



University of Dundee

High-resolution quantitative proteome analysis reveals substantial differences between phagosomes of RAW 264.7 and bone marrow-derived macrophages

Guo, Manman; Härtlova, Anetta; Dill, Brian D.; Prescott, Alan R.; Gierliski, Marek; Trost, Matthias

Published in:
Proteomics

DOI:
[10.1002/pmic.201400431](https://doi.org/10.1002/pmic.201400431)

Publication date:
2015

Document Version
Publisher's PDF, also known as Version of record

[Link to publication in Discovery Research Portal](#)

Citation for published version (APA):

Guo, M., Härtlova, A., Dill, B. D., Prescott, A. R., Gierliski, M., & Trost, M. (2015). High-resolution quantitative proteome analysis reveals substantial differences between phagosomes of RAW 264.7 and bone marrow-derived macrophages. *Proteomics*, 15(18), 3169-3174. DOI: 10.1002/pmic.201400431

General rights

Copyright and moral rights for the publications made accessible in Discovery Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from Discovery Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain.
- You may freely distribute the URL identifying the publication in the public portal.

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

DATASET BRIEF

High-resolution quantitative proteome analysis reveals substantial differences between phagosomes of RAW 264.7 and bone marrow derived macrophages

Manman Guo^{1*}, Anetta Härtlova^{1*}, Brian D. Dill¹, Alan R. Prescott², Marek Gierliński³ and Matthias Trost^{1*}

¹ MRC Protein Phosphorylation and Ubiquitylation Unit, College of Life Science, University of Dundee, Scotland, UK

² Division of Cell Signalling and Immunology, College of Life Science, University of Dundee, Scotland, UK

³ Division of Computational Biology, College of Life Sciences, University of Dundee, Scotland, UK

Macrophages are important immune cells operating at the forefront of innate immunity by taking up foreign particles and microbes through phagocytosis. The RAW 264.7 cell line is commonly used for experiments in the macrophage and phagocytosis field. However, little is known how its functions compare to primary macrophages. Here, we have performed an in-depth proteomics characterization of phagosomes from RAW 264.7 and bone marrow derived macrophages by quantifying more than 2500 phagosomal proteins. Our data indicate that there are significant differences for a large number of proteins including important receptors such as mannose receptor 1 and Siglec-1. Moreover, bone marrow derived macrophages phagosomes mature considerably faster by fusion with endosomes and the lysosome which we validated using fluorogenic phagocytic assays. We provide a valuable resource for researcher in the field and recommend careful use of the RAW 264.7 cell line when studying phagosome functions. All MS data have been deposited in the ProteomeXchange with identifier PXD001293 (<http://proteomecentral.proteomexchange.org/dataset/PXD001293>).

Received: September 8, 2014

Revised: September 8, 2014

Accepted: December 8, 2014

Keywords:

Cell biology / Macrophage / Phagosome / Proteomics RAW 264.7



Additional supporting information may be found in the online version of this article at the publisher's web-site

Macrophages are immune cells that exist in many different tissues and perform a wide range of biological functions [1, 2]. They are extremely plastic in their protein expression pattern and can become activated by a range of cytokines and pathogen-associated molecules such as lipopolysaccharide. One of the main functions of macrophages is phagocytosis, which is the active uptake of large particles (> 0.5 μm) by cells [3]. Phagocytosis is an important cellular mechanism for almost all eukaryotes, highly conserved from amoeba to human [4], in mammals, it is a key part of the innate immune response against invading microorganisms.

Moreover, macrophages phagocytose apoptotic cells and cell debris to recycle cellular building blocks during homeostasis and development [5, 6]. Phagocytosis is induced through the binding of particles to cell surface receptors such as the mannose receptor, Fc-receptors, or scavenger receptors. After internalization, newly formed phagosomes engage in a maturation process that involves fusion with endosomes, lysosomes, and other organelles [7, 8] leading to the formation of phagolysosomes in which the foreign matter is degraded. Peptide antigens from the particle are also presented via MHC class I and II molecules, bridging innate and adaptive immunity.

The macrophage-like cell line RAW 264.7 [9] was derived from Balb/c mice injected with Abelson murine leukemia virus and has been extensively used in the phagocytosis and macrophage field in the past with almost 4000 articles listed

Correspondence: Dr. Matthias Trost, MRC Protein Phosphorylation and Ubiquitylation Unit, College of Life Sciences, University of Dundee, Dow Street, Dundee, DD1 5EH, UK

E-mail: m.trost@dundee.ac.uk

Fax: +44-1382-223778.

Abbreviations: BMDMs, bone marrow-derived macrophages; MRC1, mannose receptor 1

*These authors contributed equally to this work.

Colour Online: See the article online to view Fig. 1 in colour.

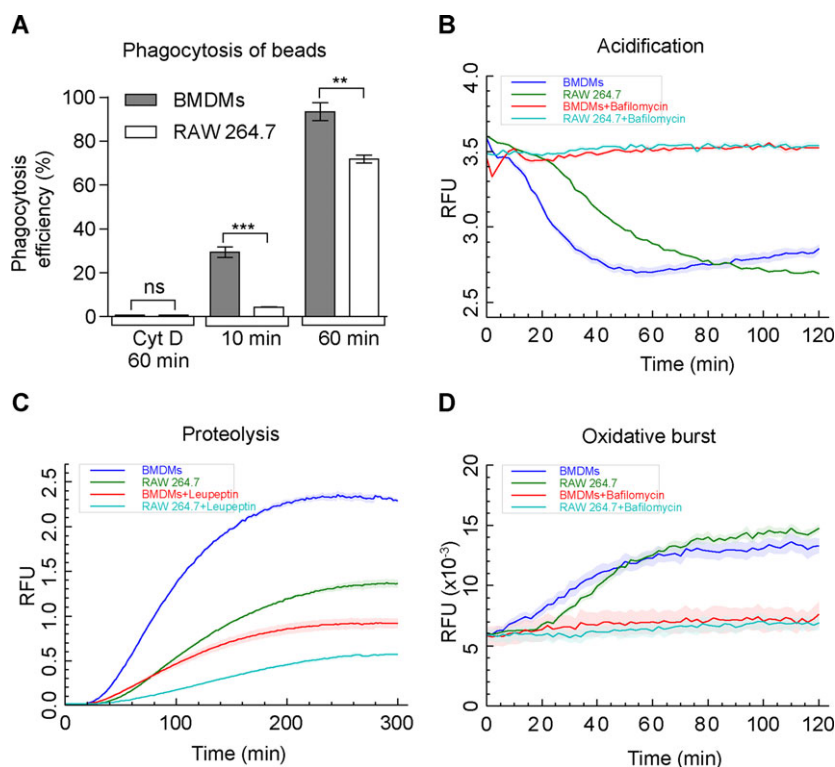


Figure 1. BMDMs display an increased rate of phagocytosis, phagosomal acidification, and proteolysis compared to RAW 264.7 macrophages. (A) BMDMs show a strongly increased efficiency of phagocytosis of AF-488-coated 3 μm beads compared to RAW 264.7 cells. Cells treated with the actin polymerization inhibitor cytochalasin D were used as non-phagocytic negative controls. Error bars represent standard deviation, pair-wise *t*-test comparison with ****p* < 0.0001 and ***p* < 0.001. (B–D) Real-time kinetics of fluorescent phagosome assays. BMDM phagosomes acidify significantly faster (B), display a much faster and stronger phagosomal proteolysis (C) but a similar oxidative burst (D) compared to RAW 264.7 cells. Shaded areas around lines are the standard error of mean (SEM) of six replicates.

in Pubmed in 2014. In recent years, the cell line has also been used extensively in proteomics experiments including a large-scale proteome [10], phagosome proteomics [4, 8, 11], responses to cytokines [12, 13], and identification of DNA receptors [14].

In this paper, we have compared the phagosomal functions and proteomes of RAW 264.7 and bone marrow derived macrophages (BMDMs) [15, 16] in order to determine if the cell line was a good substitute for primary macrophages when studying phagosome and macrophage biology.

First, we performed a set of experiments to characterize differences in phagocytic uptake of beads and phagosome maturation in RAW 264.7 and BMDMs. Phagocytic uptake assays using Alexafluor488-labeled 3 μm silica beads showed a significantly reduced phagocytosis rate in RAW 264.7 compared to BMDM, particularly at early time points (Fig. 1A). Moreover, we performed fluorogenic assays to measure pH, proteolysis, and oxidative capacity of phagosomes in real time [17–19] and identified a significant faster phagosome acidification in BMDMs (Fig. 1B), which is possibly due to the faster phagocytosis compared to RAW 264.7 cells. However, BMDMs also display a much stronger proteolytic activity, suggesting a quicker fusion with lysosomes, a reduced inactivation of lysosomal proteases, and/or a higher concentration of proteases in the lysosome (Fig. 1C). Interestingly, RAW 264.7 cells produce a similar or even slightly higher oxidative burst in phagosomes compared to BMDMs (Fig. 1D), suggesting that this function is well conserved in the cell line.

In order to get insights into the molecular differences of the RAW 264.7 and BMDM phagosome proteomes, we performed a label-free proteome analysis of phagosomes from both cell types. Phagocytosis was induced for 30 min using carboxylated 1 μm polystyrene beads (Estapor/EMD Millipore) and phagosomes were isolated using sucrose ultracentrifugation in which the phagosomes float above cell debris and other organelles [11, 20]. Protein extracts from three biological replicates were resuspended in 1% sodium 3-[(2-methyl-2-undecyl-1,3-dioxolan-4-yl)methoxy]-1-propanesulfonate (commercially available as RapiGest, Waters) in 50 mM Tris-HCl pH 8.0 with 5 mM tris(2-carboxyethyl)phosphine (TCEP, Pierce), heated at 60°C for 5 min, then alkylated using 10 mM iodoacetamide, diluted to 0.1% RapiGest and finally trypsinized overnight at 30°C. After SPE clean-up, 2 μg of each sample was injected onto an Orbitrap Fusion mass spectrometer (Thermo-Fisher Scientific) via a 50 cm long 75 μm column using a 6 h gradient delivered by a Dionex U-3000 LC system (Thermo-Fisher Scientific). Data were acquired with a resolution of 120 000 in MS1 and a scan range of 400–1600. Peptide ions were fragmented by HCD (35% collision energy) with a resolution of 15 000, an AGC target of 50 000, and a maximum injection time of 60 ms. The whole duty cycle was set to 2.5 s during which the instrument performed “top speed” analysis. Data were analyzed by label-free quantitation using MaxQuant v1.5.0.12 [21] and searched against a murine Uniprot-Trembl database (51 372 entries; downloaded February 19 2014) and a list of common contaminants. This resulted in an identification of 3535 proteins

A GO-term enrichment

enriched in BMDM phagosomes				enriched in RAW 264.7 phagosomes			
GO-#	Term	# of proteins	p-value	GO-#	Term	# of proteins	p-value
GO:06952	defense response	25	9.3e-6	GO:06412	translation	22	0.0005
GO:07155	cell adhesion	23	0.008	GO:15672	monovalent inorganic cation transport	13	0.004
GO:31012	extracellular matrix	14	0.003	GO:15293	symporter activity	7	0.025
GO:48002	antigen processing & presentation	9	0.0001	GO:33176	proton-transporting vATPase compl.	6	0.0006
GO:16209	antioxidant activity	6	0.015	GO:42641	actomyosin	4	0.044
GO:08305	integrin complex	5	0.021	GO:05544	Ca ²⁺ -dependent phospholipid binding	3	0.0001

B Phagosome network

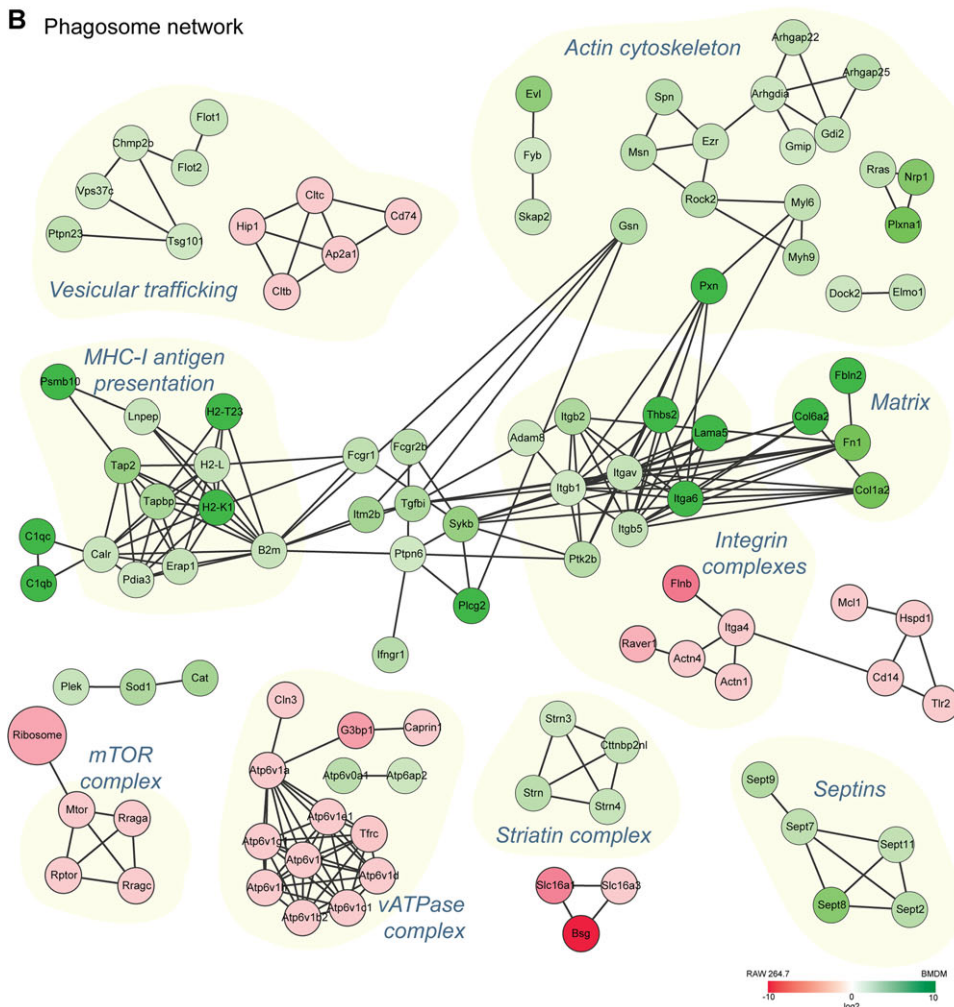


Figure 2. GO-term enrichment and phagosome network analyses. (A) Selected GO terms of proteins enriched in the phagosome proteomes isolated from either BMDM or RAW 264.7 cells. (B) The phagosome network analysis shows cellular processes that were significantly enriched in the phagosomal proteome of BMDM (green) or decreased (red) compared to RAW 264.7 cells. The connection (gray edges) between nodes represents experimentally proven physical interaction between proteins.

(<1% FDR) and quantitation of 2510 proteins (present in two of the three replicates), providing the detailed analysis of the phagosome proteome so far (Supporting Information Table 1). A total of 290 and 226 proteins were significantly enriched (twofold, $p < 0.05$) on BMDM and RAW 264.7 phagosomes, respectively. Fifty eight BMDM and 17 RAW 264.7 proteins were considered unique to their cell line (a minimum of five quantified peptides in one and no quantitative values identified in the other cell type).

GO enrichment analysis of the proteins enriched on RAW 264.7 phagosomes compared to BMDM (Fig. 2A) includes ribosomes and translation proteins, which are probably over-represented as RAW 264.7 is an actively proliferating cell

line compared to BMDMs. Moreover, the vATPase complex is strongly enriched, although our functional data (Fig. 1B) showed a clear delay of the acidification in RAW 264.7. Interestingly, two vATPase subunits, ATP6v0a1 and ATP6ap2 (Renin receptor), are more abundant on BMDM phagosomes (Fig. 2B), suggesting they have an important role in the regulation of the acidifying complex. BMDM phagosomes have protein complexes enriched in the innate immune response, such as TLR 9 and TLR13 as well as complement receptors. Furthermore, they have strongly enriched integrin complexes (including integrins alpha-5, alpha 6, beta-1, beta-2, and beta-5) and express higher levels of lectins such as mannose receptor 1 (MRC1), galectins 1, 3, 8, and 9 as well as

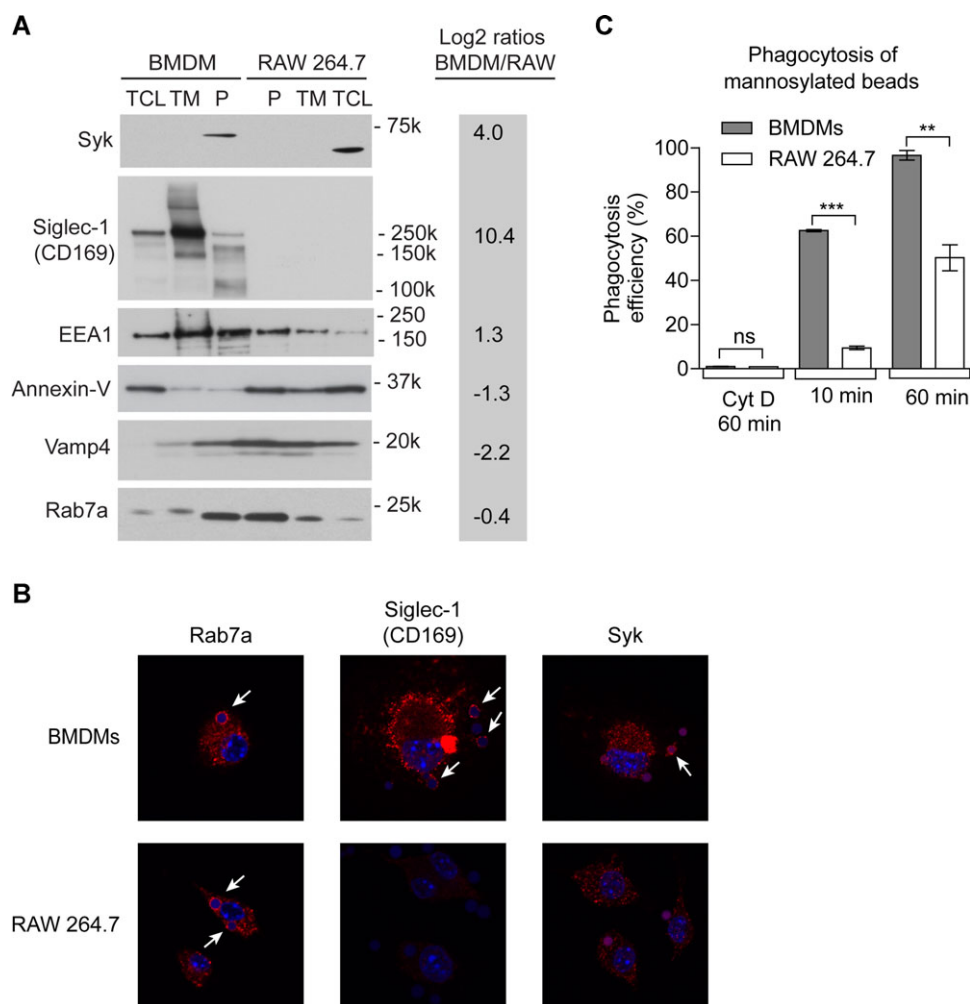


Figure 3. Validation of the proteomics results. (A) Western blotting analysis of six proteins selected from the proteomics data. P, phagosome; TM, total membrane extract; TCL, total cell lysate. Log2 ratios of proteomics results are given for convenience. (B) Fluorescence microscopy of Rab7a, Siglec-1, and Syk proteins showed much stronger expression of Siglec-1 in BMDMs than RAW 264.7 cells. Rab7a was localized to the phagosome in both cell types but Siglec-1 and Syk were only associated with the phagosome in BMDMs. (C) The efficiency of phagocytosis of mannan-coated beads is strongly increased in BMDMs compared to RAW 264.7 cells, due to the much higher expression of mannose receptor 1 (MRC1). Error bars represent standard deviation, pair-wise *t*-test comparison with ****p* < 0.0001 and ***p* < 0.001.

chitinase-like protein 3 (Chil3/Ym1) and Sialoadhesin (Siglec1). Interestingly, BMDMs express also four times more Clec10a (Mgl1) which has been used with MRC1 and Ym1 as markers for alternatively activated macrophages [22]. RAW 264.7, on the other hand, express higher levels of Clec4e (Mincle), Clec4a (Dcir), and Clec7a (Dectin 1), as well as a fourfold higher amount of interferon- γ receptor, suggesting that the basal activation state of RAW 264.7 cells is considerably more proinflammatory compared to BMDMs.

Next, we performed a STRING analysis [23] of the proteins enriched in BMDMs and RAW 264.7 cells (Fig. 2B). This showed that various protein complexes are significantly enriched on BMDM phagosomes, including septin 2, 7, 8, 9, and 11, which have recently been shown to be important for phagosome formation [24], several complexes of cytoskeleton proteins associated with ezrin, moesin and Rock2 and Evl [25], proteins of the ESCRT complexes [26], the striatin complex [27] that has not previously been shown to locate to the phagosome, the above-mentioned integrin complex, and the entire antigen crosspresentation complex associated with Tap2. One would therefore expect considerable differences in

the ability to cross-present antigens via MHC class I between RAW 264.7 cells and BMDMs.

We validated our proteomics results independently using Western blotting for a number of proteins (Fig. 3A). In RAW 264.7 cells, we confirmed a strong phagosomal enrichment of annexin-V representing a larger number of annexins, indicative of a higher phospholipid content on RAW 264.7 phagosomes. Our data further show that Siglec1 is indeed only expressed in BMDMs. Moreover, we noticed that Syk, an important kinase for phagocytosis through various receptors [28], is located strongly to the phagosome in BMDMs, while it is not enriched in RAW 264.7 cells, with the majority being found in the total cell lysate. We further validated this by fluorescence microscopy (Fig. 3B), showing strong enrichment on BMDM phagosomes for Siglec-1 and specific translocation for Syk.

Finally, as discussed above, we identified a >1000-fold larger amount of MRC1 on phagosomes from BMDMs compared to RAW 264.7. This led us to investigate if uptake of beads coated with the polysaccharide mannan (Dill et al., submitted), which is taken up through mannose receptor [29], was affected between the two cell types. Figure 3C

indeed shows an even stronger difference in phagocytic uptake of mannosylated beads than uncoated beads (Fig. 1A), indicating that MRC1 enhances the uptake of these beads in BMDMs.

In conclusion, we provide the largest dataset of the murine phagosome proteome published so far, which will be an important resource for researchers in the phagocytosis and macrophage field. Furthermore, our data indicate that several phagosomal functions such as acidification and proteolysis are significantly reduced in the RAW 264.7 cell line compared to BMDMs from C57/BL6 mice. By showing the molecular differences and similarities on the phagosome of these cell types, we allow researchers to choose the best model for their protein of interest.

The MS proteomics data in this paper have been deposited in the ProteomeXchange Consortium (<http://proteomecentral.proteomexchange.org>) via the PRIDE partner repository [30]: dataset identifier PXD001293. We would like to thank Natalia Shpiro for synthesis of sodium 3-[(2-methyl-2-undecyl-1,3-dioxolan-4-yl)methoxy]-1-propanesulfonate; Carol Clacher, Laura Frew, and Gail Gilmour in Transgenic Services for collection of murine femurs; the MRC PPU mass spec facility and the PRIDE team for support. The College of Life Sciences Data Analysis Group is funded by Wellcome Trust grant 097945/Z/11/Z. This work was funded by Medical Research Council UK and the pharmaceutical companies supporting the Division of Signal Transduction Therapy (DSTT) (AstraZeneca, Boehringer-Ingelheim, GlaxoSmithKline, Janssen Pharmaceutica, Merck KGaA and Pfizer).

The authors have declared no conflict of interest.

References

- Mosser, D. M., Edwards, J. P., Exploring the full spectrum of macrophage activation. *Nat. Rev. Immunol.* 2008, *8*, 958–969.
- Wynn, T. A., Chawla, A., Pollard, J. W., Macrophage biology in development, homeostasis and disease. *Nature* 2013, *496*, 445–455.
- Underhill, D. M., Goodridge, H. S., Information processing during phagocytosis. *Nat. Rev. Immunol.* 2012, *12*, 492–502.
- Boulais, J., Trost, M., Landry, C. R., Dieckmann, R. et al., Molecular characterization of the evolution of phagosomes. *Mol. Syst. Biol.* 2010, *6*, 423.
- Davies, L. C., Jenkins, S. J., Allen, J. E., Taylor, P. R., Tissue-resident macrophages. *Nat. Immunol.* 2013, *14*, 986–995.
- Murray, P. J., Wynn, T. A., Protective and pathogenic functions of macrophage subsets. *Nat. Rev. Immunol.* 2011, *11*, 723–737.
- Jutras, I., Desjardins, M., Phagocytosis: at the crossroads of innate and adaptive immunity. *Annu. Rev. Cell Dev. Biol.* 2005, *21*, 511–527.
- Campbell-Valois, F. X., Trost, M., Chemali, M., Dill, B. D. et al., Quantitative proteomics reveals that only a subset of the endoplasmic reticulum contributes to the phagosome. *Mol. Cell. Proteomics* 2012, *11*, M111 016378.
- Raschke, W. C., Baird, S., Ralph, P., Nakoinz, I., Functional macrophage cell lines transformed by Abelson leukemia virus. *Cell* 1978, *15*, 261–267.
- Ritorto, M. S., Cook, K., Tyagi, K., Pedrioli, P. G., Trost, M., Hydrophilic strong anion exchange (hSAX) chromatography for highly orthogonal peptide separation of complex proteomes. *J. Proteome Res.* 2013, *12*, 2449–2457.
- Trost, M., English, L., Lemieux, S., Courcelles, M. et al., The phagosomal proteome in interferon-gamma-activated macrophages. *Immunity* 2009, *30*, 143–154.
- Marcantonio, M., Trost, M., Courcelles, M., Desjardins, M., Thibault, P., Combined enzymatic and data mining approaches for comprehensive phosphoproteome analyses: application to cell signaling events of interferon-gamma-stimulated macrophages. *Mol. Cell. Proteomics* 2008, *7*, 645–660.
- Bell, C., English, L., Boulais, J., Chemali, M. et al., Quantitative proteomics reveals the induction of mitophagy in tumor necrosis factor-alpha-activated (TNFalpha) macrophages. *Mol. Cell. Proteomics* 2013, *12*, 2394–2407.
- Burckstummer, T., Baumann, C., Bluml, S., Dixit, E. et al., An orthogonal proteomic-genomic screen identifies AIM2 as a cytoplasmic DNA sensor for the inflammasome. *Nat. Immunol.* 2009, *10*, 266–272.
- Weischenfeldt, J., Porse, B., Bone marrow-derived macrophages (BMM): isolation and applications. *CSH Protoc* 2008, *2008*, pdb prot5080.
- Tomida, M., Yamamoto-Yamaguchi, Y., Hozumi, M., Purification of a factor inducing differentiation of mouse myeloid leukemic M1 cells from conditioned medium of mouse fibroblast L929 cells. *J. Biol. Chem.* 1984, *259*, 10978–10982.
- Podinovskaia, M., VanderVen, B. C., Yates, R. M., Glennie, S. et al., Dynamic quantitative assays of phagosomal function. *Curr. Protoc. Immunol.* 2013, *102*, 14.34:14.34.1–14.34.14.
- Yates, R. M., Hermetter, A., Russell, D. G., Recording phagosome maturation through the real-time, spectrofluorometric measurement of hydrolytic activities. *Methods Mol. Biol.* 2009, *531*, 157–171.
- Yates, R. M., Hermetter, A., Taylor, G. A., Russell, D. G., Macrophage activation downregulates the degradative capacity of the phagosome. *Traffic* 2007, *8*, 241–250.
- Desjardins, M., Huber, L. A., Parton, R. G., Griffiths, G., Biogenesis of phagolysosomes proceeds through a sequential series of interactions with the endocytic apparatus. *J. Cell Biol.* 1994, *124*, 677–688.
- Cox, J., Mann, M., MaxQuant enables high peptide identification rates, individualized p.p.b.-range mass accuracies and proteome-wide protein quantification. *Nat. Biotechnol.* 2008, *26*, 1367–1372.
- Martinez, F. O., Helming, L., Gordon, S., Alternative activation of macrophages: an immunologic functional perspective. *Annu. Rev. Immunol.* 2009, *27*, 451–483.
- Franceschini, A., Szklarczyk, D., Frankild, S., Kuhn, M. et al., STRING v9.1: protein-protein interaction networks, with

- increased coverage and integration. *Nucleic Acids Res.* 2013, *41*, D808–D815.
- [24] Huang, Y. W., Yan, M., Collins, R. F., Diccio, J. E. et al., Mammalian septins are required for phagosome formation. *Mol. Biol. Cell* 2008, *19*, 1717–1726.
- [25] Coppolino, M. G., Krause, M., Hagendorff, P., Monner, D. A. et al., Evidence for a molecular complex consisting of Fyb/SLAP, SLP-76, Nck, VASP and WASP that links the actin cytoskeleton to Fcγ receptor signalling during phagocytosis. *J. Cell Sci.* 2001, *114*, 4307–4318.
- [26] Hurley, J. H., The ESCRT complexes. *Crit. Rev. Biochem. Mol. Biol.* 2010, *45*, 463–487.
- [27] Hwang, J., Pallas, D. C., STRIPAK complexes: structure, biological function, and involvement in human diseases. *Int. J. Biochem. Cell Biol.* 2014, *47*, 118–148.
- [28] Tohyama, Y., Yamamura, H., Protein tyrosine kinase, syk: a key player in phagocytic cells. *J Biochem* 2009, *145*, 267–273.
- [29] Eshdat, Y., Ofek, I., Yashouv-Gan, Y., Sharon, N., Mirelman, D., Isolation of a mannose-specific lectin from *Escherichia coli* and its role in the adherence of the bacteria to epithelial cells. *Biochem. Biophys. Res. Commun.* 1978, *85*, 1551–1559.
- [30] Vizcaino, J. A., Côté, R. G., Csordas, A., Dianes, J.A. et al., The PRoteomics IDentifications (PRIDE) database and associated tools: status in 2013. *Nucleic Acids Res.* 2013, *41*, D1063–D1069.