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# Mechanisms of Tolerance: Role of the Thymus and Persistence of Antigen in Calcineurin-Induced Tolerance of Renal Allografts in MGH Miniature Swine

Joseph R. Scalea, Isabel Hanekamp and Kazuhiko Yamada\*  
*Transplantation Biology Research Center, Massachusetts General Hospital,  
Harvard Medical School,  
USA*

*(\*Address any correspondence to Kazuhiko Yamada)*

## 1. Introduction

The induction of donor-specific tolerance remains a major goal of clinical transplantation. Partially inbred MGH miniature swine, in which swine leukocyte antigens (SLA) have been defined and fixed, have been utilized extensively as a preclinical model for tolerance induction. This preclinical large-animal model is an invaluable tool for studying the mechanism of transplantation tolerance. Recently, we have investigated the role of the persistence of donor antigen in the maintenance of tolerance and the peripheral regulatory cells' ability to confer tolerance to naïve animals.

Mechanisms of tolerance can be elucidated in part by attempting to interfere with (or "break") the tolerant state. We have previously reported that a short course of calcineurin inhibition permits the uniform development of long-term, donor-specific tolerance to renal allografts in juvenile miniature swine. Once tolerant, animals that undergo graft nephrectomy and immediate retransplantation accept donor-MHC matched kidneys while maintaining stable renal function without further immunosuppression.

Utilizing this model we have attempted to break tolerance using several strategies. These have included administration of recombinant IL-2 to provide additional T-cell help, manipulation of the host thymus, and removal of donor grafts. Our data indicate that (1) presence of an intact thymus is essential for the induction, but not for the maintenance of tolerance; (2) the persistence of the donor renal graft is essential for the indefinite continuation of tolerance; and (3) the pathway of donor antigen presentation, in tolerant and previously tolerant animals, is important for preservation or loss of the tolerant state. Although obvious limitations exist with regard to animal studies, our consistently reproducible results using MHC inbred miniature swine provide a unique opportunity to study the mechanisms of transplantation in animals physiologically similar to humans. As we have previously shown, our results are highly suggestive of what will occur in clinical human transplantation protocols.

## 2. Abbreviations

APC: Antigen Presenting Cell  
CTL: Cytotoxic T-Lymphocytes  
CyA: Cyclosporine A  
FoxP3+: Forkhead Box 3  
HSC: Hematopoietic Stem Cells  
IL-2: Interleukin 2  
MGH: Massachusetts General Hospital  
MHC: Major Histocompatibility Complex  
PBL: Peripheral Blood Lymphocytes  
SLA: swine leukocyte antigens  
UNOS: United Network for Organ Sharing

## 3. Importance of tolerance

According to UNOS, over 110,000 patients are currently awaiting organ transplantation in the United States. However, in 2010 there were only 14,506 donors. Although the benefits of valued organ transplants are substantial, patients must accept the risks of the required immunosuppression. Immunosuppressive protocols generally include T-cell depletion in the perioperative period, followed by initiation of calcineurin inhibition, mycophenolic acid, and steroids. Many centers have been successful in reducing or eliminating the use of chronic steroids, but patients are still susceptible to the morbidity associated with immunosuppression (1,2).

Transplantation tolerance, or immunologic non-responsiveness to donor antigen, would reduce the morbidity observed with immunosuppression (3). Immunosuppression has been associated with malignancy, infection, end-organ damage, and economic burden. As many as 30% of renal transplant recipients develop skin cancer within 10 years of transplantation (4) similar results have been reported in the cardiac transplant patient population (4,5). Furthermore, viral and bacterial infections are more likely to occur in the immunocompromised patient and although death rates from infection have decreased every decade for the last 30 years, patients at the extremes of age are still considered high-risk for infectious complications (6). End-organ failure is also a potential complication of immunosuppression; renal failure, likely due to calcineurin inhibitors, has been associated with chronic immunosuppression (7,8). Additionally, the lifetime economic cost of immunosuppressive drugs alone following kidney transplantation range from \$68,000 to \$88,000 depending on the selected regimen (9,10).

Tolerance strategies would potentially alleviate the risks of malignancy, organ-failure, and infectious complications as well as the cost associated with anti-rejection medications. Recent clinical successes in inducing tolerance to kidney (11,12) and liver (13) grafts have proven that tolerance strategies are clinically applicable.

## 4. Antigen presentation of allogeneic antigens and tolerance

Tissue antigens are any proteins which when presented by the major histocompatibility complex lead to immunologic responses (14,15). Proof of this MHC restriction earned Zinkernagel and Doherty the Nobel Prize in 1996 for work that was done in the 1970s (15).

In transplantation, these antigens can be presented by either donor or recipient antigen presenting cells (APCs). The presentation of antigen to recipient lymphocytes potentially leads to allosensitization of recipient T-cells. In direct alloantigen presentation, donor tissue antigens are recognized by recipient T-cells in the context of donor MHC molecules. Conversely, indirect antigen presentation occurs when alloantigens are recognized in the setting of recipient MHC. Recently, one group described an additional "semi-direct" mechanism in which the recipient cells acquire the intact MHC from the donor and subsequently present antigen in the context of the newly acquired donor MHC (16).

A comprehensive understanding of antigen presentation is helpful for elucidating both transplantation rejection and tolerance. In allotransplantation, passenger lymphocytes and professional APCs within the transplanted organ present donor antigen to recipient T-cells leading to acute rejection (17,18). Indirect presentation of donor antigen may continue indefinitely and is generally associated with chronic rejection (19). These two pathways, however, are not mutually exclusive as early rejection episodes portend increased risk of future chronic rejection (20). While these alloreactive stimuli are known to play a role in rejection, small and large animal studies of tolerance have also demonstrated the importance of donor antigen presentation in the establishment of a tolerogenic milieu (21-23).

Mechanisms of transplantation tolerance have been broadly categorized as "central" or "peripheral" on the basis of whether T-cells are rendered unresponsive during their maturation in the thymus or after they have left the thymus, respectively (24-26). Deletion is thought to be the mechanism responsible for central tolerance, whereas peripheral tolerance is likely mediated by anergy, suppression, or ignorance (27). Central tolerance can be induced by exposing newly developed T-cells to alloantigens on the progeny of hematopoietic stem cells (HSC) injected either at a very early stage in the development of the immune system, either in utero or in neonates (28,29) or in adult animals following ablation of mature T cells (30-32). This tolerance is deletional and presumably utilizes the same process of negative selection that is responsible for self-tolerance during T-cell maturation in the thymus (29,33). In addition to bone marrow transplantation, another strategy to induce central tolerance is thymic transplantation. We, and others, have demonstrated successful induction of tolerance with donor thymic grafts across allogeneic and xenogeneic barriers in small and large animal models (34-36).

Peripheral tolerance to alloantigens has been induced in many ways, generally by providing alloantigen in a non-stimulatory fashion or at a time when the aggressive alloreactive response has been simultaneously averted. Peripheral tolerance has been ascribed to the same processes as those invoked to explain peripheral tolerance to self - i.e. anergy(37-41), peripheral deletion(42,43), clonal ignorance (32) and regulation (44-49).

Investigation of peripheral cellular suppression began over 40 years ago, though the implications of these findings were not immediately evident. Tada et al. demonstrated that when KLH primed T-lymphocytes were passively transferred to mice prior to immunization with DNP-KLH, antibody formation was inhibited. In contrast, when these cells were transferred after immunization with DNP-KLH, antibody formation was not inhibited. These results demonstrated that a peripheral T-cell response was suppressing immunologic response to immunization (50,51). This phenomenon was studied extensively in the 1970s and 1980s, and Castagnoli et al. found that the supernatant from antigen-specific suppressive thymoma cells was capable of suppressing an alloreactive response to the same antigen. These and other data suggested that the effects of purported T suppressor cells are mediated by one or more soluble factors (52,53). Our understanding of these suppressive

cells was broadened further when Sakaguchi et al. demonstrated that the lack of ostensible suppressor cells in nude/nude mice led to severe autoimmune disease, which could be rectified by inoculation with nude/+ thymocyte suspensions. Sakaguchi and his colleagues then successfully phenotyped these suppressor/regulatory cells as CD4<sup>+</sup>, CD25<sup>hi</sup> (and some years later, FoxP3+) and showed that these cells are likely responsible for tolerance of self, and possibly necessary for the maintenance of peripheral tolerance (47,54).

## 5. Importance of large animal models

While small animal data are valuable for defining mechanisms, large animal studies are imperative for the development of preclinical models (55). Numerous strategies for tolerance induction in rodents have proven fruitful, though few have been successfully translated to humans, non-human primates, or other large animals (55,56). Likely owing to the increased complexity of the immune system in large animals and longer-term exposure to environmental antigens, the results from tolerance induction protocols in large animals have been less successful than rodent studies (55-57). Another possible cause for this difference is that MHC class II expression on endothelial cells that is the first target in the alloreactive response to in solid organs. Murine “resting” endothelial cells do not express MHC class II while swine, whereas primate endothelial cells do (58).

## 6. MHC inbred MGH miniature swine: A unique large animal model to study mechanisms of acceptance/rejection

Miniature swine have been developed in our laboratory over the past thirty years as a model system for studies of transplantation biology. Swine were chosen for this purpose because they represent one of the few large animal species in which breeding characteristics make genetic experiments possible. Swine have a relatively large litter size (3-10 offspring) and a short gestational cycle (3 months). They reach sexual maturity at approximately 6 months of age, and sows have an estrous cycle every 3 weeks. These breeding characteristics have made it possible to develop MHC homozygous lines of miniature swine in a relatively short time and have also made it possible to isolate new MHC recombinants, to breed them to homozygosity, and to carry out short-term backcross experiments in order to identify and study the segregation of genetic characteristics (59). Our miniature swine thus represent the only large animal model in which MHC genetics can be reproducibly controlled. As such, these animals have been particularly useful in assessing the effects of MHC matching on rejection and/or tolerance induction (60,61).

At present, we maintain swine of three homozygous SLA haplotypes, SLA<sup>a</sup>, SLA<sup>c</sup>, SLA<sup>d</sup> and five lines bearing intra-SLA recombinant haplotypes as illustrated in Fig. 1. All of these lines differ by minor histocompatibility loci, thus providing a model in which most of the transplantation combinations relevant to human transplantation can be mimicked. Thus, for example, transplants within an MHC homozygous herd simulate transplants between HLA identical siblings, while transplants between herds resemble cadaveric or non-matched sibling transplants. Likewise, transplants between pairs of heterozygotes can be chosen to resemble parent into offspring or one-haplotype mismatched sibling transplants (22). In addition, we have chosen one subline of our SLA<sup>dd</sup> animals for further inbreeding, in order to produce a fully inbred line of miniature swine. This subline reached a coefficient of inbreeding of >96%, leading for the first time to long-term acceptance of reciprocal skin grafts (62). These animals

have also made it possible to carry out adoptive transfer experiments for the first time in a large animal model, as reported here (Scalea et al, manuscript in preparation).

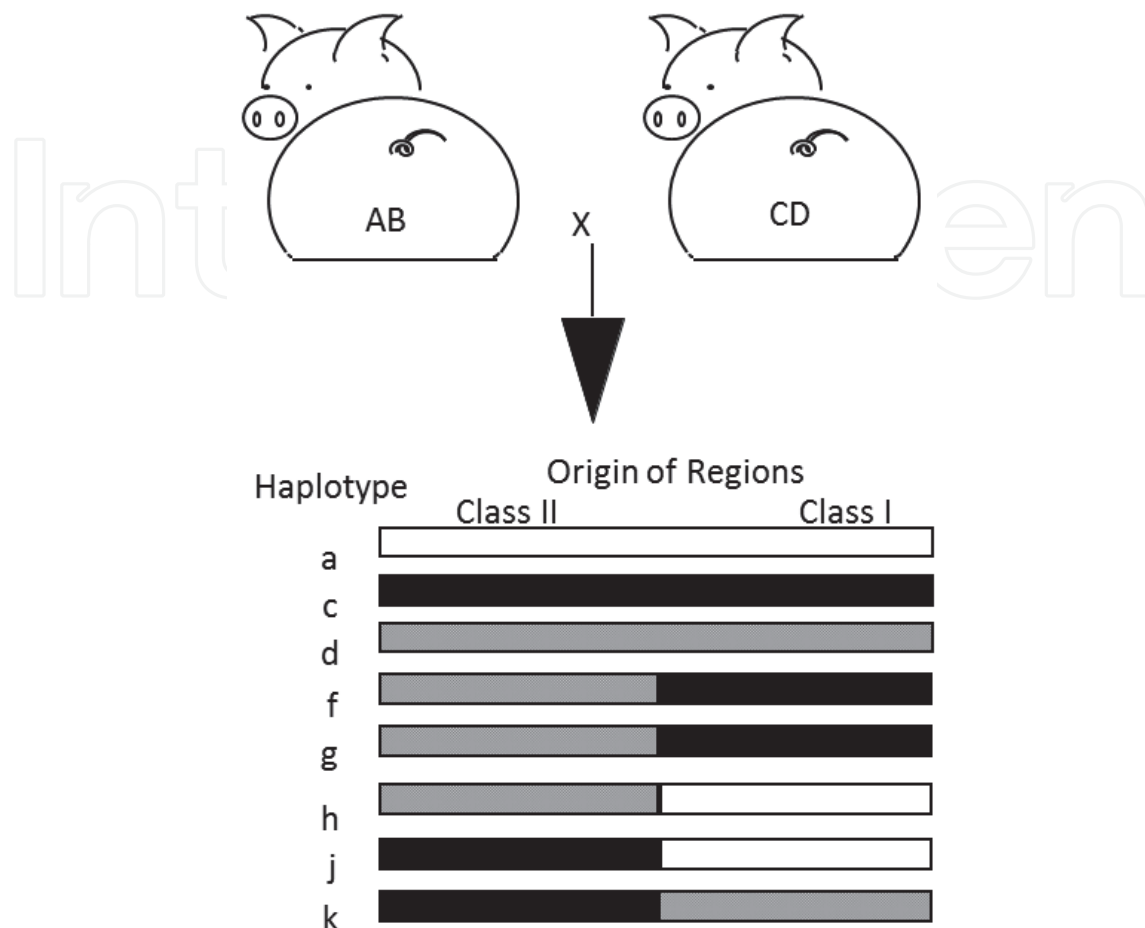


Fig. 1. The serial breeding of MGH swine has led to fixed, defined MHC classes. Transplantation between lines of these swine allows researchers to mimic living-related and cadaveric organ transplantation.

## 7. Thymic-dependent and antigen-dependent tolerance in MHC miniature swine renal transplant models

### 7.1 Induction of tolerance with a short course and high dose calcineurin inhibitor in MGH miniature swine

- i. **Effects of CyA on renal allograft survival across a selective MHC barrier:** We have attempted to induce tolerance using pharmacological limitation of T cell help. For this purpose, class I mismatched renal transplants were performed using a short course of treatment with Cyclosporin A (CyA) (63,64). We chose to study the effect of CyA across a selective *two-haplotype class I mismatched, class II matched* barrier, since without immunosuppression such recipients uniformly reject renal allografts within three weeks without immunosuppression (Fig. 2). As reported in our initial study of this treatment regimen, a twelve-day course of CyA (10-13 mg/kg/day) induced long-term, specific tolerance in eight of eight *two-haplotype class II matched, class I mismatched* recipients(64). This result has been reproduced subsequently in more than fifty additional CyA-treated



animals, 100% of which develop long-term tolerance across a class I disparity. It is important to note that although this dose and the resulting blood levels (400-800 ng/dl) are high with respect to clinically acceptable values, the toxicity caused by such levels clinically is generally reversible if discontinued after a two-week course (65).

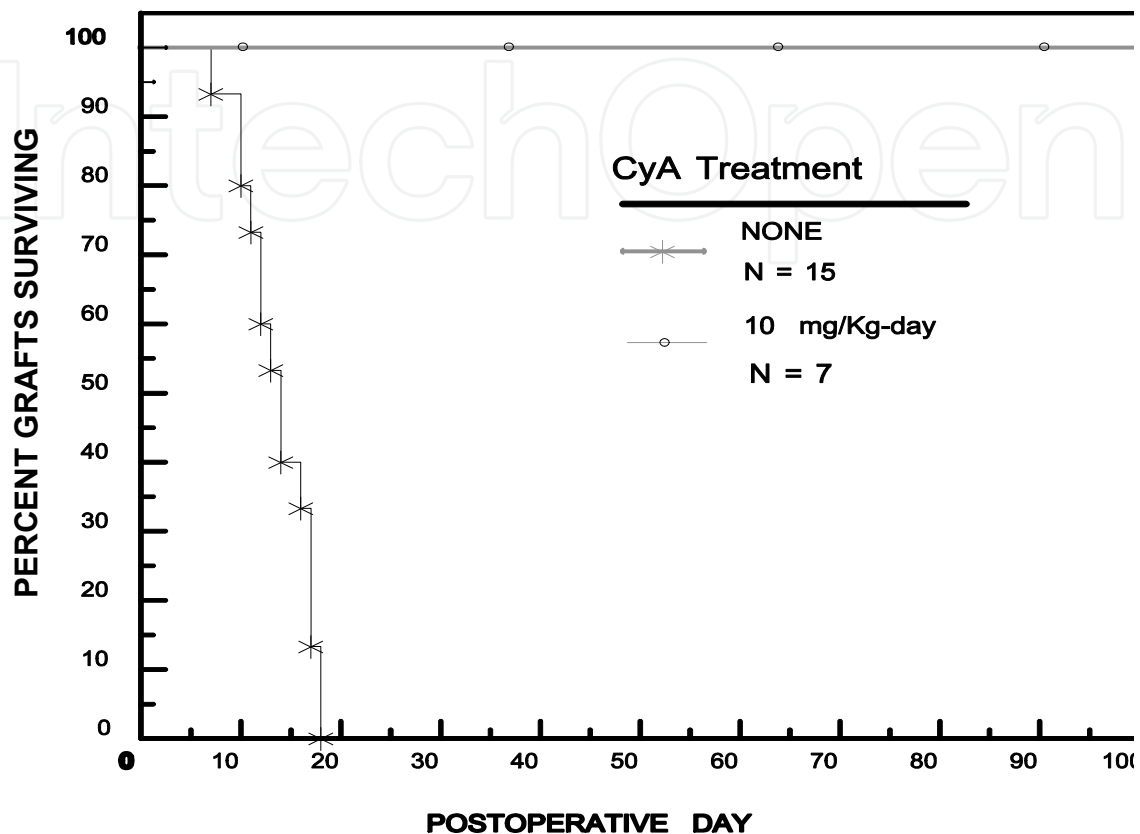


Fig. 2. With a short-course of high-dose Cyclosporine A, tolerance is uniformly established across an MHC class-I mismatch in MGH miniature swine ( $n > 50$ ).

- ii. **Effects of CyA on renal allograft survival across other selective MHC barriers:** We have also studied whether tolerance is induced with a 12-day course of CyA therapy across further immunologic disparities. Although the CyA regimen was capable of prolonging renal allograft survival across a full MHC barrier, it did not induce long-term tolerance in any of the animals tested (64). However, since pharmacologic help limitation by CyA should be possible regardless of the MHC disparity, we also tested the effect of the CyA regimen on renal transplants selectively mismatched for class II or for one full haplotype (66,67). Of seven CyA-treated class I matched, class II mismatched kidney transplants, five (71%) were accepted long-term, whereas two rejected on days 20 and 39, respectively (68). Similarly, of six CyA-treated recipients of single haplotype mismatched kidney allografts, two rejected (on days 31 and 37), while four (67%) accepted long-term. Thus, tolerance was possible across both class II and single haplotype full mismatches, but with a lower success rate and with a less stable clinical course than across a selective class I mismatch.
- iii. **Effects of Tacrolimus on renal allograft survival across other selective MHC barriers** According to the latter hypothesis, increasingly potent immunosuppression of T-cell help would be required for increasing the extent of the MHC disparity (i.e. class I < class II < one full haplotype < two full haplotypes). Our subsequent results using

Tacrolimus corroborate this finding (69). We have demonstrated that Tacrolimus at blood levels between 35 and 80ng/ml facilitates tolerance induction not only across a class I barrier, but also across a two-haplotype full MHC barrier in-vivo (69). However, like CyA-induced tolerance across class I mismatched barrier, thymic-dependent mechanisms are involved in this Tacrolimus-induced tolerance (70). Tolerance was only induced in juvenile hosts (see more details below).

## 7.2 Mechanisms of tolerance using high-dose calcineurin inhibitors

To understand better the mechanisms of tolerance induction, both the role of thymus (i.e. aging) and persistence of donor antigens in tolerance induction/maintenance have been studied.

- i. *Thymic dependent tolerance:* A series of experiments were performed using aged and thymectomized animals. The data from these studies demonstrated that when aged animals underwent class-I mismatched kidney transplantation followed by 12-days of CyA, tolerance could not be induced. Similarly, thymectomy prior to kidney transplantation interfered with the development of tolerance across the same class-I barrier (71). Interestingly, thymectomy in a maintenance period (beyond 6 weeks after transplantation) did not abrogate tolerance (70). Recent work from this laboratory has also demonstrated thymic dependent mechanisms play important role in tacrolimus induced tolerance in both miniature swine and nonhuman primates (Yamada et al manuscript in preparation). These studies indicated that that 1) tolerance induction is thymus-dependent, whereas tolerance-maintenance is not and 2) an interaction between central and peripheral mechanisms of tolerance is likely occurring in the stably tolerant animal.
- ii. *Regulatory mechanisms and stability of tolerance – Class I mismatched kidney transplantation with a 12-day course of CyA:*

- a. In vitro assays suggesting involvement of regulatory mechanisms

Because thymectomy post-transplantation did not lead to the abrogation of tolerance, investigators postulated that a suppressive peripheral regulatory mechanism had developed in class I mismatched tolerant model. When peripheral blood lymphocytes (PBL) from long-term tolerant animals were stimulated with donor PBLs, and re-cultured with naïve SLA matched PBLs plus either donor-PBLs or 3<sup>rd</sup> party PBLs, we observed suppression of the naïve-SLA anti-donor CTL response and maintenance of naïve-SLA anti-3<sup>rd</sup> party CTL responses (45). This co-culture assay demonstrated in-vitro that a peripheral cellular mechanism of suppression was present. To study activation and effector function of these purported regulatory cells, we performed co-culture assays in which PBLs taken from a tolerant animal were placed in a transwell culture system such that they were separated from donor PBLs by a permeable membrane (69). In previous co-culture CTL assays performed without the membrane barrier, we observed that naïve-recipient SLA matched PBLs were inhibited by the presence of PBLs taken from a tolerant animal. However, when the cells were separated by a permeable membrane, the cells were no longer suppressive (69,72). This demonstrated that 1) direct cell-to-cell contact is required for activation of the peripheral regulatory mechanism and 2) soluble factors produced by the peripheral regulatory cells are themselves incapable of regulatory effector activation in this class I mismatched kidney model.

- b. Exogenous T-cell help interfered with the induction of tolerance but not maintenance of tolerance

Based on these in-vivo and in-vitro data, we attempted to further define the role of regulatory cells in the MGH miniature swine model, by attempting to abrogate tolerance in



animals. Because a lack of T cell help likely plays an essential role in calcineurin-induced tolerance, we administered exogenous IL-2 either 1) during the induction period (Days 8, 9 and 10) or 2) once animals were long-term tolerant (73). Much like our thymectomy experiments, we found that whereas IL-2 administration can prohibit the induction of tolerance if administered perioperatively, treatment with exogenous IL-2 failed to abrogate tolerance in long-term tolerant (LTT) animals (73). To further distill the role of T-cell help required for the anti-donor cellular response in tolerant animals, we challenged LTT animals with skin grafts from class-I donor/class-II third party donors instead of IL-2 (74). We reasoned that the class-II disparate graft may be capable of providing the necessary T-cell help by stimulating the alloreactive CD4<sup>+</sup> population. Although recipient animals experienced a brief rejection crisis following skin grafting, they remained tolerant in the long-term (74). Thus, once established, the peripheral mechanism of tolerance is steadily stable, and capable of suppressing further stimulation with donor antigen.

c. Role of graft in maintenance of tolerance

Because 1) removal of the thymus in a tolerant animal did not lead to tolerance abrogation and 2) based on the in-vitro data suggesting that tolerance was mediated by an active cellular process, we next questioned if the graft itself was providing the tolerogenic stimulus for maintenance of tolerance. To test this hypothesis, we designed several experiments in which the tolerated graft was removed and replaced by a donor-MHC matched graft (21) (75). In the first experiment, long-term tolerant animals underwent graft nephrectomy and immediate retransplantation with a donor-MHC matched graft. As previously published, each animal uniformly accepted the retransplanted graft and never experienced rejection. In the next experiments, we introduced a period of “absence-of-donor-antigen” by removing the tolerated kidney from LTT animals and replacing it with a self-MHC matched graft to support the life of the animal. Then, at 1 and 3 months, animals were retransplanted with a donor-MHC matched graft. When animals bearing self-MHC matched grafts underwent retransplantation from actual donors immediately after primary graft nephrectomy, all animals accepted kidney with stable renal function (21). However, we observed a brief rejection crisis followed by uniform acceptance when second kidneys were transplanted at one month after the primary graft nephrectomy. Moreover, as the period of absence-of-donor-antigen was increased to 3 months, retransplantation was followed by significant rejection crisis in two of three animals. One animal completely rejected the retransplanted graft within 2 months and the other had severe rejection episodes (21). Furthermore, when skin grafts from class-I donor/class-II third party donors were transplanted onto animals during absence of donor kidneys, second kidneys transplanted 3 months after primary kidney nephrectomy were uniformly rejected in an accelerated manner (<7 days), indicating a “broken tolerance” (Yamada et al., manuscript in preparation). This series of experiments confirmed that the kidney plays an essential role in the maintenance of tolerance. Because we observed rejection of the retransplanted donor-MHC matched graft following periods without donor antigen present, it is likely that the kidney provides a constant tolerogenic stimulus via a peripheral mechanism which is required for maintenance of T-regulatory cells.

d. Current evidence for the role of T-regulatory cells in peripheral tolerance

The previous experiment clarified our understanding of the role that persistence of donor antigen plays in the maintenance of tolerance and provided indirect evidence that this process was mediated by T-regulatory cells (21,45). We then postulated that, if T-regulatory cells were responsible for maintaining tolerance then removal of T-regulatory cells from a LTT animal, may lead to abrogation. Thus, we next attempted to hasten the onset of

rejection by performing an extensive leukapheresis immediately prior to retransplantation of a donor-matched graft following a three month period of absence-of-donor-antigen. In eight of ten animals in this protocol, we observed rejection of a subsequently retransplanted donor-MHC matched graft, and complete or chronic rejection in four of ten animals (Scalea et al, manuscript being submitted). When the leukapheresed cells were evaluated in vitro, we observed that they were capable of suppressing a naïve anti-donor response in a cell mediated lympholysis (CTL) co-culture assay in a donor-specific manner (Scalea et al, manuscript in preparation).

Having determined that these cells were capable of suppressing a naïve response in-vitro (45), we then questioned whether they would be suppressive in-vivo. However, based on our absence-of-donor-antigen and leukapheresis with retransplantation experiments, it appeared that both the kidney and peripheral regulatory cells were important for the continuation of tolerance (21). To resolve the contribution of the circulating cellular compartment versus the kidney itself, we adoptively transferred leukapheresed cells from tolerant donors that were from our most highly-inbred line of miniature swine. We found that when either the LTT kidney was transplanted into a naïve animal, or leukapheresed cells were transferred along with a naïve donor graft, there was prolonged graft survival, but not long-term acceptance (Scalea et al, manuscript in preparation). Conversely, when the leukapheresed cells and kidney were transferred from the same LTT animal, the naïve recipient experienced long-term graft survival. Additionally, these recipients were unresponsive to donor in CML assay as early as day 28, and in one case, as late as 150 days following transplantation (Scalea et al, manuscript being submitted).

## 8. Summary

In summary, our understanding of the mechanisms of tolerance has blossomed over the last several decades. It is clear that while small animal data has helped elucidate the molecular basis for tolerance, large animal studies are required for the development of preclinical models. While animal models have some limitations, our unique model allows us to model accurately the clinical scenarios of clinical transplantation. Using this model, we have demonstrated the importance of the thymus for tolerance induction and the persistence of donor antigen for the maintenance of tolerance. Recent work in our laboratory has demonstrated that stable peripheral tolerance is mediated by a cellular mechanism that is best explained by the presence of T-regulatory cells.

## 9. References

- [1] Yamani MH, Taylor DO, Czerr J et al. Thymoglobulin induction and steroid avoidance in cardiac transplantation: results of a prospective, randomized, controlled study. *Clin Transplant* 2008;22: 76-81.
- [2] Sarwal MM, Yorgin PD, Alexander S et al. Promising early outcomes with a novel, complete steroid avoidance immunosuppression protocol in pediatric renal transplantation. *Transplantation* 2001;72: 13-21.
- [3] Griesemer AD, Sorenson EC, Hardy MA. The role of the thymus in tolerance. *Transplantation* 2010;90: 465-474.
- [4] Carroll RP, Segundo DS, Hollowood K et al. Immune phenotype predicts risk for posttransplantation squamous cell carcinoma. *J Am Soc Nephrol* 2010;21: 713-722.

- [5] Brewer JD, Colegio OR, Phillips PK et al. Incidence of and risk factors for skin cancer after heart transplant. *Arch Dermatol* 2009;145: 1391-1396.
- [6] George JF, Pamboukian SV, Tallaj JA et al. Balancing rejection and infection with respect to age, race, and gender: clues acquired from 17 years of cardiac transplantation data. *J Heart Lung Transplant* 2010;29: 966-972.
- [7] Scalea JR, Butler CC, Munivenkatappa RB et al. Pancreas transplant alone as an independent risk factor for the development of renal failure: a retrospective study. *Transplantation* 2008;86: 1789-1794.
- [8] Ziolkowski J, Paczek L, Senatorski G et al. Renal function after liver transplantation: calcineurin inhibitor nephrotoxicity. *Transplant Proc* 2003;35: 2307-2309.
- [9] Earnshaw SR, Graham CN, Irish WD, Sato R, Schnitzler MA. Lifetime cost-effectiveness of calcineurin inhibitor withdrawal after de novo renal transplantation. *J Am Soc Nephrol* 2008;19: 1807-1816.
- [10] Yao G, Albon E, Adi Y et al. A systematic review and economic model of the clinical and cost-effectiveness of immunosuppressive therapy for renal transplantation in children. *Health Technol Assess* 2006;10: iii-xi, 1.
- [11] Kawai T, Cosimi AB, Spitzer TR et al. HLA-mismatched renal transplantation without maintenance immunosuppression. *N Engl J Med* 2008;358: 353-361.
- [12] Scandling JD, Busque S, Dejbakhsh-Jones S et al. Tolerance and chimerism after renal and hematopoietic-cell transplantation. *N Engl J Med* 2008;358: 362-368.
- [13] Alexander SI, Smith N, Hu M et al. Chimerism and tolerance in a recipient of a deceased-donor liver transplant. *N Engl J Med* 2008;358: 369-374.
- [14] Zinkernagel RM, Doherty PC. Restriction of in vitro T cell-mediated cytotoxicity in lymphocytic choriomeningitis within a syngeneic or semiallogeneic system. *Nature* 1974;248: 701-702.
- [15] Zinkernagel RM, Doherty PC. Immunological surveillance against altered self components by sensitised T lymphocytes in lymphocytic choriomeningitis. *Nature* 1974;251: 547-548.
- [16] Smyth LA, Herrera OB, Golshayan D, Lombardi G, Lechler RI. A novel pathway of antigen presentation by dendritic and endothelial cells: Implications for allorecognition and infectious diseases. *Transplantation* 2006;82: S15-S18.
- [17] Sayegh MH. Why do we reject a graft? Role of indirect allorecognition in graft rejection. *Kidney Int* 1999;56: 1967-1979.
- [18] Giangrande I, Yamada K, Germana S, Sachs DH, LeGuern C. Tolerant cells infiltrating class I mismatched swine kidney allografts lack the CD4 single positive subset and down regulate TCR gene expression. *Transplant Proc* 1997;29: 1164.
- [19] Kuo E, Maruyama T, Fernandez F, Mohanakumar T. Molecular mechanisms of chronic rejection following transplantation. *Immunol Res* 2005;32: 179-185.
- [20] Pascual M, Theruvath T, Kawai T, Tolkoff-Rubin N, Cosimi AB. Strategies to improve long-term outcomes after renal transplantation. *N Engl J Med* 2002;346: 580-590.
- [21] Okumi M, Fishbein JM, Griesemer AD et al. Role of persistence of antigen and indirect recognition in the maintenance of tolerance to renal allografts. *Transplantation* 2008;85: 270-280.
- [22] Rosengard BR, Ojikutu CA, Fishbein J, Kortz EO, Sachs DH. Selective breeding of miniature swine leads to an increased rate of acceptance of MHC-identical, but not of class-I disparate, renal allografts. *J Immunol* 1992;149: 1099-1103.

- [23] Griesemer AD, LaMattina JC, Okumi M et al. Linked suppression across an MHC-mismatched barrier in a miniature swine kidney transplantation model. *J Immunol* 2008;181: 4027-4036.
- [24] Wekerle T, Kurtz J, Bigenzahn S, Takeuchi Y, Sykes M. Mechanisms of transplant tolerance induction using costimulatory blockade. *Curr Opin Immunol* 2002;14: 592-600.
- [25] Sachs DH. Transplantation tolerance. *Transplant Proc* 1998;30: 1627-1629.
- [26] Sachs DH. Immunologic tolerance to organ transplants. *J Gastrointestinal Surgery* 1999;3: 105-110.
- [27] Sykes M, Sachs DH. Immunobiology of transplantation. In: Dulbecco R, ed. *Encyclopedia of Human Biology*. San Diego: Academic Press, Inc., 1991: 357-365.
- [28] Anderson D, Billingham RE, Lampkin GH, Medawar PB. The use of skin grafting to distinguish between monozygotic and dizygotic twins in cattle. *Heredity* 1951;5: 379.
- [29] Inaba M, Inaba K, Hosono M et al. Distinct mechanisms of neonatal tolerance induced by dendritic cells and thymic B cells. *J Exp Med* 1991;173: 549-559.
- [30] Rayfield LS, Brent L. Tolerance, immunocompetence, and secondary disease in fully allogeneic radiation chimeras. *Transplantation* 1983;36: 183-189.
- [31] Ildstad ST, Sachs DH. Reconstitution with syngeneic plus allogeneic or xenogeneic bone marrow leads to specific acceptance of allografts or xenografts. *Nature* 1984;307(5947): 168-170.
- [32] Tomita Y, Khan A, Sykes M. Role of intrathymic clonal deletion and peripheral anergy in transplantation tolerance induced by bone marrow transplantation in mice conditioned with a non-myeloablative regimen. *J Immunol* 1994;153: 1087-1098.
- [33] Barclay AN, Mayrhofer G. Bone marrow origin of Ia-positive cells in the medulla of rat thymus. *J Exp Med* 1981;153: 1666.
- [34] Zhao Y, Swenson K, Sergio JJ, Arn JS, Sachs DH, Sykes M. Skin graft tolerance across a discordant xenogeneic barrier. *Nature Med* 1996;2: 1211-1216.
- [35] Yamada K, Shimizu A, Utsugi R et al. Thymic transplantation in miniature swine. II. Induction of tolerance by transplantation of composite thymokidneys to thymectomized recipients. *J Immunol* 2000;164: 3079-3086.
- [36] Kamano C, Vagefi PA, Kumagai N et al. Vascularized thymic lobe transplantation in miniature swine: Thymopoiesis and tolerance induction across fully MHC-mismatched barriers. *Proc Natl Acad Sci U S A* 2004;101: 3827-3832.
- [37] Charlton B, Auchincloss H, Jr., Fathman CG. Mechanisms of transplantation tolerance. *Annu Rev Immunol* 1994;12: 707-734.
- [38] Kurtz J, Shaffer J, Lie A, Anosova N, Benichou G, Sykes M. Mechanisms of early peripheral CD4 T-cell tolerance induction by anti-CD154 monoclonal antibody and allogeneic bone marrow transplantation: evidence for anergy and deletion but not regulatory cells. *Blood* 2004;103: 4336-4343.
- [39] Quezada SA, Jarvinen LZ, Lind EF, Noelle RJ. CD40/CD154 interactions at the interface of tolerance and immunity. *Annu Rev Immunol* 2004;22: 307-328.
- [40] Rocha B, Grandien A, Freitas AA. Anergy and exhaustion are independent mechanisms of peripheral T cell tolerance. *J Exp Med* 1995;181: 993-1003.



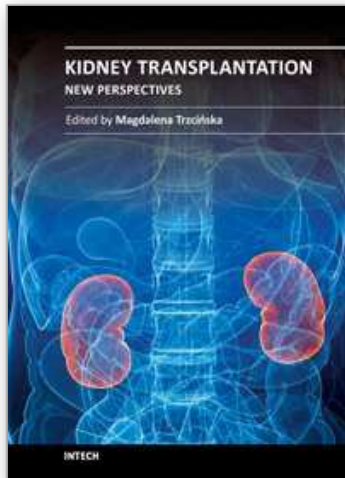
- [41] Sato K, Yamashita N, Yamashita N, Baba M, Matsuyama T. Regulatory dendritic cells protect mice from murine acute graft-versus-host disease and leukemia relapse. *Immunity* 2003;18: 367-379.
- [42] Charpentier B, Alard P, Hiesse C, Lantz O. Induction of peripheral T cell tolerance and allo/xenoimmunity. *Pathol Biol (Paris)* 1994;42: 237-240.
- [43] Wekerle T, Kurtz J, Sayegh M et al. Peripheral deletion after bone marrow transplantation with costimulatory blockade has features of both activation-induced cell death and passive cell death. *J Immunol* 2001;166: 2311-2316.
- [44] Hara M, Kingsley CI, Niimi M et al. IL-10 is required for regulatory T cells to mediate tolerance to alloantigens in vivo. *J Immunol* 2001;166: 3789-3796.
- [45] Ierino FL, Yamada K, Hatch T, Sachs DH. Preliminary in vitro evidence for regulatory cells in a miniature swine renal allograft model. *Transplant Proc* 1997;29: 1165.
- [46] Sakaguchi S. Regulatory T cells: key controllers of immunologic self-tolerance. *Cell* 2000;101: 455-458.
- [47] Sakaguchi S, Sakaguchi N, Asano M, Itoh M, Toda M. Immunologic self-tolerance maintained by activated T cells expressing IL-2 receptor alpha-chains (CD25). Breakdown of a single mechanism of self-tolerance causes various autoimmune diseases. *J Immunol* 1995;155: 1151-1164.
- [48] Wing K, Onishi Y, Prieto-Martin P et al. CTLA-4 control over Foxp3+ regulatory T cell function. *Science* 2008;322: 271-275.
- [49] Wood KJ, Sakaguchi S. Regulatory T cells in transplantation tolerance. *Nat Rev Immunol* 2003;3: 199-210.
- [50] Tada T, Takemori T. Selective roles of thymus-derived lymphocytes in the antibody response. I. Differential suppressive effect of carrier-primed T cells on hapten-specific IgM and IgG antibody responses. *J Exp Med* 1974;140: 239-252.
- [51] Takemori T, Tada T. Selective roles of thymus-derived lymphocytes in the antibody response. II. Preferential suppression of high-affinity antibody-forming cells by carrier-primed suppressor T cells. *J Exp Med* 1974;140: 253-266.
- [52] Lee ST, Paraskevas F. Soluble factors in immune sera of mice. I. Specific and non-specific suppressive activity. *Clin Exp Immunol* 1976;24: 177-184.
- [53] Ricciardi-Castagnoli P, Doria G, Adorini L. Production of antigen-specific suppressive T cell factor by radiation leukemia virus-transformed suppressor T cells. *Proc Natl Acad Sci U S A* 1981;78: 3804-3808.
- [54] Sakaguchi S, Sakaguchi N, Shimizu J et al. Immunologic tolerance maintained by CD25+ CD4+ regulatory T cells: their common role in controlling autoimmunity, tumor immunity, and transplantation tolerance. *Immunol Rev* 2001;182:18-32.: 18-32.
- [55] Kirk AD. Crossing the bridge: large animal models in translational transplantation research. *Immunol Rev* 2003;196: 176-196.
- [56] Larsen CP, Knechtle SJ, Adams A, Pearson T, Kirk AD. A new look at blockade of T-cell costimulation: a therapeutic strategy for long-term maintenance immunosuppression. *Am J Transplant* 2006;6: 876-883.
- [57] Salama AD, Remuzzi G, Harmon WE, Sayegh MH. Challenges to achieving clinical transplantation tolerance. *J Clin Invest* 2001;108: 943-948.



- [58] Madsen JC, Yamada K, Allan JS et al. Transplantation tolerance prevents cardiac allograft vasculopathy in major histocompatibility complex class I-disparate miniature swine. *Transplantation* 1998;65: 304-313.
- [59] Pennington LR, Flye MW, Kirkman RL, Thistlethwaite JR Jr, Williams GM, Sachs DH. Transplantation in miniature swine. X. Evidence for non-SLA-linked immune response gene(s) controlling rejection of SLA-matched kidney allografts. *Transplantation* 1981;32: 315-320.
- [60] Gianello PR, Yamada K, Fishbein JM et al. Long-term acceptance of primarily vascularized renal allografts in miniature swine. Systemic tolerance versus graft adaptation. *Transplantation* 1996;61: 503-506.
- [61] Fishbein JM, Gianello P, Nickleit V et al. Interleukin-2 reverses the ability of cyclosporine to induce tolerance to class I disparate kidney allografts in miniature swine. *Transplant Proc* 1993;25: 322-323.
- [62] Mezrich J, Yamada K, Sachs DH, Madsen JC. Regulatory T cells generated by the kidney may mediate the beneficial immune effects of combining kidney with heart transplantation. *Surgery* 2004;135: 473-478.
- [63] Gianello P, Fishbein JM, Sachs DH. Tolerance to primarily vascularized allografts in miniature swine. *Immunol Rev* 1993;133: 19-44.
- [64] Rosengard BR, Ojikutu CA, Guzzetta PC et al. Induction of specific tolerance to class I disparate renal allografts in miniature swine with cyclosporine. *Transplantation* 1992;54: 490-497.
- [65] Radovancevic B, Frazier OH. Treatment of moderate heart allograft rejection with Cyclosporine. *J Heart Transplant* 1986;5: 307-311.
- [66] Gianello PR, Fishbein JF, Rosengard BR et al. Tolerance to class I disparate renal allografts in miniature swine: maintenance of tolerance despite induction of specific antidonor CTL responses. *Transplantation* 1995;59: 772-777.
- [67] Gianello PR, Lorf T, Yamada K et al. Induction of tolerance to renal allografts across single- haplotype MHC disparities in miniature swine. *Transplantation* 1995;59: 884-890.
- [68] Fishbein JM, Rosengard BR, Gianello P et al. Development of tolerance to class II mismatched renal transplants following a short course of cyclosporine therapy in miniature swine. *Transplantation* 1994;57: 1303-1308.
- [69] Utsugi R, Barth RN, Lee RS et al. Induction of transplantation tolerance with a short course of tacrolimus (FK506): I. Rapid and stable tolerance to two-haplotype fully mhc-mismatched kidney allografts in miniature swine. *Transplantation* 2001;71: 1368-1379.
- [70] Vagefi PA, Ierino FL, Gianello PR et al. Role of the thymus in transplantation tolerance in miniature Swine: IV. The thymus is required during the induction phase, but not the maintenance phase, of renal allograft tolerance. *Transplantation* 2004;77: 979-985.
- [71] Yamada K, Vagefi PA, Utsugi R et al. Thymic transplantation in miniature swine: III. Induction of tolerance by transplantation of composite thymokidneys across fully major histocompatibility complex-mismatched barriers. *Transplantation* 2003;76: 530-536.
- [72] Ierino FL, Yamada K, Hatch T, Rembert J, Sachs DH. Peripheral tolerance to class I mismatched renal allografts in miniature swine: donor antigen-activated peripheral

- blood lymphocytes from tolerant swine inhibit antidonor CTL reactivity. *J Immunol* 1999;162: 550-559.
- [73] Gianello PR, Blancho G, Fishbein JF et al. Mechanism of cyclosporin-induced tolerance to primarily vascularized allografts in miniature swine. Effect of administration of exogenous IL-2. *J Immunol* 1994;153: 4788-4797.
- [74] Rosengard BR, Kortz EO, Ojikutu CA et al. The failure of skin grafting to break tolerance to class I disparate renal allografts in miniature swine despite inducing marked anti-donor cellular immunity. *Transplantation* 1991;52: 1044-1052.
- [75] Rosengard BR, Fishbein JM, Gianello PR et al. Retransplantation in miniature swine: lack of a requirement for graft adaptation for maintenance of specific renal allograft tolerance. *Transplantation* 1994;57: 794-799.

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## **Kidney Transplantation - New Perspectives**

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Although many years have passed since the first successful kidney transplantation, the method, although no longer considered a medical experiment, is still perceived as controversial and, as such, it triggers many emotions and that's why conscious educational efforts are still needed for kidney transplantation, for many people being the only chance for an active lifestyle and improved quality of life, to win common social acceptance and stop triggering negative connotations. Apart from transplantation controversies piling up over years transplantologists also have to face many other medical difficulties. The chapters selected for this book are of high level of content, and the fact that their authors come from many different countries, and sometimes even cultures, has facilitated a comprehensive and interesting approach to the problem of kidney transplantation. The authors cover a wide spectrum of transplant-related topics.

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Unit 405, Office Block, Hotel Equatorial Shanghai  
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Phone: +86-21-62489820  
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