Ife Journal of Science vol. 16, no. 3 (2014)

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# INTERACTIONS OF CROSS-LINKED AND UNCROSS-LINKED CHITOSAN HYDROGELS WITH SURFACTANTS FOR BIOMEDICAL APPLICATIONS

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#### ABSTRACT

The swelling equilibrium of Chitosan and sodium tripolyphosphate (NaTPP) cross-linked chitosan hydrogels in aqueous solutions of surfactants differing in structure and hydrophobicity at 25°C is reported. Anionic surfactant sodium dodecylsulfate (SDS), the cationic surfactant hexadecyltrimethylammonium bromide (HTAB) and neutral surfactants Triton X-100 were employed. The surfactants induced abrupt change in the gel volume. The equilibrium swelling ratio first decreased sharply as the concentration of the surfactant increased and remained almost constant up to the critical micelles concentration (CMC) of the surfactants and then increased again as the concentration increased above the CMC of the surfactants used. The equilibrium volume change of hydrogel was significantly increased from HTAB > Triton X-100 > SDS > the mixed SDS/Triton X-100 system. A decrease in equilibrium swelling ratio of the gel in SDS/TX-100 mixtures was observed with an increase in the mole ratio of SDS. The results also revealed that cross-linking with 3% or 5% w/v NaTPP exhibited lower equilibrium swelling values. This swelling study showed that cross-linking density, surfactant type, and their respective concentrations were important parameters that could affect equilibrium swelling of chitosan gel. Structural elucidation of the nanocomposites was monitored using Infra-red Spectroscopy (IR), and X-Ray Diffractography (XRD).

Keywords: Chitosan Hydrogel, Surfactants, Swelling Equilibrium and Hydrophobicity

## **INTRODUCTION**

Considerable attention has been drawn to the use of natural hydrophilic polymers as drug carriers, dialysis membranes and artificial organs from the view point of environmental pollution, biodegradability, safety and cost (Li et al. 1997). A wide range of hydrophilic polymers have been examined for preparing hydrogels and chitosan is one of them. Chitosan is biodegradable and biocompatible. It exhibits bio-adhesive characteristics. Chitosan is a copolymer of glucosamine and N-acetylglucosamine linked by β-1-4 glucosidic bonds obtained by Ndeacetylation of chitin. It is also a polycationic polysaccharide that can form complexes in acidic medium with negatively charged moieties such as sodium tri-polyphosphate, glutaraldehyde, sodium carboxymethylcellulose, alginic acid, glutamate, and citrates but such complexes formed are insoluble in alkaline medium (Remunan-Lopez and Bodmeier, 1997; Mi et al., 1999; Wang et al., 2001; Lu et al., 2007; Ahmet et al., 2010 and Leonida et al., 2011). In recent years, a number of studies have reported the use of chitosan complexes, for instance, in preparing microspheres (Jiang et al., 2010 and Wang et al., 2011), beads (Sezer and Akbuga, 1995; Chiou et al.,

2006 and Bamgbose *et al.*, 2013) and artificial films (Berger *et al.*, 2004; Wan *et al.*, 2004; Rana *et al.*, 2004 and Bamgbose *et al.*, 2012). Cross linking of chitosan are effective and convenient methods of improving the physical and mechanical properties of chitosan for practical applications (Jameela *et al.*, 1994) and considerable work has been done on the characterization and swelling behavior of hydrogel crosslinked with various crosslinking agents. (Karadag and Saraydin, 2002).

Surfactants are employed in most studies because they comprise of hydrophilic, ionic or non-ionic groups bound to a non-polar hydrophobic group (Tuncer and Melike, 2004). It is expected that both the peculiarities of charged groups on the surfactant and the network, and the hydrophobicity of the surfactants tails will influence the equilibrium extent of gel swelling. Interactions between surfactant molecule and a polymer gel network often result in a pronounced volume change of the gel with the surfactant effectively absorbed by the gel (Khokhlov et al., 1992; Kokufuta et al., 1993; Khokhov et al., 1993 and Isogai et al., 1996). Study response of the polymer network to external stimuli such as change in pH, ionic strength and temperature is of

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interest not only for the design of novel sensing, switching, drug delivery and other devices, but also for better understanding of the interaction of polymer system (Ju *et al.*, 2001; Peppas *et al.*, 2000; Hirasa *et al.*, 1991; Martinez-Ruvalcaba *et al.*, 2009 and Ju *et al.*, 2002). It has been demonstrated that polymers swell if they interact with the solvents at all, and the degree of this interaction is significantly determined by the polymer structure, its environment and the degree of cross linking which is an index of porosity.

Although the swelling characteristics of chitosan hydrogel are known in literature (Tuncer and Melike, 2004; Chiou et al., 2006; Ashok and Vikas, 2010 and Bamgbose et al., 2012,), information regarding the interaction of chitosan gel with cationic, anionic and neutral surfactants is a subject of continuing interest. The anionic surfactant sodium dodecylsulfate (SDS), the cationic surfactant hexadecyltrimethylammonium bromide (HTAB) and neutral surfactants Triton X-100 were used. These surfactants were chosen because of their extreme differences in water solubility and in sensitivity to dissolved organic matters. The effects of these surfactants on the equilibrium swelling ratio of chitosan hydrogel were studied in aqueous system as a function of surfactant type and their respective concentrations at room temperature.

As a result of the need to develop a cheap nontoxic, readily available chitosan hydrogel which could be effectively used for controlled release applications in a wide range of concentrations under ambient temperature and pressure and at a moderate dose, the swelling characteristics of cross linked chitosan films were investigated.

#### **MATERIALS AND METHODS**

# Materials

Sodium dodecylsulfate (SDS) with CMC = 8.22 x10<sup>-3</sup> moldm<sup>-3</sup> was purchased from Surechem P r o d u c t L t d E n g l a n d a n d hexadecylmethylammonium bromide (HTAB) with CMC =11.0 x 10<sup>-3</sup> moldm<sup>-3</sup>, Sodium tripolyphosphate (NaTPP), sodium citrate (NaCit) and glacial acetic acid were obtained from Sigma-Aldrich Chemical Co (St Louis, USA). The Triton X-100 (TX-100) with CMC =  $0.228 \times 10^{-3} \text{ moldm}^{-3}$  was a Rieldel-de-Hans product. All chemicals were of analytical grade and used without purification. In all the experiments, double distilled water was used. The CMC of each surfactant was determined by conductometric method using electric conductivity meter DDS-307 made by Jenway at controlled temperature  $25^{\circ}C \pm 1$ .

## **Gel Preparation**

The protocols for the preparation of the various hydrogels were carried out using literature methods of Ahmet et al., 2010. Chitosan produced from lobster shell wastes was offered as flakes from Aldrich Chemical Co. with 85% degree of deacetylation. 4% w/v chitosan was dissolved in 2% v/v acetic acid solution to prepare homogeneous chitosan solution at room temperature with stirring for 20 minutes. The solutions were cast into films in plastic petri dishes and air dried for 24 h. The dry films were immersed in 5% NaOH solution to neutralize acetic acid residues, and then washed with ethanol to remove excess NaOH. After rinsing with excess distilled water, the films were air dried for 24 hrs. Also, chitosan films were prepared by crosslinking with 3% and 5% w/v NaTPP solution of films containing 4% w/v chitosan. These films were washed with water to remove excess NaTPP.

#### Characterization

The XRD measurements were taken with Monochromatic CuK $\alpha$  radiation (wavelength = 1.5406) produced by MD Minidiffractometer model. The IR spectra were recorded on Perkin Elmer IR spectrophotometer in the spectral region of 350 to 4000 cm<sup>-1</sup>.

#### Swelling Equilibrium Measurement

For the swelling experiments, 0.123g dry film with an average thickness of  $16m\mu$  was immersed in 10 ml of aqueous surfactants solution prepared using double distilled water and equilibrated for 24 hrs. Gravimetric method was employed to study the swelling process. The equilibrium swelling ratio of hydrogel, which signifies the expanding and retracting forces between the films at equilibrium, was determined. The weight of the swollen sample was measured after blotting excessive water gently with filter paper. Aqueous surfactants solution were kept at 25°C for 24 hours to allow the film to reach the equilibrium state to examine surfactant composition effects on swelling properties of the film. The percentage swelling (S%) was calculated using the following equation (Uzum and Karadag, 2006);

$$S\% = \left(\frac{W_t - W_o}{W_o}\right) \times 100 \tag{1}$$

Where  $W_t$  is the mass of the swollen film at time t, and  $W_0$  is the mass of the dry film. The surfactants were used in a wide concentration range from the pre-micellar to the post-micellar region.

# **RESULTS AND DISCUSSION**

#### Characterization of Chitosan Hydrogel

The Chitosan hydrogel and NaTPP cross-linked hydrogel were characterized using IR

spectroscopy. The results are presented in Figures (1, 2 & 3). In Figure 1, the spectral band for chitosan appears at 3760 cm<sup>-1</sup> (axial OH group), 3271 cm<sup>-1</sup> (N-H stretch of secondary amine) and 1153-1054 cm<sup>-1</sup> (ether linkage, C-O-C band stretching). However, the peak is more pronounced at 3000 cm<sup>-1</sup> which indicates that the (C-H) peak is more intense in this compound. There are also bands at 1649 cm<sup>-1</sup> (C=O stretch of amide), 1531 cm<sup>-1</sup> (NH angular deformation in CONH plane) and 2360 cm<sup>-1</sup> (CN asymmetric band stretching). The bending vibrations between 1500 and 1000 cm<sup>-1</sup> intensity indicate some interactions of the amino group.

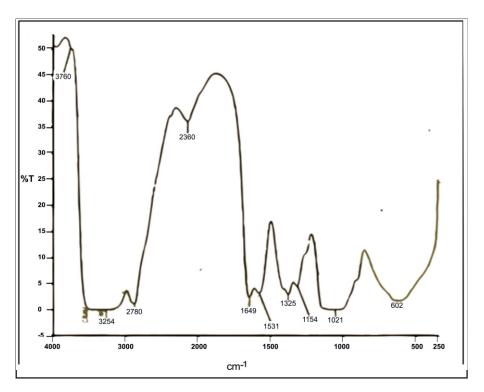


Figure 1: IR spectrum of Chitosan Hydrogel

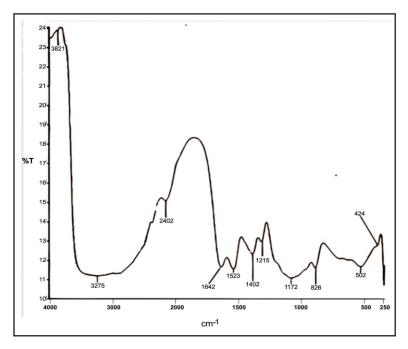


Figure 2: IR Spectrum of 3% NaTPP Cross-linked Chitosan Hydrogels.

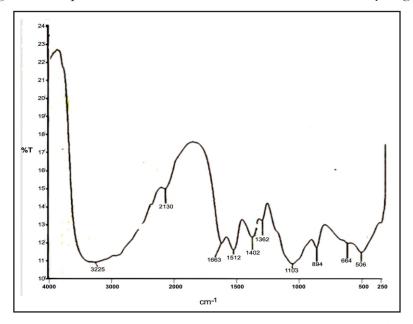


Figure 3: IR Spectrum of 5% NaTPP Cross-linked Chitosan Hydrogels.

The IR spectra of NaTPP cross-linked chitosan hydrogels showed peaks at  $1054 - 1308 \text{ cm}^{-1}$  suggesting the presence of phosphonate linkages between ammonium,  $-NH_3^+$  of chitosan and  $-PO_3^{2-}$  moieties of NaTPP during cross linking process. One peak was observed at 1195 cm<sup>-1</sup> in the film cross linked with 3% w/v NaTPP (Fig. 2) indicating no appreciable linkage. Also, the films cross-linked by 5% w/v, NaTPP showed two peaks, one at 1140 cm<sup>-1</sup> and another at 1279 cm<sup>-1</sup> (Fig. 3), indicating symmetric and asymmetric stretching of phosphonate linkage, respectively

(Ashok and Vikas, 2010). Kemp (1991) also observed that the asymmetric peak is known to occur due to restricted rotation.

The two terminals  $-PO_3^{2-}$  groups of NaTPP molecule appears to be connected with two- $NH_3^+$ ,  $-(CH_3COO^-)$  groups of two chitosan monomers.

Figure 4 (A, B, C, D) presents XRD spectra of the chitosan and chitosan hydrogels. The XRD spectrum of chitosan shows that the chitosan

exhibits diffraction peak at  $2\theta \sim 35^{\circ}$  which is a typical fingerprint of semi-crystalline chitosan. However, the X-ray diffractogram of chitosan film shows that the film has a higher crystallinity than the original chitosan and also has a better amorphous domain which allows better accessibility of the solvent. When the film was exposed to X-ray diffraction analysis, it gives a diffractogram of five peaks characteristics of

crystalline regions at 27.5°, 34°, 39°, 52°, and 68°, respectively. This suggested that the film is a porous membrane. The XRD patterns of a typical NaTPP cross-linked chitosan hydrogels synthesized in this study are almost identical regardless of experimental conditions Figure 4 (C & D), indicates a reflection of high degree of crystallinity.

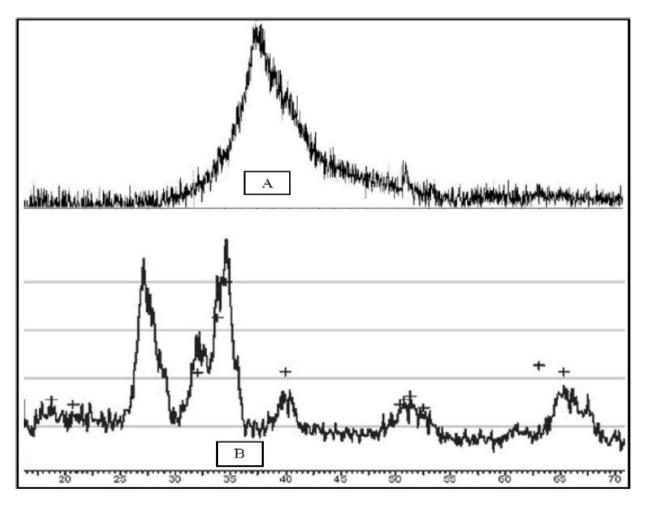
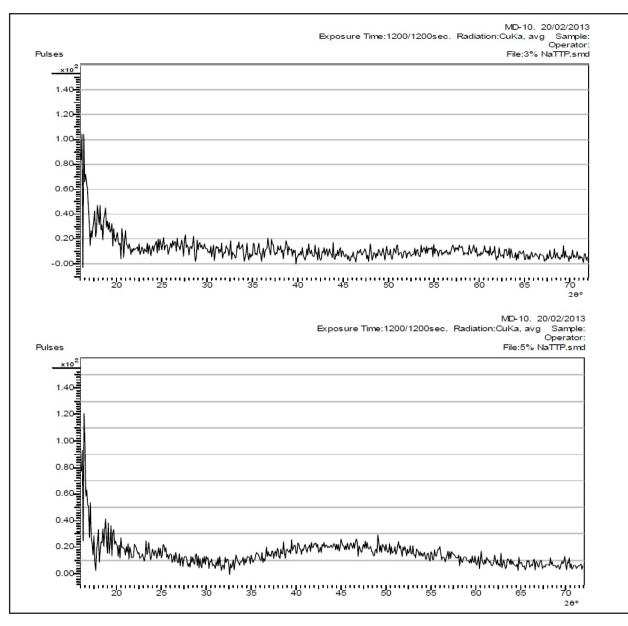


Figure 4: XRD Diffractogram of Chitosan (A) and Chitosan Hydrogel (B)



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Figure 4: XRD Diffractogram of 3% NaTPP Cross-linked Chitosan (C) and 5% NaTPP Cross-linked Chitosan Hydrogel (D)

# Effects of Surfactants on Swelling Equilibrium

Figure 5, shows the percentage equilibrium swelling volume of chitosan hydrogel in anionic surfactant (SDS), cationic surfactant (HTAB) and the neutral surfactant Triton X-100 systems, plotted as a function of the Surfactant solution concentrations below, at and above the CMC of the surfactants. All the surfactants induce abrupt change in the gel volume. The equilibrium swelling ratio first decreased sharply as the concentration of the surfactant increases and remained almost constant up to the CMC of the surfactants. It increased again as the concentration increases above the CMC of the surfactants (Figures 5 & 6). The increase is more at concentrations above the CMC of all the surfactants used in this study. The increase is more at concentrations above the critical micellar concentration (CMC =  $8.22 \times 10^{-3}$  for SDS,  $11.0 \times 10^{-3}$  for HTAB, and  $0.228 \times 10^{-3}$  moldm<sup>-3</sup> for Triton X-100) of all the surfactants. This enhancement may be attributed to the successive micellization of the heterogeneous monomer species as a result of increase in the micellar size and geometry. Surfactants are assumed to exist in monomeric state below CMC. Above CMC, the concentration of monomers remains constant at CMC while the excession.

surfactants leads to the formation of pseudophase micelles in accordance to the conventional. phase- separation model for surfactants in aqueous solution (Daniel and Cary, 1989). As shown in Figure 5, the chitosan hydrogel exhibited re-entrant conformational transitions depending on the concentrations below and above the CMC of the surfactants. The percentage equilibrium volume swelling of chitosan film first decreased with increase in the concentrations of SDS below the CMC of the surfactant from 0.00242 moldm<sup>-3</sup> to 0.00822 moldm<sup>-3</sup> (the CMC of SDS) and then increased again up to 0.0147 moldm<sup>-3</sup>. At concentrations below 0.00822 moldm<sup>-3</sup>, the interactions between the carboxylic acid groups of the chitosan hydrogel and surfactant molecule predominate over the interaction between

polymer chains. This causes the hydrogel to swell. On the other hand the alkyl group in the chain backbone is hydrophobic in nature, these groups are exposed to form aggregates above 0.00713 moldm<sup>-3</sup> concentration to the CMC giving rise to conformational changes. The polymer chains are forced into a globule conformation in which polymer-polymer interactions are dominant. Consequently, the continuous re-entrant phase transition at above the 0.00896 moldm<sup>-3</sup> results from a change in the balance between various types of interactions especially hydrogen bonding and hydrophobic interactions. These trends are clearly observed for the chitosan hydrogel in all other surfactants and in the NaTTP cross-linked chitosan but with lower swelling percentage equilibrium volume values as shown in Figure 7.

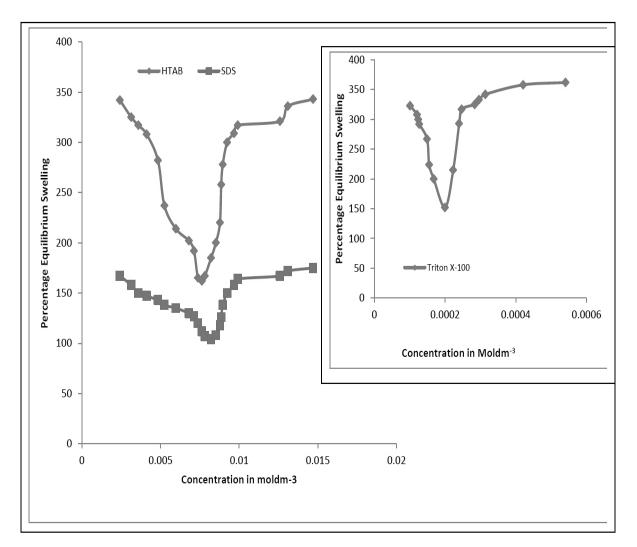
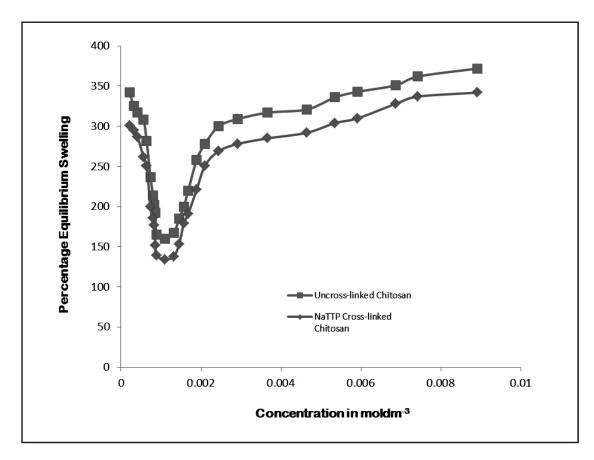


Figure 5: Percentage Equilibrium Swelling as a Function of Surfactant Concentrations. Inset (Triton X-100)



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Figure 6: The Swelling Equilibrium of both Chitosan and NaTTP Cross-linked Chitosan Hydrogels in HTAB

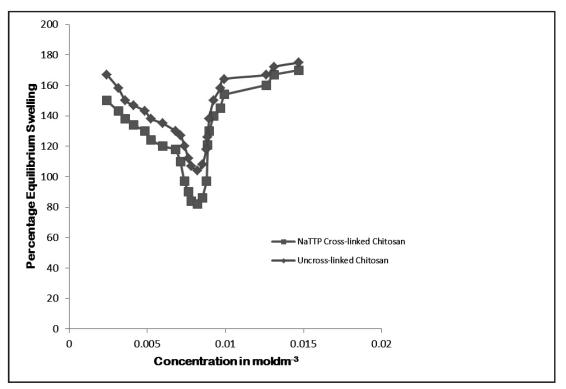


Figure 7: The Swelling Equilibrium of both Chitosan and NaTTP Cross-linked Chitosan Hydrogels in SDS

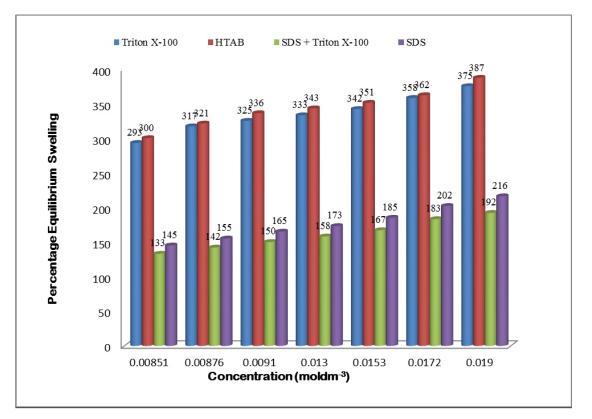


Figure 8: Percentage Equilibrium Swelling as a Function of Surfactant Concentrations

NaTPP dissolved in water and dissociate to give phosphoric ions. The cross-linking of chitosan is known to be dependent on the availability of the cationic sites and the negatively charged species. Swelling of chitosan films, mainly influenced by ionic interactions between chitosan chains, is reported to depend on the cross-linking density achieved during the formation of the network. An increase in cross-linking density is reported to induce a decrease in swelling (Mi *et al.*, 1999; Wang *et al.*, 2001 and Leonida *et al.*, 2011).

Figure 8, also clearly shows that, the percentage equilibrium volume change of chitosan hydrogel increased from HTAB > Triton X-100 > SDS > SDS/TX-100 mixed system. The results also show that the equilibrium swelling ratio of the gel decreases in SDS/TX-100 mixtures with an increase in the mole ratio of SDS from 0.00 to 0.85 and increased sharply as the concentration of the surfactant increases.

# CONCLUSION

The swelling equilibrium of Chitosan and NaTPP cross-linked hydrogels in aqueous solutions of surfactants differing in structure and hydrophobicity has been studied. The chitosan hydrogels were synthesized from a non-toxic, lowcost biocompatible material. All the surfactants induce abrupt change in the gel volume. The percentage equilibrium swelling of the chitosan hydrogel increased sharply as the concentration of the surfactant increases and it is more at concentrations above the critical micellar concentration (CMC) of all the surfactants used in this study. These trends are clearly observed for the chitosan hydrogel in all other surfactants and in the NaTTP cross-linked chitosan but with lower swelling percentage equilibrium volume values. It was also observed that the surfactants induced profoundly different gel swelling profile, and both the structure and the hydrophobicity of the surfactant are the influential factors. This swelling study shows that the cross-linking density, the surfactant types, and their respective concentrations are important parameters affecting the equilibrium swelling ratio of the chitosan gel. As drug release is known to be influenced by cross-linking density, temperature, and pH, these factors can be judiciously used in predicting and modifying controlled drug release.

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# ACKNOWLEDGEMENT

The authors are grateful to the staff of the Central Laboratory of the Federal University of Agriculture Abeokuta, Chemistry Department, University of Ibadan and Center for Energy and Research, Obafemi Awolowo University, Ile-Ife, Nigeria for providing the facilities and some of the chemicals used for this work.

# REFERENCES

- Ahmet M.K., Tshabalala M.A., Buschle-Diller G. 2010. Wood Hemicellulose/Chitosan-Based Semi-Inter-Penetrating Network Hydrogels: Mechanical, Swelling and Controlled Drug Release Properties. J. Bioresour. 5: 1036-1054.
- Ashok K. T. and Vikas R. 2010. Cross-linked Chitosan films: Effect of cross-linking Density on Swelling parameters. *Pak. J. Pharm.* Sci., 23 (4), 443-448
- Bamgbose J.T., Bamigbade A.A., Adewuyi S., Dare E.O., Lasisi A.A., and Njah A.N. 2012. Equilibrium Swelling and Kinetic Studies of Highly Swollen Chitosan Film. *J. Chem. Chem. Eng.* 6, 272-283.
- Bamgbose J.T., Bamigbade A.A. and Nkiko M.O. (2013). Adsorption kinetics and thermodynamics of malachite green onto chitosan/sodium citrate beads. *Ife J. of Science*. 15 (2): 385-398
- Berger J., Reist M., Mayer J.M., Felt O., Peppas N.A., and Gurny R. 2004. Structure and interactions in covalently and ionically cross-linked chitosan hydrogels for biomedical applications. *Eur. J Pharm. Biopharm.* 57: 19-34.
- Chiou K.C., Sahiner N., Godbey W.T., and Mcperson G.L. 2006. Microgel, nanogel and hydrogel-hydrogel semi-IPN composites for biomedical applications: synthesis and characterization. *Coll. Polym. Sci.*, 284: 1121-1129.
- Daniel E. K. and Cary T. C. 1989. Water solubility e n h a n c e m e n t o f D D T a n d Trichlorobenzene by some surfactant below and above the Critical Micellar Concentration. *Environ. Sci. Technol.* 23: 832-838.
- He F. and Zhao D. 2005. Preparation and characterization of a new class of starchstabilized bimetallic nanoparticles for

degradation of chlorinated hydrocarbons in water. *Environ Sci.*, *Technol.* 39(9): 3314–3320.

- Hirasa O., Ito S., Yamauchi A., Fujishige S., and Ichijo H. 1991. Polymer Gels. "Fundamentals and Biomedical application" Plenum Press, New York, p. 247
- Isogai N., Gong J.P., and Osada Y. 1996. Correlation analysis of diffusion of organic solvents across synthetic low polar polymeric membranes. *Macromol.*, 29: 6803-6812.
- Jameela S. R., Misra A. and Jayakriscnan A. 1994. Crosslinked chitosan microsphere as carrier for prolonged delivery of macromolecular drugs. J. Biomater. Sci., Polym. Edu. 6: 621-631
- Jiang T., Nukavarapu S.P., Deng M., Jabbarzadeh E., Kofron M.D., Doty S.B., Abdel-Fattah W.I., and Laurencin C.T. 2010. Development of new chitosan-cellulose multicore microcapsules for controlled drug delivery. *Acta Biomater.* 6: 3457-3489.
- Ju H.K., Kim S.Y., Kim S.J., and Lee Y.M. 2002. Interpenetrating polymer network hydrogels based on poly[ethylene glycol] macromer and chitosan. J. Appl polym. Sci., 83:1128-1134.
- Ju H.K., Kim S.Y., and Lee Y.M. 2001. Studies on effect of pH on cross-linking of chitosan with sodium tripolyphosphate: A technical note. *Polym*, 42: 685-697.
- Karadag E. and Saraydin D. 2002. Swelling of superabsorbent acylamide sodium acrylate hydrogels prepared using multifunctional crosslinkers. *Turk. J. Chem.* 26: 863-875.
- Kemp W. 1991. Infrared Spectroscopy. Macmillan Press, London, pp.19-56.
- Khokhlov A.R., Kramarenko E.Y., Makhaeva E.G., and Starodubtzev S.G. 1992. The influence of preparation methods on the swelling and network properties of acrylamide hydrogels with cross-linkers. *Macromol.*, 25: 4779-4786.
- Khokhov A.R., Starodubtzev S.G., Vasilevskaya V.V., and Dusek K. 1993. Advances in polymer science. Vol. 109, Berlin springer, 3451-3455.
- Kokufuta E., Zhang Y.Q., Tanaka T., and Mamada

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A. 1993. Water in hydrogels: An NMR study of water/polymer interactions in lightly cross-linked chitosan networks. *Macromol.*, 26:1053-1065.

- Leonida M.D., Banjade S., Vo T., Anderle G., Haas G.J., and Philips N. 2011. Nanocomposite materials with antimicrobial activity based on chitosan. *Int. J. Nano and Biomater.*, 3(4): 316–334.
- Li, Q., Dunn, E.T., Grandmaison, W.E. and Goosan M.F.A. 1997. Application and properties of Chitin and chitosan Goosen M.F.A. (Ed.) Technomic publishing company. Inc.USA, pp.3-9.
- Lu G.Y., Kong L.J., Sheng B.Y., Wang G., Gong Y.D., and Zhang X.F. 2007. Thermosensitivepoly [Nisopropylacrylamide] hydrogels: Synthesis, swelling and interaction with ionic surfactants. *Eur. Polym. J.* 43: 3807-3815.
- Martinez-Ruvalcaba A., J.C. Sanchez-Diaz, F. Becerra, L.E. Cruz-Barba and Gonzalaz-Alvarez A. 2009. Swelling characterization and drug delivery kinetics of polyacrylamide-co-itaconic acid/chitosan hydrogel. eXPRESS Polym. Lett. 3 (1) 25-32
- Mi F.L., Shyu S.S., Lee S.T., and Wong T.B. 1999. Kinetic study of chitosantripolyphosphate complex reaction and acid-resistive properties of the chitosantripolyphosphate gel beads prepared by in-liquid curing method. *J. Polym. Sci. Part-B Polym. Phys.*, 37: 1551-1564.
- Peppas N.A., Bures P., Leobanding W., and Lchikawa W. 2000. Hydrogels in Pharmaceutical formulations. *Eur. J Pharm. Biopharm.*, 50: 27-38
- Rana V., Babita K., Goyal D., Gorea R., and Tiwary A.K. 2004. Optimization of chitosan films as a substitute for animal and human

epidermal sheets for *in vitro* permeation of polar and nonpolar drugs. *Acta Pharm.*, 54: 287-299.

- Remunan-Lopez C., and Bodmeier R. 1997. Mechanical, water uptake and permeability properties of cross-linked chitosan glutamate and alginate films. *J. Contr. Rel.*, 44: 215-225.
- Sezer A.D., and Akbuga J. 1995. Controlled release of piroxicam from chitosan beads. *Int. J Pharm.*, 121:113-116.
- Tuncer C. and Melike D. 2004. Effects of temperature and surfactants on the equilibrium swelling behavior of polyacrylamide-co-(itaconic acid) hydrogel. *Macromol. Mater. Eng.*, 289, 548-551.
- Uzum O.B. and Karadag E. 2006. Synthetic polymeric absorbent for dye based on c h e m i c a l l y c r o s s - l i n k e d acrylamide/mesaconic acid hydrogels. J. *Appl. Polym. Sci.*, 101 (1):405-413.
- Wan Y., Creber K.A.M., Peppley B., and Bui V.T. 2004. Synthesis, characterization and ionic conductive properties of phosphorylated chitosan membranes. *Macromol. Chem. Physiol.*, 204: 850-858.
- Wang L., Khor E., Lim L.Y. 2001. Chitosanalginate-CaCl<sub>2</sub> system for membrane coat application. J. Pharm. Sci., 90:1134-1142.
- Wang M., Feng Q., Guo X., She Z., and Tan R. 2011. Methotrexate loaded chitosan and chitin microcapsules—in vitro characterization and pharmacokinetic s in mice bearing Ehrlich ascites carcinoma. J. Microencapsul. 15:581–594.
- Ziabari M., Mottaghitalab V., Haghi A.K. 2009. Smart nanoparticles: Preparation, characterization and applications. *Brazil. J. Chem. Eng.*, 26(1): 53–62.