Northumbria Research Link

Citation: Rodrigues, Antenor, Louvaris, Zafeiris, Dacha, Sauwaluk, Janssens, Wim, Pitta, Fabio, Vogiatzis, Ioannis, Gosselink, Rik and Langer, Daniel (2020) Differences in Respiratory Muscle Responses to Hyperpnea or Loaded Breathing in COPD. Medicine & Science in Sports & Exercise, 52 (5). pp. 1126-1134. ISSN 0195-9131

Published by: Lippincott Williams & Wilkins

URL:	https://doi.org/10.1249/mss.000000000002222
<https: 10.1249="" doi.org="" mss.000000000002222=""></https:>	

This version was downloaded from Northumbria Research Link: http://nrl.northumbria.ac.uk/id/eprint/42721/

Northumbria University has developed Northumbria Research Link (NRL) to enable users to access the University's research output. Copyright © and moral rights for items on NRL are retained by the individual author(s) and/or other copyright owners. Single copies of full items can be reproduced, displayed or performed, and given to third parties in any format or medium for personal research or study, educational, or not-for-profit purposes without prior permission or charge, provided the authors, title and full bibliographic details are given, as well as a hyperlink and/or URL to the original metadata page. The content must not be changed in any way. Full items must not be sold commercially in any format or medium without formal permission of the copyright holder. The full policy is available online: http://nrl.northumbria.ac.uk/policies.html

This document may differ from the final, published version of the research and has been made available online in accordance with publisher policies. To read and/or cite from the published version of the research, please visit the publisher's website (a subscription may be required.)



University Library

1	Differences in respiratory muscle responses to hyperpnea or loaded breathing in COPD
2	
3	Antenor Rodrigues ^{1,2,3*} , Zafeiris Louvaris ^{1,2,4*} , Sauwaluk Dacha ^{1,2,5} , Wim Janssens ^{1,2} , Fabio
4	Pitta ³ , Ioannis Vogiatizis ^{4,6} , Rik Gosselink ^{1,2} , Daniel Langer ^{1,2}
5	
6	¹ Faculty of Kinesiology and Rehabilitation Sciences, Department of Rehabilitation
7	Sciences, Research Group for Rehabilitation in Internal Disorders, KU Leuven.
8	² Respiratory Rehabilitation and Respiratory Division, University Hospital Leuven,
9	Belgium.
10	³ Laboratory of Research in Respiratory Physiotherapy (LFIP), Department of
11	Physiotherapy, Universidade Estadual de Londrina (UEL), Londrina, Brazil.
12	⁴ 1 st Department of Critical Care Medicine and Pulmonary Services, GP Livens and M
13	Simou Laboratories, Medical School of Athens University, Evaggelismos Hospital, Athens,
14	Greece.
15	⁵ Faculty of Associated Medical Sciences, Department of Physical Therapy, Chiang Mai
16	University, Chiang Mai, Thailand.
17	⁶ Department of Sport, Exercise and Rehabilitation, Faculty of Health and Life Sciences,
18	Northumbria University Newcastle, UK.
19	* These authors have contributed equally to the study, and share first authorship
20	
21	Address for correspondence: Dr. Daniel Langer, Department of Rehabilitation Sciences, KU
22	Leuven, Leuven 3001, Belgium; daniel.langer@kuleuven.be

23 Abstract

Introduction: To compare acute mechanical and metabolic responses of the diaphragm and rib
cage inspiratory muscle during two different types of respiratory loading in patients with COPD.

Methods: In 16 patients (age:65±13, 56% male, FEV1:60±6%pred, Pimax:82±5%pred) assessments of respiratory muscle electromyography (EMG), esophageal (Pes) and gastric (Pga) pressures, breathing pattern, and noninvasive assessments of systemic (VO₂, cardiac output, oxygen delivery and extraction) and respiratory muscle hemodynamic and oxygenation responses (blood flow index [BFI], oxygen delivery index, deoxyhemoglobin concentration [HHb] and tissues oxygen saturation [StiO₂]), were performed under two different conditions of respiratory muscle loading (hyperpnea and loaded breathing).

33 **Results:** During hyperpnea, breathing frequency, minute ventilation, esophageal and diaphragm pressure-time product (PTP)/min, cardiac output and VO2 were higher than during loaded 34 35 breathing (P < 0.05). Average inspiratory Pes and Pdi per breath scalene (SCA), 36 sternocleidomastoid (SCM), and intercostal muscle activation was higher during loading 37 breathing (P < 0.05). Higher Pdi during loaded breathing compared to hyperpnea was due to higher Pes (P < 0.05). Diaphragm activation, inspiratory and expiratory Pga and expiratory 38 39 abdominal muscle activation did not differ between the two conditions (P>0.05). SCA-BFI and 40 oxygen delivery index were lower and SCA-HHb was higher during loaded breathing. 41 Furthermore, SCA and intercostal muscle StiO₂ were lower during loaded breathing compared to 42 hyperpnea (*P*<0.05).

43 Conclusion: Greater inspiratory muscle effort during loaded breathing evoked larger ribcage and
44 neck muscle activation compared to hyperpnea. In addition, lower SCA and intercostal muscles

45 StiO₂ during loading breathing than during hyperpnea might indicates a mismatch between46 inspiratory muscle oxygen delivery and utilization.

47 Key Words: RESPIRATORY MUSCLE ACTIVATION, RESPIRATORY MUSCLE
48 LOADING, RESPIRATORY MUSCLE METABOLISM, RESPIRATORY MUSCLE
49 TRAINING.

50 INTRODUCTION

Improvements in both respiratory muscle endurance and strength can be observed in 51 patients with COPD after either whole-body, or specific respiratory muscle endurance 52 53 training.(1-3) The improvements in respiratory muscle function in response to endurance training 54 are probably mainly due to the increased ventilatory demands induced by (exercise) hyperpnea. 55 Hyperpneic training provides a high respiratory flow / low resistance stimulus to the respiratory 56 muscles during a high number of consecutive repetitions.(2, 3) It has also been demonstrated that 57 adding specific hyperpneic (i.e. endurance) respiratory muscle training can enhance the effects of 58 whole body endurance training on respiratory muscle endurance. However, larger improvements in inspiratory muscle strength (i.e., pressure generating capacity) have been reported after 59 60 specific inspiratory muscle strength training (IMT) in comparison with whole body endurance 61 training (i.e. average increases of 16 vs 6 cmH₂O in maximal inspiratory mouth pressure [MIP] During inspiratory muscle strength training loading is induced by 62 respectively).(4, 5) 63 overcoming a "high external resistance" during a limited number of breathing cycles per session (e.g. 30-40 full vital capacity breaths against loads equaling about 30-50% of MIP).(4) 64 Therefore, as much as limb muscles respond distinctively to endurance and strengthening 65 66 stimuli, (6, 7) it can also be expected that different responses are induced when the respiratory muscles are exposed to "endurance" (i.e., hyperpnea) or "strengthening" (i.e., loaded breathing) 67 68 stimuli. Differences in acute responses to either endurance or strengthening stimuli imposed on 69 the respiratory muscle in terms of muscle recruitment and activation patterns as well as local and systemic oxygenation responses have however, to the best of our knowledge, never been 70 71 comprehensively characterized. Therefore, we aimed to explore and compare the acute responses

of a number of physiological variables during these two different types of inspiratory muscleloading in patients with COPD..

74 METHODS

Subjects. Sixteen symptomatic patients (Baseline Dyspnea Index 6 ± 1),(8) with a clinical diagnosis of COPD according to the Global Initiative for Chronic Obstructive Pulmonary Disease (GOLD),(9) aged between 55 and 74 years (see online supplement) were included in the study. The study was approved by the local hospital ethics committee (reference number: S58513). Before participation in the study, all patients were informed about potential risks and discomforts associated with performing the experiments and provided written informed consent.

81 Study design. Experiments were performed on two visits. During the first visit (i.e., 82 initial testing) patients performed comprehensive pulmonary function testing.(10, 11) Maximal inspiratory muscle strength was measured by maximum inspiratory mouth pressures (MIP).(12, 83 84 13) An incremental cardiopulmonary exercise test (CPET),(14) and a constant work rate cycle 85 endurance test (CWRT),(14) were also performed during this visit (see supplemental online material for more details). During the second visit, patients performed, in random order, both a 86 Normocaphic Hyperphea trial (hyperphea), (13, 15) reproducing the ventilatory responses (i.e., 87 88 mean tidal volume, breathing frequency and minute ventilation) recorded for each patient during 89 the CWRT,(15) and a Tapered Flow Resistive Loading task (loaded breathing) reproducing 90 ventilator loading during a high-intensity IMT session. (13, 16) Both tasks were performed for 91 five minutes. Breathlessness was measured by the modified Borg scale at rest and at the end of each task.(17) Additionally, during the final 60 seconds of both the hyperpnea and loaded 92 93 breathing tasks, respiratory muscle perfusion,(18) oxygen delivery,(19) respiratory muscle 94 activation (root mean square EMG%max) and respiratory effort were assessed.(13, 20-23)

Metabolic and ventilatory variables were also assessed breath by breath during both tasks by a 95 96 metabolic cart (Vmax 229; Sensor Medics, Anaheim, CA, USA).

97 Hyperpnea. Patients were requested to maintain tidal volume, breathing frequency and minute ventilation reproducing their own breathing responses recorded during the CWRT for 98 99 five minutes.(15) Thus, during the test investigators provided continuous verbal guidance aiming 100 to maintain a maximum variation in minute ventilation of 5% throughout the test.(15) Visual 101 feedback on breathing parameters was also provided on a screen displayed in front of the patient 102 so as to adjust his/her breathing frequency and tidal volume to the level required by the 103 investigator. Normocapnia was maintained by having subjects inspire from a Douglas bag 104 containing 5% CO₂, 21% O₂ and 74% N₂ for balance, connected to a two-way non-rebreathing 105 valve (model 2700, Hans Rudolph) by a piece of tubing.(15)

106 Loaded breathing. The loaded breathing training session was performed in accordance 107 with previously published protocols of IMT using the electronic POWERbreathe KH2 108 device.(16) Subjects were requested to breathe out completely (i.e., until residual volume) 109 through a loaded breathing device (POWERbreathe KH2) followed by full vital capacity 110 inspirations against an external resistance of ~50% of patients MIP for 30 breaths or for a 111 minimum of five minutes.(16) Thereby loading the inspiratory muscles throughout their full 112 range of motion in accordance with a previously published method.(16)

113 Respiratory muscle pressures, work of breathing and activation during hyperpnea 114 and loaded breathing. Respiratory muscle pressures and diaphragm activation (EMGdi) were measured by a combined multipair esophageal electrode catheter with esophageal- and gastric-115 116 balloons (Yinghui Medical Equipment Technology Co. Ltd., Guangzhou, China) nasally inserted 117 after topical anesthesia. Procedures for optimal positioning of the catheter and signal processing 118 have already been published.(20) EMGdi was converted into root mean square (RMS), 119 normalized by its maximum activation during inspiratory capacity maneuvers (ICs) and reported 120 as percentage of maximum activation (EMGdi, %max). Continuous measurements of esophageal 121 (Pes), gastric (Pga) and transdiaphragmatic (Pdi, i.e., Pga - Pes) pressures and its derivatives 122 were performed. Inspiratory Pes, Pga and Pdi max were obtained during inspiratory sniff 123 maneuvers.(20) Expiratory Pga max, however, was obtained during forced expiratory capacity 124 maneuvers (see online supplement). Ribcage, i.e., scalene (SCA), sternocleidomastoid (SCM), parasternal intercostal and 7th intercostal (ICM and 7thICM, respectively), and abdominal (ABD) 125 126 muscle activation was measured by surface electromyography (sEMG) (Desktop Direct 127 Transmission (DTS), NORAXON, Scottsdale, USA).(21) Electrodes were placed (1) on the 128 posterior left triangle of neck at the level of cricoid process for scalene muscle measurements 129 (EMGsca), (2) at the midpoint along the long axis of the right sternocleidomastoid muscle 130 between the mastoid process and the medial clavicle for sternocleidomastoid muscle measurement (EMGscm), (3) at the right parasternal space of the 2nd and 3rd rib 3 cm lateral to 131 132 the sternum for parasternal intercostal muscle measurements (EMGpicm), (4) at the line between the 7th and 8th intercostal space at mid axillary line for 7th intercostal muscle measurements 133 (EMG 7th icm), (5) over upper 1/3 of rectus abdominis under costal cartilage level (EMGabd) 134 135 (see online supplement)..

Systemic hemodynamic and vascular responses during loaded breathing and hyperpnea. Cardiac output, heart rate and stroke volume were continuously measured by a commercial impedance cardiography device (PhysioFlowPF50; Manatec Biomedical, Macheren, France) previously validated for COPD patients (see online supplement).(24) Estimated systemic oxygen delivery was calculated by the product of cardiac output and arterial oxygen content. The latter was calculated as the product of 1.39 x hemoglobin concentration [Hb] and %SpO₂.(<u>25</u>)
Arterio-venous oxygen content (i.e., a-vO₂ diff) difference was calculated by dividing oxygen
uptake by cardiac output. The systemic oxygen extraction ratio was calculated as the ratio of the
a-vO₂ diff to arterial oxygen content. In addition, systemic vascular conductance was calculated
by dividing cardiac output by mean arterial blood pressure.

Respiratory muscles perfusion and oxygenation responses. SCA, SCM and 7thICM, 146 147 and ABD blood flow indices (BFI) were calculated by using two commercial Near-Infrared 148 Spectroscopy (NIRS; NIRO-200 and a NIRO-200NX; HAMAMATSU Photonics KK) devices in 149 combination with light-absorbing indocyanine green dye (ICG) that was injected through a 150 peripheral venous catheter as previously described and validated for patients with COPD (see 151 online supplement). For the above-mentioned respiratory muscles oxygen delivery index was 152 calculated by the product of BFI and arterial oxygen content. NIRS optodes were placed at the right posterior triangle of the neck, the left 7th intercostal space and over the upper 1/3 of rectus 153 154 abdominis below costal cartilage level to respectively measure SCA, 7thICM and rectus 155 abdominis muscle perfusion. ICG injections for calculating BFI were performed during the last 5 156 breaths during loaded breathing and during the last 30 seconds of the hyperpnea trial.

NIRS-derived changes in respiratory muscle deoxyhemoglobin concentration ([HHb])
was used as an index of respiratory muscle oxygen extraction.(26) NIRS-derived tissue oxygen
saturation index (i.e., StiO₂) was considered as a measure of the dynamic balance between local
tissue oxygen delivery and utilization (27) and, therefore, local muscle capacity to match oxygen
supply relative to its metabolic demand (see online supplement)).

162 Statistical analysis. A power >0.99 was found based on the difference between SCM
163 muscle activation (EMGscm,%max) between the three tasks (i.e., rest, hyperpnea and loaded

164 breathing, see *Data analysis section* in the online supplement). Data are expressed as mean \pm SE 165 or mean difference (95% confidence interval). Mean respiratory muscle activation, respiratory 166 pressures and its derivatives, breathing pattern variables and central hemodynamic and metabolic 167 variables during the last 60 seconds of rest, hyperpnea and loaded breathing were compared by 168 one-way repeated measures ANOVA when normal distribution was not violated. Otherwise, the 169 Friedman test was used. When statistical significance was met (P < 0.05) pairwise comparisons 170 with Holm correction were performed as post-hoc analyses. Changes in respiratory muscle 171 perfusion and oxygenation responses from rest to hyperpnea versus rest to loaded breathing were 172 compared by paired t-tests when normally distributed, or by Mann-Whitney tests if normal 173 distribution assumptions were not met (see online supplement).

174

RESULTS

Subjects characteristics. Subjects' characteristics are described in detail in Table 1. The 175 176 sample was well balanced regarding sex and composed by patients classified as having mild to 177 very severe COPD presenting resting lung hyperinflation (i.e., increased RV/TLC) (see Subjects 178 characteristics in the supplemental material for more details). Six out of the sixteen included 179 were not willing (n = 5) or able (n = 1) to undergo measurements of EMGdi, Pes and Pga with the 180 esophageal catheter system. Three patients did not have respiratory muscle perfusion measured 181 due to either technical reasons (n=1) or because of contraindications regarding ICG injections 182 (n=2). Hence, nine out of the sixteen patients had concurrent measurements of EMGdi, 183 respiratory pressures and respiratory muscle perfusion. There were no differences regarding pulmonary function, peak exercise and inspiratory muscle capacity between subjects with 184 185 EMGdi and respiratory pressures measurements versus those subjects not able or not willing to 186 undergo these specific experimental procedures.

187 **Respiratory symptoms during hyperpnea and loaded breathing tasks.** Neither 188 breathlessness nor respiratory effort sensations were statistically different between hyperpnea 189 and loaded breathing $(5 \pm 1 \text{ vs. } 4 \pm 1, P=0.15 \text{ and } 5 \pm 1 \text{ vs. } 5 \pm 1, P=0.93, \text{ respectively}).$

190 **Respiratory muscle activation.** We observed similar levels of diaphragm activation 191 (EMGdi%max) (Figure 1a) between hyperpnea and loaded breathing (P= 0.35). SCM, SCA and 192 both intercostals muscle activation (i.e., parasternal and 7th intercostal) were significantly higher 193 during loaded breathing as compared to hyperpnea (Figures 1b – 1e). There were no significant 194 differences between expiratory activation of the abdominal muscle between hyperpnea and 195 loaded breathing (EMGabd, %max: 33 ±4 vs. 30 ± 6, respectively; P=0.27).

196 **Respiratory pressures and work of breathing.** Diaphragmatic and esophageal pressures 197 per breath were significantly higher during loaded breathing in comparison to hyperpnea, gastric 198 pressure, however, was similar between the two conditions (P=0.64; Table 2). Pes PTP and Pes 199 WOB/b were significantly higher during loaded breathing in comparison to hyperpnea (Table 2). 200 Inspiratory Pga and Pdi WOB/b were significantly greater during loaded breathing as compared 201 to hyperpnea (Table 2). Pes WOB/min, and Pdi WOB/min tended to be higherduring loaded 202 breathing in comparison to hyperpnea (P=0.06 and P=0.08 respectively), but Pga WOB/min was 203 similar (P=0.96) between the two conditions. Pes, Pga and Pdi PTP/min responses during 204 hyperpnea were significantly higher as compared to loaded breathing (Table 2). There were no 205 significant differences in expiratory Pga between hyperpnea and loaded breathing (P=0.83; 206 Table 2).

Breathing pattern. In comparison to hyperpnea, absolute and relative inspiratory
 volumes were significantly higher during loaded breathing. Respiratory rate and minute
 ventilation however, was significantly lower during loading breathing compared to hyperpnea

210 (P<0.05; Table 2). Peak inspiratory flow was similar (P=0.20) and accompanied by longer 211 inspiratory time and lower duty cycle during loaded breathing in comparison to hyperpnea 212 (P<0.05; Table 2). During hyperpnea, end-inspiratory lung volume (EILV) achieved $81 \pm 2\%$ of 213 the vital capacity and during loaded breathing achieved $59 \pm 4\%$ of the vital capacity. 214 Representing an end-inspiratory reserve volume of 1.76 ± 0.12 L during hyperpnea and $2.90 \pm$ 215 0.24 during loaded breathing (P < 0.001).

Systemic hemodynamic, metabolic and cardiovascular responses. Cardiac output,
VO₂, a-vO₂ diff and systemic vascular conductance responses were significantly greater during
hyperpnea than during loaded breathing (P<0.05; Table 3). Mean arterial blood pressure did not
significantly differ between the two conditions (Table 3).

220 Respiratory muscle perfusion and oxygenation responses. Increases from rest in 221 SCABFI and oxygen delivery index were significantly less during loaded breathing as compared 222 to hyperpnea (P<0.05; Table 4). The change from rest in SCA oxygen extraction (i.e., [HHb]) 223 was significantly higher during loading breathing as compared to hyperpnea (P < 0.05; Table 4). 224 During loading breathing SCA-StiO₂ decreased from rest whilst during hyperpnea SCA-StiO₂ 225 increased leading to a significant difference in SCA-StiO₂ between the two conditions (P<0.05; Table 4). Increases from rest in 7thICMBFI and oxygen delivery index were less (but not 226 227 significant P=0.27 and P=0.26, respectively) during loaded breathing as compared to hyperpnea. The change from rest in 7thICM-HHB tended to be higher during loaded breathing as compared 228 to hyperpnea (P=0.06). During loading breathing 7thICM -StiO₂ decreased from rest whilst during 229 hyperpnea 7thICM -StiO₂ increased leading to a significant difference in 7thICM -StiO₂ between 230 231 the two conditions (P<0.05; Table 4). No significant changes in BFI (P=0.09), oxygen delivery

232 (P=0.10), [HHB] (P=0.11), and StiO₂ (P=0.50) were observed for the ABDs between loaded 233 breathing and hyperpnea.

234 DISCUSSION

235 Main findings. Our key findings are that by engaging either in hyperpnea (endurance 236 stimulus) or loaded breathing (strength stimulus) differences in both local (i.e., respiratory 237 muscle) and systemic responses are evoked in patients COPD. In both conditions the increase in 238 systemic and respiratory muscle hemodynamics from rest seems to increase in association with 239 the increase in VO₂, (Tables 3 and 4). Loaded breathing elicited greater activation of the SCA, SCM ICM and ^{7th}ICM and inspiratory muscle and reduction in SCA and 7thICM-StiO₂ (Figure 1 240 241 and Table 4, respectively) compared to hyperpnea, thus reflecting the additional burden imposed 242 on these muscles by a strengthening stimulus in comparison to an endurance loading stimulus 243 (Table 2). In addition, increases in diaphragmatic activation during hyperpnea and loaded 244 breathing relative to resting breathing were of similar magnitude in both conditions (Figure 1).

Respiratory muscle activation during loaded breathing and hyperpnea. The 245 246 contribution of SCA, SCM and intercostal muscles to the act of breathing is known to be 247 amplified with increased ventilatory demands.(28-30) Additionally, increased lung volumes are 248 known to impact on the length-tension relationship of the diaphragm, by moving it away from its 249 optimal length to generate pressure.(31-33).(31). Notably, as compared to diaphragm, increased 250 lung volumes ensuing less length-tension impairment of SCA, SCM and intercostal muscles. 251 These muscle undergo less severe length changes resulting in "less" sub-optimal length at higher volumes, (34-36) thereby relatively preserved pressure generating capacity. (36) Thus, SCA, 252 253 SCM and intercostal muscles recruitment enables the respiratory system to compensate for the 254 lost efficiency of the diaphragm by increasing lateral, dorsoventral (i.e., intercostals), and cranial 255 (i.e., SCA and SCM) displacement of the rib cage.(31), SCA, SCM and intercostal muscles 256 recruitment serves as a reserve to overcome increasing demands imposed on the respiratory 257 system under these conditions (i.e., performing faster and deeper inspiratory maneuvers as well 258 as against higher loads).(34, 35) In our study, the recruitment of SCA, SCM and both intercostal 259 muscles was further amplified during loaded breathing (Figure 1) when an additional external 260 load was imposed on the respiratory system in addition to higher inspiratory volumes and flow 261 rates. This resulted in further increases in respiratory demands (i.e., increased inspiratory 262 pressures, WOB and PTP; Table 2). Furthermore, increases in inspiratory Pdi during loaded 263 breathing (in comparison with hyperpnea) were mostly due to more negative inspiratory Pes but 264 not more positive Pga (Table 2; see online supplement). These findings suggest that SCA, SCM 265 and intercostal muscles were preferably recruited to perform the additional work imposed on the 266 inspiratory muscles during loaded breathing.

267 Systemic and respiratory muscle metabolism during loaded breathing and hyperpnea.

268 It is know that during exercise systemic responses such as cardiac output and VO₂ increases 269 proportionally to the work being performed by the working muscles per unit time.(<u>37-39</u>) Our 270 study further supports these relations by demonstrating that increases in both VO₂ and cardiac 271 output during hyperpnea and loaded breathing appeared to have strong associations with PTP 272 expressed per minute rather than per breath (Figure 2). Highlighting that increases in respiratory 273 muscle oxygen requirements (i.e., cost of breathing) seems to be associated with the cumulative 274 respiratory muscle effort that is developed during a given task rather than the respiratory muscle 275 effort of each breath of a given task (figure 2).

276 The higher levels of both systemic and respiratory muscle oxygen extraction (i.e., a-vO₂
277 difference and oxygen extraction and [HHb], respectively) during hyperpnea in comparison to

278 loaded breathing were accompanied by sufficient increase in both systemic and respiratory 279 muscle oxygen delivery (Tables 3 and 4), thereby preserving the balance between respiratory 280 muscle oxygen delivery and utilization (i.e., StiO₂; Table 4). During loaded breathing, however, 281 despite higher respiratory pressure swings and PTP per breath (Table 2), PTP/min was lower 282 than during hyperpnea (Table 3). Likewise, increases in VO₂ and in cardiac output were less 283 during loaded breathing in comparison to hyperpnea (Table 3). The lower "systemic" oxygen 284 requirements (i.e., VO₂ and a-vO₂ diff, Table 3) during loaded breathing were accompanied by a smaller increase in respiratory muscle blood flow and oxygen delivery in comparison to 285 286 hyperpnea (Table 4). These responses observed during loading breathing resulted in a mismatch between SCA and 7thICM muscles oxygen delivery and utilization (Table 4), resulting in greater 287 288 increases in muscle oxygen extraction (i.e., HHB) and lower StiO₂ as compared to hyperpnea 289 (Table 4). Higher intramuscular tensions imposed during loading breathing, might have 290 contributed to limiting increased in muscle blood flow and oxygen delivery as compared to 291 hyperpnea (Table 4).(40) The evidence of high intramuscular pressures during loading breathing 292 is supported by the results demonstrating that mean arterial pressure did not statistically differ 293 between the two conditions (Table 3) even that during loading breathing central hemodynamic 294 responses were significantly lower compared to hyperpnea (Table 3). Indeed, studies have shown 295 that increases in intramuscular pressure during dynamic exercise can reflexively increase mean 296 arterial blood pressure (via the activation of the mechanoreceptor-mediated reflex within the 297 skeletal muscle), the latter increases can be maintained throughout the exercise period. (41)

298 General considerations. Collectively, these results seem to support the notion that 299 additional inspiratory pressures generated during loaded breathing are mainly a consequence of 300 increased loading and activation of SCA, SCM and both intercostal muscles. The behavior of the 301 "respiratory effort-recruitment" ratio,(<u>42</u>) i.e., the "matching" between respiratory muscle effort 302 (e.g., Pes, %max) and the recruitment of different inspiratory muscles (EMG, %max), is 303 noteworthy. While during resting breathing a higher ratio indicates a "predominantly diaphragm 304 contribution to breathing", with increasing load (i.e., hyperpnea and loaded breathing), the ratio 305 becomes similar between diaphragm and SCA, SCM and both intercostal muscles, thereby 306 indicating that SCA, SCM and both intercostal muscles contribution to breathing becomes 307 equally important as that of the diaphragm (supplemental material Figure E1).

308 The observed acute increases in load and work being performed by the inspiratory 309 muscles during both tasks (Table 2) are known to be important determinants of muscle 310 improvements after exercise programs.(43) Furthermore, according to the specificity and 311 overload principles of training, (43) in response to a low load (i.e., pressures), high repetition 312 (i.e., breathing frequency and duration) and high exercise-volume (i.e., PTP cmH2O/s/min) 313 (Table 2) stimulus as hyperpnea, an endurance benefit would be expected. While after loaded 314 breathing, improvements in strength would be anticipated as consequence of the high load (i.e., 315 pressures), low repetition (i.e., breathing frequency and duration) and high contraction-volume 316 (PTP cmH2O/s/b) stimulus imposed by this regimen (Table 2). Noteworthy the additional 317 recruitment of only SCA, SCM and both intercostal muscles (Figure 1) as the strategy to 318 overcome the load imposed during loaded breathing in comparison to hyperpnea (Table 2) was 319 accompanied by an increased metabolic burden (Table 3 It is therefore likely these inspiratory 320 muscles will mostly benefit from this additional stimulus (i.e., increased load). $(\underline{43})$ It has 321 previously been observed that a period of high intensity inspiratory muscle strength training 322 resulted in increases in specific hypertrophy of intercostal muscle fibers.(44)

323 Implications. The differences in physiological responses evoked by these different types 324 (and intensities) of respiratory muscles loading support observations that had previously been 325 done in clinical practice. It has long been assumed that while exercise hyperpnea constitutes a 326 training load to the respiratory muscles a larger stimulus might be applied with specific 327 respiratory muscle training.(45) This is supported by data from RCTs showing that adding 328 specific inspiratory muscle strength training resulted in larger improvements in respiratory 329 muscle function (strength and endurance), exercise capacity (cycling endurance time) and 330 reduction in dyspnea(1) than standard endurance exercise training alone.(2, 4) The stimulus 331 imposed during loaded breathing in this study (resembling a specific type of inspiratory muscle 332 strength training) seems to be a good complimentary training stimulus for the respiratory 333 muscles in addition to whole body exercise training. (46) Based on our data it provides a different 334 additional load to the respiratory muscles in comparison to exercise hyperpnea. In contrast with 335 earlier hypotheses this additional load did not result in stimulating the diaphragm in exceeding a 336 plateau in motor unit recruitment that is typically observed early during exercise hyperpnea, (47)337 but by further stimulating SCA, SCM and intercostal muscle recruitment above levels observed 338 during exercise breathing. Nevertheless, it is important to stress that the hyperpnea used herein 339 resembles the load imposed to the respiratory system during exercise hyperpnea (i.e., 70% MVV 340 for several minutes) and not necessarily loads imposed during specific respiratory muscle 341 endurance training (i.e., 50 - 70% MVV for 15-30 minutes).(4) Whether higher volumes and 342 longer durations of specific respiratory muscle endurance training might also lead to differential 343 activation and recruitment patterns of respiratory muscles in comparison to the relatively short 344 exercise hyperpnea stimulus provided in our study remains to be investigated.

345 Strengths, limitations and technical considerations. The multitude of variables 346 simultaneously collected is a strength of the study. It allows the concurrent investigation of the 347 behavior of respiratory muscles activation, pressure generation and metabolism under the same 348 stimulus. Unfortunately, however, assessments of blood flow and oxygen requirements of the 349 diaphragm could not be performed due to methodological and safety issues. A limitation of our 350 study is the small sample size due to the complexity and the invasiveness of its methods and the 351 fact that not all subjects were able or willing to undergo all experimental procedures. However, 352 the sample was powered sufficiently (see *Data analysis* in the supplemental material for more 353 details) to detect differences in a wide variety of physiological markers. Moreover, while 354 reproducing the ventilatory pattern of exercise hyperpnea (i.e., breathing frequency, tidal volume 355 and ventilation), there were also no statistically significant differences between the expiratory 356 gastric pressures and expiratory ABD activation that were generated during cycling exercise in comparison to hyperpnea (Pga cycling 20 ± 3 vs Pga hyperpnea 25 ± 5 , cmH₂O; *P*=0.3; EMGabd 357 358 cycling 23 ± 4 vs EMGabd hyperpnea 33 ± 4 , %max; P=0.10, respectively). Thus, providing an 359 adequate reproducibility between exercise hyperpnea and the hyperpnea task used in our study. 360 Arterial oxygen content, a-VO₂ diff and systemic oxygen extraction were estimated using 361 continuous SpO_2 measurements at the expense of acceptable reduced accuracy in the hypoxemic 362 patients compared with invasive arterial blood sampling. In addition, it is known that the EMG signal from the costal diaphragm can generate noise on the activation of the 7thICM we measured 363 herein. However, the different pattern of diaphragm and 7thICM activation between loaded 364 breathing and hyperpnea suggested that this was not the case in our data. Nevertheless, it is 365 366 possible that the EMG signal measured at these muscles as well as at SCA and SCM could be, at 367 least in part, contaminated from nearby activity due to the use of superficial electrodes. In our

368 patients, the contribution of diaphragmatic blood flow to the overall NIRS signal on the 7th 369 intercostal space is probably limited considering that adipose tissue thickness (fat + skin layer) 370 (measurements were performed using a Harpenden skinfold caliper) indicated a mean value of 371 8.2 ± 3.7 mm. Therefore, the maximum penetration depth of NIRS light to the muscle tissue was 372 reduced to approximately 12 mm. Taking into account the substantial distance between the 373 sampling point of NIRS on the skin and the diaphragmatic appositional area compared with the 374 shorter distance to the intercostals we believe that perfusion and oxygenation measures in our 375 study at this site reflected mostly the external and internal intercostal muscles.

376

CONCLUSION

During loaded breathing there was greater respiratory muscle effort compare to hyperpnea which ensued larger ribcage and neck muscle activation during inspirations. This response reflects the additional burden imposed on these muscles by a strengthening stimulus in comparison to an endurance loading stimulus. In addition, the decrease in ribcage and neck muscle tissue oxygen saturation during loading breathing compared to hyperpnea might indicates a mismatch between inspiratory muscle oxygen delivery and utilization .

383

384

385

386 Acknowledgments

387 This research project was supported by Research Foundation Flanders (FWO):
388 G0A4516N - ZKC9570 - C22/15/035. Antenor Rodrigues is supported by the Coordination for
389 the Improvement of Higher Education Personnel (CAPES), Brazil (88881.188754/2018-01). Dr.

Zafeiris Louvaris is recipient of an ERS fellowship (LTRF 2016-6686) and a postdoctoral fellow
of Research Foundation Flanders (FWO - 12U5618N).

392

393 Conflict of interest

The authors have no conflict of interest to disclose. The results presented herein do not constitute endorsement by ACSM and are presented clearly, honestly, and without fabrication, falsification, or inappropriate data manipulation.

397 REFERENCES

Charususin N, Gosselink R, Decramer M, et al. Randomised controlled trial of adjunctive
 inspiratory muscle training for patients with COPD. Thorax. 2018;73(10):942-50. doi:
 10.1136/thoraxjnl-2017-211417. PubMed PMID: 29914940.

401 2. Mador MJ, Deniz O, Aggarwal A, Shaffer M, Kufel TJ, Spengler CM. Effect of
402 respiratory muscle endurance training in patients with COPD undergoing pulmonary
403 rehabilitation. Chest. 2005;128(3):1216-24. doi: 10.1378/chest.128.3.1216. PubMed PMID:
404 16162709.

3. Illi SK, Held U, Frank I, Spengler CM. Effect of respiratory muscle training on exercise
performance in healthy individuals: a systematic review and meta-analysis. Sports Med.
2012;42(8):707-24. Epub 2012/07/07. doi: 10.2165/11631670-00000000-00000. PubMed
PMID: 22765281.

409 4. Gosselink R, De Vos J, van den Heuvel SP, Segers J, Decramer M, Kwakkel G. Impact
410 of inspiratory muscle training in patients with COPD: what is the evidence? The European
411 respiratory journal. 2011;37(2):416-25. doi: 10.1183/09031936.00031810. PubMed PMID:
412 21282809.

413 5. Beaumont M, Forget P, Couturaud F, Reychler G. Effects of inspiratory muscle training
414 in COPD patients: A systematic review and meta-analysis. The clinical respiratory journal.
415 2018;12(7):2178-88. doi: 10.1111/crj.12905. PubMed PMID: 29665262.

416 6. Richardson RS, Sheldon J, Poole DC, Hopkins SR, Ries AL, Wagner PD. Evidence of
417 skeletal muscle metabolic reserve during whole body exercise in patients with chronic
418 obstructive pulmonary disease. American journal of respiratory and critical care medicine.
419 1999;159(3):881-5. doi: 10.1164/ajrccm.159.3.9803049. PubMed PMID: 10051266.

420 7. Shoemaker JK, Hodge L, Hughson RL. Cardiorespiratory kinetics and femoral artery
421 blood velocity during dynamic knee extension exercise. Journal of applied physiology.
422 1994;77(6):2625-32. doi: 10.1152/jappl.1994.77.6.2625. PubMed PMID: 7896601.

423 8. Mahler DA, Weinberg DH, Wells CK, Feinstein AR. The measurement of dyspnea.
424 Contents, interobserver agreement, and physiologic correlates of two new clinical indexes.
425 Chest. 1984;85(6):751-8. PubMed PMID: 6723384.

426 9. Singh D, Agusti A, Anzueto A, et al. Global Strategy for the Diagnosis, Management,
427 and Prevention of Chronic Obstructive Lung Disease: The GOLD Science Committee Report
428 2019. The European respiratory journal. 2019. doi: 10.1183/13993003.00164-2019. PubMed
429 PMID: 30846476.

Miller MR, Hankinson J, Brusasco V, et al. Standardisation of spirometry. The European
respiratory journal. 2005;26(2):319-38. doi: 10.1183/09031936.05.00034805. PubMed PMID:
16055882.

433 11. Wanger J, Clausen JL, Coates A, et al. Standardisation of the measurement of lung
434 volumes. The European respiratory journal. 2005;26(3):511-22. doi:
435 10.1183/09031936.05.00035005. PubMed PMID: 16135736.

436 12. Black LF, Hyatt RE. Maximal respiratory pressures: normal values and relationship to
437 age and sex. The American review of respiratory disease. 1969;99(5):696-702. doi:
438 10.1164/arrd.1969.99.5.696. PubMed PMID: 5772056.

Laveneziana P, Albuquerque A, Aliverti A, et al. ERS Statement on Respiratory Muscle
Testing at Rest and during Exercise. The European respiratory journal. 2019. doi:
10.1183/13993003.01214-2018. PubMed PMID: 30956204.

442 14. Puente-Maestu L, Palange P, Casaburi R, et al. Use of exercise testing in the evaluation
443 of interventional efficacy: an official ERS statement. The European respiratory journal.
444 2016;47(2):429-60. doi: 10.1183/13993003.00745-2015. PubMed PMID: 26797036.

Vogiatzis I, Athanasopoulos D, Habazettl H, et al. Intercostal muscle blood flow
limitation during exercise in chronic obstructive pulmonary disease. American journal of
respiratory and critical care medicine. 2010;182(9):1105-13. doi: 10.1164/rccm.201002-0172OC.
PubMed PMID: 20622032.

Langer D, Charususin N, Jacome C, et al. Efficacy of a Novel Method for Inspiratory
Muscle Training in People With Chronic Obstructive Pulmonary Disease. Physical therapy.
2015;95(9):1264-73. doi: 10.2522/ptj.20140245. PubMed PMID: 25858974.

452 17. Borg GA. Psychophysical bases of perceived exertion. Medicine and science in sports
453 and exercise. 1982;14(5):377-81. Epub 1982/01/01. PubMed PMID: 7154893.

Louvaris Z, Habazettl H, Wagner H, Zakynthinos S, Wagner P, Vogiatzis I. Near-infrared
spectroscopy using indocyanine green dye for minimally invasive measurement of respiratory
and leg muscle blood flow in patients with COPD. Journal of applied physiology.
2018;125(3):947-59. doi: 10.1152/japplphysiol.00959.2017. PubMed PMID: 29927736.

Piiper J, Haab P. Oxygen supply and uptake in tissue models with unequal distribution of
blood flow and shunt. Respiration physiology. 1991;84(2):261-71. PubMed PMID: 1876763.

Langer D, Ciavaglia C, Faisal A, et al. Inspiratory muscle training reduces diaphragm
activation and dyspnea during exercise in COPD. Journal of applied physiology.
2018;125(2):381-92. doi: 10.1152/japplphysiol.01078.2017. PubMed PMID: 29543134.

463 21. Hawkes EZ, Nowicky AV, McConnell AK. Diaphragm and intercostal surface EMG and
464 muscle performance after acute inspiratory muscle loading. Respiratory physiology &
465 neurobiology. 2007;155(3):213-9. doi: 10.1016/j.resp.2006.06.002. PubMed PMID: 16846758.

466 22. American Thoracic Society/European Respiratory S. ATS/ERS Statement on respiratory
467 muscle testing. American journal of respiratory and critical care medicine. 2002;166(4):518-624.
468 doi: 10.1164/rccm.166.4.518. PubMed PMID: 12186831.

469 23. Faisal A, Alghamdi BJ, Ciavaglia CE, et al. Common Mechanisms of Dyspnea in
470 Chronic Interstitial and Obstructive Lung Disorders. American journal of respiratory and critical
471 care medicine. 2016;193(3):299-309. Epub 2015/09/26. doi: 10.1164/rccm.201504-0841OC.
472 PubMed PMID: 26407036.

473 24. Louvaris Z, Spetsioti S, Andrianopoulos V, et al. Cardiac output measurement during
474 exercise in COPD: A comparison of dye dilution and impedance cardiography. The clinical
475 respiratory journal. 2019;13(4):222-31. doi: 10.1111/crj.13002. PubMed PMID: 30724023.

476 25. Borghi-Silva A, Oliveira CC, Carrascosa C, et al. Respiratory muscle unloading improves
477 leg muscle oxygenation during exercise in patients with COPD. Thorax. 2008;63(10):910-5. doi:
478 10.1136/thx.2007.090167. PubMed PMID: 18492743.

479 26. Grassi B, Quaresima V. Near-infrared spectroscopy and skeletal muscle oxidative
480 function in vivo in health and disease: a review from an exercise physiology perspective. Journal

481 of biomedical optics. 2016;21(9):091313. doi: 10.1117/1.JBO.21.9.091313. PubMed PMID:
482 27443955.

483 27. Boushel R, Langberg H, Olesen J, Gonzales-Alonzo J, Bulow J, Kjaer M. Monitoring
484 tissue oxygen availability with near infrared spectroscopy (NIRS) in health and disease.
485 Scandinavian journal of medicine & science in sports. 2001;11(4):213-22. PubMed PMID:
486 11476426.

Washino S, Kanehisa H, Yoshitake Y. Neck inspiratory muscle activation patterns during
well-controlled inspiration. European journal of applied physiology. 2017;117(10):2085-97. doi:
10.1007/s00421-017-3699-5. PubMed PMID: 28823081.

29. Campbell EJ. The role of the scalene and sternomastoid muscles in breathing in normal
subjects; an electromyographic study. Journal of anatomy. 1955;89(3):378-86. PubMed PMID:
13251968; PubMed Central PMCID: PMC1244766.

30. Raper AJ, Thompson WT, Jr., Shapiro W, Patterson JL, Jr. Scalene and sternomastoid
muscle function. Journal of applied physiology. 1966;21(2):497-502. doi:
10.1152/jappl.1966.21.2.497. PubMed PMID: 5934453.

496 31. De Troyer A, Boriek AM. Mechanics of the respiratory muscles. Comprehensive
497 Physiology. 2011;1(3):1273-300. doi: 10.1002/cphy.c100009. PubMed PMID: 23733642.

498 32. De Troyer A, Wilson TA. Effect of acute inflation on the mechanics of the inspiratory
499 muscles. Journal of applied physiology. 2009;107(1):315-23. doi:
500 10.1152/japplphysiol.91472.2008. PubMed PMID: 19265064.

501 33. De Troyer A, Leduc D, Cappello M, Mine B, Gevenois PA, Wilson TA. Mechanisms of 502 the inspiratory action of the diaphragm during isolated contraction. Journal of applied

503 physiology. 2009;107(6):1736-42. doi: 10.1152/japplphysiol.00753.2009. PubMed PMID:
504 19797686.

505 34. Decramer M, De Troyer A. Respiratory changes in parasternal intercostal length. Journal
506 of applied physiology: respiratory, environmental and exercise physiology. 1984;57(4):1254-60.
507 doi: 10.1152/jappl.1984.57.4.1254. PubMed PMID: 6501034.

- 508 35. Farkas GA, Rochester DF. Contractile characteristics and operating lengths of canine
 509 neck inspiratory muscles. Journal of applied physiology. 1986;61(1):220-6. doi:
 510 10.1152/jappl.1986.61.1.220. PubMed PMID: 3733607.
- 511 36. Farkas GA. Mechanical properties of respiratory muscles in primates. Respiration
 512 physiology. 1991;86(1):41-50. PubMed PMID: 1759052.
- 513 37. Roussos C, Macklem PT. The respiratory muscles. The New England journal of
 514 medicine. 1982;307(13):786-97. doi: 10.1056/NEJM198209233071304. PubMed PMID:
 515 7050712.
- 38. Rochester DF, Pradel-Guena M. Measurement of diaphragmatic blood flow in dogs from
 xenon 133 clearance. Journal of applied physiology. 1973;34(1):68-74. doi:
 10.1152/jappl.1973.34.1.68. PubMed PMID: 4572509.

39. Andersen P, Saltin B. Maximal perfusion of skeletal muscle in man. The Journal of
physiology. 1985;366:233-49. PubMed PMID: 4057091; PubMed Central PMCID:
PMC1193029.

40. Bellemare F, Wight D, Lavigne CM, Grassino A. Effect of tension and timing of
contraction on the blood flow of the diaphragm. Journal of applied physiology: respiratory,
environmental and exercise physiology. 1983;54(6):1597-606. doi:
10.1152/jappl.1983.54.6.1597. PubMed PMID: 6874482.

526 41. Gallagher KM, Fadel PJ, Smith SA, et al. Increases in intramuscular pressure raise
527 arterial blood pressure during dynamic exercise. Journal of applied physiology. 2001;91(5):2351528 8. doi: 10.1152/jappl.2001.91.5.2351. PubMed PMID: 11641380.

529 42. Ertl P, Kruse A, Tilp M. Detecting fatigue thresholds from electromyographic signals: A

530 systematic review on approaches and methodologies. J Electromyogr Kinesiol. 2016;30:216-30.

531 Epub 2016/08/17. doi: 10.1016/j.jelekin.2016.08.002. PubMed PMID: 27529668.

532 43. Kraemer WJ, Ratamess NA. Fundamentals of resistance training: progression and
533 exercise prescription. Medicine and science in sports and exercise. 2004;36(4):674-88. PubMed
534 PMID: 15064596.

44. Ramirez-Sarmiento A, Orozco-Levi M, Guell R, et al. Inspiratory muscle training in
patients with chronic obstructive pulmonary disease: structural adaptation and physiologic
outcomes. American journal of respiratory and critical care medicine. 2002;166(11):1491-7. doi:
10.1164/rccm.200202-075OC. PubMed PMID: 12406842.

539 45. Decramer M. Response of the respiratory muscles to rehabilitation in COPD. Journal of
540 applied physiology. 2009;107(3):971-6. Epub 2009/04/04. doi:
541 10.1152/japplphysiol.91459.2008. PubMed PMID: 19342436.

542 46. Polkey MI, Ambrosino N. Inspiratory muscle training in COPD: can data finally beat
543 emotion? Thorax. 2018;73(10):900-1. doi: 10.1136/thoraxjnl-2018-212070. PubMed PMID:
544 29945956.

545 47. Sinderby C, Spahija J, Beck J, et al. Diaphragm activation during exercise in chronic
546 obstructive pulmonary disease. American journal of respiratory and critical care medicine.
547 2001;163(7):1637-41. Epub 2001/06/13. doi: 10.1164/ajrccm.163.7.2007033. PubMed PMID:
548 11401887.

549

550 Supplemental digital content 1. doc

551 Supplemental digital content 2. tif

552

Figure 1. Comparisons between the EMG activation among the different tasks. EMGdi, %max: relative diaphragmatic activation; EMGsca, %max: relative scalenes activation; EMGscm, %max: relative sternocleidomastoid activation; EMGicm, %max: relative parasternal intercostal activation; EMG 7th icm, %max: relative 7th intercostal activation. Boxplots shows median at central line, first and third quantiles for lower and upper box's limits, respectively, and minimum and maximum values for lower and upper limits. Dots are single patients' values. Dots outside the limits are outliers' values. **P* <0.05; NS; *P* >0.05. EMGdi: n= 10; sEMG n= 16.

560

561 Figure 2. Relationship between work og breathing (WOB) and pressure-time product (PTP) with 562 oxygen consumption (VO₂; a and d, respectively) and cardiac output (CO; b and e, respectively); and between systemic oxygen delivery (O₂ del) and oxygen consumption (VO₂) and vascular 563 conductance (Vasc. cond.; c and f, respectively). r: Pearson coefficient correlations; R²: Adjusted 564 565 R squared (univariate linear regression); NA: not applicable; NS: P > 0.05 (non-significant). Lines are the best-fitting line and shadow areas are 95% confidence interval. Circles: rest; triangles: 566 loading. 567 normocapnic hyperpnea; cross: tapered flow resistive

Table 1. Subjects characteristics, pulmonary function and peak exercise and inspiratory muscle

569 capacity data

	n: 16
Demographics/Anthropometrics	
Sex, male/female	9 /7
Age, yrs	65±13
BMI, kg/m ²	27 ± 1.6
Pulmonary function	
FEV ₁ ,L	1.44±0.15
FEV1, %pred	60 ± 6
FVC,L	3.23±0.22
FVC, %pred	99±8
FEV ₁ /FVC, %	44±3
MVV, L/min	52±5
MVV, %pred	65±8
TLC,L	6.4 ± 0.46
TLC, %pred	118±5
RV,L	3.45±0.33
RV,%pred	155 ± 12
RV/TLC, %	54±2
VC,L	2.9 ± 0.2
TLCO, mmol/min/kpa	4.3±0.4
TLCO, %pred	56±4

Peak exercise data and inspiratory muscle capacity	
Wpeak, W	81±7
Wpeak, %max	71±5
VO2, peak, L/min	1.371 ± 0.116
VO ₂ , peak, %max	87±8
COpeak, L/min	12.0±0.5
MIP, cmH ₂ O	74±4
MIP, %pred	82±5
MIP <lln, n(%)<="" th=""><th>9 (56)</th></lln,>	9 (56)
Hb, g/dl	14.5 ± 0.3

Data are mean ± SE or n (%). FEV₁: forced expiratory volume in the first second; FVC: forced vital capacity; MVV: maximum voluntary ventilation; TLC: total lung capacity; RV: residual
volume; TLCO: transfer factor for carbon monoxide; MIP: maximal inspiratory pressure; Insp.
mm. weakness: maximum inspiratory pressure bellow the lower limit of normality; Wpeak; peak
exercise capacity; VO2peak: peak oxygen consumption; COpeak; peak cardiac output; LLN:
lower limit of normality.

577	Table 2. Respiratory pressures and	work of breathing and breathing pattern data during hyperphy	a and loaded breathing
••••		······································	

					Mean diff (95% Cl	0
	Rest	Hyperpnea	Loaded breathing	Hyperpnea - Rest	Loaded breathing - Rest	Loaded breathing - Hyperpnea
Respiratory pressures and work	ofbreathing	g(n=10)				
Pes, cmH ₂ O	-9±1	-15±1	-35±2	-6(-112)*	-26(-3021)*	-19(-2415)*
Pes, %max	14±2	23±2	54±5	10(-2-21)*	40(27-51)*	30(18-41)*
inspPga, cmH2O	10±2	12±2	15±4	1(-9-12)	5(-5-15)	3(-7-13)
expPga, cmH2O	10±1	21±4	21±4	10(-1-21)	11(0-22)	1(-10-12)
inspPga, %max	$21\pm$	22±4	26±6	1(-15-17)	5(-11-21)	4(-12-20)
Pdi, cmH2O	19±1	27±2	50±4	7(172)*	30(40-20)*	22(32-12)*
Pdi, %max	21±2	28±1	53±4	7(-2-16)*	32(22-41)*	24(15-34)*
Pes WOB, L/cmH2O	6±1	16±2	113±16	10(-22-42)*	108(75-140)*	97(65-130)*
inspPga WOB,, L/cmH ₂ O	3±1	9±2	33±5	6(-6–17)	30(18-41)*	24(13-36)*
Pdi WOB, L/cmH2O	7±2	14±4	104±15	7(-25-39)*	97(65-128)*	90(58-122)*
PTP Pes, cmH2O/s//b	4 ± 0	6±0	8±1	2(0-4)*	4(-2-4)*	2(0-4)*
inspPTP Pga, cmH2O/s/b	4 ± 1	4 ± 1	3±1	0(-3-3)	-1(4-2)	-1(4-2)

PTP Pdi, cmH2O/s/b	8 ± 1	10 ± 1	11±1	2(0-6)	3(0-6)	1(-2-4)
Pes WOB, L/cmH2O/min	95±11	495±62	624±71	400(209-591)*	529 (337-720)*	129(-62-320)
inspPga WOB, L/cmH2O/min	52±7	276±58	198±38	224(83-365)*	147(6-288)*	-77(-218-64)
Pdi WOB, L/cmH2O/min	109±16	430±107	567±66	321 (64-578)*	458 (200-715)*	136(-120-394)
PTP Pes, cmH2O/s/min	71 ± 12	184±16	49±9	112(69–157)*	-21 (-66-22)	-135(-179–91)*
inspPTP Pga, cmH2O/s/min	84±18	142±28	21±7	58(-12-127)*	-62(-132-7)*	-120(-19051)*
PTPPdi, cmH2O/s/min	154±26	325±35	68±13	171 (79–262)*	-85(-177-6)*	-256(-3481654.73)*

Breathing pattern (n=16)

	Rest	hyperpnea	loaded breathing	hyperpnea - Rest	loaded breathing - Rest	loaded breathing - hyperpnea
VE,L	13±1	38±3	12±1	25(18-32)*	-1(-8-5)	-26(-3319)*
Insp. vol., L	0.74±0.06	1.17 ± 0.11	1.9±0.21	0.43(-0.05-0.91)*	1.16(0.68-1.64)*	0.73 (0.25-1.21)*
Bf, b/min	20±1	34±1	$7{\pm}1$	14(10-18)*	-13(-178)*	-27(-3122)*
Insp. peak flow, L/sec	0.91 ± 0.05	2.47 ± 0.18	2.23±0.2	1.56(1.03-2.09)*	1.32(0.80-1.85)*	-0.24(-0.77-0.28)
Insp. time, s	1.27±0.1	0.67±0.04	2.26±0.22	-0.60(-1.09-0.11)*	0.99(0.50-1.47)*	1.58(1.10-2.07)*
Ti/Ttot, %	38±1	37±1	24±2	-2(-6-4)	-14(-198)*	-12(-187)*

578 Data are mean ± SE or mean difference (95% confidence interval). Ti/Ttot: duty cycle of respiration; Bf: breathing frequency; Pes:

579 Esophageal pressure; Pdi: Transdiaphragmatic pressure; WOB: work of breathing; PTP: Pressure Time Product. * *P* <0.05.

					Mean diff (95%)	CI)
	Rest	hyperpnea	loaded breathing	hyperpnea - Rest	loaded breathing - Rest	loaded breathing - hyperpnea
HR, bpm	76±3	90±4	89±4	14(2-26)*	13(1-25)*	-1(-13–11)
SV,ml	70±4	84±6	73±4	15(-1-31)*	4(-13-20)	-11(-27–5)*
CO,L/min	5.2±0.3	7.5±0.5	6.5±0.4	2.3(0.9–3.7)*	1.1 (0.2–2.6)*	-1.1 (4–0.3)*
CO, %max	44±3	62±4	54±4	19(6-32)	10(-3-23)	-8(-21-5)
VO2, ml/min	283±20	625±42	443±34	342 (229-454)*	161 (46–275)*	-181 (-29667)*
VO2, %max	25±4	54±7	39±5	29(10-48)*	13(-6-32)*	-16(-34–3)*
VCO ₂ , ml/min	224±14	412±69	409±32	188(35-341)*	185 (29-340)*	-4(-159-151)
CaO2mlO2/100ml	18.9±0.5	19.2±0.5	19.2±0.4	0.3(-1.2–1.9)	0.3(-1.2–1.9)	0(-1.5–1.6)
O2 delivery, LO2/min	0.98±0.05	1.42 ± 0.1	1.23±0.07	0.44 (0.17–0.71)*	0.25(-0.01-0.52)*	-0.18(-0.45-0.08)*
O ₂ extraction, %	29±2	46±4	38±3	16(6-26*	8(-2-19)*	-8(-18-3)*
a-vO2 difference , mlO2/100ml	5.6±03	8.73±0.75	7.3±0.7	3.2(1.1-5.3)*	1.7(-0.4-3.9)*	-1.4(-3.5-0.7)*
SVC,ml/min/mmHg	56±3	74±5	63±4	18(5-32)*	7(-7-21)*	-11(-26-3)*
SpO ₂ ,%	94±1	95±1	94±1	2(-1-4)	0(-2-3)	-1(-4-1)

SBP,mmHg	120±3	139±6	133±5	19(4-33)*	13(-3-28)*	-5.9(-22-10)
DBP,mmHg	80±2	88±2	90±4	8(-1-17)	10(1-20)*	2(-8-12)
MAP	93±2	105±3	104±4	12(2-22)*	11(1-21)*	-1 (-11-10)

Data are mean \pm SE or mean difference (95% confidence interval). HR: heart rate; SV: stroke volume; CO: cardiac output; VO₂: oxygen consumption; VCO₂: carbon dioxide production; CaO₂; arterial oxygen content; a-vO₂ difference: arterio-venous oxygen difference; SVC: systemic vascular conductance SpO₂: peripheral oxygen saturation; SBP: systolic blood pressure; DBP: diastolic blood pressure; Vasc. Cond.: systemic vascular conductance. **P* <0.05.

586

588	Table 4. Respiratory muscles perfusion and oxygenation responses during hyperpna and loaded
589	breathing

			Mean diff (95% CI)
	hyperpnea	loaded breathing	loaded breathing - hyperpnea
Respiratory muscle perf	iusion, n= 13		
ASCA BFI, nmol/L	4.67±1.3	2.81 ± 1.17	-1.86(-3.20.5)*
Δ7 th IC BFI, nmol/L	0.76±0.2	0.5 ± 0.2	0.27 (-0.78 - 0.2)
AABD BFI, nmol/L	1.2 ± 0.5	0.4 ± 0.3	-0.8(-1.7-0.2)
Respiratory muscle O2d	lelivery		
ΔSCA O2del, au	90±24	54±22	-36(-1162)*
Δ7 th IC O2del, au	14 ± 4	10±5	-5 (414)
AABD O2del, au	23 ± 10	8±6	-14(3-33)
Respiratory muscle oxy	gen saturation	,n=15	
ΔSCA StiO2, %	1.25 ± 0.9	-2.84±1.27	-4.1 (-62.1)*
Δ7 th IC StiO2, %	1.5 ± 0.71	-1.52 ± 0.86	-3(-4.91.3)*
AABD StiO2, %	1.00 ± 1.00	-0.40 ± 1.52	-1.38 (-3.6 - 0.9)
Respiratory muscle oxy	gen extraction	,n=15	
ΔSCA [HHb], µmol/L	2.94±1.33	7.68 ± 2.08	4.73 (1.88 - 7.58)*
Δ7 th IC [HHb], µmol/L	0.42 ± 0.61	1.9 ± 0.87	1.48 (-0.05 - 3)
ΔABD [HHb], μmol/L	-1.67 ± 0.86	0.03 ± 1.1	1.70(-0.82 - 3.48)

590 Data are mean \pm SE or mean difference (95% confidence interval). Δ : changes from rest; SCA: 591 Scalenes; 7th IC: 7th Intercostal; ABD: Rectus Abdominis; [HHb]: deoxyhemoglobin 592 concentration; StiO₂: Tissue oxygen saturation index; BFI: blood flow index. **P* <0.05.



597 Figure 2.

