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Bisphosphonate-Related Osteonecrosis of the Jaw Around Dental Implants

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1. Introduction

Bisphosphonate is the collective name for compounds in which the backbone of pyrophosphoric acid, a P-O-P structure, is converted to chemically stable P-C-P, and this structure shows the affinity of bisphosphonate for bone hydroxyapatite in the body (Fleisch et al. 2002). Administered bisphosphonate preparations (BPs) transfer to and deposit in bone, exhibiting a bone resorption-inhibitory effect. The chemical structure of the side chain bound to the carbon atom of P-C-P markedly influences bisphosphonate activity, and, particularly, side chains containing nitrogen molecules not only markedly increase the affinity for HA but also bone resorption-inhibitory activity (Migliorati et al. 2005). Because of this potent bone resorption-inhibitory effect, BPs are the first-choice treatment for osteoporosis worldwide (Russell et al. 2007), and their efficacy for malignant tumorassociated hypercalcemia (Body et al.1999), ostealgia complicating bone metastasis of solid tumors (Hortobagyi et al. 1998), and multiple myeloma accompanied by bone destruction (Berenson et al. 1996) has been shown. Adverse effects of BPs were previously considered to be relatively mild, such as digestive symptoms induced by oral preparations and fever induced by injections (Berenson et al. 1996), but the association of BPs with osteonecrosis of the jaw has been frequently described since Marx (2003) initially reported it (Ruggiero et al. 2004; Marx et al. 2005; Khosla et al. 2007)

Bisphosphonate-related osteonecrosis of the jaw (BRONJ) is clinically diagnosed when the following 3 conditions are met: 1) current or previous treatment with BPs, 2) persistent exposure of necrotized bone in the maxillofacial region for 8 weeks or longer, 3) no past medical history of radiotherapy for the jaw bone (AAOMS 2007; Ruggiero et al. 2009). BPs inducing BRONJ mostly contain nitrogen (NBPs). BRONJ frequently develops in patients under NBP treatment upon dental treatment, such as tooth extraction, implant placement, and surgical periodontal treatment accompanied by bone invasion, and inflammatory diseases, such as periodontal disease and abscess (Ruggieri et al. 2009; Vahtsevanos et al. 2009; Yoneda et al. 2010). BRONJ shows poor responses to the standard treatment for common osteomyelitis of the jaw, such as curettage/resection of necrotized tissue and antimicrobial drugs, and its intractability is the most serious problem.

Stack & Epker (1995) initially reported the influence of BPs on dental implant treatment. They introduced a case in which all 5 favorably functioning implants in the lower anterior tooth region were lost after the 5-month administration of a BP containing no nitrogen, etidronatedisodium (Didronel; Procter and Gamble, Cincinnati, OH), for osteoporosis. The details of the bone wound healing process after implant loss or whether BRONJ developed were not described, but the author concluded that Didronel administration is a risk factor for implant treatment. However, several recent studies reported that implant treatment in the presence of NBPs was possible when the NBPs were oral drugs. Fugazzotto et al (2007) evaluated that a retrospective analysis of case records of patients with a history of oral BPs therapy treated as part of routine periodontal and implant treatment. They reported that no osteonecrosis was noted immediately postoperatively or during follow-up period 61 patients, and all implants were functioning successfully 12 to 24 months postinsertion.Grant et al. (2008) evaluated the clinical results of 115 patients who received oral BPs during implant treatment. They showed that there was no significant difference in treatment results between patients with and without oral BPs during implant treatment. It was also reported that no patients developed ONJ after implant treatment. Bell & Bell (2008) reported that bone grafting for implant treatment in 42 patients who received oral BPs for osteoporosis treatment was conducted safely. Therefore, no conclusion has been reached regarding whether oral BPs are contraindicated for implant treatment. On the other hand, it has been clarified that the risk of developing BRONJ upon open dental treatment rises in patients treated with NBP intravenous injection showing a potent bone adsorptioninhibitory action, and implant treatment is considered contraindicated for such cases (Khan et al. 2008; Ruggiero et al. 2009). However, there have been fewer reports on the influence of intravenous BPs on implants which have already acquired osseointegration and are favorably functioning, and the actual state is unclear.

We experienced the case of a patient who developed ONJ around implants in the maxilla, considered to be caused by intravenous BP administration for osteometastasis treatment of breast cancer, and underwent partial maxillectomy to relieve inflammation (Shirota et al. 2009). This chapter introduces clinical findings, the treatment and its outcome, and periimplant histopathological findings in the patient, and the developmental mechanism of periimplant BRONJ is discussed with a literature review.

2. Patient and methods

2.1 Outline of the patient

The patient was a 54-year-old female who visited our hospital for chief complaints of periimplant pain in the upper left molar region and bone exposure. The patient developed breast cancer about 6 years ago and underwent left mastectomy followed by irradiation and chemotherapy. However, multiple cancer metastases occurred about 2 years ago, and the patient underwent cancer chemotherapy and intravenous BPs injection.

The patient underwent implant treatment for the upper left molar region at a dental clinic immediately before the development of breast cancer, in which 2 implants were placed. These implants functioned without a problem for about 6 years, but peri-implant gingival recession, bone exposure, and pain appeared about one month ago, and the patient received antimicrobial drug administration, irrigation, and disinfection at a clinic, but the symptoms did not improve. Thus, the patient visited our department. On the first examination, brown-colored necrotized bone was exposed in the upper left first molar region on the buccal side.

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The peri-implant gingiva was painful and swollen with slight flare, but no implant mobility was observed (Fig.1). On panoramic radiography, mild bone resorption was noted in alveolar bone in the implant neck. Computed tomography (CT) revealed sequestration with an irregular trabecular bone structure around the implant (Fig.2). Biopsy was performed in consideration of metastasis of breast cancer to the oral cavity, but only inflammatory cell infiltration was observed without tumor cells.

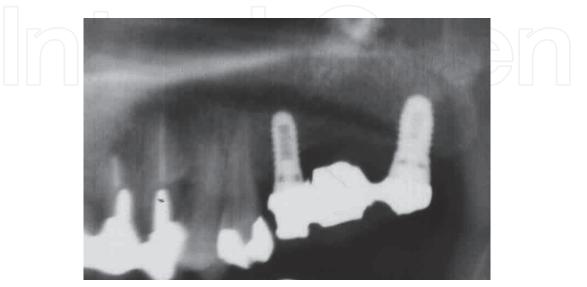


Fig. 1. Radiograph showing slight bone resorption around the implants.

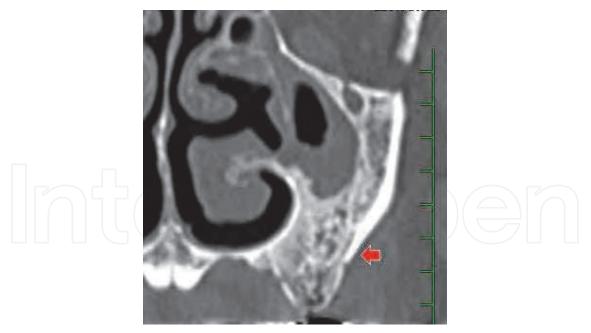


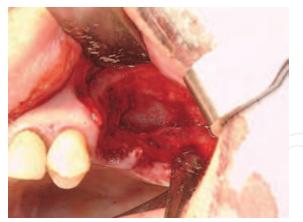
Fig. 2. Computed tomography (CT) image around the implants (arrow).

2.2 Diagnosis and treatment

It is well-known that curative chemotherapy or long-term adrenocortical steroid administration for lymphoblastic leukemia and Hodgkin and non-Hodgkin lymphomas induces osteonecrosis of the femoral head (Kubo et al. 2001; Bojko et al 2003; Karimova et al.

2008). Many anticancer drugs were administered for breast cancer metastasis in this patient, but no osteonecrosis of the jaw caused by these drugs has been reported. In addition, the clinical symptoms of this patient met all diagnostic criteria of BRONJ proposed by AAOMS (AAOMS 2007; Ruggiero et al. 2009). Based on these findings, BP-induced osteonecrosis was diagnosed.

In the treatment, morbid bone tissue including implants was resected in consideration of the following 3 conditions: 1) early chemotherapy for metastatic cancer was necessary, for which removal of the infected lesion was necessary, 2) there was a possibility that continuation of the withdrawal of BPs would aggravate metastatic cancer-associated bone destruction and hypercalcemia, and 3) it was difficult to remit inflammation by employing conservative treatment proposed by the American Association of Oral and Maxillofacial Surgeons (AAOMS 2007; Ruggiero et al. 2009). After considering these points, sequestrectomy of the left maxilla was selected. The gingival margin was incised between the upper left first premolar and implant towards the maxillary tuberosity, and the lesion area was exposed by forming a buccal and palatal mucoperiosteal flap. The bone cortex of the lesion was brownish black, and necrotic bone was confirmed between the distal surface of the second premolar and mesial surface of the implant placed in the second molar area, but particularly around the implant placed in the first molar area (Fig.3a). The area for resection around the living bone in the lesion was determined after the extraction of the second premolar, and the pathological bone tissue including the implant was removed with a Lindeman bar and bone chisel, followed by the grinding of the bone surface of the resection stump until bleeding was confirmed. As a result, the instrument partially perforated the maxillary sinus floor, and the thickened maxillary sinus mucosa was exposed. This sinus mucosa was spared, and the surgical wound was completely closed by suturing the mucoperiosteal flaps (Fig.3b)



(a) Pathological bone including the implants was removed, and maxillary sinus mucosa was exposed.



(b) The surgical wound was completely closed by suturing the mucoperiosteal flap.

Fig. 3. Intraoperative view of the sequestomy of the left maxilla.

After surgery, antibiotic was intravenously administered for 3 days and oral antibiotic was administered for 3 days. In addition, hyperbaric oxygen therapy was performed. The efficacy of hyperbaric oxygen therapy for BRONJ is controversial (Ruggiero et al. 2004; Freiberger et al 2009), but, at least, it may be effective to promote mucoepithelial wound healing in the

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sequestration-resected region. The wound was completely covered with healthy mucosa 2 weeks after surgery (Fig.4.). Since the remission of inflammation could be confirmed, anticancer drug and BP administration was initiated for metastatic cancer and bone metastasis-associated bone destruction. The patient died of multiorgan failure associated with multiple cancer metastasis 8 months after surgery, but BRONJ did not recur before death.



Fig. 4. Intraoral view at 4 weeks after surgery.

2.3 Macroscopic and μ CT findings of the resected specimen

Implants were strongly fixed to the bone, and areas of brown sequestration were observed around them (Fig.5.). On μ CT, no apparent abnormality, such as trabecular destruction, was observed, and the implant surface was in direct contact with the trabecula at various sites (Fig.6.).



Fig. 5. The resected specimen shows a portion of the left maxilla including two implants.

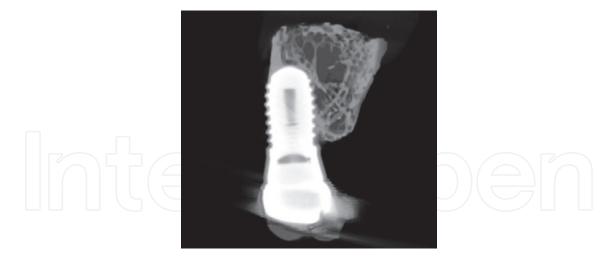


Fig. 6. μ CT findings of the resected specimen.

2.4 Histopathological analysis 2.4.1 Histologic preparation

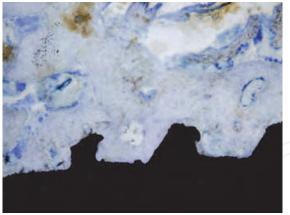
The resected specimen was divided into the lesion including the implants and peri-lesion, i.e., healthy tissue in the resected stump. The bone tissue around the implants was fixed, dehydrated, and embedded in polyester resin (Ligorac®, Wako Pure Chemical Industries, Ltd., Tokyo, Japan). Undecalcified ground sections 15 µm in thickness were cut along the long axis of the implant using the EXAKT Cutting-Grinding System (EXAKT, Norderstedt, Germany) and stained with 1% toluidine blue (pH 7.2), then the bone tissue around the lesion was dehydrated and embedded in paraffin according to the conventional procedure. Some of the sections were stained with hematoxylin-eosin (HE) and AZAAN. Other sections were processed for immunohistochemical analysis by immunostaining with anti-human CD3, CD20, CD45 and CD68 (Daco, Carpinteria, CA).

2.4.2 Peri-implant histopathological findings in undecalcified ground sections

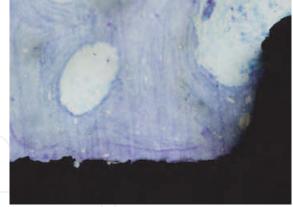
The implant surface was mostly in direct contact with bone tissue, suggesting that osseointegration was maintained until BRONJ development (Fig.7a). No osteocyte was present in the cavity of bone contacting implants or the surrounding trabeculae, nor were there cellular components on the bone surface, and bone tissue mostly showed sequestration (Fig.7b). The marrow cavity was filled with aggregates of actinomycete-like microorganisms, neutrophils, and other necrotized soft tissue (Fig.8a). In addition, adsorption cavities with a worm-eaten appearance were widely present. Normally, bone tissue necrotized in the body is absorbed by osteoclasts, shows sequestration, and is naturally excreted. However, no osteoclasts were present in the resorption cavity, suggesting that active resorption of the bone occurred in the BRONJ developmental process, rather than the resorption of necrotized bone by osteoclasts (Fig.8b).

2.4.3 Histopathological findings around lesions in decalcified sections

Osteocytes were present in the bone cavity (Fig.9a,b). The trabecular surface was smooth, and osteoid formation and osteoblast arrangement were observed. Reactive bone formation was also observed at various trabecular sites. Granulation tissue rich in capillary blood

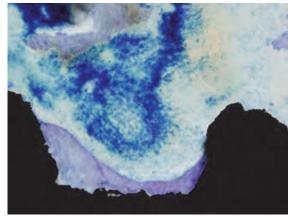


(a) Most of the implant surfaces are in direct contact with the bone (× 40).

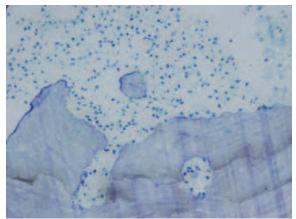


(b) The bone in contact with the implant sureface is necrotic (× 200).

Fig. 7. Histological findings of the tissue from the implants in an undecalcofied sections stained with toluidine blue.



(a) Growth of a bacterial colony into the marrow cavity is observed (× 200).



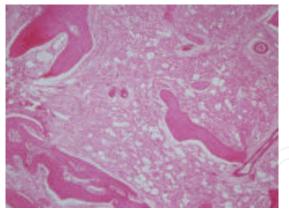
(b) Marked bone destruction with a moth eaten appearance is observed on the necrotic trabecular bone surface (× 200).

Fig. 8. Histological findings of the tissue from the implants in an undecalcofied sections stained with toluidine blue.

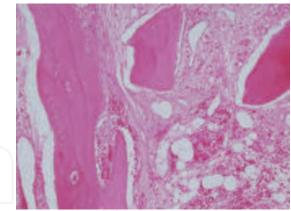
vessels invaded the bone marrow region (Fig.10a,b). On immunohistological staining, lymphocytes infiltrating the bone marrow were mostly CD3-positive, showing that the cells were derived from T cells. In contrast, only a few lymphocytes were positive for CD20 and CD45 (Fig.11a,b), but they were considered not to be significant. Multinucleated giant cells strongly positive for CD68, assumed to be osteoclasts, were present at various sites on the trabecular surface (Fig.12a,b).

3. Discussion on the peri-implant BRONJ developmental mechanism

No viewpoint regarding the pathogenesis of BRONJ has been established. Based on clinical experience, mucosal injuries caused by poorly fitting dentures and oral surgical treatment, such as that for periodontal disease and tooth extraction, have been reported as local risk factors, but this patient received no surgical dental treatment during the BP administration



(a) The trabecular bone surface are smooth (HE stain; × 40)



b) Growth of fibrous connective tissue and vascular formation are observed in the marrow(HE stain, × 100).

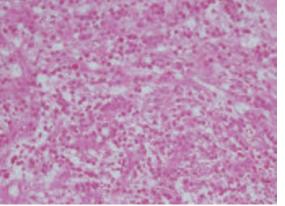
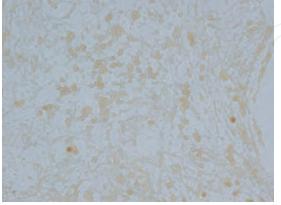


Fig. 9. Histological findings of the tissue around the lesion in a decalcified thin section.

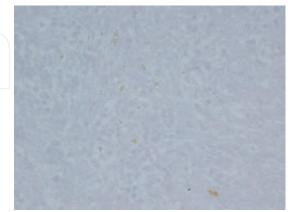
(a) Numerous lymphoid cells are observed in the marrow (HE stain; × 200).

(b) Lining cells and osteoid formation are observed (AZAN; × 100).

Fig. 10. Histological findings of the tissue around the lesion in a decalcified thin section.



(a) Many lymphoid cells are positive for anti-human CD3 (× 200).

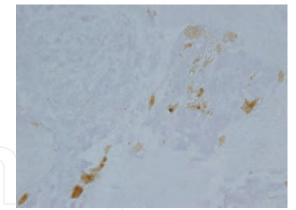


(b) Only a few lymphoctyes are positive for CD20 (× 200).

Fig. 11. Immunohistological staining of hte tissue around the lesion in a decalcified thin section.



(a) Only a few lymphoctyes are positive for CD45 (× 200).



(b) Multinuclear cells are strongly positive for anti-human CD68 antibody (× 200).

Fig. 12. Immunohistological staining of the tissue around the lesion in a decalcified thin section.

period, and no implant mobility was observed in the BRONJ site. Histologically, oseointegration was widely maintained on the implant surface, and no deep overgrowth of gingival epithelium was noted, suggesting that BRONJ in this patient suddenly occurred around favorably functioning implants, and was not induced by peri-implant inflammation. Necrosis of the femoral head (Kerachian et al. 2006) and irradiation-induced osteonecrosis of the jaw (Marx 1983) are widely known as osteonecrotic lesions, and the cause is insufficiency of vascular formation in both diseases. Reportedly, zoledronate exhibits a potent vascularization-inhibitory effect (Giraudo et al. 2004), and the BP-induced obstruction of blood vessels in bone and necrosis of osteocytes around them have been reported (Hansen et al. 2007). Based on these findings, it is hypothesized that insufficiency of the blood supply is also involved in BRONJ, but, histologically, the absence of vascularization inhibition in BRONJ has been reported (Bedogni et al. 2008). Another important hypothesis suggests an influence of reduced bone remodeling due to the BP-induced inhibition of bone resorption by osteoclasts. It has been reported that osteocyte mortality increases when bone remodeling declines, i.e., BPs reduce bone remodeling, increasing osteocyte death, which is involved in BRONJ (Allen et al. 2009). However, many resorption cavities were present on the periimplant-sequestered bone surface, and the growth of granulation tissue containing abundant capillary blood vessels and reactive bone formation in the resected stump were confirmed in this patient. Thus, it is difficult to explain the cause of BRONJ with insufficient blood supply and the bone remodeling-inhibitory effect of BPs alone.

Pamidronate and zoledronate administered intravenously to this patient are BPs that incorporate nitrogen. BPs containing nitrogen (NBPs) induce an inflammatory reaction by stimulating cells (Hewit et al. 2005; Endo et al. 1993; Nakamura et al. 1996). NBPs also exist on the surface of gram-negative cell walls, and increase inflammatory reactions to lipopolysaccharide (LPS), which has a strong inflammatory effect (Yamaguchi et al. 2000). Therefore, BRONJ may be caused by interactions among several factors differing among cases, not by a single cause. For example, in this patient, the following conditions may have influenced each other and caused BRONJ: 1) the excessive inhibition of bone turnover due to the NBP-related inhibition of bone resorption; 2) the occlusal force loaded on implants was directly transmitted to the bone, which activated local bone metabolism in the surrounding

region, releasing BP accumulated in the bone at a high level; 3) free NBP-induced inflammatory reactions, induction of osteoclast apoptosis (Roelofs et al. 2006), and inhibition of epithelial activity (Landesberg et al. 2009); 4) increased risk of bacterial infection due to suppression of the immune system; 5) enhanced inflammatory reactions due to interactions between gram-negative bacterial lipopolysaccharides and NBPs and 6) reduced local blood flow by vascularization inhibition and vascular obstruction (Fig.13).

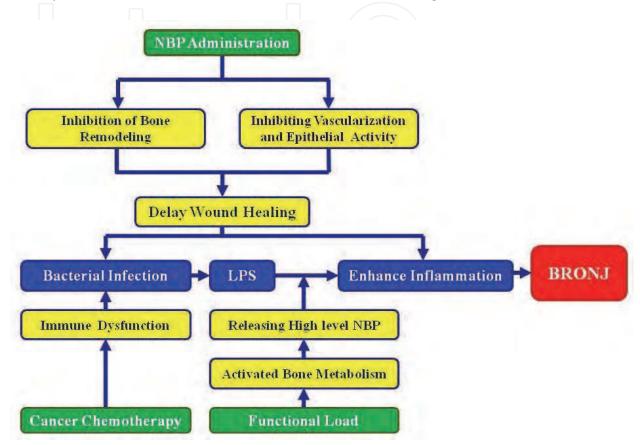


Fig. 13. Hypothetical mechanism of BRONJ aroud the implants.

4. Conclusion

BRONJ is intractable, and no treatment method has been established. Only guidelines mainly concerning prevention and conservative treatment have been proposed. When periimplant BRONJ occurs, firstly, conservative treatment should be selected following the AAOMS treatment guidelines, placing great importance on the prevention of exacerbation of BRONJ, but when NBPs are administered to control bone metastasis, it should be a priority to re-start BP administration and prevent any delay in cancer chemotherapy. Accordingly, although the appropriateness and timing of sequestration site resection should be carefully investigated, active surgical treatment may be one choice for some patients. Implant treatment is no longer special treatment. It is widely introduced into routine dental practice as a method to recover the occlusal function of defective teeth. Therefore, NBP intravenous injections will be increasingly administered to patients with a past medical history of implant treatment, for which the establishment of a practical clinical policy for the prevention and treatment of BRONJ is strongly desired.

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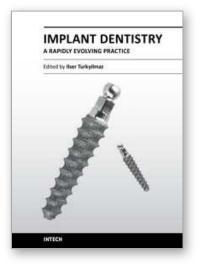
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Implant Dentistry - A Rapidly Evolving Practice

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Implant dentistry has come a long way since Dr. Branemark introduced the osseointegration concept with endosseous implants. The use of dental implants has increased exponentially in the last three decades. As implant treatment became more predictable, the benefits of therapy became evident. The demand for dental implants has fueled a rapid expansion of the market. Presently, general dentists and a variety of specialists offer implants as a solution to partial and complete edentulism. Implant dentistry continues to evolve and expand with the development of new surgical and prosthodontic techniques. The aim of Implant Dentistry - A Rapidly Evolving Practice, is to provide a comtemporary clinic resource for dentists who want to replace missing teeth with dental implants. It is a text that relates one chapter to every other chapter and integrates common threads among science, clinical experience and future concepts. This book consists of 23 chapters divided into five sections. We believe that, Implant Dentistry: A Rapidly Evolving Practice, will be a valuable source for dental students, post-graduate residents, general dentists and specialists who want to know more about dental implants.

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