

# **Bridging Worlds:**

## From macro- and micro-scale to prototype and product

#### Dr. Simon Coleman

Insight Centre for Data Analytics, Dublin City University, Dublin 9, Ireland.

## Lab to Life

lab-based development

#### Bench top breakthrough to real-life application

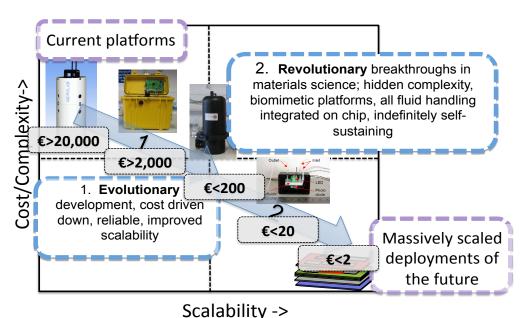


Bulb image: http://gtnetwork.ie/index.php/gifted-adults/lightbulb/ <accessed 12/11/14>
Production line: http://en.wikipedia.org/wiki/Production\_line, accessed 12/11/14>
Money: http://www.dreamstime.com/stock-photo-business-man-pile-money-vector-illustration-cartoon-image35110170
<accessed 12/11/14>

# **Our Challenge**



- Development of low cost, autonomous, deployable environmental sensor platforms.
- Innovative sampling and target preconcentration strategies for more comprehensive analysis.
- Merging novel materials and microfluidic platforms



- Highly specific target detection methods for determination of bacterial and chemical contaminants.
- Commercialisation of platform in parallel to research activities.

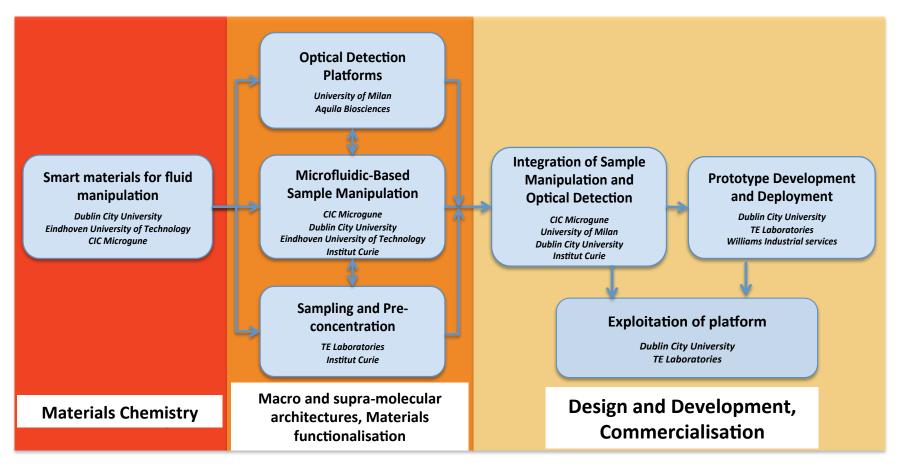




# NAPES Research and productisation NAPES

#### **MAM-14 Topics**





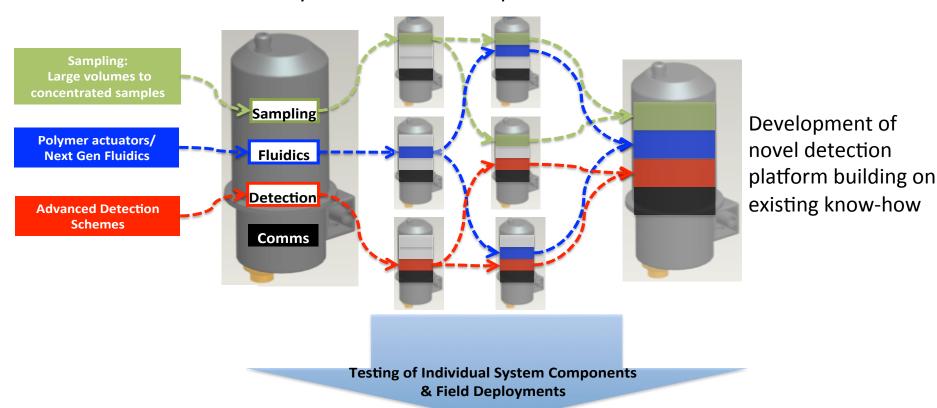




# **NAPES** Research strategy



"Plug and Play" use of existing systems to test novel platforms



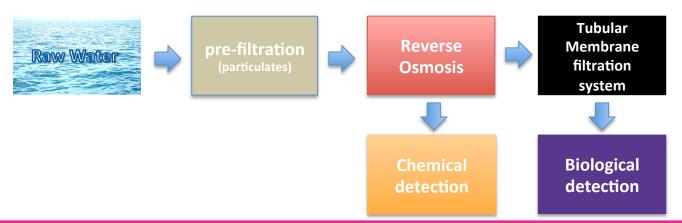




# **Sampling and Pre-Concentration**



- Portable deployable platforms commonly take small, mL scale volumes for analysis
- Milli- and micro-litre samples may not truly represent large water bodies
- NAPES sampling systems will facilitate intake of much larger volumes with sample reduction and significant pre-concentration using:
  - Chemical concentration will employ Reverse Osmosis (RO).
  - > Bacterial concentration will use a tubular membrane based filtration unit (TF).











# **Sampling and Pre-Concentration**

Reverse Osmosis (RO) chemical preconcentration



	Contaminants (ppm)					
	Nitrate	Nitrite	Phosphate	Ammonia	Iron	Manganese
Initial Sample	23.38	0.75	1.79	0.11	0.076	0.024
Run 1	191.86	7.51	3.11	0.21	0.103	0.08
Run 2	74.59	2.42	2.88	0.12	0.039	0.03
Run 3	90.13	2.94	3.07	0.12	0.094	0.028
Run 4	102.08	3.33	2.9	0.12	0.085	0.041
RO Processed water	5.13	0.24	<1	0	0.034	0.013
Conc. factor	4.4	4.4	1.6	1.09	1.1	1.7





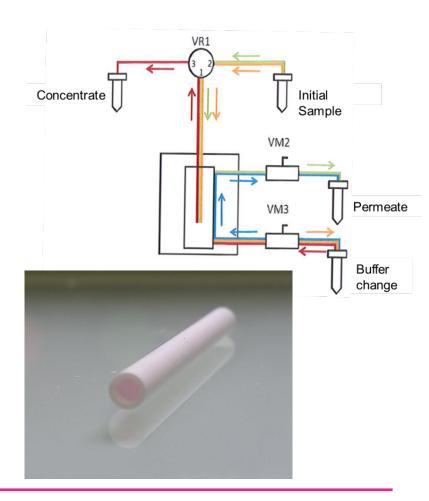


# **Sampling and Pre-Concentration**

**Tubular Membrane Filtration (TF)** 



- Institut Curie developing a TF system will employ a ceramic membrane system.
- Bacterial concentration up to 30X
- Volume throughput of system 1mL/minute
- 10 mL initial volume reduced to approx. 100-500 μL
- Up to **100 fold reduction** in volume in 10 minutes (related sample volume, pressure and pore size).
- Potential for chemical concentration.



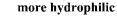


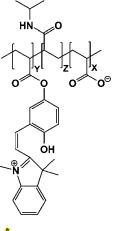




Photoresponsive polymer hydrogels

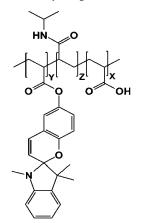




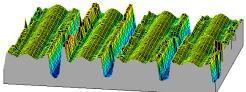




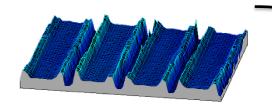
less hydrophilic



Molecular switching













Macro-scale change



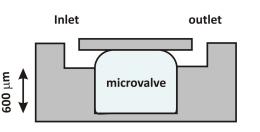


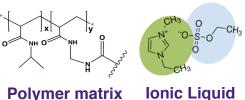


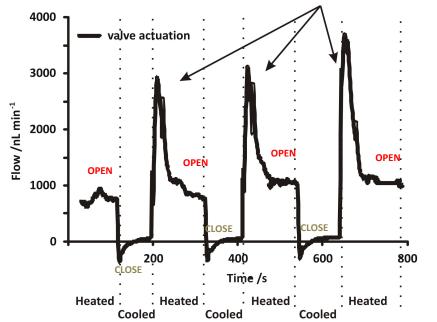
#### Microfluidic platforms incorporating smart materials

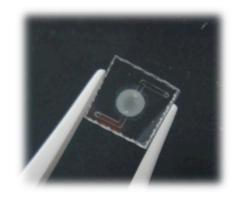


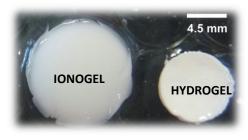
Burst of the flow sensor











- Actuation in 4s ± 2s
- Recovery in 32s ± 2s
- > 8 cycles

- 33.5 % volume change compared to hydrogels
- Low evaporation process
- Less brittle, Soft (plasticizer)





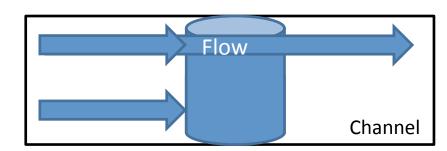


#### Light actuated polymer valves

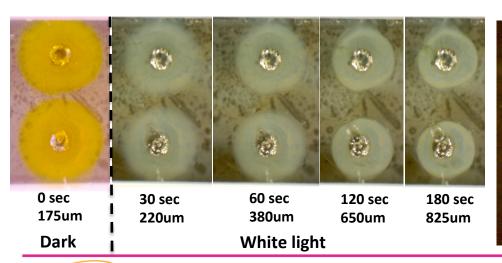


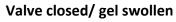
Valve open/gel shrunk

- DCU developed polymer valve systems using light actuated materials.
- Reproducible actuation effects over several cycles
- Approx. 10- 40% shrinking (depending on gel size and light).



In-situ polymerisation of valves





No Flow



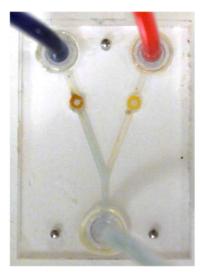




#### Polymer valve integrated within microfluidic chips

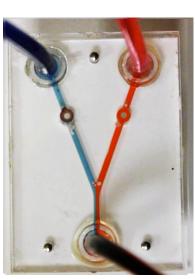


- Integration of valves within microfluidic chips allows for non-contact control of valving system
- Surface mount, low power LED light source allows for precise illumination of valves with no mechanical components on chip
- Modular nature of fluidic platform allows for ease of replacement of fluidic components when replacement required











Valves closed

Valves open

Multiple channel flow

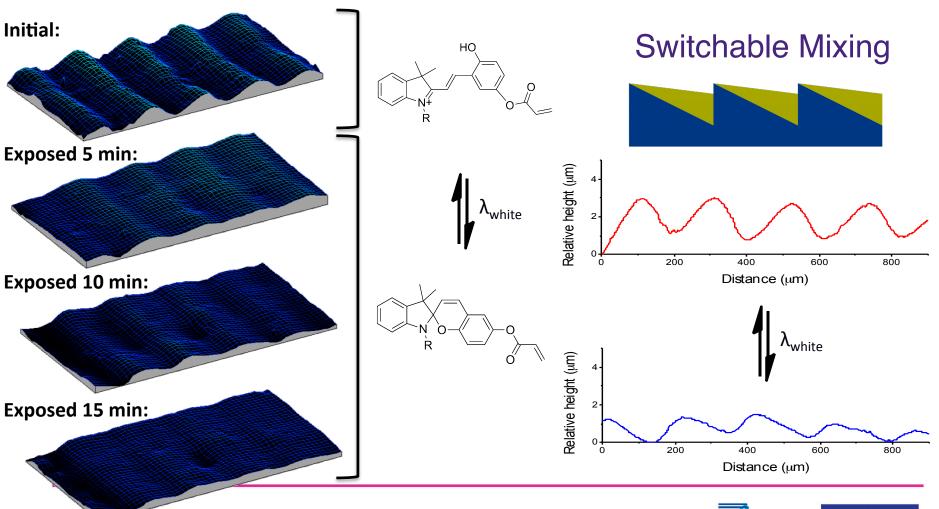






Photoresponsive ratchets





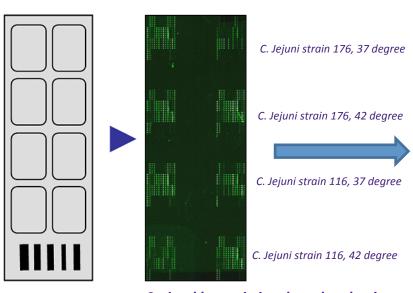
**Challenge: Increasing actuation speeds** 





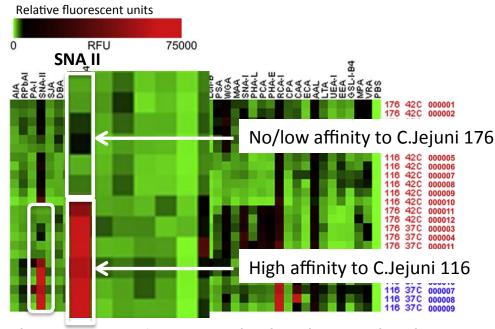
#### Lectin microarray analysis of two strains of C. Jejuni





Glass slide: 8 lectin arrays Per slide. Stained bacteria incubated on lectin array resulting in a specific glycosignature

42 lectins in 6x replications; 256 spots per array



FACE Patenteinteinties. Equatateathe 126 bindbeatleonin SNACH with Rifigh affinity (red) however strain 176 is not binding to this lectin at all (green/black).

Each pixel is the RFU from one lectin. Each column is a lectin that can detect a specific Glycan.

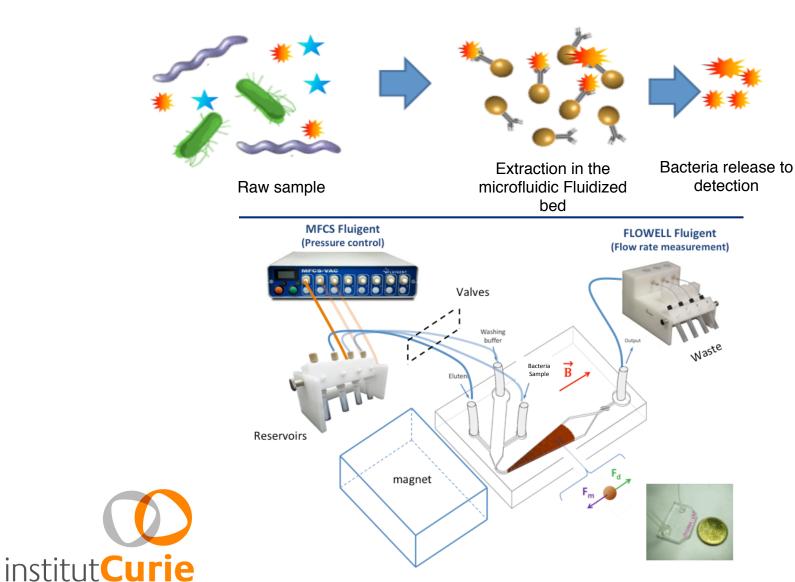






On-chip bacterial detection using magnetic bead system

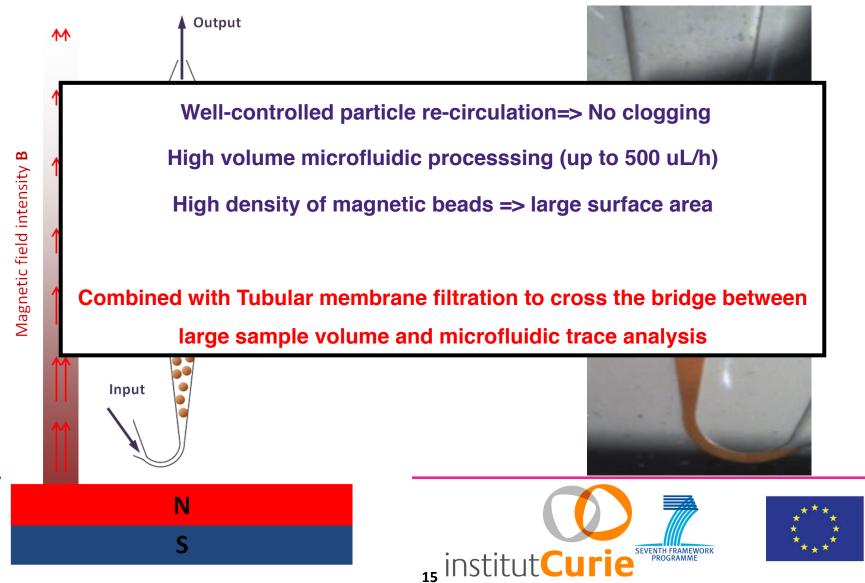




Microfluidic Magnetic fluidized bed

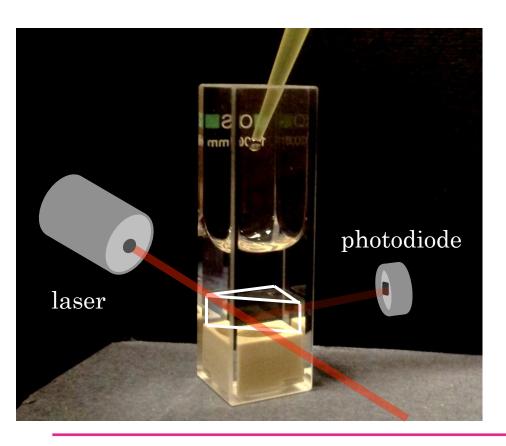


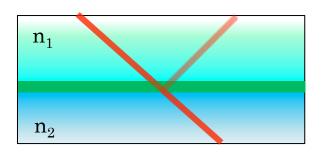
Flow rate : 2 μL/min



Reflective Phantom Interface (RPI) Method







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- F. Giavazzi et al., Proc Natl Acad Sci USA, 110 (2013) 9350-9355
- F. Giavazzi et al., Biosensors and Bioelectronics, in press (2014), DOI: 10.1016/j.bios.2014.02.077



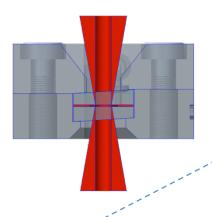


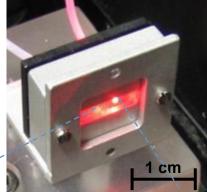
#### **Bacterial detection by RPI surface**

Design and construction of fluidic module

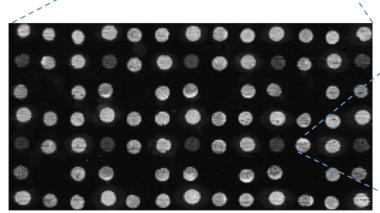


 Aquila Bioscience and University of Milan producing RPI surfaces with strain specific bonding based upon lectin coatings.

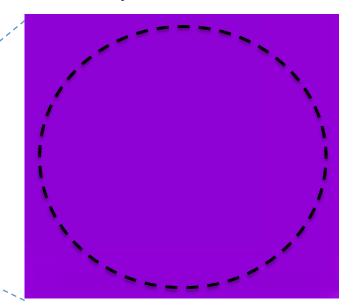








Evaluation of the optical platform







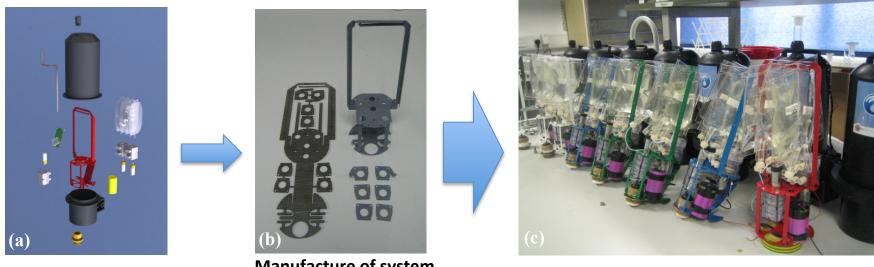




# **Prototype Development**



- Dublin City University(DCU) will lead the production of prototype platforms
- Design and engineering of platform for scalable production
- Commercialisation activities in parallel with TEL to carry out market research and influence final system construction.



Design (CAD, Solidworks)

Manufacture of system (3d printing, laser cut frames, components)

Small scale Prototype production line (test scalability)







# **Prototype Field deployment**



- Industry-Academic Collaborative activity
- TE Laboratories and Williams Industrial Services and DCU will lead Prototype testing and deployment.
- Phase 1: Real water samples will be collected for lab based testing of prototypes
- Phase 2: Field trials at Waste water treatment plants and water supply reservoirs
- Potential for deployments outside of Ireland with partners (e.g Italy, Spain)



Example of DCU coordinated deployment of autonomous phosphate system in Irish river.











# **DCU Led Commercialisation: A Case Study**



#### **Funding history**

2011-2013: Concept & validation

2013-2014: Commercialisation

2014: Incorporation and seed capital









A globally deployed technology: Ireland, UK, Australia, Brazil... more to come...

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# **Concluding remarks**



- NAPES aims to develop novel materials science and microfluidic platforms while developing a commercial strategy in parallel
- Use of **readily sourced electronics and raw materials** to ensure **Scalability** of modules and economic design of final device.
- Combination of **new materials technologies**, **biochemistry and microfluidics** to produce **low cost sensing platforms** allowing greater accessability to technology.





#### **NAPES Consortium**























# **Acknowledgements**

#### **Project partners:**

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