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From Pluripotency to Early Differentiation of Human Embryonic Stem Cell Cultures Evaluated by Electron Microscopy and Immunohistochemistry

Janus Valentin Jacobsen, Claus Yding Andersen,
Poul Hyttel and Kjeld Møllgård
*University of Copenhagen
Denmark*

1. Introduction

Under appropriate culture conditions human embryonic stem cells (hESCs) can retain an undifferentiated state during numerous passages (Thomson et al., 1998). In the undifferentiated state, hESCs express characteristic markers like NANOG, OCT4, TDGF1, DNMT3B, GABRB3, and GDF3, and are maintained by plating undifferentiated cells or colonies of cells into new culture dishes with fresh medium every 7 to 10th day (Adewumi et al., 2007). Following periods exceeding 7 to 10 days in culture without passage, the cell population tends to become heterogeneous with differentiation starting to occur within a given colony or in various parts of a culture dish. The tendency for undergoing differentiation is independent of whether feeder cells, protein matrixes, or special plastic surfaces are used and what specific hESC medium is employed. Although an increasing density of cells during culture has been suggested to be one reason for spontaneous differentiation of cells to occur, it is also well known that morphologically perfect undifferentiated hESCs often appear in very high density in the same culture dish, even when differentiation has started to occur (Laursen et al., 2007). The transition from the undifferentiated state to more differentiated cell types appears to take place as a gradual process in colonies of hESCs, and it is currently not known how the ultrastructural organization of cells changes along with the differentiation process as defined from immunohistochemical differentiation markers. In the present study we have performed a spatiotemporal investigation on the differentiation of hESC colonies by electron microscopical and immunohistochemical approaches.

2. Materials and methods

2.1 Culture of human embryonic stem cell lines LRB008, LRB010 and LRB017

The hESC lines LRB008, LRB010, and LRB017 were established at the Laboratory of Reproductive Biology, Copenhagen University Hospital from surplus embryos donated by couples undergoing IVF treatment after informed consent. The study was approved by the regional ethical committee of Copenhagen and Frederiksberg Municipalities (permission no.

KF 01-188/03). Donated embryos that developed to the blastocyst stage, were used to derive hESC lines following isolation of the inner cell mass isolated by manual dissection using hypodermic needles. The zona pellucida was digested by pronase (1mg/mL) (Sigma-Aldrich, P8811), and the inner cell masses (ICMs) were isolated by immunosurgery. The immunosurgery was performed by incubation in rabbit anti-whole serum antibody (Sigma-Aldrich, H8765) diluted 1:3 in KnockOut Dulbeccos's modified Eagles medium (KO-DMEM) (Invitrogen, 10829-018) for 30 min, followed by three successive washes in KO-DMEM and incubation in guinea pig serum (State Serum Institute, Copenhagen) diluted in 1:5 KO-DMEM. The ICMs and the derived cell lines were cultured on mitotically inactivated mouse embryonic fibroblast (MEF) cells in hESC medium based on KO-DMEM supplemented with knockout serum replacement (15% of final concentration), L-glutamine (2 mmol/L), β -mercaptoethanol (0.1 mmol/L), MEM non-essential aminoacids (0.1 mmol/L), Penicillin/Streptomycin (50U/g/mL), and basic fibroblast growth factor (bFGF, 4 ng/mL). The cells were cultured in a humidified atmosphere consisting of 6% CO₂, 7% O₂ and 87% N₂ at 37°C. Cell lines were normally passaged onto fresh MEF feeders once a week, when confluence reached around 70-80%. Passage of cells was carried out by mild trypsin treatment (0.05% trypsin in EDTA). An exception to MEFs as supportive feeders was the use of mitotically inactivated human foreskin fibroblast (hFF) in experiments where cell lines LRB008 and LRB017 were harvested for scanning electron microscopy (SEM).

2.2 Parallel sampling of colonies from the same culture dishes

For optimal concurrency in the collection of material for electron microscopy and immunohistochemistry, the experimental design was based on parallel sampling from the same culture dishes as illustrated in Figure 1. Colonies from the hESC line LRB010 were cultured for a period of either 6, 8, 11, 14 or 17 days. Samples were pooled in groups of *early* stage (6 and 8 days), *intermediate* stage (11 and 14 days), and *late* stage (17 days) colonies, according to duration of cultures at harvest. In addition, colonies of two other hESC lines, LRB008 and LRB017, were cultured and harvested after 7 and 14 days, for immunohistochemistry and investigation of colony surface ultrastructure by scanning electron microscopy (SEM).

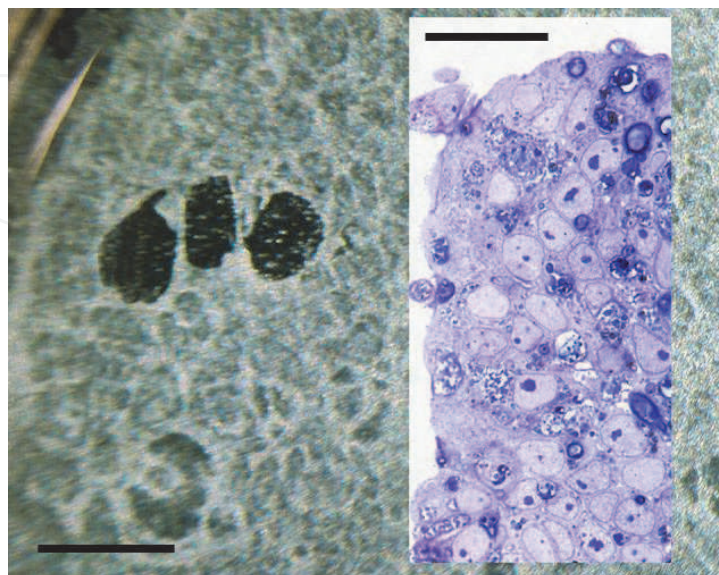


Fig. 1. The principle of parallel sampling.

The large greyscale phase contrast micrograph shows the bottom of a culture dish with growth of the hESC line LRB010 on a HFF feeder cell layer. Three black holes can be observed next to each other, where hESC colonies have been removed manually by hypodermic needles for fixation in glutaraldehyde and subsequent ultrastructural studies. The light micrograph shown in the inset depicts a semithin survey section (2 μm) with part of a fixed colony embedded in epoxy resin and stained with toluidine blue. After removal of tissue samples for electron microscopy, the remaining layer within the culture dish was fixed with Bouin's fixative for immunohistochemistry. Phase contrast micrograph: Scale bar = 5 mm. Light micrograph insert: Scale bar = 25 μm .

2.3 Processing of tissue for transmission electron microscopy (TEM)

Cultured hESC lines were detached using hypodermic needles and fixed in 3% glutaraldehyde in 0.1 M Na-phosphate buffer (pH 7.3) for 1-2 h at 4°C. Following fixation, specimens were transferred into 0.1 M Na-phosphate buffer (pH 7.3) and stored at 4°C for later processing. Specimens were embedded in 4% agar at 45°C (Bacto-agar, Difco Laboratories, Detroit, USA) under stereo microscopy and post-fixed in 1% OsO₄ in 0.1 M Na-phosphate buffer (pH 7.3) for 1 h at room temperature followed by wash in 0.1 M Na-phosphate buffer (pH 7.3) for 5 min. Tissue samples were stained en-bloc with 1% uranyl acetate in MilliQ water (MilliRO Plus and MilliQ PF Plus Water Purifications Systems, Millipore A/S, Hedehusene, Denmark), dehydrated in a series of ascending concentrations of ethanol (50% for 10 min, 70% for 10 min, 96% for 10 min, 99% for 3x20 min). Following the dehydration steps, samples were embedded in Epon (TAAB 812 Embedding resin, Medium), using propylene oxide as an intermedium, and polymerized for 48 h at 60°C. Semithin sections (2 μm) were cut on an ultramicrotome (Reichert Ultracut UCT, Leica) using glass knives prepared on a knifemaker (LKB Bromma 7800). Sections were then stained with 1% basic toluidine blue for evaluation by bright-field light microscopy until a satisfactory part of the colony was exposed. Sections of interest were re-embedded (Hyttel and Madsen, 1987) for further ultrathin sectioning (70 nm) on an ultramicrotome (Reichert Ultracut UCT, Leica) using a diamond knife (Jumdi, 2 mm). The ultrathin sections were contrast stained using 2% uranyl acetate in MilliQ water and lead citrate (Reynolds, 1963), collected on 150 mesh copper grids covered with a parlodion/amylacetate film, and examined using a transmission electron microscope (CM100, Philips, Darmstadt, Netherlands).

2.4 Processing of tissue for scanning electron microscopy (SEM)

Cultures of hESCs grown on glass coverslips were fixed in 3% glutaraldehyde diluted in 0.1 M Na-phosphate buffer (pH 7.3) for approximately 1 h at 4°C, then transferred into 0.1 M Na-phosphate buffer (pH 7.3) and stored at 4°C. On the day of further processing the specimens were washed 3x5 min in 0.1 M Na-phosphate buffer (pH 7.3), post-fixed in 1% OsO₄ in 0.1 M Na-phosphate buffer (pH 7.2) followed by additional 3x5 min washings in 0.1 M Na-phosphate buffer (pH 7.3). Glass coverslips with hESC colonies were then transferred from plastic growth plates into glass dishes and dehydrated in a series of ascending concentrations of acetone (25% for 10 min, 40% for 10 min, 60% for 10 min, 75% for 10 min, 90% for 10 min, 100% for 3x20 min). Glass coverslips with colonies were too large to fit into chambers of the critical point dryer and therefore had to be fractioned into smaller pieces under stereo microscope. The intermediate fluid acetone, was eliminated from the cells using a critical point dryer (EMS850, Electron Microscopy Sciences, Hatfield, Pennsylvania,

USA) by several flushings with liquid CO₂. Subsequently, critical point drying was performed. Pieces of coverslips with dried colonies were then mounted on specimen holders and coated with 5 nm gold/palladium in a sputter coater (SC7640 Suto/Manual High Resolution Sputter Coater, Quorum Technologies, Newhaven, UK). Specimens were evaluated using a scanning electron microscope (FEI Quanta 200, FEI Company, Eindhoven, The Netherlands).

2.5 Processing of tissue for immunohistochemistry

Colonies of the three hESC lines (LRB008, LRB010 and LRB017) were cultured under conditions as described above for 6, 7, 8, 11, 14 and 17 days and fixed in Bouin's fixative *in situ* in the culture dish following removal of material intended for electron microscopy. After 1 to 2 hours of fixation the fixative was replaced with 70 % ethanol and 24 – 48 hours later the 70 % ethanol was replaced with 90 % ethanol. Following overnight dehydration in 90 % ethanol, the samples were exposed to 99 % ethanol for 12 hours. Then the colonies were gently dissected free from the bottom using a Cell Scraper (Nunc) starting from the periphery. Colonies were then lifted carefully from the bottom of the culture dish to a small metal embedding mould into which xylene was pipetted. After 1 hour's exposure to xylene, paraffin was gently added to the embedding mould. Following paraffin embedding specimens were cut in 3-5 µm thick serial horizontal sections, strictly in parallel to the bottom of the cultures (Brøchner et al., 2010). Every 10th section was stained by toluidine blue.

Prior to immunohistochemistry, non-specific binding was inhibited by incubation for 30 minutes with blocking buffer (ChemMate antibody diluent S2022, DakoCytomation, Glostrup, Denmark) at room temperature. The sections were incubated overnight at 4°C with rabbit polyclonal or mouse monoclonal antibodies against OCT4, occludin, ZO-1, claudin-5, β-catenin, E-cadherin, N-cadherin, vimentin and nestin (details in Table 1). The sections were then washed with TBS and incubated for 30 minutes with the REAL EnVision™ Detection System, Peroxidase/DAB+, Rabbit/Mouse, (K5007) from DakoCytomation. The sections were counterstained with Mayer's hematoxylin and dehydrated in graded alcohols followed by xylene and coverslipped with DPX mounting media. Control sections were incubated with mouse IgG1, IgG2a or irrelevant rabbit antibodies, as well as subjected to omission of primary or secondary antibodies. Regarding OCT4, preincubation was performed 1 hour before incubation with the corresponding peptide in the proportion 1 to 5.

Antigen	Manufacturer	Code nr.	Antibody-species		
β-catenin	TriChem	17C2	Mouse IgG2a	÷	1:1000
Claudin-5	Abcam	Ab 5106	Rabbit	M6	1:100
E-cadherin	Abcam	Ab1416	Rabbit	M6	1:50
N-cadherin	Abcam	Ab12221	Rabbit	M6	1:500
Nestin	Chemicon	MAB 5326	Mouse, IgG1	÷	1:500
Occludin	Abcam	Ab64482	Rabbit	M6	1:75
OCT4	Abcam	Ab 19857	Rabbit	TEG	1:400
Vimentin	Dako	M0725	Mouse	M6	1:100
ZO-1	Zymed	Z-R1	Rabbit	÷	1:400

Table 1. Primary antibodies, manufactures, code numbers and dilutions.

3. Results

The cells and colonies grown on MEF feeders or hFF feeders in a culture medium with bFGF added developed well, with a typical morphology of undifferentiated stem cells showing areas of large single nucleated cells with scanty cytoplasm interspersed with cells of more differentiated morphology depending on culture age.

3.1 Transmission electron microscopy

Early stage colonies: In 2 μm survey sections from Epon-embedded material, the colonies appeared multi-layered with the highest numbers of cell layers in the center, flattening towards the periphery. Three different cell compartments within the colony were chosen for closer ultrastructural examinations, as illustrated by rectangles in Fig. 2, and consisted of: 1) an apical compartment with cells facing the culture medium; 2) a basal compartment containing cells anchoring the colony to the feeder cell layer, and; 3) a compartment in the center of the colony demarcated by apical and basal cells. In the apical compartment (Fig. 2C-E), cells were polarized with a large apical cytoplasm, rich in organelles, and nuclei located basally. Cells were connected to each other by apical junctional complexes – zonulae occludentes (tight junctions) and zonulae adherentes (adherens junctions), with abundant association of intermediary filaments of the cytoskeleton. Microvilli were observed projecting from the cell surfaces. Below the apical epithelial covering of the colony, a population of cells with a large nucleus to cytoplasm ratio and poor in organelles, representing a typical undifferentiated ESC-like morphology, resided in the center of the colony (Fig. 2B). Cell membranes of the basal cell compartment did not show any sign of junctional specializations, and higher magnification confirmed a continuous even spacing between the plasma membranes of the basal cells (Fig. 2F and G).

Intermediate stage colonies: The 2 μm survey sections showed a colony thickness of one or few cell layers. The colony harvested the 11th day of culture was still attached to a monolayer of MEF feeder cells with a distinct basement membrane separating them. Towards the apical surfaces, well developed tight and adherens junctions at the lateral plasma membrane connected cells of the apical compartment. Below the junctional complex, well-developed gap junctions and maculae adherentes (desmosomes) could be observed. All junctional complexes were closely associated with extensive bundles of intermediate filaments (Fig. 3C-D and 4D). In basal regions, adjacent cells were interdigitating and showed many gap junctions, whereas tight junctions, adherens junctions, and desmosomes were lacking (Figs. 3A, B, E & F and 4A, B, C & E).

Late stage colonies: In 2 μm survey sections, basal and center compartments were easily identified. Remnants of the basement membrane, but no feeder cells, were observed beneath the basal cells of the colony (Fig. 5A). The apical compartment contained a population of cells with obvious epithelial specializations such as junctional complexes with associated desmosomes, polarized organelle rich cytoplasm with basally located nuclei, and microvilli covering the apical surfaces. As was the case for early and intermediate stage colonies, a network of intermediary filaments radiated from sites of junctional complexes, where gap junctions were also present (Fig. 5C-E). At this late stage, a population of cells with ESC-like undifferentiated morphology similar to the ones previously described was still present (Fig. 5B).

Specializations linking adjacent plasma membranes of the basal compartment apart from obvious gap junctions were not observed (Fig. 5F-G).

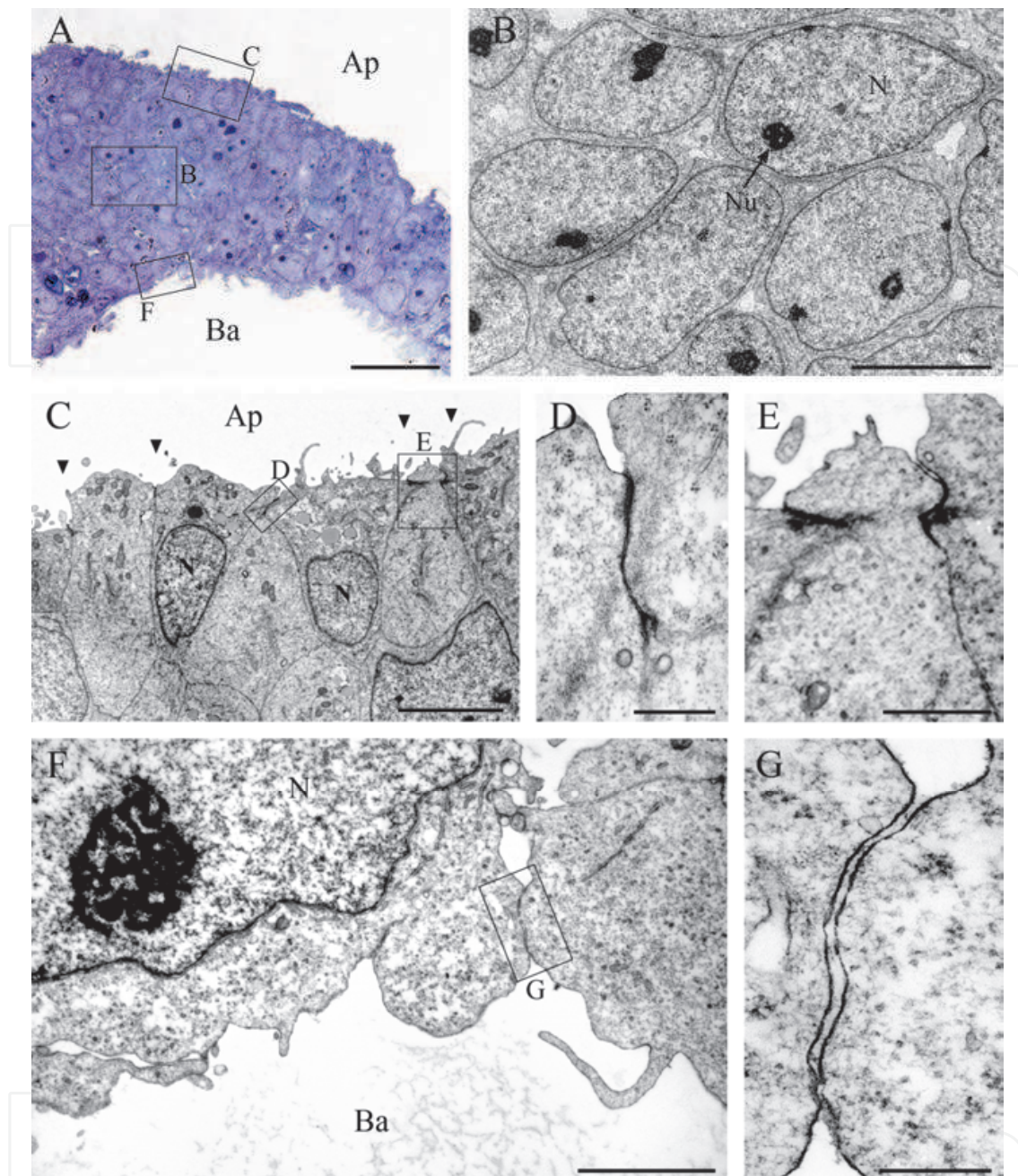


Fig. 2. Transmission electron micrographs of hESC line LRB010 cultured for 6 days (early). **(A)** Semithin cross section illustrating 3 different spatial compartments within the colony: Upper rectangle represents the localization of an apical compartment facing the culture medium *in vitro*, lower rectangle illustrates a population of basal cells anchoring the colony to the feeders, and the rectangle in the centre represents cells in between the apical and the basal compartment. Stained with toluidine blue. Ap, apical; Ba, basal. Scale bar = 25 μm . **(B)** Representative area from the center compartment with cells of typical undifferentiated morphology. N, nucleus; Nu, nucleolus. Scale bar = 5 μm . **(C)** Apical region of the colony with tight junction specializations shown in inserts. Scale bar = 5 μm . **(D)** Tight junction complex from insert in C (left). Scale bar = 500 nm. **(E)** Tight junctions from inset in C (right). Scale bar = 1 μm . **(F)** Basal region of the colony with a representative junction framed. Scale bar = 2 μm . **(G)** High magnification of inset in (F) showing no signs of junction specializations. Scale bar = 250 nm.

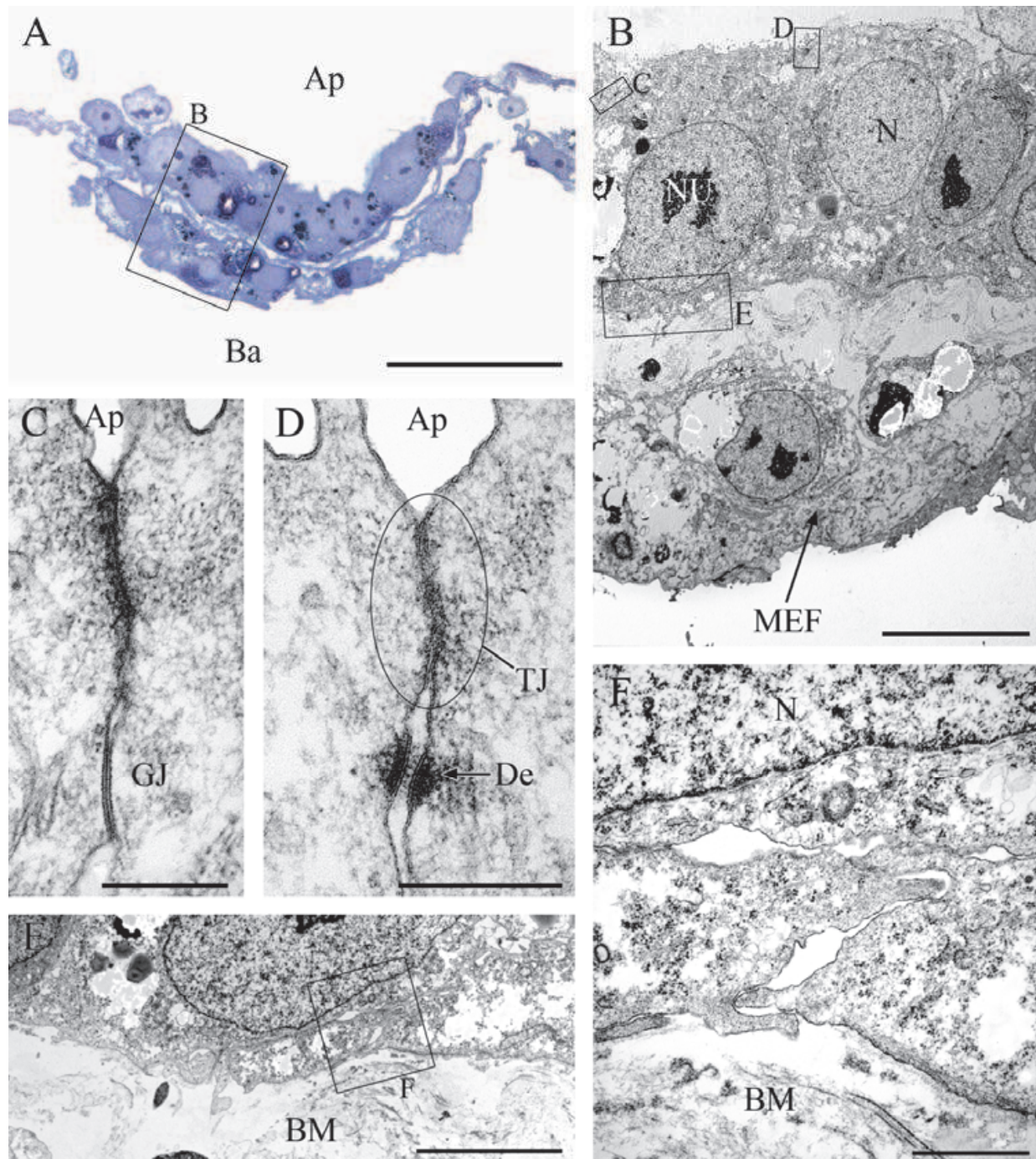


Fig. 3. Transmission electron micrographs of hESC line LRB010 cultured for 11 days (intermediate).

(A) Light microscopy of semithin cross section of colony with framed area spanning the apical and basal compartments magnified in B-F. Ap, apical; Ba, basal. Scale bar = 50 μm . (B) Apical and basal regions further magnified are framed. MEF, mouse embryonic fibroblast feeder layer; N, nucleus; Nu, nucleolus. Scale bar = 10 μm . (C) Junctional complex with distinct gap junction (GJ). Scale bar = 250 nm. (D) Tight- and adherens junction (TJ) with associated desmosome (De). Scale bar = 250 nm. (E) Basal region showing interdigitating processes without junctional specializations. BM, basement membrane. Scale bar = 10 μm . (F) Higher magnification of insert in E. Scale bar = 1 μm .

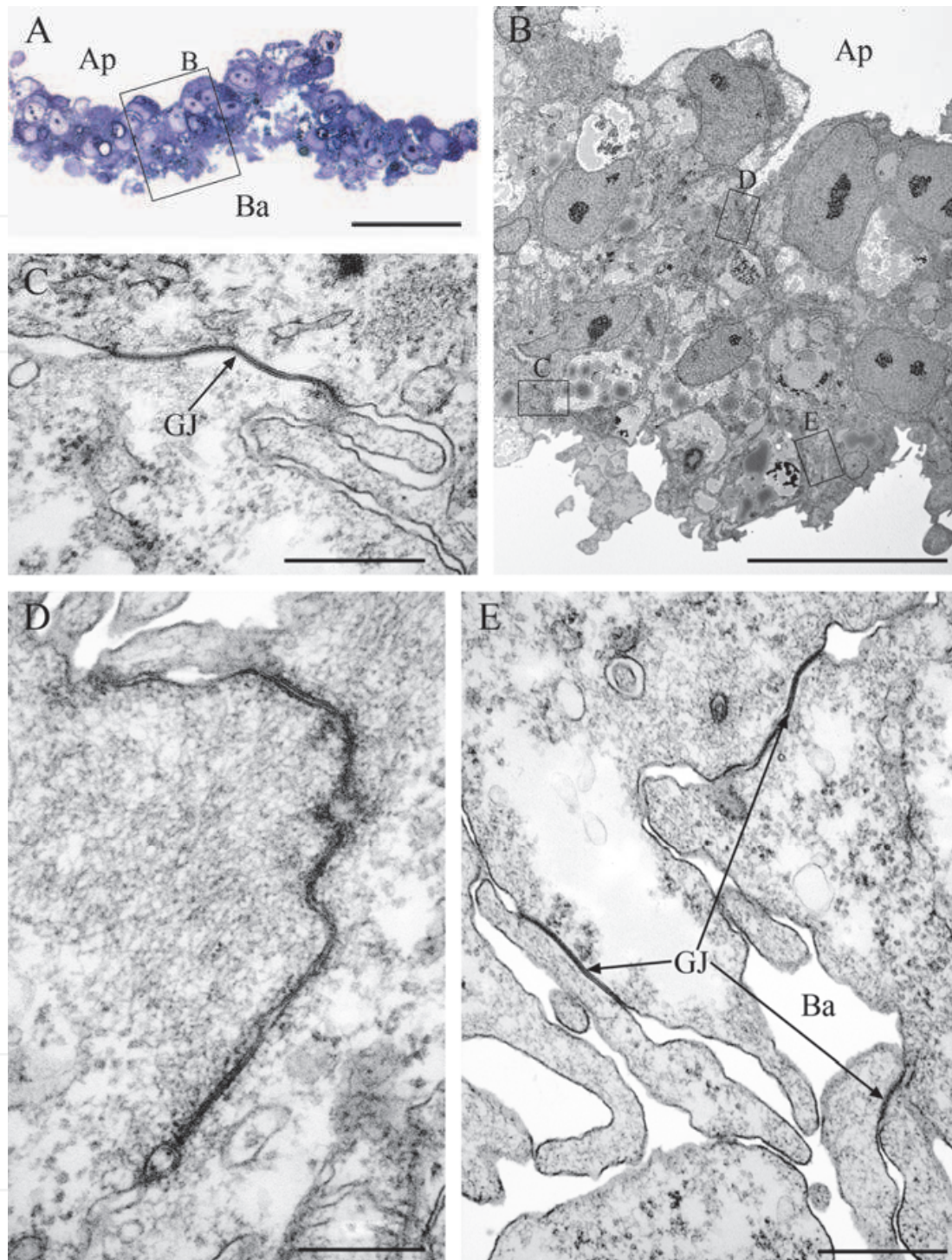


Fig. 4. Transmission electron micrographs of hESC line LRB010 cultured for 14 days (intermediate).

(A) Light microscopy of semithin cross section of colony marking representative area (frame) chosen for further analysis. Ap, apical; Ba, basal. Scale bar = 50 μm . **(B)** Marked locations (frame) of junctional areas seen in (C-E). BM, basement membrane. Scale bar = 20 μm . **(C)** Gap junction (GJ) in connection to interdigitations in the basal region of the colony. Scale bar = 500 nm. **(D)** Tight- and adherens junction with prominent intermediary filament cytoskeleton. Scale bar = 250 nm. **(E)** Abundant gap junctions (GJ) at the basal region. Scale bar = 500 nm.

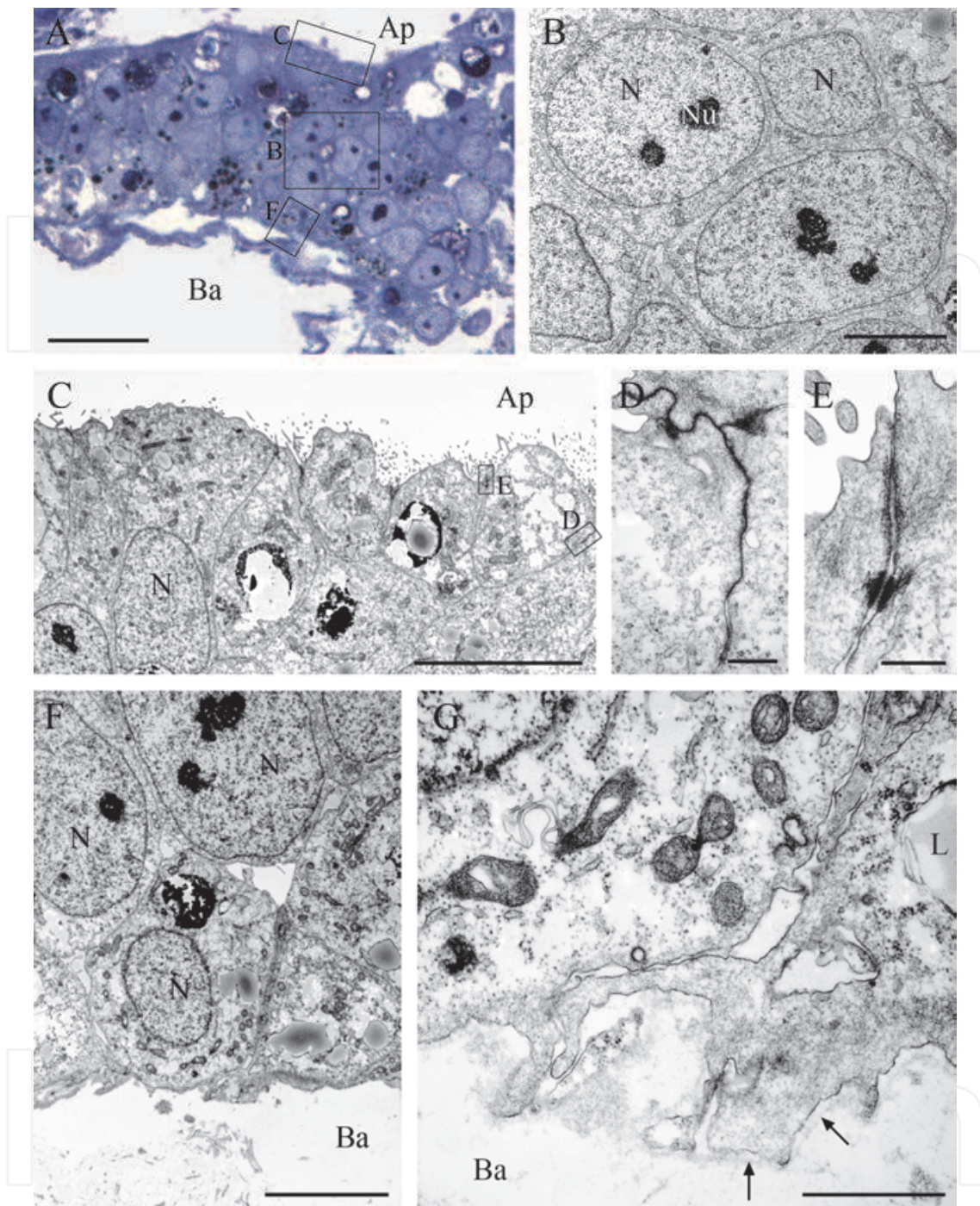


Fig. 5. Transmission electron micrographs of hESC line LRB010 cultured for 17 days (late). (A) Light microscopy of semithin overview section showing areas representative of apical, central and basal compartments of the colony. Ap, apical; Ba, basal. Scale bar = 20 μm . (B) Central compartment containing cells of typical undifferentiated morphology. N, nucleus. Scale bar = 5 μm . (C) Apical compartment where top layer of cells are bound together by apical junctional complexes (arrows). Two of the junctions can be seen in D-E. Scale bar = 10 μm . (D) Tight junction (C, left). Scale bar = 500 nm. (E) Tight- and adherens junction (C, right) with distinct desmosome. Scale bar = 250 nm. (F) Basal compartment. Scale bar = 5 μm . (G) Basal region without specializations. Basement membrane is marked by arrows. M, mitochondria; L, lipid droplet. Scale bar = 1 μm .

3.2 Scanning electron microscopy

Although 7 day-old colonies (LRB008 and LRB017) and feeders presented cracks resulting from the dehydration and critical drying processes, plenty of undamaged colony surface area was available for further examination. At low magnification, the colonies appeared as a flattened sheet of cells with a distinct boundary to the feeder cell layer they were grown on (Fig. 6A-B). The majority of the colony surface consisted of a fairly homogenous, flattened cell layer, but marked differences could be noted between individual cells in the flattened cell layer, with some cells having smooth surfaces, while others appeared with more rough surfaces covered by microvilli (Fig. 6C). Other surface areas displayed a more uniform

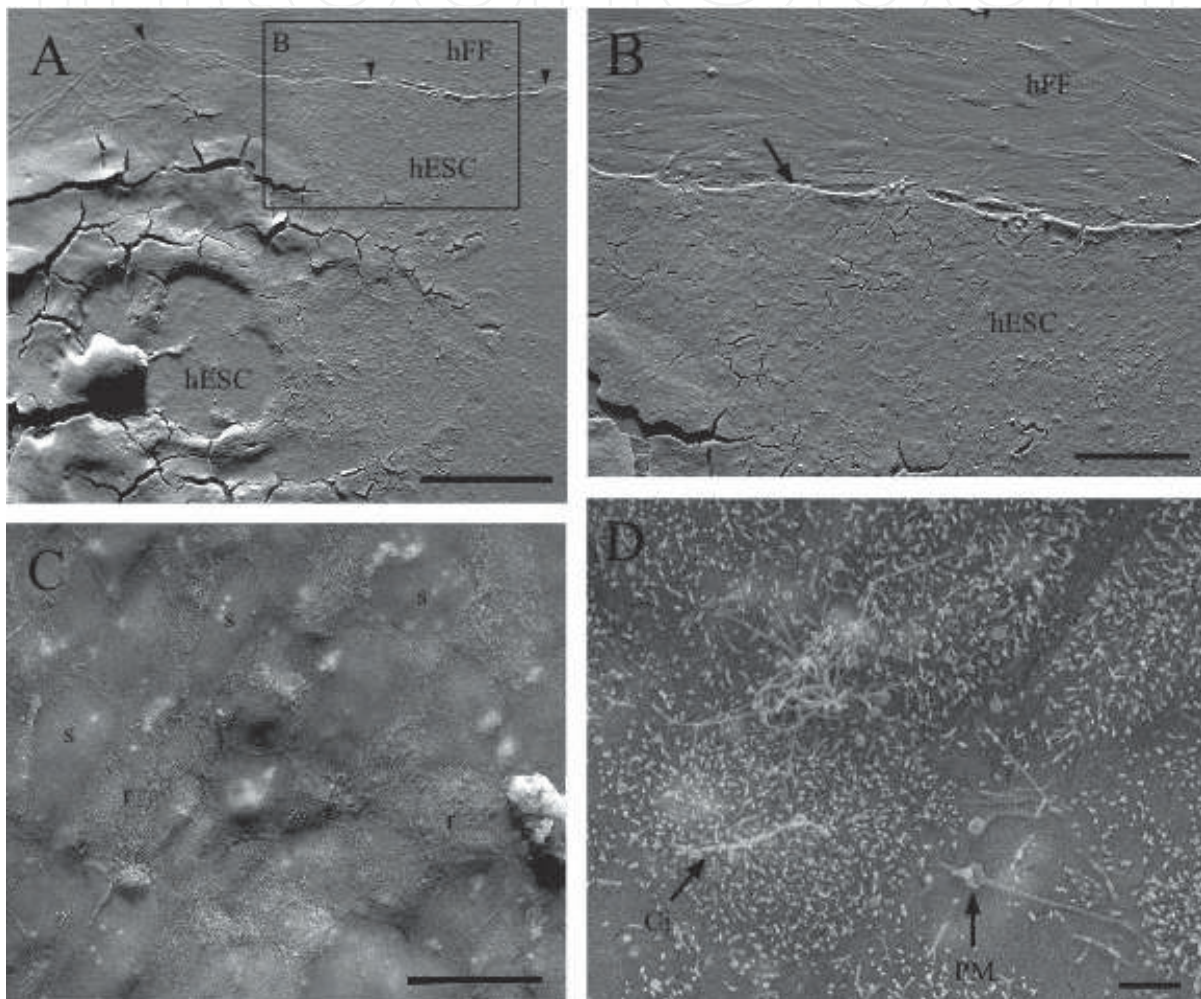


Fig. 6. Scanning electron micrographs of hESC lines LRB008 and LRB017 grown on human foreskin fibroblasts for 7 days in culture.

(A) Colony with arrowheads indicating border between colony and feeder layer. hFF, human foreskin fibroblasts; hESC, human embryonic stem cells. Scale bar = 500 μm . **(B)** Inset in (A) showing higher magnification of the periphery with arrow indicating border between colony and feeder layer. Scale bar = 200 μm . **(C)** Colony showing marked differences in cell surfaces. Some cells appear smooth surfaced (s), while others have a rougher surface (r) covered by microvillous extensions. Scale bar = 20 μm . **(D)** Microvillous surface in the periphery of the colony showing a cilium (Ci) extending from it, and a polar midbody (PM) still connecting dividing daughter cells. Scale bar = 2 μm .

distribution of microvilli, and the presence of individual cilia could be documented (Fig. 6D). Commonly, both cell lines exhibited superficial cytoplasmic bridges still connecting dividing daughter cells, with distinct polar midbodies (Fig. 6D).

Low magnification images of the 14 day-old colonies revealed great differences in the overall morphology of the two different cell lines investigated. One colony (LRB008) formed big bulging areas of cells growing in layers into the culture medium, while the other (LRB017) retained the flattened sheet-like appearance even after 14 days in culture (Fig. 7A-D). In the bulging areas of LRB008, separate cells clearly grew on top of each other, some forming structures protruding from the surface (Fig. 7A-B). Superficial cytoplasmic bridging between dividing cells was seen in LRB008, but not in LRB017. The rather uniform cell layer of LRB017 occasionally showed rounded protrusions of the plasma membrane, often in clusters (Fig. 7D). Furthermore, a varied microvillous covering could be observed throughout the colonies of both cell lines on bulging as well as homogenous flat areas.

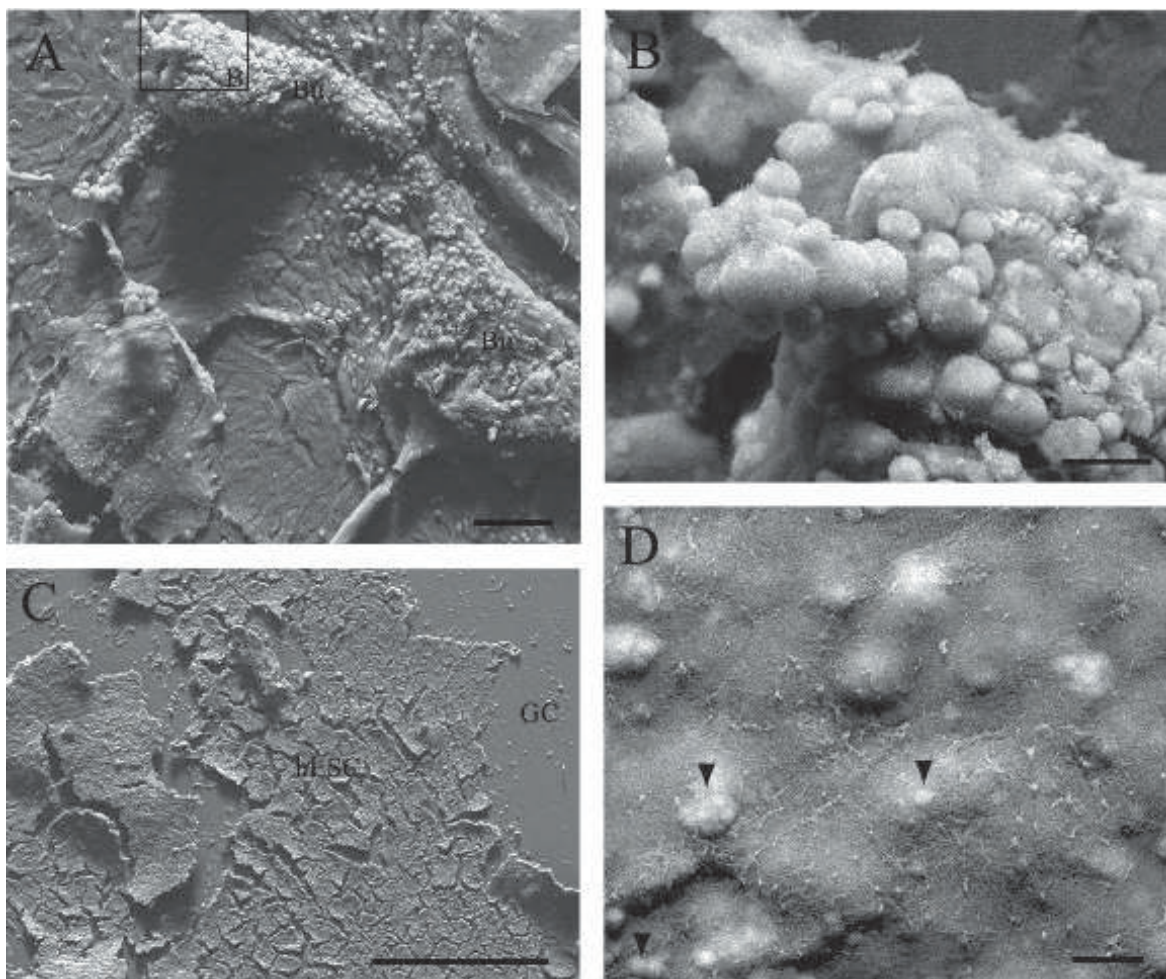


Fig. 7. Scanning electron micrographs of hESC lines LRB008 and LRB017 grown on human foreskin fibroblasts for 14 days in culture.

(A) Bulging (Bu) areas of differentiation on LRB008 after 14 days of culture. Scale bar = 100 μm . (B) Higher magnification of inset in A showing differentiating cells protruding from the surface. Scale bar = 20 μm . (C) Colony of LRB017 partly detached from the glass coverslip (GC) they were grown on. Scale bar = 400 μm . (D) Higher magnification of hESCs from (C) with areas of membrane protrusions marked by arrowheads. Scale bar = 5 μm .

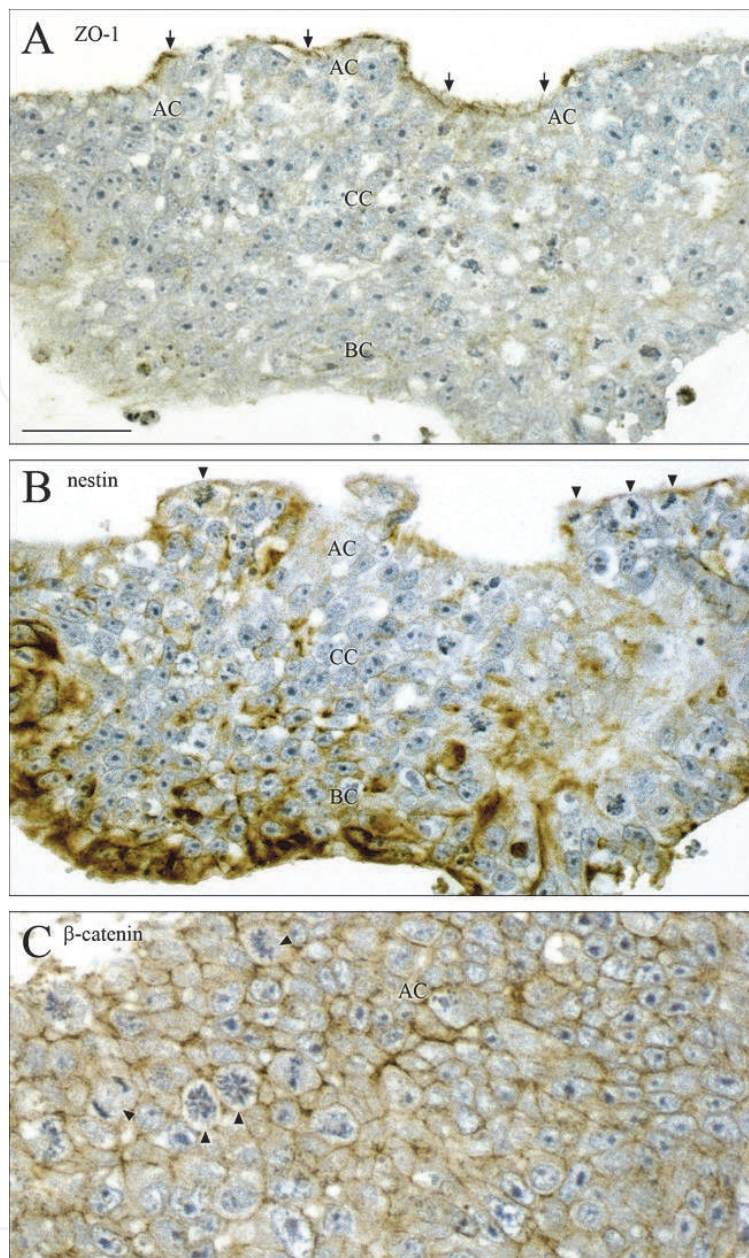


Fig. 8. Expression of ZO-1, nestin and β -catenin shown by immunohistochemistry in sections from hESC line LRB010 cultured for 6 days. In **(A)** stained for ZO-1 the apical compartment (AC) is separated from the culture medium by a tight junctional complex (arrows). The central compartment (CC) and the basal compartment (BC) show no distinct reactivity. **(B)** The basal compartment (BC) is strongly positive for nestin, whereas the central compartment (CC) is characterized by cells of typical undifferentiated morphology. Note the pattern of differentiation from the center towards the basal compartment indicating a gradual change from hESCs to nestin-positive neuroectodermal cells. Some of the many mitotic figures in the apical compartment (AC) situated at the apical surface facing the culture medium are indicated by arrowheads. **(C)** A tangential section through the apical compartment (AC) illustrates the strong β -catenin-reactivity associated with the adherens junctions. Note the many mitotic figures (arrowheads), which are also seen in the same region in cross-sectioned colonies (arrowheads in **B**). A, B & C: same magnification. Scale bar = 25 μ m.

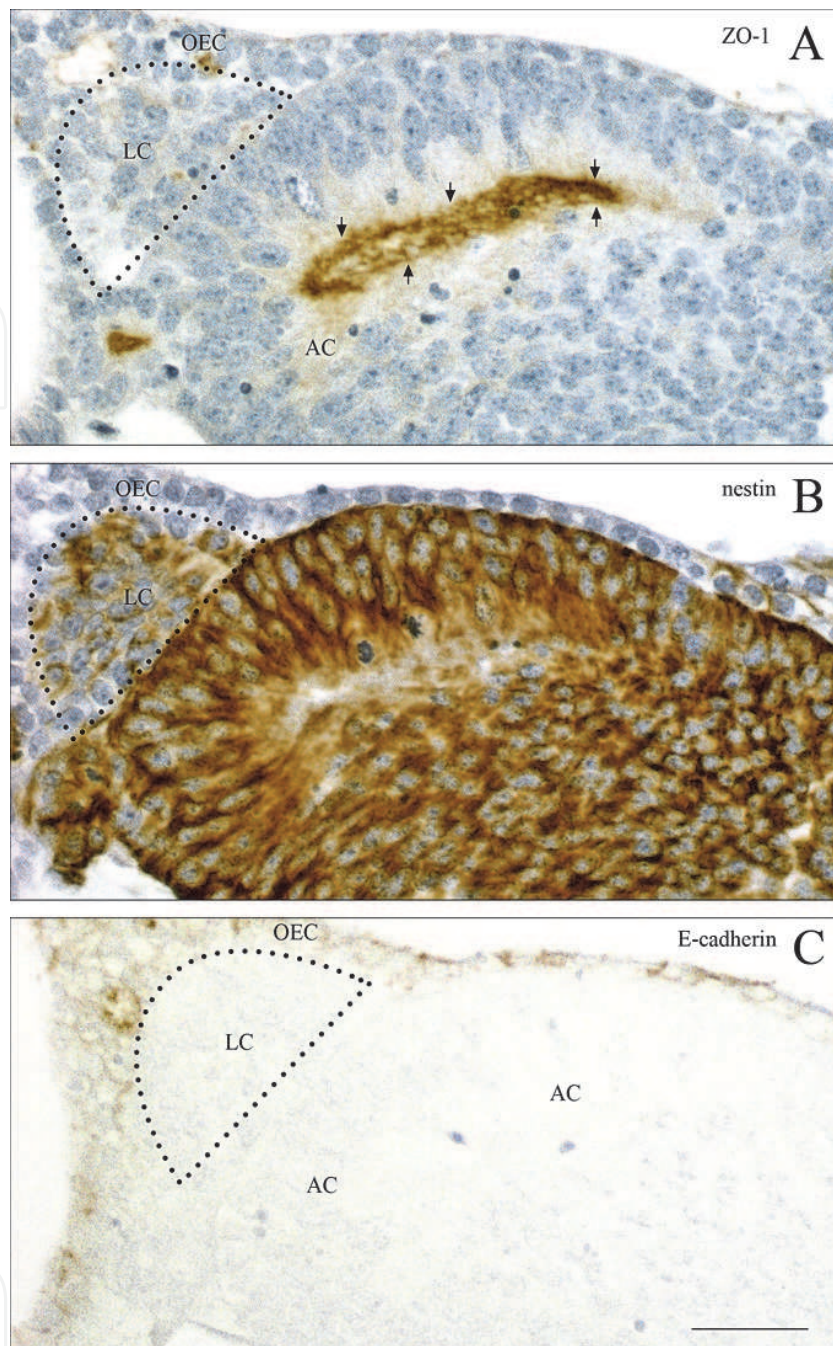


Fig. 9. Expression of ZO-1, nestin and E-cadherin shown by immunohistochemistry in tangential sections from hESC line LRB010 cultured for 17 days.

(A): The apical compartment (AC) in the center of the figure, stained for ZO-1 shows a well-developed tight junctional complex (arrows). A lateral compartment (LC) marked by a punctuated line and an outer epithelial compartment (OEC) are not stained. **(B)** The entire pseudo-stratified epithelium of the apical compartment is strongly positive for nestin. A new lateral compartment (LC) marked by a punctuated line is slightly positive for nestin, but strongly positive for vimentin and N-cadherin (not shown). **(C)** A single-layered outer epithelial compartment (OEC) and its extension towards the upper left of the figure show positive reactivity for E-cadherin, whereas both apical (AC) and lateral compartments (LC) are unstained. A,B & C: same magnification. Scale bar = 25 μ m.

3.3 Immunohistochemistry

It was possible to distinguish three different compartments in most colonies already from day 6: (1) An apical epithelial-like polarized layer facing the culture medium; (2) a basal cell layer facing the feeder cell layer; and (3) a central compartment enclosed by the apical and basal cell layers consisting of 'classical' hESCs with 'undifferentiated morphology' (Fig. 8 A-B). The youngest colony showed the presence of markers for tight junctions (occludin, claudin-5 and ZO-1 - Fig. 8A), and adherens junctions (E-cadherin associated with β -catenin - Fig. 8C) in the apical polarized epithelial-like cell layer. This was in marked contrast to the epithelioid 'classical' hESCs of the central compartment characterized by absence of markers for tight junctions. All cells exhibited positive OCT4-staining of their nuclei pointing to heterogeneity of the early hESCs, i.e. an epithelial-like cell type with junctional complexes facing the outside compartment (the culture medium) and an epithelioid cell type without classical junctional complexes, but with strong OCT4 reactivity in their nuclei. The basal cell layer, on the other hand, exhibited a distinct reactivity for the neuroectodermal marker nestin (Fig. 8B), but markers for tight junctions were absent. A gradual change in the pattern of differentiation from a nestin-negative, but OCT4-positive central compartment towards a strongly nestin-positive basal compartment indicating a gradual change from an OCT4-positive hESC population to a neuroectodermal cell population existed as illustrated in Fig. 8B.

In older colonies (17 days) immunohistochemical analysis revealed that the majority of the differentiating cells had lost their OCT4-positivity, indicating that the central compartment with 'classical' hESCs showing 'undifferentiated morphology' had disappeared in most colonies. An apical epithelial-like pseudostratified layer facing the culture medium was still present. The apical regions of the lateral cell membranes were joined by tight junctions as shown by a positive reactivity for occludin, claudin-5, and ZO-1 (Fig. 9A). In some cases the entire apical layer was densely stained for nestin (Fig. 9B), whereas the apical regions of other colonies showed a very strong staining for vimentin with some more basally located nestin-positive cells (not shown). A single layered outer compartment was characterized by a lack of staining for vimentin and nestin but by a positive staining reaction for E-cadherin (Fig. 9C). In a newly developed lateral compartment adjacent to the apical and above the central compartment cells showed a strongly positive N-cadherin and vimentin-staining but no staining for E-cadherin (Fig. 9C) suggesting an epithelial-mesenchymal transition (EMT) resulting in migratory vimentin-positive mesenchymal-like cells.

4. Discussion

Although living hESCs often appear homogeneously undifferentiated in their colony formation, as observed in phase contrast microscopy, the present study demonstrates that various differentiation events occur already within a few days after passage both at the ultrastructural and light microscopic level. Usually it is sufficient to maintain hESCs in an undifferentiated state by a passage of cells every 5-7 days. However, here we demonstrate that already within the early stage colonies, around the time of normal passage, a pronounced compartmentalization has taken place within colonies, suggesting spatial and temporal dimensions of differentiation. In all colonies examined, irrespective of the culture period and spatial configuration of the colony, the apical layer of cells, which is in direct

contact with the culture medium, formed an apico-basolateral polarized sheet of tightly interconnected cells resembling a columnar epithelium and thus providing solid documentation for a morphological specialization linking adjacent cells by junctional complexes consisting of tight and adherens junctions, shown by EM and immunohistochemistry. Such an epithelial coating, forming a barrier to the external environment, might provide deeper lying cells with a selective transcellular uptake of nutrients and other media components, thus creating a microenvironment supporting the undifferentiated state. A central compartment, containing cells with typical undifferentiated ESC-morphology, was indeed observed with EM and immunohistochemistry in both *early* and *medium* stage colonies, indicating long-term retention of an environment permissive of undifferentiated cell growth, regardless of surrounding peripheral differentiation. Indeed these observations might also explain why morphological undifferentiated hESC in culture dishes that has been grown for a prolonged period of time of perhaps two weeks or more is often found in the central and basal compartments which are in direct contact with bottom of the culture dish without tight junctions underneath layers of more differentiated cells on top of the colony.

Based upon studies of short-term feeder free cultures, Ullmann *et al.* (2007) proposed a colony structure almost identical to the *early* stage colony in this study. The immunostainings for E-cadherin, a marker for epithelial adherens junctions, were localized only to the uppermost cell layer of a multilayer, whereas it disappeared peripherally, where vimentin positive cells began to appear. This provides evidence for an epithelial-mesenchymal transition (EMT) in the peripheral regions where the colonies flattens into a monolayer. However, according to our TEM investigations, these peripheral monolayers of the *intermediate* stage colonies grown on MEF exhibited junctional complexes, hence suggesting an epithelial nature.

Colonies organized with confined undifferentiated cells and peripheral differentiation was confirmed by a shift in immunoreactivity from the pluripotency marker SSEA-4 to a marker of differentiation SSEA-1 towards the periphery (Johkura *et al.*, 2004). This down-regulation in pluripotency markers was to some extent also indicated in the short-term feeder free cultures referred to above, where the intensity in immunostainings of the key pluripotency markers OCT4 and NANOG decreased in peripheral regions (Ullmann *et al.*, 2007). Despite an apparent decrease in the amount of the key transcription factors, expression levels were still substantial, raising the question whether a deviation from the undifferentiated morphology to a more epithelial-like morphology is accompanied by decrease in potency. Additionally, increased amounts of laminin in the periphery of the colonies confirmed accumulation of extracellular matrices under standard culture conditions (Johkura *et al.*, 2004), most likely as a constituent of the basement membrane secreted by differentiated epithelial cells. One of the *intermediate* (11 days in culture) and the *late* stage colonies had distinct basement membranes. The basement membrane of the *intermediate* stage colony harvested at the 14th day of culture was supposedly teared of at harvest. This was indicated by the damaged appearance basally. Neither did our observations indicate presence of a basement membrane beneath the most basal cells of the multilayered *early* stage colony, possibly explaining colony detachment from feeders without damaging structures of the basal cell layer or lifting up accompanying feeders. The location of the basement membrane in the *late* stage colony (Fig. 5), with its

multilayered appearance, suggests that it could be a single-cell layer cut in a transverse section deviating somewhat from the perpendicular plane, thus increasing the interface of the section. On the other hand, a model of a multilayered colony consisting of several layered compartments cannot be excluded.

Gap junctions were identified both as part of the apical junctional complex (Fig. 4D), spanning all stages, but also in basal regions of the presumed monolayer in one of the *intermediate* stage colonies (Fig. 4E). A definite cell-to-cell coupling is demonstrated between epithelial cells of the apical compartment of murine cells of the ICM which are connected to each other (Lo & Gilula, 1979). The present study suggests that this also may apply to human ICMs. In accordance, Sathananthan *et al.* (2002) points out that a high degree of ultrastructural similarity exists between cells of the ICM and hESCs with undifferentiated morphology. Hence a gap junctional coupling could be expected between cells within the compartment of cells with undifferentiated morphology. Indeed, gap junctions were found by TEM within the undifferentiated compartment (micrographs not included), also demonstrating cell-to-cell coupling to exist between these cells. Whether the apical epithelial compartment is gap junctionally coupled to the underlying cells with undifferentiated morphology remains to be determined. Differentiation processes are normally associated with the progressive loss of communication between different tissues, for example as observed in between the ICM and the trophectoderm (Lo & Gilula, 1979). This suggests the possibility of two separate compartments intra-connected by gap junctions without coupling to each other. Huettner *et al.* (2006) demonstrated extensive gap junction communication in hESC colonies by dye injections and physiological recordings together with documentation of different connexins by immunocytochemistry and RT-PCR, assuming colonies to consist of uniform monolayers of undifferentiated cells corresponding to apical epithelial cells.

In order to document possible differences among different hESC lines, two other cell lines, LRB008 and LRB017, were included to improve our knowledge about the general colony outline and surface structures from a SEM point of view. At least after 7 days, this analysis confirmed that the periphery is composed of a primarily homogenous flat monolayer comparable to single-cell layers visualised by TEM. Cell line LRB017 showed a distinct accumulation of cells in the center of the colony. This was more difficult to identify in cell line LRB008, because the area in question had suffered considerable damage from processing of the sample, which in itself could indicate a thicker cell layer more vulnerable to shrinkage during dehydration. These observations agreed very well with the previously proposed colony structure described by Ullmann *et al.* (2007), but contradicted the saucer-shaped colony structure described by Sathananthan *et al.* (2002), in spite of identical culture conditions and time of harvest for the latter. The impact of duration of cultures is well illustrated by the 14-day old cultures. These showed a more chaotic growth of differentiating cells protruding from the surface in a highly irregular pattern in LRB008 (Fig. 7A-B), while LRB017 maintained a flattened growth. Single or clustered rounded protrusions in the cell membrane, in particular the 14-day-old cultures (Fig. 7D), showed similarity to the coarse particles and halos previously identified by phase contrast microscopy consisting of apoptotic cells and bodies (Johkura *et al.*, 2004). This suggests a pronounced cell death at this late stage. The fact that the entire colonies, in general, were

covered with microvilli with varying heterogeneity, and often with microvilli-free cells interspersed, suggests that the extent of epithelial specialization of the medium-facing cell layer. Heterogeneous cell surfaces between groups of cells, as well as between single adjacent cells, very well correspond to results from a study demonstrating regional differences in expression of specific markers for hESCs (Laursen *et al.*, 2007). Recently, one group published findings of primary cilia on surfaces of 33 and 90 percent of the hESCs of two different cell lines, H1 and LRB003, respectively (Kiprilov *et al.*, 2008). Additionally, cilia were shown to co-localize with OCT4 staining, thus indicating their pluripotency with the assumption of the associated hedgehog signalling machinery to be important in the maintenance of the undifferentiated/self-renewable state. Following 7 days in culture, with the same culture conditions and using two other hESC lines derived also derived in our laboratory, only one single cilium was found. This raises the question of importance of cilia in regulation of hESCs as proposed by Kiprilov *et al.* (2008), and definitely demonstrates differences among hESC lines.

To EMT, or not to EMT – in vivo and in culture. During *in vivo* embryogenesis and organ development the process initially termed epithelial-mesenchymal transformation (Hay, 2005), now referred to as epithelial-mesenchymal transition (EMT), (Kalluri & Weinberg, 2009; Thiery *et al.*, 2009) is a key event. EMT is a biological process that allows an epithelial cell to undergo multiple biochemical changes that enable it to assume a mesenchymal cell phenotype, which includes enhanced migratory capacity, invasiveness, elevated resistance to apoptosis, and increased production of extracellular matrix components (Kalluri and Weinberg, 2009). Early in the third week of human embryonic development, epiblast cells lateral to the primitive streak begin to move into the streak, where they undergo an EMT, i.e. the epithelial epiblast cells elongate and detach from their neighbours and then migrate away below the primitive streak as individual loosely organized mesendodermal progenitors. This process of gastrulation represents the first EMT after implantation (Thiery *et al.*, 2009). After gastrulation the remaining epithelial epiblast cells, which did not enter the primitive streak and therefore did not undergo the initial EMT, form epidermal and neural territories progressively along the rostrocaudal axis. The differentiating neural plate is flanked by the prospective neural crest and further laterally by the prospective surface ectoderm, which constitutes the remaining areas of the ectoderm. Neural crest cells from the neural crest territory in the dorsal neural tube then delaminate in a process of primary EMT (Thiery *et al.*, 2009). However, what has not really been emphasized so far, is the lack of EMT in the initial differentiation of epiblast cells directly into the remaining ecto- and neuroectodermal germ layer – an *in vivo* process of Epiblast-ectodermal transformation (EET) corresponding to an *in vitro* process of ESC-ectodermal transformation.

In a previous *in vitro* study of regional differences in the expression of markers during differentiation of hESC lines (LRB001-004), we noticed a distinct and direct transformation from undifferentiated OCT4 positive hESCs to SSEA1-positive fully differentiated epithelial cells (Laursen *et al.*, 2007), and in a very recent investigation of six different LRB-cell lines we observed a similar abrupt transformation between OCT4-positive hESC regions and areas with either ectodermal p63-positive cells or neuroectodermal nestin-positive cells, while a direct transition into either meso- or endodermal marker positive cells, was not seen (Brøchner *et al.*, 2011). In the present *in vitro* study we found an OCT4-positive

epithelial-like cell type with a well-defined junctional complex facing the outside compartment which strongly resemble an *in vivo* embryonic epiblast cell type, whereas the pluripotent cells without junctional complexes in the central compartment of early cultures resemble an ICM-like cell type. The gradual pattern of differentiation from the center towards both apical and basal compartments indicates a direct shift from hESCs to columnar epithelial epiblast-like cells and to nestin-positive neuroectodermal cells in the basal compartment.

5. Conclusion

In conclusion, the epithelial coating of the colony might serve two different purposes: (1) The tight junction component of the junctional complex forms a barrier for transepithelial transport between the culture medium and the internal environment of the colony; thus the epithelial cells could provide cells in central and basal compartments with nutrients and other media components, probably via a specific receptor mediated transcellular epithelial transport, in order to create a microenvironment supporting the undifferentiated stage of underlying cells within the central compartment. (2) There seems to be a direct transformation of pluripotent hESCs into ecto- and neuroectodermal germ layer cells, while the adherens component of the junctional complex is instrumental in directing the further differentiation of hESCs into mesodermal and endodermal lineages via an epithelial-mesenchymal transition (EMT).

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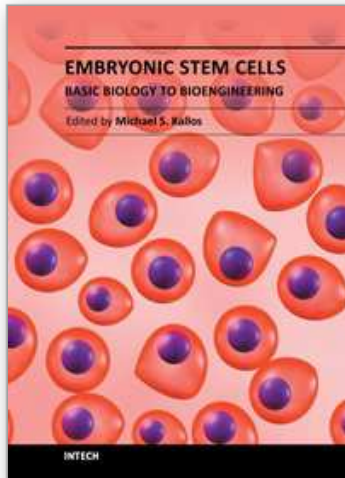
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Phone: +86-21-62489820
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