A comparison of two methods for determining titanium dioxide marker content in broiler digestibility studies

N. K. Morgan[1a], D. V. Scholey[1] and E. J. Burton[1]

[1a] *School of Animal, Rural and Environmental Science, Nottingham Trent University, Southwell, Nottinghamshire, England, United Kingdom.*

Corresponding author: Natalie Morgan Email: nat.morgan@ntu.ac.uk

Short title: Titanium dioxide broiler digestibility methodology

Abstract

The use of inert markers in broiler diets eliminates the need to quantitatively evaluate feed intake and excreta output to determine diet digestibility, and enables nutrient uptake at specific points along the gastrointestinal tract to be examined. Titanium dioxide (TiO_2) is commonly used for this purpose and measured using a UV spectrophotometric assay. Two experiments were conducted to observe whether an inductively coupled plasma optical emission spectrophotometer (ICP-OES) assay is able to replace the UV-spectroscopy assay for rapid analysis of TiO_2 in broiler feed and ileal digesta samples. In the first experiment, TiO_2 was added at 5g/kg to 19 broiler diets. Ross 308, male broilers (n=452) fed these diets were involved in a series of digestion studies to determine ileal digesta recovery of TiO_2. In the second experiment, defined amounts of TiO_2 were added to ileal digesta samples from Ross 308, male broilers (n=176) and TiO_2 recoveries were determined. The feed and ileal samples from both experiments were analysed by both UV-spectroscopy and ICP-

OES, and relatedness of the findings from the two assays was determined. Overall relatedness of the two assays was strong for determination of TiO_2 concentration in both the broiler diets and ileal digesta samples (r = 0.908 and r = 0.884 respectively). Overall recovery of supplemented TiO_2 was 97.62% by the UV-spectroscopy assay and 98.77% by the ICP-OES assay. The ICP-OES assay in this study was as accurate as spectrophotometric determination for quantification of TiO_2 content. The ICP-OES method can also be used to analyse several elements within one assay, with a single preparation step, so the measurement of TiO_2 may be incorporated into the analysis of other minerals. Time and resources dedicated to determining diet digestibility in broilers could be minimised by using the ICP-OES assay to replace the UV-spectroscopy assay when measuring TiO_2 concentration.

Keywords: Broiler, Titanium Dioxide, Digestibility, Methodology

Implications

Titanium dioxide (TiO_2) is commonly added as an inert marker to broiler diets to enable diet digestibility to be determined. This study demonstrates that an ICP-OES assay could replace the commonly used UV-spectroscopy assay for the determination of TiO_2 concentration in poultry diets and ileal digesta. This is advantageous because the ICP-OES assay used in this study has comparatively greater detection limits and sensitivity than the UV-spectroscopy assay. Additionally the ICP-OES assay enables TiO_2 determination to be incorporated into other mineral concentration analyses.

Introduction

Inert digestibility markers added to broiler diets eliminate the need to evaluate quantitative feed intake and excreta output, and enable nutrient utilisation to be examined along the gastrointestinal tract (Short *et al.,* 1996). Inert markers must maintain digestive transit at the same speed as other dietary nutrients in the tract and be physiologically inactive, as well as being non-toxic, easily analysed, able to be homogenously mixed into a diet, indigestible and non-absorbed (Jagger *et al.,* 1992; Titgemeyer *et al*., 2001). Titanium dioxide (TiO_2) has some advantages over the commonly used chromic oxide (Cr_2O_3), with studies showing improvements in reproducibility and homogeneity (Jagger *et al*., 1992). TiO_2 is also approved for use as a feed additive by the Food and Drug Administration, unlike Cr_2O_3 (Titgemeyer *et al*., 2001). Another commonly used marker is acid insoluble ash, but it has been suggested that its digestive transit does not accurately reflect that of feed passage (Cheng and Coon 1990).

The method most widely used to determine TiO_2 concentration is UV-spectroscopy, primarily based around the method of Short *et al*. (1996). This method involves the initial hydrolysis of the sample with sulphuric acid followed by a colour reaction. An intense orange/yellow colour results from the addition of hydrogen peroxide to an acidic titanium solution, and the colour intensity can be quantified by UV-spectrometry. This method has been used successfully in several species including poultry (Short *et al*., 1996), cattle (Titgemeyer *et al*., 2001) and pigs (Jagger *et al*., 1992), but some authors reported being unable to achieve reliable results using this process (Myers *et al.,* 2004).

In poultry research TiO_2 as a dietary marker has been used successfully to determine calcium and phosphorus utilisation (Walk *et al*., 2012). Mineral digestibility

and utilisation in poultry is frequently analysed by induced coupled plasma optical emission spectrophotometer (ICP-OES) in preference to UV methods as the ICP-OES assay can be used to analyse many elements in one preparation. Titanium concentration can be detected by ICP-OES, which suggests that there is potential for TiO_2 measurement to be made concurrently with mineral content, thus reducing analysis time and resource use.

A comparison between a UV-spectroscopy assay and ICP-OES assay for determination of TiO_2 has previously been investigated by Boguhn *et al.* (2009) in turkey diets and digesta. In this paper it was suggested that there was incomplete recovery of TiO_2 for both assays used, and hence values read to be lower than expected. However, detailed inspection of the results of the turkey data presented by Boguhn *et al.* (2009) confirms that for some of the samples the readings were higher than expected when the UV-spectroscopy assay was used, and lower than expected when the ICP-OES assay was used. This suggests that potentially that neither, or just one, of the assays is producing values that are representative of the TiO_2 concentration in the sample. It is possible that the UV-spectroscopy assay is amplifying the value, and the ICP-OES assay is not detecting all the TiO_2 in the sample. The conclusion made by Boguhn *et al.* (2009) that both assays can be used to determine TiO_2 may therefore be questionable. Rodehutscord *et al*. (2012) have subsequently used ICP-OES to analyse TiO_2 concentration in broiler ileal digesta indicating that the new ICP methodology is an attractive prospect to workers in the field, but highlighting that this is an area that requires further validation. The aim of this study therefore was to investigate consistency of TiO_2 recovery from an ICP-OES and a UV-spectroscopy assay, and evaluate if the ICP-OES assay can be used as an

alternative to the UV-spectroscopy assay for the determination of TiO_2 as a marker in poultry digestibility studies.

Material and methods

Birds and Husbandry

For experiment 1, Ross 308, male broilers (n=452) were involved in a series of digestion studies to determine ileal digesta recovery of TiO_2 either by UV-spectroscopy by the method of Short *et al*. (1996), or by an inductively coupled plasma optical emission spectrophotometer (ICP-OES) assay. Birds were fed one of 19 experimental diets in mash form, each with TiO_2 added at 5g/kg; 6 semi-synthetic starch dextrose based diets, and 13 more commercial style diets based on cereals including wheat, rapeseed, maize and rye and soya bean meal. All 19 diets were analysed for TiO_2 concentration. Each diet was fed to a minimum of 20 birds. All birds were from breeder flocks aged 42-45 weeks old and were obtained from a commercial hatchery at day of hatch. Chicks were randomised by weight and placed in 0.64 m^2 floor pens in groups of four, bedded on clean wood shavings. Birds were allowed ad libitum access to the treatment diets and water for the duration of the trials; which spanned between two and four weeks. The room was thermostatically controlled to produce an initial temperature of 32°C and reduced to 21°C by day 21. The lighting regimen used was 24 hours light on day 1, with darkness increasing by 1 hour per day until 6 hours of darkness was reached and this was maintained throughout the remainder of the study. Birds were euthanised by cervical dislocation. Digesta sample collection was carried out on a total of 144 14 day-old birds, 144 21 day-old birds and 164 28 day-old birds. At each bird age, digesta was pooled per pen of four birds, and averaged across diet. Digesta content was removed from the

intestinal section distal to the Meckel's diverticulum and proximal to the ileo-ceco-colonic junction of each bird. The digesta samples were then freeze-dried and ground through a 1mm screen.

For experiment 2, Ross 308 male broilers (n=176) were fed a diet that contained no TiO_2 from d0-42. The birds were from a breeder flock age of 43 weeks old, and were obtained from a commercial hatchery at day of hatch. Chick placing, room temperature and lighting regime were as previously described. Birds were allowed ad libitum access to the treatment diets and water for the duration of the trial. Digesta content was removed from the intestinal section distal to the Meckel's diverticulum and proximal to the ileo-ceco-colonic junction of each bird. The samples were freeze-dried and ground through a 1mm screen. TiO_2 was subsequently added to the digesta samples at 0, 5, 10, 15 and 20g/kg to encompass the range found in poultry digestibility studies.

All feed and digesta samples from both experiment 1 and experiment 2 were analysed for TiO_2 concentration by both the UV-spectroscopy and ICP-OES assays described below.

Calibration Standards

250mg titanium dioxide was dissolved in 100ml of 7.4M sulphuric acid (H_2SO_4) and diluted to 500ml with distilled water to produce a standard titanium solution of 0.5mg/ml. This standard solution was used to prepare the calibration curve for both the UV-spectroscopy and ICP-OES assays. For the ICP-OES assay, the TiO_2 standard solution was diluted with ultra-pure water in varying increments to produce

standards between 0 and 10ppm. These standards were measured on an ICP-OES (Optima 2100 DV ICP-OES, model PQ Excell VG Elemental, Perkin-Elmer, USA) set to detect Ti at wavelength 334.936nm, and a calibration curve was derived from the readings. For the UV-spectroscopy assay, graded volumes of TiO_2 standard solution was pipetted into individual 100ml volumetric flasks and made up to 10ml with 7.4M H_2SO_4. 10ml 30% hydrogen peroxide (H_2O_2) was then added to the solutions and the contents were made up to 100ml with distilled water before measurement on a spectrophotometer (Unicam Helios, Berkshire, UK) set at 410 nm.

UV-Spectroscopy Assay

The UV-spectroscopy assay was based on that of Short *et al*. (1996). Briefly, triplicate aliquots (approximately 0.3g) of each digesta sample and 5 replicates of each of the 19 feed samples were ashed in porcelain crucibles for 16 hours at 650°C. Once cooled, 10ml H_2SO_4 (7.4 M) was added to each crucible and the samples were heated for approximately 1 hour until completely dissolved. The contents were then transferred quantitatively into 100ml volumetric flasks via filter papers (Whatman 541) using distilled water. 10ml of 30% H_2O_2 was then added to each flask and the flasks made to volume with distilled water. Solutions were thoroughly mixed prior to reading on a spectrophotometer set at 410nm. Sample analysis was repeated if the Z-value between the same samples exceeded 5%.

ICP-OES Assay

For the ICP-OES assay an aqua regia digestion step was carried out according to AOAC 985.01. Briefly, 10ml of aqua regia (35.5-37.5% hydrochloric acid (HCl) and 68-72% nitric acid (HNO_3) at a ratio of 3:1) was added to 50ml glass conical flasks

containing triplicate aliquots (approximately 0.5g) of each digesta sample and 5 replicates of each feed sample, and left at room temperature (14.4°C +/- 0.15 SEM) for a minimum of 12 hours. The samples were then boiled until completely dissolved, for approximately 1 hour. The contents were then filtered through Whatman 541 filter papers into 50ml volumetric flasks and made to volume with ultra-pure water, before transferral into 15ml tubes. The samples were assayed on an ICP-OES set to detect Ti at wavelength 334.936. Sample analysis was repeated if the Z-value between the same samples exceeded 5%. Four digesta samples were repeated using a reduced sample size (approximately 0.2g) with 8 replicates to assess whether smaller quantities of material were viable for the assay.

Statistical Analysis

All data was analysed using IBM SPSS statistics version 21. T-Tests were conducted to differentiate between means. The relatedness of the readings from each assay was investigated using Pearson product-moment correlation coefficient and interpretations of the strength of the relationship between the two methods was based on guidelines by Cohen (1988); weak relationship r = 0.10 to 0.29, medium relationship r = 0.30 to 0.49 and strong relationship r = 0.50 to 1.0. Linear regressions were calculated using the true and measured titanium concentrations. Significance was accepted at $P < 0.05$.

Results and Discussion

There were no significant differences between any TiO_2 concentrations measured by the UV-spectroscopy assay and the ICP-OES assay. There were consistently strong relationships between the two methods for analysis of TiO_2

concentration in the diets (Table 1) and ileal digesta (Table 2). This suggests that the ICP-OES assay used in this study is successful at identifying diet and ileal digesta TiO_2 concentration, and hence has the potential to replace the widely used UV-spectroscopy assay.

The ICP-OES assay had to be modified to analyse ileal digesta samples in experiment 1 as some of the samples contained TiO_2 levels that saturated the ICP-OES detector, which compromised the sensitivity of the measurement. When a smaller sample size (0.2g) was analysed, the samples all read in the optimum necessary range for detection by the ICP-OES, so smaller quantities can be universally used to avoid any need to dilute the samples with ultrapure water. Coefficients of variation for the smaller sample size were less than 5%.

Relatedness between the two methods in determination of ileal digesta TiO_2 was numerically greater when phytase was included in the diets (Table 2). Phytase improves digestibility and therefore increases TiO_2 digesta content (Rutherfurd *et al*., 2004). The sensitivity of the UV-spectroscopy assay decreases as TiO_2 concentration decreases (Boghurn *et al.,* 2009), whereas the sensitivity of the ICP-OES assay is consistent and not dictated by concentration in the sample. This suggests that in the presence of high TiO_2 concentration, such as in the digesta samples from birds fed phytase, the two assays were similar in sensitivity, but in the samples with lower TiO_2 concentration the similarity in sensitivity between the two assays reduced, and the UV-spectroscopy assay was comparatively less reliable. This also potentially explains why observed deviances in TiO_2 level in the diet away from the supplemented 5g/kg were greater when analysed by UV-spectroscopy than by ICP-OES. The observed deviances are likely because dietary TiO_2 levels were measured per kg feed.

In this study there were no significant differences between the measured values, or between the calculated slopes determined by the two assays for the analytical recoveries of TiO_2, whereas previous research has shown marked differences between the two assays (Boguhn *et al.,* 2009). Also, Boguhn *et al.* (2009) found that values from the ICP-OES assay were lower than the expected values, which was not the case in this study (Table 1 and 2). This may be due to the shorter digestion time used (25 minutes in contrast to 60 minutes), so there may have been incomplete dissolution of the samples. Further verification of full Ti recovery was made in the second study where known amounts of Ti were added to digesta before quantification analysis via both methods. This found consistently strong relationships between the two methods at the different TiO_2 supplementation levels in the digesta samples (Table 3) and that the slopes produced by both methods were almost identical. The observed recovery of supplemented TiO_2 was 97.62% by the UV-spectroscopy assay and 98.77% by the ICP-OES assay in this study.

The main advantage of the ICP-OES assay when compared to the UV-spectroscopy is that the former has been shown to be more sensitive at quantitative analysis with improved detection limits. The ICP-OES assay is also less time-consuming, and the ICP-OES enables several elements to be detected in parallel which reduces preparation time and the amount of sample, and hence potentially the number of birds, required.

There are however, some advantages to the UV-spectroscopy assay compared with the ICP-OES assay. The ICP-OES assay is more expensive due to the cost to run the ICP-OES and to maintain the argon gas supplies, although this is mitigated by the potential for concurrent mineral analysis. The ICP-OES assay is also more hazardous as involves the use of aqua regia which is moderately more

corrosive than sulphuric acid. Furthermore the detection range is greater in the UV-spectroscopy method which reduces any potential need for dilution of samples, but in this study, a reduced sample weight (0.2g) was shown to overcome any requirement for dilution with the ICP-OES method.

In conclusion, the ICP-OES assay used in this study was successful at determining TiO_2 added as an inert marker in broiler digestibility studies, and could replace the widely used UV-spectroscopy assay. The ICP-OES assay is more sensitive at quantitatively analysing TiO_2 concentration, consumes less time than the UV-spectroscopy assay, and allows the TiO_2 determination to be carried out concurrently with other mineral analysis by ICP-OES. However it is essential that the current sample weight (0.2g digesta) is used for detection.

References

Boguhn J, Baumgärtel T, Dieckmann A and Rodehutscord, M 2009. Reaserch Note: Determination of titanium dioxide supplements in different matrices using two methods involving photometer and inductively coupled plasma optical emission spectrometer measurements. Archives of Animal Nutrition 63, 337-342.

Cheng TK and Coon CN 1990. Research Note: Calcium digestibility studies utilizing acid-insoluble ash measurements. Poultry Science 69, 2228-2230.

Cohen, J.W 1988. Statistical power analysis for the behavioural sciences (2nd edn). Hillsdale, Lawrence Erlbaum Associates, NJ, pp 79-81.

Jagger S, Wiseman J, Cole DJA and Craigon J 1992. Evaluation of inert markers for the determination of ileal and faecal apparent digestibility values in the pig. British Journal of Nutrition 68, 729-739.

Myers WD, Ludden PA., Nayigihugu V and Hess BW 2004 Technical Note: A procedure for the preparation and quantitative analysis of samples for titanium dioxide. Journal of Animal Science 82, 179-183.

Rodehutscord M, Dieckmann A, Witzig M and Shastak Y 2012. A note on sampling digesta from the ileum of broilers in phosphorus digestibility studies. Poultry Science 91, 965-971.

Rutherfurd SM, Chung TK, Morel PCH and Moughan PJ 2004. Effect of microbial phytase on ileal digestibility of phytate phosphorus, total phosphorus, and amino acids in a low-phosphorus diet for broilers. Poultry Science 83, 61–68.

Short FJ, Gorton, P and Wiseman J 1996. Determination of titanium dioxide added as an inert marker in chicken digestibility studies. Animal Feed Science and Technology 59, 215–221.

Titgemeyer EC , Armendariz CK, Bindel DJ, Greenwood RH and Löest C A 2001. Evaluation of titanium dioxide as a digestibility marker for cattle. Journal of Animal Science 79, 1059–1063.

Walk CL, Bedford MR and McElroy AP 2012. Influence of limestone and phytase on broiler performance, gastrointestinal pH, and apparent ileal nutrient digestibility. Poultry Science 91, 1371-1378.

Table 1 Relatedness of an ICP-OES assay and UV-spectroscopy assay for determination of TiO_2 concentration in broiler diets[a] (Experiment 1)

	Method of TiO_2 Determination (g/kg)		
Diet	ICP-OES	UV-spectroscopy	Relatedness[b]
Semi-synthetic starch dextrose[c]	6.03	6.29	0.684
Wheat Soyabean[d]	5.93	5.69	0.794
Wheat Soyabean 0FTU/kg phytase	5.85	5.97	0.778
Wheat Soyabean 500FTU/kg phytase	5.71	6.08	0.759
Wheat Soyabean 5000FTU/kg phytase	6.64	6.97	0.708
Wheat Rapeseed 0FTU/kg phytase	6.11	6.53	0.886
Wheat Rapeseed 500FTU/kg phytase	4.90	5.08	0.866
Wheat Rapeseed 5000FTU/kg phytase	6.49	6.53	0.963
Maize Rapeseed	6.87	6.98	0.995
Maize Soyabean	4.99	4.88	0.956
Maize, Rye, Wheat, Soyabean	4.87	5.16	0.758
Maize, Rye, Soyabean	5.75	5.47	0.689
SEM	0.14	0.23	

[a] Represent the average of a minimum of 5 replicates per diet, measured as per kg feed.
[b] Strength of the relationship between the ICP-OES and UV-Spectroscopy method for Ti measured in each diet where confidence in the result is $P<0.05$.
[c] Represents the average measured TiO_2 content of 6 semi-synthetic starch dextrose based diets
[d] Represents the average measured TiO_2 content of 3 wheat soyabean meal based diets

Table 2 Relatedness of an ICP-OES assay and UV-spectroscopy assay for determination of TiO_2 concentration in broiler ileal digesta[a] (Experiment 1)

	Method of TiO_2 Determination (g/kg)		
	ICP-OES	UV-spectroscopy	Relatedness[b]
Semi-synthetic starch dextrose[c]	13.58	13.40	0.776
Wheat Soyabean[d]	13.99	13.53	0.550
Wheat Soyabean 0FTU/kg phytase	13.43	13.65	0.512
Wheat Soyabean 500FTU/kg phytase	15.63	15.87	0.822
Wheat Soyabean 5000FTU/kg phytase	13.32	12.42	0.887
Wheat Rapeseed 0FTU/kg phytase	13.16	12.48	0.529
Wheat Rapeseed 500FTU/kg phytase	14.19	14.95	0.613
Wheat Rapeseed 5000FTU/kg phytase	12.92	12.71	0.858
Maize Rapeseed	12.23	12.01	0.584
Maize Soyabean	12.49	12.99	0.726
Maize, Rye, Wheat, Soyabean	12.33	12.04	0.563
Maize, Rye, Soyabean	12.19	12.06	0.646
SEM	0.20	0.26	

[a] Represent the average response of a minimum of 20 birds per diet, 452 birds in total, with digesta samples collected at age 14, 21 or 28 days post-hatch. Analysis was replicated a minimum of 3 times per digesta sample.

[b] Strength of the relationship between the ICP-OES and UV-Spectroscopy method for Ti measured in each digesta sample where confidence in the result is $P<0.05$.

[c] Represents the average measured TiO_2 content of ileal digesta from birds fed one of 6 semi-synthetic starch dextrose based diets, from 32 birds per diet, 192 birds in total, fed as 8 pens of 4 birds per diet

[d] Represents the average measured TiO_2 content of ileal digesta from birds fed one of 3 wheat soyabean meal based diets, from 64 birds per diet, 192 birds in total, fed as 16 pens of 4 birds per diet

Table 3 Calculated slopes of linear regressions and relatedness of an ICP-OES assay and UV-spectroscopy assay for determination of TiO_2 recovery at different levels in broiler ileal digesta[a] (+/- SEM) (Experiment 2)

TiO_2 added to sample (g/kg)	Method of TiO_2 Determination (g/kg)		Relatedness[b]
	ICP-OES	UV-spectroscopy	
0	0.13 (+/- 0.01)	0.15 (+/- 0.03)	0.952
5	4.94 (+/- 0.24)	4.79 (+/- 0.32)	0.745
10	10.06 (+/- 0.29)	9.84 (+/- 0.21)	0.868
15	14.80 (+/- 0.23)	14.63 (+/- 0.27)	0.918
20	20.04 (+/- 0.20)	19.74 (+/- 0.44)	0.734
Slope[c]	0.999	0.998	

[a] Represents the average response of spiked digesta pooled from 176 birds aged 42 days post-hatch. Analysis was replicated 10 times per sample.

[b] Strength of the relationship between the ICP-OES and UV-Spectroscopy method for Ti measured in each digesta sample where confidence in the result is $P<0.05$.

[c] Linear regressions where y was the measured titanium concentration and x was the true titanium concentration.