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1,25(OH)₂D₃ and Cyclooxygenase-2: Possible Targets for Breast Cancer?

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1. Introduction

Currently, breast cancer is the most common malignancy in women. In the U.S. in 2005, approximately 211,240 patients were newly diagnosed with primary breast cancer and 58,490 women were diagnosed with ductal carcinoma in situ (DCIS). Of these, 58,490 deaths are estimated. Therefore breast cancer takes second place following only behind lung cancer [1,2,3]. Because of this, it is necessary to develop new strategies and treatment options that may improve the prognosis.

Besides the classic histo-pathological parameters used to estimate the prognosis of malignant diseases, the identification of additional molecular prognostic parameters would be very helpful in planning treatment by evaluating protein or mRNA expression in tumor tissue. One of these potential molecular prognostic parameters might be the cyclooxygenase-2 (COX-2) [4, 5]. New treatment strategies using compounds that attack well defined proteins in the tumor require verification of the expression of these target proteins. Many similarities exist between tumor tissue and inflammatory modified tissue and normally, inflammatory reaction is self limiting, however, in tumor tissue the inflammatory reaction is persistent. An increased angiogenesis and an elevated production of cytokines, chemokines and proteases lead to good conditions for cell proliferation and invasion in the tumor tissue [6].

Targeted strategies might eliminate this inflammatory reaction that promotes tumor growth and tumorigenesis and there is already promising data around the use of COX-2-inhibitors. The antiproliferative effects of vitamin D may be another starting point; however the data on vitamin D intake or to the exertion of vitamin D analogs is occasionally inconsistent.

The important role that vitamin D and calcium adopt in the human metabolism was recognized as early as the nineteen-twenties as it was used to prevent the bone disease, rickets which was widespread in children at that time [7]. In the last 20 years non-classical effects of vitamin D and its influence on physiology followed because it's potentially anticarcinogen impacts made it more and more interesting. Besides stable calcium-homeostasis by the renal expressed 1- α -hydroxylase functionality, extra renal expressed 1- α -hydroxylase also is also known to have antiproliferative and immune-modulating features

[8-10]. This fact has led to the development of new treatment strategies in the clinical use of $1,25(\text{OH})_2\text{D}_3$ (calcitriol). The goal was to affect and treat cancer, psoriasis, autoimmune diseases and host-graft-rejection [11-14]. Implementation of these new treatment options in vivo was conspicuously hindered as $1,25(\text{OH})_2\text{D}_3$ has a potentially hypercalcaemic side effect. Finally the application of synthetic $1,25(\text{OH})_2\text{D}_3$ analogs led to several successful results due to its less calciotropic effects [15, 16]. The implementation of vitamin D, primarily cancer and autoimmune diseases appears to play a more preventative role as opposed to therapeutic [17, 18].

Observational studies showed an association between vitamin D intake and $25(\text{OH})_2\text{D}_3$ plasma levels as well as a reduced risk of breast cancer [19, 20]. Studies that tried to elucidate the correlation between sunlight and cancer prevention demonstrated that long sunlight exposure was associated with a low rate of primary breast cancer and consecutively a low mortality rate [21-24]. $1,25(\text{OH})_2\text{D}_3$ is the biologically active form of vitamin D that binds as a ligand to the nuclear vitamin D receptor (VDR) of the genes that are important for vitamin D metabolism (1- α -hydroxylase, 24-hydroxylase) [25]. $1,25(\text{OH})_2\text{D}_3$ and its analogs are able to inhibit the proliferation of breast cancer cells in vitro and in vivo [26-29].

2. Prostaglandin metabolism

The cyclooxygenase system consists of two different isoenzymes, COX-1 and COX-2. This system is an integral part of the prostaglandin synthetase complex and is involved controlling inflammatory processes (1). After transformation of arachidonic acid to prostaglandin G_2 (PGG_2), a glutathione dependent peroxidase converts PGG_2 to PGH_2 by an oxi- and peroxidation. PGH_2 acts as basic substrate for the synthesis of different prostaglandins by the microsomal and cytosolic prostaglandin synthase, which are tissue- and cell-specific. Based on the cellular enzyme setting, different prostaglandins are synthesized in different tissues where they act in an auto- or paracrine manner [30]. Prostaglandin E_2 (PGE_2) is one of the best known prostaglandins and is generated by the prostaglandin E synthase. These consist of three different forms: two microsomal prostaglandin E synthases and the cytosolic prostaglandin synthase E [31].

The 15-hydroxyprostaglandin dehydrogenase (15-PGDH) belonging to the oxidoreductases family, inactivates all generated prostaglandins by oxidation to 15-keto metabolites which then have greatly reduced biological activity [32].

3. PG-receptors

The physiological effects of many prostaglandins are mediated by binding to G protein coupled receptors. These effects regulate inflammatory mediations, control hormone regulation, constrict or dilate in vascular smooth muscle cells and regulate calcium movement and their specific receptors activate signal transduction pathways which could induce chronic processes like angiogenesis. For example, PGE_2 interacts with four cell surface receptors - EP_{1-4} and the EP_2 receptor subtype is involved in the G_s /cAMP/proteinkinase which is a signalling pathway leading to an increased VEGF expression. PGI_2 and PGA_2 interact with nuclear receptors, belonging to the peroxisome proliferator-activated receptors (PPARs) family. After dimerization with the 9-cis-Retinoid receptor (RXR) and then binding to a sequence specific responsive element located at the promoter of its target gene, they directly induce gene expression [33].

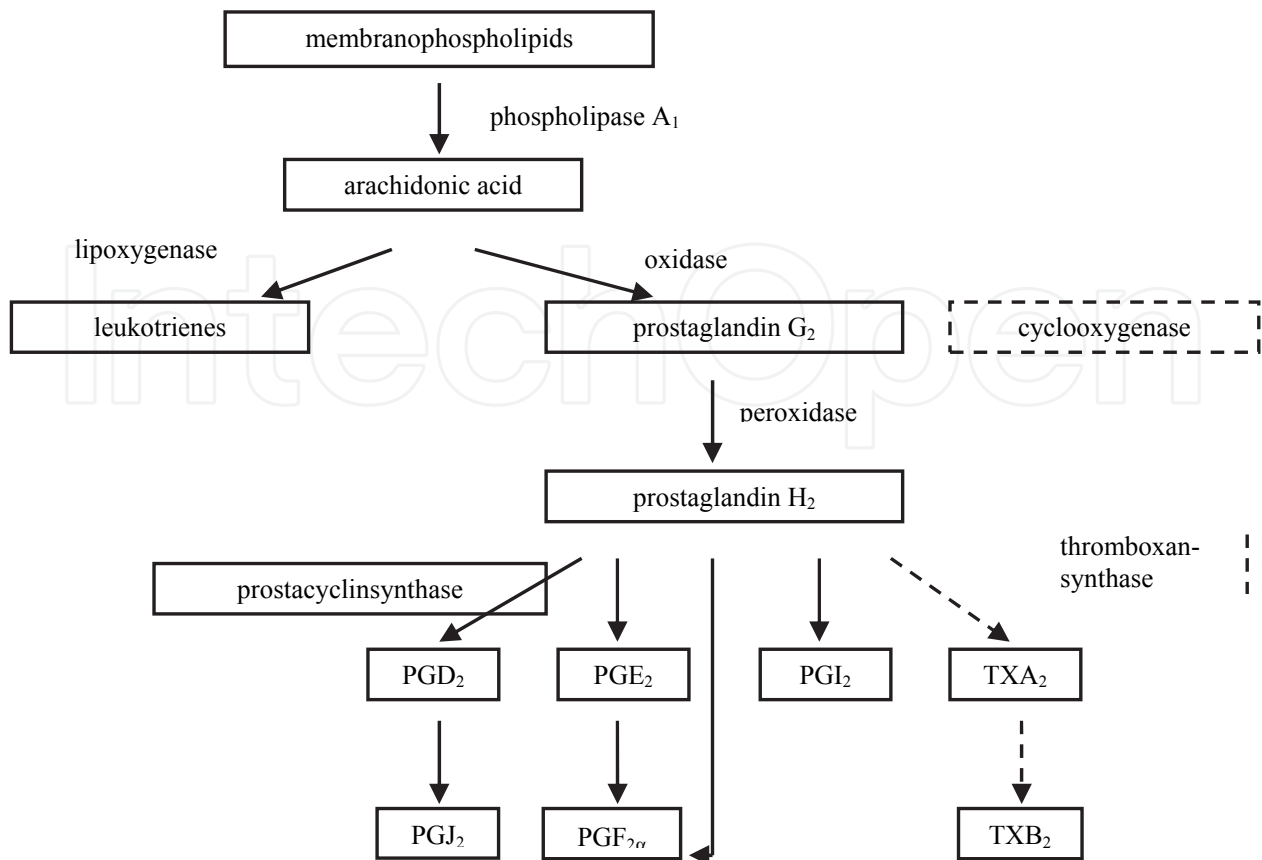


Fig. 1. Prostaglandin metabolism.

4. Isoenzymes COX-1 und COX-2

COX-1 is ubiquitous and not a relevant prognostic factor [34]. In contrast, the COX-2 enzyme is not constitutively expressed. The COX-2 gene expression is stimulated by many growth factors, cytokines and prostaglandins and is associated with inflammation [35]. COX-2 is predominantly a pro-inflammatory enzyme but late in the inflammatory phase, the enzyme is involved in limiting inflammation.

Studies with COX-1 and COX-2 knockout mice lead to new consolidated findings about the function of these enzymes concerning ovarian functionality and reproduction as well as cardiovascular development [36 - 39]. The COX enzymes are the main target of non-steroidal anti-inflammatory drugs (NSAID) where isoenzymes specifically inhibit the biological activity of COX enzymes. Celecoxib and rofecoxib are selective COX-2 inhibitors whereas acetylsalicylic acid, ibuprofen and indomethacin are non-specific and target both isoenzymes.

5. Role of COX-2 in carcinogenesis

The cyclooxygenases, especially COX-2, play an important role in the development and progression of malignant tumours. The over expression of COX-2 is associated with the differentiation of tumor cells by several mechanisms [40] and can be detected in various epithelial carcinomas such as in colon [41, 42], gastric [43] and esophageal cancers [44] as well as in prostate [45], liver, pancreas and lung cancers [46]. One of the mechanisms that are modulated during carcinogenesis is the neoangiogenesis [47 - 55].

Epidemiological studies have shown that a continuous intake of NSAIDs protects against the incidence of breast cancer [56-58].

Increased PGE₂ levels can be detected in cultivated human breast cancer cell lines as well as in invasive human breast cancer cells [59-62] and are associated with both a negative hormone receptor status and an escalated metastatic potential [59].

As mentioned previously, PGE₂ is the ligand for at least four cell surface receptors - EP₁₋₄ and several studies have presented the impact of the EP₁-receptor in carcinogenesis of colon and breast cancer [63]. A blockage of the EP₂-receptor leads to a reduction and a diminishment of intestinal polyposis in APC^{A716}-knock-out mice [64]. There was an increased detection of EP₂- and EP₄-receptors in the breast tumors of COX-2-MMTV mice; therefore, it appears that the EP-receptors play an important role in mediating PG functions and in promoting carcinogenesis.

6. Role of 15-PGDH in carcinogenesis

Increased PGE₂ levels in context to mammary carcinomas are associated with an enhanced cell proliferation, invasiveness, resistance to apoptosis and angiogenesis [65, 66]. The regulation of plasma PGE₂ level results from its synthesis and its biological inactivation through 15-PGDH, the key enzyme for the biological inactivation of PGs [32]. Recent studies hypothesized 15-PGDH as a tumor suppressor gene in correlation to colon, bladder and bronchial carcinomas [67-69]. Wolf and co-workers [70] assumed antiproliferative effects of 15-PGDH in breast cancer cells. The estrogen receptor (ER) positive and well differentiated MCF-7 breast cancer cell line had an increased 15-PGDH expression compared to poorly differentiated, ER negative MDA-MB-231 cells, which express COX-2 and lead to primary breast cancer. Different studies reported that MCF-7 cells are the only breast cancer cell line with an enhanced 15-PGDH expression and low levels of 15-PGDH are accompanied by poorly prognostic factors [70]. This data attended by a microarray analysis of van't Veer and co-workers [71] supports the advice that a loss of 15-PGDH expression plays a pivotal role in the development of poorly differentiated mammary carcinomas. Data generated from genetically modified MDA-MB-231 cells that over express the enzyme and MCF-7 where 15-PGDH was knockout, corroborates the hypothesis that 15-PGDH acts as a tumor suppressor gene in breast cancer [70]. MDA-MB-231 cells showed a decreased invasiveness similar to studies in colon [67] and bronchial carcinomas [68]. Yan and co-workers [67] reported that 15-PGDH is naturally expressed in colon tissues and was dramatically reduced in colon carcinomas. The reconstitution of 15-PGDH in immunodeficiency mice prevented the colon cancer cells from generating tumors and so the authors concluded, that 15-PGDH acts as tumor suppressor and inhibits the angiogenic and proliferative effects of COX-2 *in vivo*.

7. COX-2 expression in breast cancer

Experimental immunochemical studies of COX-2 expression in breast cancer have produced varying and sometimes controversial and inconsistent data. Generally the consensus is that COX-2 is expressed by invasive ductal and lobular carcinoma and that the proportion of COX-2 positive tumors varies between studies (Table 1). In studies where poor prognostic tumor characteristics were examined, a correlation was found between prognostic parameters such as hormone receptor negativity, HER2 positivity, increased tumor size, high nuclear grade, development of distant metastases and a reduced survival rate (Table 1) [5]. Moreover COX-2 expression correlates with aromatase expression.

An explanation for the variable findings of COX-2 protein expression may be caused by the different scoring systems and cut-offs used for COX-2 immunoreactivity.

Half and co-workers [72] examined immunochemical human breast cell lines of normal and neoplastic breast tissue and detected a COX-2 expression in breast cancer cells in 43%, in DCIS in 62.5% and benign breast cells had a COX-2 expression in 81%. The more elevated COX-2 expression in DCIS in terms of a premalignant lesion might mean that an up-regulation or over-expression of COX-2 occurs relatively early in the carcinogenesis of breast cancer [72]. Contrary to Half and co-workers [72], Denkert and co-workers [73] could not detect a COX-2 expression in benign breast tissue and this may support the partially conflicting data. Denkert and co-workers [73] detected a COX-2 expression in 41% in invasive ductal breast cancer, however detected it in only 14% of invasive lobular tumors and 21% in other breast carcinomas (Table1). The COX-2 expression was associated with positive axillary lymph nodes (>50% node positive, just 16% in node negative breast cancer), extensive tumor growth (58% in tumors >20mm, in 24% in tumors <20mm), poor nuclear grading, vascular invasion and hormone receptor negativity.

Not all the studies have determined a correlation between COX-2 expression and clinicopathological parameters. Half and co-workers [72] could not demonstrate a significant correlation but Ristimäki and co-workers [74] certainly did show a significant correlation between COX-2 expression and hormone receptor negativity, extensive tumor growth, high nuclear grading and HER2 positivity. In a recently published paper by Singh-Ranger and co-workers [75], a correlation to distant metastases was described and Nassar and co-workers [76] demonstrated a correlation to nuclear grading and tumor size; however a correlation to important clinical goals such as eradicating the disease and enhancing overall survival rate have not yet been found.

Therefore COX-2 over-expression correlates in a different manner depending upon its aggressiveness the invasive potential of tumor cells, and then consequently exhibiting a higher incidence of distant metastases [40].

Reference	N =	COX-2 positive (%)			Correlation of COX-2 expression and clinicopathological parameters						
		Carcinoma	DCIS	Benign tissue	Angio-genesis	HR-status	HER2	Grading	Age	Node +	Big tumor
[77]	44	2/44 (4,5%)	-	-	Not examined						
[78]	27	7/17 (42%)	8/10 (80%)	-	Not examined						
[72]	106	18/42 (43%)	10/16 (63%)	39/48 (81%)	-	No	No	-	-	-	-
[73]	221	80/221 (36%)	-	0%	Yes	Yes	-	Yes	No	Yes	Yes
[79]	46	50%	-	-	Yes	No	No	No	No	Yes	
[74]	1576	589/1576 (37,4%)	-	-	-	Yes	Yes	Yes	-	Yes	Yes
[80]	106	90/106 (85%)	-	-	No	No	No	No	No	No	No
[81]	128	41%	-	-	Yes	-	-	Yes	-	Yes	Yes
[82]	192	40,6%	-	-	-	Yes	Yes	Yes	-	-	Yes
[83]	65	41/65 (63%)	-	-	-	Yes	Yes	-	-	-	-
[76]	43	41/43 (95%)	-	-	-	No	No	Yes	-	-	Yes

Table 1. Immunochemical examinations of COX-2 expression and correlation with selected clinicopathological parameters in breast tissue.

Transcriptional studies have also revealed a distinct variation in their results regarding COX-2 expression. The detection rate varies between 50 and 100% in the literature (Table 2). There is a comparable relationship between COX-2 immunoreactivity and mRNA expression in tumor tissue [77]. Zhao and co-workers [84] demonstrated an increased mRNA expression in hormone receptor positive breast cancer; a result that was confirmed by Singh and co-workers [85] in breast cancer with positive progesterone receptors. However, only a small number of studies have examined the correlation between mRNA expression and clinico-pathological parameters. These results are summarized in Table 2.

Reference	N =	COX-2-mRNA positive (%)	Clinicopathological correlation of COX-2 with						
			Angio-genesis	HR-status	HER2	Grading	Age	Node +	Big tumor
[86]	40	40/40 (100%)	Not examined						
[72]	9	9/9 (100%)	Not examined						
[87]	7	7/7 (100%)	-	-	-	-	-	-	-
[88]	20	10/20 (50%)	-	-	-	-	-	-	-
[85]	18	18/18 (100%)	Not examined	Yes (PR)	Not examined				
[84]	30	27/30 (90%)	-	Yes (ER+)	-	Not examined			

Table 2. COX-2 mRNA expression and correlation with selected clinicopathological parameters in breast cancer.

These results are contrary to the immunochemically evaluated data, which show an association to hormone receptor negative tumors. This could be explained because before the genetic information of COX-2 is translated into a biologically active protein, COX-2 mRNA is post transcriptionally modified in the nucleus. Thus, we speculate that the COX-2 mRNA is destabilised by its AU rich sequences and no COX-2 protein is generated. Therefore, the correlation between the hormone receptor status and COX-2 mRNA levels is not obvious in studies where the COX-2 protein expression was investigated [5]. There are some well known factors which affect the COX-2 mRNA levels like interleukin-1 stabilises the highly unstable COX-2 mRNA transcript [89], however steroids may destabilise the COX-2 mRNA [90]. Furthermore, it might be possible that genetically different subtypes of breast cancer express COX-2 and are then associated with both hormone receptor-negative and receptor-positive tumors [91]. Additionally, Ristimäki and co-workers [74] reported that hormone receptor-positive patients who express COX-2 had a poor survival rate.

7.1 COX-2 and hormone receptors

There is concurrent evidence about the interaction of PGE₂/COX-2 and the estrogen receptor signalling pathway. For example, COX-2 expression is correlated with the expression of the aromatase [92] and in vitro studies support this data. It has been shown that COX-2 promotes the aromatase transcription, whereas COX-2 inhibitors diminish it [93]. Based on the elevated synthesis of prostaglandins in cells that express COX-2, the aromatase expression and activity is increased in breast cells [94, 95]. Expression of aromatase leads to estrogen production and from cell line studies; we know that hormone receptor expression can be induced by sex steroid hormones [96]. All the data supports the

close correlation between COX-2 and hormone receptors. Wolf and co-workers [70] reported a link between the estrogen signalling pathway and 15-PGDH by a negative feedback mechanism. High levels of this hormone reduced the 15-PGDH expression but the activity of the ERE (estrogen responsive element) and the activity of the aromatase increased. New studies suggest a synergism between selective COX-2 and aromatase inhibitors.

7.2 Results from in vivo studies

The impact of COX-2 in carcinogenesis of breast tumors has been shown in transgenic mice models [97]. It has been reported that the over-expression of COX-2 in breast tissues is associated with decreased BAX and Bcl-xL (pro apoptotic) and increased Bcl-2 (anti apoptotic) protein levels. Therefore, the author suggested that induction of carcinogenesis is COX-2 dependent [97]. In contrast, the resistance to apoptosis is associated with increased COX-2 levels [98]. The importance of COX-2 in correlation to the tumor formation has been investigated in COX-2 knockout mice. The COX-2 knockout mice lead to an 86% reduction of intestinal adenoids [99].

7.3 COX-2 and tumorigenesis

The expression of COX-2 is regulated by post-transcriptional and -translational mechanisms. Different cytokines, growth factors and oncogenes have been shown to induce the COX-2 expression which is associated with carcinogenesis [46, 100].

7.3.1 Influence of COX-2 on angiogenesis and apoptosis

Angiogenesis is the development of new blood vessels and is an important factor in tumor proliferation, invasion and metastasis. Davies and co-workers [101] showed a significant positive correlation between COX-2 expression and the endothelial surface marker CD31. Other reports confirmed a positive correlation between COX-2 and the vascular endothelial growth factor (VEGF) [102, 103]. During carcinogenesis COX-2 modulates the neoangiogenesis and seems to stimulate the production of proangiogenic factors such as VEGF, basic fibroblast growth factor (bFGF), transforming growth factor 1 (TGF1), platelet derived growth factor (PDGF) and endothelin [104, 105]. The application of selective COX-2 inhibitors decreased the angiogenesis in different vivo models [106]. Recently, an angiogenesis independent mechanism, so called Vasculogenic Mimicry (VM), was described where poorly differentiated breast cancer cells were nourished without the mechanisms of classic neoangiogenesis [107-109]. VM is a phenomenon of vessel formation of epithelial tumor cells without any participation of endothelial cells and it is a mechanism independent of or simultaneous to neoangiogenesis thus ensuring the tumor perfusion [110]. Hence, the VM might be an important factor for new antiangiogenic therapeutical approaches. The existence of VM in breast cancer patients is associated with a poor 5-year survival rate compared to patients without. [111]. Basu and co-workers [112] reported that highly invasive MDA-MB-231 breast cancer cells and the less invasive subtype MDA-MB-435 cells that over express COX-2, formed new micro vessels. In contrast, non-invasive MCF-7- and ZR-75-1-breast cancer cells which had a lower COX-2 expression did not. The application of the COX-2 inhibitor celecoxib (40 und 60 $\mu\text{mol/l}$, $p > 0.001$), inhibits the formation of new vessels. This effect was restored with PGE₂. This data was confirmed by an in vivo xenograft model. VEGF, GRO, IL-6, IL-8, TIMP1, TIMP2 were the main angiogenic proteins which were inhibited by celecoxib [112].

7.4 Breast cancer and NSAIDs

The rationale for using NSAIDs is their non-selective (ASS, ibuprofen etc.) or selective (COX-2 inhibitors such as celecoxib) suppression of the COX-system. In a meta-analysis consisting of 14 epidemiological studies (6 cohort studies and 8 case control studies) breast cancer risk was reduced by 18% due to constant intake of NSAIDs [113]. An extensive Canadian study including 5882 patients reported a reduction of breast cancer incidence by 24% due to the NSAIDs intake for 2 – 5 years [58]. Another case control study demonstrated a 40% reduction after 5 years of NSAIDs intake [57]. These results seemingly justify the preventive use of NSAIDs, however, contrarian results were delivered by the Nurses Health Study. This trial showed no difference during the intake of ASS (100mg) in neither women with breast cancer nor in healthy women [114]. On the contrary it was in patients with colon cancer who led the continuous intake of NSAIDs to a reduction of incidence in 40-50% [56, 115, 116].

Data of animal models supports the use of selective COX-2 inhibitors both for the therapeutic and preventive uses. For instance, the use of celecoxib in rats led to an averaged downsizing of breast tumor volume by 32%, however, a tumor volume enlargement of 518% was observed in the control group [117]. Harris and co-workers [118] examined the influence of celecoxib in 120 rats. Three groups of rats were formed. In one group the food was enriched with celecoxib. The other two groups obtained either ibuprofen or nothing. After seven days 7,12-Dimethylbenz(a)anthracene (DMBA) was applied intragastrically and the described food was continued for another 105 days. A distinct reduction in tumor incidence, variety and tumor volume was shown in the celecoxib treated group [118]. In a recently published paper Barnes and co-workers [119] could induce breast tumors in mice by injecting estrogen-positive MCF7/HER2-18- and estrogen-negative MDAMB231 breast cancer cells. The application of celecoxib resulted in a significant growth reduction of the MCF7/HER-18 tumors (58.7%) and the MDAMB231 tumors (46.3%) in comparison to the control group. Therefore, celecoxib dropped the COX-2 expression and enhanced the apoptosis significantly [119]. Yoshinaka and co-workers [120] also showed that the use of celecoxib significantly reduced tumor sizes, increased apoptosis and that a reduced DNA synthesis in the tumor tissue of mice induced breast carcinomas. Moreover the neoangiogenesis was influenced as VEGF-A-mRNA levels were found to be reduced [120].

7.5 COX-2-inhibitors in systemic treatment

Several studies have evaluated the significance of COX-2 inhibitors in combination with systemic treatment. A phase II study observed a clinical benefit of 47.5% for the combination of capecitabine and celecoxib in patients with metastatic breast cancer. The combination was well tolerated [121].

Recently published data about COX-2 and its significance on the aromatase and influence on the female hormonal balance are of strong interest. Besides finding an increased effect on estrogen synthesis in malignant breast tissue, a strong correlation between COX-2 and aromatase mRNA expression were found. This data supports the assumption that COX-2 is able to regulate aromatase activity in breast tissue [92]. A possible synergism between COX-2 and aromatase-inhibitors is even more interesting and so a prospective randomised phase III multicenter trial (REACT-trial) was conducted that included primary breast cancer patients in order to evaluate the combination of celecoxib and exemestane, an aromatase

inhibitor, in an adjuvant setting. The combination of celecoxib and exemestane was already well tolerated and had shown a clinical benefit of 74% [122] or had led to a benefit extension (median 96.6 weeks vs. 49.1 weeks) in patients with metastatic breast cancer [123].

Other malignancies were also proven on the benefit of selective and non-selective COX-2 inhibitors in combination with other compounds such as chemotherapy [124, 125], tyrosinekinase inhibitors [126] and other new approaches [127]. Some of them are encouraging, like the results of the ASCENT trial [124] and some are disappointing. Further work is required to establish how NSAIDs can be best applied for therapeutic benefit.

8. Vitamin D

8.1 Vitamin D metabolism

Vitamin D, a secosteroid hormone, is assimilated by food (milk, fish, liver), multi-vitamin preparations and dietary supplements [128]. Vitamin D is also synthesised from 7-dehydrocholesterol and provitamin D₃ after skin exposure with sunlight (ultraviolet spectrum 290-315nm) [129]. Based on its animal or herbal origin, there are two existing vitamin D metabolites: cholecalciferol (vitamin D₃) and ergocalciferol (vitamin D₂) [17], which is less efficient in increasing the 25-hydroxyvitaminD [25(OH)₂D₃] serum levels [130]. Cholecalciferol attains to the liver through the bloodstream and is transformed to 25(OH)₂D₃ (25-hydroxyvitamin D₃, calcediol, 25-hydroxycholecalciferol) by a hydroxylation on the C25 position [131, 132]. 25(OH)₂D₃, a circulating metabolite, correlates with the vitamin D balance. The hydroxylation of cholecalciferol on the C25 position is inadequately regulated. 25(OH)₂D₃ level increased with the vitamin D intake, therefore, the 25(OH)₂D₃ serum level is normally used as an indicator of the vitamin D balance [133]. The serum level range of 25(OH)₂D₃ is between 10 and 50 ng/ml and round about 30pg/ml for 1,25(OH)₂D₃ [134]. 25(OH)₂D₃ is renally converted to the biologically active metabolite 1,25-dihydroxycholecalciferol [1,25(OH)₂D₃] by the 1- α -hydroxylase (CYP27B1). 1,25(OH)₂D₃ is 100-1000 fold more active than the other natural metabolites [135]. The 1- α -hydroxylase, a mitochondrial enzyme, which belongs to the P450 enzyme family is located in the renal proximal tubule. Besides the renal expression of the enzyme, many studies reported an extra renal expression of 1- α -hydroxylase and thus an extra renal synthesis of 1,25(OH)₂D₃. This enzyme has been detected in many cell types and tissues, e.g. prostate, breast, lung, pancreas, parathyroid and monocytes [136]. The extra renal synthesised 1,25(OH)₂D₃ has cell specific functions and as a result acts as local auto- and paracrine factors. In this context, many extra renal effects of 1,25(OH)₂D₃, e.g. cell cycle arrest, induction of apoptosis and cell differentiation, have been reported [136]. The fine tuned activity of 1- α -hydroxylase correlates inversely with the calcium metabolism and thus the circulating levels of 1,25(OH)₂D₃ correlates inversely with the ingested amount of calcium [137]. 1,25(OH)₂D₃ Serum levels are maintained in pmol/l range by a classic negative feedback mechanism. The decrease of calcium or phosphate levels leads to an increase of the 1- α -hydroxylase activity and an enhanced synthesis of 1,25(OH)₂D₃ which in turn promotes the intestinal resorption of calcium and phosphate and the calcium mobilisation from the bones. The activity of 1- α -hydroxylase decreased with increasing 1,25(OH)₂D₃ levels, which leads to 24 hydroxylase activation. This enzyme degrades 1,25(OH)₂D₃ to its inactive metabolite 24,25(OH)₂D₃ [138, 139], which is subsequently converted to calcitroic acid and excreted. Hence, the nutritive intake of

calcium directly regulates 1- α -hydroxylase activity and indirectly modifies parathormone levels. This hormone produced in the parathyroids increases the phosphate excretion in the proximal tubule but promotes the sodium, potassium and calcium resorbtion in the distal tubule. Under normocalcemic conditions, the activity of 1- α -hydroxylase is inhibited. These regulations are necessary to synthesize 1,25(OH)₂D₃ even though much is needed to cover the calcium and phosphate demand and to avoid a 1,25 (OH)₂D₃ intoxication [139]. The circulating vitamin D level depends on many different factors such as: the vitamin D content in either the ingested nutrition or the dietary supplements, and the endogenous production and degeneration via vitamin D metabolising enzymes. A simplified scheme of vitamin D metabolism is presented in figure (2).

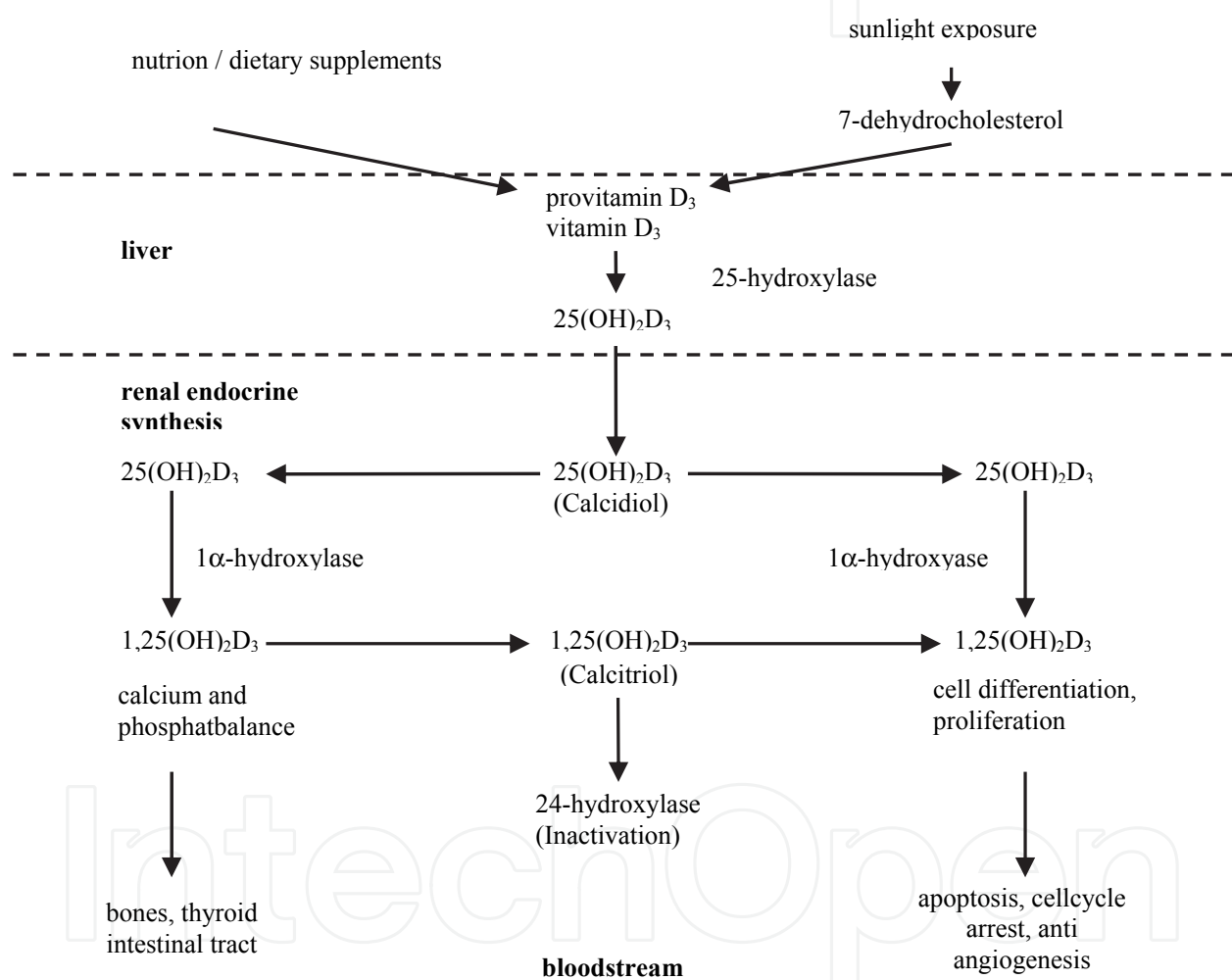


Fig. 2. Simplified scheme of the vitamin D metabolism. Vitamin D (food intake, synthesis in skin) is metabolised in liver to 25(OH)₂D₃, then via the renal 1- α -hydroxylase (endocrine signalling pathway or extra renal in tissues (autocrine/paracrine signalling pathway) to 1,25(OH)₂D₃.

8.2 Extrarenal vitamin D metabolizing enzymes

The biologically active metabolite is produced after a series of hydroxylations through cytochrome P450 enzymes which belong to the cytochrome p450 super family. The different enzymes are handled as follows:

8.2.1 1- α -hydroxylase (CYP27B1)

The 25-hydroxyvitamin-D₃-[25(OH)₂D₃]1- α -hydroxylase (1- α -hydroxylase) is encoded by the CYP27b1 gene and catalyzes the synthesis of 1,25(OH)₂D₃ from 25(OH)₂D₃. 1,25(OH)₂D₃ is the most important regulator of the enzyme that leads to a decreased enzyme expression. The regulation of the extra renal 1- α -hydroxylase depends on local factors like cytokines (interleukins, interferones and tumor necrosis) and growth. The optimal 1,25(OH)₂D₃-level tuning mechanism is not yet completely understood [139]. The reduced expression of the enzyme suggests the involvement of a negative vitamin D responsive element (VDRE) and Turunen and co-workers [140] showed that the enzyme's response to 1,25(OH)₂D₃ is a cell specific event with participation of many VDREs. The suppression of cell proliferation, the induction of apoptotic events and the modulation of immune responses are counted among the classical features of 1,25(OH)₂D₃. After binding to the vitamin D receptor, 1,25(OH)₂D₃ is able to arrest the cell cycle of a tumor cell in the G1-G0 phase via specific mechanisms [139]. In prostate and colon cancer the tumor protective effects of vitamin D is correlated to vitamin D deficiency [141]. Much data reports that both the renal and extra renal 1- α -hydroxylase are based on the expression of the same gene product. In contrast to the renal 1- α -hydroxylase, the extra renal enzyme is not subjected to the auto regulation as mentioned above [136, 142]. Therefore the enzyme's tissue specific expression might be a key mechanism in connecting the vitamin D metabolism to the anticarcinogenic effects of 1,25(OH)₂D₃.

Although the enzyme's cytokine and growth factor related regulation is not completely understood, it has been shown that different cytokines stimulate the 1- α -hydroxylase in different cell types [139, 143-146]. Another potential mechanism of gene regulation is the incidence of different gene polymorphisms [147] and inactive variants due to alternative splicing of the 1- α -hydroxylase mRNA. But this mechanism's function is not completely clarified. Alternative splicing within the post transcriptional modification is a normal process of gene expression in breast cancer cells and based on the pre mRNA, different mature mRNAs are generated when introns or exons are deleted or added. Thus, the translation of these mRNAs leads to different enzyme proteins, however, mis-spliced mRNAs are usually quickly degraded although it appears that this mechanism has failed in various cells. It has been reported that different protein variants of 1- α -hydroxylase might have diverse biological functions. Fischer and co-workers [148] showed 6 different variants of the enzyme in MCF10F via nested touchdown PCR, but in MCF-7, these variants appeared weakly expressed. Based on this data, the authors concluded that because alternative splicing regulates the level of the active enzyme extrarenally it therefore regulates the local production of 1,25(OH)₂D₃ [149]. The activity of the extrarenally expressed 1- α -hydroxylase is an important factor of the tumor pathophysiology because of an accumulation of 1,25(OH)₂D₃ in many tissues. Studies of prostate [150, 151], colon [152-154] and breast cancer [148, 155, 156] have shown the expression of 1- α -hydroxylase in healthy as well as in malignant tissues. Thus, 1,25(OH)₂D₃, which is produced extrarenally might have autocrine behaviour to protect cells against transformation and supports the suggestion of its carcinoprotective effects. Accordingly, low 1- α -hydroxylase levels correlate with the risk of prostate-, colon- [157] or breast cancer [158, 159]. Moreover, the extra renal production of 1,25(OH)₂D₃ inhibits cell proliferation and promotes cell differentiation in xenograft models [160]. Besides the expression of the 1- α -hydroxylase in breast [155, 161], endometrial [162], cervical and ovarian carcinomas [163], the induction of the enzyme has also been shown in

lymphomas [164] and dysgerminomas [165]. In these reports, the local synthesis of $1,25(\text{OH})_2\text{D}_3$ was mediated by the $1\text{-}\alpha$ -hydroxylase expression of tumor associated macrophages. The expression of the enzyme mammary gland tissue occurs in lobules and ductus, primarily in the cancer tissue and invasive tumor cells and inflammatory infiltrate. Thus, it might be possible that the enzyme activity and the vitamin D receptor (VDR) expression are considerably higher than in the benign tissue compared to aggressive tumor cell lines (MCF-7res, MDA-MB231) [166]. Townsend and co-workers [166] compared breast cancer and benign tissue samples via reverse transcription PCR. They reported a 27-fold induction of the $1\text{-}\alpha$ -hydroxylase expression and 7-fold induction of the VDR expression in tumor samples. Because 80% of the tumor tissues had an increased $1\text{-}\alpha$ -hydroxylase and VDR, they concluded that there was a closed coupling of both gene products. These results are in compliance with Segersten and co-workers [161]. The capacity of $1\text{-}\alpha$ -hydroxylase to synthesize $1,25(\text{OH})_2\text{D}_3$ within the mammary gland parenchyma results in, on the one hand, the available amount of $25(\text{OH})_2\text{D}_3$, and is dependent on sun light exposure and the season [167-170] - normally there is no definite correlation between $25(\text{OH})_2\text{D}_3$ and $1,25(\text{OH})_2\text{D}_3$ - yet on the other hand, the level of the extra renal production of $1,25(\text{OH})_2\text{D}_3$ is limited by the expression of the $1,25(\text{OH})_2\text{D}_3$ - decomposing enzyme 24-hydroxylase, which is stimulated by $1,25(\text{OH})_2\text{D}_3$ in VDR expressing tissues. Based on the missing correlation of 24-hydroxylase and VDR or the $1\text{-}\alpha$ -hydroxylase in breast cancer tissues, it seems that 24-hydroxylase is independently regulated. Kemmis and co-workers [171] demonstrated the expression of a functioning VDR and an inhibition of proliferation via $1,25(\text{OH})_2\text{D}_3$ in benign breast cells and MCF-7. The VDR expression in HMEC breast cells was higher than in MCF7 cells. Furthermore, the authors showed an expression of $25(\text{OH})_2\text{D}_3$ metabolizing $1\text{-}\alpha$ -hydroxylase and 24-hydroxylase in these cell types, whereas the $1\text{-}\alpha$ -hydroxylase expression was higher in MCF-7. In contrast to renal HKC8 cells, the expression of $1\text{-}\alpha$ -hydroxylase was not inhibited by $1,25(\text{OH})_2\text{D}_3$. Based on the strong induction of the 24-hydroxylase through the $1,25(\text{OH})_2\text{D}_3$ application, the authors showed that MCF7 cells were more sensitive in response to $1,25(\text{OH})_2\text{D}_3$ compared to HKC-8 and HMEC cells. From this data, they concluded that there is a functional vitamin D receptor as well as intact signalling transduction pathways in MCF-7 cells. The data suggests that the synthesis of $1,25(\text{OH})_2\text{D}_3$ and the activation of the VDR inhibits the cell proliferation in breast cells. Thus, the treatment of benign breast cells with $25(\text{OH})_2\text{D}_3$ leads to an activation of the VDR transcription and the regulation of its target genes (CYP27B1, CYP24) and finally to an inhibition of cell proliferation. According to that, CYP27B1 lords it over CYP24 which means a transformation of $25(\text{OH})_2\text{D}_3$ to $1,25(\text{OH})_2\text{D}_3$. Kemmis and co-workers [171] have shown for the first time that physiological $1,25(\text{OH})_2\text{D}_3$ levels (30-100nmol/L) are able to inhibit cell proliferation in benign HMEC cells and in MCF7 breast cancer cells. Interestingly, aging process and the associated lack of estrogens correlate with decreased $25(\text{OH})_2\text{D}_3$ levels. The reason is that the ability of estrogen to stimulate the renal CYP27B1 activity [172]. Accordingly, the lack of estrogens leads to decreased $1,25(\text{OH})_2\text{D}_3$ levels and presents the highest risk for breast cancer in postmenopausal women [273].

8.2.2 24-hydroxylase (CYP24)

The 25-hydroxyvitamin D_3 -24 hydroxylase (24-OHase, 24-hydroxylase) encoded by the CYP24 gene is induced by $1,25(\text{OH})_2\text{D}_3$ in breast cell lines where the enzyme is time and dose dependently stimulated by $1,25(\text{OH})_2\text{D}_3$ [174]. An increased enzyme expression in

ovarian, cervical and breast cancer compared to healthy tissue samples has been shown by immunochemistry and real time PCR [163]. In contrast, Townsend and co-workers [166] showed a 4-fold increase of the enzyme expression in malignant breast tissues compared to healthy tissue samples using the same technique. Additionally, the expression of 24 hydroxylase increased in breast cancer cells foremost in hormone resistant MCF-7 Res and the aggressive MDA-MB231 cells compared to benign MCF-12A cells [166]. Kemmis and co-workers [171] reported the highest 24 hydroxylase expression in MCF-7 cells and Segersten and co-workers [161] showed a 2-fold enzyme expression in tumor tissues compared to benign tissue samples. The authors concluded that the conversion of 1,25(OH)₂D₃ into the inactive metabolite 1,24,25(OH)₃D₃ is significantly higher in malignant tissues. Furthermore Townsend and co-workers [166] detected the enzyme only in breast cancers with an increased 1- α -hydroxylase and VDR expression. Further analysis showed that in a healthy tissue sample expression of 24-hydroxylase correlated with both 1 α -hydroxylase and VDR. There was not such a correlation in breast tumors. Hypothetically, the 24 hydroxylase acts as a part of a well organized feedback mechanism and is transcriptionally modulated to increase the local 1,25(OH)₂D₃ and VDR level [166]. The synthesis of 1,25(OH)₂D₃ via the 1- α -hydroxylase has been shown in benign and malignant mammary gland tissues but this mechanism's efficiency in tumor tissues might be affected by a dysregulated 24 hydroxylase expression.

8.3 Vitamin D-receptor (VDR / mVDR)

The vitamin D receptor (VDR) is an ubiquitarily expressed steroid hormone receptor. Like other steroid, thyroid and retinoid receptors, the VDR is a member of the nuclear hormone receptor family. The receptor binds to its ligand 1,25(OH)₂D₃, interacts with other receptors by dimerization and binds as homodimers or heterodimers to specific DNA sequences. So called vitamin D responsive elements (VDRE) recruit additional co-activators (such as SRC-1, GRIP-1/TIF2, ACTR) and interact with the transcriptional processing order to initiate or inhibit the transcription of its target genes [25]. It is well known, that steroid receptors consist of different variants with distinct specificities. Sunn and co-workers [175] described an N-terminal variant of the VDR. 1,25(OH)₂D₃ mediates its genomic effects as a VDR ligand and via the directed binding to the VDRE [176]. Besides its function in bone metabolism and in the calcium/phosphate balance, the VDR interacts with different signalling pathways, e.g. with p21, a cyclin dependent kinase inhibitor which is involved in cell cycle regulation and inhibition of the cancer cell proliferation [26]. There are some suggestions about the existence of a membrane VDR (mVDR) [177]. The mVDR mediates its signals through the change of the intracellular calcium concentrations and through interactions with the protein kinase C and enzymes of the MAPK family [178-183]. Although the mVDR seems unrelated to the nuclear VDR, Marcinkowska and co-workers [184] reported an interaction of both receptors. The function of this mechanism is not clearly defined and the cloning of the mVDR has failed until today.

Many studies reported that extra renal VDR expression is associated with the non-classical effects of 1,25(OH)₂D₃. The VDR expression has been shown in healthy breast tissues and in more than 80% of the breast cancer tissues [185]. The natural ligand of the VDR, 1,25(OH)₂D₃ and many new developed synthetic vitamin D analogues inhibit cell proliferation and induce apoptosis in breast cancer cell lines [186, 187]. Furthermore, in animal models, vitamin D analogues retard the tumor growth and lead to a regression of breast tumors [12].

8.4 Vitamin D-receptor gene polymorphism

The gene that encodes the VDR has various polymorphisms. It has been hypothesised that the genetic VDR polymorphism influences the breast cancer risk due to its potential effects on VDR gene expression and protein function [188, 189]. Many polymorphisms of the VDR gene have been identified and several, such as FOK1, Bsm1, APA1, TAQ1 and Poly(A) are well analysed [190, 191]. The studies that were conducted had conflicting results [191]. Curran and co-workers [192] showed a significant association of the VDR polymorphism APA1 und TAQ with the breast cancer risk. A significant increased breast cancer risk in women with the ff genotype FOK1 was observed by Chen and co-workers [193]. Sinotte and co-workers [194] detected a significant link between familial breast cancer disposition and FOK1. Other data came from Trabert and co-workers [195] who found a correlation between a higher breast cancer risk and the genotype Bsm1 bb in postmenopausal women although there is also published data without any evidence for a link between VDR polymorphisms and breast cancer risk [196-199]. An analysis of the last 13 published studies in which different VDR polymorphisms and its relation to breast cancer were examined leads to the suggestion that the modification of breast cancer risk is associated with certain VDR polymorphisms and therefore $1,25(\text{OH})_2\text{D}_3$ might modify the risk of breast cancer [200]. A recently published paper by McCullough and co-workers [201] presented certain VDR gene polymorphisms associated with a decreased breast cancer risk in women who ingested high doses of calcium (no calcitriol) concluding that nutritive influences might modify the link between gene polymorphisms and breast cancer. This data could shed light on breast cancer risk evaluation or could even be used in a predictive manner to answer the question about which women are strongly endangered to develop distant metastases.

9. Calcium

Like vitamin D, humans ingest calcium through food or dietary supplements. 99% of calcium is bound as hydroxyl phosphatide in bones and teeth [202]. Only 1% calcium is extracellularly located. Plasma levels of calcium (Ca^{2+}) are limited by the intestinal absorption, the renal secretion and the reabsorption. Additionally, the skeletal calcium storage and resorption keep the plasma levels of calcium in a closed range (3.5–5mmol/l) [202].

10. Vitamin D, calcium and breast cancer risk

10.1 Dietary and supplemental vitamin D intake

For $1,25(\text{OH})_2\text{D}_3$ several studies have shown both an antiproliferative effect and an inhibition of angiogenesis in malignant and healthy breast cancer cells [17, 185, 203-206]. In mouse models, an increased intake of vitamin D led to the suppression of epithelial hyperproliferation and tumorigenesis of the mammary gland that was caused by rich nutrition [207, 208].

Last but not least, it has been proven by the First National Health and Nutrition Examination Survey Epidemiologic Follow-Up Study that sunlight exposure is inversely correlated with breast cancer risk [209, 210]. In this study, the female population in the north-eastern parts of the U.S. have a higher risk of contracting breast cancer compared to the other states of the U.S. This leads to the suggestion that sunlight induced vitamin D production has a positive influence in avoiding breast cancer [20].

In contrast, in the Nurses' Health Study, there was an inverse association between vitamin D intake and breast cancer risk among premenopausal women but no association among postmenopausal women [20]. Consistent with this observation, a study published a few years ago was based on the Cancer Prevention Study II Nutrition Cohort and observed no associations between breast cancer and total and dietary vitamin D intakes among postmenopausal women [211]. Another Italian study recently showed an inverse association between vitamin D intake (in the study >143 IU) and breast cancer in 2569 breast cancer patients [212]. Two other studies that concentrated on vitamin D deficiency and its susceptibility for breast cancer incidence approved that a deficiency conditional on nutrition in adolescence does not lead to an increased breast cancer risk [213, 214]. The proper dose of vitamin D remains unclear and a recommendation does not exist, however a meta-analysis gives evidence towards a dose of >400 IU per day to reduce breast cancer risk [213].

10.2 Role of vitamin D in breast cancer

To date, there have been several epidemiologic studies of the association between vitamin D and breast cancer risk, however, their results have not been consistent. Several studies observed an association between 25(OH)₂D₃ plasma levels and breast cancer incidence [19, 215-217]. The predictive value of 25(OH)₂D₃ plasma levels depends upon the time they have been measured. Plasma levels that have been measured within a few years before breast cancer diagnosis are less predictive than plasma level measured many years before [217]. Furthermore, plasma levels that have been measured around 15 years before diagnosis do not have any aetiological value for the genesis of breast cancer [216].

Bertone-Johnson and co-workers [217] found a marginally significant reduction of breast cancer risk in women >60 years who had elevated 25(OH)₂D₃ and 1,25(OH)₂D₃ plasma levels. In contrast, published data by Shin and co-workers [20] demonstrated a significantly decreased breast cancer incidence in premenopausal, but not in postmenopausal women, who had continuous vitamin D intake.

Furthermore, a case control study observed that women with plasma 25(OH)₂D₃ concentration <50nmol/l had >5 times higher risk of breast cancer than those with plasma concentrations exceeding >150 nmol/l [158]. Janowsky and co-workers [19] also showed an inverse association between 1,25(OH)₂D₃ plasma levels to the point of diagnosis and breast cancer risk in patients with breast cancer. However, there was no difference in 1,25(OH)₂D₃ plasma levels between patients with breast cancer and those with DCIS. The authors suggested that the grade of invasion was not correlated with the extent of 1,25(OH)₂D₃ level. Another nested case-control study with 96 breast cancer cases and 96 controls found no association between prediagnostic 1,25(OH)₂D₃ levels and levels at the time of diagnosis and breast cancer risk among postmenopausal women [216].

The circulating concentration of 25(OH)₂D₃ is considered to be an excellent measure of the availability of vitamin D from the diet, supplements and from synthesis in the skin [218]. Its potential importance in breast carcinogenesis is due to the fact that 25(OH)₂D₃ can be metabolised to 1,25(OH)₂D₃ by 1- α -hydroxylase in breast tissue [155]. Thus, 25(OH)₂D₃ levels may be more representative of intracellular levels of 1,25(OH)₂D₃ than circulating levels of 1,25(OH)₂D₃ [217].

To date, no studies have been published investigating intracellular or tissue levels of 1,25(OH)₂D₃ and 25(OH)₂D₃ in association with breast cancer risk.

10.3 Dietary and supplemental calcium intake

Many studies about the importance of calcium and its association to breast cancer have already been published. Most of them are case-control studies and nearly all of them are relatively small and there is insufficient documentation regarding risk factors for breast cancer in multivariate analyses.

Calcium is participating on carcinogenesis via regulation of cell proliferation, differentiation and apoptosis [219-221]. Cell proliferation and differentiation of breast cells can be increased by elevated calcium levels [208, 222, 223]. Boyapati and co-workers [224] observed a non-significant inverse association between calcium intake and breast cancer risk among pre- and postmenopausal women but the Nurses' Health Study has shown this association only for premenopausal women [20].

The anti-carcinogenic effects of calcium are last but not least, mediated by vitamin D, therefore calcium is one of the key mediators of the vitamin D induced apoptosis in breast cancer cells [208].

11. Calcitriol and prostaglandins in cancer

The stimulation of the renal calcitriol [1,25(OH)₂D₃] synthesis in vitro is well known as well as the inhibition of acetylsalicylic acid as a non-selective NSAID [225]. This justifies the clinical use of NSAIDs in treating arthritis for example. Hayes and co-workers [226] observed an inhibition of calcitriol synthesis caused by PGE₁ and PGE₂ in synovial fluid macrophages from arthritic joints and with that he proved the link between vitamin D and prostaglandin metabolism. Several published studies have proven the anti-carcinogenic effects shown in different signalling pathways on prostate cancer cells [227-229]. The team around David Feldman examined the influence of calcitriol in established human prostate cancer cell lines (androgen dependent LNCaP cells and androgen independent PC-3 cells) and in primary normal prostatic epithelial cells derived from normal and cancerous human prostate tissue. They showed that calcitriol regulates biologically active prostaglandin levels and prostaglandin actions by three mechanisms: calcitriol suppresses the COX-2 expression and moreover it up-regulates the expression of 15-PGDH. This dual influence of calcitriol was associated with a decrease of PGE₂ secretion in prostate cancer cells. Calcitriol reduces the mRNA expression of prostaglandin receptors EP₂ and FP, additionally a mechanism to inhibit the biological activity of prostaglandins.

The combination of calcitriol and NSAIDs led to a significant growth inhibition in prostate cancer cells via its synergistic effects. These findings might postulate that calcitriol and NSAIDs are definitely a useful combination in chemo preventive and/or therapeutic strategies in prostate cancer [230]. Unpublished own data support these results as we showed an inverse correlation between VDR- and COX-2 expression in breast cancer cells and a downregulation of COX-2 and an upregulation of 15 PGDH by calcitriol. Therefore we propose that these findings and suggest a possible link between VDR, associated target genes and the prostaglandin metabolism.

12. Concluding remarks

In conclusion, there is promising preclinical data inhibiting COX-2 in breast cancers, therefore the chance exists to innovatively disturb carcinogenesis of those gynecological oncological neoplasms Phase II trials have already been conducted to clear the safety of a

celecoxib treatment in metastatic breast cancer. Furthermore, calcitriol and calcium have shown anti-carcinogenic effects in experimental studies and several epidemiological studies have demonstrated an inverse association between vitamin D and calcium intake and breast cancer. Other studies have detected an inverse association between plasma and serum levels and breast cancer risk. Experimental studies support the hypothesis that the reduction of breast cancer risk is more significant among premenopausal women than among postmenopausal women and mPGES-1 and EP receptors might be important targets for the development of new anti-inflammatory and antiproliferative tumor therapies.

Questions that remain unanswered are: has calcitriol as antiproliferative effects in breast cancer as was proven in prostate cancer? Does a link exist between vitamin D and prostaglandin metabolism in breast cancers? These questions have to be answered as the increasing incidence of breast cancer have yet to be solved. Innovative treatment strategies fall on fruitful ground. Thus we need further studies that elucidate the importance of COX-2 inhibitors in the preventive as well as in the adjuvant settings in breast cancers and finally that will evaluate the promising importance in the neoangiogenesis in detail.

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14. Abbreviations

bFGF	=	basic fibroblast growth factor
COX	=	cyclooxygenase
DCIS	=	ductal carcinoma in situ
DMBA	=	Dimethylbenz(a)anthracene
ER	=	estrogen receptor
ERE	=	estrogen responsive element
FIGO	=	Fédération Internationale de Gynécologie et d'Obstétrique
GRO	=	growth related protein
HER2	=	human epidermal growth factor receptor 2
IL	=	interleukin
NSAID	=	non-steroidal anti-inflammatory drug
mPGES-1	=	microsomal prostaglandin E synthase-1
mRNA	=	messenger ribonucleic acid
PCR	=	polymerase chain reaction
PDGF	=	platelet derived growth factor
PG	=	prostaglandin
PGDH	=	prostaglandin dehydrogenase
PPAR	=	peroxisomes proliferator-activated receptor
TGF1	=	transforming growth factor 1
TIMP	=	tissue inhibitor of matrix metalloproteinase
VDR	=	vitamin D receptor
VDRE	=	vitamin D responsive element
VEGF	=	vascular endothelial growth factor
VM	=	vasculogenic mimicry

15. References

- [1] American Cancer Society. *Cancer facts and figures 2005*. Atlanta, American Cancer Society 2005, p. 9. (<http://www.cancer.org/downloads/STT/CAFF2005f4PWSecured.pdf>)
- [2] Ozols, R. F.; Rubins, S. C.; Dembo, A. J.; Robboy, S. In *Gynecologic oncology: Epithelial ovarian cancer*; Hoskins, W. J.; Perez, C. A.; Young, R. C.; Ed.; Lippincott Williams & Wilkins: Philadelphia, 1992 pp. 731-781.
- [3] FIGO (International Federation of Gynecology and Obstetrics) annual report on the results of treatment in gynecological cancer. *Int. J. Gynaecol. Obstet.* 2003, *83 Suppl 1: ix-xxii*, 1-229.
- [4] Ferrandina, G.; Lauriola, L.; Zannoni, G. F.; Fagotti, A.; Fanfani, F.; Legge, F.; Maggiano, N.; Gessi, M.; Mancuso, S.; Ranelletti, F. O.; Scambia, G. Increased cyclooxygenase-2 (COX-2) expression is associated with chemotherapy resistance and outcome in ovarian cancer patients. *Ann. Oncol.* 2002, *13*, 1205-1211.
- [5] Singh-Ranger, G.; Salhab, M.; Mokbel, K. The role of cyclooxygenase-2 in breast cancer: review. *Breast Cancer Res. Treat.* 2008, *109(2)*, 189-198.
- [6] Coussens, L. M.; Werb, Z. *Inflammation and cancer*. *Nature* 2002, *420(6917)*, 860-867.
- [7] Park, E. The etiology of rickets. *Physiol. Rev.* 1923 (Baltimore) *iii*, 106-163.
- [8] Deluca, H. F.; Cantorna, M. T. Vitamin D: its role and uses in immunology. *FASEB J.* 2001, *15*, 2579-2585.
- [9] Guyton, K. Z.; Kensler, T. W.; Posner, G. H. Vitamin D and vitamin D analogs as cancer chemopreventive agents. *Nutr. Rev.* 2003, *61*, 227-238.
- [10] Jones, G.; Strugnell, S. A.; Deluca, H. F. Current understanding of the molecular actions of vitamin D. *Physiol. Rev.* 1998, *78*, 1193-1231.
- [11] Adorini L. Immunomodulatory effects of vitamin D receptor ligands in autoimmune diseases. *Int. Immunopharmacol.* 2002, *2*, 1017-1028.
- [12] Colston, K. W.; Hansen, C. M. Mechanisms implicated in the growth regulatory effects of vitamin D in breast cancer. *Endocr. Relat. Cancer* 2002, *9(1)*, 45-59.
- [13] Johnson, C. S.; Hershberger, P. A.; Trump, D. L. Vitamin D-related therapies in prostate cancer. *Cancer Metastasis Rev.* 2002, *21*, 147-158.
- [14] Mathieu, C.; Adorini, L. The coming of age of 1,25-dihydroxyvitamin D₃ analogs as immunomodulatory agents. *Trends Mol. Med.* 2002, *8*, 174-179.
- [15] O'Kelly, J.; Koeffler, H. P. Vitamin D analogs and breast cancer. *Recent Results Cancer Res.* 2003, *164*, 333-348.
- [16] van den Bemd, G. J.; Chang, G. T. Vitamin D and vitamin D analogs in cancer treatment. *Curr. Drug Targets* 2002, *3*, 85-94.
- [17] Welsh, J.; Wietzke, J. A.; Zinser, G. M.; Byrne, B.; Smith, K.; Narvaez, C. J. Vitamin D₃ receptor as a target for breast cancer prevention. *J. Nutr.* 2003, *133*, 2425-2433S.
- [18] Garland, C. F.; Garland, F. C.; Gorham, E. D. Calcium and vitamin D. Their potential roles in colon and breast cancer prevention. *Ann. N. Y. Acad. Sci.* 1999, *889*, 107-119.
- [19] Janowsky, E. C.; Lester, G. E.; Weinberg, C. R.; Millikan, R. C.; Schildkraut, J. M.; Garrett, P. A.; Hulka, B. S. Association between low levels of 1,25-dihydroxyvitamin D and breast cancer risk. *Public Health Nutr.* 1999, *2*, 283-291.
- [20] Shin, M. H.; Holmes, M. D.; Hankinson, S. E.; Wu, K.; Colditz, G. A.; Willett, W. C. Intake of dairy products, calcium, and vitamin D and risk of breast cancer. *J. Natl. Cancer Inst.* 2002, *94*, 1301-1311.

- [21] Studzinski, G. P.; Moore, D. C. Sunlight – can it prevent as well as cause cancer? *Cancer Res.* 1995, 55, 4014–4022.
- [22] Gorham, E. D.; Garland, F. C.; Garland, C. F. Sunlight and breast cancer incidence in the USSR. *Int. J. Epidemiol.* 1990, 19, 820–824.
- [23] Garland, F. C.; Garland, C. F.; Gorham, E. D.; Young J. F. Geographic variation in breast cancer mortality in the United States: a hypothesis involving exposure to solar radiation. *Prev. Med.* 1990, 19, 614–622.
- [24] Blot, W. J.; Fraumeni J. F. Jr.; Stone B. J. Geographic patterns of breast cancer in the United States. *J. Natl. Cancer. Inst.* 1977, 59, 1407–1411.
- [25] Malloy, P. J.; Pike, J. W.; Feldman, D. The vitamin D receptor and the syndrome of hereditary 1,25-dihydroxyvitamin D-resistant rickets. *Endocr. Rev.* 1999, 20, 156–188.
- [26] Liu, M.; Lee, M. H.; Cohen, M.; Bommakanti, M.; Freedman, L. P. Transcriptional activation of the Cdk inhibitor p21 by vitamin D₃ leads to the induced differentiation of the myelomonocytic cell line U937. *Genes Dev.* 1996, 10, 142–153.
- [27] Polly, P.; Carlberg, C.; Eisman, J. A.; Morrison, N. A. Identification of a vitamin D₃ response element in the fibronectin gene that is bound by a vitamin D₃ receptor homodimer. *J. Cell. Biochem.* 1996, 60, 322–333.
- [28] Polly, P.; Carlberg, C.; Eisman, J. A.; Morrison, N. A. 1 α ,25-dihydroxyvitamin D₃ receptor as a mediator of transrepression of retinoid signaling. *J. Cell. Biochem.* 1997, 67, 287–296.
- [29] Berger, U.; Wilson, P.; McClelland, R. A.; Colston, K.; Haussler, M. R.; Pike, J. W.; Coombes, R. C. Immunocytochemical detection of 1,25-dihydroxyvitamin D receptors in normal human tissues. *J. Clin. Endocrinol. Metab.* 1988, 67, 607–613.
- [30] Stack, E.; DuBois, R. N. Regulation of cyclo-oxygenase-2. *Best Pract. Res. Clin. Gastroenterol.* 2001, 15 (5), 787–800.
- [31] Murakami, M.; Kudo, I. Recent advances in molecular biology and physiology of the prostaglandin E₂-biosynthetic pathway. *Prog. Lipid. Res.* 2004, 43(1), 3–35.
- [32] Anggard, E. The biological activities of three metabolites of prostaglandin E₁. *Acta Physiol. Scand.* 1966, 66, 509–510.
- [33] Tsuboi, K.; Sugimoto, Y.; Ichikawa, A. Prostanoid receptor subtypes. *Prostaglandins Other Lipid. Mediat.* 2002, 68–69, 535–556.
- [34] Williams, C. S.; DuBois, R. N. Prostaglandin endoperoxide synthase: why two isoforms? *Am. J. Physiol.* 1996, 270, 393–400.
- [35] Herschman, H. R. Prostaglandin synthase 2. *Biochim. Biophys. Acta.* 1996, 1299(1), 125–140.
- [36] Langenbach, R.; Morham, S. G.; Tiano, H. F.; Loftin, C. D.; Ghanayem, B. I.; Chulada, P. C.; Mahler, J. F.; Lee, C. A.; Goulding, E. H.; Kluckman, K. D.; Kim, H. S.; Smithies, O. Prostaglandin synthase 1 gene disruption in mice reduces arachidonic acid-induced inflammation and indomethacin-induced gastric ulceration. *Cell* 1995, 83(3), 483–492.
- [37] Lim, H.; Paria, B. C.; Das, S. K.; Dinchuk, J. E.; Langenbach, R.; Trzaskos, J. M.; Dey, S. K. Multiple female reproductive failures in cyclooxygenase 2-deficient mice. *Cell* 1997, 91(2), 197–208.
- [38] Davis, B.J.; Lennard, D. E.; Lee, C. A.; Tiano, H. F.; Morham, S. G.; Wetsel, W. C.; Langenbach, R. Anovulation in cyclooxygenase-2 deficient mice is restored by prostaglandin E₂ and interleukin-1 beta. *Endocrinology* 1999, 140(6), 2685–2695.

- [39] Loftin, C. D.; Trivedi, D. R.; Tiano, H. F.; Clark, J.A.; Lee, C. A.; Epstein, J. A.; Morham, S. G.; Breyer, M. D.; Nguyen, M.; Hawkins, B.M. Failure of ductus arteriosus closure and remodelling in neonatal mice deficient in cyclooxygenase-1 and -2. *Proc. Natl. Acad. Sci. U.S.A.* 2001, 98(3), 1059-1064
- [40] Rozic, J. G.; Chakraborty, C.; Lala, P. K. Cyclooxygenase inhibitors retard murine mammary tumor progression by reducing tumor cell migration, invasiveness and angiogenesis. *Int. J. Cancer.* 2001, 93, 497-506.
- [41] Eberhart, C. E.; Coffey, R. J.; Radhika, A.; Giardiello, F. M.; Ferreback, S.; Dubois, R. N. Up-regulation of cyclooxygenase gene expression in human colorectal adenomas and adenocarcinomas. *Gastroenterology* 1994, 107, 1183-1188.
- [42] Sinicrope, F. A.; Lemoine, M.; Xi, L.; Lynch, P. M.; Cleary, K. R.; Shen, Y.; Frazier, M. L. Reduced expression of cyclooxygenase 2 proteins in hereditary nonpolyposis colorectal cancers relative to sporadic cancers. *Gastroenterology* 1999, 117, 350-358.
- [43] Ristimaki, A.; Honkanen, N.; Jankala, H.; Sipponen, P.; Harkonen, M. Expression of cyclooxygenase-2 in human gastric carcinoma. *Cancer Res.* 1997, 57, 1276-1280.
- [44] Zimmermann, K. C.; Sarbia, M.; Weber, A.; Bochar, F.; Gabbert, H. E.; Schror, K. Cyclooxygenase-2 expression in human esophageal carcinoma. *Cancer Res.* 1999, 5, 198-204.
- [45] Swami, S.; Krishnan, A. V.; Moreno, J.; Bhattacharyya, R. B.; Peehl, D.; Feldman, D. Calcitriol and enistein actions to inhibit the prostaglandin pathway: potential combination therapy to treat prostate cancer. *J. Nutr.* 2007, 137, 205S-210S.
- [46] Subbaramaiah, K.; Dannenberg, A. J. Cyclooxygenase 2: a molecular target for cancer prevention and treatment. *Trends. Pharmacol. Sci.* 2003, 24, 96-102.
- [47] Heinonen, P. K.; Metsa-Ketela, T. Prostaglandin and thromboxane production in ovarian cancer tissue. *Gynecol. Obstet. Invest.* 1984, 18(5), 225-229.
- [48] Ylikorkala, O.; Kauppila, A.; Viinikka, L. Prostacyclin and thromboxane in ovarian cancer: effect of cytostatics and prostaglandin synthesis inhibitors. *Gynecol. Oncol.* 1983, 16(3), 340-345.
- [49] Munkarah, A. R.; Morris, R.; Baumann, P.; Deppe, G.; Malone, J.; Diamond, M. P.; Saed, G. M. Effects of prostaglandin E(2) on proliferation and apoptosis of epithelial ovarian cancer cells. *J. Soc. Gynecol. Investig.* 2002, 9(3), 168-173.
- [50] Gupta, R. A.; Tejada, L. V.; Tong, B. J.; Das, S. K.; Morrow, J. D.; Dey, S. K.; DuBois, R. N. Cyclooxygenase-1 is overexpressed and promotes angiogenic growth factor production in ovarian cancer. *Cancer Res.* 2003, 63(5), 906-911.
- [51] Ali-Fehmi, R.; Morris, R. T.; Bandyopadhyay, S.; Che, M.; Schimp, V.; Malone, J.M. Jr.; Munkarah, A. R. Expression of cyclooxygenase-2 in advanced stage ovarian serous carcinoma: correlation with tumor cell proliferation, apoptosis, angiogenesis, and survival. *Am. J. Obstet. Gynecol.* 2005, 192(3), 819-825.
- [52] Daikoku, T.; Wang, D.; Tranguch, S.; Morrow, J. D.; Orsulic, S.; DuBois, R. N.; Dey, S. K. Cyclooxygenase-1 is a potential target for prevention and treatment of ovarian epithelial cancer. *Cancer Res.* 2005, 65(9), 3735-3744.
- [53] Dore, M.; Cote, L. C.; Mitchell, A.; Sirois, J. Expression of prostaglandin G/H synthase type 1, but not type 2, in human ovarian adenocarcinomas. *J. Histochem. Cytochem.* 1998, 46(1), 77-84.
- [54] Daikoku, T.; Tranguch, S.; Trofimova, I. N.; Dinulescu, D. M.; Jacks, T.; Nikitin, A. Y.; Connolly, D. C.; Dey, S.K. Cyclooxygenase-1 is overexpressed in multiple

- genetically engineered mouse models of epithelial ovarian cancer. *Cancer Res.* 2006, 66(5), 2527-2531.
- [55] Rodriguez-Burford, C.; Barnes, M.; N.; Oelschlager, D. K.; Myers, R. B.; Talley, L. I.; Partridge, E. E.; Grizzle, W. E. Effects of nonsteroidal anti-inflammatory agents (NSAIDs) on ovarian carcinoma cell lines: preclinical evaluation of NSAIDs as chemopreventive agents. *Clin. Cancer Res.* 2002, 8(1), 202-209.
- [56] Schreinemachers, D. M., Everson, R. B. Aspirin use and lung, colon, and breast cancer. Incidence in a prospective study. *Epidemiology* 1994, 5, 138-146.
- [57] Harris, R. E., Namboodiri, K. K., Farrar, W. B. Non-steroidal anti-inflammatory drugs and breast cancer. *Epidemiology* 1996, 7, 203-205.
- [58] Sharp, C. R., Collet, J. P., McNutt, M., Belzile, E., Boivin, J. F., Hanley, J. A. Nested case control study of the effect of non-steroidal anti-inflammatory drugs on breast cancer risk and stage. *Br. J. Cancer* 2000, 83, 112-120.
- [59] Liu, X.; Rose, D. Differential expression and regulation of cyclooxygenase-1 and -2 in two human breast cancer cell lines. *Cancer Res.* 1996, 56, 5125-5127.
- [60] Schrey, M. P.; Patel, K. V. Prostaglandin E₂ production and metabolism in human breast cancer cells and breast fibroblasts. Regulation by inflammatory mediators. *Br. J. Cancer* 1995, 72, 1412-1419.
- [61] Rolland, P. H.; Martin, P. M.; Jacquemier, J.; Rolland, A.; Toga, M. Prostaglandins in human breast cancer: evidence suggesting that elevated prostaglandin production is a marker of high metastatic potential for neoplastic cells. *J. Natl. Cancer Inst. (Bethesda)* 1980, 64, 1061-1070.
- [62] Karmali, R. A.; Welts, S.; Thaler, H. T.; Lefevre, F. Prostaglandins in breast cancer. Relationship to disease stage and hormone status. *Br. J. Cancer* 1983, 48, 689-696.
- [63] Kawamori, T.; Wakabayashi, K. COX-2 and prostanoid receptors: good targets for chemoprevention. *J. Environ. Pathol. Toxicol. Oncol.* 2002, 21(2), 149-153.
- [64] Sonoshita, M.; Takaku, K.; Sasaki, N.; Sugimoto, Y.; Ushikubi, F.; Narumiya, S.; Oshima, M.; Taketo, M. M. Acceleration of intestinal polyposis through prostaglandin receptor EP2 in Apc(Delta 716) knockout mice. *Nat. Med.* 2001, 7, 1048-1051.
- [65] Zha, S.; Yegnasubramanian, V.; Nelson, W. G.; Isaacs, W. B.; De Marzo, A. M. Cyclooxygenases in cancer: progress and perspective. *Cancer Lett.* 2004, 215, 1-20.
- [66] Chang, S. H.; Liu, C. H.; Conway, R.; Han, D. K.; Nithipatikom, K.; Trifan, O. C.; Lane, T. F.; Hla, T. Role of prostaglandin E₂-dependent angiogenic switch in cyclooxygenase 2-induced breast cancer progression. *Proc. Natl. Acad. Sci. U S A* 2004, 101, 591-596.
- [67] Yan, M.; Rerko, R. M.; Platzer, P.; Dawson, D.; Willis, J.; Tong, M.; Lawrence, E.; Lutterbaugh, A.; Lu, S.; Lu, S.; Willson, J. K.; Luo, G.; Hensold, J.; Tai, H. H.; Wilson, K.; Markowitz, S. D. 15-Hydroxyprostaglandin dehydrogenase, a COX-2 oncogene antagonist, is a TGF-beta-induced suppressor of human gastrointestinal cancers. *Proc. Natl. Acad. Sci. USA* 2004, 101, 17468-17473.
- [68] Ding, Y.; Tong, M.; Liu, S.; Moscow, J. A.; Tai, H. H. NAD⁺-linked 15-hydroxyprostaglandin dehydrogenase (15-PGDH) behaves as a tumor suppressor in lung cancer. *Carcinogenesis* 2005, 26, 65-72.
- [69] Gee, J. R.; Montoya, R. G.; Khaled, H. M.; Sabichi, A. L.; Grossman, H. B. Cytokeratin 20, AN43, PGDH, and COX-2 expression in transitional and squamous cell carcinoma of the bladder. *Urol. Oncol.* 2003, 21, 266-270.

- [70] Wolf, I.; O'Kelly, J.; Rubinek, T.; Tong, M.; Nguyen, A.; Lin, B. T.; Tai, H. H.; Karlan, B. Y.; Koeffler, H. P. 15-hydroxyprostaglandin dehydrogenase is a tumor suppressor of human breast cancer. *Cancer Res.* 2006, 66(15), 7818-7823.
- [71] van 't Veer, L. J.; Dai, H.; van de Vijver, M. J.; He, Y. D.; Hart, A. A.; Mao, M.; Peterse, H. L.; van der Kooy, K.; Marton, M. J.; Witteveen, A. T.; Schreiber, G. J.; Kerkhoven, R. M.; Roberts, C.; Linsley, P. S.; Bernards, R.; Friend, S. H. Gene expression profiling predicts clinical outcome of breast cancer. *Nature* 2002, 415, 530-536.
- [72] Half, E.; Tang, X. M.; Gwyn, K.; Sahin, A.; Wathen, K.; Sinicrope, F. A. Cyclooxygenase-2 expression in human breast cancers and adjacent ductal carcinoma in situ. *Cancer Res.* 2002, 62, 1676-1681.
- [73] Denkert, C.; Winzer, K. J.; Müller, B. M.; Weichert, W.; Pest, S.; Köbel, M.; Kristiansen, G.; Reles, A.; Siegert, A.; Guski, H.; Hauptmann, S. Elevated expression of cyclooxygenase-2 is a negative prognostic factor for disease free survival and overall survival in patients with breast carcinoma. *Cancer* 2003, 97(12), 2978-2987.
- [74] Ristimäki, A.; Sivula, A.; Lundin, J.; Lundin, M.; Salminen, T.; Haglund, C.; Joensuu, H.; Isola, J. Prognostic significance of elevated cyclooxygenase-2 expression in breast cancer. *Cancer Res.* 2002, 62, 632-635.
- [75] Singh Ranger, G.; Thomas, V.; Jewell, A.; Mokbel, K. Elevated cyclooxygenase-2 expression correlates with distant metastases in breast cancer. *Anticancer Res.* 2004, 24(4), 2349-2351.
- [76] Nassar, A.; Radhakishnan, A.; Cabero, I. A.; Cotsonis, G.; Cohen, C. COX-2 expression in invasive breast cancer: Correlation with prognostic parameters and outcome. *Appl. Immunohistochem. Mol. Morphol.* 2007, 15(3), 255-259.
- [77] Hwang, D., Scollard, D., Byrne, J., Levine, E. Expression of cyclooxygenase-1 and cyclooxygenase-2 in human breast cancer. *J. Natl. Cancer Inst.* 1998, 90, 455-460.
- [78] Soslow, R., Dannenberg, A., Rush, D., Woerner, B. M., Khan, K. N., Masferrer, J., Koki, A. COX-2 is expressed in human pulmonary, colonic, and mammary tumors. *Cancer* 2000, 2637-2645.
- [79] Costa, C.; Soares, R.; Reis-Filho, J. S.; Leitão, D.; Amendoeira, I., Schmitt, F. C. Cyclooxygenase 2 expression is associated with angiogenesis and lymph node metastasis in human breast cancer. *J. Clin. Pathol.* 2002, 55, 429-434.
- [80] Kelly, L. M.; Hill, A. D. K.; Kennedy, S.; Connolly, E. M.; Ramanath, R.; The, S.; Dijkstra, B.; Purcell, R.; McDermott, E. W.; O'Higgins, N. Lack of prognostic effect of COX-2 expression in primary breast cancer on short term follow-up. *Eur. J. Surg. Oncol.* 2002, 29, 707-710.
- [81] Lim, S. C. Role of COX-2, VEGF and cyclin D1 in mammary infiltrating duct carcinoma. *Oncol. Rep.* 2003, 10 (5), 1241-1249.
- [82] Wülfing, P.; Diallo, R.; Müller, C.; Wülfing, C.; Poremba, C.; Heinecke, A.; Rody, A.; Greb, R. R.; Böcker, W.; Kiesel, L. Analysis of cyclooxygenase-2 expression in human breast cancer: high throughput tissue microarray analysis. *J. Cancer Res. Clin. Oncol.* 2003, 129, 375-382.
- [83] Boland, G. P.; Butt, I. P.; Prasad, R.; Knox, W. F.; Bundred, N. J. COX-2 expression is associated with an aggressive phenotype in ductal carcinoma in situ. *Br. J. Cancer* 2004, 90, 423-429.
- [84] Zhao, X. Q.; Pang, D.; Xue, Y. Expression of the cyclooxygenase-2 gene in human breast carcinoma. *Zhonghua Wai Ke Za Zhi* 2003, 41(6), 427-429.

- [85] Singh Ranger, G.; Kirkpatrick, K. L.; Clark, G. M.; Mokbel, K. Cyclooxygenase-2 (COX-2) mRNA expression correlates with progesterone receptor positivity in human breast cancer. *Curr. Med. Res. Opin.* 2003, 19(2), 131-134.
- [86] Kirkpatrick, K.; Ogunkolade, W.; Bustin, S.; Jenkins, P.; Ghilchik, M.; Mokbel, K. The mRNA expression of cyclooxygenase-2 and vascular endothelial growth factor in human breast cancer. *Breast Cancer Res. Treat.* 2001, 69(3), 373.
- [87] Watanabe, O.; Shimizu, T.; Imamura, H.; Kinoshita, J.; Utada, Y.; Okabe, T.; Kimura, K.; Hirano, A.; Yoshimatsu, K.; Aiba, M.; Ogawa, K. Expression of cyclooxygenase-2 in malignant and benign breast tumours. *Anticancer Res.* 2003, 23(4), 3215-3221.
- [88] Yoshimura, N.; Sano, H.; Okamoto, M.; Akioka, K.; Ushogome, H.; Kadotani, Y.; Yoshimura, R.; Nobori, S.; Higuchi, A.; Ohmori, Y.; Nakamura, K. Expression of cyclooxygenase-1 and -2 in human breast cancer. *Surg. Today* 2003, 33, 805-811.
- [89] Ristimäki, A.; Garfinkel, S.; Weesendorf, J.; Maciag, T.; Hla, T. Induction of cyclooxygenase-2 by interleukin-1 alpha. Evidence for post-transcriptional regulation. *J. Biol. Chem.* 1994, 269(16), 11769-11775.
- [90] Evett, G. E.; Xie, W.; Chipman, J. G.; Robertson, D. L.; Simmons, D. L. Prostaglandin GH Synthase isoenzyme 2 expression in fibroblasts: regulation by dexamethasone, mitogens and oncogenes. *Arch. Biochem. Biophys.* 2003, 306, 169-177.
- [91] Sorlie, T.; Perou, C. M.; Tibshirani, R. Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *PNAS USA* 2001, 98, 10869-10874.
- [92] Salhab, M.; Singh-Ranger, G.; Mokbel, R.; Jouhra, F.; Jiang, W.G.; Mokbel, K. Cyclooxygenase-2 mRNA expression correlates with aromatase expression in human breast cancer. *J. Surg. Oncol.* 2007, 96(5), 424-428.
- [93] Diaz-Cruz, E. S.; Shapiro, C. L.; Brueggemeier, R. W. Cyclooxygenase inhibitors suppress aromatase expression and activity in breast cancer cells. *J. Clin. Endocrinol. Metab.* 2005, 90, 2563-2570.
- [94] Richards, J. A.; Petrel, T. A.; Brueggemeier, R. W. Signaling pathways regulating aromatase and cyclooxygenases in normal and malignant breast cells. *J. Steroid. Biochem. Mol. Biol.* 2002, 80, 203-212.
- [95] Brueggemeier, R. W.; Richards, J. A.; Petrel, T. A. Aromatase and cyclooxygenases: enzymes in breast cancer. *J. Steroid. Biochem. Mol. Biol.* 2003, 86, 501-507.
- [96] Vienonen, A.; Syvala, H.; Miettinen, S.; Tuohimaa, P.; Ylikomi, T. Expression of progesterone receptor isoforms A and B is differentially regulated by estrogen in different breast cancer cell lines. *J. Steroid. Biochem. Mol. Biol.* 2002, 80, 307-313.
- [97] Liu, C. H., Chang, S., Narko, K., Trifan, O. C., Wu, M., Smith, E., Haudenschild, C., Lane, T. F., Hla, T. Overexpression of cyclooxygenase-2 is sufficient to induce tumorigenesis in transgenic mice. *J. Biol. Chem.* 2001, 276, 18563-18569.
- [98] Tsujii, M.; DuBois, R. N. Alterations in cellular adhesion and apoptosis in epithelial cells overexpressing prostaglandin endoperoxide synthase 2. *Cell* 1995, 93, 705-716.
- [99] Oshima, M., Dinchuk, J. E., Kargman, S. L., Oshima, H., Hancock, B., Kwong, E., Trzaskos, J. M., Evans, J. F., Taketo, M. M. Suppression of intestinal polyposis in Apc716 knockout mice by inhibition of cyclooxygenase-2 (COX-2). *Cell* 1996, 80, 803-809.
- [100] Mestre, J. R.; Subbaramaiah, K.; Sacks, P. G.; Schantz, S. P.; Tanabe, T.; Inoue, H.; Dannenberg, A. J. Retinoids suppress epidermal growth factor-induced

- transcription of cyclooxygenase-2 in human oral squamous carcinoma cells. *Cancer Res.* 1991, 57(14), 2890-2895.
- [101] Davies, G.; Salter, J.; Hills, M.; Martin, L.-A.; Sacks, N.; Dowsett, M. Correlation between cyclooxygenase-2 expression and angiogenesis in human breast cancer. *Clin. Cancer Res.* 2003, 9, 2651-2656.
- [102] Lim, S. C.; Park, S. Y.; Do, N. Y. Correlation of cyclooxygenase-2 pathway and VEGF expression in head and neck squamous cell carcinoma. *Oncol. Rep.* 2003, 10 (5), 1073-1079.
- [103] Chu, J.; Lloyd, F. L.; Trifan, O. C.; Knapp, B.; Rizzo, M. T. Potential involvement of the cyclooxygenase-2 pathway in the regulation of tumor-associated angiogenesis and growth in pancreatic cancer. *Mol. Cancer Ther.* 2003, (1), 1-7.
- [104] Tsujii, M.; Kawano, S.; Tsujii, S.; Sawaoka, H.; Hori, M.; DuBois, N. Cyclooxygenase regulates angiogenesis induced by colon cancer cells. *Cell* 1998, 93, 705-716.
- [105] Gately, S. The contributions of cyclooxygenase-2 to tumour angiogenesis. *Cancer Metastasis Rev.* 2001, 19(1-2), 19-27.
- [106] Daniel, T. O.; Liu, H.; Morrow, J. D.; Crews, B. C.; Marnett, L.J. Thromboxane A2 is a mediator of cyclooxygenase-2-dependent endothelial migration and angiogenesis. *Cancer Res.* 1999, 59, 4574-4577.
- [107] Pezzella, F.; Pastorino, U.; Tagliabue, E.; Andreola, S.; Sozzi, G.; Gasparini, G.; Menard, S.; Gatter, K. C.; Harris, A. L.; Fox, S.; Buyse, M.; Pilotti, S.; Pierotti, M.; Rilke, F. Non-small-cell lung carcinoma tumor growth without morphological evidence of neo-angiogenesis. *Am. J. Pathol.* 1997, 151, 1417-1423.
- [108] Maniotis, A. J.; Folberg, R.; Hess, A.; Seftor, E. A.; Gardner, L. M.; Pe'er, J.; Trent, J. M.; Meltzer, P. S.; Hendrix, M. J. Vascular channel formation by human melanoma cells in vivo and in vitro: vasculogenic mimicry. *Am. J. Pathol.* 1999, 155, 739-752.
- [109] Shirakawa, K.; Shibuya, M.; Heike, Y.; Takashima, S.; Watanabe, I.; Konishi, F.; Kasumi, F.; Goldman, C. K.; Thomas, K. A.; Bett, A.; Terada, M.; Wakasugi, H. Tumor-infiltrating endothelial cells and endothelial precursor cells in inflammatory breast cancer. *Int. J. Cancer* 2002, 99, 344-351.
- [110] Hendrix, M. J.; Seftor, E. A.; Hess, A. R.; Seftor, R. E. Vasculogenic mimicry and tumour-cell plasticity: lessons from melanoma. *Nat. Rev. Cancer* 2003, 3, 411-421.
- [111] Shirakawa, K.; Kobayashi, H.; Heike, Y.; Kawamoto, S.; Brechbiel, M. W.; Kasumi, F.; Iwanaga, T.; Konishi, F.; Terada, M.; Wakasugi, H. Hemodynamics in vasculogenic mimicry and angiogenesis of inflammatory breast cancer xenograft. *Cancer Res.* 2002, 62, 560-566.
- [112] Basu, G. D.; Liang, W. S.; Stephan, D. A.; Wegener, L. T.; Conley, C. R.; Pockaj, B. A.; Mukerjee, P. A novel role for cyclooxygenase-2 in regulating vascular channel formation by human breast cancer cells. *Breast Cancer Res.* 2006, 8(6), R69.
- [113] Khuder, S.A.; Mutgi, A. B. Breast cancer and NSAID use: a metaanalysis. *Br. J. Cancer.* 2001, 84, 1188-1192.
- [114] Egan, K., Stampfer, M., Giovannucci, E., Rosner, B., Colditz, G. Prospective study of regular aspirin use and the risk of breast cancer. *J. Natl. Cancer Inst.* 1996, (Bethesda), 88, 988-993.
- [115] Rosenberg, L.; Palmer, J. R.; Zauber, A. G.; Warshaver, M. E.; Stolley, P.D.; Shapiro, S. A hypothesis: nonsteroidal anti-inflammatory drugs reduce the incidence of large-bowel cancer. *J. Natl. Cancer Inst.* 1991, 83, 355-358.

- [116] Thun, M. J.; Namboodiri, M. M.; Heath, C. W. Jr. Aspirin use and reduced risk of fatal colon cancer. *N. Engl. J. Med.* 1991, 325, 1593-1596.
- [117] Alshafie, G. A., Abou-Issa, H., Seibert, K., Harris, R. Chemotherapeutic evaluation of celecoxib, a cyclooxygenase-2 inhibitor, in rats mammary tumor model. *Oncol. Rep.* 2000, 7, 1377-1381.
- [118] Harris, R. E.; Alshafie, G. A.; Abou-Issa, H.; Seibert, K. Chemoprevention of breast cancer in rats by celecoxib, a cyclooxygenase 2 inhibitor. *Cancer Res.* 2000, 60, 2101-2103.
- [119] Barnes, N. L.; Warnberg, F.; Farnie, G.; White, D.; Jiang, W.; Anderson, E.; Bundred, N. J. Cyclooxygenase-2 inhibition: effects on tumour growth, cell cycling and lymphangiogenesis in a xenograft model of breast cancer. *Br. J. Cancer* 2007, 96(4), 575-582.
- [120] Yoshinaka, R.; Shibata, M. A.; Morimoto, J.; Tanigawa, N.; Otsuki, Y. COX-2 inhibitor celecoxib suppresses tumor growth and lung metastasis of a murine mammary cancer. *Anticancer Res.* 2006, 26(6B), 4245-4254.
- [121] Fabi, A.; Metro, G.; Papaldo, P.; Mottolese, M.; Melucci, E.; Carlini, P.; Sperduti, I.; Russillo, M.; Gelibter, A.; Ferretti, G.; Tomao, S.; Milella, M.; Cognetti, F. Impact of celecoxib on capecitabine tolerability and activity in pretreated metastatic breast cancer: results of a phase II study with biomarker evaluation. *Cancer Chemother. Pharmacol.* 2008, 62(4), 717-725.
- [122] Canney, P. A.; Machin, M. A.; Curto, J. A feasibility study of the efficacy and tolerability of the combination of Exemestane with the COX-2 inhibitor Celecoxib in post-menopausal patients with advanced breast cancer. *Eur. J. Cancer* 2006, 42(16), 2751-2756.
- [123] Dirix, L. Y.; Ignacio, J.; Nag, S.; Bapsy, P.; Gomez, H.; Raghunadharao, D.; Paridaens, R.; Jones, S.; Falcon, S.; Carpentieri, M.; Abbattista, A.; Lobelle, J. P. Treatment of advanced hormone-sensitive breast cancer in postmenopausal women with exemestane alone or in combination with celecoxib. *J. Clin. Oncol.* 2008, 10, 26(8), 1253-1259.
- [124] Beer, T. M.; Ryan, C. W.; Venner, P. M.; Petrylak, D. P.; Chatta, G. S.; Ruether, J. D.; Chi, K. N.; Young, J.; Henner, W.D.; ASCENT(AIPC Study of Calcitriol ENhancing Taxotere) Investigators. Intermittent chemotherapy in patients with metastatic androgen-independent prostate cancer: results from ASCENT, a double-blinded, randomized comparison of high-dose calcitriol plus docetaxel with placebo plus docetaxel. *Cancer* 2008, 112(2), 326-330.
- [125] Dragovich, T.; Burris, H. 3rd, Loehrer, P.; Von Hoff, D. D.; Chow, S.; Stratton, S.; Green, S.; Obregon, Y.; Alvarez, I.; Gordon, M. Gemcitabine plus celecoxib in patients with advanced or metastatic pancreatic adenocarcinoma: results of a phase II trial. *Am. J. Clin. Oncol.* 2008, 31(2), 157-162.
- [126] Agarwala, A.; Fisher, W.; Bruetman, D.; McClean, J.; Taber, D.; Titzer, M.; Juliar, B.; Yu, M.; Breen, T.; Einhorn, L. H.; Hanna, N. Gefitinib plus celecoxib in chemotherapy-naïve patients with stage IIIB/IV non-small cell lung cancer: a phase II study from the Hoosier Oncology Group. *J. Thorac. Oncol.* 2008, 3(4), 374-379.
- [127] Xiao, H.; Zhang, Q.; Lin, Y.; Reddy, B. S.; Yang, C. S.; Combination of atorvastatin and celecoxib synergistically induces cell cycle arrest and apoptosis in colon cancer cells. *Int. J. Cancer* 2008, 122(9), 2115-2124.

- [128] Giovanucci, E. The epidemiology of vitamin D and cancer incidence and mortality: a review (United States). *Cancer causes control* 2005, 16, 83-95.
- [129] Holick, M. F. Sunlight and vitamin D for bone health and prevention of autoimmune diseases, cancers, and cardiovascular disease. *Am. J. Clin. Nutr.* 2004, 80, 1678-1688S.
- [130] Trang, H. M.; Cole, D. E.; Rubin, L. A.; Pierratos, A.; Siu, S.; Vieth, R. Evidence that vitamin D₃ increases serum 25-hydroxyvitamin D more efficiently than does vitamin D₂. *Am. J. Clin. Nutr.* 1998, 68, 854-858.
- [131] Hollis, B. W.; Wagner, C. L. Normal serum vitamin D levels. *N. Engl. J. Med.* 2005, 352(5), 515-516.
- [132] Reichrath, J. Vitamin D and the skin: an ancient friend, revisited. *Experimental Dermatology* 2007, 16(7), 618-625
- [133] Holick, M. F. The cutaneous photosynthesis of previtamin D₃: a unique photoendocrine system. *J. Invest. Dermatol.* 1981, 77, 51-58.
- [134] Feskanich, D.; Ma, J.; Fuchs, C. S.; Kirkner, G. J.; Hankinson, S. E.; Hollis, B. W.; Giovannucci, E. L. Plasma vitamin D metabolites and risk of colorectal cancer in women. *Cancer Epidemiol. Biomarkers Prev.* 2004, 13(9), 1502-1508.
- [135] Ohyama, Y., Yamasaki, T. Eight cytochrome P450s catalyze vitamin D metabolism. *Front. Biosci.* 2004, 9, 3007-3018.
- [136] Hewison, M.; Zehnder, D.; Chakraverty, R.; Adams, J. S. Vitamin D and barrier function: a novel role for extra-renal 1 α -hydroxylase. *Mol. Cell Endocrinol.* 2004, 215, 31-38.
- [137] Bell, N. H. Renal and nonrenal 25-hydroxyvitamin D-1 α -hydroxylases and their clinical significance. *J. Bone Miner. Res.* 1998, 13, 350-353.
- [138] Omdahl, J. L.; Morris, H. A.; May, B. K. Hydroxylase enzymes of the vitamin D pathway: expression, function, and regulation. *Annu. Rev. Nutr.* 2002, 22, 139-166.
- [139] Dusso, A. S.; Brown, A. J.; Slatopolsky, E. Vitamin D. *Am. J. Physiol. Renal Physiol.* 2005, 289(1), F8-28.
- [140] Turunen, M. M.; Dunlop, T. W.; Carlberg, C.; Väisänen, S. Selective use of multiple vitamin D response elements underlies the 1 α ,25-dihydroxyvitamin D₃-mediated negative regulation of the human CYP27B1 gene. *Nucleic Acids Res.* 2007, 35(8), 2734-2747.
- [141] Zitterman, A. Vitamin D in preventive medicine: are we ignoring the evidence? *Br. J. Nutr.* 2003, 89, 552-572.
- [142] Hewison, M.; Zehnder, D.; Bland, R.; Stewart, P. M. 1 α -Hydroxylase and the action of vitamin D. *J. Mol. Endocrinol.* 2000, 25, 141-148.
- [143] Zehnder, D.; Bland, R.; Chana, R. S.; Wheeler, D. C.; Howie, A. J.; Williams, M. C.; Stewart, P. M.; Hewison, M. Synthesis of 1,25-dihydroxyvitamin D(3) by human endothelial cells is regulated by inflammatory cytokines: a novel autocrine determinant of vascular cell adhesion. *J. Am. Soc. Nephrol.* 2002, 13(3), 621-629.
- [144] Stoffels, K.; Overbergh, L.; Giuliatti, A.; Verlinden, L.; Bouillon, R.; Mathieu, C. Immune regulation of 25-hydroxyvitamin-D₃-1 α -hydroxylase in human monocytes. *J. Bone Miner. Res.* 2006, 21(1), 37-47.
- [145] Overbergh, L.; Stoffels, K.; Waer, M.; Verstuyf, A.; Bouillon, R.; Mathieu, C. Immune regulation of 25-hydroxyvitamin D-1 α -hydroxylase in human monocytic THP1

- cells: mechanisms of interferon-gamma-mediated induction. *J. Clin. Endocrinol. Metab.* 2006, 91(9), 3566-3574.
- [146] Stoffels, K.; Overbergh, L.; Bouillon, R.; Mathieu, C. Immune regulation of 1 α -hydroxylase in murine peritoneal macrophages: unravelling the IFN γ pathway. *J. Steroid. Biochem. Mol. Biol.* 2007, 103(3-5), 567-571.
- [147] Lopez, E. R.; Zwermann, O.; Segni, M.; Meyer, G.; Reincke, M.; Seissler, J.; Herwig, J.; Usadel, K. H.; Badenhop, K. A promoter polymorphism of the CYP27B1 gene is associated with Addison's disease, Hashimoto's thyroiditis, Graves' disease and type 1 diabetes mellitus in Germans. *Eur. J. Endocrinol.* 2004, 151, 193-197.
- [148] Fischer, D.; Seifert, M.; Becker, S.; Ludders, D.; Cordes, T.; Reichrath, J.; Friedrich, M. 25-Hydroxyvitamin D₃ 1 α -hydroxylase splice variants in breast cell lines MCF-7 and MCF-10. *Cancer Genomics Proteomics* 2007, 4(4), 295-300.
- [149] Cordes, T.; Diesing, D.; Becker, S.; Fischer, D.; Diedrich, K.; Friedrich, M. Expression of splice variants of 1 α -hydroxylase in mcf-7 breast cancer cells. *J. Steroid. Biochem. Mol. Biol.* 2007, 103(3-5), 326-329.
- [150] Barreto, A. M.; Schwartz, G. G.; Woodruff, R.; Cramer, S. D. 25-Hydroxyvitamin D₃, the prohormone of 1,25-dihydroxyvitamin D₃, inhibits the proliferation of primary prostatic epithelial cells. *Cancer Epidemiol. Biomarkers Prev.* 2000, 9, 265-270.
- [151] Schwartz, G. G.; Whitlatch, L. W.; Chen, T. C.; Lokeshwar, B. L.; Holick, M. F. Human prostate cells synthesize 1,25-dihydroxyvitamin D₃ from 25-hydroxyvitamin D₃. *Cancer Epidemiol. Biomarkers Prev.* 1998, 7, 391-395.
- [152] Tangpricha, V.; Flanagan, J. N.; Whitlatch, L. W.; Tseng, C. C.; Chen, T. C.; Holt, P. R.; Lipkin, M. S.; Holick, M. F. 25-Hydroxyvitamin D-1 α -hydroxylase in normal and malignant colon tissue. *Lancet* 2001, 357, 1673-1674.
- [153] Bareis, P.; Bises, G.; Bischof, M. G.; Cross, H. S.; Peterlik, M. 25-hydroxy-vitamin D metabolism in human colon cancer cells during tumor progression. *Biochem. Biophys. Res. Commun.* 2001, 285, 1012-1017.
- [154] Ogunkolade, B. W.; Boucher, B. J.; Fairclough, P. D.; Hitman, G. A.; Dorudi, S.; Jenkins, P. J.; Bustin, S. A. Expression of 25-hydroxyvitamin D-1 α -hydroxylase mRNA in individuals with colorectal cancer. *Lancet* 2002, 359, 1831-1832.
- [155] Friedrich, M.; Diesing, D.; Cordes, T.; Fischer, D.; Becker, S.; Chen, T. C.; Flanagan, J. N.; Tangpricha, V.; Gherson, I.; Holick, M. F.; Reichrath, J. Analysis of 25-hydroxyvitamin D₃-1 α -hydroxylase in normal and malignant breast tissue. *Anticancer Res.* 2006, 26(4A), 2615-2620.
- [156] Garland, C. F.; Comstock, G. W.; Garland, F. C.; Helsing, K. J.; Shaw, E. K.; Gorham, E. D. Serum 25-hydroxyvitamin D and colon cancer: eight-year prospective study. *Lancet* 1989, 2, 1176-1178.
- [157] Holt, P. R.; Arber, N.; Halmos, B.; Forde, K.; Kissileff, H.; McGlynn, K. A.; Moss, S. F.; Kurihara, N.; Fan, K. Colonic epithelial cell proliferation decreases with increasing levels of serum 25-hydroxy vitamin D. *Cancer Epidemiol. Biomarkers Prev.* 2002, 11, 113-119.
- [158] Lowe, L. C.; Guy, M.; Mansi, J. L.; Peckitt, C.; Bliss, J.; Wilson, R. G.; Colston, K. W. Plasma 25-hydroxy vitamin D concentrations, vitamin D receptor genotype and breast cancer risk in a UK Caucasian population. *Eur. J. Cancer* 2005, 41, 1164-1169.

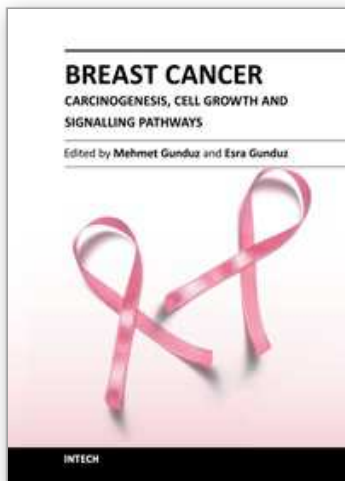
- [159] Berube, S.; Diorio, C.; Verhoek-Oftedahl, W.; Brisson, J. Vitamin D, calcium and mammographic breast densities. *Cancer Epidemiol. Biomarkers Prev.* 2004, 13, 1466-1472.
- [160] Huang, D. C.; Papavasiliou, V.; Rhim, J. S.; Horst, R. L.; Kremer, R. Targeted disruption of the 25-hydroxyvitamin D3 1 α -hydroxylase gene in ras-transformed keratinocytes demonstrates that locally produced 1 α ,25-dihydroxyvitamin D3 suppresses growth and induces differentiation in an autocrine fashion. *Mol. Cancer Res.* 2002, 1, 56-67.
- [161] Segersten, U.; Holm, P. K.; Björklund, P.; Hessman, O.; Nordgren, H.; Binderup, L.; Akerström, G.; Hellman, P.; Westin, G. 25-Hydroxyvitamin D3 1 α -hydroxylase expression in breast cancer and use of non-1 α -hydroxylated vitamin D analogue. *Breast Cancer Res.* 2005, 7(6), 980-986.
- [162] Agic, A.; Xu, H.; Altgassen, C.; Noack, F.; Wolfler, M. M.; Diedrich, K.; Friedrich, M.; Taylor, R. N.; Hornung, D. Relative expression of 1,25-dihydroxyvitamin D3 receptor, vitamin D 1 α -hydroxylase, vitamin D 24-hydroxylase, and vitamin D 25-hydroxylase in endometriosis and gynecologic cancers. *Reprod. Sci.* 2007, 14(5), 486-497.
- [163] Friedrich, M.; Rafi, L.; Mitschele, T.; Tilgen, W.; Schmidt, W.; Reichrath, J. Analysis of the vitamin D system in cervical carcinomas, breast cancer and ovarian cancer. *Recent Results Cancer Res.* 2003, 164, 39-46.
- [164] Hewison, M.; Kantorovich, V.; Liker H. R.; van Herle, A. J.; Cohan, P.; Zehnder, D.; Adams, J. S. Vitamin D-mediated hypercalcemia in lymphoma: evidence for hormone production by tumoradjacent macrophages. *J. Bone Miner. Res.* 2003, 18, 579-582.
- [165] Evans, K. N.; Taylor, H.; Zehnder, D.; Kilby, M. D.; Bulmer, J. N.; Shah, F.; Adams, J. S.; Hewison, M. Increased expression of 25-hydroxyvitamin D-1 α -hydroxylase in dysgerminomas: a novel form of humoral hypercalcemia of malignancy. *Am. J. Pathol.* 2004, 165, 807-813.
- [166] Townsend, K.; Banwell, C. M.; Guy, M.; Colston, K. W.; Mansi, J. L.; Stewart, P. M.; Campbell, M. J.; Hewison, M. Autocrine metabolism of vitamin D in normal and malignant tissue. *Clin. Cancer Res.* 2005, 11(9), 3579-3586.
- [167] Ainsleigh, H. G. Beneficial effects of sun exposure on cancer mortality. *Prev. Med.* 1993, 22, 132-140.
- [168] Gorham, E. D.; Garland, F. C.; Garland, C. F. Sunlight and breast cancer incidence in the USSR. *Int. J. Epidemiol.* 1990, 19, 820-824.
- [169] Garland, F. C.; Garland, C. F.; Gorham, E. D.; Young, J. F. Geographic variation in breast cancer mortality in the United States: a hypothesis involving exposure to solar radiation. *Prev. Med.* 1990, 19, 614-622.
- [170] Grant, W. B. An estimate of premature cancer mortality in the U.S. due to inadequate doses of solar ultraviolet-B radiation. *Cancer* 2002, 94, 1867-1875.
- [171] Kemmis, C. M.; Salvador, S. M.; Smith, K. M.; Welsh, J. Human mammary epithelial cells express CYP27B1 and are growth inhibited by 25-hydroxyvitamin D-3, the major circulating form of vitamin D-3. *J. Nutr.* 2006, 136, 887-892.
- [172] Holick, M. F.; Siris, E. S.; Binkley, N.; Beard, M. K.; Khan, A.; Katzer, J. T.; Petruschke, R. A.; Chen, E.; de Papp, A. E. Prevalence of vitamin D inadequacy among

- postmenopausal women. North American women receiving osteoporosis therapy. *J. Clin. Endocrinol. Metab.* 2005, 90, 3215-3224.
- [173] Sowers, M. R.; Wallace, R. B.; Hollis, B. W.; Lemke, J. H. Parameters related to 25-OH-D levels in a population-based study of women. *Am. J. Clin. Nutr.* 1986, 43, 621-628.
- [174] Diesing, D.; Cordes, T.; Fischer, D.; Diedrich, K.; Friedrich, M. Vitamin D--metabolism in the human breast cancer cell line MCF-7. *Anticancer Res.* 2006, 26(4A), 2755-2759.
- [175] Sunn, K. L.; Cock, T. A.; Crofts, L. A.; Eismann, J. A.; Gardiner, E. M. Novel N-terminal variant of human VDR. *Mol. Endocrinol.* 2001, 15(9), 1599-1609.
- [176] Tsai, M. J.; O'Malley, B. W. Molecular mechanisms of action of steroid/thyroid receptor superfamily members. *Annu. Rev. Biochem.* 1994, 63, 451-486.
- [177] Nemere, I.; Szego, C. M. Early actions of parathyroid hormone and 1,25-dihydroxycholecalciferol on isolated epithelial cells from rat intestine. *Endocrinology* 1981, 108, 1450-1462.
- [178] Studzinski, G. P.; McLane, J. A.; Uskokovic, M. R. Signaling pathways for vitamin D-induced differentiation: implications for therapy of proliferative and neoplastic diseases. *Crit. Rev. Eukaryot. Gene Expr.* 1993, 4, 279-312.
- [179] Marcinkowska, E.; Wiedlocha, A.; Radzikowski, C. 1,25-dihydroxyvitamin D₃ induced activation and subsequent nuclear translocation of MAPK is upstream regulated by PKC in HL-60 cells. *Biochem. Biophys. Res. Commun.* 1997, 241, 410-426.
- [180] Marcinkowska, E.; Wiedlocha, A.; Radzikowski, C. Evidence that phosphatidylinositol 3-kinase and p70S6K protein are involved in differentiation of HL-60 cells induced by calcitriol. *Anticancer Res.* 1998, 18, 3507-3514.
- [181] Marcinkowska, E. Evidence that activation of MEK1,2/erk1,2 signal transduction pathway is necessary for calcitriol-induced differentiation of HL-60 cells. *Anticancer Res.* 2001, 21, 499-504.
- [182] Mehta, R. G.; Mehta, R. R. Vitamin D and cancer. *J. Nutr. Biochem.* 2002, 13, 252-264.
- [183] Boland, R.; De Boland, A. R.; Buitrago, C.; Morelli, S.; Santillan, G.; Vazquez, G.; Capiati, D.; Baldi, C. Non-genomic stimulation of tyrosine phosphorylation cascades by 1,25(OH)₂D₃ by VDR-dependent and -independent mechanisms in muscle cells. *Steroids* 2002, 67, 477-482.
- [184] Marcinkowska, E.; Wiedlocha, A. Steroid signal transduction activated at the cell membrane: from plants to animal. *Acta Biochimica Polonica* 2002, 49, 735-745.
- [185] Colston, K. W.; Berger, U.; Coombes, R. C. Possible role for vitamin D in controlling breast cancer cell proliferation. *Lancet* 1989, 1, 188-191.
- [186] James, S. Y.; Mackay, A. G.; Colston, K. W. Vitamin D derivatives in combination with 9-cis retinoic acid promote active cell death in breast cancer cells. *J. Mol. Endocrinol.* 1995, 14(3), 391-394.
- [187] Welsh, J. E. Induction of apoptosis in breast cancer cells in response to vitamin D and antiestrogens. *Biochem. Cell. Biol.* 1995, 72(11-12), 537-545.
- [188] Berger, U.; Wilson, P.; McClelland, R. A.; Colston, K.; Haussler, M. R.; Pike, J. W.; Coombes, R. C. Immunocytochemical determination of estrogen receptor, progesterone receptor, and 1,25-dihydroxyvitamin D₃ receptor in breast cancer and relationship to prognosis. *Cancer Res.* 1991, 51, 239-244.
- [189] Welsh, J. Targets of vitamin D receptor signaling in the mammary gland. *J. Bone Miner. Res.* 2007, 22 Suppl 2, V86-90.

- [190] Uitterlinden, A. G.; Fang, Y.; Van Meurs, J. B.; Pols, H. A.; Van Leeuwen, J. P. Genetics and biology of vitamin D receptor polymorphisms. *Gene* 2004, 338(2), 143-156.
- [191] Guy, M.; Lowe, L. C.; Bretherton-Watt, D.; Mansi, J. L.; Peckitt, C.; Bliss, J.; Wilson, R. G.; Thomas, V.; Colston, K. W. Vitamin D receptor gene polymorphisms and breast cancer risk. *Clin. Cancer Res.* 2004, 15, 10(16), 5472-5481.
- [192] Curran, J. E.; Vaughan, T.; Lea, R. A.; Weinstein, S. R.; Morrison, N. A.; Griffiths, L. R. Association of A vitamin D receptor polymorphism with sporadic breast cancer development. *Int. J. Cancer* 1999, 83, 723-726.
- [193] Chen, W. Y.; Bertone-Johnson, E. R.; Hunter, D. J.; Willett, W. C.; Hankinson, S. E. Associations between polymorphisms in the vitamin D receptor and breast cancer risk. *Cancer Epidemiol. Biomarkers Prev.* 2005, 14(10), 2335-2339.
- [194] Sinotte, M.; Rousseau, F.; Ayotte, P.; Dewailly, E.; Diorio, C.; Giguere, Y.; Berube, S.; Brisson, J. Vitamin D receptor polymorphisms (FokI, BsmI) and breast cancer risk: association replication in two case-control studies within French Canadian population. *Endocr. Relat. Cancer.* 2008, 15(4), 975-83.
- [195] Trabert, B.; Malone, K. E.; Daling, J. R.; Doody, D. R.; Bernstein, L.; Ursin, G.; Marchbanks, P. A.; Strom, B. L.; Humphrey, M. C.; Ostrander, E. A. Vitamin D receptor polymorphisms and breast cancer risk in a large population-based case-control study of Caucasian and African-American women. *Breast Cancer Res.* 2007, 9(6), R84.
- [196] Lundin, A. C.; Soderkvist, P.; Eriksson, B.; Bergman-Jungstrom, M.; Wingren, S. Association of breast cancer progression with a vitamin D receptor gene polymorphism. South-East Sweden Breast Cancer Group. *Cancer Res.* 1999, 59, 2332-2334.
- [197] Ruggiero, M.; Pacini, S.; Aterini, S.; Fallai, C.; Ruggiero, C.; Pacini, P. Vitamin D receptor gene polymorphism is associated with metastatic breast cancer. *Oncol. Res.* 1998, 10, 43-46.
- [198] Dunning, A. M.; McBride, S.; Gregory, J.; Durocher, F.; Foster, N. A.; Healy, C. S.; Smith, N.; Pharoah, P. D.; Luben, R. N.; Easton, D. F.; Ponder, B. A. No association between androgen or vitamin D receptor gene polymorphisms and risk of breast cancer. *Carcinogenesis* 1999, 20, 2131-2135.
- [199] Newcomb, P. A.; Kim, H.; Trentham-Dietz, A.; Farin, F.; Hunter, D.; Egan, K. M. Vitamin D receptor polymorphism and breast cancer risk. *Cancer Epidemiol. Biomarkers Prev.* 2002, 11, 1503-1504.
- [200] Cui, Y.; Rohan, T. E. Vitamin D, Calcium, and breast cancer risk: A review. *Cancer Epidemiol. Biomarkers Prev.* 2006, 15(18), 1427-1437.
- [201] McCullough, M. L.; Stevens, V. L.; Diver, W. R.; Feigelson, H. S.; Rodriguez, C.; Bostick, R. M.; Thun, M. J.; Calle, E. E. Vitamin D pathway gene polymorphisms, diet, and risk of postmenopausal breast cancer: a nested case-control study. *Breast Cancer Res.* 2007, 9(1), R9.
- [202] Cashman, K. D. Calcium intake, calcium bioavailability and bone health. *Br. J. Nutr.* 2002, 87 Suppl 2, S169-177.
- [203] Welsh, J. Vitamin D and breast cancer: insights from animal models. *Am. J. Clin. Nutr.* 2004, 80, 1721-1724S.

- [204] Mantell, D. J.; Owens, P. E.; Bundred, N. J.; Mawer, E. B.; Canfield, A. E. 1,25-Dihydroxyvitamin D(3) inhibits angiogenesis in vitro and in vivo. *Circ. Res.* 2000, 87, 214–220.
- [205] Saez, S.; Falette, N.; Guillot, C.; Meggouh, F.; Lefebvre, M. F.; Crepin, M.; William, L. McGuire. Memorial Symposium. 1,25(OH)₂D₃ modulation of mammary tumor cell growth in vitro and in vivo. *Breast Cancer Res. Treat.* 1993, 27, 69–81.
- [206] Eisman, J. A.; Sutherland, R. L.; McMenemy, M. L.; Fragonas, J. C.; Musgrove, E. A.; Pang, G. Y. Effects of 1,25-dihydroxyvitamin D₃ on cell-cycle kinetics of T 47D human breast cancer cells. *J. Cell. Physiol.* 1989, 138(3), 611–616.
- [207] Jacobson, E. A.; James, K. A.; Newmark, H. L.; Carroll, K. K. Effects of dietary fat, calcium, and vitamin D on growth and mammary tumorigenesis induced by 7,12-dimethylbenz(a)anthracene in female Sprague-Dawley rats. *Cancer Res.* 1989, 49, 6300–6303.
- [208] Xue, L.; Lipkin, M.; Newmark, H.; Wang, J. Influence of dietary calcium and vitamin D on diet-induced epithelial cell hyperproliferation in mice. *J. Natl. Cancer Inst.* 1999, 91, 176–181.
- [209] John, E. M.; Schwartz, G. G.; Dreon, D. M.; Koo, J. Vitamin D and breast cancer risk: the NHANES I epidemiologic follow-up study, 1971–1975 to 1992. National Health and Nutrition Examination Survey. *Cancer Epidemiol. Biomarkers Prev.* 1999, 8, 399–406.
- [210] Knight, J. A.; Lesosky, M.; Barnett, H.; Raboud, J. M.; Vieth, R. Vitamin D and reduced risk of breast cancer: A population-based control study. *Cancer Epidemiol. Biomarkers Prev.* 2007, 16, 422–429.
- [211] McCullough, M. L.; Rodriguez, C.; Diver, W. R.; Feigelson, H. S.; Stevens, V. L.; Thun, M. J.; Calle, E. E. Dairy, calcium, and vitamin D intake and postmenopausal breast cancer risk in the Cancer Prevention Study II Nutrition Cohort. *Cancer Epidemiol. Biomarkers Prev.* 2005, 14, 2898–2904.
- [212] Rossi, M.; McLaughlin, J. K.; Lagiou, P.; Bosetti, C.; Talamini, R.; Lipworth, L.; Giacosa, A.; Montella, M.; Franceschi, S.; Negri, E.; La Vecchia, C. Vitamin D intake and breast cancer risk: a case-control study in Italy. *Ann. Oncol.* 2009, 20(2), 374–378.
- [213] Frazier, A. L.; Ryan, C. T.; Rockett, H.; Willett, W. C.; Colditz, G. A. Adolescent diet and risk of breast cancer. *Breast Cancer Res.* 2003, 5, R59–64.
- [214] Frazier, A. L.; Li, L.; Cho, E.; Willett, W. C.; Colditz, G. A. Adolescent diet and risk of breast cancer. *Cancer Causes Control* 2004, 15, 73–82.
- [215] Gissel, T.; Rejnmark, L.; Mosekilde, L.; Vestergaard P. Intake of vitamin D and risk of breast cancer—a meta-analysis. *J. Steroid. Biochem. Mol. Biol.* 2008, 111(3–5), 195–199.
- [216] Hiatt, R. A.; Krieger, N.; Lobaugh, B.; Drezner, M. K.; Vogelmann, J. H.; Orentreich, N. Prediagnostic serum vitamin D and breast cancer. *J. Natl. Cancer Inst.* 1998, 90, 461–463.
- [217] Bertone-Johnson, E. R.; Chen, W. Y.; Holick, M. F.; Hollis, B. W.; Colditz, G. A.; Willett, W. C.; Hankinson, S. E. Plasma 25-hydroxyvitamin D and 1,25-dihydroxyvitamin D and risk of breast cancer. *Cancer Epidemiol. Biomarkers Prev.* 2005, 14, 1991–1997.
- [218] Mawer, E. B.; Walls, J.; Howell, A.; Davies, M.; Ratcliffe, W. A.; Bundred, N. J. Serum 1,25-dihydroxyvitamin D may be related inversely to disease activity in breast cancer patients with bone metastases. *J. Clin. Endocrinol. Metab.* 1997, 82, 118–122.

- [219] Whitfield, J. F.; Boynton, A. L.; MacManus, J. P.; Sikorska, M.; Tsang, B. K. The regulation of cell proliferation by calcium and cyclic AMP. *Mol. Cell Biochem.* 1979, 27, 155–179.
- [220] Mathiasen, I. S.; Sergeev, I. N.; Bastholm, L.; Elling, F.; Norman, A. W.; Jaattela, M. Calcium and calpain as key mediators of apoptosis-like death induced by vitamin D compounds in breast cancer cells. *J. Biol. Chem.* 2002, 277, 30738–30745.
- [221] Sergeev, I. N. Calcium as a mediator of 1,25-dihydroxyvitamin D₃-induced apoptosis. *J. Steroid. Biochem. Mol. Biol.* 2004, 89–90, 419–425.
- [222] McGrath, C. M.; Soule, H. D. Calcium regulation of normal human mammary epithelial cell growth in culture. *In Vitro* 1984, 20, 652–662.
- [223] Russo, J.; Russo, I. H. The pathway of neoplastic transformation of human breast epithelial cells. *Radiat. Res.* 2001, 155, 151–154.
- [224] Boyapati, S. M.; Shu, X. O.; Jin, F.; Dai, Q.; Ruan, Z.; Gao, Y. T.; Zheng, W. Dietary calcium intake and breast cancer risk among Chinese women in Shanghai. *Nutr. Cancer* 2003, 46, 38–43.
- [225] Wark, J. D.; Larkins, R. G.; Eisman, J. A.; Wilson, K. R. Regulation of 25-hydroxyvitamin-D-1 α -hydroxylase in chick isolated renal tubules: effects of prostaglandin E₂, frusemide and acetylsalicylate. *Clin. Sci.* 1981, 62, 53–59.
- [226] Hayes, M. E.; Rai, A.; Cooper, R. G.; Bayley, D.; Freemont, A. J.; Mawer, E. B. Inhibition by prostaglandin E₁ and E₂ of 1,25-dihydroxyvitamin D₃ synthesis by synovial fluid macrophages from arthritic joints. *Ann. Rheum. Dis.* 1992, 51, 632–637.
- [227] Miller, G. J. Vitamin D and prostate cancer: biologic interactions and clinical potentials. *Cancer Metastasis Rev.* 1998, 17, 353–360.
- [228] Konety, B. R.; Getzenberg, R. H. Vitamin D and prostate cancer. *Urol Clin. North Am.* 2002, 29, 95–106.
- [229] Zhuang, S. H.; Burnstein, K. L. Antiproliferative effect of 1 α ,25-dihydroxyvitamin D₃ in human prostate cancer cell line LNCaP involves reduction of cyclin-dependent kinase 2 activity and persistent G₁ accumulation. *Endocrinology* 1998, 139, 1197–1207.
- [230] Moreno, J.; Krishnan, A. V.; Swami, S.; Nonn, L.; Peehl, D. M.; Feldman, D. Regulation of prostaglandin metabolism by calcitriol attenuates growth stimulation in prostate cancer cells. *Cancer Res.* 2005, 65(17), 7917–7925.



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Cancer is the leading cause of death in most countries and its consequences result in huge economic, social and psychological burden. Breast cancer is the most frequently diagnosed cancer type and the leading cause of cancer death among females. In this book, we discussed various aspects of breast cancer carcinogenesis from clinics to its hormone-based as well as genetic-based etiologies for this deadly cancer. We hope that this book will contribute to the development of novel diagnostic as well as therapeutic approaches.

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