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Acaricides – Biological Profiles, Effects and Uses in Modern Crop Protection

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1. Introduction

Acaricide is a pesticide designed to control harmful species of mites (Acari)1. In crop protection practices, acaricides are used against phytophagous mites, pests causing economic injuries to agricultural crops and ornamental plants. Until mid-twentieth century, in agroecosystems of low-level productivity, phytophagous mite populations usually stayed below economic injury levels, due to natural regulation by predatory mites and insects, their natural enemies. The concept of secondary pest outbreak was introduced on spider mites (Tetranychidae), the most important plant-feeding mites, as a paradigm. Advances in agricultural production after World War II, based on the extensive use of pesticides and fertilizers, irrigation and other cultural practices, induced increase in spider mite populations far above economic threshold (Huffaker et al., 1970; McMurtry et al., 1970; Jeppson et al., 1975; Metcalf, 1980). Grown under favourable conditions, host plants became high quality food sources for the mite pests, which gave rise to outbreaks of their populations and made it possible to compensate for the losses caused by predators' activity. Moreover, widespread use of neuroactive insecticides (synthetic organic compounds used against insects as target pests, but toxic to other non-target insect and mite species as well) destroyed spider mite predators, generally more susceptible than their prey; on the other hand, heavy selection pressure by neuroactive insecticides caused emergence of tetranychid mite populations resistant to these compounds. Besides the resistance of spider mites and the elimination of their predators, as the primary causes, outbreaks are influenced by sublethal effects of pesticides on behaviour and physiology of pests and/or predators (Metcalf, 1980; Hardin et al., 1995; Dutcher, 2007).

Spider mites, mostly polyphagous species, are common pests in modern agroecosystems worldwide, and some of them are among the most important crop pests. After Tetranychidae,

¹ Mites (subclassis Acari), morphologically and ecologically very diverse assemblage of tiny invertebrates, belongs to class Arachnida (together with spiders and scorpions), subphylum Chelicerata and phylum Arthropoda. The arthropods also include insects, from which mites differ, beside being eight-legged animals (insects are hexapods) by the lack of true head and conspicuous body segmentation. There are some 50.000 mite species known today, but it is estimated that the true number is 20 times higher. Besides agricultural pests and their natural enemies (predators), mites include species of medical and veterinary importance (house dust mites, scabies mites, ticks), while the species living in soil and water are important environmental indicators.

the second most important mite pests are gall and rust mites (Eriophyoidea), while the other economically harmful species can be found among false spider mites (Tenuipalpidae), tarsonemid mites (Tarsonemidae) and acarid mites (Acaridae). Phytophagous mites feed on the liquid content of plant cells, thus disrupting the physiology of a host plant and causing various damages to plant tissues and organs, while some of the species can also act as vectors of plant viruses. In spite of relatively small size (100-400 µm), plant-feeding mites can cause considerable crop yield and quality losses, because they have short life span and under favourable conditions their populations quickly reach high abundance (Helle & Sabelis, 1985a,b; Lindquist et al., 1996; Zhang, 2003; van Leeuwen et al., 2010).

The use of acaricides has increased substantially over the past half of the 20th century. Since the first serious and widespread outbreaks of spider mites populations, during the 1950s, organophosphorous and other neuroactive insecticides were replaced by specific acaricides i.e. compounds exclusively or primarily effective against mites. Several generations of structurally diverse synthetic acaricides, directed against various biochemical and physiological targets, have been commercialized until now. Besides specific acaricides, a number of insecticides with considerable acaricidal activity (pyrethroids, avermectins, benzoylureas) have also been used, while some older neuroactive compounds are still available for the control of phytophagous mites (Jeppson et al., 1975; Knowles, 1997; Dekeyser, 2005; van Leeuwen et al., 2010). Most of the modern acaricides exert their effects through disruption of respiratory processes. Another approach in the development of synthetic acaricides launched compounds that act on growth and development (Dekeyser, 2005; Krämer & Schirmer, 2007). On the other hand, various natural bioactive products with acaricidal activity (botanical and microbial pesticides, essential oils, horticultural spray oils, mycopesticides) have become important alternatives to synthetic acaricides (Beattie et al., 2002; Copping & Duke, 2007; Faria & Wraight, 2007).

Acaricide resistance in phytophagous mites is a seriously increasing phenomenon, especially in spider mites which have a remarkable intrinsic potential for rapid evolution of resistance (Croft & van de Baan, 1988; van Leeuwen et al., 2009). Their populations have often developed a very high degree of resistance to a newly introduced compound after few years of use, with cross-resistance to other compounds with the same mode of action. According to APRD (Arthropod Pesticide Resistance Database) more than 700 cases of acaricide resistance in phytophagous mites have been reported. About 93% of these reports refer to spider mites resistance, and almost a half of spider mite resistance cases is related to the twospotted spider mite (Tetranychus urticae), highly polyphagous species, one of the most important pests in greenhouses throughout the world (Whalon et al., 2008, 2010). Therefore, there is a continual need for development and application of new acaricides with novel biochemical modes of action, but also for optimization of their use in order to prevent or delay the evolution of resistence and prolong their life span (Dekeyser, 2005). Considering biorational pest control as key approach to modern crop protection (Horowitz et al., 2009) new acaricides should be selective, that is, effective against the target pests and compatible with their natural enemies. Moreover, these compounds must be safe products with respect to human health, beneficial and non-target organisms (mammals, birds, earthworms, bees, aquatic organisms) and the environment in order to meet the regulatory requirements.

This chapter focuses on biological profiles of acaricides that have been commercialized at the end of the 20th and beginning of the 21st century, acaricide resistance in phytophagous mites, bioactive products of natural origin as alternatives to synthetic acaricides, compatibility of acaricides with the biological control agents, and other current issues related to acaricide uses in modern crop protection.

2. Summary of acaricide development

As already remarked, phytophagous mites became important pests of cultivated plants in the mid-20th century, during the "golden age of insecticide discovery" (Casida & Quistad, 1998) that was marked by intensive use of **organochlorines**, **organophosphates** and **carbamates**, broad spectrum insecticides which, as it was later discovered, included many acaricidal compounds. Those were the neuroactive compounds which disrupt the transmission of impulses between nerve cells of an insect by blocking the action of the enzyme acetylcholinesterase (organophosphates, carbamates) or interferring with ion channels in the nerve membrane (organochlorines) (Ishaaya, 2001). Figure 1 shows chlorpyrifos, probably the most common commercialized organophosphate today, carbaryl, the first synthesized carbamate, and endosulfan, one of the rare organochlorine compounds still in use.

Fig. 1. Neuroactive insecticides with acaricidal activity: chlorpyrifos (organophosphate), carbaryl (carbamate) and endosulfan (organochlorine)

The first serious and widespread spider mite outbreaks following applications of neuroactive insecticides, observed at the end of 1940s and beginning of 1950s, initiated the research and development of specific acaricides. These compounds, exclusively or primarily effective against mites, were gradually taking over the organochlorines, organophosphates and carbamates. Bridged diphenyls (bromopropylate, chloropropylate, chlorobenzilate, chlorfenethol, dicofol, tetradifon), the first specific acaricides, established themselves on the market in the 1950s. During the 1960s and early 1970s, the second generation of structurally rather different specific acaricides emerged, the most important of which were propargite, organotins (cyhexatin, fenbutatin-oxide) and formamidines (amitraz, chlordimeform). Most of first and second generation acaricides are not used any longer. Specific acaricides of the third generation are represented by mite growth inhibitors (clofentezine, hexythiazox), commercialized in the first half of the 1980s (Fig. 2) In addition to specific acaricides, structurally diverse synthetic acaro-fungicides (dinocap, chinomethionate, dichlofluanid) were introduced; on the other hand, the use of sulfur products (that had been exploited as acaro-fungicides since 19th century) was largely displaced by novel synthetic compounds.

Introduction of specific acaricides reduced the adverse impact on beneficial insects (predators of insect and mite pests, polinators) to the minimum; at the same time, many specific acaricides proved to be selective, i.e. less toxic to predaceous mites than phytophagous mites. These acaricides effectively control populations of phytophagous mites resistant to neuroactive compounds, since they are compounds having different biochemical modes of action, with targets mostly being outside the nervous system (March, 1976; Knowles, 1976, 1997; Ishaaya, 2001; Krämer & Schirmer, 2007). Moreover, specific acaricides are far more safer for humans, non-target organisms and the environment in

$$CI \longrightarrow CH_3 \longrightarrow CH_3 \longrightarrow CH_3$$

$$CI \longrightarrow CH_3 \longrightarrow CH_3$$

$$CH_3 \longrightarrow CH_3$$

$$CH_3$$

Fig. 2. Representatives of the first (dicofol), second (amitraz, propargite, cyhexatin) and third (clofentezine) generation of specific acaricides

comparison to neuroactive compounds, in particular organochlorines that were almost all severely restricted or banned in developed countries in the 1970s. Organophosphates and carbamates, however, remain to be the predominant group of insecticides by accounting for 35% of the global market (van Leeuwen et al., 2009).

In addition to specific acaricides, two new groups of synthetic insecto-acaricides were placed on the market in the 1970s and 1980s: **pyrethroids** (neuroactive compounds, sodium channel modulators) and **benzoylureas** (compounds acting on growth and development by inhibition of biosynthesis of chitin, a biopolymer present in the cuticle of arthropods). Another new commercial product was **abamectin**, neuroactive insecto-acaricide (chloride channel activator), a mixture of macrocyclic lactones avermectin B_{1a} and avermectin B_{1b}, natural products isolated from the fermentation of *Streptomyces avermitilis*, a soil Actinomycete (Fig. 3), (Ishaaya, 2001; Krämer & Schirmer, 2007). These compounds increased the biochemical diversity of acaricides and insecto-acaricides, but beside the partly expected resistance, some other problems emerged, such as the pyrethroid-induced spider mite outbreaks (Gerson & Cohen, 1989; van Leeuwen et al., 2009).

In the last two decades, a considerable number of non-neuroactive synthetic acaricides and insecto-acaricides emerged on the global market, but there is also a growing interest to find new and reinstate the already known acaricidal compounds of natural origin (Dekeyser, 2005; Copping & Duke, 2007; Krämer & Schirmer, 2007). The search for new chemistries that act on novel target sites and determination of an efficient strategy for use of acaricides that have different biochemical modes of action is presently the only sustainable solution that can prevent or delay the evolution of resistance and prolong the life span of acaricides (Dekeyser, 2005).

Nowadays, acaricides are developed under conditions marked by growing demand of the public opinion for safer, "greener" pesticides, and increasingly stricter toxicological and

$$(Z)-(1R)-cis-$$

$$CF_3$$

$$CH_3$$

Fig. 3. Structural formulas of bifenthrin (pyrethroid), flucycloxuron (benzoylurea) and abamectin (natural product)

eco-toxicological criteria for market circulation of the existing pesticides and registration of the new ones imposed by the regulatory agencies and issues (Casida & Quistad, 1998; Dekeyser, 2005). In the USA, the passage of the Food Quality Protection Act (FQPA) of 1996 brought about significant changes in the way in which pesticides are registered by the U.S. EPA (Environmental Protection Agency). Besides re-evaluation of registered pesticides, priority in registration program has been given to "reduced-risk *pesticides*", i.e. pesticides with reduced risk to human health, non-target organisms and environment as a replacement for older and potentially riskier chemicals. The list of reduced-risk pesticides includes several new acaricides and insecto-acaricides (EPA, 2009, 2010). In the European Union, implementation of Directive 91/414 that requires science-based assessment of pesticide risk to human health and the environment, has seriously impacted the EU acaricide portfolio. Nevertheless, new Regulation (EC) 1107/2009, revises the Directive and introduces hazardbased cut-off criteria, thus increasing the safety level (Balderacchi & Trevisan, 2010; van Leeuwen et al., 2010). When looking at the acaricides, from 103 substances, only 26 are currently included in a 'positive' list of compounds (Annex I) and the status of another four is pending (EU, 2010).

3. New synthetic acaricides

3.1 Acaricides acting on respiration targets

Similar to nervous system of insects, nervous system of mites has also long been the target for most chemicals used for their control (Casida & Quistad, 1998). The situation has somewhat changed during the last two decades due to commercialization of large number of acaricidal compounds acting on mitochondrial respiration process, that produces most of the energy in cells. This process includes two coupled parts: mitochondrial electron transport (MET) and oxidative phosphorylation. Although some of the older acaricides were known to inhibit respiration, the real exploitation of this target started no sooner than after the 1990s, with the prospects for expanding and developing new, more effective and safer products (Dekeyser, 2005; Lűmmen, 2007; Krämer & Schirmer, 2007).

Throughout the mitochondrial electron transport chain there are various potential sites for inhibition, but only three have been used so far as target sites of acaricidal activity, at transmembrane enzyme complexes. In the period 1991-93, four compounds from different chemical classes were successively commercialized: fenpyroximate (pyrazole), pyridaben (pyridazinone), fenazaquin (quinazoline) and tebufenpyrad (pyrazolecarboxamide) (Fig. 4), whose mode of action was inhibition of MET at complex I. These compounds, also known as METI acaricides, quickly gained the popularity worldwide owing to the high efficacy against both tetranychid and eriophyoid mites, quick knockdown effect and long-lasting impact. In addition, these substances have low to moderate mammalian toxicity and short to moderate environmental persistence (Dekeyser, 2005; Krämer & Schirmer, 2007, van Leewen et al., 2010). Fenpyroximate and tebufenpyrad are included in Annex I, while fenazaquin and pyridaben applications have been resubmitted for inclusion (EU, 2010). Fenpyroximate is also on the list of reduced risk and organophosphorus alternative pesticides (EPA, 2009). Complex I inhibitors also include pyrimidifen (pyrimidinamine), commercialized in 1995, as well as insecto-acaricide tolfenpyrad, another pyrazolecarboxamide, commercialized in 2002, and **flufenerim**, more recent derivative of pyrimidifen (Krämer & Schirmer, 2007).

The only known complex II inhibitor is the recently introduced insecto-acaricide cyenopyrafen, a compound from the acrylonitrile class of chemistry (Lűmmen, 2007). Complex III inhibition is mode of action of acequinocyl, fluacrypyrim and bifenazate. Acequinocyl, a naphthoquinone compound (Fig. 5) commercialized in 1999, is a proacaricide which is bioactivated via deacetylation. It is effective against all stages of spider mites, with low mammalian toxicity and short environmental persistence (Dekeyser, 2005). It is included in the EPA list of reduced risk pesticides, while the decision on its status under Directive 91/414 is pending (EPA, 2009; EU, 2010). Bifenazate, a carbazate compound (Fig. 5) is highly effective against immatures and adults of spider mites, with rapid knockdown effect (Ochiai et al., 2007). Although it was first considered to be a neurotoxin, more recent experimental results indicate complex III as target site (van Nieuwenhuyse et al., 2009). Bifenazate is a pro-acaricide which is bioactivated via hydrolysis of ester bonds, so the organophosphorous compounds, as inhibitors of esterase hydrolitic activity, can antagonize the toxicity of this acaricide (van Leeuwen et al., 2007). Bifenazate, introduced in 1999, is a compound of low mammalian toxicity and short environmental persistence; it is classified as a reduced risk and organophosphate alternative pesticide, and it is included in Annex I (Dekeyser, 2005; EPA, 2009; EU, 2010). Fluacrypyrim, introduced in 2002, shows acaricidal effect against all stages of tetranychids. This is the first strobilurin not commercialized as a fungicide (Dekeyser, 2005; Krämer & Schirmer, 2007), and more compounds with acaricidal effect from this group are anticipated (Li et al., 2010).

$$CH_3 \qquad N \qquad CH_3 \qquad CH_3 \qquad CH_2 \qquad CH_3 \qquad CH_2 \qquad CH_3 \qquad C(CH_3)_3 \qquad C(CH_2)_1 \qquad C(CH_2)_1 \qquad C(CH_2)_2 \qquad$$

Fig. 4. Structural formulas of acaricides acting on respiration targets: METI acaricides

Insecto-acaricide **diafenthiuron**, a novel thiourea compound (Fig. 5) launched in 1991, is the only modern representative of compounds that disrupt oxidative phosphorylation by inhibition of the mitochondrial ATP synthase, an enzyme with essential role in cellular bioenergetics (this mode of action has been recognized in propargites, tetradifons and organotin compounds). Diafenthiuron is a pro-acaricide, its carbodiimide metabolite inhibits the enzyme. It is effective against motile stages of spider mites and also provides good eriophyoid control. Diafenthiuron has low mammalian toxicity and short environmental persistence (Krämer & Schirmer, 2007; van Leewen et al., 2010).

Fig. 5. Structural formulas of acaricides acting on respiration targets: complex III inhibitors (acequinocyl, bifenazate) and inhibitors of oxidative phosphorylation (diafenthiuron, chlorfenapyr)

Another insecto-acaricide, **chlorfenapyr**, a pyrrole compound (Fig. 5) commercialized in 1995, at biochemical level acts as uncoupler of oxidative phosphorylation via disruption of the proton gradient. Chlorfenapyr is effective against all stages of spider mites and eriophyoid mites. This compound is a pro-acaricide activated by N-dealkylation. Chlorfenapyr is a compound of moderate mammalian toxicity, but long environmental persistence (Krämer & Schirmer, 2007; Van Leeuwen et al., 2010). It is included in the EPA list as an alternative to organophosphorus compounds (EPA, 2009).

3.2 Acaricides acting on growth and development targets

Another direction in research and development of synthetic acaricides is directed towards compounds affecting developmental processes. **Etoxazole**, a oxazoline compound (Fig. 6), is acaricide highly effective against eggs and immatures of spider mites, non-toxic to adults, but it considerably reduces fertility of treated females (Kim & Yoo, 2002; Dekeyser, 2005). This acaricide, launched in 1998, is usually classified among mite growth inhibitors, together with clofentezine and hexythiazox, older acaricides that cause similar symptoms (Marčić, 2003; Krämer & Schirmer, 2007), but whose exact mode of action is unknown. On the other hand, Nauen & Smagghe (2006) provided experimental evidence that etoxazole acts as a chitin synthesis inhibitor similar to benzoylureas. Etoxazole is on the EPA list of reduced risk and organophosphorus alternative pesticides (EPA, 2009).

Discovery of **spirodiclofen** and **spiromesifen**, tetronic acid derivatives (Fig. 6) launched in 2002-2004, broadened the biochemical diversity of acaricides by introducing a completely new mode of action. These compounds act as inhibitors of acetyl-CoA-carboxylase, a key enzyme in fatty acid biosynthesis. Spirodiclofen and spiromesifen are highly toxic to eggs and immatures of spider mites, while their effects on adult females are slower with fecundity and fertility reduction; their acaricidal effect is long-lasting and stable (Krämer &

FOCH₂CH₃

$$C(CH_3)_3$$

$$CH_3$$

$$CH_3$$

$$CH_3$$

$$CH_3$$

$$CH_2$$

$$CH_3$$

$$CH_3$$

$$CH_2$$

$$CH_3$$

$$CH_$$

Fig. 6. Structural formulas of acaricides acting on growth and development

Schirmer, 2007; Marčić, 2007; Marčić et al., 2007; Van Pottelberge et al., 2009; Marčić et al., 2010). These two acaricides are the only new compounds used for control of eriophyoid mites as well (van Leeuwen et al., 2010). In addition to acaricidal effect, spirodiclofen has also shown considerable insecticidal activity against eggs and larvae of pear psylla and scales (Krämer & Schirmer, 2007; Marčić et al., 2007), while spiromesifen provides effective control of whiteflies (Krämer & Schirmer, 2007; Kontsedalov et al., 2008). Both compounds have low mammalian toxicity and short environmental persistence. Spirodiclofen is included in Annex I, and evaluation of spiromesifen is in progress (EU, 2010). Spirotetramat, a tetramic acid derivate recently introduced, belongs to inhibitors of acetyl-CoA-carboxylase. Although initially developed for control of whiteflies and aphids (Brück et al., 2009), the studies of its effects on *T. urticae* (Marčić et al., unpublished data) indicate that spirotetramat could potentially be an effective acaricide as well.

4. Natural acaricides and other alternative solutions

The use of natural products for plant and crop protection dates back to times long before the introduction of synthetic pesticides which imposed themselves as the main means for suppression of harmful organisms. In recent times, the significance of natural pesticides is constantly growing, primarily in organic agriculture, but also in the framework of biorational pest control programs which insist on use of environmentally-friendly pesticides and exploatation of novel biochemical modes of action (Isman, 2006; Isman & Akhtar, 2007; Copping & Duke, 2007; Horowitz et al., 2009). Some of the natural products are substances that have significant acaricidal effect.

Probably the most studied botanical insecticide in the last twenty years is a triterpenoid azadirachtin (Fig. 7), the major active ingredient of extracts, oils and other products derived from the seeds of the Indian neem tree (Azadirachta indica). Neem-products are registered in over 40 countries as products for suppression of arthropod pests important in growing of fruit, vegetables and ornamental plants (Kleeberg, 2004; Milenković et al., 2005). The effects of azadirachtin on treated insects manifest slowly and they include complete or partial antifeedant response, delayed and/or disrupted moulting, inhibited reproduction (Copping & Duke, 2007; Isman & Akhtar, 2007). The studies on spider mites (Sundaram & Sloane, 1995; Mansour et al., 1997; Martinez-Villar et al., 2005) indicate that azadirachtin, in addition to being toxic to various development stages, acts as antifeedant, reduces fecundity and fertility and shortens the life span of adult insects. Beside on spider mites, azadirachtin also exhibits acaricidal effect on some acarid and tarsonemid mites (Collins, 2006; Venzon et al., 2008). Azadirachtin is considered to be non-toxic to mammals and is not expected to have any adverse effects on the environment (Copping & Duke, 2007); its Annex I application is resubmitted (EU, 2010). Many neem/azadirachtin-based products are approved for use in organic crop production (Zehnder et al., 2007; EU, 2008; Dayan et al., 2009).

Products isolated from soil actinomycetes are an important source for deriving natural insecticides and acaricides. In early 1990s, several years after introduction of abamectin, another fermentation product, **milbemectin**, was commercialized. Milbemectin is a mixture of milbemycin A₃ and milbemycin A₄, natural products isolated from the fermentations of *Streptomyces hygroscopicus* subsp. *aureolacrimosus* (Fig. 7). Milbemectin is a neuroactive acaricide (chloride channel activator), effective against tetranychid and eriophyoid mites, relatively safe compound owing to the rapid uptake into treated plants combined with fast degradation of surface residues (Copping & Duke, 2007; Krämer & Schirmer, 2007). Like abamectin, milbemectin is also included in Annex I (EU, 2010).

$$CH_3 \xrightarrow{C} CH_3 \xrightarrow{H} CH_3$$

$$CH_3 \xrightarrow{C} CH_3 \xrightarrow{H} CH_3$$

$$CH_3 \xrightarrow{C} CH_3 \xrightarrow{C} CH_3$$

$$CH_3 \xrightarrow{C} CH_3 \xrightarrow{C}$$

Fig. 7. Natural products with considerable acaricidal activity

The more recent example is **spinosad**, a mixture of spinosyn A and spinosyn D, secondary metabolites of *Saccharopolyspora spinosa*, (Fig. 7), introduced in 1997 as neuroactive insecticide, nicotinic acetylcholine receptor agonist (Copping & Duke, 2007; Krämer & Schirmer, 2007). This insecticide exerts significant acaricidal effect. Van Leeuwen et al. (2005) found out that the residual toxicity of spinosad to female *T. urticae* is equal to the level of toxicity resulting from application of dicofol, bromopropylate or fenbutatin oxide, while Villanueva & Walgenbach (2006) demonstrated that spinosad affects larvae and adults of this tetranychid, but relatively slowly, with the assumption that negative results of the previous testing of acaricidal properties were based on experiments that did not provide enough time for response. Spinosad shows systemic acaricidal effect, if used for substrate watering, such as rockwool, where the absorption level is reduced to the minimum (Van Leeuwen et al. (2005). This is a compound of very low mammalian toxicity and highly favourable environmental profile; it is included in the EPA list of organophosphorus alternative pesticides, and also in Annex I (EPA, 2009; EU, 2010). Spinosad is approved for use as an organic insecticide (EU, 2008; Dayan et al., 2009).

Essential oils, secondary metabolites abundant is some aromatic plants from families Lamiaceae, Apiaceae, Rutaceae, Myrtaceae and others, have been suggested as alternative sources for pest control products. Predominant bioactive ingredients of essential oils are monoterpenes and sesquiterpenes. Besides exerting acute toxicity to insects and mites, essential oils show sublethal effect as repellents, antifeedants and reproduction inhibitors. Lethal and sublethal effects of essential oils are the consequence of direct contact and/or

uptake of gas-phase via respiratory system. Insect octopaminergic nervous system is considered to be the target site of action of some essential oil constituents, but this may not be the case considering their acaricidal activity. Moreover, there is a possibility that essential oils, as complex mixtures, act at multiple target sites (Isman, 2006; Miresmailli et al., 2006; Isman & Akhtar, 2007; Shaaya & Rafaeli, 2007). Essential oils extracted from caraway seeds, eucalyptus, mint, rosemary, basil, oregano, thyme, and other plants have shown a significant acaricidal activity (Aslan et al., 2004; Choi et al., 2004; Miresmailli et al., 2006). These oils could be useful as fumigants in the control of phytophagous mites in greenhouses; however, for improving acaricidal activity, their commercial formulations need to be developed (Choi et al., 2004; Han et al., 2010). Essential oils are mostly nontoxic to mammals; being volatile products, they have limited environmental persistence (Isman, 2006). Rosemary oil, thyme oil and some other essential oils are available for pest control in organic farming (Dayan et al., 2009).

Petroleum oils have been used for more than a century to control a wide range of crop pests, including spider mites. Because of their high phytotoxicity, the use of petroleum oils was limited to dormant or delayed dormant application against overwintering pest stages, to avoid injury to green plant tissue. Advances in petroleum chemistry considerably reduced phytotoxicity in newer, highly-refined petroleum-derived spray oils (PDSO), which are recognized today as an important alternative to synthetic pesticides. PDSO are environmentally-friendly products with negligible impact on human health and the environment. The most widely accepted theory on their mode of action is that PDSO primarily act physically by blocking the spiracles in insects (or the stigmata in mites) and thus causing suffocation, but it can not be presumed as the only mode of action (Taverner, 2002). At least some modern oils cause a range of cellular disruption leading to rapid insect death (Najar-Rodriguez et al., 2008). PDSO are highly effective against spider mites and eriophyoid mites in various field and greenhouse crops (Agnello et al., 1994; Nicetic et al., 2001; Marčić et al., 2009; Chueca et al., 2010). Beside mineral, plant oils proved to be effective acaricides as well, such as cottonseed oil (Rock & Crabtree, 1987) soybean oil (Lancaster et al., 2002; Moran et al., 2003) and rapeseed oil (Kiss et al., 1996; Marčić et al., 2009). PDSO and plant spray oils are considered compatible with organic farming (Zehnder et al., 2007; EU, 2008). Rapeseed oil is included in Annex I (EU, 2010).

Numerous studies indicate that entomopathogenic fungi, especially ascomycetes, can play an important role in regulation of harmful arthropod populations if used in biological control (Hajek & Delalibera, 2010), or applied as **mycoinsecticides** and/or **mycoacaricides** (Maniania et al., 2008; Jackson et al., 2010). Among the entomopathogenic fungi, the most potent pathogens of tetranychids and other pest mite species are *Beauveria basssiana*, *Hirsutella thompsonii*, *Lecanicillium* sp., *Metharizium anisopliae*, *Isaria fumosorosea*, *Neozygites floridana* (Chandler et al., 2000; Maniania et al., 2008), whose conidia and blastospores are used for formulation of fungal-based biopesticides. At the beginning of the 1980s, only one mycoacaricide was available (Mycar), formulated from conidia of *H. thompsoni* and intended for supression of citrus rust mite. Quarter of century later, there are some 30 commercial products acting against tetranychid, eriophyoid, and tarsonemid mites, mostly formulated as wettable powder or oil dispersion, and one third of which is made from conidia of *B. bassiana* (Faria & Wraight, 2007).

5. Acaricide resistance in phytophagous mites

As a result of exceptional intrinsic potential of mites for rapid development of resistance (Cranham & Helle, 1985; Croft & van de Baan, 1988; van Leeuwen et al., 2009) and often not

so rational actions of humans, the acaricide resistance in mites, in particular the species from Tetranychidae family, has become a global phenomenon. Arthropod Pesticide Resistance Database (APRD) - managed by scientist from Michigan State University and supported by Insecticide Resistance Action Committee (IRAC), a specialist technical group of the industry association CropLife – contains published data on resistance in insects and mites important for agriculture, veterinary medicine and public health, from 1914 to date (Whalon et al., 2008, 2010). This database, which involves a large number of scientists and experts from around the world who work on its administration and upgrading, is useful for comprehension of acaricide resistance in mites on a global level.

In the mid-2010, APRD contained 9394 reports on resistance developed in 572 species of arthropods, of which 1130 reports refer to 82 species from Acari subclass. Out of this number, 745 reports concern 39 species belonging to four families of phytophagous mites: Tetranychidae, Acaridae, Eriophyidae and Tenuipalpidae. Approximately 93% of reports deal with the resistance of spider mites, with two predominant species two-spotted spider mite, *Tetranychus urticae* (53% of spider mite reports) and European red mite, *Panonychus ulmi* (26% of spider mite reports) (Tab. 1). The authors of the APRD created the list of the "top 20" resistant arthropod pests in the world, ranked by number of compounds with reported resistance. On this list, *T. urticae* and *P. ulmi* rank first and ninth, respectively, by data for 92 and 42 compounds for which the information about resistant populations exist (Whalon et al., 2008, 2010).

For both species, the majority of reports refer to resistance to organophosphates documented during the 1950s, 1960s, and 1970s. Together with carbamates, organophosphate compounds account nowadays for more than 35% of global insecticide market, so that the reports on resistant tetranychid populations/strains are still coming (Herron et al., 1998; Stumpf et al., 2001; Tsakaragkou et al., 2002; Kumral et al., 2009). The important part of the APRD database concerns pyrethroids resistance in tetranychids. Today, this class of compounds accounts for 20% of the market, but the increasing number of cases of resistance to bifenthrin and other pyrethroids has been registered in the recent past (Herron et al., 2001; Ay and Gűrkan, 2005; Kumral et al., 2009; Tsakaragkou et al., 2009). As for other specific acaricides and insecto-acaricides, there is practically no active substance without documented cases of resistance, but there is an obvious difference in the scope of phenomenon between certain acaricides or groups of acaricides. For instance, global popularity of METI-acaricides contributed to relatively fast development of resistant spider mite populations in Japan, South Korea, Australia, Brazil, California and some European countries. On the other hand, there are only few reports on resistance to fenbutatin-oxide and other organotin compounds which have been used for four decades now (Stumpf & Nauen, 2001; Auger et al., 2004; van Leeuwen et al., 2009; Stavrinides et al., 2010).

Another phytophagous mite on the list of "top 20", bulb mite *Rhizoglyphus robini* (Acaridae), ranks 19th with 22 reports on resistance to almost exclusively organophosphate compounds. Cases of resistance to organophosphates are also registered for other species of acarids listed in the APRD. Among eriophyoid mites, resistance in citrus rust mite, *Phyllocoptruta oleivora*, to dicofol has been best documented and most studied (Omoto et al., 1994); other cases of resistance in Eriophyoidea also refer to organophosphorous compounds.

In addition to comprehensive documenting of acaricidal resistance in mites, the factors affecting this phenomenon of microevolution were also studied, as well as its physiological, biochemical ang genetic mechanisms. The results of these studies were summarized by Cranham & Helle (1985), Croft & van de Baan (1988), Messing & Croft (1996), Knowles

Mite species	No. of cases	No. of compounds
Tetranychus urticae	367	92
Panonychus ulmi	181	42
Panonychus citri	26	20
Tetranychus cinnabarinus	26	16
Tetranychus mcdanieli	19	13
Tetranychus kanzawai	12	12
Tetranychus viennensis	7	7
Tetranychus atlanticus	7	5 5
Tetranychus pacificus	7	5
Oligonychus pratensis	6	6
Tetranychus turkestani	5	5
Tetranychus hydrangaea	5	4
Tetranychus arabicus	3	3
Tetranychus crataegi	3	3
Tetranychus desertorum	3	3
Tetranychus ludeni	3	3
Tetranychus bimaculatus	2	2
Tetranychus cucurbitacearum	2	2
Tetranychus schoenei	2	2
Tetranychus tumidus	2	2
Eotetranychus hicoriae	1	1
Tetranychus althaeae	1	1
Tetranychus canadensis	1	1
Tetranychidae	691	
Rhizoglyphus robini	22	22
Rhizoglyphus echinopus	6	5
Acarus siro	$\frac{6}{4}$	3
Acarus chaetoxysilus	2	2
Acarus farris	1	1
Tyrophagus palmarum	1	1
Tyrophagus putrescentiae	1	1
Acaridae	37	
Acandae	37	
Phyllocoptruta oleivora	3	2
Aculus cornutus —	3	3
Aculus pelekassi	3 / / (3
Aculus fockeui	1	
Aculus lycopersici	1	
Aculus malivagrans	1	1
Aculus schlechtendali	1	1
Eriophyidae	13	
Brevipalpus chilensis	3	3
Brevipalpus phoenicis	1	1
Tenuipalpidae	4	•
Tenuipaipiuae	4	

Tab. 1. Reported cases of acaricide resistance in phytophagous mites (Whalon et al., 2010)

(1997), van Leeuwen et al. (2009), and the largest number of data refers to populations/strains of *T. urticae*. As in other arthropods, the resistance in mites is caused by a less sensitive target site (target site resistance) and/or enhanced detoxification (metabolic resistance). The insensivity of acetylcholinesterase is the most common type of organophosphorous resistance in *T. urticae* (Cranham & Helle, 1985; Stumpf et al., 2001, Tsagkarakou et al., 2002; van Leeuwen et al., 2009; Khajehali et al., 2010; Kwon et al., 2010a). Metabolic resistance mediated by carboxylesterases was found in majority of cases of resistance development to pyrethroids in this species (Ay & Gűrkan, 2005; van Leeuwen et al., 2005b, van Leeuwen & Tirry, 2007), while the oxidative metabolism appears to play a major role in resistance to METI-acaricides (Stumpf & Nauen, 2001; Kim et al., 2004, 2006; van Pottelberge et al., 2009).

The results of numerous conventional genetic studies indicate that in most cases single major gene controls inheritance of resistance in spider mites (Cranham & Helle, 1985; van Leeuwen et al., 2009). Although the monogenic and dominant resistance has been expected due to intense selection pressure to which the populations under the open field or greenhouse conditions are exposed (Roush & McKenzie, 1987), some major exceptions occur, such as the monogenic-recessive resistance to dicofol (Rizzieri et al., 1988), propargite (Keena & Granett, 1990), pyridaben (Goka, 1998) and etoxazole (Uesugi et al., 2002), and poligenic resistance to cyhexatin (Mizutani et al., 1988). Lately, several studies dealing with molecular basis of the target site resistance to pyrethroids (Tsagkarakou et al., 2009; Kwon et al., 2010b), organophosphates (Khajehali et al., 2010; Kwon et al., 2010a) and bifenazate (van Leeuwen et al., 2008, van Nieuwenhuyse et al., 2009) have been published. Especially interesting discovery is that the bifenazate resistance in *T. urticae* is inherited only maternally, which is the first occurrence of non-Mendelian inheritance since the beginning of genetic studies on pesticide resistance in arthropods (van Leeuwen et al., 2008, van Nieuwenhuyse et al., 2009).

Biological, biochemical and genetic characterization of resistance is one of the essential elements in defining the strategy for management of acaricide resistance in phytophagous mites. An effective acaricide resistance management program could be based on general resistance management principles endorsed by IRAC (Krämer & Schirmer, 2007). The key recommendation is reduction of the selection for resistance which is possible to attain if there were available as many as possible acaricides with different modes of action. The history of resistance in *T. urticae* best illustrates the importance of the above: the first resistant populations can emerge as soon as after two or three years from the start of a new acaricide application, causing an obvious pest control failure (Cranham & Helle, 1985; Knowles, 1997; van Leeuwen et al., 2009).

In the European Union, the implementation of Directive 91/414 reduced the EU acaricide portfolio by more than 70% (EU, 2010; van Leeuwen et al., 2010). On the other hand, it is Directive 91/414 that requires pesticide registrants to address the risk of resistance development as part of dossiers submitted for EU registration (Thompson et al., 2008). In "Declaration of Ljubljana" (Bielza et al., 2008) a group of leading resistance management experts expressed strong concern that further loss of active ingredients resulting from the implementation of Directive 91/414 (and its revision) could endanger the sustainability of European farming, increasing the risk of developing resistance to the relatively few remaining substances. The scientists concluded that the resistance management requires

access to a diversity of chemistries with different modes of action. Considering the fact that every year only few new active substances are registered in the EU, it is clear that the pesticide industry is unable to offer enough replacements for the products which are being withdrawn from the market (Thompson et al., 2008).

6. Acaricides and integrated control of phytophagous mites

Biological control of phytophagous mites by predatory mites (Phytoseiidae) and other predators proved to be a successful alternative to conventional chemical control, especially on greenhouse crops (Gerson & Weintraub, 2007). In spite of undoubtful advantages, biological control includes significant limitations as well (Gerson et al., 2003), which makes the use of acaricides still indispensable. In modern crop protection, these acaricides should be biorational compounds: highly effective against mite pests and relatively safe to their predators (i.e. selective), with low risk to human health and the environment. Biorational acaricides are important element of integrated control of phytophagous mites which is based on combination of chemical, biological and other control measures.

Therefore, it is very important to study the effects of acaricides and other pesticides on phytoseiid mites, other predatory mites and insect predators of phytophagous mites. Predators come into contact with pesticides if treated with them directly or exposed to their residues, if they feed on contaminated prey or pollen. Beside lethal effects (mortality), pesticides also cause a variety of sublethal effects, by changing the biological parameters and/or behaviour of survivors (Blűmel et al., 1999; Desneux et al., 2007). International Organization for Biological Control/Western Palearctic Regional Section (IOBC/WPRS) offered one of the most comprehensive programs to test lethal and sublethal side-effects of pesticides on beneficial organism, which comprise laboratory, semi-field and field trials. IOBC/WPRS working group "Pesticides and beneficial organisms" organized and carried out several joint testing programs for most of the predators, parazitoids and other beneficial organisms, including phytoseiid mites (Blűmel et al., 1999; Blűmel & Hausdorf, 2002). However, some methodological solutions within the IOBC procedures (way of exposure, choice of doses/concentrations, evaluation criteria) have been criticized as insufficiently realistic (Bakker & Jacas, 1995; Amano & Haseeb, 2001). In order to acquire an in-depth knowledge on sublethal effects of pesticides on biological control agents, the population level-toxicity approach was proposed; it is based on creation of life tables and calculation of population growth parameters, and/or projection of population growth rate based on matrix model (Stark & Banks, 2003; Stark et al., 2007).

The most frequently encountered on the lists of non-selective active substances are organochlorines, organophosphates, carbamates, pyrethroids and other broad-spectrum insecto-acaricides, which are *per definitionem* toxic to large number of insect and mite species, including Phytoseiidae and majority of other arthropods, predators of phytophagous mites (Croft & Brown, 1975; Knowles, 1997; Blűmel et al., 1999; Gerson et al., 2003). On the other side, abamectin and milbemectin, which are also broad-spectrum insecto-acaricides, are considered safe to beneficial arthropods under field conditions due to their short environmental persistance, rapid uptake into treated plants and fast degradation of surface residues (Krämer & Schirmer, 2007). Although beneficials may be killed when treated directly by spray oils or exposed to the vapor phase of essential oils, their short-term residual activity does not severely affect populations of phytoseiid mites and other predators (Chueca et al., 2010; Han et al., 2010).

It should be noted that certain fungicides (benomyl, dithocarbamates) are partly harmful to predatory mites (Blűmel et al., 1999; Gerson et al., 2003; Alston & Thomson, 2004). Sulfur, acaro-fungicide approved in organic farming (Zehnder et al., 2007; EU, 2008; Milenkovic et al. 2010) have been identified as disruptive to integrated mite control (Beers et al., 2009). Also, there are records of adverse effects of neonicotinoids (new class of neuroactive insecticides which is in great expansion in the last two decades), on survival and/or fecundity (James, 2003; Duso et al., 2008), predator activity (Poletti et al., 2007) and population growth (Stavrinides & Mills, 2009) of phytoseiid mites.

Specific acaricides are considered harmless to majority of predatory insects, while their toxicity to various development stages of the same mite species, and to different mite species, varies to a certain extent. From the standpoint of selectivity, it is essential to be aware of the comparative toxicity of acaricides to phytophagous mites and predatory mites (Knowles, 1997). Compounds, such as organotins, mite growth inhibitors and regulators, acequinocyl, diafenthiuron, some METI acaricides, bifenazate, spirodiclofen, spiromesifen, are usually graded as selective acaricides, much more toxic to phytophagous mites than to phytoseiid and other predatory mites (Blűmel et al., 1999; Knowles, 1976, Dekeyser, 2005; Krämer & Schirmer, 2007). Spinosad and azadirachtin appear to be compatible with predatory mites (Spollen & Isman, 1996; Williams et al., 2003; Raguraman et al., 2004).

Both positive and negative evaluation results are based on smaller or larger number of experimental data, but they should not be taken as general and final conclusions on (non)selectivity. Besides expected intrinsic differences among predatory species in susceptibility to the same pesticide, the literature provides different, and sometimes even contrasting results on compatibility for the same active substance and the same predatory species, due to different test procedures (applied doses/concentrations, way of treatment and exposure of test organisms, observed parameters, laboratory or field experiments); on the other hand, the results obtained by standardized methods are affected by the product formulation type, origin of test organism (autochthonous population or commercialized strain) and other factors (Blűmel et al., 1993; Duso et al., 2008).

Physiological selectivity, i.e. reduced susceptibility due to pesticide metabolism is the most desired testing result of pesticide effects to phytophagous mite predators. But, the non-selective compounds can be made safer for use by special application technology (Blűmel et al., 1999; van Leeuwen et al., 2005a), by reducing the doses/concentrations (Rhodes et al., 2006), by releasing the predators so that they would be exposed to older residues (Lilly & Campbell, 1999), by using the strain of predators with developed resistance to acaricides and other pesticides (Sato et al., 2007).

Application of selective acaricides (synthetic or natural) with releases of commercialized strains of phytoseiid mites and other predators is a sustainable alternative to an approach based on chemical measures only (Lilly & Campbell, 1999; Rhodes et al., 2006; Sato et al., 2007). According to Kogan (1998), a pest control program reaches the level at which it can be qualified as an integrated pest management (IPM) program only when biorational pesticides (acaricides) and release of predators are integrated with other control tactics, preventive and remedial (crop rotation, host plant resistance, cultural practices, mechanical and physical control measures etc). Higher IPM levels entail transfer from species/population level integration (the control of single species or species complexes), via community level integration (multiple pest categories e.g. insects, mites, pathogens, weeds and their control) to ecosystem level integration (the control of multiple pest impacts within the context of the total cropping system).

World-wide, IPM has become the accepted model for crop protection over the past decades, but the adoption of IPM programs has been generally slow in both the developed and the developing countries (Peshin et al., 2009). In the European Union, Directive 91/414 encourages Member States to take the principles of IPM into account, but the implementation is voluntary (Freier & Boller, 2009). Success of IPM has often been measured by the reduction in pesticide usage, which is not necessarily a reliable indicator (Kogan, 1998). Transition from conventional pest control to IPM actually changes the role of pesticides (acaricides) in modern crop protection: within the principles of IPM, pesticides are applied highly rationally and in interaction with other control tactics.

7. References

- Alston, D.G. & Thomson, S.V. (2004). Effects of fungicide residues on the survival, fecundity and predation of the mites *Tetranychus urticae* (Acari: Tetranychidae) and *Galendromus occidentalis* (Acari: Phytoseiidae). *Journal of Economic Entomology*, 97, 3, 950-956, ISSN 0022-1493
- Amano, H & Haseeb, M. (2001). Recently-proposed methods and concepts of testing the effects of pesticides on the beneficial mite and insect species: study limitations and implications in IPM. *Applied Entomology and Zoology*, 36, 1, 1-11, ISSN 0003-6862
- Agnello, A.; Reissig, W.H. & Harris, T. (1994). Management of summer populations of European red mite (Acari: Tetranychidae) on apple with horticultural oil. *Journal of Economic Entomology*, 87, 1 148-161, ISSN 0022-1493
- Aslan, I.; Özbek, H.; Çalmaşur, Ö. & Şahin, F. (2004). Toxicity of essential oil vapours to two greenhouse pests, *Tetranychus urticae* Koch and *Bemisia tabaci* Genn. *Industrial Crops and Products*, 19, 2, 167-173, ISSN 0296-6690
- Auger, P.; Bonafos, R.; Guichou, S. & Kreiter, S. (2004). Resistance to fenazaquin and tebufenpyrad in *Panonychus ulmi* Koch (Acari: Tetranychidae) populations from South of France apple orchards. *Crop Protection*, 22, 8, 1039-1044, ISSN 0261-2194
- Ay, R. & Gűrkan, M.O. (2005). Resistance to bifenthrin and resistance mechanisms of different strains of the two-spotted spider mite (*Tetranychus urticae*) from Turkey. *Phytoparasitica*, 33, br, 237-244, ISSN 0334-2123
- Bakker, F.M. & Jacas, J.A. (1995). Pesticides and phytoseiid mites strategies for risk assessment. *Ecotoxicology and Environmental Safety*, 32, 1, 58-67, ISSN 0147-6513
- Balderacchi, M. & Trevisan, M. (2010). Comments on pesticide risk assessment by the revision of Directive EU 91/414. *Environmental Science and Polllution Research*, 17, 3, 523-528, ISSN 0944-1344
- Beers, E.H.; Martinez-Rocha, L.; Talley, R.R. & Dunley, J.E. (2009). Lethal, sublethal, and behavioral effects of sulfur-containing products in bioassays of three species of orchard mites. *Journal of Economic Entomology*, 102, 1, 324-335, ISSN 0022-1493
- Bielza, P.; Denholm, I.; Ioannidis, P.; Sterk, G.; Leadbeater, A.; Leonard, P. & Jørgensen, L.N. (2008). Declaration of Ljubljana the impact of a declining European pesticide Portfolio on resistance management. *Outlooks on Pest Management*, 19, 6, 246-248, ISSN 1743-1026
- Blümel, S. & Hausdorf, H. (2002). Results of the 8th and 9th IOBC joint pesticide testing programme: Persistance test with *Phytoseiulus persimilis* Athias Henriot (Acari: Phytoseiidae). *IOBC/WPRS Bulletin* 25, 11 43-51, ISSN 92-9067-148-X

- Blümel, S.; Baker, F. & Grove, A. (1993). Evaluation of different methods to assess the side-effects of pesticides on *Phytoseiulus persimilis* A.-H. *Experimental and Applied Acarology*, 17, 3, 161-169, ISSN 1572-9702
- Blümel, S.; Matthews, G.A.; Grinstein, A. & Elad, Y.(1999). Pesticides in IPM: selectivity, side-effects, application and resistance problems. In: *Integrated Pest and Disease Management in Greenhouse Crops*, Albajes, R.; Gullino, M.A.; van Lenteren, J.C. & Elad, Y. (ed.), 150-167, Kluwer Academic Publishers, ISBN 0-7923-5631-4, Dordrecht, the Netherlands
- Brück, E.; Elbert, A.; Fischer, R.; Krueger, S.; Kühnhold, J.; Klueken A.M.; Nauen, R.; Niebes, J.F.; Reckman, U.; Schnorbach, J.J.; Steffens, R. & van Waetermeulen, X. (2009). Movento®, an innovative ambimobile insecticide for sucking insect pest control in agriculture: biological profile and field performance. *Crop Protection*, 28, 10, 838-844, ISSN 0261-2194
- Casida, J.E. & Quistad, G.B. (1998). Golden age of insectificde research: past, present, or future? *Annual Review of Entomology*, 43, 1-16, ISSN 0066-4170
- Chandler, D.; Davidson, G.; Pell, J.K.; Ball, B.V.; Shaw, K. & Sunderland, K.D. (2000). Fungal biocontrol of Acari. *Biocontrol Science and Technology*, 10, 3, 357-384, ISSN 0958-3157
- Chueca, P.; Garcera, C.; Molto, E.; Jacas, J.A.; Urbaneja, A. & Pina, T. (2010). Spray deposition and efficacy of four petroleum-derived oils used against *Tetranychus urticae*. *Journal of Economic Entomology*, 103, 2, 386-393, ISSN 0022-1493
- Choi, W.I.; Lee, S.G.; Park, H.M. & Ahn, Y.J. (2004). Toxicity of plant essential oils to *Tetranychus urticae* (Acari: Tetranychidae) and *Phytoseiulus persimilis* (Acari: Phytoseiidae). *Journal of Economic Entomology*, 97, 2, 553-558, ISSN 0022-1493
- Collins, D.A. (2006). A review of alternatives to organophosphorus compounds for the control of storage mites. *Journal of Stored Products Research*, 42, 4, 395-426, ISSN 0022-474X
- Copping, L.G. & Duke, S.O. (2007). Natural products that have been used commercially as crop protection agents a review. *Pest Management Science*, 63, 6, 524-554, ISSN 1526-498X
- Cranham, J.E. & Helle W. (1985). Pesticide Resistance in Tetranychidae. In: *Spider Mites: Their Biology, Natural Enemies and Control*, Vol. 1B, Helle, W. & Sabelis, M.W. (ed.), 405-421, Elsevier, ISBN 0-444-42374-5, Amsterdam, the Netherlands
- Croft, B.A. & van de Baan, H.E. (1988). Ecological and genetic factors influencing evolution of pesticide resistance in tetranychid and phytoseiid mites. *Experimental and Applied Acarology*, 4, 3, 277-300, ISSN 1572-9702
- Croft, B.A. & Brown, A.W.A. (1975). Responses of arthropod natural enemies to insecticides. *Annual Review of Entomology*, 20, 285-335, ISSN 0066-4170
- Dayan, F.E.; Cantrell, C.L. & Duke, S.O. (2009). Natural products in crop protection. *Bioorganic and Medicinal Chemistry*, 17, 12, 4022-4034, ISSN 0968-0896
- Dekeyser, M.A. (2005). Acaricide mode of action. *Pest Management Science*, 61, 2, 103-110, ISSN 1526-498X
- Desneux, N.; Decourtye, A. & Delpuech, J.M. (2007). The sublethal effects of pesticides on beneficial arthropods. *Annual Review of Entomology*, 52, 81-106, ISSN 0066-4170
- Duso, C.; Malagnini, V.; Pozzebon, A.; Castagnoli, M.; Liguori, M. & Simoni, S. (2008). Comparative toxicity of botanical and reduced-risk insecticides to Mediterranean

- populations of *Tetranychus urticae* and *Phytoseiulus persimilis* (Acari: Tetranychidae, Phytoseiidae). *Biological Control* 47, 1, 16-21, ISSN 1049-9644
- Dutcher, J.D. (2007). A review of resurgence and replacement causing pest outbreaks in IPM, In: *General Concepts in Integrated Pest and Disease Management*, Ciancio, A. & Mukerji, K.G. (ed.), 27-43, Springer, ISBN 978-1-4020-6060-1, Dordrecht, the Netherlands
- EPA (2009). Reduced Risk/Organophosphate Alternative Decisions for Conventional Pesticides. www.epa.gov/opprd001/workplan/completionsportrait.pdf
- EPA (2010). Implementation of Requirements under the FQPA.

 www.epa.gov/pesticides/regulating/laws/fqpa/fqpa_implementation.htm
 (Accessed July 31, 2010)
- EU (2008). Comission Regulation (EC) No 889/2008. Official Journal of the European Union, 51, L 250, ISSN 1725-2555
- EU (2010). EU Pesticide Database: http://ec.europa.eu/sanco_pesticides/public/index.cfm (Accessed July 31, 2010)
- Faria, M.R. & Wraight, S.P. (2007). Mycoinsecticides and mycoacaricides: a comprehensive list with worldwide coverage and international classification of formulation types. *Biological Control*, 43, 3, 237-256, ISSN 1049-9644
- Freier, B. & Boller, E.F. (2009). Integrated Pest Management in Europe History, Policy, Achievements and Implementation. In: *Integrated Pest Management: Dissemination and Impact*, Peshin, R. & Dhawan, A.K. (ed.), 435- 454, Springer Science+Business Media B.V. ISBN 978-1-4020-8989-3, Dordrecht, the Netherlands
- Gerson, U. & Cohen, E. (1989). Resurgences of spider mites (Acari: Tetranychidae) induced by synthetic pyretrhroids. *Experimental and Applied Acarology*, 6, 1, 29-46, ISSN 1572-9702
- Gerson, U.; Smiley, R.L. & Ochoa, R. (2003). Mites (Acari) for Pest Control. Blackwell Science Ltd., ISBN 978-0-632-05658-3, Oxford, UK
- Gerson, U. & Weintraub, P.G. (2007). Mites for the control of pests in protected cultivation. *Pest Management Science*, 63, 7, 658-676, ISSN 1526-498X
- Goka, K. (1998). Mode of inheritance of resistance to three new acaricides in the Kanzawa spider mite, *Tetranychus kanzawai* Kishida (Acari: Tetranychidae). *Experimental and Applied Acarology*, 22, 12, 699-708, ISSN 1572-9702
- Hajek, A.E. & Delalibera Jr., I. (2010). Fungal pathogens as classical biological control agents against arthropods. *BioControl*, 55, 1, 147-158, ISSN 1386-6141
- Han, J.; Choi, B.R.; Lee, S.G.; Kim, S.I. & Ahn, Y.J. (2010). Toxicity of plant essential oils to acaricide-susceptible and -resistant *Tetranychus urticae* (Acari: Tetranychidae) and *Neoseiulus californicus* / Acari: Phytoseiidae). *Journal of Economic Entomology*, 103, 4, 1293-1298, ISSN 0022-0493
- Hardin, M.R.; Benrey, B.; Coll, M.; Lamp, W.O.; Roderick, G.K. & Barbosa, P. (1995). Arthropod pest resurgence: an overview of potential mechanisms. *Crop Protection*, 14, 1, 3-18, ISSN 0261-2194
- Helle, W. & Sabelis, M.W. (1985a). *Spider Mites: Their Biology, Natural Enemies and Control, Vol. 1A*, Elsevier, ISBN 0-444-42372-9, Amsterdam, the Netherlands
- Helle, W. & Sabelis, M.W. (1985b). *Spider Mites: Their Biology, Natural Enemies and Control, Vol. 1B*, Elsevier, ISBN 0-444-42374-5, Amsterdam, the Netherlands

- Herron, G.A.; Edge, V.E.; Wilson, L.J. & Rophail, J. (1998). Organophosphate resistance in spider mites (Acari: Tetranychidae) from cotton in Australia. *Experimental and Applied Acarology*, 22, 1, 17-30, ISSN 1572-9702
- Herron, G.A.; Rophail, J. & Wilson, L.J. (2001). The development of bifenthrin resistance in two-spotted spider mite (Acari: Tetranychidae) from Australina cotton. *Experimental and Applied Acarology*, 25, 4, 301-310, ISSN 1572-9702
- Horowitz, A.R.; Ellsworth, P.C. & Ishaaya, I. (2009). Biorational Pest Control An Overview, In: *Biorational Control of Arthropod Pests*, Ishaaya, I. & Horowitz, A.R. (ed.), 1-20, Springer, ISBN 978-90-481-2315-5, Dordrecht, the Netherlands
- Huffaker, C.B.; van de Vrie, M. & McMurtry, J.A. (1970). Ecology of tetranychid mites and their natural enemies: a review. II. Tetranychid populations and their possible control by predators: an evaluation. *Hilgardia*, 40, 391-458, ISSN 0073-2230
- Ishaaya, I. (2001). Biochemical Sites of Insecticide Action and Resistance. Springer-Verlag, ISBN 3-540-67625-2, Berlin, Heidelberg and New York
- Isman, M.B. (2006). Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world. *Annual Review of Entomology*, 51, 45-66, ISSN 0066-4170
- Isman, M.B. & Akhtar, Y. (2007). Plant Natural Products as a Source for Developing Environmentaly Acceptable Insecticides. In: *Insecticide Design Using Advanced Technologies*, Ishaaya I.; Nauen R. & Horowitz R. (ed.), 235-248, Springer-Verlag, ISBN 10: 3-540-46904-4, Berlin, Heidelberg
- Jackson, M.A.; Dunlap, C.A. & Jaronski, S.T. (2010). Ecological considerations in producing and formulating fungal entomopathogens for use in insect biocontrol. *BioControl*, 55, 1, 129-145, ISSN 1386-6141
- James, D.G. (2003). Toxicity of imidacloprid to *Galendromus occidentalis, Neoseiulus fallacis* and *Amblyseius andersoni* (Acari: Phytoseiidae) from hops in Washington State, USA. *Experimental and Applied Acarology*, 31, 3-4, 275-281, ISSN 1572-9702
- Jeppson, L.R.; Keifer, H.H. & Baker, E.W. (1975). *Mites Injurious to Economic Plants*, University of California Press, ISBN 0-520-02381-1, Berkeley and Los Angeles.
- Keena, M.A. & Granett, J. (1990). Genetic analysis of propargite resistance in Pacific spider mites and twospotted spider mites (Acari: Tetranychidae). *Journal of Economic Entomology*, 83, 3, 655-661, ISSN 0022-1493
- Kim, Y.J.; Lee, S.H.; Lee, S.W. & Ahn, Y.J. (2004). Fenpyroximate resistance in *Tetranychus urticae* (Acari: Tetranychidae): cross resistance and biochemical resistance mechanisms. *Pest Management Science*, 60, 10, 1001-1006, 1526-498X
- Kim, Y.J.; Park, H.M.; Cho, J.R. & Ahn, Y.J. (2006). Multiple resistance and biochemical mechanisms of pyridaben resistance in *Tetranychus urticae* (Acari: Tetranychidae). *Journal of Economic Entomology*, 99, 3, 954-958, ISSN 0022-1493
- Kim, S.S. & Yoo, S. (2002). Comparative toxicity of some acaricides to the predatory mite, *Phytoseiulus persimilis* and the twospotted spider mite, *Tetranychus urticae*. *BioControl*, 47, 5, 563-573, ISSN 1386-6141
- Kiss, J.; Szendrey, L.; Schlosser, E. & Kotlar, I. (1996). Application of natural oil in IPM of grapevine with special regard to predatory mites. *Journal of Environmental Science and Health, Part B*, 31, 5, 421-425, ISSN 0360-1234
- Khajehali, J.; van Leeuwen, T.; Grispou, M.; Morou, E.; Alout, H.; Weill, M.; Tirry, L.; Vontas, J. & Tsagkarakou, A. (2010), Acetylcholinesterase point mutations in European

- strains of *Tetranychus urticae* (Acari: Tetranychidae) resistant to organophosphates. *Pest Management Science*, 66, 2, 220-228, 1526-498X
- Kleeberg, H. (2004). Neem based products registration requirements, regulatory processes and global implications. In: *Neem, Today and in The New Millenium*, Koul O. & Wahab S. (ed.), 109-123, Kluwer Academic Publishers, ISBN-10: 1402012292, Dordrecht, the Netherlands
- Knowles, C.O. (1976). Chemistry and toxicology of quinoxaline, organotin, organofluorine, and formamidine acaricides. *Environmental Health Perspectives*, 14, 93-102, ISSN 0091-6765
- Knowles, C.O. (1997). Mechanisms of resistance to acaricides, In: *Molecular Mechanisms of Resistance to Agrochemicals*, Sjut, V. & Butters, J.A. (ed.), 58-78, Springer Verlag, ISBN 3-540-62461-9, Berlin Heidelberg
- Kogan, M. (1998). Integrated pest management historical perspectives and contemporary developments. *Annual Review of Entomology*, 43, 243-270, ISSN 0066-4170
- Kontsedalov, S.; Gottlieb, Y.; Ishaaya, I.; Nauen, R.; Horowitz, R. & Ghanim, M. (2008). Toxicity of spiromesifen to the developmental stages of *Bemisia tabaci* biotype B. *Pest Management Science*, 65, 1, 5-13, ISSN 1526-498X
- Krämer, W. & Schirmer, U. (2007). *Modern Crop Protection Compounds, Vol. 3,* WILEY VCH Verlag GmbH & Co., ISBN 978-3-527-31496-6, Weinheim, Germany
- Kumral, N.A.; Susurluk, H.; Gencer N.S. & Gűrkan, M.O. (2009). Resistance to chlorpyrifos and lambda- cyhalothrin along with detoxifying enzyme activities in field collected female populations of European red mite. *Phytoparasitica*, 37, 1, 1-15, ISSN 0334-2123
- Kwon, D.H.; Im, J.S.; Ahn, J.J.; Lee, J.H.; Clark, J.M. & Lee, S.H. (2010a). Acetylcholinesterase point mutations putatively associated with monocrotophos resistance in the two-spotted spider mite. *Pesticide Bioochemistry and Physiology*, 96, 1, 36-42, ISSN 0048-3575
- Kwon, D.H.; Clark, J.M. & Lee, S.H. (2010b). Cloning of a sodimu channel gene and identification of mutations putatively associated with fenpropathrin resistance in *Tetranychus urticae*. *Pesticide Bioochemistry and Physiology*, 97, 2, 93-100, ISSN 0048-3575
- Lancaster, A.L.; Deyton, D.E.; Sams, C.E.; Cummins, J.C.; Pless, C.D. & Fare, D.C. (2002). Soybean oil controls two-spotted spider mites on burning bush. *Journal of Environmental Horticulture*, 20, 2, 86-92, ISSN 0738-2898
- Li, M.; Liu, C.L.; Li, L.; Yang, H.; Li, Z.N.; Zhang, H. & Li, Z.M. (2010). Design, synthesis and biological activities of new strobilurin derrivatives containing substituted pyrazoles. *Pest Management Science*, 66, 1, 107-112, ISSN 1526-498X
- Lilly, R. & Campbell, C.A.M. (1999). Biological, chamical and integrated control of two-spotted spider mite *Tetranychus urticae* on dwarf hops. *Biocontrol Science and Technology*, 9, 4, 467-473, ISSN 0958-3157
- Lindquist, E.E.; Sabelis, M.W. & Bruin, J. (1996). *Eriophyoid Mites: Their Biology, Natural Enemies and Control*, Elsevier, ISBN 0-444-88628-1, Amsterdam, the Netherlands
- Lűmmen, P. (2007). Mitochondrial electron transport complexes as biochemical target sites for insecticides and acaricides, In: *Insecticide Design Using Advanced Technologies*, Ishaaya, I.; Nauen, R. & Horowitz, R. (ed.), 197-215, Springer-Verlag, ISBN 10: 3-540-46904-4, Berlin Heidelberg

- Maniania, N.K.; Bugeme, D.M.; Wekesa, V.W.; Delalibera, I.Jr. & Knapp, M. (2008). Role of entomopathogenic fungi in the control of *Tetranychus evansi* and *Tetranychus urticae* (Acari:Tetranychidae), pests of horticultural crops. *Experimental and Applied Acarology*, 46, 1-4, 259-274, ISSN 1572-9702
- Mansour, F.A.; Ascher, K.R.S. & Abo-Moch, F. (1997). Effects of Neemgard on phytophagous and predacious mites and on spiders. *Phytoparasitica*, 25, 4, 333-336, ISSN 0334-2123
- Marčić, D. (2003). The effects of clofentezine on life-table parameters in two-spotted spider mite *Tetranychus urticae*. *Experimental and Applied Acarology*, 30, 3, 249-263, ISSN 1572-9702
- Marčić, D. (2007). Sublethal effects of spirodiclofen on life history and life-table parameters of two-spotted spider mite (*Tetranychus urticae*). *Experimental and Applied Acarology*, 42, 2, 121-129, ISSN 1572-9702
- Marčić, D.; Ogurlić, I.; Mutavdžić, S. & Perić, P. (2010). The effects of spiromesifen on life history traits and population growth of two-spotted spider mite (Acari: Tetranychidae). *Experimental and Applied Acarology*, 50, 3, 255-267, ISSN 1572-9702
- Marčić, D.; Perić, P.; Prijović, M.; Ogurlić, I. & Andrić, G. (2007). Effectiveness of spirodiclofen in the control of European red mite (*Panonychus ulmi*) on apple and pear psylla (*Cacopsylla pyri*). *Pesticides and Phytomedicine*, 22, 3, 301-309, ISSN 1820-3949
- Marčić, D.; Perić, P.; Prijović, M. & Ogurlić, I. (2009). Field and greenhouse evaluation of rapeseed spray oil against spider mites (Acari: Tetranychidae), green peach aphid (Homoptera: Aphididae) and pear psylla (Hemiptera: Psyllidae) in Serbia. *Bulletin of Insectology*, 62, 2, 159-167, ISSN 1721-8861.
- March, R.B. (1976). Properties and actions of bridged diphenyl acaricides. *Environmental Health Perspectives*, 14, 83-91, ISSN 0091-6765
- Martinez-Villar, E.; Saenz-de-Cabezon, F.J.; Moreno-Grijalba, F.; Marco, V. & Perez-Moreno, I. (2005). Effects of azadirachtin on the two-spotted spider mite, *Tetranychus urticae* (Acari: Tetranychidae). *Experimental and Applied Acarology*, 35, 3, 215-222, ISSN 1572-9702
- McMurtry, J.A.; Huffaker, C.B. & van de Vrie, M. (1970). Ecology of tetranychid mites and their natural enemies: a review. I. Tetranychid enemies, their biological characters and the impact of spray practices. *Hilgardia*, 40, 331-370, ISSN 0073-2230
- Messing, R.H. & Croft, B.A. (1996). Pesticide Resistance in Eriophyoid Mites, Their Competitors and Predators. In: *Eriophyoid Mites Their Biology, Natural Enemies and Control,* Lindquist, E.E.; Sabelis, M.W. & Bruin, J. (ed.), 695-726, Elsevier, ISBN 0-444-88628-1, Amsterdam, the Netherlands
- Metcalf, R.L. (1980). Changing role of insecticides in crop protection. *Annual Review of Entomology*, 25, 219-256, ISSN 0066-4170
- Milenković, S.; Tanasković, S. & Lazić, T. (2005). [Azadirachtin the possibilities of use in plant protection]. *Voćarstvo*, 39, 1, 61-69, ISSN 1820-5054 (in Serbian)
- Milenković, S.; Tanasković, S. & Sretenović, D. (2010). Organic raspberry production in Serbia, Proceedings of VII International Conference on Integrated Fruit Production, (IOBC/WPRS Bulletin, Vol. 54), pp. 413-416, ISBN 978-92-9067-213-5, Avignon, France, October 2008, IOBC/WPRS
- Miresmailli, S.; Bradbury, R. & Isman, M.B. (2006). Comparative toxicity of *Rosmarinus officinalis* L. essential oil and blends of its major constituents against *Tetranychus*

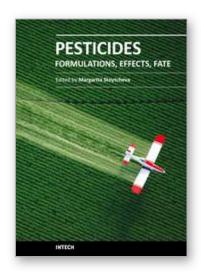
- urticae Koch (Acari: Tetranychidae) on two different host plants. *Pest Management Science*, 62, 4, 366-371, ISSN 1526-498X
- Mizutani, A.; Kumayama, F.; Ohra, K.; Ishiguro, T. & Hayashi, Y. (1988). Inheritance of resistance to cyhexatin in the Kanzawa spider mite, *Tetranychus kanzawai* Kishida (Acari: Tetranychidae). *Applied Entomology and Zoology*, 23, 3, 251-255, ISSN 0003-6862
- Moran, R.E.; Deyton, D.E.; Sams, C.E.; Pless, C.D. & Cummins, J.C. (2003). Soybean oil as a summer spray for apple: European red mite control, net CO₂ assimilation and phytotoxicity. *HortEcience*, 38, 2, 234-238, ISSN 0018-5345
- Najar-Rodriguez, A.J.; Lavidis, N.A.; Mensah, R.K.; Choy, P.T. & Walter, G.H. (2008). The toxicological effects of petroleum spray oils on insects evidence for an alternative mode of action and possible new control options. *Food and Chemical Toxicology*, 46, 9, 3003-3014, ISSN 0278-6915
- Nauen, R. & Smagghe, G. (2006). Mode of action of etoxazole. *Pest Management Science*, 62, 5, 379-382, ISSN 1526-498X
- Nicetic, O.; Watson, D.M.; Beattie, G.A.C.; Meats, A. & Zheng, J. (2001). Integrated pest management of two-spotted mite *Tetranychus urticae* on greenhouse roses using petroleum spray oil and the predatory mite *Phytoseiulus persimilis*. *Experimental and Applied Acarology*, 25, 1, 37-53, ISSN 1572-9702
- Ochiai, N.; Mizuno, M.; Mimori, N.; Miyake, T.; Dekeyser, M.; Canlas, L.J. & Takeda, M. (2007). Toxicity of bifenazate and its principal active metabolite, diazene, to *Tetranychus urticae* and *Panonychus citri* and their relative toxicity to the predaceous mites, *Phytoseiulus persimilis* and *Neoseiulus californicus*. *Experimental and Applied Acarology*, 43, 3, 181-197, ISSN 1572-9702
- Omoto, C.; Dennehy, T.J.; McCoy, C.W.; Crane, S.E. & Long, J.W. (1994). Detection and characterisation of the interpopulation variation of citrus rust mite resistance to dicofol in Florida citrus. *Journal of Economic Entomology*, 87, 3, 566-572, ISSN 0022-1493
- Peshin, R.; Bandral, R.S.; Zhang, W.J.; Wilson, L. & Dhawan, A. (2009). Integrated Pest Management: a Global Overview of History, Programs and Adoption. In: *Integrated Pest Management: Innnovation-Development Process*, Peshin, R & Dhawan, A.K. (ed.), 1-49, , Springer, ISBN 978-1-4020-8991-6, Dordrecht, the Netherlands
- Poletti, M.; Maia, A.H.N. & Omoto, C. (2007). Toxicity of neonicotinoid insecticides to Neoseiulus californicus and Phytoseiulus macropilis (Acari: Phytoseiidae) and their impact on functional response to Tetranychus urticae (Acari: Tetranychidae). Biological Control 40, 1, 30-36, ISSN 1049-9644
- Raguraman, S.; Ganapathy, N. & Venkatesan, T. (2004). Neem versus entomopathogens and natural enemies of crop pests: the potential impact and strategies. In: *Neem, Today and in The New Millenium*, Koul O. & Wahab S. (ed.), 109-123, Kluwer Academic Publishers, ISBN-10: 1402012292, Dordrecht, the Netherlands
- Rhodes, E.M.; Liburd, O.E.; Kelts, C.; Rondon, S.I. & Francis, R.R. (2006). Comparison of single and combination treatments of *Phytoseiulus persimilis*, *Neoseiulus californicus* and Acramite (bifenazate) fro control of two-spotted spider mites in strawberries. *Experimental and Applied Acarology*, 39, 3-4, 213-225, ISSN 1572-9702
- Rizzieri, D.A.; Dennehy, T.J. & Glover, T.J. (1988). Genetic analysis of dicofol resistance in two populations of twospotted spider mite (Acari: Tetranychidae) from New York apple orchards. *Journal of Economic Entomology*, 81, 5, 1271-1276, ISSN 0022-1493

- Rock, G.C. & Crabtree, K.W. (1987), Biological activity of petroleum and cottonseed oils against two tetranychid mite species and tortricid insect species found on apple. *Journal of Agricultural Entomology*, 4, 3, 247-253, ISSN 0735-939X
- Roush, R.T. & McKenzie, J.A. (1987). Ecological genetics of insecticide and acaricide resistance. *Annual Review of Entomology*, 32: 361-380, ISSN 0066-4170
- Sato, M.E.; da Silva, M.Z.; de Souza Filho, M.F.; Matioli, A.L. & Raga, A. (2007). Management of *Tetranychus urticae* (Acari: Tetranychidae) in strawberry fields with *Neoseiulus californicus* (Acari: Phytoseiidae) and acaricides. *Experimental and Applied Acarology*, 42, 2, 107-120, ISSN 1572-9702
- Shaaya, E. & Rafaeli, A. (2007). Essential Oils as Biorational Insecticides Potency and Mode of Action. In: *Insecticide Design Using Advanced Technologies*, Ishaaya, I.; Nauen, R. & Horowitz, R. (ed.), 249-261, Springer, ISBN 10: 3-540-46904-4, Berlin Heidelberg
- Spollen, K.M. & Isman, M.B. (1996). Acute and sublethal effects of a neem insecticide on the commercial biological control agents *Phytoseiulus persimilis* and *Amblyseius cucumeris* (Acari: Phytoseiidae) and *Aphidoletes aphidimyza* (Diptera: Cecidomyiidae). *Journal of Economic Entomology*, 89, 6, 1379-1386, ISSN 0022-1493
- Stark, J.D. & Banks, J.E. (2003). Population-level effects of pesticides and other toxicants on arthropods. *Annual Review of Entomology*, 48, 505-519, ISSN 0066-4170
- Stark, J.D.; Sugayama, R.L. & Kovaleski, A. (2007). Why demographic and modeling approaches should be adopted for estimating the effects of pesticides on biocontrol agents. *BioControl*, 52, 3, 365-374, ISSN 1386-6141
- Stavrinides, M.C. & Mills, N.J. (2009). Demographic effects of pesticides on biological control of Pacific spider mite (*Tetranychus pacificus*) by the western predatory mite (*Galendromus occidentalis*). *Biological Control*, 48, 3, 267-273, ISSN 1049-9644
- Stavrinides, M.C.; van Niewenhuyse, P.; van Leeuwen, T. & Mills, N.J. (2010). Development of acaricide resistance in Pacific spider mite (*Tetranychus pacificus*) from California vineyards. *Experimental and Applied Acarology*, 50, 3, 243-254, ISSN 1572-9702
- Stumpf, N. & Nauen, R. (2001). Cross-resistance, inheritance and biochemistry of mitochondrial electron transport inhibitor-acaricide resistance in *Tetranychus urticae* (Acari: Tetranychidae). *Journal of Economic Entomology*, 94, 6, 1577-1583, ISSN 0022-1493
- Stumpf, N.; Zebitz, C.P.W.; Kraus, W.; Moores, G. & Nauen, R. (2001). Resistance to organophosphates and biochemical genotyping of acetylcholinesterases in *Tetranychus urticae* (Acari: Tetranychidae). *Pesticide Biochemistry and Physiology*, 69, 2, 131-142, ISSN 0048-3575
- Sundaram, K.M.S. & Sloane, L. (1995). Effects of pure and formulated azadirachtin, a neembased biopesticide, on the phytophagous spider mite, *Tetranychus urticae* Koch. *Journal of Environmental Science and Health (B)*, 30, 6, 801-814, ISSN 0360-1234
- Taverner, P. (2002). Drowning or just waving? A perspective on the ways petroleum-derrved oils kill arthropod pests of plants. In: *Spray Oils beyond 2000 Sustainable Pests and Diseases Management*, Beattie, G.A.C.; Watson, D.M.; Stevens, M.L.; Rae, D.J. & Spooner-Hart, R.N. (ed.)., 78-88, University of Western Sydney, ISBN 0-7341-0205-4, Sydney.
- Thompson, G.D.; Matten, S.; Denholm, I.; Whalon, M.E. & Leonard, P. (2008). The Politics of Resistance Management: Working Towards Pesticide Resistance Management Globally. In: *Global Pesticide Resistance in Arthropods*, Whalon, M.E.; Mota-Sanchez,

- D. & Hollingworth, R.M. (ed.), 146-165, CAB International, ISBN 978-1-84593-353-1, Wallingford, UK
- Tsagkarakou, A.; Navajas, M.; Cuany, A.; Chevillon, C. & Pasteur, N. (2002). Mechanisms of resistance to organophosphates in *Tetranychus urticae* (Acari: Tetranychidae) from Greece. *Insect Biochemistry and Molecular Biology*, 32, 4, 417-424, ISSN 0965-1748
- Tsagkarakou, A.; van Leeuwen, T.; Khajehali, J.; Ilias, A.; Grispou, M.; Williamson, M.S.; Tirry, L. & Vontas, J. (2009). Identification of pyrethroid resistance mutations in the *para* sodium channel of the twospotted spider mite *Tetranychus urticae* (Acari: Tetranychidae). *Insect Molecular Biology*, 18, 5, 583-593, ISSN 0962-1075
- Uesugi, R.; Goka, K. & Osakabe M. (2002). Genetic basis of resistance to chlorfenapyr and etoxazole in the two-spotted spider mite (Acari: Tetranychidae). *Journal of Economic Entomology*, 95, 6, 1267-1274, ISSN 0022-1493
- van Leeuwen, T. & Tirry. L. (2007). Esterase-mediated bifenthrin resistance in a multiresistant strain of the two-spotted spider mite, *Tetranychus urticae*. *Pest Management Science*, 63, 2, 150-156, ISSN 1526-498X
- van Leeuwen, T.; Dermauw, W.; van de Veire, M. & Tirry, L. (2005a). Systemic use of spinosad to control the two-spotted spider mite (Acari: Tetranychidae) on tomatoes grown in rockwool. *Experimental and Applied Acarology*, 37, 1, 93-105, ISSN 1572-9702
- van Leeuwen, T.; van Pottelberge, S. & Tirry. L. (2005b). Comparative acaricide susceptibility and detoxifying enzyme activities in field-collected resistant and susceptible strains of *Tetranychus urticae*. *Pest Management Science*, 61, 5, 499-507, 1526-498X
- van Leeuwen, T.; Vanholme, B.; van Pottelberge, S.; van Nieuwenhuyse, P.; Nauen, R.; Tirry, L. & Denholm, I. (2008). Mitochondrial heteroplasmy and the evolution of insecticide resistance: non-Mendelian inheritance in action. *Proceedings of the National Academy of Science*, 105, 16, 5980-5985, ISSN 0027-8424
- van Leeuwen, T.; van Pottelberge, S. & Tirry, L. (2007). Organophosphate insecticides and acaricides antagonise bifenazate toxicity through esterase inhibition in *Tetranychus urticae*. *Pest Management Science*, 63, 12, 1172-1177, ISSN 1526-498X
- van Leeuwen, T.; Vontas, J.; Tsagkarakou, A. & Tirry. L. (2009). Mechanisms of Acaricide Resistance in the Two-Spotted Spider Mite *Tetranychus urticae*, In: *Biorational Control of Arthropod Pests*, Ishaaya, I. & Horowitz, A.R. (ed.), 347-393, Springer, ISBN 978-90-481-2315-5, Dordrecht, the Netherlands
- van Leeuwen, T.; Witters, J.; Nauen, R.; Duso, C. and Tirry, L. (2010). The control of eriophyoid mites: state of the art and future challenges. *Experimental and Applied Acarology*, 51, 1-3, 205-224, ISSN 1572-9702
- van Nieuwenhuyse, P.; van Leeuwen, T.; Khajehali, J.; Vanholme, B. & Tirry, L. (2009). Mutations in the mitochondrial cytochrome b of *Tetranychus urticae* Koch (Acari: Tetranychidae) confer cross-resistance between bifenazate and acequinocyl. *Pest Management Science*, 65,4, 404-412, ISSN 1526-498X
- van Pottelberge, S.; van Leeuwen, T.; Nauen, R. & Tirry, L. (2009). Resistance mechanisms to mitochondrial electron transport inhibitors in a field-collected strain of *Tetranychus urticae* Koch (Acari: Tetranychidae). *Bulletin of Entomological Research*, 99, 1, 23-31, ISSN 0007-4853

- Venzon, M.; Rosado, M.C.; Molina-Rugama, A.J.; Duarte, V.S.; Dias, R. & Pallini, A. (2008). Acaricidal efficacy of neem against *Polyphagotarsonemus latus* (Banks) (Acari: Tarsonemidae). *Crop Protection*, 27, 3-5, 869-872, ISSN 0261-2194
- Villanueva, R.T. & Walgenbach, J.F. (2006). Acaricidal properties of spinosad against *Tetranychus urticae* and *Panonychus ulmi* (Acari: Tetranychidae). *Journal of Economic Entomology*, 99, 3, 843-849, ISSN 0022-1493
- Whalon, M.E.; Mota-Sanchez, D. & Hollingworth, R.M. (2008). *Global Pesticide Resistance in Arthropods*, CAB International, ISBN 978-1-84593-353-1, Wallingford, UK
- Whalon, M.E.; Mota-Sanchez, D.; Hollingworth, R.M. & Duynslager, L. (2010). *Arthropod Pesticide Resistance Database*. www.pesticideresistance.com (Accessed: July 21, 2010)
- Williams, T.; Valle, J. & Viňuela, E. (2003). Is the naturally derived insecticide spinosad compatible with insect natural enemies? *Biocontrol Science and Technology*, 13, 5, 459-475, ISSN 0958-3157
- Zehnder, G.; Gurr, G.M.; Kũhne, S.; Wade, M.R.; Wartten, S.D. & Wyas, E. (2007). Arthropod pest management in organic crops. *Annual Review of Entomology*, 52, 57-80 (supplemental material), ISSN 0066-4170
- Zhang, Z.O. (2003). *Mites of Greenhouses, Identification, Biology and Control*, CAB International, ISBN 0-85199-590-X, Wallingford, UK





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This book provides an overview on a large variety of pesticide-related topics, organized in three sections. The first part is dedicated to the "safer" pesticides derived from natural materials, the design and the optimization of pesticides formulations, and the techniques for pesticides application. The second part is intended to demonstrate the agricultural products, environmental and biota pesticides contamination and the impacts of the pesticides presence on the ecosystems. The third part presents current investigations of the naturally occurring pesticides degradation phenomena, the environmental effects of the break down products, and different approaches to pesticides residues treatment. Written by leading experts in their respective areas, the book is highly recommended to the professionals, interested in pesticides issues.

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