during synaptic transmission through neurons (Hertz & Zielke, 2004). This is, perhaps, the most common astrocytic dysfunction that likely occurs in some neurodegenerative states.

Astrocytes react to various neurodegenerative insults rapidly, leading to vigorous astrogliosis. This reactive gliosis is associated with alteration in morphology and structure of activated astrocytes along with its functional characteristics (Eddleston & Mucke, 1993). The astrocytic processes construct a bushy network surrounding the injury site, thus secluding the affected part from the rest of the CNS area. Subsequently, astrogliosis has been implicated in the pathogenesis of a variety of chronic neurodegenerative diseases, including Alzheimer's disease (AD), Parkinson's disease, Amyotrophic Lateral Syndrome (ALS), acute traumatic brain injury, stroke, and neuroinflammatory brain diseases (Axelsson *et al.*, 2011; Ciesielska *et al.*, 2009; Garcia-Matas *et al.*, 2010; Heales *et al.*, 2004; Li *et al.*, 2011; Simpson *et al.*, 2010; Sofroniew, 2000).

3.2 Alzheimer's disease

AD is characterized clinically by cognitive loss in two or more domains, including memory, language, calculations, orientation and judgment; the loss must be of sufficient severity to cause social or occupational disability. These clinical features are the result of neuronal death and dysfunction in the cerebral cortex, entorhinal area, hippocampus, ventral striatum and basal forebrain, eventually resulting in severe dementia. Pathologically, the two hallmark findings of the disorder are neurofibrillary tangles and amyloid plaques (Veerhuis, 2011).

Senile plaques, a pathologic hallmark of Alzheimer's disease, are associated with GFAP-positive activated astrocytes (Nagele *et al.*, 2004). It is reported that in various neuropathological states, the increased GFAP expression corresponds to the severity of astroglial activation (Axelsson *et al.*, 2011; Kashon *et al.*, 2004; Notturmo *et al.*, 2009; Pelinka *et al.*, 2004; Simpson *et al.*, 2010; Toft-Hansen *et al.*, 2011).

Concerning astrocytes, recent findings suggest that they play a role in the clearance of the A β - peptide and thus in preventing plaque formation (Li *et al.*, 2011). Similarly, this peptide decreases glutamate uptake in cultured astrocytes, thus increasing oxidative stress and activation of mitogen-activated protein kinase cascades (Agostinho *et al.*, 2010; Matos *et al.*, 2008). High levels of pro-inflammatory cytokines such as interleukin 1 β , interleukin 6 and TNF α , mostly produced by reactive astrocytes, are detected in the brain of AD subjects, so the consequences of this phenomenon are unclear, also because pro-inflammatory cytokines have varied effects depending on the biological context (Veerhuis, 2011).

A previous study indicated that activated astrocytes were closely associated with amyloid plaques in the molecular layer of the cerebral cortex (Wisniewski & Wegiel, 1991). Astrocytes might be activated by human amyloid- β (A β) (DeWitt *et al.*, 1998), indicating a correlation between this protein and subsequent alterations in astrocyte function. Astrocytes also accumulate neuron-derived amyloid material resulting from local neurodegeneration. Once substantial accumulation of this debris occurs, the astrocytes themselves might undergo cell death, resulting in the formation of GFAP⁺ amyloid plaques (Nagele *et al.*, 2004). *In vitro* analyses also indicate that treatment of astrocytes with A β results in an increase in calcium-wave signaling between these cells (Haughey & Mattson, 2003). In cells expressing the familial AD presenilin 1 (*PSEN1*) mutation, calcium oscillations in astrocytes were found to occur at lower ATP and glutamate concentrations than in wild-type astrocytes (Johnston *et al.*, 2006). These data support a model in which calcium signaling between astrocytes is altered by the disease process, which might, in ways that are not fully understood, contribute to dysfunction or death of neurons.

Either prooxidant agents or amyloid beta peptide did not cause deleterious effects in the astrocytes, but the combined treatment led to oxidative stress and apoptosis *in vitro* and inflammation and degenerative traits *in vivo*. Therefore, a reduced oxidative stress defense capacity in frail aged astrocytes may contribute to neuron death by failure of astrocyte support. To preserve astrocyte function and reduce oxidative stress in old age is a new goal against AD (Aliev *et al.*, 2009a; Aliev *et al.*, 2009b; Garcia-Matas *et al.*, 2010).

3.3 Parkinson's disease

PD is the second most prevalent neurodegenerative disease, after AD. PD is estimated to affect about 1 million Americans, or about 1% of the population over 60 years of age. PD is caused by the disruption of dopaminergic neurotransmission in the basal ganglia. On pathological examination, the numbers of dopaminergic neurons in the substantia nigra are markedly reduced, and Lewy bodies (cytoplasmic inclusions) are present in the residual dopaminergic neurons (Nutt & Wooten, 2005). The focus has always been on the loss of these dopaminergic neurons and subsequent depletion of dopamine, but a role for non-neuronal cells in producing neuropathological or neuroprotective functions in PD is becoming increasingly recognized.

The studies that have been carried out to date appear to support a neuroprotective role for astrocytes in PD. From pathological examinations, an increase in the number of astrocytes as well as in GFAP expression is observed in PD, (Ciesielska *et al.*, 2009; Muramatsu *et al.*, 2003), as with other neurodegenerative disorders. The pathological evidence indirectly indicates that antioxidant pathways might contribute to this neuroprotective effect, because in control brains the density of glutathione-peroxidase-positive cells was higher in the vicinity of the dopaminergic cell groups known to be resistant to the pathological process of PD. The increase in glutathione-peroxidase-containing cells was inversely correlated with the severity of dopaminergic cell loss in the respective cell groups in patients with PD. The quantity of glutathione-peroxidase-containing cells, therefore, might be critical for a protective effect against oxidative stress (Damier *et al.*, 1993). Conversely, the presence of synuclein-positive astrocytes in pathological samples has been shown to correlate with nigral neuronal cell death (Wakabayashi *et al.*, 2000).

Nitric oxide production and glutathione depletion also appear as consistent features in human PD. The release of glutathione represents another pathway by which astrocytes might be neuroprotective in PD models. Glutathione production appears to be increased by exposure of astrocytes to nitric oxide, and the increase in glutathione release by astrocytes might increase its availability to neurons, thereby making them less susceptible to reactive nitrogen species. This pattern is consistent with the data in PD patients, in whom glutathione-containing cells are in regions with preserved dopaminergic neurons (Heales *et al.*, 2004).

Evidence regarding regulation of glutamate transporter expression and function in PD has been somewhat mixed, with downregulation of glutamate transporters being reported in some studies and upregulation being reported in others. The differences in these studies might be related to the methods by which the lesions were induced (Maragakis & Rothstein, 2004).

3.4 Amyotrophic Lateral Syndrome (ALS)

Amyotrophic Lateral Syndrome is an inexorably progressive motor neuron disease, in which both the upper motor neurons and the lower motor neurons degenerate leading to

muscle atrophy. Patients eventually experience respiratory failure, usually within three to five years from diagnosis. However, the onset of ALS may be subtle and early symptoms are frequently overlooked.

Common to familial and sporadic ALS is the loss of the astrocyte glutamate transporter EAAT2. Studies of the EAAT2 transporter in tissue from individuals with sporadic ALS showed a marked loss of up to 95% of astroglial EAAT2 protein expression and activity in affected areas of the CNS (Bristol & Rothstein, 1996). A clue to a possible mechanism for EAAT2 reduction or dysfunction was provided by the finding of aberrant EAAT2 RNA species, which has been implicated in multiple neurodegenerative diseases. The production of truncated EAAT2 protein results in reduced function, and the retention of normal EAAT2 protein within the cytoplasm (Lin *et al.*, 1998). The significance of these aberrant EAAT2 RNA species continues to be debated, however, as they have also been found in some normal controls (Flowers *et al.*, 2001; Meyer *et al.*, 1999).

In both human tissue and transgenic models of ALS, there is abundant evidence that astroglial abnormalities and physiological dysfunction precede clinical disease. These changes include reactive astrogliosis that can be seen many months before motor neuron degeneration (G85R) (Bruijn *et al.*, 1997), and loss of glutamate transport and GLT1 protein expression before the onset of clinical disease or overt motor neuron degeneration (Howland *et al.*, 2002). Similarly, increased astrocytes activation and expression of immune/inflammatory markers are hallmark of this pathology (Chiu *et al.*, 2008; Chiu *et al.*, 2009). Is the reduction in GLT1 protein in astrocytes significant? Guo and colleagues addressed this question by overexpressing the EAAT2 protein in astrocytes in the mSOD1 mouse model, and demonstrated an increase in motor neuron survival and a delay in disease onset; similar outcomes are seen with drugs that increase GLT1 expression (Guo *et al.*, 2003). This evidence indicates that EAAT2 expressed in astrocytes - and probably also glutamate- influences the timing of disease onset and motor neuron survival (Guo *et al.*, 2003). Other changes associated with ALS include increased expression of various proteins in astrocytes, including inducible nitric oxide synthase (iNOS), the copper chaperone CCS, and metallothioneins. Pathologically, early cytosolic proteinaceous aggregates have been found in spinal cord astrocytes from the entire mSOD1 mouse lines examined to date (Patel & Maragakis, 2002).

3.5 Multiple Sclerosis

Multiple Sclerosis is a chronic inflammatory demyelinating disease of the central nervous system in which glial cells play a prominent role. In murine experimental autoimmune encephalomyelitis (EAE), an established animal model of multiple sclerosis, astrocyte hypertrophy coincided with manifestation of axonal damage (Wang *et al.*, 2005). Astrocytes in multiple sclerosis plaques produce IL-6 (Okuda *et al.*, 1998), lack β -2 adrenergic receptors, and potentially serve as antigen-presenting cells (Zeinstra *et al.*, 2000b), thus facilitating T-cell invasion and activation. Repeated exposure of these astrocytes to inflammatory cytokines triggers unregulated inflammatory responses and increased noradrenalin levels, leading to focal areas of myelin and axonal damage (De Keyser *et al.*, 1999; Zeinstra *et al.*, 2000a).

Concerning the immune system, class II MHC expressing astrocytes have been shown to process and present antigens and activate both naïve and memory T cells (Nikcevich *et al.*, 1997; Soos *et al.*, 1998). In contrast, other investigators have shown that class II MHC expressing astrocytes are not capable of stimulating T-cell proliferation and instead induce

apoptosis or down-regulation of T cells (Matsumoto *et al.*, 1992; Weber *et al.*, 1994). Such a response may be beneficial for astrocyte suppression of CNS autoimmunity like in multiple sclerosis.

4. Conclusions

Astrocytes play a critical role in normal function of the mammalian nervous system. Astrocytes regulate K^+ buffering, glutamate clearance, brain antioxidant defense, close metabolic coupling with neurons, and modulation of neuronal excitability. In numerous pathological states, such as AD, PD, ALS and ME, astrocytes are involved in both exacerbation of damage and neuroprotective mechanisms. As discussed in this chapter, they support neurons in many ways, all of which are essential for repair and regeneration. Disturbances in astrocytic functions are implicated in neurodegenerative diseases pathogenesis, therefore, modulation of astrocyte functioning may prove to be an efficient therapeutic strategy in many chronic CNS disorders.

5. References

- Agostinho, P.; Cunha, R.A. & Oliveira, C. (2010). Neuroinflammation, oxidative stress and the pathogenesis of Alzheimer's disease. *Curr Pharm Des* 16, 2766-2778.
- Aliev, G.; Palacios, H.H.; Lipsitt, A.E.; Fischbach, K.; Lamb, B.T.; Obrenovich, M.E.; Morales, L.; Gasimov, E. & Bragin, V. (2009a). Nitric oxide as an initiator of brain lesions during the development of Alzheimer disease. *Neurotox Res* 16, 293-305.
- Aliev, G.; Palacios, H.H.; Walrafen, B.; Lipsitt, A.E.; Obrenovich, M.E. & Morales, L. (2009b). Brain mitochondria as a primary target in the development of treatment strategies for Alzheimer disease. *Int J Biochem Cell Biol* 41, 1989-2004.
- Anderson, C.M. & Swanson, R.A. (2000): Astrocyte glutamate transport: review of properties, regulation, and physiological functions. *Glia* 32, 1-14.
- Axelsson, M.; Malmstrom, C.; Nilsson, S.; Haghighi, S.; Rosengren, L. & Lycke, J. (2011). Glial fibrillary acidic protein: a potential biomarker for progression in multiple sclerosis. *J Neurol* 258, 882-888.
- Barreto, G.; Santos-Galindo, M.; Diz-Chaves, Y.; Pernia, O.; Carrero, P.; Azcoitia, I. & Garcia-Segura, L.M. (2009). Selective estrogen receptor modulators decrease reactive astrogliosis in the injured brain: effects of aging and prolonged depletion of ovarian hormones. *Endocrinology* 150, 5010-5015.
- Bartnik-Olson, B.L.; Oyoyo, U.; Hovda, D.A. & Sutton, R.L. (2010). Astrocyte oxidative metabolism and metabolite trafficking after fluid percussion brain injury in adult rats. *J Neurotrauma* 27, 2191-2202.
- Bentzer, P.; Davidsson, H. & Grande, P.O. (2000). Microdialysis-based long-term measurements of energy-related metabolites in the rat brain following a fluid percussion trauma. *J Neurotrauma* 17, 441-447.
- Beschorner, R.; Adjodah, D.; Schwab, J.M.; Mittelbronn, M.; Pedal, I.; Mattern R.; Schluesener, H.J. & Meyermann, R. (2000). Long-term expression of heme oxygenase-1 (HO-1, HSP-32) following focal cerebral infarctions and traumatic brain injury in humans. *Acta Neuropathol* 100, 377-384.
- Bristol, L.A. & Rothstein, J.D. (1996). Glutamate transporter gene expression in amyotrophic lateral sclerosis motor cortex. *Ann Neurol* 39, 676-679.

- Bruijn, L.I.; Becher, M.W.; Lee, M.K.; Anderson, K.L.; Jenkins, N.A.; Copeland, N.G.; Sisodia, S.S.; Rothstein, J.D.; Borchelt, D.R.; Price, D.L. & Cleveland, D.W. (1997). ALS-linked SOD1 mutant G85R mediates damage to astrocytes and promotes rapidly progressive disease with SOD1-containing inclusions. *Neuron* 18, 327-338.
- Bushong, E.A.; Martone, M.E.; Jones, Y.Z. & Ellisman, M.H. (2002). Protoplasmic astrocytes in CA1 stratum radiatum occupy separate anatomical domains. *J Neurosci* 22, 183-192.
- Caesar, K.; Hashemi, P.; Douhou, A.; Bonvento, G.; Boutelle, M.G.; Walls, A.B. & Lauritzen, M. (2008). Glutamate receptor-dependent increments in lactate, glucose and oxygen metabolism evoked in rat cerebellum in vivo. *J Physiol* 586, 1337-1349.
- Ciesielska, A.; Joniec, I.; Kurkowska-Jastrzebska, I.; Cudna, A.; Przybylkowski, A.; Czlonkowska, A. & Czlonkowski, A. (2009). The impact of age and gender on the striatal astrocytes activation in murine model of Parkinson's disease. *Inflamm Res* 58, 747-753.
- Cliffe, E.E. & Waley, S.G. (1961). The mechanism of the glyoxalase I reaction, and the effect of ophthalmic acid as an inhibitor. *Biochem J* 79, 475-482.
- Chen, K.; Gunter, K. & Maines, M.D. (2000). Neurons overexpressing heme oxygenase-1 resist oxidative stress-mediated cell death. *J Neurochem* 75, 304-313.
- Chen, Y. & Swanson, R.A. (2003). Astrocytes and brain injury. *J Cereb Blood Flow Metab* 23, 137-149.
- Chiu, I.M.; Chen, A.; Zheng, Y.; Kosaras, B.; Tsiftoglou, S.A.; Vartanian, T.K.; Brown, R.H.Jr. & Carroll, M.C. (2008). T lymphocytes potentiate endogenous neuroprotective inflammation in a mouse model of ALS. *Proc Natl Acad Sci U S A* 105, 17913-17918.
- Chiu, I.M.; Phatnani, H.; Kuligowski, M.; Tapia, J.C.; Carrasco, M.A.; Zhang, M.; Maniatis, T. & Carroll, M.C. (2009). Activation of innate and humoral immunity in the peripheral nervous system of ALS transgenic mice. *Proc Natl Acad Sci U S A* 106, 20960-20965.
- Damier, P.; Hirsch, E.C.; Zhang, P.; Agid, Y. & Javoy-Agid, F. (1993). Glutathione peroxidase, glial cells and Parkinson's disease. *Neuroscience* 52, 1-6.
- De Keyser, J.; Wilczak, N.; Leta, R. & Streetland, C. (1999). Astrocytes in multiple sclerosis lack beta-2 adrenergic receptors. *Neurology* 53, 1628-1633.
- DeWitt, D.A.; Perry, G.; Cohen, M.; Doller, C. & Silver, J. (1998). Astrocytes regulate microglial phagocytosis of senile plaque cores of Alzheimer's disease. *Exp Neurol* 149, 329-340.
- Dringen, R. (2000): Metabolism and functions of glutathione in brain. *Prog Neurobiol* 62, 649-671.
- Dringen, R.; Gutterer, J.M. & Hirrlinger, J. (2000). Glutathione metabolism in brain metabolic interaction between astrocytes and neurons in the defense against reactive oxygen species. *Eur J Biochem* 267, 4912-4916.
- Dringen, R. & Hirrlinger, J. (2003). Glutathione pathways in the brain. *Biol Chem* 384, 505-516.
- Eddleston, M. & Mucke, L. (1993): Molecular profile of reactive astrocytes--Implications for their role in neurologic disease. *Neuroscience* 54, 15-36.
- Espada, S.; Ortega, F.; Molina-Jijon, E.; Rojo, A.I.; Perez-Sen, R.; Pedraza-Chaverri, J.; Miras-Portugal, M.T. & Cuadrado, A. (2010). The purinergic P2Y(13) receptor activates the Nrf2/HO-1 axis and protects against oxidative stress-induced neuronal death. *Free Radic Biol Med* 49, 416-426.

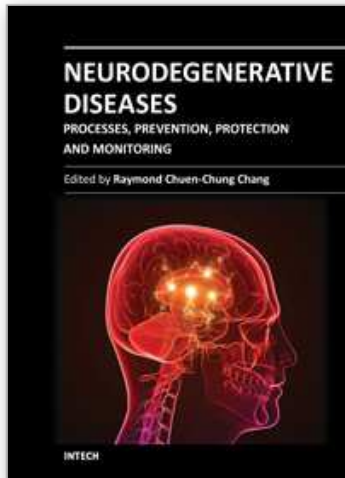
- Fernandez-Checa, J.C.; Fernandez, A.; Morales, A.; Mari, M.; Garcia-Ruiz, C. & Colell, A. (2010). Oxidative stress and altered mitochondrial function in neurodegenerative diseases: lessons from mouse models. *CNS Neurol Disord Drug Targets* 9, 439-454.
- Flowers, J.M.; Powell, J.F.; Leigh, P.N.; Andersen, P. & Shaw, C.E. (2001). Intron 7 retention and exon 9 skipping EAAT2 mRNA variants are not associated with amyotrophic lateral sclerosis. *Ann Neurol* 49, 643-649.
- Floyd, C.L. & Lyeth, B.G. (2007). Astroglia: important mediators of traumatic brain injury. *Prog Brain Res* 161, 61-79.
- Garcia-Matas, S.; de Vera, N.; Aznar, A.O.; Marimon, J.M.; Adell, A.; Planas, A.M.; Cristofol, R. & Sanfeliu, C. (2010). In vitro and in vivo activation of astrocytes by amyloid-beta is potentiated by pro-oxidant agents. *J Alzheimers Dis* 20, 229-245.
- Giordano, G.; Kavanagh, T.J. & Costa, L.G. (2009). Mouse cerebellar astrocytes protect cerebellar granule neurons against toxicity of the polybrominated diphenyl ether (PBDE) mixture DE-71. *Neurotoxicology* 30, 326-329.
- Gordon, G.R.; Mulligan, S.J. & MacVicar, B.A. (2007). Astrocyte control of the cerebrovasculature. *Glia* 55, 1214-1221.
- Guo, H.; Lai, L.; Butchbach, M.E.; Stockinger, M.P.; Shan, X.; Bishop, G.A. & Lin, C.L. (2003). Increased expression of the glial glutamate transporter EAAT2 modulates excitotoxicity and delays the onset but not the outcome of ALS in mice. *Hum Mol Genet* 12, 2519-2532.
- Halassa, M.M.; Fellin, T.; Takano, H.; Dong, J.H. & Haydon, P.G. (2007). Synaptic islands defined by the territory of a single astrocyte. *J Neurosci* 27, 6473-6477.
- Haughey, N.J. & Mattson, M.P. (2003). Alzheimer's amyloid beta-peptide enhances ATP/gap junction-mediated calcium-wave propagation in astrocytes. *Neuromolecular Med* 3, 173-180.
- Haydon, P.G. & Carmignoto, G. (2006). Astrocyte control of synaptic transmission and neurovascular coupling. *Physiol Rev* 86, 1009-1031.
- Heales, S.J.; Lam, A.A.; Duncan, A.J. & Land, J.M. (2004). Neurodegeneration or neuroprotection: the pivotal role of astrocytes. *Neurochem Res* 29, 513-519.
- Hertz, L. & Zielke, H.R. (2004). Astrocytic control of glutamatergic activity: astrocytes as stars of the show. *Trends Neurosci* 27, 735-743.
- Howland, D.S.; Liu, J.; She, Y.; Goad, B.; Maragakis, N.J.; Kim, B.; Erickson, J.; Kulik, J.; DeVito, L.; Psaltis, G.; DeGennaro, L.J.; Cleveland, D.W. & Rothstein, J.D. (2002). Focal loss of the glutamate transporter EAAT2 in a transgenic rat model of SOD1 mutant-mediated amyotrophic lateral sclerosis (ALS). *Proc Natl Acad Sci U S A* 99, 1604-1609.
- Iadecola, C. & Nedergaard, M. (2007). Glial regulation of the cerebral microvasculature. *Nat Neurosci* 10, 1369-1376.
- Imuta, N.; Hori, O.; Kitao, Y.; Tabata, Y.; Yoshimoto, T.; Matsuyama, T. & Ogawa, S. (2007). Hypoxia-mediated induction of heme oxygenase type I and carbon monoxide release from astrocytes protects nearby cerebral neurons from hypoxia-mediated apoptosis. *Antioxid Redox Signal* 9, 543-552.
- Jayakumar, A.R. & Norenberg, M.D. (2010). The Na-K-Cl Co-transporter in astrocyte swelling. *Metab Brain Dis* 25, 31-38.
- Johnston, J.M.; Burnett, P.; Thomas, A.P. & Tezapsidis, N. (2006). Calcium oscillations in type-1 astrocytes, the effect of a presenilin 1 (PS1) mutation. *Neurosci Lett* 395, 159-164.

- Jones, E.G. (1970). On the mode of entry of blood vessels into the cerebral cortex. *J Anat* 106, 507-520.
- Kashon, M.L.; Ross, G.W.; O'Callaghan, J.P.; Miller, D.B.; Petrovitch, H.; Burchfiel, C.M.; Sharp, D.S.; Markesbery, W.R.; Davis, D.G.; Hardman, J.; Nelson, J. & White, L.R. (2004). Associations of cortical astrogliosis with cognitive performance and dementia status. *J Alzheimers Dis* 6, 595-604; discussion 673-581.
- Koehler, R.C.; Roman, R.J. & Harder, D.R. (2009). Astrocytes and the regulation of cerebral blood flow. *Trends Neurosci* 32, 160-169.
- Kontos, H.A.; Raper, A.J. & Patterson, J.L. Jr. (1971). Mechanisms of action of CO₂ on pial precapillary vessels. *Eur Neurol* 6, 114-118.
- Ku, B.M.; Joo, Y.; Mun, J.; Roh, G.S.; Kang, S.S.; Cho, G.J.; Choi, W.S. & Kim, H.J. (2006). Heme oxygenase protects hippocampal neurons from ethanol-induced neurotoxicity. *Neurosci Lett* 405, 168-171.
- Lambert, S. & Oberwinkler, J. (2005). Characterization of a proton-activated, outwardly rectifying anion channel. *J Physiol* 567, 191-213.
- Lang, F.; Busch, G.L.; Ritter, M.; Volkl, H.; Waldegger, S.; Gulbins, E. & Haussinger, D. (1998). Functional significance of cell volume regulatory mechanisms. *Physiol Rev* 78, 247-306.
- Le, W.D.; Xie, W.J. & Appel, S.H. (1999). Protective role of heme oxygenase-1 in oxidative stress-induced neuronal injury. *J Neurosci Res* 56, 652-658.
- Lee, C.K.; Weindruch, R. & Prolla, T.A. (2000). Gene-expression profile of the ageing brain in mice. *Nat Genet* 25, 294-297.
- Li, C.; Zhao, R.; Gao, K.; Wei, Z.; Yin, M.Y.; Lau, L.T.; Chui, D. & Hoi Yu, A.C. (2011). Astrocytes: implications for neuroinflammatory pathogenesis of Alzheimer's disease. *Curr Alzheimer Res* 8, 67-80.
- Lin, C.L.; Bristol, L.A.; Jin, L.; Dykes-Hoberg, M.; Crawford, T.; Clawson, L. & Rothstein, J.D. (1998). Aberrant RNA processing in a neurodegenerative disease: the cause for absent EAAT2, a glutamate transporter, in amyotrophic lateral sclerosis. *Neuron* 20, 589-602.
- Lindenau, J.; Noack, H.; Possel, H.; Asayama, K. & Wolf, G. (2000). Cellular distribution of superoxide dismutases in the rat CNS. *Glia* 29, 25-34.
- Magistretti, P.J. (2006). Neuron-glia metabolic coupling and plasticity. *J Exp Biol* 209, 2304-2311.
- Magistretti, P.J. & Pellerin, L. (1999). Cellular mechanisms of brain energy metabolism and their relevance to functional brain imaging. *Philos Trans R Soc Lond B Biol Sci* 354, 1155-1163.
- Maragakis, N.J. & Rothstein, J.D. (2004). Glutamate transporters: animal models to neurologic disease. *Neurobiol Dis* 15, 461-473.
- Matos, M.; Augusto E.; Oliveira, C.R. & Agostinho, P. (2008). Amyloid-beta peptide decreases glutamate uptake in cultured astrocytes: involvement of oxidative stress and mitogen-activated protein kinase cascades. *Neuroscience* 156, 898-910.
- Matsumoto, Y.; Ohmori, K. & Fujiwara, M. (1992). Immune regulation by brain cells in the central nervous system: microglia but not astrocytes present myelin basic protein to encephalitogenic T cells under in vivo-mimicking conditions. *Immunology* 76, 209-216.
- Meyer, T.; Fromm, A.; Munch, C.; Schwalenstocker, B.; Fray, A.E.; Ince, P.G.; Stamm, S.; Gron, G.; Ludolph, A.C. & Shaw, P.J. (1999). The RNA of the glutamate transporter

- EAAT2 is variably spliced in amyotrophic lateral sclerosis and normal individuals. *J Neurol Sci* 170, 45-50.
- Muramatsu, Y.; Kurosaki, R.; Watanabe, H.; Michimata, M.; Matsubara, M.; Imai, Y. & Araki, T. (2003). Cerebral alterations in a MPTP-mouse model of Parkinson's disease--an immunocytochemical study. *J Neural Transm* 110, 1129-1144.
- Nagele, R.G.; Wegiel, J.; Venkataraman, V.; Imaki, H. & Wang, K.C. (2004). Contribution of glial cells to the development of amyloid plaques in Alzheimer's disease. *Neurobiol Aging* 25, 663-674.
- Nedergaard, M. & Dirnagl, U. (2005). Role of glial cells in cerebral ischemia. *Glia* 50, 281-286.
- Nichols, N.R. (1999). Glial responses to steroids as markers of brain aging. *J Neurobiol* 40, 585-601.
- Nikcevich, K.M.; Gordon, K.B.; Tan, L.; Hurst, S.D.; Kroepfl, J.F.; Gardinier, M.; Barrett, T.A. & Miller, S.D. (1997). IFN-gamma-activated primary murine astrocytes express B7 costimulatory molecules and prime naive antigen-specific T cells. *J Immunol* 158, 614-621.
- Notturmo, F.; Capasso, M.; DeLauretis, A.; Carpo, M. & Uncini, A. (2009). Glial fibrillary acidic protein as a marker of axonal damage in chronic neuropathies. *Muscle Nerve* 40, 50-54.
- Nutt, J.G. & Wooten, G.F. (2005). Clinical practice. Diagnosis and initial management of Parkinson's disease. *N Engl J Med* 353, 1021-1027.
- Obara, M.; Szeliga, M. & Albrecht, J. (2008). Regulation of pH in the mammalian central nervous system under normal and pathological conditions: facts and hypotheses. *Neurochem Int* 52, 905-919.
- Okuda, Y.; Sakoda, S.; Bernard, C.C. & Yanagihara, T. (1998). The development of autoimmune encephalomyelitis provoked by myelin oligodendrocyte glycoprotein is associated with an upregulation of both proinflammatory and immunoregulatory cytokines in the central nervous system. *J Interferon Cytokine Res* 18, 415-421.
- Patel, S.A. & Maragakis, N.J. (2002). Amyotrophic lateral sclerosis: pathogenesis, differential diagnoses, and potential interventions. *J Spinal Cord Med* 25, 262-273.
- Peinado, M.A.; Quesada, A.; Pedrosa, J.A.; Torres, M.I.; Martinez, M.; Esteban, F.J.; Del Moral, M.L.; Hernandez, R.; Rodrigo, J. & Peinado, J.M. (1998). Quantitative and ultrastructural changes in glia and pericytes in the parietal cortex of the aging rat. *Microsc Res Tech* 43, 34-42.
- Pelinka, L.E.; Kroepfl, A.; Leixnering, M.; Buchinger, W.; Raabe, A. & Redl, H. (2004). GFAP versus S100B in serum after traumatic brain injury: relationship to brain damage and outcome. *J Neurotrauma* 21, 1553-1561.
- Pilegaard, K. & Ladefoged, O. (1996). Total number of astrocytes in the molecular layer of the dentate gyrus of rats at different ages. *Anal Quant Cytol Histol* 18, 279-285.
- Ransom, B.; Behar, T. & Nedergaard, M. (2003). New roles for astrocytes (stars at last). *Trends Neurosci* 26, 520-522.
- Ridet, J.L.; Malhotra, S.K.; Privat, A. & Gage, F.H. (1997). Reactive astrocytes: cellular and molecular cues to biological function. *Trends Neurosci* 20, 570-577.
- Romera, C.; Hurtado, O.; Botella, S.H.; Lizasoain, I.; Cardenas, A.; Fernandez-Tome, P.; Leza, J.C.; Lorenzo, P. & Moro, M.A. (2004). In vitro ischemic tolerance involves upregulation of glutamate transport partly mediated by the TACE/ADAM17-tumor necrosis factor-alpha pathway. *J Neurosci* 24, 1350-1357.

- Rossi, D. & Volterra, A. (2009). Astrocytic dysfunction: insights on the role in neurodegeneration. *Brain Res Bull* 80, 224-232.
- Rozovsky, I.; Finch, C.E. & Morgan, T.E. (1998). Age-related activation of microglia and astrocytes: in vitro studies show persistent phenotypes of aging, increased proliferation, and resistance to down-regulation. *Neurobiol Aging* 19, 97-103.
- Salminen, A.; Ojala, J.; Kaarniranta, K.; Haapasalo, A.; Hiltunen, M. & Soininen, H. (2011). Astrocytes in the aging brain express characteristics of senescence-associated secretory phenotype. *Eur J Neurosci* 34, 3-11.
- Scafidi, S.; O'Brien, J.; Hopkins, I.; Robertson, C.; Fiskum, G. & McKenna, M. (2009). Delayed cerebral oxidative glucose metabolism after traumatic brain injury in young rats. *J Neurochem* 109 Suppl 1, 189-197.
- Schipper, H.M.; Cisse, S. & Stopa, E.G. (1995). Expression of heme oxygenase-1 in the senescent and Alzheimer-diseased brain. *Ann Neurol* 37, 758-768.
- Schipper, H.M.; Liberman, A. & Stopa, E.G. (1998). Neural heme oxygenase-1 expression in idiopathic Parkinson's disease. *Exp Neurol* 150, 60-68.
- Schousboe, A. & Waagepetersen, H.S. (2006). Glial modulation of GABAergic and glutamatergic neurotransmission. *Curr Top Med Chem* 6, 929-934.
- Serviddio, G.; Romano, A.D.; Cassano, T.; Bellanti, F.; Altomare, E. & Vendemiale, G. (2011). Principles and Therapeutic Relevance for Targeting Mitochondria in Aging and Neurodegenerative Diseases. *Curr Pharm Des*.
- Shibata, N. & Kobayashi, M. (2008). [The role for oxidative stress in neurodegenerative diseases]. *Brain Nerve* 60, 157-170.
- Simard, M.; Arcuino, G.; Takano, T.; Liu, Q.S. & Nedergaard, M. (2003). Signaling at the gliovascular interface. *J Neurosci* 23, 9254-9262.
- Simpson, J.E.; Ince, P.G.; Lace, G.; Forster, G.; Shaw, P.J.; Matthews, F.; Savva, G.; Brayne, C. & Wharton, S.B. (2010). Astrocyte phenotype in relation to Alzheimer-type pathology in the ageing brain. *Neurobiol Aging* 31, 578-590.
- Sims NR, Nilsson M & Muyderman H. (2004). Mitochondrial glutathione: a modulator of brain cell death. *J Bioenerg Biomembr* 36, 329-333.
- Slemmer, J.E.; Shacka, J.J.; Sweeney, M.I. & Weber, J.T. (2008). Antioxidants and free radical scavengers for the treatment of stroke, traumatic brain injury and aging. *Curr Med Chem* 15, 404-414.
- Sofroniew, M.V. (2000). Astrocyte failure as a cause of CNS dysfunction. *Mol Psychiatry* 5, 230-232.
- Sofroniew, M.V. (2009). Molecular dissection of reactive astrogliosis and glial scar formation. *Trends Neurosci* 32, 638-647.
- Sofroniew, M.V. & Vinters, H.V. (2010). Astrocytes: biology and pathology. *Acta Neuropathol* 119, 7-35.
- Soos, J.M.; Morrow, J.; Ashley, T.A.; Szente, B.E.; Bikoff, E.K. & Zamvil, S.S. (1998). Astrocytes express elements of the class II endocytic pathway and process central nervous system autoantigen for presentation to encephalitogenic T cells. *J Immunol* 161, 5959-5966.
- Swanson, R.A.; Ying, W. & Kauppinen, T.M. (2004). Astrocyte influences on ischemic neuronal death. *Curr Mol Med* 4, 193-205.
- Takano, T.; Tian, G.F.; Peng, W.; Lou, N.; Libionka, W.; Han, X. & Nedergaard, M. (2006). Astrocyte-mediated control of cerebral blood flow. *Nat Neurosci* 9, 260-267.

- Takeda, A.; Perry, G.; Abraham, N.G.; Dwyer, B.E.; Kutty, R.K.; Laitinen, J.T.; Petersen, R.B. & Smith, M.A. (2000). Overexpression of heme oxygenase in neuronal cells, the possible interaction with Tau. *J Biol Chem* 275, 5395-5399.
- Tilleux, S. & Hermans, E. (2007). Neuroinflammation and regulation of glial glutamate uptake in neurological disorders. *J Neurosci Res* 85, 2059-2070.
- Toft-Hansen, H.; Fuchtbauer, L. & Owens, T. (2011). Inhibition of reactive astrocytosis in established experimental autoimmune encephalomyelitis favors infiltration by myeloid cells over T cells and enhances severity of disease. *Glia* 59, 166-176.
- Veerhuis, R. (2011). Histological and direct evidence for the role of complement in the neuroinflammation of AD. *Curr Alzheimer Res* 8, 34-58.
- Volterra, A. & Meldolesi, J. (2005). Astrocytes, from brain glue to communication elements: the revolution continues. *Nat Rev Neurosci* 6, 626-640.
- Wakabayashi, K.; Hayashi, S.; Yoshimoto, M.; Kudo, H. & Takahashi, H. (2000). NACP/alpha-synuclein-positive filamentous inclusions in astrocytes and oligodendrocytes of Parkinson's disease brains. *Acta Neuropathol* 99, 14-20.
- Wang, D.; Ayers, M.M.; Catmull, D.V.; Hazelwood, L.J.; Bernard, C.C. & Orian, J.M. (2005). Astrocyte-associated axonal damage in pre-onset stages of experimental autoimmune encephalomyelitis. *Glia* 51, 235-240.
- Weber, F.; Meinl, E.; Aloisi, F.; Nevinny-Stickel, C.; Albert, E.; Wekerle, H. & Hohlfeld, R. (1994). Human astrocytes are only partially competent antigen presenting cells. Possible implications for lesion development in multiple sclerosis. *Brain* 117 (Pt 1), 59-69.
- Wilson, J.X. (1997). Antioxidant defense of the brain: a role for astrocytes. *Can J Physiol Pharmacol* 75, 1149-1163.
- Wisniewski, H.M. & Wegiel, J. (1991). Spatial relationships between astrocytes and classical plaque components. *Neurobiol Aging* 12, 593-600.
- Xu, H.L.; Koenig, H.M.; Ye, S.; Feinstein, D.L. & Pelligrino, D.A. (2004). Influence of the glia limitans on pial arteriolar relaxation in the rat. *Am J Physiol Heart Circ Physiol* 287, H331-339.
- Yi, J.H. & Hazell, A.S. (2006). Excitotoxic mechanisms and the role of astrocytic glutamate transporters in traumatic brain injury. *Neurochem Int* 48, 394-403.
- Zeier, Z.; Madorsky, I.; Xu, Y.; Ogle, W.O.; Notterpek, L. & Foster, T.C. (2011). Gene expression in the hippocampus: regionally specific effects of aging and caloric restriction. *Mech Ageing Dev* 132, 8-19.
- Zeinstra, E.; Wilczak, N. & De Keyser, J. (2000a). [3H]dihydroalprenolol binding to beta adrenergic receptors in multiple sclerosis brain. *Neurosci Lett* 289, 75-77.
- Zeinstra, E.; Wilczak, N.; Streefland, C. & De Keyser, J. (2000b). Astrocytes in chronic active multiple sclerosis plaques express MHC class II molecules. *Neuroreport* 11, 89-91.
- Zonta, M.; Angulo, M.C.; Gobbo, S.; Rosengarten, B.; Hossmann, K.A.; Pozzan, T. & Carmignoto, G. (2003). Neuron-to-astrocyte signaling is central to the dynamic control of brain microcirculation. *Nat Neurosci* 6, 43-50.
- Zou, J.; Wang, Y.X.; Dou, F.F.; Lu, H.Z.; Ma, Z.W.; Lu, P.H. & Xu, X.M. (2010). Glutamine synthetase down-regulation reduces astrocyte protection against glutamate excitotoxicity to neurons. *Neurochem Int* 56, 577-584.



Neurodegenerative Diseases - Processes, Prevention, Protection and Monitoring

Edited by Dr Raymond Chuen-Chung Chang

ISBN 978-953-307-485-6

Hard cover, 558 pages

Publisher InTech

Published online 09, December, 2011

Published in print edition December, 2011

Neurodegenerative Diseases - Processes, Prevention, Protection and Monitoring focuses on biological mechanisms, prevention, neuroprotection and even monitoring of disease progression. This book emphasizes the general biological processes of neurodegeneration in different neurodegenerative diseases. Although the primary etiology for different neurodegenerative diseases is different, there is a high level of similarity in the disease processes. The first three sections introduce how toxic proteins, intracellular calcium and oxidative stress affect different biological signaling pathways or molecular machineries to inform neurons to undergo degeneration. A section discusses how neighboring glial cells modulate or promote neurodegeneration. In the next section an evaluation is given of how hormonal and metabolic control modulate disease progression, which is followed by a section exploring some preventive methods using natural products and new pharmacological targets. We also explore how medical devices facilitate patient monitoring. This book is suitable for different readers: college students can use it as a textbook; researchers in academic institutions and pharmaceutical companies can take it as updated research information; health care professionals can take it as a reference book, even patients' families, relatives and friends can take it as a good basis to understand neurodegenerative diseases.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

George E. Barreto, Janneth Gonzalez, Francisco Capani and Ludis Morales (2011). Role of Astrocytes in Neurodegenerative Diseases, Neurodegenerative Diseases - Processes, Prevention, Protection and Monitoring, Dr Raymond Chuen-Chung Chang (Ed.), ISBN: 978-953-307-485-6, InTech, Available from: <http://www.intechopen.com/books/neurodegenerative-diseases-processes-prevention-protection-and-monitoring/role-of-astrocytes-in-neurodegenerative-diseases>

INTECH
open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

www.intechopen.com

www.intechopen.com

IntechOpen

IntechOpen

© 2011 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen