

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

4,800

Open access books available

122,000

International authors and editors

135M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.

For more information visit www.intechopen.com



Neoplasia in IBD

Joel Pekow and Marc Bissonnette

*Department of Medicine, Section of Gastroenterology, University of Chicago
USA*

1. Introduction

Longstanding inflammation of the colonic mucosa places patients with ulcerative colitis (UC) at increased risk for the development of colon cancer. As such, there has been significant research over the last 20 years into efforts to understand the natural history of neoplastic lesions in UC in order to modify this risk. This chapter will focus on the epidemiology of neoplasia in UC, the biology of IBD-associated cancer, outcomes after a diagnosis of a dysplastic lesion, as well as strategies for surveillance and chemoprevention.

2. Epidemiology

The majority of studies examining the development of cancer in IBD have demonstrated an increased risk for neoplasia in patients with long-standing UC (1-6). A 2001 meta-analysis involving 116 studies and over 50,000 patients calculated the cumulative risk of CRC in UC as 8.3% at 20 years and 18.4% at 30 years (7). However, two recent population based studies, one from Denmark and the other from Olmstead County in Minnesota, did not find a significant increase in risk (3, 8). The discrepancy in results between these more recent studies and older analyses may be secondary to the effects of newer, more effective anti-inflammatory agents for IBD or the implementation of surveillance programs and removal of colons with dysplastic lesions prior to the development of cancer.

3. Risk factors

It is postulated that cancer develops in patients with IBD secondary to prolonged inflammation. The evidence to support inflammation driving neoplastic transformation stems from studies demonstrating that patients with longer disease duration, extensive colitis, and uncontrolled inflammation are at increased risk for neoplastic changes. Risk of colorectal cancer (CRC) development rises with increased interval from diagnosis of IBD-associated colitis (2, 9). In fact, CRC is uncommon in patients who have had colitis for less than 7 years, and more commonly develops in patients who are diagnosed with IBD at a younger age (5, 7). Several studies have also demonstrated that cancer develops more frequently in those with an increased extent of colitis (2, 4). A Swedish population-based study using barium enemas quantified this risk by standardized incidence ratio as 1.7 for individuals with proctitis, 2.8 for those with left-sided colitis, and 14.8 for those with pancolitis (2).

Two recent publications have established that severity of inflammation is associated with an increased risk for cancer in IBD. In a retrospective cohort from the St. Mark's hospital,

severity of inflammation by histology was significantly associated with neoplasia in patients with extensive colitis (10). Interestingly, in this multivariate analysis only histologic inflammatory activity and not endoscopic inflammation was associated with neoplasia. A second retrospective study from Mt. Sinai hospital in New York confirmed severity of inflammation over time as a risk for neoplasia (11).

Two other well-described risk factors for neoplastic development in IBD include a family history of colorectal cancer and a history of primary sclerosing cholangitis (PSC). Several retrospective analyses have reported that patients with IBD who develop neoplasia have an odds ratio between 2.3 and 5.0 for having a family history of CRC (5, 12-14). A large population-based cohort also demonstrated a relative risk for the development of neoplasia of 2.5 for patients with a family history of colon cancer (15). As in the case of sporadic colon cancer with positive family history, this association was stronger for patients with a first-degree relative with CRC less than 50 years of age (15). Potentially, the most significant risk factor for neoplasia in patients with UC is a concomitant diagnosis of PSC. Although the reported frequency of neoplastic changes in this population varies among studies, patients with UC and PSC have consistently demonstrated a markedly increased risk for the development of both dysplasia and colon cancer compared to patients with UC without PSC (5, 16-20). The overall incidence of CRC in patients with UC and PSC was between 16% and 25% in a Swedish population-based study after 10 years of disease duration (17). This risk of neoplastic changes in UC patients with PSC has also been noted to occur earlier in the disease course than UC patients without PSC.

4. Definition of dysplasia

In UC, the term dysplasia is defined as neoplastic changes confined to the colonic epithelium. Tissue that is positive for dysplasia is most commonly identified as either low grade (LGD) or high grade (HGD) (21). Dysplasia is also characterized based on its endoscopic appearance, and outcomes of progression to cancer are associated with this endoscopic classification. Historically, flat dysplasia has been defined as dysplasia identified only by histological and not endoscopic features. However, recent studies have demonstrated that most lesions classified as flat dysplasia were obtained from targeted biopsies of visible lesions (22, 23). Raised lesions that are not endoscopically resectable are termed dysplasia associated mass or lesion or DALMs. The term ALM (adenoma-like mass) refers to a raised, endoscopically resectable lesion that resembles a sporadic adenoma by endoscopic and histological characteristics.

5. Biology of IBD-associated cancer

Although initiating mechanisms of carcinogenesis in IBD remain unknown, neoplastic lesions likely result from a combination of genetic alterations and inflammatory mediators that activate cell-signaling pathways. These pathways in turn promote deregulations in growth and apoptosis. Several molecular changes occurring in IBD-CRC are similar to those seen in sporadic CRC. In contrast to solitary lesions in sporadic colon cancer, however, neoplastic lesions in IBD are often multifocal. This finding likely reflects the widespread field defects throughout the UC involved mucosa that increase the risk for neoplastic changes. Moreover, expression changes in coding and non-coding (microRNA) genes that

are seen in malignant transformation, also occur in chronic UC, further supporting this hypothesis (24-26).

Genomic instability characterized by either chromosomal instability or microsatellite instability occurs in both sporadic and IBD-associated CRC. In fact, frequencies of these genetic abnormalities (chromosomal instability - 85%, microsatellite instability - 15%) are similar in IBD-CRC and sporadic CRC (27-30).

Genetic changes in the tumor suppressor, p53, are believed to play an important role in the development of IBD-associated neoplasia. Loss of heterozygosity and p53 mutations have both been reported in colons with IBD-associated neoplasia (31-33). It is believed that changes in p53 may occur prior to the development of dysplastic lesions in 'at risk' mucosa (32). Moreover, reactivity of p53 antibodies increase with histologic progression from UC patients without dysplasia to those with dysplasia and CRC (34). Positive p53 immunostaining can also occur prior to the development of dysplasia in chronic UC mucosa (35, 36).

The WNT pathway is deregulated in IBD-associated cancer development as occurs in sporadic colorectal carcinogenesis. Similar to genetic changes in p53, it appears that up-regulation of WNT signaling occurs early in UC-associated neoplastic progression (37). In addition to overexpression of proteins in the WNT signaling cascade, hypermethylation of WNT-suppressor genes in this pathway occur during neoplastic development in IBD (38). Such methylations could lead to silencing of tumor suppressor genes. In contrast to sporadic colon cancer, however, it appears that APC loss of function mutations play a less significant role in initiating WNT signaling (39-41). Increased mutations in the oncogene, *K-ras*, have also been described in IBD-associated colon cancer (31). However, the timing of *K-ras* mutations in neoplastic progression needs to be clarified in larger studies.

In addition to genetic changes, previous studies in animal models of ulcerative colitis and colitis-associated colon cancer have demonstrated involvement of other key signaling pathways including the vitamin D receptor, NF κ B, transforming growth factor beta (TGF β), cyclooxygenase-2 (COX2), toll-like receptor-4 (TLR4), and the epidermal growth factor receptor (EGFR). Several mouse studies have shown that active vitamin D or its analogues inhibit progression in murine models of inflammation-associated colitis (42, 43). Furthermore, one retrospective analysis identified decreased expression of VDR in IBD-associated dysplastic lesions (44). NF κ B controls a vast array of functions and is a master regulator of many pro-inflammatory cytokines including TNF- α and IL-1 β . NF κ B overexpression is known to contribute to both inflammation and malignant transformation in several cancers (45). NF κ B has also been demonstrated to contribute to malignant transformation in a mouse model of inflammation-associated cancer (46). In the study by Greten et al, NF κ B in epithelial cells was essential for survival signals, allowing mutant clones to expand, whereas NF κ B in stromal cells increased cytokines and growth factors required for tumor growth (46). Furthermore, NF κ B mediates TNF- α activation of cytidine deaminase in human colonic epithelial cells and colitis-associated cancers. Activation induced cytidine deaminase plays a critical role in physiological antibody diversification, but also contributes to malignant lymphocytic transformation (47). In addition to NF κ B up-regulation, TLR4 overexpression occurs in colitis-associated colon cancer that enhances Cox-2 expression via an EGFR-dependent mechanism (48). Recent studies from our laboratory have demonstrated that EGFR signals were required for Cox-2 up-regulation in this model (49).

6. Surveillance for IBD-associated neoplasia

There are no randomized controlled trials investigating the mortality benefit of surveillance colonoscopy in patients with UC. The best evidence to support routine endoscopic surveillance comes from retrospective case-controlled studies. In a retrospective analysis of patients with CRC, Choi et al. reported that patients who underwent surveillance had a carcinoma detected at an earlier Dukes stage and improved 5-year survival rate (50). A second analysis by Lashner and colleagues found that in 186 patients with extensive UC who underwent surveillance, patients had an improved survival and delayed time to colectomy, although the decrease in mortality was not related to cancer free survival (51). A Swedish population-based nested case-control study examining patients who died from CRC reported that two of 40 patients with UC who had died from colon cancer had undergone at least one screening exam, compared to 18 of 102 controls with UC who did not die from colon cancer (52). Similar protective effects of surveillance were seen in a second retrospective cohort (13). In a Cochrane database analysis of these studies published in 2006, the authors concluded that there was indirect evidence that surveillance is likely to show a cost benefit and be effective in reducing the risk of death from IBD-associated CRC (53).

The current standard of care recommended for the prevention of cancer in IBD is regular surveillance colonoscopy. The ability to prevent cancer with this strategy relies on the early detection of precancerous lesions. Most strategies for early detection involve both random biopsies and targeted biopsies of suspicious lesions. The major challenge with this strategy is sampling error. With random biopsies, it has been estimated 33 biopsies are needed to exclude dysplasia with 90% certainty and 64 biopsies are needed for a 95% certainty. Most gastroenterologists do not approach such numbers of biopsies during surveillance exams (54, 55). Several recent studies, however, indicate that the yield of targeted biopsies is much greater than random biopsies of the colon. One possible explanation for these findings was suggested by three recent retrospective analyses. These studies concluded that most dysplastic lesions can be visualized with white light colonoscopy (22, 23, 56).

Recent experience with chromoendoscopy, however, has consistently shown superior detection of dysplastic lesions with super vital staining compared to uncontrasted white light examinations (57-61). Chromoendoscopy is typically done with either indigo carmine or methylene blue dye. In a recent meta-analysis of six studies, the difference in proportion of lesions detected by chromoendoscopy vs. white light only was 44% (62). Autofluorescence with narrow band imaging (NBI) has been suggested to improve detection of dysplastic lesions in UC as well, although studies testing the benefit of NBI compared to high definition colonoscopy have been inconclusive (63-65).

Several recommendations have been published to guide surveillance strategies in patients with UC (66-69). The most recent consensus statement was released by the American Gastroenterological Association (AGA) and recommended initiating surveillance no later than 8 years of disease duration for patients with left-sided or pancolitis (69). During surveillance examinations, multiple biopsies should be obtained from each anatomic location in the colon. This statement included chromoendoscopy as a recommended alternative to random biopsies by endoscopists who have expertise with the technique. The AGA recommended repeat examinations every 1-3 years and to decrease the interval to every 1-2 years after 20 years of disease duration. For patients with PSC, surveillance exams should be performed at the time of diagnosis and then yearly thereafter, because of an increased risk earlier in the disease course.

7. Outcome after a diagnosis of dysplasia

After a diagnosis of flat HGD, colectomy has been universally recommended because there is a significant risk of harboring a synchronous CRC. One systematic review calculated this risk as 42% (70). A subsequent prospective analysis from St. Marks Hospital reported a 45% incidence of synchronous carcinomas in patients undergoing immediate colectomy after diagnosis of HGD (9). In this analysis, eight patients underwent surveillance. Of these eight, one developed CRC and seven developed further dysplasia (6 HGD, 1 LGD) (9). There appears to be a similar risk of development of cancer in patients with endoscopically unresectable DALMs (70, 71). Because of the high risk of CRC development, colectomy is warranted for any patient with a DALM or flat HGD.

In contrast to HGD, the management of patients with IBD-associated indeterminate dysplasia (IND) or LGD remains controversial. Previous studies have varied in their reported rates of progression from low-grade lesions to advanced neoplasia from 16% to 54% (9, 70, 72-76). The discrepancy in reported rates of progression to advanced neoplasia is likely secondary to the population heterogeneity of these studies. Within the classification of LGD, outcomes are different for flat dysplastic lesions and adenoma-like dysplastic lesions (ALMs). For patients who have an ALM in the absence of surrounding dysplasia, the risk of development of cancer appears to be minimal (77-79). For this reason, patients with polypoid lesions that resemble a sporadic adenoma without surrounding flat dysplasia can be managed with endoscopic resection and surveillance. Conversely, flat dysplastic lesions carry a higher risk of malignant progression and of harboring a synchronous CRC at the time of diagnosis (72, 74, 78). Total abdominal colectomy should be discussed with patients following a diagnosis of flat LGD. For patients with controlled disease who elect to undergo surveillance of flat LGD lesions, close follow up with endoscopic evaluations, initially at 3 to 6 month intervals is warranted.

Although neoplastic changes may develop in the pouch or in the anal transition zone, the risk of dysplasia appears to be low for patients with UC who undergo a restorative proctocolectomy with ileoanal anastomosis. One large analysis of 23 observational studies and over 2000 patients estimated that only slightly more than 1% of patients have confirmed dysplasia in the pouch or anal transition zone at follow up (80). A more recent analysis of over 3000 patients from the Cleveland Clinic reported the incidence of neoplasia to be 0.9%, 1.3%, 1.9%, 4.2%, and 5.1% at 5, 10, 15, 20, and 25 years after surgery, respectively (81). In both these studies, the risk of neoplastic transformation was significantly higher in patients who had dysplasia or cancer as their indication for initial colectomy. Although there are no published guidelines for surveillance after restorative proctocolectomy, many clinicians recommend a surveillance program because there remains a risk of neoplastic transformation, albeit low. It is postulated that performing a hand-sewn ileoanal anastomosis may decrease the risk of neoplasia. However, published studies have reported no difference between a stapled technique and hand-sewn anastomosis with mucosectomy (81, 82).

8. Chemoprevention

The primary goal of chemoprevention is to decrease the incidence of neoplastic lesions in those at increased risk. An effective chemopreventive agent offers the theoretical advantage over surveillance endoscopy alone by decreasing the frequency, cost, and risk of colonoscopy, as well as reducing need for colectomy.

The majority of studies examining chemopreventive agents in IBD have focused on the use of 5-aminosalicylates (5-ASA). There are several postulated mechanisms by which 5-ASA inhibits malignant transformation. These include inhibition of NF κ B, increased apoptosis of mutant clones, decreased proliferation, and prevention of oxygen-radical induced DNA damage (83-85). A meta-analysis of nine case control studies examining the efficacy of 5-ASA in preventing dysplasia or cancer revealed a pooled odds ratio of 0.51 (95% CI, 0.38-0.69) (86). However, several studies that have been published subsequent to this meta-analysis have not found a protective effect of 5-ASA therapy (87-90). Taken together, data to support 5-ASA chemoprevention in IBD is inconclusive, likely due to the heterogeneity of individuals in these studies. Furthermore, it is not known what effect 5-ASA has on CRC risk in patients who have achieved mucosal healing with other therapies.

The bile acid, ursodeoxycholic acid, has been used as a chemopreventive agent in UC patients with PSC. In animal models, UDCA is protective against the development of colon cancer (91-94). The mechanism of UDCA's chemopreventive activity remains uncertain, although it is likely multifactorial (92, 94). There have been two retrospective analyses of UDCA in patients with UC and PSC with conflicting results (93, 95-97). In a randomized placebo-controlled trial of UDCA at the dose of 13-15mg/kg-body wt/day, the relative risk for dysplasia or cancer in the group receiving UDCA was 0.26 (95% CI, 0.06-0.92). However, a more recent randomized placebo-controlled trial examining high dose UDCA (28-30 mg/kg-body wt/day) in UC patients with PSC found that patients taking UDCA had a higher risk of developing colorectal neoplasia (98). Currently, the American Association for the Study of Liver Diseases (AASLD) does not recommend UDCA for chemoprevention in patients with UC and PSC as larger prospective studies of low dose UDCA are needed to further evaluate this potential chemopreventive agent (99).

Other chemopreventive agents that have been studied in IBD-associated colitis include folic acid, immunomodulators, and vitamin D. Although a recent analysis of thiopurines in IBD found their use to be protective, the majority of studies investigating the chemopreventive efficacy of immunomodulators have not shown a benefit (10, 75, 100, 101). While folic acid deficiency is associated with decreased risk of sporadic CRC in epidemiological studies and folic acid is protective of other malignancies, the studies examining folic acid in the chemoprevention of CRC in patients with IBD have not demonstrated a benefit (102-104). The data on chemoprevention with folic acid in IBD comes from small retrospective analyses that have failed to show a statistical difference in the risk of dysplasia (105, 106). Finally, vitamin D has shown chemopreventive efficacy in murine models of sporadic and inflammation-associated colon cancer (42, 43, 107). Although vitamin D supplementation has not been examined in humans with UC, decreased vitamin D receptor expression is seen in cancers of patients with IBD and vitamin D appears to be chemopreventive of human CRC in epidemiological studies (44, 108). For this reason, vitamin D might offer a potential benefit to patients with chronic UC, although there have been no controlled studies in an IBD population.

9. Conclusion

Patients with chronic ulcerative colitis are at increased risk for the development of colon cancer. Because of this risk, colonoscopic surveillance is recommended for early detection of precancerous lesions. Successful implementation of surveillance programs has likely limited the mortality from CRC in this high-risk population. The outcome after detection of

dysplastic lesions needs to be better defined in future studies as endoscopic imaging techniques improve our ability to detect early neoplastic changes. Identification of effective chemopreventive agents against CRC development in UC could decrease the incidence and morbidity of colitis-associated neoplasia. To date, however, there is a lack of data to recommend the use of any specific chemopreventive agents in UC. Because of the cost and morbidity in the detection and treatment of neoplastic lesions in chronic UC, future research is urgently needed to identify efficacious, safe, and cost-effective chemopreventive agents and to establish clinical and biological predictors of dysplasia in order to tailor personalized surveillance strategies.

10. References

- [1] Prior P, Gyde SN, Macartney JC, Thompson H, Waterhouse JA, Allan RN. Cancer morbidity in ulcerative colitis. *Gut*. 1982;23(6):490-7.
- [2] Ekobom A, Helmick C, Zack M, Adami HO. Ulcerative colitis and colorectal cancer. A population-based study. *N Engl J Med*. 1990;323(18):1228-33.
- [3] Winther KV, Jess T, Langholz E, Munkholm P, Binder V. Long-term risk of cancer in ulcerative colitis: a population-based cohort study from Copenhagen County. *Clin Gastroenterol Hepatol*. 2004;2(12):1088-95.
- [4] Gyde SN, Prior P, Allan RN, Stevens A, Jewell DP, Truelove SC, Lofberg R, Brostrom O, Hellers G. Colorectal cancer in ulcerative colitis: a cohort study of primary referrals from three centres. *Gut*. 1988;29(2):206-17.
- [5] Bergeron V, Vienne A, Sokol H, Seksik P, Nion-Larmurier I, Ruskone-Fourmestreaux A, Svrcek M, Beaugerie L, Cosnes J. Risk factors for neoplasia in inflammatory bowel disease patients with pancolitis. *Am J Gastroenterol*. 2010;105(11):2405-11.
- [6] Devroede GJ, Taylor WF, Sauer WG, Jackman RJ, Stickler GB. Cancer risk and life expectancy of children with ulcerative colitis. *N Engl J Med*. 1971;285(1):17-21.
- [7] Eaden JA, Abrams KR, Mayberry JF. The risk of colorectal cancer in ulcerative colitis: a meta-analysis. *Gut*. 2001;48(4):526-35.
- [8] Jess T, Loftus EV, Jr., Velayos FS, Harmsen WS, Zinsmeister AR, Smyrk TC, Schleck CD, Tremaine WJ, Melton LJ, 3rd, Munkholm P, Sandborn WJ. Risk of intestinal cancer in inflammatory bowel disease: a population-based study from olmsted county, Minnesota. *Gastroenterology*. 2006;130(4):1039-46.
- [9] Rutter MD, Saunders BP, Wilkinson KH, Rumbles S, Schofield G, Kamm MA, Williams CB, Price AB, Talbot IC, Forbes A. Thirty-year analysis of a colonoscopic surveillance program for neoplasia in ulcerative colitis. *Gastroenterology*. 2006;130(4):1030-8.
- [10] Rutter M, Saunders B, Wilkinson K, Rumbles S, Schofield G, Kamm M, Williams C, Price A, Talbot I, Forbes A. Severity of inflammation is a risk factor for colorectal neoplasia in ulcerative colitis. *Gastroenterology*. 2004;126(2):451-9.
- [11] Gupta RB, Harpaz N, Itzkowitz S, Hossain S, Matula S, Kornbluth A, Bodian C, Ullman T. Histologic inflammation is a risk factor for progression to colorectal neoplasia in ulcerative colitis: a cohort study. *Gastroenterology*. 2007;133(4):1099-105; quiz 340-1.
- [12] Velayos FS, Loftus EV, Jr., Jess T, Harmsen WS, Bida J, Zinsmeister AR, Tremaine WJ, Sandborn WJ. Predictive and protective factors associated with colorectal cancer in ulcerative colitis: A case-control study. *Gastroenterology*. 2006;130(7):1941-9.

- [13] Eaden J, Abrams K, Ekbom A, Jackson E, Mayberry J. Colorectal cancer prevention in ulcerative colitis: a case-control study. *Aliment Pharmacol Ther.* 2000;14(2):145-53.
- [14] Nuako KW, Ahlquist DA, Mahoney DW, Schaid DJ, Siems DM, Lindor NM. Familial predisposition for colorectal cancer in chronic ulcerative colitis: a case-control study. *Gastroenterology.* 1998;115(5):1079-83.
- [15] Askling J, Dickman PW, Karlen P, Brostrom O, Lapidus A, Lofberg R, Ekbom A. Family history as a risk factor for colorectal cancer in inflammatory bowel disease. *Gastroenterology.* 2001;120(6):1356-62.
- [16] Shetty K, Rybicki L, Brzezinski A, Carey WD, Lashner BA. The risk for cancer or dysplasia in ulcerative colitis patients with primary sclerosing cholangitis. *Am J Gastroenterol.* 1999;94(6):1643-9.
- [17] Kornfeld D, Ekbom A, Ihre T. Is there an excess risk for colorectal cancer in patients with ulcerative colitis and concomitant primary sclerosing cholangitis? A population based study. *Gut.* 1997;41(4):522-5.
- [18] Marchesa P, Lashner BA, Lavery IC, Milsom J, Hull TL, Strong SA, Church JM, Navarro G, Fazio VW. The risk of cancer and dysplasia among ulcerative colitis patients with primary sclerosing cholangitis. *Am J Gastroenterol.* 1997;92(8):1285-8.
- [19] Brentnall TA, Haggitt RC, Rabinovitch PS, Kimmey MB, Bronner MP, Levine DS, Kowdley KV, Stevens AC, Crispin DA, Emond M, Rubin CE. Risk and natural history of colonic neoplasia in patients with primary sclerosing cholangitis and ulcerative colitis. *Gastroenterology.* 1996;110(2):331-8.
- [20] D'Haens GR, Lashner BA, Hanauer SB. Pericholangitis and sclerosing cholangitis are risk factors for dysplasia and cancer in ulcerative colitis. *Am J Gastroenterol.* 1993;88(8):1174-8.
- [21] Riddell RH, Goldman H, Ransohoff DF, Appelman HD, Fenoglio CM, Haggitt RC, Ahren C, Correa P, Hamilton SR, Morson BC, et al. Dysplasia in inflammatory bowel disease: standardized classification with provisional clinical applications. *Hum Pathol.* 1983;14(11):931-68.
- [22] Rubin DT, Rothe JA, Hetzel JT, Cohen RD, Hanauer SB. Are dysplasia and colorectal cancer endoscopically visible in patients with ulcerative colitis? *Gastrointest Endosc.* 2007;65(7):998-1004.
- [23] Rutter MD, Saunders BP, Wilkinson KH, Kamm MA, Williams CB, Forbes A. Most dysplasia in ulcerative colitis is visible at colonoscopy. *Gastrointest Endosc.* 2004;60(3):334-9.
- [24] Noble CL, Abbas AR, Cornelius J, Lees CW, Ho GT, Toy K, Modrusan Z, Pal N, Zhong F, Chalasani S, Clark H, Arnott ID, Penman ID, Satsangi J, Diehl L. Regional variation in gene expression in the healthy colon is dysregulated in ulcerative colitis. *Gut.* 2008;57(10):1398-405.
- [25] Okahara S, Arimura Y, Yabana T, Kobayashi K, Gotoh A, Motoya S, Imamura A, Endo T, Imai K. Inflammatory gene signature in ulcerative colitis with cDNA macroarray analysis. *Aliment Pharmacol Ther.* 2005;21(9):1091-7.
- [26] Pekow JR, Dougherty U, Mustafi R, Zhu H, Kocherginsky M, Rubin DT, Hanauer SB, Hart J, Chang EB, Fichera A, Joseph LJ, Bissonnette M. miR-143 and miR-145 are downregulated in ulcerative colitis: Putative regulators of inflammation and protooncogenes. *Inflamm Bowel Dis.* 2011. epub ahead of print.
- [27] Tahara T, Inoue N, Hisamatsu T, Kashiwagi K, Takaishi H, Kanai T, Watanabe M, Ishii H, Hibi T. Clinical significance of microsatellite instability in the inflamed mucosa

- for the prediction of colonic neoplasms in patients with ulcerative colitis. *J Gastroenterol Hepatol*. 2005;20(5):710-5.
- [28] Umetani N, Sasaki S, Watanabe T, Shinozaki M, Matsuda K, Ishigami H, Ueda E, Muto T. Genetic alterations in ulcerative colitis-associated neoplasia focusing on APC, K-ras gene and microsatellite instability. *Jpn J Cancer Res*. 1999;90(10):1081-7.
- [29] Xie J, Itzkowitz SH. Cancer in inflammatory bowel disease. *World J Gastroenterol*. 2008;14(3):378-89.
- [30] Goel GA, Kandiel A, Achkar JP, Lashner B. Molecular pathways underlying IBD-associated colorectal neoplasia: therapeutic implications. *Am J Gastroenterol*. 2011;106(4):719-30.
- [31] Leedham SJ, Graham TA, Oukrif D, McDonald SA, Rodriguez-Justo M, Harrison RF, Shepherd NA, Novelli MR, Jankowski JA, Wright NA. Clonality, founder mutations, and field cancerization in human ulcerative colitis-associated neoplasia. *Gastroenterology*. 2009;136(2):542-50 e6.
- [32] Brentnall TA, Crispin DA, Rabinovitch PS, Haggitt RC, Rubin CE, Stevens AC, Burner GC. Mutations in the p53 gene: an early marker of neoplastic progression in ulcerative colitis. *Gastroenterology*. 1994;107(2):369-78.
- [33] Burner GC, Rabinovitch PS, Haggitt RC, Crispin DA, Brentnall TA, Kolli VR, Stevens AC, Rubin CE. Neoplastic progression in ulcerative colitis: histology, DNA content, and loss of a p53 allele. *Gastroenterology*. 1992;103(5):1602-10.
- [34] Yoshizawa S, Matsuoka K, Inoue N, Takaishi H, Ogata H, Iwao Y, Mukai M, Fujita T, Kawakami Y, Hibi T. Clinical significance of serum p53 antibodies in patients with ulcerative colitis and its carcinogenesis. *Inflamm Bowel Dis*. 2007;13(7):865-73.
- [35] Gerrits MM, Chen M, Theeuwes M, van Dekken H, Sikkema M, Steyerberg EW, Lingsma HF, Siersema PD, Xia B, Kusters JG, van der Woude CJ, Kuipers EJ. Biomarker-based prediction of inflammatory bowel disease-related colorectal cancer: a case-control study. *Cell Oncol (Dordr)*. 2011;34(2):107-17.
- [36] van Schaik FD, Oldenburg B, Offerhaus GJ, Schipper ME, Vleggaar FP, Siersema PD, van Oijen MG, Ten Kate FJ. Role of immunohistochemical markers in predicting progression of dysplasia to advanced neoplasia in patients with ulcerative colitis. *Inflamm Bowel Dis*. 2011. epub ahead of print.
- [37] Claessen MM, Schipper ME, Oldenburg B, Siersema PD, Offerhaus GJ, Vleggaar FP. WNT-pathway activation in IBD-associated colorectal carcinogenesis: potential biomarkers for colonic surveillance. *Cell Oncol*. 2010;32(4):303-10.
- [38] Dhir M, Montgomery EA, Glockner SC, Schuebel KE, Hooker CM, Herman JG, Baylin SB, Gearhart SL, Ahuja N. Epigenetic regulation of WNT signaling pathway genes in inflammatory bowel disease (IBD) associated neoplasia. *J Gastrointest Surg*. 2008;12(10):1745-53.
- [39] You XJ, Bryant PJ, Journak F, Holcombe RF. Expression of Wnt pathway components frizzled and disheveled in colon cancer arising in patients with inflammatory bowel disease. *Oncol Rep*. 2007;18(3):691-4.
- [40] Kukitsu T, Takayama T, Miyanishi K, Nobuoka A, Katsuki S, Sato Y, Takimoto R, Matsunaga T, Kato J, Sonoda T, Sakamaki S, Niitsu Y. Aberrant crypt foci as precursors of the dysplasia-carcinoma sequence in patients with ulcerative colitis. *Clin Cancer Res*. 2008;14(1):48-54.

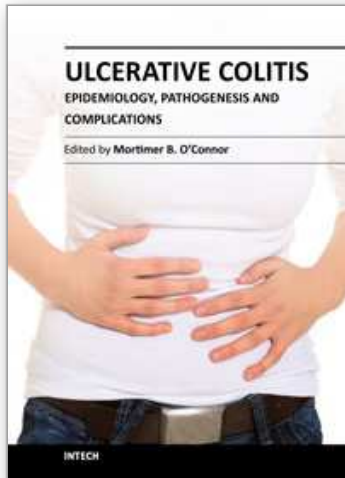
- [41] Aust DE, Terdiman JP, Willenbacher RF, Chang CG, Molinaro-Clark A, Baretton GB, Loehrs U, Waldman FM. The APC/beta-catenin pathway in ulcerative colitis-related colorectal carcinomas: a mutational analysis. *Cancer*. 2002;94(5):1421-7.
- [42] Fichera A, Little N, Dougherty U, Mustafi R, Cerda S, Li YC, Delgado J, Arora A, Campbell LK, Joseph L, Hart J, Noffsinger A, Bissonnette M. A vitamin D analogue inhibits colonic carcinogenesis in the AOM/DSS model. *J Surg Res*. 2007;142(2):239-45.
- [43] Kikuchi H, Murakami S, Suzuki S, Kudo H, Sassa S, Sakamoto S. Chemopreventive effect of a vitamin D(3) analog, alfacalcidol, on colorectal carcinogenesis in mice with ulcerative colitis. *Anticancer Drugs*. 2007;18(10):1183-7.
- [44] Wada K, Tanaka H, Maeda K, Inoue T, Noda E, Amano R, Kubo N, Muguruma K, Yamada N, Yashiro M, Sawada T, Nakata B, Ohira M, Hirakawa K. Vitamin D receptor expression is associated with colon cancer in ulcerative colitis. *Oncol Rep*. 2009;22(5):1021-5.
- [45] Mantovani A. Molecular pathways linking inflammation and cancer. *Curr Mol Med*. 2010;10(4):369-73.
- [46] Greten FR, Eckmann L, Greten TF, Park JM, Li ZW, Egan LJ, Kagnoff MF, Karin M. IKKbeta links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. *Cell*. 2004;118(3):285-96.
- [47] Clevers H. At the crossroads of inflammation and cancer. *Cell*. 2004;118(6):671-4.
- [48] Fukata M, Chen A, Vamadevan AS, Cohen J, Breglio K, Krishnareddy S, Hsu D, Xu R, Harpaz N, Dannenberg AJ, Subbaramaiah K, Cooper HS, Itzkowitz SH, Abreu MT. Toll-like receptor-4 promotes the development of colitis-associated colorectal tumors. *Gastroenterology*. 2007;133(6):1869-81.
- [49] Dougherty U, Cerasi D, Taylor I, Kocherginsky M, Tekin U, Badal S, Aluri L, Sehdev A, Cerda S, Mustafi R, Delgado J, Joseph L, Zhu H, Hart J, Threadgill D, Fichera A, Bissonnette M. Epidermal growth factor receptor is required for colonic tumor promotion by dietary fat in the azoxymethane/dextran sulfate sodium model: roles of transforming growth factor- α and PTGS2. *Clin Cancer Res*. 2009;15(22):6780-9.
- [50] Choi PM, Nugent FW, Schoetz DJ, Jr., Silverman ML, Haggitt RC. Colonoscopic surveillance reduces mortality from colorectal cancer in ulcerative colitis. *Gastroenterology*. 1993;105(2):418-24.
- [51] Lashner BA, Kane SV, Hanauer SB. Colon cancer surveillance in chronic ulcerative colitis: historical cohort study. *Am J Gastroenterol*. 1990;85(9):1083-7.
- [52] Karlen P, Kornfeld D, Brostrom O, Lofberg R, Persson PG, Ekbohm A. Is colonoscopic surveillance reducing colorectal cancer mortality in ulcerative colitis? A population based case control study. *Gut*. 1998;42(5):711-4.
- [53] Collins PD, Mpofu C, Watson AJ, Rhodes JM. Strategies for detecting colon cancer and/or dysplasia in patients with inflammatory bowel disease. *Cochrane Database Syst Rev*. 2006(2):CD000279.
- [54] Rubin CE, Haggitt RC, Burmer GC, Brentnall TA, Stevens AC, Levine DS, Dean PJ, Kimmey M, Perera DR, Rabinovitch PS. DNA aneuploidy in colonic biopsies predicts future development of dysplasia in ulcerative colitis. *Gastroenterology*. 1992;103(5):1611-20.
- [55] Bernstein CN, Weinstein WM, Levine DS, Shanahan F. Physicians' perceptions of dysplasia and approaches to surveillance colonoscopy in ulcerative colitis. *Am J Gastroenterol*. 1995;90(12):2106-14.

- [56] Blonski W, Kundu R, Lewis J, Aberra F, Osterman M, Lichtenstein GR. Is dysplasia visible during surveillance colonoscopy in patients with ulcerative colitis? *Scand J Gastroenterol.* 2008;43(6):698-703.
- [57] Kiesslich R, Fritsch J, Holtmann M, Koehler HH, Stolte M, Kanzler S, Nafe B, Jung M, Galle PR, Neurath MF. Methylene blue-aided chromoendoscopy for the detection of intraepithelial neoplasia and colon cancer in ulcerative colitis. *Gastroenterology.* 2003;124(4):880-8.
- [58] Kiesslich R, Goetz M, Lammersdorf K, Schneider C, Burg J, Stolte M, Vieth M, Nafe B, Galle PR, Neurath MF. Chromoscopy-guided endomicroscopy increases the diagnostic yield of intraepithelial neoplasia in ulcerative colitis. *Gastroenterology.* 2007;132(3):874-82.
- [59] Marion JF, Waye JD, Present DH, Israel Y, Bodian C, Harpaz N, Chapman M, Itzkowitz S, Steinlauf AF, Abreu MT, Ullman TA, Aisenberg J, Mayer L, Chromoendoscopy Study Group at Mount Sinai School of M. Chromoendoscopy-targeted biopsies are superior to standard colonoscopic surveillance for detecting dysplasia in inflammatory bowel disease patients: a prospective endoscopic trial. *Am J Gastroenterol.* 2008;103(9):2342-9.
- [60] Hurlstone DP, Sanders DS, Lobo AJ, McAlindon ME, Cross SS. Indigo carmine-assisted high-magnification chromoscopic colonoscopy for the detection and characterisation of intraepithelial neoplasia in ulcerative colitis: a prospective evaluation. *Endoscopy.* 2005;37(12):1186-92.
- [61] Rutter MD, Saunders BP, Schofield G, Forbes A, Price AB, Talbot IC. Pancolonoscopic indigo carmine dye spraying for the detection of dysplasia in ulcerative colitis. *Gut.* 2004;53(2):256-60.
- [62] Subramanian V, Mannath J, Ragunath K, Hawkey CJ. Meta-analysis: the diagnostic yield of chromoendoscopy for detecting dysplasia in patients with colonic inflammatory bowel disease. *Aliment Pharmacol Ther.* 33(3):304-12.
- [63] van den Broek FJ, Fockens P, van Eeden S, Reitsma JB, Hardwick JC, Stokkers PC, Dekker E. Endoscopic tri-modal imaging for surveillance in ulcerative colitis: randomised comparison of high-resolution endoscopy and autofluorescence imaging for neoplasia detection; and evaluation of narrow-band imaging for classification of lesions. *Gut.* 2008;57(8):1083-9.
- [64] Dekker E, van den Broek FJ, Reitsma JB, Hardwick JC, Offerhaus GJ, van Deventer SJ, Hommes DW, Fockens P. Narrow-band imaging compared with conventional colonoscopy for the detection of dysplasia in patients with longstanding ulcerative colitis. *Endoscopy.* 2007;39(3):216-21.
- [65] van den Broek FJ, Fockens P, van Eeden S, Stokkers PC, Ponsioen CY, Reitsma JB, Dekker E. Narrow-band imaging versus high-definition endoscopy for the diagnosis of neoplasia in ulcerative colitis. *Endoscopy.* 2011;43(2):108-15.
- [66] Carter MJ, Lobo AJ, Travis SP, Ibd Section BSoG. Guidelines for the management of inflammatory bowel disease in adults. *Gut.* 2004;53 Suppl 5:V1-16.
- [67] Kornbluth A, Sachar DB. Ulcerative colitis practice guidelines in adults (update): American College of Gastroenterology, Practice Parameters Committee. *Am J Gastroenterol.* 2004;99(7):1371-85.
- [68] Itzkowitz SH, Present DH, Crohn's, Colitis Foundation of America Colon Cancer in IBD SG. Consensus conference: Colorectal cancer screening and surveillance in inflammatory bowel disease. *Inflamm Bowel Dis.* 2005;11(3):314-21.

- [69] Farraye FA, Odze RD, Eaden J, Itzkowitz SH, McCabe RP, Dassopoulos T, Lewis JD, Ullman TA, James T, 3rd, McLeod R, Burgart LJ, Allen J, Brill JV. Diagnosis AGAIMPPo, Management of Colorectal Neoplasia in Inflammatory Bowel D. AGA medical position statement on the diagnosis and management of colorectal neoplasia in inflammatory bowel disease. *Gastroenterology*. 2010;138(2):738-45.
- [70] Bernstein CN, Shanahan F, Weinstein WM. Are we telling patients the truth about surveillance colonoscopy in ulcerative colitis? *Lancet*. 1994;343(8889):71-4.
- [71] Blackstone MO, Riddell RH, Rogers BH, Levin B. Dysplasia-associated lesion or mass (DALM) detected by colonoscopy in long-standing ulcerative colitis: an indication for colectomy. *Gastroenterology*. 1981;80(2):366-74.
- [72] Ullman T, Croog V, Harpaz N, Sachar D, Itzkowitz S. Progression of flat low-grade dysplasia to advanced neoplasia in patients with ulcerative colitis. *Gastroenterology*. 2003;125(5):1311-9.
- [73] Jess T, Loftus EV, Jr., Velayos FS, Harmsen WS, Zinsmeister AR, Smyrk TC, Tremaine WJ, Melton LJ, 3rd, Munkholm P, Sandborn WJ. Incidence and prognosis of colorectal dysplasia in inflammatory bowel disease: a population-based study from Olmsted County, Minnesota. *Inflamm Bowel Dis*. 2006;12(8):669-76.
- [74] Ullman TA, Loftus EV, Jr., Kakar S, Burgart LJ, Sandborn WJ, Tremaine WJ. The fate of low grade dysplasia in ulcerative colitis. *Am J Gastroenterol*. 2002;97(4):922-7.
- [75] Connell WR, Lennard-Jones JE, Williams CB, Talbot IC, Price AB, Wilkinson KH. Factors affecting the outcome of endoscopic surveillance for cancer in ulcerative colitis. *Gastroenterology*. 1994;107(4):934-44.
- [76] Lim CH, Dixon MF, Vail A, Forman D, Lynch DA, Axon AT. Ten year follow up of ulcerative colitis patients with and without low grade dysplasia. *Gut*. 2003;52(8):1127-32.
- [77] Odze RD, Farraye FA, Hecht JL, Hornick JL. Long-term follow-up after polypectomy treatment for adenoma-like dysplastic lesions in ulcerative colitis. *Clin Gastroenterol Hepatol*. 2004;2(7):534-41.
- [78] Pekow JR, Hetzel JT, Rothe JA, Hanauer SB, Turner JR, Hart J, Noffsinger A, Huo D, Rubin DT. Outcome after surveillance of low-grade and indefinite dysplasia in patients with ulcerative colitis. *Inflamm Bowel Dis*. 16(8):1352-6.
- [79] Engelsgerd M, Farraye FA, Odze RD. Polypectomy may be adequate treatment for adenoma-like dysplastic lesions in chronic ulcerative colitis. *Gastroenterology*. 1999;117(6):1288-94; discussion 488-91.
- [80] Scarpa M, van Koperen PJ, Ubbink DT, Hommes DW, Ten Kate FJ, Bemelman WA. Systematic review of dysplasia after restorative proctocolectomy for ulcerative colitis. *Br J Surg*. 2007;94(5):534-45.
- [81] Kariv R, Remzi FH, Lian L, Bennett AE, Kiran RP, Kariv Y, Fazio VW, Lavery IC, Shen B. Preoperative colorectal neoplasia increases risk for pouch neoplasia in patients with restorative proctocolectomy. *Gastroenterology*. 2010;139(3):806-12, 12 e1-2.
- [82] Al-Sukhni W, McLeod RS, MacRae H, O'Connor B, Huang H, Cohen Z. Oncologic outcome in patients with ulcerative colitis associated with dysplasia or cancer who underwent stapled or handsewn ileal pouch-anal anastomosis. *Dis Colon Rectum*. 53(11):1495-500.

- [83] Kaiser GC, Yan F, Polk DB. Mesalamine blocks tumor necrosis factor growth inhibition and nuclear factor kappaB activation in mouse colonocytes. *Gastroenterology*. 1999;116(3):602-9.
- [84] Reinacher-Schick A, Seidensticker F, Petrasch S, Reiser M, Philippou S, Theegarten D, Freitag G, Schmiegel W. Mesalazine changes apoptosis and proliferation in normal mucosa of patients with sporadic polyps of the large bowel. *Endoscopy*. 2000;32(3):245-54.
- [85] Allgayer H, Kolb M, Stuber V, Kruis W. Modulation of base hydroxylation by bile acids and salicylates in a model of human colonic mucosal DNA: putative implications in colonic cancer. *Dig Dis Sci*. 1999;44(4):761-7.
- [86] Velayos FS, Terdiman JP, Walsh JM. Effect of 5-aminosalicylate use on colorectal cancer and dysplasia risk: a systematic review and metaanalysis of observational studies. *Am J Gastroenterol*. 2005;100(6):1345-53.
- [87] Terdiman JP, Steinbuch M, Blumentals WA, Ullman TA, Rubin DT. 5-Aminosalicylic acid therapy and the risk of colorectal cancer among patients with inflammatory bowel disease. *Inflamm Bowel Dis*. 2007;13(4):367-71.
- [88] Jess T, Loftus EV, Jr., Velayos FS, Winther KV, Tremaine WJ, Zinsmeister AR, Scott Harmsen W, Langholz E, Binder V, Munkholm P, Sandborn WJ. Risk factors for colorectal neoplasia in inflammatory bowel disease: a nested case-control study from Copenhagen county, Denmark and Olmsted county, Minnesota. *Am J Gastroenterol*. 2007;102(4):829-36.
- [89] Bernstein CN, Nugent Z, Blanchard JF. 5-aminosalicylate is not chemoprophylactic for colorectal cancer in IBD: a population based study. *Am J Gastroenterol*. 2011;106(4):731-6.
- [90] Ullman T, Croog V, Harpaz N, Hossain S, Kornbluth A, Bodian C, Itzkowitz S. Progression to colorectal neoplasia in ulcerative colitis: effect of mesalamine. *Clin Gastroenterol Hepatol*. 2008;6(11):1225-30; quiz 177.
- [91] Earnest DL, Holubec H, Wali RK, Jolley CS, Bissonette M, Bhattacharyya AK, Roy H, Khare S, Brasitus TA. Chemoprevention of azoxymethane-induced colonic carcinogenesis by supplemental dietary ursodeoxycholic acid. *Cancer Res*. 1994;54(19):5071-4.
- [92] Wali RK, Frawley BP, Jr., Hartmann S, Roy HK, Khare S, Scaglione-Sewell BA, Earnest DL, Sitrin MD, Brasitus TA, Bissonette M. Mechanism of action of chemoprotective ursodeoxycholate in the azoxymethane model of rat colonic carcinogenesis: potential roles of protein kinase C-alpha, -beta II, and -zeta. *Cancer Res*. 1995;55(22):5257-64.
- [93] Khare S, Mustafi R, Cerda S, Yuan W, Jagadeeswaran S, Dougherty U, Tretiakova M, Samarel A, Cohen G, Wang J, Moore C, Wali R, Holgren C, Joseph L, Fichera A, Li YC, Bissonette M. Ursodeoxycholic acid suppresses Cox-2 expression in colon cancer: roles of Ras, p38, and CCAAT/enhancer-binding protein. *Nutr Cancer*. 2008;60(3):389-400.
- [94] Batta AK, Salen G, Holubec H, Brasitus TA, Alberts D, Earnest DL. Enrichment of the more hydrophilic bile acid ursodeoxycholic acid in the fecal water-soluble fraction after feeding to rats with colon polyps. *Cancer Res*. 1998;58(8):1684-7.
- [95] Tung BY, Emond MJ, Haggitt RC, Bronner MP, Kimmey MB, Kowdley KV, Brentnall TA. Ursodiol use is associated with lower prevalence of colonic neoplasia in

- patients with ulcerative colitis and primary sclerosing cholangitis. *Ann Intern Med.* 2001;134(2):89-95.
- [96] Wolf JM, Rybicki LA, Lashner BA. The impact of ursodeoxycholic acid on cancer, dysplasia and mortality in ulcerative colitis patients with primary sclerosing cholangitis. *Aliment Pharmacol Ther.* 2005;22(9):783-8.
- [97] Khare S, Cerda S, Wali RK, von Lintig FC, Tretiakova M, Joseph L, Stoiber D, Cohen G, Nimmagadda K, Hart J, Sitrin MD, Boss GR, Bissonnette M. Ursodeoxycholic acid inhibits Ras mutations, wild-type Ras activation, and cyclooxygenase-2 expression in colon cancer. *Cancer Res.* 2003;63(13):3517-23.
- [98] Eaton JE, Silveira MG, Pardi DS, Sinakos E, Kowdley KV, Luketic VA, Harrison ME, McCashland T, Befeler AS, Harnois D, Jorgensen R, Petz J, Lindor KD. High-Dose Ursodeoxycholic Acid Is Associated With the Development of Colorectal Neoplasia in Patients With Ulcerative Colitis and Primary Sclerosing Cholangitis. *Am J Gastroenterol.* 2011. epub ahead of print.
- [99] Chapman R, Fevery J, Kalloo A, Nagorney DM, Boberg KM, Shneider B, Gores GJ, American Association for the Study of Liver D. Diagnosis and management of primary sclerosing cholangitis. *Hepatology.* 51(2):660-78.
- [100] van Schaik FD, van Oijen MG, Smeets HM, van der Heijden GJ, Siersema PD, Oldenburg B. Thiopurines prevent advanced colorectal neoplasia in patients with inflammatory bowel disease. *Gut.* 2011. epub ahead of print.
- [101] Matula S, Croog V, Itzkowitz S, Harpaz N, Bodian C, Hossain S, Ullman T. Chemoprevention of colorectal neoplasia in ulcerative colitis: the effect of 6-mercaptopurine. *Clin Gastroenterol Hepatol.* 2005;3(10):1015-21.
- [102] Heimburger DC, Alexander CB, Birch R, Butterworth CE, Jr., Bailey WC, Krumdieck CL. Improvement in bronchial squamous metaplasia in smokers treated with folate and vitamin B12. Report of a preliminary randomized, double-blind intervention trial. *JAMA.* 1988;259(10):1525-30.
- [103] Freudenheim JL, Graham S, Marshall JR, Haughey BP, Cholewinski S, Wilkinson G. Folate intake and carcinogenesis of the colon and rectum. *Int J Epidemiol.* 1991;20(2):368-74.
- [104] Butterworth CE, Jr., Hatch KD, Gore H, Mueller H, Krumdieck CL. Improvement in cervical dysplasia associated with folic acid therapy in users of oral contraceptives. *Am J Clin Nutr.* 1982;35(1):73-82.
- [105] Lashner BA, Heidenreich PA, Su GL, Kane SV, Hanauer SB. Effect of folate supplementation on the incidence of dysplasia and cancer in chronic ulcerative colitis. A case-control study. *Gastroenterology.* 1989;97(2):255-9.
- [106] Lashner BA, Provencher KS, Seidner DL, Knesebeck A, Brzezinski A. The effect of folic acid supplementation on the risk for cancer or dysplasia in ulcerative colitis. *Gastroenterology.* 1997;112(1):29-32.
- [107] Wali RK, Bissonnette M, Khare S, Hart J, Sitrin MD, Brasitus TA. 1 alpha,25-Dihydroxy-16-ene-23-yne-26,27-hexafluorocholecalciferol, a noncalcemic analogue of 1 alpha,25-dihydroxyvitamin D₃, inhibits azoxymethane-induced colonic tumorigenesis. *Cancer Res.* 1995;55(14):3050-4.
- [108] Gorham ED, Garland CF, Garland FC, Grant WB, Mohr SB, Lipkin M, Newmark HL, Giovannucci E, Wei M, Holick MF. Optimal vitamin D status for colorectal cancer prevention: a quantitative meta analysis. *Am J Prev Med.* 2007;32(3):210-6.



Ulcerative Colitis - Epidemiology, Pathogenesis and Complications

Edited by Dr Mortimer O'Connor

ISBN 978-953-307-880-9

Hard cover, 280 pages

Publisher InTech

Published online 14, December, 2011

Published in print edition December, 2011

This book is intended to act as an up-to-date reference point and knowledge developer for all readers interested in the area of gastroenterology and in particular, Ulcerative Colitis. All authors of the chapters are experts in their fields of publication, and deserve individual credit and praise for their contributions to the world of Ulcerative Colitis. We hope that you will find this publication informative, stimulating, and a reference point for the area of Ulcerative colitis as we move forward in our understanding of the field of medicine.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Joel Pekow and Marc Bissonnette (2011). Neoplasia in IBD, Ulcerative Colitis - Epidemiology, Pathogenesis and Complications, Dr Mortimer O'Connor (Ed.), ISBN: 978-953-307-880-9, InTech, Available from: <http://www.intechopen.com/books/ulcerative-colitis-epidemiology-pathogenesis-and-complications/neoplasia-in-ibd>

INTECH
open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2011 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen