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Circular Single-Stranded DNA Virus (*Microviridae*: Gokushovirinae: Jodiemicrovirus) Associated with the Pathobiome of the Flat-Back Mud Crab, Eurypanopeus depressus

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ABSTRACT A single-stranded DNA (ssDNA) virus is presented from a metagenomic data set derived from Alphaproteobacteria-infected hepatopancreatic tissues of the crab Eurypanopeus depressus. The circular virus genome (4,768 bp) encodes 14 hypothetical proteins, some similar to other bacteriophages (Microviridae). Based on its relatedness to other Microviridae, this virus represents a member of a novel genus.

icroviridae is a viral family with two subfamilies and 6 genera (1). It contains viruses that infect prokaryotes exclusively. Metagenomic techniques have unearthed the diversity of this family through evaluations of environmental, culture stock, and animal specimens (1-3). Microviruses have not been identified from Rickettsiales endosymbionts of Crustacea. Other bacteriophages isolated from crustacean microbiomes/pathobiomes show potential for phage therapy, avoiding the overuse of antibiotics in aquaculture (4). Few models exist to easily test this scenario in the laboratory.

We obtained DNA using a Zymo kit (D4070) on homogenized hepatopancreatic tissues of Eurypanopeus depressus (n = 1), a panopeid crab from meso- and euryhaline locations across the Gulf of Mexico and Atlantic North America. The specimen was collected from a euryhaline site in North Carolina (Hoop Pole Creek, Atlantic Beach) in December 2018. A total of 1 µg of DNA was used to prepare a NEBNext Ultra DNA library for Illumina HiSeq (10×) sequencing (NEB, USA) with a PE150 cartridge. This resulted in 11 million reads (50 to 150 bp) that were assembled using SPAdes v.3.13.0 (using default parameters and k-mer lengths of 21, 33, 55, 77, 99, and 127) (5) from trimmed reads using Trimmomatic (LEADING:3 TRAILING:3 SLIDINGWINDOW:4:15 MINLEN:36) (6). This resulted in 523,047 contigs (>500 bp) (N_{50} , 2,133; N_{75} , 1,340; L_{50} , 100,989; L_{75} , 211,827). The genome (4,768 bp) of a *Microviridae* sp. was identified based on high coverage (>1,000×), with a GC content of 33% and 14 hypothetical open reading frames (ORFs) (Fig. 1 and Table 1). The genome was annotated using ExPASy (standard genetic code) (7) and GeneMarkS (virus) (8). The relatedness of the genes and their function was identified using BLASTP (E value < 10) and InterProScan (9). Phylogenetics were conducted using IQ-Tree (10) after MAFFT alignment (11) of the capsid protein (ORF-1). The virus is genetically related to the Gokushovirinae subfamily of the Microviridae and represents a basal member to the three genera Bdellomicrovirus, Chlamydiamicrovirus, and Spiromicrovirus, as well as multiple other undescribed isolates associated with bacterial endosymbionts of tortoises, marine invertebrates, and insects (Fig. 1).

Of the 14 hypothetical ORFs, 5 showed similarity to other proteins in GenBank (Table 1). The proteins included a major capsid protein, DNA pilot protein, and the replicator initiator protein, which showed 32 to 44% amino acid similarity to other

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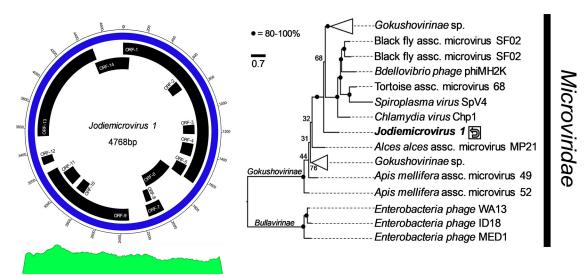


FIG 1 Circular genome of *Jodiemicrovirus 1*, consisting of 4,768 bp, and phylogenetic comparison to other *Microviridae* using the capsid protein (ORF-1). The genome contains 14 hypothetical open reading frames. The chart at the bottom identifies the read coverage across the circular genome, representing 934,456 reads mapped to the genome, providing >1,000× coverage using CLC Genomics Workbench. The phylogenetic comparison included the MAFFT-aligned (11) capsid protein (716 positions) from multiple *Microviridae*. The maximum likelihood tree was inferred from 36 *Microviridae* spp. and was developed with the LG+F+G4 evolutionary model and 1,000 bootstraps in IQ-Tree (10). The final consensus tree (shown) had a log likelihood of -28,362.192 and scale of 0.7 units. The accession numbers used were AXL15123, AXQ65957, QCS36953, AXH77578, AXL15643, AZL82997, AZL82921, AXL14929, YP_009218802, AYQ58216, AXL14945, AZL82910, AZL82729, AZL83022, AZL83017, QCS36934, QCS36961, QCS37361, AZL82956, AZL82992, AZL82871, YP_00951424, AZL82946, QCQ84972, AZL82926, QCS37201, QCQ84913, AZL82717, AZL82837, YP_512416, YP_512796, AII27899, NP_073538, NP_044312, and NP_598320.

Microviridae (Table 1). One virus was from an environmental sphagnum peat soil sample (12), and two viruses derived from the intestinal tract of Ciona robusta (Tunicata) (marine) (2). Two genes showed closest similarity to hypothetical bacterial genes (Table 1). Seven ORFs were identified internally to other ORFs, indicating the presence of putative overlapping genes recently discovered for the Microviridae (13). Based on its relatedness to known Microviridae, this genome might represent a novel genus (suggested, Microviridae: Gokushovirinae: Jodiemicrovirus).

To conclude, we present the genome of a bacteriophage likely to infect an undescribed member of the *Anaplasmataceae* which parasitizes the host hepatopancreas, identified via histology, electron microscopy, and genomics (our unpublished data). It may constitute a useful model system for understanding the effect of phage therapy relative to an intracellular bacterium causing disease in crustaceans.

TABLE 1 Similarity and predicted function of the 14 hypothetical ORFs found in *Jodiemicrovirus* 1^a

ORF	Predicted function	Closest hit (accession no.)	Similarity (%)	Coverage (%)	E value
1	Viral capsid	Microviridae sp. (AXL15123)	43.06	96	4e-138
2	Transmembrane	_	_	_	_
3	Signal peptide	Acidimicrobiaceae (MBB33698)	55.56	84	2.9
4	Unknown	_	_	_	_
5	Transmembrane	_	_	_	_
6	Unknown	_	_	_	_
7	Unknown	_	_	_	_
8	Unknown	_	_	_	_
9	DNA pilot protein	Microviridae sp. (YP_009160339)	33.33	40	1e-7
10	Unknown	_	_	_	_
11	Transmembrane	_	_	_	_
12	Signal peptide	_	_	_	_
13	Replication initiator protein	Microviridae sp. (AXL15534)	32.00	72	3e-28
14	Unknown	Bacteria (EKD64965)	40.38	41	6.6

^aThe coding orientation is positive in all cases. Data were analyzed using InterProScan and BLASTP. —, lack of significant protein similarity to any other known sequence data.

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Data availability. The complete genome, annotation, and associated forward and reverse reads for this novel virus can be found under accession number MN335165, BioProject number PRJNA574411, and BioSample number SAMN12567204.

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REFERENCES

- Adriaenssens EM, Wittmann J, Kuhn JH, Turner D, Sullivan MB, Dutilh BE, Jang HB, van Zyl LJ, Klumpp J, Lobocka M, Moreno Switt Al, Rumnieks J, Edwards RA, Uchiyama J, Alfenas-Zerbini P, Petty NK, Kropinski AM, Barylski J, Gillis A, Clokie MRC, Prangishvili D, Lavigne R, Aziz R, Duffy S, Krupovic M, Poranen MM, Knezevic P, Enault F, Tong Y, Oksanen HM, Brister JR. 2018. Taxonomy of prokaryotic viruses: 2017 update from the ICTV Bacterial and Archaeal Viruses Subcommittee. Arch Virol 163: 1125–1129. https://doi.org/10.1007/s00705-018-3723-z.
- Creasy A, Rosario K, Leigh B, Dishaw L, Breitbart M. 2018. Unprecedented diversity of ssDNA phages from the family *Microviridae* detected within the gut of a protochordate model organism (*Ciona robusta*). Viruses 10:404. https://doi.org/10.3390/v10080404.
- Walters M, Bawuro M, Christopher A, Knight A, Kraberger S, Stainton D, Chapman H, Varsani A. 2017. Novel single-stranded DNA virus genomes recovered from chimpanzee feces sampled from the Mambilla Plateau in Nigeria. Genome Announc 5:e01715-16. https://doi.org/10.1128/genomeA .01715-16.
- Lomelí-Ortega CO, Martínez-Díaz SF. 2014. Phage therapy against Vibrio parahaemolyticus infection in the whiteleg shrimp (Litopenaeus vannamei) larvae. Aquaculture 434:208–211. https://doi.org/10.1016/ j.aquaculture.2014.08.018.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- 6. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for

- Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10.1093/bioinformatics/btu170.
- Gasteiger E, Hoogland C, Gattiker A, Wilkins MR, Appel RD, Bairoch A. 2005. Protein identification and analysis tools on the ExPASy server, p 571–607. In Walker JM (ed), The proteomics protocols handbook. Humana Press. Totowa. N.I.
- Besemer J, Lomsadze A, Borodovsky M. 2001. GeneMarkS: a self-training method for prediction of gene starts in microbial genomes. Implications for finding sequence motifs in regulatory regions. Nucleic Acids Res 29:2607–2618. https://doi.org/10.1093/nar/29.12.2607.
- Quevillon E, Silventoinen V, Pillai S, Harte N, Mulder N, Apweiler R, Lopez R. 2005. InterProScan: protein domains identifier. Nucleic Acids Res 33:W116-W120. https://doi.org/10.1093/nar/qki442.
- Trifinopoulos J, Nguyen LT, von Haeseler A, Minh BQ. 2016. W-IQ-TREE: a fast online phylogenetic tool for maximum likelihood analysis. Nucleic Acids Res 44:W232–W235. https://doi.org/10.1093/nar/gkw256.
- 11. Miller MA, Pfeiffer W, Schwartz T. 2011. The CIPRES science gateway: a community resource for phylogenetic analyses, p 41. Proceedings of the 2011 TeraGrid Conference: extreme digital discovery. Association for Computing Machinery, 18 to 21 July 2011, Salt Lake City, UT.
- Quaiser A, Dufresne A, Ballaud F, Roux S, Zivanovic Y, Colombet J, Sime-Ngando T, Francez A-J. 2015. Diversity and comparative genomics of *Microviridae* in Sphagnum-dominated peatlands. Front Microbiol 6:375. https://doi.org/10.3389/fmicb.2015.00375.
- Pavesi A. 2006. Origin and evolution of overlapping genes in the family *Microviridae*. J Gen Virol 87:1013–1017. https://doi.org/10.1099/ vir.0.81375-0.

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