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Food and Chemical Toxicology 50 (2012) 4062-4067





journal homepage: www.elsevier.com/locate/foodchemtox

# Effects of fibre-enriched diets on tissue lipid profiles of MSG obese rats

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#### ARTICLE INFO

Article history: Received 2 November 2011 Accepted 1 August 2012 Available online 8 August 2012

Keywords: MSG Fibre Obesity Dyslipidemia HMG-CoA reductase

#### ABSTRACT

In order to investigate the influence of some fibre-enriched diets on tissue lipids in an animal model of obesity induced by the administration of monosodium glutamate (MSG), obese rats were fed diets containing 30% of Acha, Cassava, Maize and Plantain for five weeks and weight gain, feed intake and lee index were recorded. The lipid profiles of plasma, erythrocytes, kidney, heart and liver as well as hepatic 3hydroxyl-3-methylglutaryl-CoA (HMG-CoA) reductase activity were measured. The diets significantly (p < 0.05) reduced weight gain and lee index in the obese rats. Obesity-induced increase in plasma and erythrocytes lipid levels was significantly (p < 0.05) reduced by these diets. MSG-induced obesity also resulted in a significant increase (p < 0.05) in hepatic cholesterol level which was reduced by the diets. MSG-obesity was characterised by a significant (p < 0.05) increase in cholesterol, triacylglycerol and phospholipids in kidney and this was reversed by the diets except Maize which did not reverse the increased cholesterol level. Only Acha reversed the obesity-induced increase in heart cholesterol and phospholipids. The increased activity of hepatic HMG-CoA reductase associated with obesity was also significantly (p < 0.05) reduced by the diets. In conclusion, dyslipidemia associated with MSG-induced obesity could be attenuated by consumption of fibre-enriched diets.

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## 1. Introduction

Increased food consumption in excess of energy requirements, coupled with excessive body fat present a growing human health problem (Mozes et al., 2004). Obesity, which is defined as body fat excess, usually develops slowly as a result of long-term alterations in energy balance. In general, when food intake exceeds energy expenditure, the retained energy is deposited as fat (Dolnikoff et al., 2001). Obesity is a highly prevalent disorder associated with decreased life expectancy and increased morbidity because of its combination with a variety of other disorders including hyperglycemia, hyperlipidemia, hypertension and consequently cardiovascular diseases carrying significant economic cost (Guven et al., 1999). These disorders are often associated with both qualitative and quantitative changes in lipid composition of several tissues in the body.

Dietary habits are considered one of the factors contributing to obesity; hence, a scrupulous dietary intervention is indicated in its management (De Filippo et al., 2010). This dietary intervention should include diets that regulate food intake and energy balance.

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African diets are known to be high-fibre and low-fat (De Filippo et al., 2010; Thian et al., 2006). While the rural African population live agrarian life style, urban population is becoming westernized (De Filippo et al., 2010). In Nigeria, for example, the rural populations consume more of carbohydrate and fibre-rich food like Cassava, Acha, Plantain and Maize more than western-type processed food and beverages (Olumakaiye et al., 2010). Thian et al. (2006) reported that in sub-saharan Africa, obesity is becoming a problem with 10-30% of men and 15-45% of women in West Africa being either overweight or obese.

In general, it is believed that humans are more suited to resist famine than overabundance of food (called the "thrift gene hypothesis") and; hence, it has been argued that the easy and related inexpensive availability of energy-dense food is responsible for the current obesity epidemic (Das, 2010). The energy balance is very tightly controlled by hypothalamic factors. Hence, the gut-brain axis and the cross-talk between gut hormones and hypothalamic factors are important in the regulation of food intake, energy balance, and development of obesity (Das, 2010). This knowledge has been under utilised in developing experimental murine model of obesity by intraperitoneal administration of neonatal rats with monosodium glutamate (MSG). This study was therefore aimed at studying the effects of some fibre-enriched diets on MSG - obesity - induced perturbations in lipid metabolism.

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O.A. Rotimi et al./Food and Chemical Toxicology 50 (2012) 4062-4067

#### 2. Materials and methods

#### 2.1. Animals and diets

Male neonatal albino rats were used for the experiment and obesity was induced as described by Nakagawa et al. (2000) with slight modification. Briefly, the neonates were treated with MSG (4 mg/g body weight) intraperitoneally five times (2,4,6,8 and 10 days after birth). Physiological saline was administered in a similar fashion to the control animals. The rats were weaned on the 21st day and raised normally thereafter, and studied at the age of 10 weeks. At the tenth week, Lee indices of the animals were calculated as  $\frac{\sqrt[3]{body weight(g)}}{nasal-anal length(cm)}$ . 25 rats with Lee index of 0.3 or more were considered obese. A total of 30 rats (25 obese surviving rats, 5 normal rats (not treated with MSG)) were used and they were divided into 6 groups of 5 rats each and randomly assigned to the experimental diets (Table 1). The 6 groups of rats were designated as follows:

Group 1. Normal-C: Rats fed normal diet.

Group 2. MSG-C: Obese rats fed normal diet.

Group 3. MSG-Acha: Obese rats fed Acha.

Group 4. MSG-Cassava: Obese rats fed Cassava flakes.

Group 5. MSG-Maize: Obese rats fed Maize.

Group 6. MSG-Plantain: Obese rats fed unripe Plantain flour.

The rats were fed for five weeks on the experimental diets. The composition of the experimental diet (Table 1) was based on the AIN-93 semisynthetic diet and were prepared to contain 30% of the fibre enriched diets. The animals were given food and distilled water ad libitum during the experimental period. Food consumption was measured daily while weight gain was measured weekly. The Ethical Committee for Conduction of Animal Studies at the Department of Biochemistry, University of Agriculture, Abeokuta, Ogun State approved the experimental protocol and all animals were cared for in accordance with the principles and guidelines of the committee.

## 2.2. Tissue collection

At the end of the experimental period, blood was collected from the animals into heparinised tubes by cardiac puncture under light ether anesthesia after an overnight fast. The plasma was seperated from the erythrocytes by centrifuging the whole blood at 5000 rpm for 10 min. Erythrocytes were washed three times with normal saline. The organs were excised, rinsed with normal saline, blotted dry and weighed immediately. The erythrocytes, plasma and organs were stored at -20 °C until analyzed.

## 2.3. Biochemical analyses

#### 2.3.1. Plasma lipid profiles

Plasma concentrations of total cholesterol and triglycerides were determined with commercial kits (Cromatest linear chemicals, Montgat Spain). HDL cholesterol and triglycerides were determined in plasma with same commercial kits for total cholesterol and triglycerides after very low density lipoproteins (VLDL) and LDL were precipitated with heparin-MnCl<sub>2</sub> solution as described by Gidez et al.

#### Table 1

Composition of diet in g/100 g.

Composition	Control	Acha	Cassava	Maize	Plantain
Fish Meal	20	20	20	20	20
Groundnut oil	5	5	5	5	5
Mineral mix*	3.5	3.5	3.5	3.5	3.5
Vitamin mix*	1	1	1	1	1
Cellulose	5	5	5	5	5
Maize starch	50	25	25	25	25
Sucrose	15	10	10	10	10
Choline bitartate	0.2	0.2	0.2	0.2	0.2
DL-Methionine	0.3	0.3	0.3	0.3	0.3
Acha	-	30	-	-	-
Cassava	-	-	30	-	-
Maize	-	-	-	30	-
Unripe Plantain flour	-	-	-	-	30

\* Mineral mix and vitamin mix contains the following in g/100 g:

Calcium phosphate 49.50, sodium powder 11.80, potassium sulphate 5.20, sodium chloride 7.40, magnesium oxide 2.40, potassium citrate 22.40, ferric citrate 0.60, manganous carbonate 0.35, cupric carbonate 0.03, zinc carbonate 0.16, chromium potassium sulfate 0.055, potassium iodate 0.001, sodium selenate 0.001, choline chloride 0.50, thiamine HCl 0.06, riboflavin 0.06, niacine 0.30, calcium pantothenate 0.16, biotin 0.01, vit D3 0.025, vit B12 0.10, vit E acetate 1.00, pyridoxine 0.07, folic acid 0.02, vit A acetate 0.08.

(1982). Total phospholipids in plasma and HDL were extracted with chloroformmethanol mixture (2:1, v/v) as described by Folch et al. (1957). Phospholipid content was then determined according to the method of Stewart (1980), which is based on the formation of a complex between phospholipids and ammonium ferrothiocyanate.

## 2.3.2. Erythrocyte lipid profile

Lipids were extracted from the erythrocytes as described by Rose and Oklander (1965) using chloroform-isopropanol (7:11, v/v). Aliquots of the chloroform-isopropanol extract were then used for the determination of cholesterol and triglyceride. Determination of total phospholipids in the chloroform-isopropanol extract of the erythrocyte followed the same procedure as described for plasma (Stewart, 1980).

#### 2.3.3. Organ lipid profiles

Organ lipids were extracted according to the method of Folch et al. (1957). After washing with 0.05 M KCl solution, aliquots of the chloroform-methanol extract were then used for the determination of cholesterol, triglycerides and phospholipids concentrations. Cholesterol determination followed the same procedure as described for erythrocytes while determination of phospholipids followed the same procedure as described for plasma. Triacylglycerol concentrations in aliquots of the chloroform-methanol extracts of each organ were determined following the procedure described by Kriketos et al. (2003).

#### 2.3.4. 3-Hydroxy-3-methylglutaryl (HMG) coenzyme A (CoA) reductase activity

The activity of HMG-CoA reductase was determined using an indirect method described by Rao and Ramakrishnan (1975). In this method, the concentrations of HMG-CoA and mevalonate are determined in the liver homogenate and the ratio of HMG-CoA/mevalonate is taken as an index of the activity of HMG-CoA reductase. An increase in the ratio indicates decrease whereas a decrease in the ratio indicates increased HMG-CoA reductase activity.

#### 2.3.5. Estimation of VLDL-cholesterol and LDL-cholesterol

The concentrations of very low density lipoproteins (VLDL) Cholesterol and Low Density (LDL) Cholesterol were calculated by a modification of the Friedewald formular (Sandkamp et al., 1990).

VLDL-Cholesterol was calculated as triacylglycerols/5 and LDL-cholesterol was calculated by the equation: LDL-cholesterol = Total plasma cholesterol-(HDL + VLDL).

## 2.3.6. Estimation of atherogenic and coronary risk indexes

Atherogenic Index (AI) was calculated as the ratio of LDL-Cholesterol:HDL-Cholesterol while Coronary Risk Index (CRI) was estimated as the ratio of plasma total Cholesterol:HDL-Cholesterol as described by Ademuyiwa et al. (2008).

#### 2.4. Statistical analysis

Calculations were made using the software "SPSS 13.0 for Windows" and data were expressed as Mean  $\pm$  SEM of five readings. Analysis of Variance (ANOVA) was carried out to test for the level of homogeneity at p < 0.05 among the groups using Duncan's Multiple Range Test (DMRT) to separate the heterogeneous groups.

## 3. Results

The MSG treated rats gained more weight and consumed less feed than the control rats. There was no significant (p < 0.05) difference in the Lee indices among all the MSG treated groups before treatment with the diets. However, the diets brought about a significant (p < 0.05) decrease in the Lee indices of the animals. *Acha* and unripe Plantain flour containing diets brought the lee index to levels similar that of the normal rats (Table 2). There was no significant (p < 0.05) difference in heart weight among all the groups. Kidney weight of the MSG-treated rats and that on Cassava containing diet were also significantly (p < 0.05) lower than the normal rats. Liver weight of the MSG treated rats were significantly (p < 0.05) lower than that of the normal rats and the other groups (Table 2).

MSG intake induced dyslipidemia in both plasma and lipoproteins and this was characterised by increased cholesterol, triacylglycerol and phospholipids. Intake of the diets attenuated the MSG-induced dyslipidemia (excluding total and HDL-phospholipids where *Acha* further significantly (p < 0.05) increased their concentrations). Unripe Plantain flour containing diet had the most

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## 4064

Table 2

#### O.A. Rotimi et al. / Food and Chemical Toxicology 50 (2012) 4062-4067

Group	Normal-C	MSG-C	MSG-Acha	MSG-Cassava	MSG-Maize	MSG-Plantair
Initial weight (g)	135.02 ± 2.10 <sup>a</sup>	154.40 ± 3.20 <sup>b</sup>	150.40 ± 5.20 <sup>b</sup>	156.50 ± 3.20 <sup>b</sup>	155.50 ± 5.60 <sup>b</sup>	151.6 ± 4.30 <sup>b</sup>
Final weight (g)	$148.80 \pm 1.30^{a}$	$196.00 \pm 2.50^{b}$	$199.60 \pm 3.20^{b}$	$180.93 \pm 2.40^{\circ}$	191.94 ± 1.50 <sup>b</sup>	$139.6 \pm 3.40^{d}$
Weight gain (g) <sup>B</sup>	$13.80 \pm 2.20^{a}$	$41.60 \pm 5.04^{cd}$	$49.20 \pm 5.31^{d}$	24.43 ± 3.64 <sup>ac</sup>	$36.44 \pm 7.21^{ac}$	$-12.00 \pm 4.35^{t}$
Feed intake (g/day)	$16.19 \pm 0.87^{a}$	$10.85 \pm 0.22^{b}$	$14.80 \pm 0.58^{cd}$	$13.40 \pm 0.46^{\circ}$	$17.20 \pm 0.12^{e}$	$8.77 \pm 0.47^{f}$
Initial Lee-index	$0.29 \pm 0.03^{a}$	$0.33 \pm 0.02^{b}$	$0.34 \pm 0.01^{b}$	$0.33 \pm 0.02^{b}$	$0.34 \pm 0.01^{b}$	$0.33 \pm 0.01^{b}$
Final Lee-index	$0.29 \pm 0.02^{a}$	$0.33 \pm 0.03^{d}$	$0.30 \pm 0.03^{ab}$	$0.31 \pm 0.03^{bc}$	0.31 ± 0.03 <sup>c</sup>	$0.29 \pm 0.04^{a}$
Heart <sup>A</sup>	$0.32 \pm 0.01^{a}$	$0.28 \pm 0.01^{a}$	$0.32 \pm 0.01^{a}$	$0.31 \pm 0.01^{a}$	$0.31 \pm 0.02^{a}$	$0.32 \pm 0.03^{a}$
Kidney <sup>A</sup>	$0.52 \pm 0.02^{a}$	$0.48 \pm 0.02^{b}$	$0.56 \pm 0.03^{a}$	$0.50 \pm 0.02^{b}$	$0.56 \pm 0.02^{a}$	$0.59 \pm 0.04^{\circ}$
Liver <sup>A</sup>	$2.9 \pm 0.08^{a}$	2.8 ± 0.13 <sup>b</sup>	$3.3 \pm 0.01^{\circ}$	3.3 ± 0.17 <sup>c</sup>	$3.2 \pm 0.07^{a}$	$3.8 \pm 0.26^{d}$

Each value represents the mean  $\pm$  S.E.M. n = 5. Values within the same row with different alphabets are significantly (p < 0.05) different from each other. <sup>A</sup> Values expressed in organ weight/100 g Body Weight

<sup>B</sup> Weight gain is the difference between the weight at the beginning of the treatment and the weight at the end of the treatment (5 weeks)

Effects of the diets on blood lipid profiles.	Tuble J	
	Effects of the diets or	ı blood lipid profiles.

Group	Normal-C	MSG-C	MSG-Acha	MSG-Cassava	MSG-Maize	MSG-Plantain
Erythrocyte cholesterol	$47.82 \pm 1.59^{a}$	85.80 ± 1.03 <sup>e</sup>	65.11 ± 1.47 <sup>b</sup>	66.59 ± 1.16 <sup>bc</sup>	70.11 ± 1.67 <sup>dc</sup>	$73.33 \pm 1.27^{d}$
Erythrocyte triacylglycerol	$30.74 \pm 0.85^{a}$	67.27 ± 1.17 <sup>c</sup>	$30.16 \pm 0.94^{a}$	$19.09 \pm 0.78^{b}$	19.28 ± 1.39 <sup>b</sup>	$22.06 \pm 1.09^{b}$
Erythrocyte phospholipids	$157.40 \pm 2.75^{a}$	$184.47 \pm 2.09^{d}$	155.27 ± 3.57 <sup>a</sup>	139.02 ± 0.83 <sup>bc</sup>	145.76 ± 3.39 <sup>b</sup>	131.69 ± 3.00 <sup>c</sup>
Plasma total cholesterol	81.17 ± 0.59 <sup>a</sup>	$112.42 \pm 2.82^{d}$	92.06 ± 1.99 <sup>c</sup>	90.51 ± 2.89 <sup>cb</sup>	98.11 ± 3.47 <sup>c</sup>	83.71 ± 2.21 <sup>a</sup>
Plasma total triacylglycerol	57.69 ± 1.89 <sup>a</sup>	$156.4 \pm 3.63^{d}$	87.3 ± 2.71 <sup>c</sup>	80.95 ± 1.24 <sup>c</sup>	71.92 ± 1.58 <sup>b</sup>	$67.39 \pm 1.00^{b}$
Plasma total phospholipids	$108.59 \pm 2.04^{a}$	131.74 ± 3.46 <sup>d</sup>	166.69 ± 1.16 <sup>e</sup>	$102.96 \pm 2.36^{a}$	124.39 ± 1.00 <sup>c</sup>	$80.73 \pm 0.89^{b}$
HDL cholesterol	36.9 ± 1.85 <sup>a</sup>	$17.63 \pm 2.82^{d}$	30.08 ± 1.11 <sup>c</sup>	37.09 ± 1.21 <sup>a</sup>	49.83 ± 0.77 <sup>e</sup>	$25.70 \pm 0.34^{b}$
HDL triacylglycerol	23.45 ± 1.06 <sup>a</sup>	32.07 ± 1.86 <sup>b</sup>	33.64 ± 1.33 <sup>b,c</sup>	35.49 ± 0.96 <sup>b,c</sup>	36.22 ± 0.61 <sup>c</sup>	$23.70 \pm 1.44^{a}$
HDL phospholipids	$9.91 \pm 0.92^{a}$	$37.48 \pm 1.09^{d}$	60.21 ± 0.92 <sup>e</sup>	16.32 ± 1.30 <sup>b</sup>	21.11 ± 0.44 <sup>c</sup>	$10.41 \pm 0.89^{a}$
LDL cholesterol	32.71 ± 1.74 <sup>a</sup>	63.50 ± 2.46 <sup>c</sup>	44.52 ± 2.78 <sup>b</sup>	37.22 ± 2.76 <sup>ab</sup>	$33.89 \pm 2.30^{a}$	44.53 ± 2.55 <sup>b</sup>
VLDL cholesterol	$11.54 \pm 0.37^{a}$	$31.28 \pm 0.72^{d}$	$17.46 \pm 0.54^{\circ}$	$16.20 \pm 0.25^{\circ}$	$14.38 \pm 0.32^{b}$	$13.47 \pm 0.20^{b}$

Values expressed as mg/dL and each value represents the mean ± S.E.M. n = 5. Values within the same row with different alphabets are significantly (p < 0.05) different from each other.

lowering effect, except in the case of LDL-cholesterol where Maize was the most effective. HDL-cholesterol level of the MSG-treated was significantly (p < 0.05) lower than that of the control. Intake of the diet resulted in a significant (p < 0.05) increase in HDL-cholesterol concentration with Maize being significantly (p < 0.05)higher than the control and Cassava similar to the control animals (Table 3).

Cholesterol, triacylglycerol and phospholipid levels in the erythrocytes were significantly (p < 0.05) higher in the MSG treated rats when compared to the control rats. Intake of the diets, significantly (p < 0.05) decreased their concentrations when compared to the MSG treated rats, the phospholipid concentrations of the animals taking the diets being significantly lower than the normal animals (Table 3).

Renal cholesterol and triacylglycerol levels in the MSG-treated rats were significantly (p < 0.05) higher than the normal. Intake

of the diets resulted in a significant (p < 0.05) decrease in their concentrations. However MSG treatment brought about a significant (p < 0.05) decrease in the level of phospholipid in the kidney when compared to the normal control. This reduction was significantly (p < 0.05) reversed by Maize-containing diets while other diets led to a further significant (p < 0.05) drop in the concentration of phospholipid in the kidney (Table 4).

Cholesterol and phospholipid levels in the heart of the MSGtreated rats were significantly (p < 0.05) higher than that of the normal rats. While other diets resulted in a significant increase in heart cholesterol and phospholipid, Acha containing diets consumption gave a significant decrease in the heart cholesterol while Acha and unripe Plantain flour reduced heart phospholipid. The MSG treated rats had a triacylglycerol level significantly (p < 0.05) lower than the normal rats. Consumption of Acha and Cassava containing diets gave triacylglycerol levels similar to that

Table 4				
Effects	of diets	on	organ	lipids.

Group	Normal-C	MSG-C	MSG-Acha	MSG-Cassava	MSG-Maize	MSG-Plantain
Renal cholesterol	$2.02 \pm 0.02^{a}$	$2.89 \pm 0.07^{d}$	$1.10 \pm 0.02^{e}$	$1.09 \pm 0.02^{e}$	$2.21 \pm 0.10^{\circ}$	$2.00 \pm 0.04^{\rm b}$
Renal triacylglycerol	$5.53 \pm 0.07^{a}$	6.07 ± 0.13 <sup>e</sup>	$3.05 \pm 0.02^{d}$	$5.03 \pm 0.07^{\circ}$	$5.25 \pm 0.12^{\circ}$	$3.91 \pm 0.05^{b}$
Renal phospholipids	$11.19 \pm 0.19^{a}$	$8.79 \pm 0.08^{\circ}$	$7.72 \pm 0.11^{b}$	$5.47 \pm 0.16^{d}$	$12.10 \pm 0.16^{a}$	8.31 ± 0.19 <sup>b</sup>
Heart cholesterol	$0.54 \pm 0.01^{a}$	0.76 ± 0.01 <sup>c</sup>	$0.46 \pm 0.01^{b}$	$0.88 \pm 0.03^{d}$	0.98 ± 0.01 <sup>e</sup>	$0.85 \pm 0.01^{d}$
Heart triacylglycerol	$2.21 \pm 0.04^{a}$	$1.86 \pm 0.02^{b}$	$2.27 \pm 0.12^{d}$	$2.08 \pm 0.08^{\circ}$	$3.61 \pm 0.07^{e}$	$1.45 \pm 0.02^{f}$
Heart phospholipids	$15.81 \pm 0.16^{a}$	$19.76 \pm 0.10^{d}$	$8.88 \pm 0.03^{b}$	21.99 ± 1.01 <sup>e</sup>	21.51 ± 0.55 <sup>e</sup>	17.72 ± 0.18 <sup>c</sup>
Hepatic cholesterol	$1.91 \pm 0.07^{a}$	$2.59 \pm 0.08^{b}$	$1.24 \pm 0.08^{a}$	$1.63 \pm 0.06^{\circ}$	$2.27 \pm 0.06^{d}$	$1.88 \pm 0.04^{a}$
Hepatic triacylglycerol	$0.86 \pm 0.06^{a}$	$1.11 \pm 0.06^{a}$	$0.95 \pm 0.08^{a}$	$1.30 \pm 0.07^{b}$	$1.81 \pm 0.08^{\circ}$	$1.05 \pm 0.08^{a}$
Hepatic phospholipids	$22.93 \pm 0.1^{a}$	22.17 ± 0.49 <sup>bc</sup>	$24.20 \pm 1.00^{d}$	31.86 ± 0.79 <sup>e</sup>	$20.39 \pm 0.52^{b}$	$18.33 \pm 0.45^{a}$

Values expressed as mg/g tissue and each value represents the mean  $\pm$  S.E.M. n = 5. Values within the same row with different alphabets are significantly (p < 0.05) different from each other.

of the normal rats while Maize containing diets gave levels significantly (p < 0.05) higher than the normal rats (Table 4).

Hepatic cholesterol levels in the MSG-treated rats were significantly (p < 0.05) higher than that of the normal rats. Consumption of the diets however resulted in a significant (p < 0.05) decrease in the cholesterol concentrations. Acha had the most pronounced effect in lowering hepatic cholesterol while Maize had the least effect. MSG treatment brought about a significant decrease in hepatic phospholipids. Consumption of unripe Plantain flour brought about significant (p < 0.05) decrease while Acha and Cassava significantly (p < 0.05) increased phopholipid levels. Hepatic TG level in MSG-treated rats was similar to that of the normal rats, however Acha and Cassava significantly (p < 0.05) increased the concentration of TG while the concentration of TG in the other diets was similar to that of the normal and MSG-treated rats (Table 4).

The MSG treated rats had a coronary risk index value of about 2.9 times higher than the normal rats (p < 0.05). Rats fed with Cas-

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sava and Maize containing diets brought the ratio to levels similar to that of the normal rats. Treatment with MSG led to about 4-fold increase in the ratio of atherogenic risk index when compared with the normal control. Intake of the diets reduced the ratio, except the animals fed Acha and unripe Plantain flour, to levels similar to that of the normal animals. (Fig. 1) HMG-CoA/Mevalonate ratio is inversely proportional to the activity of HMG CoA reductase in the liver of the animals. The HMG CoA reductase activities of MSG-treated rats were significantly higher than others while Acha had the lowest HMG CoA reductase activity. The activity of the enzyme of the obese rats fed the diets was between 1.1 and 4.5 times lower when compared with the obese rats not fed the diets. (Fig. 2)

## 4. Discussion

natally treated with MSG exhibited obesity with increased Lee in-





Fig. 1A. Coronary Risk Index (CRI) of the animals exposed to different experimental diets. Bars represent the means ± SEM (n = 5). Bars with different alphabets are significantly (p < 0.05) different from each other.



Fig. 1B. Atherogenic Index (AI) of the animals exposed to different experimental diets. Bars represent the means ± SEM (n = 5). Bars with different alphabets are significantly (p < 0.05) different from each other.

O.A. Rotimi et al. / Food and Chemical Toxicology 50 (2012) 4062-4067



Fig. 2. HMG CoA/Mevalonate ratio of the animals exposed to different experimental diets. Bars represent the means  $\pm$  SEM (n = 5). Bars with different alphabets are significantly (p < 0.05) different from each other.

dex. This observation is consistent with the results of earlier studies (Martinkova et al., 2000; Novelli et al., 2007; Nakagawa et al., 2000). It is noteworthy that throughout the duration of the study, MSG-obese rats consumed less food compared with the normal rats. In contrast to other forms of obesity which are associated with hyperphagia, the obesity model of neonatal MSG-treated rodents is associated with increased plasma levels of corticosterone (Guimaraes et al., 2002) as well as increased lipogenesis and reduced lipolysis in adipose tissues (Dolnikoff et al., 2001) occurring despite their normophagia or even hypophagia (Zhang et al., 1994; Martinkova et al., 2000).

When the hypothalamic ventromedial nucleus and arcuate nucleus are destroyed in rats by treatment with MSG in the neonatal stage, there is development of obesity as the rats grew (Nakagawa et al., 2000). Novelli et al. (2007) reported that adult rats (age 60 days) model of obesity experience a cessation of growth and development with a concomitant accumulation of fat in the abdominal region, leading to an increase in weight without a correspondent increase in the body length. This may lead to chronic exposure of tissues to elevated concentrations of lipids which may contribute to tissue dysfunction and obesity-related disease such as diabetes, hypertension, cardiovascular disease and metabolic syndrome (Muoio and Newgard, 2006).

The present investigation shows that unripe Plantain flour consumption by the obese rats led to weight loss while Cassava and Maize had reduced weight gain as compared with the MSG-obese rats fed on control diets. Unripe Plantain has been reported to be useful in managing diabetes and obesity due to its low glycemic index which is the consequence of its small concentration of free sugars and rapidly digestible starch (Ramdath et al., 2004). The potential mechanisms of weight reduction by these diets may include antihyperglycemic effects, inhibition of lipid droplet accumulation in fat cells without affecting adipose conversion, inhibition of fatty acid synthase, or inhibition of adipocyte differentiation (Kim et al., 2003). In particular, high-fibre diet feeding is accompanied by greater satiety and less hunger than low-fibre weight-reducing diets. Hence, an explanation for the reduced weight gain in rats on fibre-enriched diets is that nutrient absorption is limited and its caloric utilisation is different (Mazur et al., 1990).

As also observed in this study, MSG-induced obesity was associated with dyslipideamia in the rat tissues. The response of plasma and erythrocyte lipid profile presented in this work is in general agreement with data from other studies (Nakagawa et al., 2000; Nagata et al., 2006). Dyslipidemia was expressed as an increase in plasma total cholesterol, plasma triacylglycerol, RBC cholesterol and RBC triacylglycerol with a decrease in HDL cholesterol. Also total cholesterol:HDL cholesterol and LDL cholesterol:HDL cholesterol ratios were increased. In man at least, these ratios are considered to increase the risk of cardiovascular dysfunction, especially when total cholesterol level is elevated (Dabai et al., 1996). It was therefore noteworthy that the diets used in this study were able to improve plasma and RBC dyslipidemia and reduce atherogenic and coronary heart disease risk index in MSG-obese rats. This observation is consistent with the findings of Adamson and Mbajiorgu (1985) who reported that the serum lipid levels were reduced in rabbits fed Cassava and Unripe Plantain flourbased diets. They attributed their observations to the type and amount of dietary fibre in the diets they used.

Several mechanisms have been suggested to be responsible for the influence of dietary fibre on blood lipids. One is ascribed to the water soluble fibres (Kritchevsky, 1988; Madar and Odes, 1990), which when present in gastrointestinal tract increases viscosity and interferes with micelle formation and lipid absorption (Gallaher et al., 1993; Superko et al., 1988). In addition, certain fibre varieties bind or adsorb bile acids and neutral sterols, enhancing their removal from enterohepatic circulation leading to lower cholesterol in the various pools. If the absorption of lipids in the small intestine was decreased, it would be expected that the amount of cholesterol, triacylglycerol and perhaps phospholipids in the pools would also decrease but all pools of lipid are not affected in the same way and often times, the effect of different fibres is to shift the balance of lipid among the various compartments and pools (Adamson and Mbajiorgu, 1985; Mehta and Kaur, 1992). Shortchain fatty acids (SCFA)-acetate, propionate and butyrate are produced in large quantities from the fermentation of dietary fibres in the colon. Studies have indicated that there might be a connection between SCFA and lipid metabolism. Wolever et al. (1991), using rectal infusions of acetate alone, propionate alone, or a combination of acetate and propionate, showed that propionate inhibited the utilisation of acetate for cholesterol synthesis. It is possible that the blood levels of these SCFAs in circulation and the amount in the organs examined in this study were responsible for the variation in the amount of lipids present in them.

The fibre-enriched diets used in this study also resulted in a reduction of hepatic HMG-CoA reductase activity of the rats. SCFAs produced by bacterial fermentation of dietary fibre, in particular propionate, have been suggested to down regulate the activity of this enzyme (Chen et al., 1984) leading to the hypolipedemic effect

of dietary fibres. It has been established that the intake of fermentable fibres increases SCFA levels in the hepatic portal blood (Hara et al., 1998).

This study therefore established that all of the fibre-enriched diets were able to improve dyslipidemia in MSG-induced obesity in rats. The unripe Plantain flour diet was the most effective in bringing about body mass reduction in the obese rats. Also, the study provided an unequivocal demonstration of the lipid-lowering effects of the fibre-enriched diets used in this study in dyslipidemic MSG-induced obese rats. These effects were associated with modifications of plasma lipoprotein distribution, RBC lipid levels and HMG-CoA reductase activity.

## **Conflict of interest**

The authors declare that there is no conflict of interest.

#### References

- Adamson, I., Mbajiorgu, N., 1985. Tissue lipids and intestinal ATPases in rabbits fed cassava and plantain diets. Nutr. Res. 5, 1359–1373.
- Ademuyiwa, O., Ugbaja, R.N., Rotimi, S.O., 2008. Plasma lipid profile, atherogenic and coronary risk indices in some residents of Abeokuta in south-western Nigeria. Biokemistri 20 (2), 85–91.
- Chen, W.J., Anderson, J.W., Jennings, D., 1984. Propionate may mediate the hypocholesterolemic effects of certain soluble plant fibers in cholesterol-fed rats. Proc. Soc. Exp. Biol. Med. 175, 215–218.
- Dabai, F.D., Walker, A.F., Sambrook, I.E., Welch, V.A., Owen, R.W., Abeyasekera, S., 1996. Comparative effects on blood lipids and faecal steroids of five legume species incorporated into a semi-purified, hypercholesterolaemic rat diet. Br. J. Nutr. 75, 557–571.
- Das, U.N., 2010. Obesity: genes, brain, gut, and environment. Nutrition 26, 459-473.
- De Filippo, C., Cavalieri, D., Di Paola, M., Ramazzotti, M., Poullet, J.B., Massart, S., Collini, S., Pieraccini, G., Lionetti, P., 2010. Impact of diet in shaping gut microbiota revealed by a comparative study in children from Europe and rural Africa. Proc. Natl. Acad. Sci. USA 107, 14691–14696.
- Dolnikoff, M., Martin-Hidalgo, A., Machado, U.F., Lima, F.B., Herrera, E., 2001. Decreased lipolysis and enhanced glycerol and glucose utilization by adipose tissue prior to development of obesity in monosodium glutamate (MSG) treated-rats. Int. J. Obes. Relat. Metab. Disord. 25, 426–433.
- Folch, J., Lees, M., Sloane Stanley, G.H., 1957. A simple method for the isolation and purification of total lipides from animal tissues. J. Biol. Chem. 226, 497–509.
- Gallaher, D.D., Hassel, C.A., Lee, K.J., Gallaher, C.M., 1993. Viscosity and fermentability as attributes of dietary fiber responsible for the hypocholesterolemic effect in hamsters. J. Nutr. 123, 244–252.
  Gidez, L.I., Miller, G.J., Burstein, M., Slagle, S., Eder, H.A., 1982. Separation and
- Gidez, Ll., Miller, G.J., Burstein, M., Slagle, S., Eder, H.A., 1982. Separation and quantitation of subclasses of human plasma high density lipoproteins by a simple precipitation procedure. J. Lipid Res. 23, 1206–1223.
- Guimaraes, R.B., Telles, M.M., Coelho, V.B., Mori, R.C., Nascimento, C.M., Ribeiro, E.B., 2002. Adrenalectomy abolishes the food-induced hypothalamic serotonin release in both normal and monosodium glutamate-obese rats. Brain Res. Bull. 58, 363–369.
- Guven, S., El-Bershawi, A., Sonnenberg, G.E., Wilson, C.R., Hoffmann, R.G., Krakower, G.R., Kissebah, A.H., 1999. Plasma leptin and insulin levels in weight-reduced obese women with normal body mass index: relationships with body composition and insulin. Diabetes 48, 347–352.

- Hara, H., Haga, S., Kasai, T., Kiriyama, S., 1998. Fermentation products of sugar-beet fiber by cecal bacteria lower plasma cholesterol concentration in rats. J. Nutr. 128, 688–693.
- Kim, M.S., Kim, J.K., Kim, H.J., Moon, S.R., Shin, B.C., Park, K.W., Yang, H.O., Kim, S.M., Park, R., 2003. Hibiscus extract inhibits the lipid droplet accumulation and adipogenic transcription factors expression of 3T3-L1 preadipocytes. J. Altern. Complement Med. 9, 499–504.
- Kriketos, A.D., Furler, S.M., Gan, S.K., Poynten, A.M., Chisholm, D.J., Campbell, L.V., 2003. Multiple indexes of lipid availability are independently related to whole body insulin action in healthy humans. J. Clin. Endocrinol. Metab. 88, 793–798. Kritchevsky, D., 1988. Dietary fiber. Annu. Rev. Nutr. 8, 301–328.
- Madar, Z., Odes, H.S., 1990. Dietary fiber research. Prog. Biochem. Pharmacol. 24 (III-VIII), 1–137.
- Martinkova, A., Lenhardt, L., Mozes, S., 2000. Effect of neonatal MSG treatment on day-night alkaline phosphatase activity in the rat duodenum. Physiol. Res. 49, 339–345.
- Mazur, A., Remesy, C., Demigne, C., 1990. The effect of high-fibre diet on plasma lipoproteins and hormones in genetically obese Zucker rats. Eur. J. Clin. Invest. 20, 600–606.
- Mehta, K., Kaur, A., 1992. Reviews: Dietary fibre. Intl. J. Diab. Dev. Countries 12, 12– 18.
- Mozes, S., Sefcikova, Z., Lenhardt, L., Racek, L., 2004. Obesity and changes of alkaline phosphatase activity in the small intestine of 40- and 80-day-old rats subjected to early postnatal overfeeding or monosodium glutamate. Physiol. Res. 53, 177– 186.
- Muoio, D.M., Newgard, C.B., 2006. Obesity-related derangements in metabolic regulation. Annu. Rev. Biochem. 75, 367–401.
- Nagata, M., Suzuki, W., Iizuka, S., Tabuchi, M., Maruyama, H., Takeda, S., Aburada, M., Miyamoto, K., 2006. Type 2 diabetes mellitus in obese mouse model induced by monosodium glutamate. Exp. Anim. 55, 109–115.
- Nakagawa, T., Ukai, K., Ohyama, T., Gomita, Y., Okamura, H., 2000. Effects of chronic administration of sibutramine on body weight, food intake and motor activity in neonatally monosodium glutamate-treated obese female rats: relationship of antiobesity effect with monoamines. Exp. Anim. 49, 239–249.
- Novelli, E.L., Diniz, Y.S., Galhardi, C.M., Ebaid, G.M., Rodrigues, H.G., Mani, F., Fernandes, A.A., Cicogna, A.C., Novelli Filho, J.L., 2007. Anthropometrical parameters and markers of obesity in rats. Lab Anim. 41, 111–119.
- Olumakaiye, M.F., Atinmo, T., Olubayo-Fatiregun, M.A., 2010. Food consumption patterns of Nigerian adolescents and effect on body weight. J. Nutr. Educ. Behav. 42, 144–151.
- Ramdath, D.D., Isaacs, R.L., Teelucksingh, S., Wolever, T.M., 2004. Glycaemic index of selected staples commonly eaten in the Caribbean and the effects of boiling v. crushing. Br. J. Nutr. 91, 971–977.
- Rao, A.V., Ramakrishnan, S., 1975. Indirect assessment of hydroxymethylglutaryl-CoA reductase (NADPH) activity in liver tissue. Clin. Chem. 21, 1523–1525.
- Rose, H.G., Oklander, M., 1965. Improved procedure for the extraction of lipids from human erythrocytes. J. Lipid Res. 6, 428–431.
  Sandkamp, M., Funke, H., Schulte, H., Kohler, E., Assmann, G., 1990. Lipoprotein(a) is
- Sandkamp, M., Funke, H., Schulte, H., Kohler, E., Assmann, G., 1990. Lipoprotein(a) is an independent risk factor for myocardial infarction at a young age. Clin. Chem. 36, 20–23.
- Stewart, J.C., 1980. Colorimetric determination of phospholipids with ammonium ferrothiocyanate. Anal. Biochem. 104, 10–14.
- Superko, H.R., Haskell, W.L., Sawrey-Kubicek, L., Farquhar, J.W., 1988. Effects of solid and liquid guar gum on plasma cholesterol and triglyceride concentrations in moderate hypercholesterolemia. Am. J. Cardiol. 62, 51–55.
- Thian, I., Samba, K., Lwanga, D., 2006. Diet related chronic disease in West African region. SCN News 33, 5.
- Wolever, T.M., Spadafora, P., Eshuis, H., 1991. Interaction between colonic acetate and propionate in humans. Am. J. Clin. Nutr. 53, 681–687.
- and propionate in humans. Am. J. Clin. Nutr. 53, 681–687. Zhang, W.M., Kuchar, S., Mozes, S., 1994. Body fat and RNA content of the VMH cells in rats neonatally treated with monosodium glutamate. Brain Res. Bull. 35, 383– 385.