Int.J.Curr.Res.Aca.Rev.2014; 2(8):138-158



International Journal of Current Research and Academic Review

ISSN: 2347-3215 Volume 2 Number 8 (August-2014) pp. 138-158 www.ijcrar.com



Probiotics in the Management of Diseases: A Review

S.Oranusi*, O.M.Adedeji, and B.K. Olopade

Department of Biological Sciences, Covenant University, Ota, Ogun State, Nigeria **Corresponding author*

KEYWORDS	A B S T R A C T
Probiotics, Management of Diseases, Immunomodulations, Saccharomyces boulardii, Clostridium butyricum, and Streptococcus salivarius	In recent years, there has been an increase in the application of probiotics for the treatment of some diseases and to alleviate the symptoms of many others. Diseases and ailments such as diarrhea, pouchitis, cancer, ulcerative colitis, irritable bowel disease, and a host of others have experienced an increase in the use of certain probiotics bacteria to combat them. The complete mechanisms of action of probiotics in disease management and enhancement of the health of the host remain largely unknown, but the major activity thus appear to be via modulation of immune responses (immunomodulations) and colonization competitive shielding off of pathogens. This paper is set to review some of the various ailments for which probiotics have been used. With an upsurge in the use of probiotics, also comes an increase in skeptism on the safety of their use for medical purpose, the safety concerns that may obstruct effective usage and thus judicious application of probiotics in disease management warrants further investigations.

Introduction

defined Probiotics are as live microorganisms which when administered in adequate amounts confer a health benefit on the host (FAO/WHO, 2002). They are live nonpathogenic preparation administered to improve and restore the microbial balance of gastrointestinal tract. Probiotics. as biological factors, control the gut microbiota and result in its progression. They are organisms which are generally regarded as safe (GRAS) and consumed without the risk of infections (FAO/WHO, 2002). Most organisms used as probiotics belong to the lactic acid bacilli, Lactobacillus and Bifidobacterium, a nonpathogenic E. coli

strain (*E. coli* Nissle 1917), *Saccharomyces boulardii*, *Clostridium butyricum*, and *Streptococcus salivarius* subspecies *thermophilus*, genetically engineered bacteria that secrete immunosuppressive substances such as interleukin-10 (IL-10) have been studied (Sartor, 2004).

Probiotics are commonly consumed as part of fermented foods with specially added active live cultures, such as in yogurt, soy yogurt, cheese or as dietary supplements. Probiotics may beneficially affect the host by augmenting its intestinal microbial population beyond the amount already existing, thus possibly inhibiting pathogens. Specific attributes that position an organism to be an effective probiotics include acid tolerance, bile tolerance, cell surface hydrophobicity, protoplast regeneration, antimicrobial activity, cholesterol removal and bile salt deconjugation, gut colonization, lactose removal, protease and amino peptidase activity (Sartor, 2004; Sudah et al., 2009; Scaldaferri et al., 2013). To be functional in the intestinal tracts, probiotics are expected to be viable and in a certain number. As such the modes of delivery and production should be targeted towards maintaining the viability of the organism after production and even during storage.

Probiotic organisms may be naturally occurring microbes (as is the case for all used in food), or microbes that have been genetically altered for a specific effect. The complete mechanism of action of probiotics in disease management is not known, however, the major activity thus appear to be via colonization competitive shielding off of pathogens and immunomodulations. Several research effort have explained these mechanism of activities to include improving gastrointestinal tract health via modifying gut pH, antagonizing pathogens through production of antibacterial compounds, competitive exclusions of pathogens at the binding and receptor sites, enhancing the immune system, synthesizing enhancing the bioavailability and of nutrients, competing for available nutrients, symptoms reducing of the lactose intolerance, decreasing the prevalence of allergy in susceptible individuals, and reducing risks of certain cancers through binding of deleterious mutagens and carcinogens (Hirayama and Rafter 2006; Jain et al., 2010).

In the present review article we highlight some of the diseases for which probiotics have been used to manage and ameliorate conditions with a view to steering interests for further probe into specific mechanisms of activity of probiotics and its possible use in the management of diseases hitherto not perceived/conceived. For more in-depth view on the diseases highlighted, the reader is kindly asked to refer to specific publications.

Probiotics in the treatment/ management of diarrheal diseases

Diarrhea has been defined as the passage of 3 or more loose or liquid stools per day, or more frequently than is normal for the individual. It is usually a symptom of gastrointestinal infection, which can be caused by a variety of bacterial, viral and parasitic organisms, diarrhea could also be antibiotic associated (WHO, 2013). Diarrhea leads to fluid loss, and may be lifethreatening, particularly in young children and people who are malnourished or have impaired immunity (WHO, 2013). Reports abound on the use of probiotics for treatment of antibiotic associated diarrhea (AAD) and the management of chronic and acute enteric infections and their associated diarrheal complexes.

The rationale for using probiotics for treatment of diarrhea is based on the observation that they act either by modifying the composition of colonic microbial flora or by modulating the immune response. Lemberg et al. (2007) penned that probiotics modify the composition of microbial flora by several mechanisms which include: Competing with pathogens for nutrients and receptors; Inducing hydrolysis of toxins and Inducing receptors: production of substances antimicrobial including peptides of the innate immune system; Inducing production of organic acids and modulation of nitric oxide synthesis. Seth et

al. (2008) opined regulating intestinal permeability by modulating the epithelial tight junctions as a mechanism of action while Walker (2008) noted the exerting of a tropic action on the intestinal mucosa, which leads to brush border enzyme activation, stimulation of glucose absorption and anti-apoptotic effects on the enterocyte as one of the action mechanisms of probiotics in the management of diarrhea.

McFarland (2009) submitted that antibiotic associated diarrhea (AAD) is a common complication of most types of antibiotics, especially for broad-spectrum antibiotics such as clindamycin, beta-lactams and 3rd generation cephalosporins. The nonselective action of antibiotics perturb the normal microbial flora of GIT, which leads overgrowth and multiplication to of pathogenic microorganisms i.e, Clostridium difficle, Clostridium perfringens, Staphylococcus aureus, Klebsiella oxytoca, Candida spp and Salmonella spp., which ultimately result in the production of toxin, leading to diarrhea (McFarland, 2009). .

Perturbation of normal microbial flora leads to the decrease in the number of bacteria involved in the production of short chain fatty acids. These fatty acids are important for the nutrition of the enterocyte and for water and electrolyte absorption, and their decrease may result in watery diarrhea. Erythromycin special is case of antimicrobial agents which directly stimulate the motilin and induce the diarrhea (McFarland, 2009).

Kotowsha *et al.* (2005) observed that oral administration of selected probiotics strain along with antibiotics can be the better option to handle the situation of antimicrobial associated diarrhea. In a clinical study, Kotowsha *et al.* enrolled 269 children who were taking antibiotics for ear or respiratory infections and randomized them to either *Saccharomyces boulardii* (500 mg/d) or placebo for the duration of the antibiotic treatment. Even though the follow-up time was short (two weeks post-antibiotic), the frequency of diarrhea in the probiotic group was significantly less (3.4%) compared to 17.3% in the placebo group (Kotowsha *et al.*, 2005).

Clostridium difficile-associated diarrhoea (CDAD) is most often caused bv cephalosporins, clindamycin, ampicillin, amoxicillin. Clostridium difficile and been associated with symptomatic has diarrhoea since being identified as the responsible pathogen for pseudomembranous colitis (Santosa et al., 2006). Total flora replacement or faecal bacterio therapy has been described as an effective treatment alternative in severe C. difficile infections. It is based on transfer of faecal flora from a healthy individual to a severely ill patient. Total flora replacement has also been used to manage severe constipation, irritable bowel syndrome, and inflammatory bowel disease. Homologous faecal enemas have been used in recalcitrant cases of CDAD usually with stool donated by the patient's partner. L. acidophilus and L. rhamnosus are probiotics that have been detected in these fecal samples. It is an adjunctive therapy in sporadic clinical use. (Borody et al., 2003; 2004).

Traveler's diarrhea occurs in about half of travelers who visit high-risk areas. Although most cases are mild and self-limiting, there is considerable morbidity. Antibiotics are an effective means of prophylaxis but are not recommended for widespread use. Hence there is a need for cost-effective alternative treatments. The efficacy of probiotics in traveler's diarrhea has been reported with *Saccharomyses boulardii* and *Lactobacillus rhamnosus* GG shown to have significant effects. *S. boulardii* seems to prevent bacterial diarrhoea more effectively, while *Lactobacillus rhamnosus* GG has been shown to be more effective against viral and idiopathic diarrhea (Goldin and Gorbach, 2008).

Infective diarrhea due to Rotavirus is the of infantile leading cause diarrhea worldwide and rapid oral rehydration is the primary treatment. Several potential mechanisms have been proposed for how lactobacilli reduce the duration of rotavirus diarrhea through competitive blockage of receptor sites in which lactobacilli bind to receptors, enhanced immune response, and signal(s) from lactobacilli that regulates the secretary and motility defenses designed to remove perceived noxious substances and lactobacilli that produce substances meant to inactivate the viral particles. The probiotics most frequently studied for treating acute diarrhea include Lactobacillus rhamnosus GG and Lactobacillus reuteri (Santosa et al., 2006, Goldin and Gorbach, 2008).

Probiotics in inflammatory bowel disease (IBD)

Inflammatory bowel disease (IBD) is a collective term, used for ulcerative colitis (UC), Crohn's disease (CD) and Pouchitis. IBD is an abnormal immune response against luminal antigen of commensal genetically predisposed bacteria in individuals (Sartor, 2004; Fedorak et al., 2004; Gionchetti et al., 2005; Marteau et al., 2009). Traditionally known medication used in IBD includes 5-aminosalicylic acid (5-ASA) and corticosteroids. Limited clinical trials suggest that selected probiotics species, alone or in combination, can prevent recurrent intestinal inflammation and possibly treat active IBD, with best results in pouchitis, and, to a lesser extent, ulcerative colitis and Crohn's disease (Table 1 adapted from Sartor, 2004).

Several probiotics mechanisms of action, relative to inflammatory bowel disease, have been elucidated: (1) competitive exclusion, whereby probiotics compete with microbial pathogens i. e. colonization resistanceoccupy ecologic niche. (2)immunomodulation and/or stimulation of an immune response. Alter immunoregulation by induce IL-10, transforming growth factor expression and secretion, stimulate secretory immunoglobulin A production, decrease tumor necrosis factor expression ; (3) antimicrobial activity and suppression of pathogen growth, inhibit pathogenic enteric bacteria via decrease luminal pH, secrete bacteriocidal proteins,; (4) enhancement of barrier activity. Improve epithelial and mucosal barrier function, (Produce short chain fatty acids, including butyrate. Enhance mucus production and increase barrier integrity; and (5) induction of T cell apoptosis, block epithelial bindinginduction of MUC 2 inhibit epithelial invasion (Rioux and Fedorak, 2006; Marteau et al., 2009; Scaldaferri et al., 2013).

Fujmori et al.(1997) reported that in Crohn's disease, commensal E. coli strain stimulates the release of tumor necrosis factor- α (TNF- α) and interleukin-8 (IL-8) by inflamed mucosa. However, some lactobacilli strains including L. casei down regulate the spontaneous releases of TNF- α and the inflammatory response induced by E.coli. Therefore, it makes sense either to eliminate some bacteria with antibiotics or to alter the gut flora, in favor of more beneficial bacteria, by the use of probiotics and prebiotics (prebiotics are dietary non-digested substances usually carbohydrates that stimulate the growth and metabolism of protective commensal enteric bacteria) (Fujmori et al., 1997; Scaldaferri et al., 2013).

Author (date)	Probiotic	Clinical situation	Result
Crohn's disease			
Plein, Hotz (1993)135	Saccharomyces boulardii	Maintenance of remission	↓ diarrhea vs. placebo
Malchow (1997)93	<i>E. coli</i> Nissle 1917	Maintenance of remission	↓ relapse vs. placebo
Guslandi (2000)136	S. boulardii	Maintenance of remission (probiotic mesalamine vs. mesalamine alone)	 ↓ relapse vs. ↓ mesalamine alone
Prantera (2002)94	Lactobacillus GG	Postoperative prevention	No benefit
Ulcerative colitis		-	
Kruis (1997)85	<i>E. coli</i> Nissle 1917	Maintain remission	Equal to mesalamine (1.6 g)
Rembacken (1999)86	<i>E. coli</i> Nissle 1917	Maintain remission	Equal to mesalamine
Kruis (2001)87	<i>E. coli</i> Nissle 1917	Maintain remission	Equal to mesalamine
Ishikawa (2003)88	Bifidobacteria- fermented milk	Maintain remission	Superior to placebo
Pouchitis			
Gionchetti (2000)77	VSL#3	Maintain remission chronic pouchitis	Superior to placebo
Mimura (2002)83	VSL#3	Maintain remission chronic pouchitis	Superior to placebo
Gionchetti (2003)84	VSL#3	Prevention after ileostomy closure	Superior to placebo

Table.1 Randomized Double-Blind Trials of Probiotic Agents in IBD

The proposed mechanisms of action of probiotics in the management of Crohn's disease include changes in short chain fatty acids (SCFA) production patterns; reduction in pro-inflammatory cytokine secretion, improving Th1/Th2 ratios; Eliminating pathogens; enhancement of barrier function (Fujmori *et al.*, 1997).

In a study by Malchow, 28 patients of active CD were treated with a tapering dose of prednisolone and either placebo or *E. coli* Nissle (*E. coli* Nissle is a nonpathogenic *E. coli*, which colonize the intestine and inhibits the growth of enteropathogenic and

other enteric bacteria. It is proposed that this organism, by suppressing enteropathogenic bacteria, may have a long-term effect to suppress remission). The *E. coli* Nissle was given in an increasing dosage over 24 days to the final dose of 5×10^{10} bacteria per day for a year. The patients are assessed for the remission .There was higher relapse rate in the placebo group (63.6%) as compared to the *E. coli* group (33.3%) (Malchow, 1997).

Fujimori *et al.*(1997) treated 10 patients of Chorn's disease not responding to 5-ASA and prednisolne therapy with a synbiotic therapy, (Probiotic - *Bifidobacterium* and *Lactobacillus* and Prebiotic- psyllium), for 12 months. Six patients went into remission, one had a partial response with improvement of the number bowel movements, and three patients were non-responders. There was no significant difference between C-reactive protein (CRP) and erythrocyte sedimentation rate (ESR) values were observed before and after the therapy (Fujimori *et al.*, 1997).

In ulcerative colitis (UC), the composition of microflora is imbalanced associated with increased number of pro-inflammatory bacteria, including Enterobacteriaceae, and Bacteroides fragilis and decreased count of protective bacteria, including lactobacilli and bifidobacteria. Furrie et al. (2005) documented the management of ulcerative colitis via probiotics. In a double blind study for patients with active UC, employing a combination probiotics of strain. Bifidobacterium longum, with a prebiotic composed of an inulin-oligofructose (growth substrate) on 18 patients. Nine patients assigned to the treatment group and nine to the placebo group. Patients assessed with a activity index, clinical level of gut inflammatory markers and histological sore. The patients were treated with the synbiotic mixture (probiotics- prebiotic) of placebo twice-daily for 4 weeks. At the end of the month, the patients were reassessed. The patients receiving the synbiotic mixture exhibited reduced mucosal inflammatory markers in colonic biopsies. There was an improvement microscopically .There is significant reduction in TNF- α and IL- α levels in mucosal biopsies in patients treated with the active therapy as compared to placebo (Furrie et al., 2005).

Pouchitis is an idiopathic inflammatory disease of the ileal pouch that occurs in 15% – 53% of patients who undergo total abdominal colectomy with ileal pouch-anal anastomosis (IPAA) for ulcerative colitis. Fecal stasis with immunologic reactivity

important in the appears to be pathogenesis of this disease (Tursi et al., 2004). Welters et al. (2002) and Kim and Hans (2004) demonstrated that using the dietary fiber Inulin (24 g/day), for the short period of 3 weeks significantly decreases the pouchitis as measured by endoscopic and histologic score. The results correlated with a significant alteration of fecal pH, fecal butyrate, and secondary bile acid concentration.

In a randomized, double-blind, placebocontrolled study 40 patients were randomized within a week after surgery to receive either VSL#3 (3 g, 9 x 10^{11} bacteria/day) or placebo for 12 months (Welters et al., 2002). Two of 20 (10%) of the patients treated with VSL#3 developed an episode of acute pouchitis compared to 8/20 patients (40%) in the placebo group (P < 0.01). Patients treated with VSL#3 and no signs of pouchitis had a median stool frequency of 5 (range, 3–9) at the end of the trial compared to 8 (range, 6 - 12) in the placebo group (with no signs of pouchitis; P < 0.001) (Welters *et al.*, 2002).

Probiotics in Genitourinary tract infection

The genital tract of premenopausal woman mainly composed of Lactobacilli, especially Lactobacillus crispatus and Lactobacillus iners. These lactobacilli engaged in the production of bacteriocins, lactic acid and hydrogen peroxide, which maintain the pH acidic and keep away the pathogenic colonies from proliferating. Imbalanced microbial flora offer an opportunity for the proliferation of pathogenic microorganism such as E. coli, Gardnerella vaginalis. **Bacteroides** sp, beta-Streptococci Falcivibrio Mobiluncus or spp. and Candida albicans (Hillier et al., 1993; Cauci et al., 2003; Anukam et al., 2006a.b).

Unnecessary receive of broad-spectrum antibiotic (excreted out from urine), disrupt normal flora, and causes symptomatic urinary tract infections such as bacterial vaginosis and candidal vulvovaginitis ((Cauci *et al.*, 2003) Restoring the normal flora with lactobacilli may help to treat these genital infection.

The treatment of UTI usually is with oral antimicrobial regimens of clindamycin, metronidazole or nofloxacin for bacterial vaginitis and fluconazole for candidal vaginitis. Nevertheless, their nonselective antimicrobial action decrease the lactobacilli count, and alter the pH (shift to alkaline), which ultimately associated with recurrence of the infection. Use of effective probiotics could be the best option to manage the above condition and to break the cycle of recurrence.

In a clinical study conducted on 28 women suffering from recurrent candidal vulvovaginitis, Hilton et al. (1992) reported that administration of vaginal suppositories cantaining Lactobacillus rhamnosus GG twice daily for seven days, all of the women reported an improvement vaginal in and reduction vaginal symptoms in erythema and discharge.

Studies have shown that women with bacterial vaginosis (no lactobacilli) are at significantly increased risk of HIV (Falagas et al., 2006; Anukam et al., 2006a,b).The prevention or resolution of bacterial vaginosis is particularly important in women at risk of human immunodeficiency virus (HIV) infection. Thus, treatment of bacterial and promotion of vaginosis vaginal lactobacilli may reduce a woman's risk of acquiring HIV-1, gonorrhea, and trichomoniasis. A recent publication has shown that a human vaginal probiotic strain (Lactobacillus reuteri RC-14) can express potent functional viral inhibitors which may potentially lower the sexual transmission of HIV (Liu *et al.*, 2006).

A study constituting 13 women showed that consumption of yogurt containing L. acidophilus decreased the incidence of C. albicans yeast infections (Klebanoff et al., 1991). Hydrogen peroxide production is a key factor in resisting BV disease (Klebanoff et al., 1991). Hydrogen peroxide producing strains of lactobacilli have been found in 61% of pregnant women with normal flora and in only 5% of women with BV. Hydrogen peroxide has been shown to be toxic to BV causing organisms, namely, Gardnerella vaginalis and Prevotella bivia (Hillier et al., 1993). Comparable results were obtained in open and placebo controlled studies in which lyophilized L. acidophilus was applied locally or L. acidophilus yogurt was given orally (Parent et al., 1996, Hallén et al., 1992).

In these studies, success rates for control of BV or Candida vaginitis ranged from 57% to 87% in the probiotic group and from 0 to 22% in the control group (Hallén et al., 1992). Various molecular methods have shown L. crispatus and L. johnsonii to be the most common vaginal isolates from "normal" women of child- bearing age (Reid et al., 1996) The administration of L. rhamnosus GR-1 in combination with L. fermentum B-54 and RC-14 by mouth and intravaginally has been shown to be safe and to reduce the risk of UTIs, BV, and yeast vaginitis (Reid et al., 1996). As with urogenital pathogens, lactobacilli ascend from the rectum into the vagina and subsequently alter the microenvironment and potentially modulate the immunologic status of the host such that a normal vaginal flora is more often restored and retained (Gardiner et al., 2002, Cadieux et al., 2002).

Probiotics in Hypercholesterolemia and Hypertension

Cholesterol is a precursor to certain hormones and vitamins and is a component of cell membranes and nerve cells. However, elevated levels of total blood cholesterol (Hypercholesterolemia) or other blood lipids are considered to be a high risk factor for coronary heart disease one of the leading of causes death. Different mechanisms by which probiotics preparations could control blood cholesterol level have been proposed to include assimilation of cholesterol in the small intestine by probiotics and incorporation of cholesterol into the cell membrane of probiotics (Noh et al., 1997; Taranto et al., 1999;2000; Liong and Shah, 2005a).The cholesterol-lowering effect of lactic acid bacteria (Streptococcus, Lactobacillus, and *Bifidobacterium*) is well established (Nguyen et al., 2007). It has been found that the isolated lactic acid bacteria had an excellent hypocholesterolemic effect. Some strains of lactobacilli have been found to remove cholesterol via various mechanisms and can be used as a dietary adjunct to lower serum cholesterol in vivo (Liong and Shah., 2005b).

L. acidophilus deconjugates bile acids into free acids that are rapidly excreted from the intestinal tract. Because free bile salts are excreted from the body, the synthesis of new bile acids from cholesterol lowers its concentration in the body. Further, it has been suggested by Andersson et al.(1995). that the bile flow is stimulated by regular milk consumption (1 L/day). Isolates of L. acidophilus from human intestine are better able to assimilate cholesterol and actively deconjugate bile salts than commercially cultures of L. acidophilus. used Lactobacillus plantarum PH04 and L. cholesterol-lowering reuteri showed

activities (De Smet *et al.*, 1998; Nguyen *et al.*, 2007). Clinical investigation have suggested a decrease in serum cholesterol concentrations during consumption of very large amounts (8 L/day) of yogurt or fermented milk per day (Hepner *et al.*, 1979) Hypercholesterolemia is one of the major causes for hypertension (Liong and Shah, 2005b).

Probiotics have potential to regulate blood cholesterol level in hypercholesterolemia. The elevation of blood pressure is found to be greatly induced when total cholesterol level exceeds 6.4 mmol/L. This may increase cardiac output and peripheral vascular resistance that causes an elevated blood pressure (Noh et al., 1997; Taranto et al., 1999; Liong and Shah, 2005b). Therefore, lipid metabolism disorders are often the causes of hypertension. A variety of in vitro experiments and in vivo trials have provided experimental evidence to support the roles of probiotics in lowering serum cholesterol and improving lipid profiles, which subsequently leads to a reduced risk of hypertension (Noh et al., 1997; Taranto et al., 1999; Liong and Shah, 2005b).

Probiotics in oral and dental diseases

The application of probiotics in the management of dental caries (Meurman *et al., 1995;* Näse *et al., 2001;* Ahola *et al., 2002;* Kang *et al., 2005;* Stamatova *et al., 2007;* Caglar *et al., 2006; 2007; 2008a,b;* Cildir *et al., 2009),* periodontal diseases (Grudianov *et al., 2002;* Volozhin *et al., 2004;*), halitosis (Kazor *et al., 2006; Burton et al., 2006;* Kang *et al., 2006;*) and oral candidiasis (Elahi *et al., 2005;* Hatakka *et al., 2007)*have been reported.

The proposed mechanisms of action of probiotics might be due to the competition

for binding sites in oral biofilms. strengthening the mucosal barrier via tropic effects on the epithelium, and stimulating both the innate and adaptive immune system. The ability of probiotics in the reduction of S. mutans and other oral streptococci with cariogenic potential abound.(Meurman et al., 1995; Kang et al., 2005; Stamatova et al., 2007; Caglar et al., 2008a, b; Çaglar et al., 2007; Çaglar et al., 2006; Cildir et al., 2009.)

Α randomized. double-blind, placebocontrolled intervention study examined the effect of milk containing L. rhamnosus GG on caries and the risk of caries in children when compared with normal milk (Näse et al., 2001), probiotic milk was able to reduce S. mutans counts at the end of the trial and a significant reduction of caries risk was also observed. The putative caries prophylactic effect of probiotics has been also confirmed by daily intake of cheese containing L. rhamnosus GG and L. rhamnosus LC 705(Ahola et al., 2002). The probiotic cheese significantly reduced S. mutans counts in the intervention group during the post-treatment period when compared with the controls. Another probiotic species, Bifidobacterium DN-173 010, ingested once daily with yogurt demonstrated a significant reduction of salivary S. mutans, whereas no reduction was significant found in lactobacilli levels (Çaglar et al., 2005).

Periodontal inflammation has been reduced and also positively affected by the administration of two probiotic tablet forms Bifidum bacteria and lactic acid bacteria available on the Russian market (Grudianov et al., 2002). Studies have also shown that a periodontal dressing containing *L. casei* can reduce the number of most common periodontal pathogens and extend remission up to 10–12 months (Volozhin et al., 2004).

Bad breath in the oral cavity (Halitosis or foetor ex ore} is mainly ascribed to the production of volatile sulfur compounds (VSC) predominantly by Gram negative anaerobes residing in periodontal pockets and on the tongue dorsum. The replacement of bacteria implicated in halitosis by colonization with probiotic bacterial strains from the indigenous oral microbiota of healthy humans may have potential application as adjuncts for the prevention and treatment of halitosis. Kazor et al. (2003) reported that L. salivarius was the most predominant species detected in healthy subjects, whereas it was detected in only one of the subjects with halitosis at very low levels .The rationale of probiotic implementation in cases of halitosis has been documented in several studies. S. salivarius K12 taken in a lozenge after a mouthwash could reduce oral VSC levels in 85% of the subjects in the test group (Burton et al., 2005). Weissela cibaria was also reported to being able to reduce VSC production both in vitro and in vivo (Kang et al.,2006a). A contributing factor to malodor reduction can be the ability of W. cibaria to co-aggregate with species renowned for their VSC production (F. nucleatum, for example), thus reducing the source for malodorous compounds in the oral cavity (Kang *et al.*,2006b).

Hatakka et al.(2007) reported that probiotic applications in the oral cavity have the potential to alleviate symptoms and reduce pathogenic potential of Candida species. They observed that a 16-week probiotic intervention demonstrated study а significant reduction by 75% of high yeast counts in the elderly and Hyposalivation reduction was also observed by the intake of GG containing L. rhamnosus cheese associated with control of oral Candida.

Elahi *et al.*(2005), Wagner *et al.*(2000) *in vivo* studies on mice have shown that lactobacilli might indeed be effective in controlling oral candidiasis. A higher clearance of *C.albicans* in mice fed with *L. acidophilus* compared to control group was demonstrated, however, no noticeable delay in colonization of the oral cavity by *C. albicans* of immunocompromized mice was achieved when heat killed *L. casei* and *L. acidophilus* cells were given

and Stamatova Meurman, (2009)in controlled trials randomized have nevertheless shown that probiotics may control dental caries in children due to their inhibitory action against cariogenic observed streptococci. They that less evidence exists on their role in periodontal disease or oral yeast infections.

Probiotics in Kidney diseases

The number of patients with chronic kidney disease (CKD) is rising worldwide and it is now being recognized as a major public health concern. Natarajan et al. (2009; 2010) observed oral administration of a probiotic formulation of selected microbial strains extend renoprotection via intraintestinal extraction of toxic waste solutes in patients with chronic kidney disease (CKD). The main outcomes of this investigation include a significant reduction of BUN (blood urea nitrogen, serum creatinine, and uric acid.), enhanced well-being, and absence of serious adverse effects (Natarajan et a.l., 2010), thus supporting the use of the chosen probiotic formulation for bowel-based toxic solute extraction.

Lieske *et al.*(2005), Hoesl and Altwein,(2005) noted that a probiotic preparation able to degrade oxalate in vitro was shown to reduce oxalate fecal excretion. A high level of oxalate in the urine is a risk

factor for development of kidney stones. Several probiotics preparations induce protective cytokines, including IL-10, and suppress proinflammatory cytokines, such as TNF- α and IL-6. Intestinal microflora is deranged in hemodialysis (HD) patients as an increase in aerobic bacteria such as E. coli and a decrease in anaerobic bacteria such as Bifidobacterium. One study reported that oral administration of Bifidobacterium longum in a gastroresistant seamless capsule decreases the the pre-HD serum levels of homocysteine and indoxyl sulfate. It has also been shown that synbiotics containing lactobacilli can reduce serum level of p-Cresol in HD patients. High-serum p-cresyl sulfate and indoxyl sulfate levels were associated with renal progression. Serum concentrations of p-cresol are independently associated with overall mortality and cardiovascular disease in HD patients.

In-vitro and in-vivo investigations undertaken by Kibow Biotech (2012) suggest that oral administration of a probiotic formulation comprised of selected microbial strains may extend renoprotection via intra intestinal extraction of toxic solutes in patients with CKD stages 3 and 4. They opined that science has defined more than 100 uremic toxins that may be involved in Chronic Kidney Disease and as well in CKD Stage 5, also known as end-stage renal disease (ESRD), and that certain probiotic microorganisms can utilize urea, uric acid and creatinine and other toxins as its nutrients for growth. Overloaded and impaired kidneys have a buildup of these poisonous wastes in the bloodstream, probiotics microorganisms multiply, thereby creating a greater diffusion of these uremic toxins from the circulating blood across the lining of the intestinal walls into the bowel. This increased microbial growth is excreted along with the feces (which is normally 50% microbes by weight) (Kibow Biotech, 2012)

Probiotics in Chronic fatigue syndrome (CFS)

CFS is a medically unexplained illness, characterized by persistent and relapsing fatigue, in addition to cognitive dysfunction, headaches, joint pains, and central nervous system disturbances (Komaroff et al., 1996). Aaron et al. (2000) observed that many CFS patients also complain of gastrointestinal (GI) disturbances and are more likely to report a previous diagnosis of irritable bowel syndrome (IBS), meet diagnostic criteria for IBS and experience IBS related symptoms. Whitehead et al. (2002) noted that while CFS is neither a gastrointestinal nor psychiatric disorder per se, over 50 percent of patients with CFS meet the diagnostic criteria of IBS, and anxiety itself is often a hallmark symptom in those with IBS. This corroborate the discovery that gut pathogens in the GI tract can communicate with the central nervous system and influence behavior associated with emotion, anxiety in particular, even at extremely low levels and in the absence of an immune response (Lyte et al., 1998; Goehler et al., 2007).

Probiotics, or live microorganisms which confer a health benefit on the host, have the potential to influence mood-regulating systemic inflammatory cytokines, decrease oxidative stress and improve nutritional status when orally consumed (Logan et al., 2003). Several researches indicate that there are marked alterations in the intestinal microflora of CFS patients, including a lowered level of bifidobacteria and small intestinal bacterial overgrowth (Butt et al., 1998: Logan et al., 2003: Rao et al. 2009) It been observed the has that oral administration of Lactobacillus casei strain Shirota (LcS) caused a significant rise in fecal Bifidobacteria spp and Lactobacillus spp in CFS and the administration of Lactobacillus casei strain Shirota (LcS) or

placebo to otherwise healthy adults, reveals probiotics helped for mood regulation. Those with the lowest scores in the depressed/elated dimension at baseline had significant improvement in mood scores after taking the probiotic compared to the placebo group. The probiotic bacteria and placebo were unable to make a difference in those with the highest baseline mood scores (Benton et al., 2007; Rao et al., 2009). In a similar study using the animal model of depression, the oral administration of a probiotics was shown to increase plasma tryptophan levels, decrease serotonin metabolite concentrations in the frontal dopamine metabolite cortext and concentrations in the amygdaloid cortex (Desbonnet et al., 2008).

Probiotics in Diabetes

Diabetes mellitus has been described as a metabolic disease associated with a series of multiple risk factors that can be effectively managed by multifactorial interventions including dietary manipulations. Several research findings (Esteve et al., 2011; Kootte et al., 2012; Panwar et al., 2013; Stachowicz and Kiersztan, 2013), observed that in addition to risk factors such as genetic predisposition, epigenetic changes unhealthy lifestyle, and altered gut microbiota is a major risk factor because it adiposity. cause increased β-cell hyperglycemia, dysfunction, hypercholesterolemia, dyslipidaemia, metabolic systemic endotoxemia, inflammation, intestinal permeability (leaky gut), defective secretion of incretins and oxidative stress associated with type 2 diabetes (T2D). It is also a major risk factor in type 1 diabetes (Calcinaro et al., 2005). The influence of gut microbiota on health and disease has been established (Ulisse et al., 2001; Schultz et al., 2002; Vaarala, 2003; Sing et al., 2009), results from several

genomic, metagenomic and metabolomic studies have provided substantial information to target gut microbiota by dietary interventions for the management of T2D (Panwar et al., 2013). Probiotics particularly lactobacilli and bifidobacteria have emerged the as prospective biotherapeutics with proven efficacy to abrogate progression and development of diabetes through improving the altered gut microbial composition and by targeting all the possible risk factors.

Calcinaro et al. (2005) reported that orally administered probiotic compound VSL#3 (a probiotic compound containing $3 \times 10^{11}/g$ lyophilised viable bacteria, including bifidobacteria (B. longum, B. infantis and B. breve), lactobacilli (L. acidophilus, L. casei, L. delbrueckii subsp. L. bulgaricus and L. plantarum) and Streptococcus salivarius *thermophilus.*) prevented subsp. autoimmune diabetes induces and immunomodulation by a reduction in insulitis severity in a mice. They opined that the results provide a sound rationale for future clinical trials of the primary prevention of type 1 diabetes in man by oral VSL#3 administration. Similarly, Matsuzaki et al.(1997) reported that oral administration of heat killed Lactobacillus casei to nonobese diabetic (NOD) mice reduces the incidence of diabetes, but the mechanism underlying this finding has not been elucidated.

The consumption of probiotics was reported to prevent or delay the onset of diabetes and subsequently reducing the incident of hypertension. In a study conducted by Yadav *et al.* (2007), it was found that the administration of Dahi (an Indian fermented milk product) containing *Lactobacillus acidophilus*, *L. casei* and *L. lactis* to high fructose-induced diabetic rats for eight weeks decreased the accumulation of

glycogen in the liver of rats compared to the control that was not fed the probiotics. High fructose diets induce type II diabetes that is associated with insulin resistance. hyperinsulinemia, hypertriglyceridemia and hypertension. This caused by the mobilization and accumulation of fructose in the liver that increases the rate of lipogenesis and synthesis of triacylglycerol. The probiotic dahi-supplemented diet significantly delayed the onset of glucose hyperglycemia, intolerance, hyperinsulinemia, dyslipidemia, and oxidative stress in high fructose-induced diabetic rats, indicating a lower risk of diabetes and its complications. The catabolism of fructose ultimately induces insulin resistance (Yadav et al., 2007). Similarly, Eitahed et al (2011) reported that probiotic yogurt improved total cholesterol and LDL-C concentrations in type 2 diabetic people and may contribute to the improvement of cardiovascular disease risk factors.

The beneficial effects of co-consumption of probiotics with diabetic drugs on controlling diabetes have been reported. Gliclazide is an oral anti-diabetic sulfonylurea drug that has beneficial extra pancreatic effects when therapy is insufficient. insulin Such findings, point toward the beneficial effects of probiotics for treating diabetes in synergism with other diabetes drug and thereby reduces the incidence of diabetes related hypertension. Amar et al. (2011), Cani et al. (2007) reported that dietary modulation of gut microbiota with a view to bifidobacteria increasing reduced endotoxaemia and improved glucose tolerance and insulin secretion, as well as reducing inflammation development in high fat-diet-fed mice. Together, these findings suggest that the gut microbiota contribute to pathophysiological the regulation of endotoxaemia and set the tone of

inflammation, glucose tolerance and insulin secretion. Thus, specific strategies for modifying gut microbiota in favour of bifidobacteria could be useful tools for reducing the impact of high-fat feeding on the occurrence of metabolic diseases.

Probiotics in Cancer

Cancer is a complex disorder, characterized by the uncontrolled growth and spread of abnormal cells. There is not yet a cure for cancer largely because the exact causes of most types of cancer are still not known but however, it is a combination of various metabolic and physiologic disturbances in the cell, which are directly or indirectly related to the involvement of genetic makeup (Giovannucci, 2007; Jain et al., 2010;). It has been reported that diet makes an important contribution to cancer, e.g., up to 75% of colorectal cancer cases are thought to be associated with diet, implying that risks of cancer are potentially reducible. Evidence from a wide range of sources supports the view that the colonic microflora is involved in the etiology of cancer (Hirayama and Rafter, 2006). This has led to intense interest in factors such as probiotics that can modulate gut microflora and its metabolism.

It is known that the risk of developing many types of cancer can be reduced by adopting certain lifestyle changes, such as quitting smoking and eating a nutritional balanced diet. The environment delivers risk factors that cause mutations and initiate cancer or enhance growth by genetic and epigenetic mechanisms (Ferguson, 1999). Nutrition may supply products that may counteract the causative factors (Johnson et al., 1994) and that can be recommended on the basis of a wholesome and complete diet (Pool-Zabel, 2005). Much attention has focused on decreasing cancer risk through diet

alterations, particularly consumption of probiotics and increasing intake of dietary fiber (prebiotics). Several case control studies of cancer has reported inverse association for vogurt, cultured milk and other fermented milk (Le et al., 1986; Young and Wolf, 1988; Veer et al., 1989; Peters et al., 1992; Boutron et al., 1996;). However, Kampman et al. (1994a, b) reported a weak non significant inverse association for fermented dairy products and colorectal cancer and no association for intake of dairy products and decreased risk of colon cancer. Reports abound on reduction of mutagenic activity by L. acidophilus, L. casei and Bifidobacterium bifidum cultures (Biasco et al., 1991; Lidbeck et al., 1992; Aso and Akaza 1992; Hayatsu and Hayatsu, 1993; Aso et al., 1995)

It has been reported that ingestion of probiotics, prebiotics, or combinations of both (synbiotics) plays an important role in the prevention of colorectal cancer (Jain et al., 2010). Goldin and Gorbach have demonstrated that dietary administration of specific lactobacilli some strains significantly decreased the incidence of 1, 2dimethylhydrazine-induced experimental colon cancer (Goldin et al., 1996; Goldin and Gorbach, 2008). The microbial flora and the immune system of the body play an important role in the modulation of carcinogenesis. Both may be influenced by the probiotics. The overall mechanism of probiotic action in the regulation of cancer is not known, however, some of the deduced mechanisms (Hirayama and Rafter, 2006; Jain et al., 2010) include:

- Decreasing the exposure of microbial flora to chemical carcinogens
- Detoxifying ingested carcinogens,/ inhibition of carcinogens and/or procarcinogens

- Decreasing the population or metabolic activity of bacteria that may generate carcinogenic compounds ie inhibition of bacteria that convert pro- carcinogens to carcinogens
- Producing compounds that inhibit the growth of tumor cells
- Stimulating the immune system to defend better against cancer cell proliferation
- Producing metabolic products (e.g. butyrate) which improve programmed cell death (apoptosis).
- reduction of intestinal pH to reduce microbial activity
- alteration of colonic motility and transit time.

The preventions or delay in development of intestinal tumors by lactobacilli is ascribed to its binding to mutagenic compounds in the intestine and suppressing the growth of causative bacteria. which convert procarcinogens into carcinogens. The ability of lactobacilli to reduce the risk of cancers has also been based on their ability to modify gut microflora and to decrease Bglucoronidase and other carcinogen levels (Hirayama and Rafter, 2006; Jain et al., Reddy et al. (1997) developed 2010). azoxymethane-induced aberrant crypt foci in colon of rats and found that a stimulated growth of bifidobacteria in the colon could lead inhibition to the of colon carcinogenesis. The authors suggested pHlowering effect of bifidobacteria in the colon, which subsequently inhibited the growth of E. coli and clostridia.

A decrease in growth of such pathogenic microorganisms may also produce the modulation of bacterial enzymes such as beta-glucuronidase that can convert procarcinogens to proximate carcinogens. There has been evidence that some

probiotics produce butyric acid and that this molecule can influence the rate of apoptosis in enterocytes. Probiotics also neutralize the activity of mutagens such as 4nitroquinoline-N-oxide, 2-nitrofluorene, and benzopyrene (Wollowski et al., 2001). Some probiotics may decrease the fecal concentration of enzymes, mutagens, and secondary bile salts that may be involved in colon carcinogenesis. L. casei Shirota strain is reported to possess promising potential for cancer chemoprevention (Morotomi, 1996). L. rhamnosus GG can protect against the formation of dimethylhydrazine-induced colon cancer in rats (Goldin et al., 1996).

Conclusion

Various in vitro, animal model and case control studies proved the potential for and prebiotics to exert therapeutic effects. Certain combinations of pro- and prebiotics (synbiotics) have revealed greater efficacy than either treatment alone, although there is, however, few randomized controlled clinical trials and epidemiological studies demonstrating the "anti-disease" effects of probiotics in human, ie studies in humans have been less definitive, the few clinical studies do provide evidences that dietary probiotics interact with the host and possibly with the intestinal microbiota and dietary content to exert protective effects in the etiology of some diseases. Great care must be exercised in extrapolating the results of *in* vitro and animal studies to the human system. It also must be kept in mind that the composition and metabolic activities of intestinal flora of experimental animals are significantly different from those of humans. Indeed, it has been demonstrated that human intestinal microflora had different effects than mouse microflora concerning DNA adduct formation after exposure to mutagens (Hirayama et al., 2000). Further research is required to be done to identify the specific

strains and strain characteristics responsible for specific disease treatment, antitumor effects and the mechanisms by which these effects are mediated. However, even with the above reservations in mind and mindful of the limited number of human studies available, the use of probiotics for human disease management and cancer suppression is interesting, holds promise, and certainly deserves more scrutiny. Research works to identify which probiotic, prebiotic, or synbiotic will be most efficacious for a specific treatment should be a research focus for the future, the safety of their use for medical purpose and the safety concerns that may obstruct effective usage and thus judicious application of probiotics in disease management warrants serious investigations.

References

- Aaron, L.A., Burke, M.M. and Buchwald, D. 2000. Overlapping conditions among patients with chronic fatigue syndrome, fibromyalgia and temporomandibular disorder. *Arch. Intern. Med.* 160:221–227.
- Ahola, A.J., Yli-Knuuttila, H., Suomalainen, T., Poussa, T., Ahlström, A., Meurman, J.H. and Korpela, R. 2002. Short-term consumption of probiotic-containing cheese and its effect on dental caries risk factors. *Arch. Oral. Biol.* 47:799-804.
- Amar, J., Chantal, C., Aure'lie, W., Pascale, K., Christelle, V., Luis, G., Bermu'dez, H., Natalia, S., Mathieu, B., Thierry, S., Sampo, L., Arthur, O., Philippe, L., Nina, R.,Philippe, J., Sansonetti, O. and Re'my, B. 2011. Intestinal mucosal adherence and translocation of commensal bacteria at the early onset of type 2 diabetes: molecular mechanisms and probiotic treatment. *EMBO Molecular Medicine* 3: 559–572.
- Andersson, H., Bosaeus, I. and Ellegard, L. 1995. Effect of low-fat milk and fermented low-fat milk on cholesterol absorption and excretion in ileostomy subjects. *European Journal of Clinical Nutrition* 49:274-281.
- Anukam, K.C., Osazuwa, E., Osemene, G.I., Ehigiagbe, F., Bruce, A.W. and Reid, G. 2006a. Clinical study comparing probiotic

Lactobacillus GR-1 and RC-14 with metronidazole vaginal gel to treat symptomatic bacterial vaginosis. *Microbes Infect.* 8:2772-2776.

- Anukam, K., Osazuwa, E. and Ahonkhai, I. 2006b. Augmentation of antimicrobial metronidazole therapy of bacterial vaginosis with oral probiotic *Lactobacillus rhamnosus* GR-1 and *Lactobacillus reuteri* RC-14: randomized, double-blind, placebo controlled trial. *Microbes Infect.* 8:1450-1454.
- Aso, Y. and Akaza, V. 1992. Prophylactic effect of a *Lactobacillus casei* preparation on the recurrence of superficial bladder cancer. BLP Study Group. *Urol. Int.* 49: 125–129.
- Aso, Y., Akaza, H., Kotake, T., Tsukamoto, T., Imai, K. and Naito, S. 1995. Preventive effect of a *Lactobacillus casei* preparation on the recurrence of superficial bladder cancer in a double-blind trial. The BLP Study Group. *Eur. Urol.* 27: 104–109.
- Benton, D., Williams, C. and Brown, A. 2007. Impact of consuming a milk drink containing a probiotic on mood and cognition. *Eur. J. Clin. Nutr.* 61:355–361.
- Biasco, G., Paganelli, G.M., Brandi, G., Brillanti,
 S., Lami, F., Callegari, C. and Gizzi, G. 1991.
 Effect of *Lactobacillus acidophilus* and *Bifidobacterium bifidum* on rectal cell kinetics and fecal pH. *Ital. J. Gastroenterol*. 23: 142-148
- Borody, T.J., Warren, E.F. and Leis, S. 2003. Treatment of ulcerative colitis using fecal bacteriotherapy. J. Clin. Gastroenterol. 37:42-47.
- Borody, T.J., Warren, E.F. and Leis, S.M. 2004.Bacteriotherapy using fecal flora: toying with human motions. *J. Clin. Gastroenterol.* 38:475-483.
- Boutron, M.C., Faivre, J., Marteau, P., Couillault, C., Senesse, P. and Quipourt, V. 1996. Calcium, phosphorous, vitamin D, dairy products and colorectal carcinogenesis: a French case-control study. *Br. J. Cancer.* 74: 145–151.
- Burton, J., Chilcott, C. and Tagg, J. 2005. The rationale and potential for the reduction of oral malodour using *Streptococcus salivarius* probiotics. *Oral Dis.* 11(1): 29–31.
- Burton, J.P., Chilcott, C.N., Moore, C.J., Speiser, G.and Tagg, J.R. 2006. A preliminary study of the effect of probiotic *Streptococcus*

salivarius K12 on oral malodour parameters. *J. Appl. Microbiol*.100:754-764.

- Butt, H.L., Dunstan, R.H. and McGregor, N.R. 1998. Faecal microbial growth inhibition in chronic fatigue/pain patients. In: Proceedings of the AHMF International Clinical and Scientific Conference. Alison Hunter Memorial Foundation, Sydney, Australia.
- Cadieux, P., Burton, J. and Kang, C.Y. 2002. Lactobacillus strains and vaginal ecology. *JAMA* 287:1940-1941.
- Çaglar, E., Sandalli, N., Twetman, S., Kavaloglu, S., Ergeneli, S. and Selvi, S. 2005. Effect of yogurt with Bifidobacterium DN-173 010 on salivary *mutans streptococci* and lactobacilli in young adults. *Acta. Odontol. Scand.* 63:317-320).
- Çaglar, E., Cildir, S.K., Ergeneli, S., Sandalli, N. and Twetman, S. 2006. Salivary *mutans streptococci* and lactobacilli levels after ingestion of the probiotic bacterium *Lactobacillus reuteri* ATCC 55730 by straws or tablets. *Acta. Odontol. Scand.*64:314-318.
- Çaglar, E., Kavaloglu, S.C., Kuscu, O.O., Sandalli, N., Holgerson, P.L. and Twetman, S. 2007. Effect of chewing gums containing xylitol or probiotic bacteria on salivary *mutans streptococci* and lactobacilli. *Clin. Oral. Investig.* 11:425-429.
- Caglar, E., Kuscu, O.O., Selvi, K. S., Kavaloglu, C.S., Sandalli, N. and Twetman, S. 2008a. Short-term effect of ice-cream containing *Bifidobacterium lactis* Bb-12 on the number of salivary mutans streptococci and lactobacilli. *Acta Odontol. Scand.* 66:154-158.
- Çaglar, E., Kuscu, O.O., Cildir, S.K., Kuvvetli, S.S. and Sandalli, N. 2008b. A probiotic lozenge administered medical device and its effect on salivary *mutans streptococci* and lactobacilli. *Int. J. Paediatr. Dent.*18:35-39.
- Calcinaro, F., Dionisi, S., Marinaro, M., Candeloro, P., Bonato, V., Marzotti, S., Corneli, R. B., Ferretti, E., Gulino, A., Grasso, F., De Simone, C., Di Mario, U., Falorni, A., Boirivant, M. and Dotta, F. 2005. Oral probiotic administration induces interleukin-10 production and prevents spontaneous autoimmune diabetes in the nonobese diabetic mouse *Diabetologia* 48: 1565– 1575.
- Cani, P.D., Neyrinck, A. M., Fava, F., Knauf, C., Burcelin, R. G., Tuohy, K. M., Gibson, G. R.

and Delzenne, N. M. 2007. Selective increases of bifidobacteria in gut microflora improve high-fat-diet-induced diabetes in mice through a mechanism associated with endotoxaemia *Diabetologia* 50:2374–2383.

- Cauci, S., Guaschino, S. and Aloysio, D.D. 2003. Interrelationships of inter- leukin-8 with interleukin-1B and neutrophils in vaginal fluid of healthy and bacterial vaginosis positive women. *Molecular Human Reprodroduction* 9:53-58.
- Cildir, S.K., Germec, D., Sandalli, N., Ozdemir, F.I., Arun, T., Twetman, S. and Caglar, E. 2009. Reduction of salivary *mutans streptococci* in orthodontic patients during daily consumption of yoghurt containing probiotic bacteria. *Eur. J. Orthod.* 31:407-411.
- Desbonnet, L., Garrett, L., Clarke, G., Bienenstock, J. and Dinan, T. 2008. The probiotic *Bifidobacteria infantis*: an assessment of potential antidepressant properties in the rat. *J. Psychiatr. Res.* 43(2):164-74.
- De Smet, I., De Boever, P. and Verstraete, W. 1998. Cholesterol lowering in pigs through enhanced bacterial bile salt hydrolase activity. *British Journal of Nutrition* 79:185-194.
- Ejtahed, H.S., Mohtadi-Nia, J., Homayouni-Rad,
 A.'Niafar, M., Asghari-Jafarabadi, M.,
 Mofid, V. and Akbarian-Moghari, A. 2011.
 Effect of probiotic yogurt containing Lactobacillus acidophilus and Bifidobacterium lactis on lipid profile in individuals with type 2 diabetes mellitus. Journal of Dairy Science 94 (7): 3288–3294.
- Elahi, S., Pang, G., Ashman, R. and Clancy, R. 2005. Enhanced clearance of Candida albicans from the oral cavities of mice following oral administration of *Lactobacillus acidophilus. Clin. Exp. Immunol*.141:29-36.
- Esteve, E., Ricart, W., Fernández, R. and Jose, M. 2011. Gut microbiota interactions with obesity, insulin resistance and type 2 diabetes: did gut microbiote co-evolve with insulin resistance?. *Current Opinion in Clinical Nutrition and Metabolic Care* 14(5): 483–490.
- Falagas, M.E., Betsi, G.I. and Athanasiou, S. 2006. Probiotics for prevention of recurrent vulvovaginal candidiasis: a review. *Journal*

of Antimicrobial Chemotherapy 58(2):266-72.

- FAO/WHO, 2002. Guidelines for the evaluation of probiotics in food. http://www.who.int/foodsafety/fs_manageme nt/en/probiotic_guidelines.pdf. Accessed 01, August 2014.
- Fedorak, R.N. and Madsen, K.L. 2004. Probiotics and the Management of Inflammatory Bowel Disease *Inflamm. Bowel Dis.* 10 (3): 286-299.
- Ferguson, L.R. 1999. Natural and man-made mutagens and carcinogens in the diet. Introduction to special issue of mutation research. *Mutat. Res.* 443: 1–10.
- Fujimori, S., Tatsuguchi, A., Gudis, K., Kishida, T., Mitsui, K., Ehara, A., Kobayashi, T., Sekita, Y., Seo, T. and Sakamoto, C. 2007. High dose probiotic and prebiotic co-therapy for remission induction of active Crohn's disease. J. Gastroenterol. Hepatol. 22: 1199– 1204.
- Furrie, E., Macfarlane, S., Kennedy, A., Cummings, J.H., Walsh, S.V., O'Neil D, A. and Macfarlane, G.T. 2005. Synbiotic therapy (*Bifidobacterium longum* /Synergy 1) initiates resolution of inflammation in patients with active ulcerative colitis: A randomised controlled pilot trial. *Gut* 54: 242–249.
- Gardiner, G., Heinemann, C. and Baroja, M.L. 2002. Oral administration of the probiotic combination *Lactobacillus rhamnosus* GR-1 and *L. fermentum* RC-14 for human intestinal applications. *International Dairy* 12:191-196.
- Gionchetti, P., Rizzello, F., Venturi, A., Brigidi, P., Matteuzzi, D., Bazzocchi, G., Poggioli, G., Miglioli, M. and Campieri, M. 2000. Oral bacterio-therapy as maintenance treatment in patients with chronic pouchitis: a doubleblind, placebo-controlled trial. *Gastroenterology* 119:305–309.
- Gionchetti, P., Rizzello, F., Helwig, U., Venturi, A., Lammers, K.M., Brigidi, P., Vitali, B., Poggioli, G., Miglioli, M. and Campieri, M. 2003. Prophylaxis of pouchitis onset with probiotic therapy: a double-blind placebo controlled trial. *Gastroenterology* 124:1202– 1209.
- Gionchetti, P., Lammers, K. M., Rizzello, F. and Campieri, M. 2005. Probiotics and barrier function in colitis, *Gut* 54(7): 898–900.
- Giovannucci, E. 2007. Metabolic syndrome, hyperinsulinemia, and colon cancer: A review, *Am. J. Clin. Nutr.* 86: 836-45.

- Goehler LF, Lyte M, Gaykema RP(2007): Infection-induced viscerosensory signals from the gut enhance anxiety: implications for psychoneuroimmunology. Brain Behav Immun, 21:721–6).
- Goldin, B.R., Gualtieri. L. and Moore, R.P. 1996. The effect of Lactobacillus GG on the initiation and promotion of dimethylhydrazine induced intestinal tumours in the rat. *Nutrition in Cancer* 25: 197-204.
- Goldin, B.R.. and Gorbach, S.L. 2008. Clinical indications for probiotics: An Overview. *Clinical Infectious Diseases* 46(2): S96-S100.
- Grudianov, A.I., Dmitrieva, N.A.and Fomenko, E.V. 2002. Use of probiotics Bifidumbacterin and Acilact in tablets in therapy of periodontal inflammations. *Stomatologiia* (*Mosk*)81:39-43.
- Guslandi, M., Mezzi, G., Sorghi, M. and Testoni, P.A. 2000. *Saccharomyces boulardii* in maintenance treatment of Crohn's disease. *Dig.Dis. Sci.*45:1462–1464.
- Hallén, A., Jarstrand, C. and Påhlson, C. 1992. Treatment of bacterial vaginal disease with lactobacilli. *Sexually Transmitted Diseases* 19: 146-148.
- Hatakka, K., Ahola, A.J., Yli-Knuuttila, H., Richardson, M., Poussa, T., Meurman, J.H.and Korpela, R..2007. Probiotics reduce the prevalence of oral candida in the elderly. A randomized controlled trial. *J. Dent. Res*.86:125-130.
- Hayatsu, H. and Hayatsu, T. 1993. Suppressing effect of *Lactobacillus casei* administration on the urinary mutagenicity arising from ingestion of fried ground beef in the human. *Cancer Lett.* 73: 173–179.
- Hepner, G., Fried, R., St. Jeor, S., Fusetti, L. and Morin. R. 1979. Hypercholesterolemic effect of yoghurt and milk. *Am. J. Clin. Nutr.* 32:19–24.
- Hilton, E., Isenberg, H.D., Alperstein, P. 1992. Ingestion of yogurt containing *Lactobacillus acidophilus* as prophylaxis for candidal vaginitis. *Ann. Intern. Med.* 116:353–357.
- Hillier, S.L., Krohn, M.A., Rabe, L.K., Klebanoff, S.J. and Eschenbach, D.A. 1993. The normal vaginal flora H₂O₂-producing lactobacilli and bacterial vaginosis in pregnant woman. *Clinical Infectious Diseases* 16 (4):S273-S281.
- Hirayama. K., Baranczewski, P., Åkerland, J.E., Midtvedt, T., Möller, L. and Rafter, J. 2000. Effects of human intestinal flora on

mutagenicity of and DNA adduct formation from food and environmental mutagens. *Carcinogenesis* 21: 2105–2111.

- Hirayama, K. and Rafter, J. 2006. Probiotics in Cancer Prevention In Probiotics *in* Food Safety *and* Human Health, Ipek G., Vijay, K.J. and Mohamed, A. (Edts). CRC press London. Pp 365-376.
- Hoesl, C.E. and Altwein, J.E. 2005. The probiotic approach: an alternative treatment option in urology. *European Urology* 47:288-296.
- Ishikawa, H., Akedo, I., Umesaki, Y., Tanaka, R., Imaoka, A. and Otani, T. 2003. Randomized controlled trial of the effect of bifidobacteriafermented milk on ulcerative colitis. *J. Am. Coll. Nutr.* 22:56–63.
- Jain, S., Yadav, M., Menon, S., Yadav, H., and Marotta, F. 2010. Anticarcinogenic effects of probiotics, prebiotics, and synbiotics. In hand book of prebiotics and probiotics ingredients health benefits and food applications, Sungsoo, S.C. and Terry, F.E. (Edts). CRC press, London. Pp273-292.
- Johnson, I.T., Williamson, G. and Musk, S.R.R. 1994. Anticarcinogenic factors in plant foods: A new class of nutrients? *Nutr. Res. Rev.* 7: 175–204.
- Kampman, E., Goldbohm, R.A., van den Brandt, P.A., Veer, P.V. 1994a. Fermented dairy products, calcium, and colorectal cancer in the Netherlands cohort study. *Cancer Res.* 54: 3186–3190.
- Kampman, E., Giovannucci, E., Veer, P.V., Rimm, E., Stampfer, M.J., Colditz, G.A., Kok, F.J. and Willett, W.C. 1994b. Calcium, vitamin D, dairy foods, and the occurrence of colorectal adenomas among men and women in two prospective studies. *Am. J. Epidemiol.* 139: 16–29.
- Kang, M.S., Na, H.S. and Oh, J.S. 2005. Coaggregation ability of *Weissella cibaria* isolates with *Fusobacterium nucleatum* and their adhesiveness to epithelial cells. *FEMS Microbiol. Lett.*253:323-329.
- Kang, M.S., Chung, J., Kim, S.M., Yang, K.H. and Oh, J.S. 2006a. Effect of Weissella cibaria isolates on the formation of *Streptococcus mutans* biofilm. Caries Res. 40:418-425.
- Kang, M.S., Kim, B.G., Chung, J., Lee, H.C. and Oh, J.S. 2006b. Inhibitory effect of *Weissella cibaria* isolates on the production of volatile

sulphur compounds. J. Clin. Periodontol. 33:226-232.

- Kazor, C.E., Mitchel, P.M., Lee, A,M., Stokes, L.N., Loesche, W.J., Dewhirst, F.E. and Paster, B.J. 2003. Diversity of bacterial populations on the tongue dorsa of patients with halitosis and healthy patients. J. Clin. Microbiol. 41:558-563.
- Kibow Biotech 2012. Probiotics and Kidney Health. http://www.kibowbiotech. Com /probiotics-and-kidney-health.html. Accessed 31/07/2014
- Klebanoff, S.J., Hillier, S.L., Eschenbach, D.A. and Waltersdorph, A.M. 1991. Control of the microbial flora of vagina by H₂O₂-generating lactobacilli. *Journal of Infectious Diseases* 164:94-110.
- Komaroff, A.L., Fagioli, L.R. and Geiger, A.M. 1996. An examination of working case definition of chronic fatigue syndrome. *American Journal of Medicine* 100:56-64.
- Kootte, R.S., Vrieze, A., Holleman, F., Dallinga-Thie, G.M., Zoetendal, E.G. and deVos, W.M. 2012. The therapeutic potential of manipulating gut microbiota in obesity and type 2 diabetes mellitus. *Diabetes Obes.Metab.* 14: 112–120.
- Kotowska, M., Albrecht, P. and Szajewska, H. 2005. *Saccharomyces boulardii* in the prevention of antibiotic-associated diarrhoea in children: a randomized double-blind placebo-controlled trial. *Aliment Pharmacol. Ther*.21:583–90.
- Kruis, W., Schutz, E., Fric, P., Fixa, B., Judmaier, G. and Stolte, M. 1997. Doubleblind comparison of an oral *Escherichia coli* preparation and mesalazine in maintaining remission of ulcerative colitis. *Aliment. Pharmacol. Ther*.11:853–858.
- Kruis, W., Kalk, E.K., Fric, P. and Stolte, M. 2001. Maintenance of remission in ulcerative colitis is equally effective with *Escherichia coli* Nissle 1917 and with standard mesalamine. *Gastroenterology* 120:A127-34.
- Le, M.G., Moulton, L.H., Hill, C. and Kramar, A. 1986. Consumption of dairy produce and alcohol in a case-control study of breast cancer. J. Natl. Cancer Inst. 77: 633–636.
- Lemberg, D.A., Ooi, C.Y. and Day, A.S. 2007. Probiotics in paediatric gastrointestinal diseases. J. Paediatr. Child. Health 43: 331– 336.

- Lidbeck, A., Övervik, E., Rafter, J., Nord, C.E. and Gustafsson, J.Å. 1992. Effect of *Lactobacillus acidophilus* supplements on mutagen excretion in feces and urine in humans. *Microb. Ecol. Health Dis.* 5: 59–67.
- Lieske, J.C., Goldfarb, D.S., De Simone, C. and Regnier, C. 2005. Use of a probiotic to decrease enteric hyperoxaluria. *Kidney International* 68: 1244-1249.
- Liong, M.T. and Shah, N.P. 2005a. Acid and bile tolerance and cholesterol removal ability of lactobacilli strains. *Journal of Dairy Sciences* 88:55-66.
- Liong, M.T. and Shah, N.P. 2005b. Bile salt deconjugation ability, bile salt hydrolase activity and cholesterol co-precipitation ability of lactobacilli strains. *International Dairy Journal* 15 (4): 391-398.
- Liu, T., Wang, B.Q., Wang, C.S. and Yang, P.C. 2006. Concurrent exposure to thermal stress and oral Ag induces intestinal sensitization in the mouse by a mechanism of regulation of IL-12 expression. *Immunol. Cell Biol.* 84: 430–439.
- Logan, A., Rao, V. and Irani, D. 2003. Chronic fatigue syndrome: lactic acid bacteria may be of therapeutic value. *Med. Hypotheses* 60:915–23.
- Lyte, M., Varcoe, J.J. and Bailey, M.T. 1998. Anxiogenic effect of subclinical bacterial infection in mice in the absence of overt immune activation. *Physiol. Behav.* 65:63–8.
- Malchow, H.A. 1997. Crohn's disease and *Escherichia coli*. A new approach in therapy to maintain remission of colonic Crohn's disease? *J. Clin. Gastroenterol*.25:653–658.
- Marteau, P., Sokol, H., Dray, X. and Seksik, P. 2009. Bacteriotherapy for inflammatory bowel disease: therapeutic tool and/or pharmacological vectors?. *Gastroenterol. Clin. Biol.* 33 (3): 228-234.
- McFarland, L.V. 2009. Evidence-based review of probiotics for antibiotic-associated diarrhea and *Clostridium difficile* infections. *Anaerobe* 15:274-280.
- Meurman, J.H., Antila, H., Korhonen, A. and Salminen, S. 1995. Effect of *Lactobacillus rhamnosus* strain GG (ATCC 53103) on the growth of *Streptococcus* sobrinus in vitro. *Eur. J. Oral. Sci.*103:253-258.
- Morotomi, M. 1996. Properties of *Lactobacillus casei* Shirota strain as probiotics. *Asia Pacific Journal of Clinical Nutrition* 5:29-30.

- Natarajan, R., Eli, A., Friedman, P.T., Venkat, Rao., Parimalam, R. and Rahul, D.2009. Probiotics dietary supplementation in patients with stage 3 and 4 chronic kidney disease: A 6-month pilot scale trial in Canada. *Current Medical Research and Opinion* 25 (8): 1919– 1930.
- Natarajan, R., Pari, R., Eli, A., Friedman, P.T., Anthony, Joseph., Barbara, D., David, S., Gold, A., Paul, T.A., Venketeshwer, R., Emmanuel, A. and Carlos, G. M. 2010. Pilot study of probiotic dietary supplementation for promoting healthy kidney function in patients with chronic kidney disease. *Adv. Ther*. 27(9):634-647.
- Nase, L., Hatakka, K., Savilahti, E., Saxelin, M., Pönkä, A., Poussa, T., Korpela, R. and Meurman, J.H. 2001. Effect of long-term consumption of a probiotic bacterium, *Lactobacillus rhamnosus* GG, in milk on dental caries and caries risk in children. *Caries Res.* 35(6): 412–420.
- Mimura, T., Rizzello, F., Schreiber, S., Talbot, I.C., Nicholls, R.J.,Gionchetti, P., Campieri, M. and Kamm, M.A. 2002. Once daily high dose probiotic therapy maintains remission and improved quality of life in patients with recurrent or refractory pouchitis: a randomised, placebo-controlled, double-blind trial. *Gastroenterology* 122:A81-87.
- Nguyen, T.D., Kang, J.H. and Lee, M.S. 2007. Characterization of *Lactobacillus plantarum* PH04, a potential probiotic bacterium with cholesterol-lowering effects. *International Jornal of Food Microbiology* 113:358-361.
- Noh, D. O., Kim, S. H. and Gilliland. S. E.1997. Incorporation of cholesterol into the cellular membrane of *Lactobacillus acidophilus* ATCC 43121. J. Dairy Sci. 80:3107–3113.
- Panwar, H., Rashmi, H.M., Batish, V.K. and Grover, S. 2013. Probiotics as potential biotherapeutics in the management of type 2 diabetes – prospects and perspectives. *Diabetes/Metabolism Research and Reviews* 29(2): 103–112.
- Parent, D., Bossens, M. and Bayot, D. 1996. Therapy of bacterial vaginosis using exogenously-applied *Lactobacillus acidophili* and a low dose of estriol: a placebocontrolled multicentric clinical trial. *Arzneimittelforschung* 46:68-73.
- Peters, R.K., Pike, M.C., Garabrant, D. and Mack, T.M. 1992. Diet and colon cancer in Los

Angeles County, California. *Cancer Causes Control* 3: 457–473.

- Plein, K. and Hotz, J. 1993. Therapeutic effects of *Saccharomyces boulardii* on mild residual symptoms in a stable phase of Crohn's disease with special respect to chronic diarrhea—a pilot study. *Gastroenterol*.31:129–134.
- Pool-Zobel, B.L. 2005. Inulin-type fructans and reduction in colon cancer risk: Review of experimental and human data. *Br. J. Nutr.* 93: S73–S90.
- Prantera, C., Scribano, M.L., Falasco, G., Andreoli, A. and Luzi, C. 2002. Ineffectiveness of probiotics in preventing recurrence after curative resection for Crohn's disease: a randomised controlled trial with *Lactobacillus GG. Gut* 51:405–409.
- Rao, A.V., Bested, A.C., Beaulne, T.M., Katzman, M.A., Iorio, C., Berardi, J.M. and Logan, A.C. 2009. A randomized, double-blind, placebo-controlled pilot study of a probiotic in emotional symptoms of chronic fatigue syndrome. *Gut Pathogens* 1: 1-6.
- Reddy, B.S., Hamid, R. and Rao, C.V. 1997. Effect of dietary oligofructose and inulin on colonic preneoplastic aberrant crypt foci inhibition. *Carcinogenesis* 18: 1371–74.
- Reid, G., McGroarty, J. and Tomeczek, L. 1996. Identification and plasmid profiles of Lactobacillus species from the vagina of 100 healthy women. *FEMS Immunology and Medical Microbiology* 15:23-26.
- Rembacken, B.J., Snelling, A.M., Hawkey, P.M., Chalmers, D.M. and Axon, A.T. 1999. Nonpathogenic *Escherichia coli* versus mesalazine for the treatment of ulcerative colitis: a randomised trial. Lancet 354:635– 639.
- Rioux, K.P. and Fedorak, R.N. 2006. Probiotics in the treatment of inflammatory bowel disease. J. Clin. Gastroenterol. 40(3):260-3.
- Santosa, S., Farnworth, E. and Jones, P. 2006. Probiotics and their potential health claims. *Nutrition Review* 64:265–274.
- Sartor, R.B. 2004. Therapeutic manipulation of the enteric microflora in Inflammatory Bowel Diseases: Antibiotics, Probiotics, and Prebiotics. *Gastroenterology* 126:1620–1633.
- Scaldaferri, F., Viviana, G., Loris, R.L., Fabio, D.Z., Francesca, M., Ivo, B., Giovanni, B., Valentina, P., Lucrezia, L., Giovanni, C., Eleonora, G., Alessandro, S. and Antonio, G.

2013. Gut microbial flora, prebiotics and probiotics in IBD: Their current usage and utility. *BioMed Research International* 2013:1-10.

- Schultz, M., Veltkamp, C. and Dieleman, L.A. 2002. *Lactobacillus plantarum* 299V in the treatment and prevention of spontaneous colitis in interleukin-10-deficient mice. *Inflamm. Bowel Dis.* 8:71–80.
- Seth, A., Yan, F. and Rao, R.K. 2008. Probiotics ameliorate the hydrogen peroxide-induced epithelial barrier disruption by a PKC- and MAP kinase-dependent mechanism. *American Journal of Physiology*. *Gastrointestinal and Liver Physiology* 294(4):G1060-G1069.
- Singh, V., Kiran, S., Sarika, A., Desh, D. S., Parul, T., Ganda, L. S., and Hariom, Y. 2009. Innate and specific gut-associated immunity and microbial interference. *FEMS Immunology* and Medical Microbiology 55 (1): 6–12.
- Stachowicz, N. and Kiersztan, A. 2013. The role of gut microbiota in the pathogenesis of obesity and diabetes. *Postepy. Hig. Med. Dosw.* 67:288-303.
- Stamatova, I., Kari, K.and Meurman, J.H. 2007. *In vitro* evaluation of antimicrobial activity of putative probiotic lactobacilli against oral pathogens. *Int. J. Probiotics and Prebiotics* 2:225-232.
- Stamatova, I. and Meurman, J. 2009. Probiotics: Health benefits in the mouth. *American Journal of Dentistry* 22(6): 329-338.
- Sudha, M. R., Chauhan. P.1., Dixit, K.1., Babu, S. and Jamil, K. 2009. Probiotics as complementary therapy for hypercholesterolemia. *Biology and Medicine* 1(4): 1-13
- Taranto, M. P. and Font de Valdez, G. 1999. Localization and primary characterization of bile salt hydrolase from *Lactobacillus reuteri*. Biotechnol. Lett. 21:935–938.
- Taranto, M.P., Medici, M., Perdigon, G. 2000. Effect of *Lactobacillus reuteri* on the prevention of hypercholesterolemia in mice. *J. Dairy Sci.* 83:401–403.
- Tursi, A., Brandimarte, G., Giorgetti, G.M., Forti, G., Modeo, M.E. and Gigliobianco, A. 2004. Low-dose balsalazide plus a high-potency probiotic preparation is more effective than balsalazide alone or mesalazine in the treatment of acute mild-to-moderate

ulcerative colitis. *Med. Sci. Monit.* 10 (11):I126-131.

- Ulisse, S., Gionchetti, P., and D'Alo, S. 2001. Expression of cytokines, inducible nitric oxide synthase, and matrix metalloproteinases in pouchitis: effects of probiotic treatment. *Am. J. Gastroenterol.* 96:2691–2699.
- Vaarala, O. 2003. Immunological effects of probiotics with special reference to lactobacilli. *Clin. Exp. Allergy* 33:1634–1640.
- Veer, P.V., Dekker, J.M., Lamers, J.W., Kok, F.J., Schouten, E.G., Brants, H.A., Sturmans, F. and Hermus, R.J. 1989. Consumption of fermented milk products and breast cancer: a case-control study in the Netherlands. *Cancer Res.* 49: 4020–4023.
- Volozhin, A.I., Il'in, V.K., Maksimovski, IuM., Sidorenko, A.B., Istranov, L.P., Tsarev, V.N., Istranova, E.V. and Aboiants, R.K. 2004. Development and use of periodontal dressing of collagen and *Lactobacillus casei* cell suspension in combined treatment of periodontal disease of inflammatory origin (a microbiological study). *Stomatologiia (Mosk)* 83:6-8.
- Wagner, R.D., Pierson, C., Warner, T., Dohnalek, M., Hilty, M. and Balish, E. 2000. Probiotic effects of feeding heat-killed *Lactobacillus* acidophilus and *Lactobacillus casei* to *Candida albicans*-colonized immunodeficient mice. J. Food Prot.63:638-644.
- Walker, W.A. 2008. Mechanisms of action of probiotics. *Clinical Infectious Diseases* 46 (2): S87-S91.
- Welters, C. F. M., Heineman, E., Thunnissen, F. B. J. M., Van den Bogaard, A. E. J. M., Soeters, P. B. and Baeten, C. G. M. I. 2002. Effect of dietary inulin supplementation on inflammation of pouch mucosa in patients with an ileal pouch-anal anastomosis. *Diseases of the Colon and Rectum* 45(5) 621– 627.
- Whitehead, W.E., Palsson, O. and Jones, K.R. 2002. Systematic review of the co-morbidity of irritable bowel syndrome with other disorders: what are the causes and implications?. *Gastroenterology* 122:1140–56.
- WHO, 2013. Ending preventable deaths from pneumonia and diarrhoea by 2025, Integrated global action plan for the prevention and control of pneumonia and diarrhoea (GAPPD) 12 April 2013.

- Wollowski, I., Rechkemmer, G. and Pool-Zobel, B.L. 2001. Protective role of probiotics and prebiotics in colon cancer. *American Journal of Clinical Nutrition* 73(2):451S-455S.
- Yadav, H., Jain, S. and Sinha, P.R. 2007. Antidiabetic effect of probiotic dahi containing *Lactobacillus acidophilus* and *Lactobacillus casei* in high fructose fed rats. *Nutrition* 23 (1): 62–68.
- Young, T.B. and Wolf, D.A. 1988. Case-control study of proximal and distal colon cancer and diet in Wisconsin. *Int. J. Cancer.* 42: 167– 175.