

Manuscript version: Author's Accepted Manuscript

The version presented in WRAP is the author's accepted manuscript and may differ from the published version or Version of Record.

Persistent WRAP URL:

<http://wrap.warwick.ac.uk/135295>

How to cite:

Please refer to published version for the most recent bibliographic citation information. If a published version is known of, the repository item page linked to above, will contain details on accessing it.

Copyright and reuse:

The Warwick Research Archive Portal (WRAP) makes this work by researchers of the University of Warwick available open access under the following conditions.

Copyright © and all moral rights to the version of the paper presented here belong to the individual author(s) and/or other copyright owners. To the extent reasonable and practicable the material made available in WRAP has been checked for eligibility before being made available.

Copies of full items can be used for personal research or study, educational, or not-for-profit purposes without prior permission or charge. Provided that the authors, title and full bibliographic details are credited, a hyperlink and/or URL is given for the original metadata page and the content is not changed in any way.

Publisher's statement:

Please refer to the repository item page, publisher's statement section, for further information.

For more information, please contact the WRAP Team at: wrap@warwick.ac.uk.

Structural disorganization and chain aggregation of high-amylose starch in different chloride salt solutions

Ying Li^{†,‡,#}, Peng Liu^{*,†,‡,#}, Cong Ma^{†,‡}, Na Zhang^{†,‡}, Xiaoqin Shang^{†,‡}, Liming Wang^{†,‡}, Fengwei Xie^{*,§,¶}

[†]School of Chemistry and Chemical Engineering, and [‡]Fine Chemical Research Institute, Guangzhou

University, 230 Wai Huan Xi Road, Guangzhou Higher Education Mega Center, Guangzhou, Guangdong

510006, China

[§]International Institute for Nanocomposites Manufacturing (IINM), WMG, University of Warwick, Coventry

CV4 7AL, United Kingdom

[¶]School of Chemical Engineering, The University of Queensland, Brisbane, Qld 4072, Australia

Corresponding Authors

* Email address: liu_peng@gzhu.edu.cn (P. Liu)

* Email address: f.xie@uq.edu.au; fwhsieh@gmail.com (F. Xie)

KEYWORDS: *High-amylose starch; Biopolymer; Metal chloride salt; Structural disorganization; Dissolution; Starch-metal ion coordinated complex; Nanoparticles; Rheology*

ABSTRACT: As high-amylose starch (HAS) has a higher content of linearly structured chains than other types of starch, it is more scientifically interesting to realize enhanced properties or new functions

1
2
3
4 for food and materials applications. However, the full dissolution of the compact granule structure of
5
6 HAS is challenging under moderate conditions, which limits its applications. Here, we have revealed
7
8 that the granule structure of HAS can be easily destructed by certain concentrations of acidic ZnCl_2 ,
9
10 neutral MgCl_2 and alkaline CaCl_2 solutions (43 wt%, 34 wt%, and 31 wt%, respectively) at a moderate
11
12 temperature (under 50 °C). The ZnCl_2 and CaCl_2 solutions resulted in complete dissolution of HAS
13
14 granules, whereas small amounts of HAS granule remnants still existed in the MgCl_2 solution. The
15
16 regenerated starch from the CaCl_2 solution was completely amorphous, that from the ZnCl_2 solution
17
18 only presented a weak peak at 17°, and that from the MgCl_2 solution contained V-type crystallites. No
19
20 new reflections were found on the FTIR spectra indicating all these three chloride solutions can be
21
22 considered as a non-derivatizing solvent for starch. In all the three cases, nanoparticles were formed
23
24 in the regenerated starch, which could be due to the aggregation of starch chains or their complexation
25
26 with the metal cation. In addition, their water absorption ratio was 1.5 to 3 times that of the control
27
28 (treated in water).
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

INTRODUCTION

Starch is a natural polymer formed by the photosynthesis in plants and is one of the major forms for plants to store energy. It is an abundant, cheap, renewable and environmentally-friendly resource.¹

Normal or wild-type starches consist of two types of glucose polymers, the predominately-linear amylose with rare branches, and the highly-branched amylopectin.² Amylose has a molecular mass of about 10^6 Daltons with long branches. Amylopectin contains a large number of short branches and is one of the largest biopolymers (about 10^8 Daltons).³

Starch with a higher content of amylose, compared to typical wild-type lines, can be termed as high-amylose starch (HAS). HAS can be biosynthesized in mutant cereal grains such as wheat, maize, rice, and barley and have some special properties such as heat resistance and digestion resistibility.² The granule structure of HAS has been extensively studied. Its inner region is composed of mainly loosely-packed amylopectin growth rings with semicrystalline lamellae,⁴ while its compact periphery is supposed to be composed of entangled amylose chains.⁵

HAS has been used in many applications such as nutritional food, food processing, drug release, and biodegradable materials.⁶ Due to the suitable molecular weight, linear molecular structure and densely populated hydroxyl groups, amylose has high chemical reactivity, which can be used to prepare modified starch,⁷ starch-ion inclusion complexes⁸ and starch-based materials.⁹ However, the compact granule structure and the high gelatinization temperature of HAS limit its applications. As a result, how to destroy HAS granules and release starch chains is both scientifically and practically interesting.

Some solvents such as dimethyl sulfoxide (DMSO) and alkali solutions have been found to be capable of dissolving HAS. However, starch needs to be heated in 90% aqueous DMSO for about 24

h for the full dissolution; and alkaline solutions may induce depolymerization or oxidation of starch. Recent research has shown that ionic liquids (ILs) can dissolve starch and improve the degree of substitution (DS) and reaction efficiency for starch modification.¹⁰ Some examples of these modified starch include acetylated starch,¹¹ cationic starch,¹² esterified starch,¹³ carboxymethyl starch in 1-butyl-3-methylimidazolium chloride ([Bmim]Cl),¹⁴ fatty-acid starch esters in 1-ethyl-3-methylimidazolium acetate ([Emim][OAc]),¹⁵ and oxidized starch in a polyoxometalate IL.¹⁶ However, the toxicity of ILs remains uncertain and their high prices also restrict their applications.¹⁷

Besides ILs, some inorganic salt solutions can also destruct starch granules.¹⁸ Under certain concentrations, LiCl, KSCN, KI, BaF₂ and BaBr₂ solutions can be used to gelatinize starch granules at room temperature.¹⁹ ZnCl₂ solution has been proved to be as an effective solvent for cassava starch²⁰ and a capable plasticizer and reinforcing agent for starch-based materials.²¹ For the structural disorganization mechanism, Jane et al.¹⁸ have indicated that, due to the electronegative nature of starch, anions tend to repel the —OH groups of starch and stabilize starch granules; cations, on the other hand, attract the —OH groups of starch and destabilize starch granules. However, Shimizu et al.²² suggested that at high salt concentrations, the anion may enter the inside of the starch granule, breaking the granules and reducing the gelatinization temperature.

In this work, we compared the effects of ZnCl₂, MgCl₂ and CaCl₂ solutions, which are acid, neutral and alkaline solutions respectively, on the structural changes of HAS under a moderate temperature (50 °C). Based on that, we investigated the structural disorganization process of HAS granules and the aggregation of starch chains in the resulting salt solutions.

EXPERIMENTAL

Materials. Gelose 80 corn starch (G80) (about 80% amylose content, as determined by the manufacturer) was supplied by National Starch Pty Ltd. (Lane Cove, NSW 2066, Australia). Anhydrous zinc chloride (ZnCl_2), magnesium chloride (MgCl_2), and anhydrous calcium chloride (CaCl_2) were supplied by Aladdin Reagent (Shanghai) Co., Ltd. All these three chloride salts were chemically pure. Ethanol (analytical grade) was purchased from Guangzhou Chemical Reagent Factory (Guangzhou, China). All solutions were prepared with distilled water.

Sample Preparation. A certain amount of anhydrous ZnCl_2 , MgCl_2 or CaCl_2 were added to the beaker to prepare 43 wt% ZnCl_2 , 34 wt% MgCl_2 or 31 wt% CaCl_2 aqueous solutions (the concentrations were determined according to the discussion of Fig. 1). These salt solutions were then mixed with starch to achieve a starch concentration of 2 wt% (dry weight). After that, the solutions were located in a shaking bath at 50 °C. After a certain time, regenerated starch was obtained from the above solutions by adding absolute ethanol with continuous stirring (the volume of ethanol was about 2–5 times) and centrifuged under 3000 r/min for 5 min. Regenerated starch samples were washed three times by ethanol, and then dried and smashed into powder for further analysis. For comparison purposes, native G80 starch was also heated in a shaking bath at 50 °C for 4 h, which is taken as the control sample.

Light Microscopy. A polarized microscope (Axioskop 40 Pol/40A Pol, ZEISS) equipped with a 35mm SLA camera was used in the experimental work. The magnification used was 500× (50×10). Both normal and polarized lights were used for observation of starch suspensions (0.5 wt% concentration).

1
2
3
4 **X-ray Diffraction (XRD).** The crystallinity of starch samples was analyzed using an Xpert PRO
5
6 diffractometer (PANalytical B.V., Netherlands) operated at 40 mA and 40 kV with Cu K α radiation
7
8 (wavelength 0.1542 nm). The scanning was performed from 5° to 50° 2 θ at a speed of 10°/min and a
9
10 step size of 0.033°. The degree of crystallinity (X_c) was estimated using MDI Jade 6 software according
11
12 to a previous study.²³
13
14
15

16
17 **Fourier-Transform Infrared (FTIR).** FTIR spectra in the range of 400–4000 cm⁻¹ were obtained
18
19 using a Bruker Tensor-27 FT-IR Spectrometer (Bruker, Billerica, MA, USA). The samples were mixed
20
21 with KBr and well ground before being pressed into wafers.
22
23
24

25
26 **Scanning Electron Microscopy (SEM).** The morphology was imaged by a scanning electron
27
28 microscope (JEOL JSM-7001F, Tokyo, Japan) with an accelerating voltage of 10 kV and a spot size
29
30 of 6 nm. The samples were coated with platinum using an Eiko sputter coater.
31
32
33

34 **Residual cations in regenerated starch.** The amount of cation was detected by atomic absorption
35
36 spectrometry (AAS, Thermo Scientific iCE 3500) and the sample preparation was followed by Chinese
37
38 National Standards GB 5009.14-2017, GB 5009.92-2016, and GB 5009.241-2017. Specifically, 10 mL
39
40 of 69 wt% hydrogen nitrate solution and 0.5 mL of 70 wt% perchloric acid solution were added to 5 g
41
42 of the regenerated starch, and the obtained solution was heated to 220 °C for 2 h. After cooling, the
43
44 volume of the solution was adjusted to 50 mL by distilled water, and the content of cation was detected
45
46 using the AAS facility.
47
48
49
50
51

52
53 **Intrinsic viscosity.** Intrinsic viscosity was detected following a previous study.²⁰ Specifically, the
54
55 samples were dispersed in 1M KOH solution and stirred in a boiling water bath for 10 min. The
56
57 solutions were then cooled to room temperature and left overnight. After that, the solutions were
58
59
60

centrifuged under 3000 r/min for 6 min. The final concentrations of the solutions were 2.6–6.0 mg/mL.

The intrinsic viscosity was measured by an Ubbelohde dilution capillary viscometer (size 37, Shanghai Liangjing Glass Instrument Factory, China), which was immersed in a water bath maintained at 30.0±0.1 °C. The efflux time of solvent and solutions were measured in triplicate and averaged. Then, intrinsic viscosity $[\eta]$ can be calculated using the equation:

$$[\eta] = \lim_{c \rightarrow 0} \frac{t-t_0}{c \times t_0} \quad (1)$$

where t is the efflux time of the starch solution (s), t_0 is the efflux time of the KOH solution (s), and c represents the concentration of the starch solution (g/mL).

Water Absorption Measurement. The samples were soaked in water (5 wt% concentration) for 24 h and then vacuum filtrated. The water absorption was then calculated based on the weights before and after soaking. Triplicate tests were performed for each sample.

Rheology. After shaking at 50 °C for 4 h (see the section of Sample Preparation), the starch/chloride salt solutions were cooled to room temperature and tested immediately using a MCR 92 rheometer (Anton Paar GmbH, Austria) with a 60-mm-diameter cone-plate geometry and a Peltier temperature control system. For the salt solutions of different concentrations, the steady shear tests were carried out with the shear rate from 10 s⁻¹ to 500 s⁻¹ at 25 °C and 60 °C. Silicone oil (DC 200, Sigma–Aldrich) was placed around the edge of the measuring cell to maintain the moisture content of the sample. Silicone oil would hardly affect the experimental results, as it is immiscible with polysaccharide solutions and has a relatively lower viscosity (9.5 mPa/s at 20 °C). At least duplicate tests were performed for each sample.

The power-law model was used to describe the rheological behaviors:

$$\eta = K \cdot \dot{\gamma}^{n-1} \quad (2)$$

where η is the viscosity (Pa·s) of the solution, $\dot{\gamma}$ is the shear rate (s^{-1}), K is the consistency (Pa·s), and n is the power-law index.

Statistical Analysis. Microsoft Excel software was used for regression fitting based on a modified power-law equation and obtaining the correlation coefficients (R^2). Data were analyzed based on one-way analysis of variance (ANOVA) and Duncan's test for a statistical significance of $p \leq 0.05$ using IBM SPSS Statistics software (v19.0).

RESULTS

Structural Disorganization of Starch in Chloride Salt Solutions. Fig. 1a shows the birefringence (Maltese cross) of G80 starch granules in ZnCl_2 , MgCl_2 , and CaCl_2 solutions at 50 °C for 4 h. Native G80 starch granules exhibited clear birefringence. The birefringence dimmed gradually with increasing salt concentration. When these salt solutions reached a certain concentration, namely 33% ZnCl_2 , 27% MgCl_2 and 25% CaCl_2 , the birefringence disappeared completely, suggesting the disruption of crystalline structure in the granules. However, as shown in the corresponding normal-light images, although some granules were swollen, most of them were still in their original shape. During the structural disorganization, their semicrystalline lamellae were destroyed by ions firstly, but their outer layer (which is formed by entangled amylose chains and is compact) could not be destroyed under these salt concentrations.⁸ The remaining granule remnants were stable under these salt concentrations, even after 8 h of treatment.

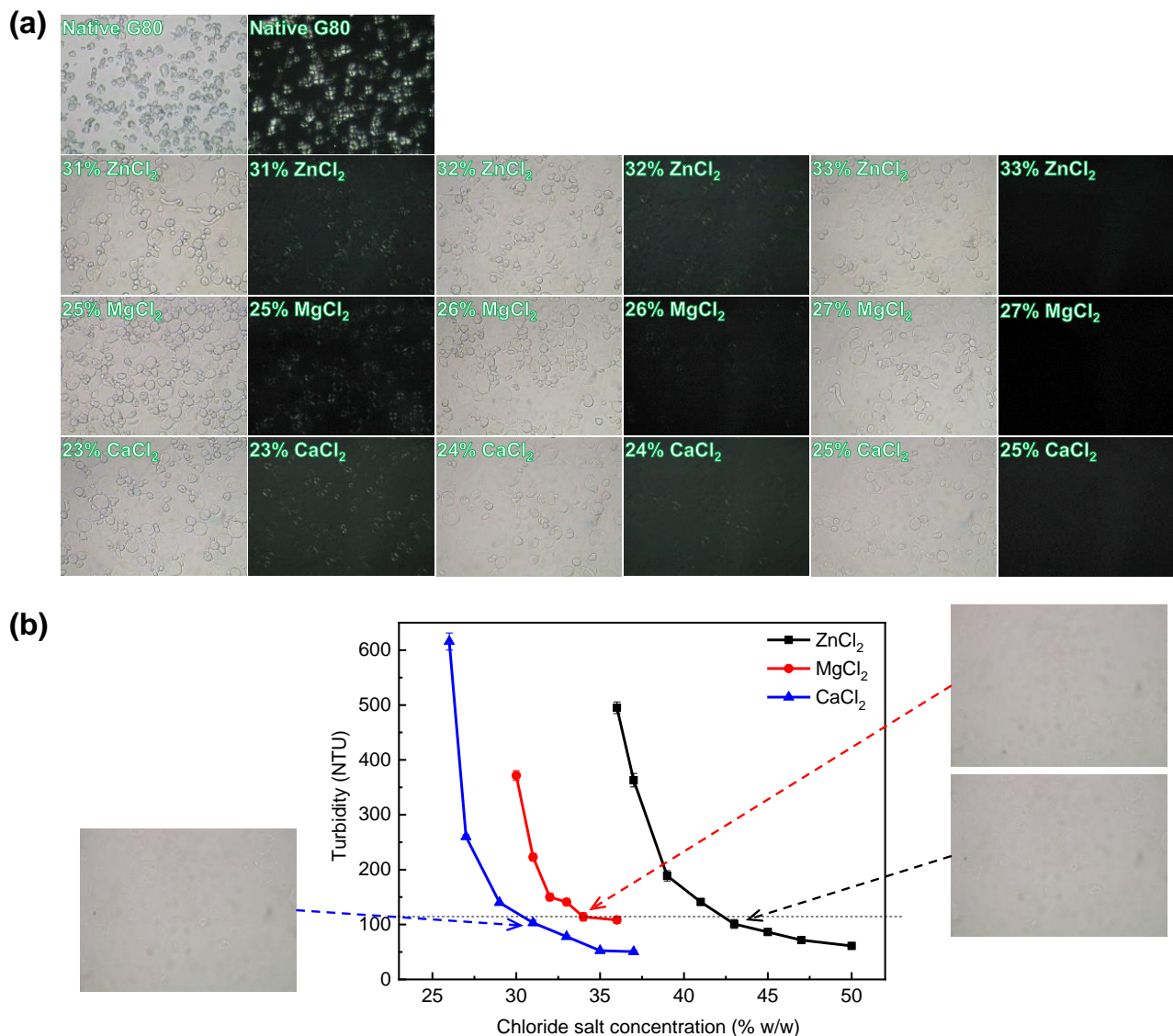


Fig. 1. (a) Microscopic images of G80 starch dissolved in chloride salt solutions under normal and polarized light; (b) Turbidity of G80 starch dissolved in three chloride salt solutions. The error bars represent standard deviations.

With increasing salt concentration, the birefringence of starch disappeared and the swollen granules became more and more ambiguous. The transparency of the solutions was characterized by turbidity (Fig. 1b). In previous studies,^{17,20} when cassava starch was dissolved in ZnCl₂ solution with increasing concentration, there was an abrupt decrease in turbidity at a certain salt concentration (critical

concentration); after this point, the turbidity became stable. However, in this study, for G80 starch in ZnCl₂ and CaCl₂ solutions with increasing concentration, the turbidity kept decreasing. For the starch in MgCl₂ solution with increasing concentration, the turbidity was stable at around 110 NTU (36% is its saturated concentration under room temperature) and no obvious granules could be observed under microscopy. Thus, this value was considered as the threshold for a homogeneous solution.

Based on these results, the salt concentrations to destruct G80 starch were determined to be 43 wt% ZnCl₂, 34 wt% MgCl₂, and 31 wt% CaCl₂ (Table 1). Under these salt concentrations, the starch-salt solutions were transparent and all the granules vanished under microscopy. Table 1 listed the parameters for G80 starch to be destructed in three chloride salt solutions. The salt concentrations for the disappearance of both birefringence and granules follow the order of ZnCl₂ > MgCl₂ > CaCl₂.

Table 1. Parameters of G80 starch dissolved in three chloride salt solutions.

	ZnCl ₂	MgCl ₂	CaCl ₂	Control
Salt concentration at birefringence disappearance (wt%)	33	27	25	–
pH of salt solution	4.21	7.10	9.52	–
Salt concentration at granule disappearance (wt%)	43	34	31	–
pH of salt solution	3.65	6.54	9.17	–
Content of cations in regenerated starch (mg/g)	53.0±1.3 ^a	18.2±0.9	21.3±0.7	–
Water absorption ratio of regenerated starch (wt%)	152±9.6	222±8.7	144±12	60.9±5

^a Average ± standard deviations.

1
2
3
4 **Morphology of Regenerated Starch.** Fig. 2 shows the SEM images of different regenerated starch
5
6 samples and the control sample. The controlled samples displayed a nearly spherical shape with a
7
8 smooth surface. For the regenerated starch from the MgCl₂ solution, the granules could be observed
9
10 within 10 min of treatment; and after 30 min, some nanoparticles emerged on the surface of intact
11
12 granules. After 4 h, some granules still remained with greater amounts of nanoparticles located on their
13
14 surfaces. In other words, the MgCl₂ solution not only destructed the starch granules but also resulted
15
16 in the formation of nanoparticles. The residual granules contributed to the high turbidity of the
17
18 starch/MgCl₂ solution, which was stable at around 110 NUT (Fig. 1b).
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

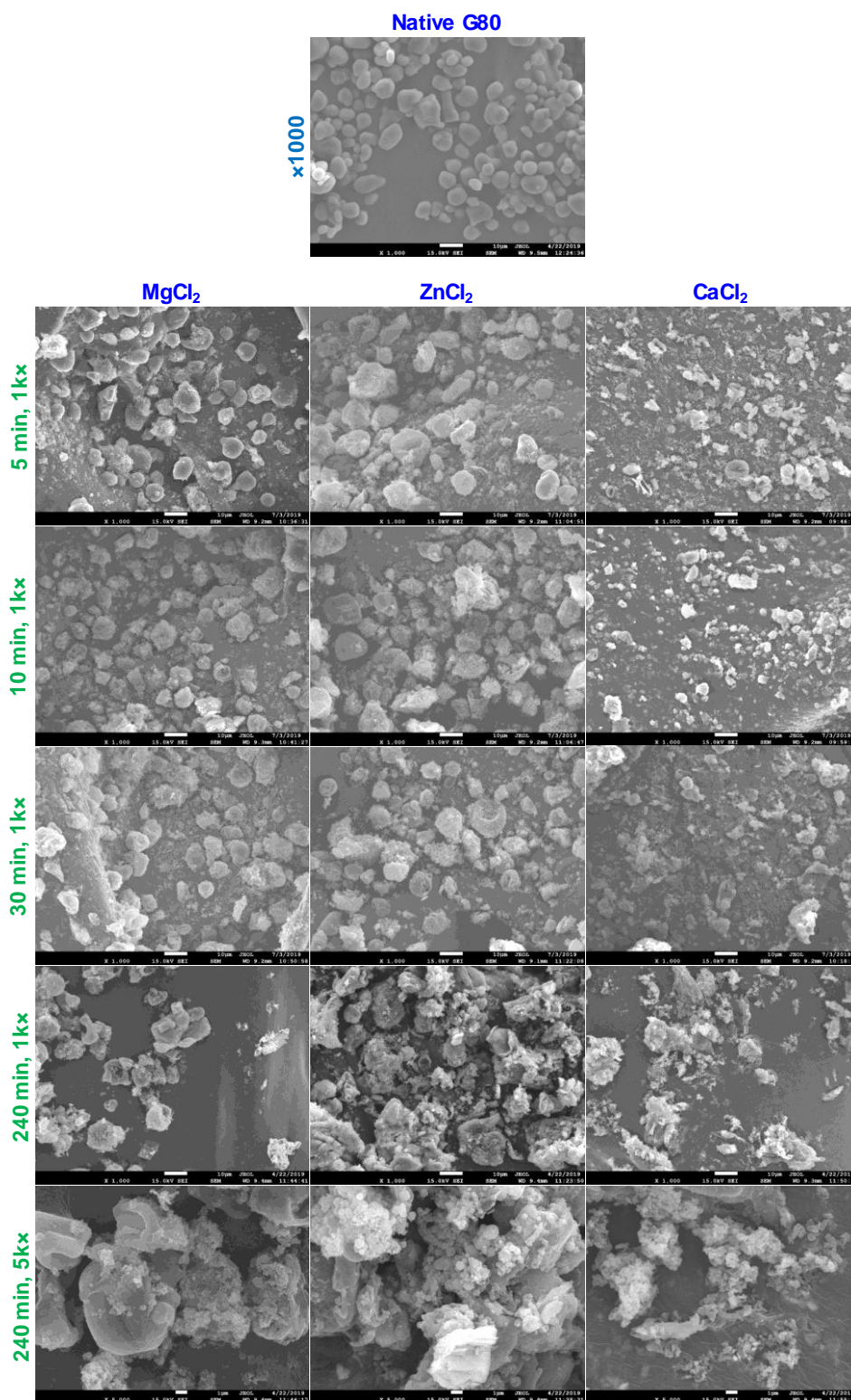


Fig. 2 Morphology of regenerated G80 starch from 34 wt% MgCl₂, 43 wt% ZnCl₂ and 31 wt% CaCl₂ solutions (magnification: 1k× or 5k×)

For the regenerated starch from the ZnCl₂ solution, 30 min of treatment led to some degree of

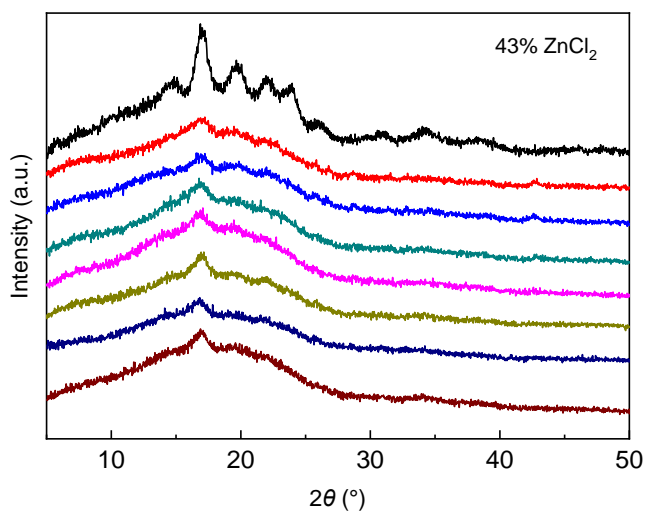
1
2
3
4 granule structural disorganization and the formation of nanoparticles. After 4 h, only nanoparticles
5
6 (with some in the form of agglomerates) could be seen without original starch granules. For the
7
8 regenerated starch from the CaCl_2 solution, all the starch granules changed into tiny fragments within
9
10 10 min. After 30 min, only agglomerated nanoparticles could be seen. This shows the efficient
11
12 structural disorganization of G80 starch in the CaCl_2 solution.
13
14
15

16
17 Besides, we can see that the granules from the MgCl_2 and ZnCl_2 solutions within 10 min were larger
18
19 than the controlled ones, which could be due to swelling. This result is in agreement with the
20
21 microscopic observation (Fig. 1b) and with the previous results in hot water²⁴ and an IL.²⁵ However,
22
23 after 30 min of treatment, the granules became smaller than the controlled ones due to the
24
25 disintegration of granules.
26
27
28
29

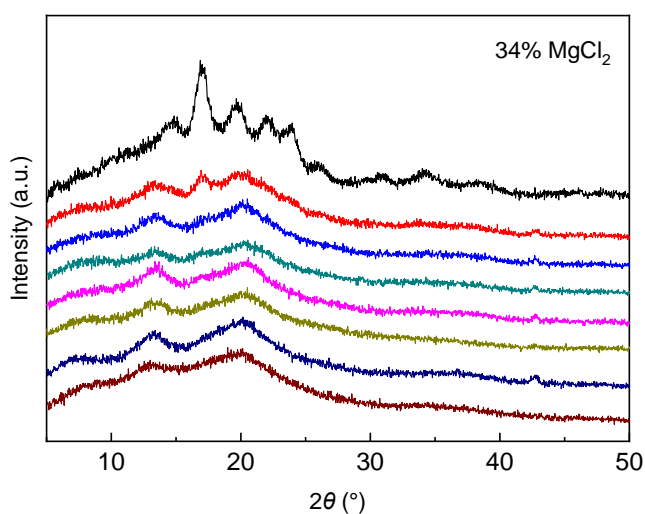
30
31 **Starch Structural Changes.** Fig. 3 illustrates the XRD curves and the degree of crystallinity (X_c)
32
33 of regenerated starch. For native G80 starch, the crystalline peaks located at 5.6° , 15° , 17° , 19.5° , 22°
34
35 and 23.5° 2θ , which indicate the B-type pattern.²⁶⁻²⁷ Its X_c was 14.7 ± 0.2 %, similar to previous
36
37 results.^{21,28} For the regenerated starch from the ZnCl_2 solution (Fig. 3a), the crystalline pattern became
38
39 very weak with only the reflection at 17° 2θ being apparent. For the samples from the MgCl_2 solution
40
41 (Fig. 3b), only the peaks at 13° and 20° 2θ remained. These reflections are characteristic of the V-type
42
43 crystalline pattern, which is due to the complexation of starch long chains with small molecules such
44
45 as fat and iodine to form a single helical structure.²⁹ Besides, the peak in 17° 2θ remained within the
46
47 first 5 min but disappeared after 10 min. For starch from the CaCl_2 solution (Fig. 3c), all peaks vanished
48
49 within 5 min. The different crystalline patterns of regenerated starch show different interactions
50
51 between starch chains and metal ions. Moreover, no new peaks emerged on the XRD curves, indicating
52
53
54
55
56
57
58
59
60

that the nanoparticles in the regenerated starch (Fig. 2) were not inorganic salt crystals.

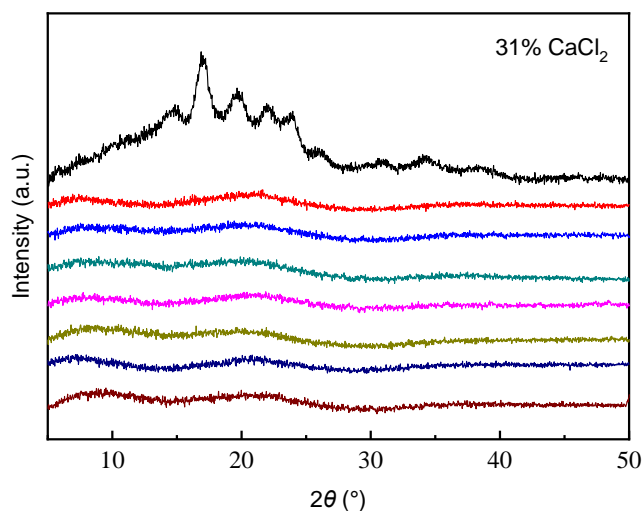
Fig. 3d shows that in all these cases, X_c decreased sharply within 5 min. For the regenerated starch from CaCl_2 solution, X_c decreased to 0 within 5 min. For the samples from the ZnCl_2 and MgCl_2 solutions, X_c dropped sharply within 30 min, to $5.7 \pm 0.2\%$ and $10.2 \pm 0.2\%$ respectively. After that, the decrease in X_c was much slower. After 4 h of treatment, X_c was $3.6 \pm 0.3\%$ and $7.3 \pm 0.3\%$ for the samples in the ZnCl_2 and MgCl_2 solution respectively.



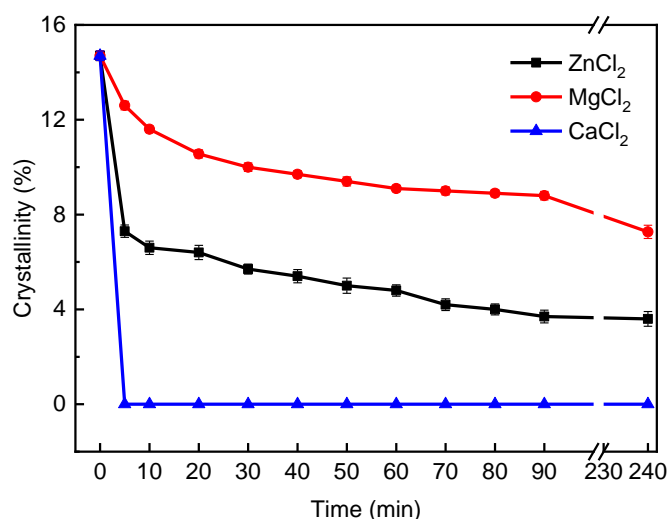
(a)



(b)



(c)



(d)

Fig. 3. XRD patterns for the regenerated G80 starch samples from 43 wt% ZnCl₂ (a), 34 wt% MgCl₂ (b), and 31 wt% CaCl₂ (c) solutions at different times of treatment (0 min, 5 min, 10 min, 20 min, 30 min, 60 min, 90 min and 240 min, from top to bottom). (d) shows the degree of crystallinity of the regenerated starch samples from 43 wt% ZnCl₂, 34 wt% MgCl₂, and 31 wt% CaCl₂ solutions as a function of time. The error bars represent standard deviations.

All FTIR spectra (Fig. 4) were similar without the appearance of new peaks, irrespective of native or regenerated starch samples, indicating no chemical changes in the regenerated starch. This is

1
2
3
4 different from previous studies,^{20,30} where for cassava starch dissolved in a ZnCl_2 solution, the O—H
5
6 bending of adsorbed water at 1647 cm^{-1} was shifted to 1627 cm^{-1} , indicating the water absorption
7
8 function of the regenerated starch became weak. However, here for G80 starch, this shift was not
9
10 apparent. Also, the water absorption ratios of the regenerated starch were much higher (nearly 1.5 to
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

3.5 times) than the controlled G80 starch (Table 1).

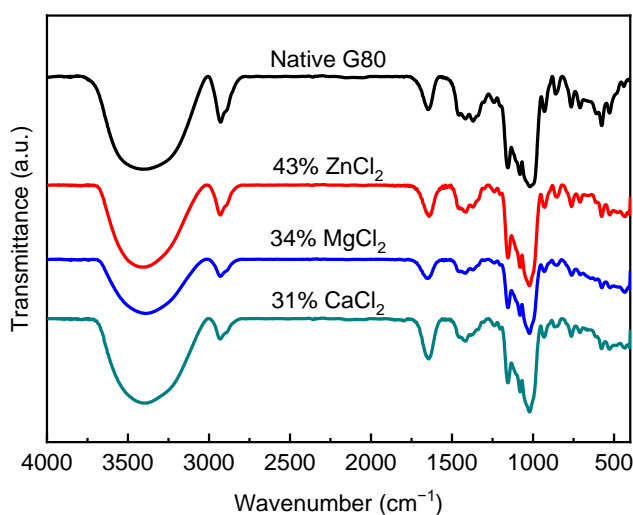


Fig. 4. FTIR spectra for native and regenerated G80 starch samples from 43 wt% ZnCl_2 solutions (a), 34 wt% MgCl_2 solutions (b), and 31 wt% CaCl_2 solutions (c) after 4 h of dissolution.

Intrinsic Viscosity of Regenerated Starch. Fig. 5 shows the intrinsic viscosity ($[\eta]$) of native and regenerated G80 starch. Intrinsic viscosity is an important parameter reflecting the size of a macromolecule in a given solvent at a certain temperature.³¹ Native starch exhibited the highest $[\eta]$ value. The sample from 34% MgCl_2 solution had a $[\eta]$ value very close to that of native starch. The $[\eta]$ value of regenerated starch from the CaCl_2 solution was higher than that from the ZnCl_2 solution, but both of them were significantly lower than that from the MgCl_2 solution.

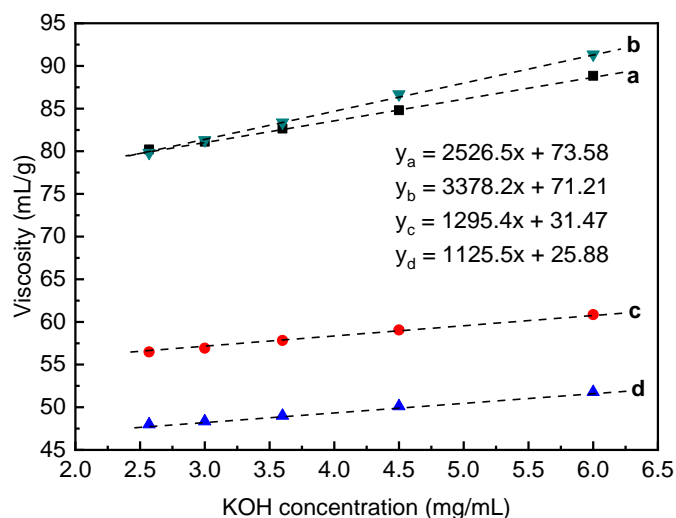


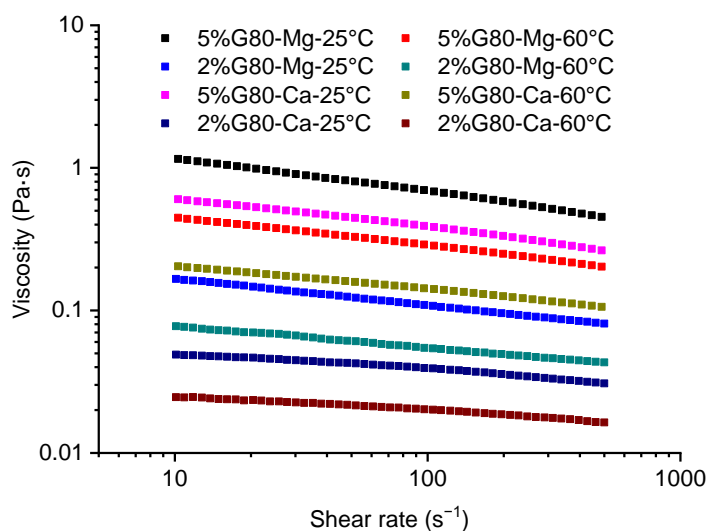
Fig. 5. Intrinsic viscosity of native starch (a), and regenerated starch samples from 34 wt% MgCl₂ solutions (b), 31 wt% CaCl₂ solutions (c), and 43 wt% ZnCl₂ solutions (d).

The intrinsic viscosity of polymers is caused by the friction force between the solvent and the solute in a dilute solution and can be used to reflect the size of the macromolecular conformation in the certain solution.³²⁻³³ For starch, the size of their macromolecular conformation was caused by the degree of polymerization (DP), namely the molecular weight.³⁴ Therefore, a lower $[\eta]$ value may indicate that the structural disorganization of starch granules in the ZnCl₂ and CaCl₂ solutions was accompanied by degradation, which was mainly caused by H⁺ and OH⁻ ions. Moreover, since the pH value of 43 wt% ZnCl₂ solution was 3.65 and that of 31 wt% CaCl₂ solution was 9.17 (Table 1), the $[\eta]$ value of the regenerated starch from the ZnCl₂ solution was lowest. The MgCl₂ solution is pH-neutral, so starch chains were hardly degraded, with similar $[\eta]$ as that of native starch.

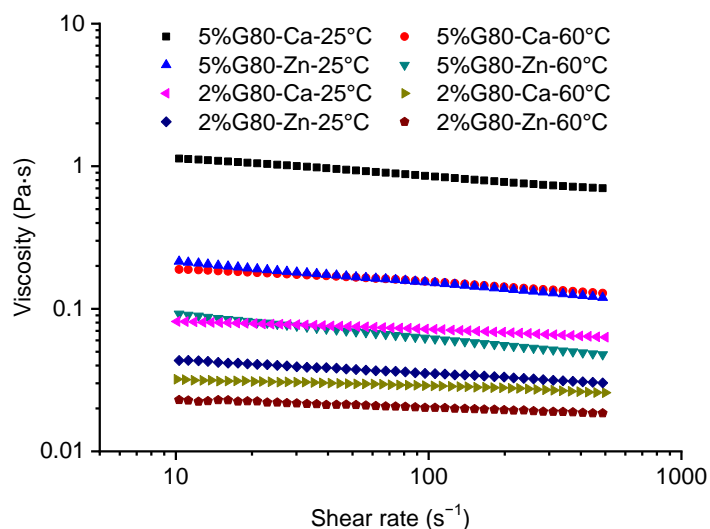
Since the degradation of molecular mass during the granule structural disorganization is mainly caused by the H⁺ or OH⁻ ions in the solution, this kind of hydrolysis should have no selectivity on α -

(1-4) or α -(1-6) glycosidic bonds. In other words, chain degradation should occur to both amylose and amylopectin.

Rheological Properties of Starch/Chloride Salt Solutions. Since the concentration of saturated MgCl_2 solution at 25 °C is 36 wt% and the threshold concentration for ZnCl_2 solution to dissolve starch is 43 wt%, we compared the rheological properties of Gelose 80 starch in 34% MgCl_2 and 34% CaCl_2 solutions (Fig. 6a) and those of Gelose 80 starch in 43% ZnCl_2 and 43% CaCl_2 solutions (Fig. 6b). Fig. 6a shows that the viscosity of starch/ MgCl_2 solution was higher than that of starch/ CaCl_2 solution at 34% concentration. Fig. 6b shows that the viscosity of starch/ CaCl_2 solution was higher than that of starch/ ZnCl_2 solution at 43% concentration. Thus, the viscosity of different starch solutions should follow the order of $\text{MgCl}_2 > \text{CaCl}_2 > \text{ZnCl}_2$ under the same conditions.



(a)



(b)

Fig. 6. Viscosity–shear rate curves for G80 starch in 34% MgCl₂ and CaCl₂ solutions (a) and in 43% ZnCl₂ and CaCl₂ solutions (b).

Table 2 shows the calculated parameters of the power-law equation at different temperatures and concentrations. Their correlation coefficients (R^2) were all higher than 0.99, showing a strong power-law dependence of viscosity on shear rate. The power-law index n for the starch/MgCl₂ solution is lower than that for the starch/CaCl₂ solution, and the n value of the starch/CaCl₂ solution is lower than that of the starch/ZnCl₂ solution. For a pseudo-plastic solution, a lower n value indicates a greater shear-thinning behavior and a larger molecular configuration of the solute.³⁵⁻³⁷ Therefore, the n result reflects that the configuration of starch chains in the solutions followed the order of MgCl₂ > CaCl₂ > ZnCl₂, which is in agreement with the intrinsic viscosity results.

Table 2. Power-law parameters for starch in three chloride solutions

Sample	n	K	R^2
--------	-----	-----	-------

5%G80-34%MgCl ₂ -25°C	0.751 ^a	2.1249 ^o	0.9961 ^b
5%G80-34%MgCl ₂ -60°C	0.791 ^c	0.7438 ^k	0.9972 ^c
2%G80-34%MgCl ₂ -25°C	0.845 ^f	0.1856 ^g	0.9998 ^d
2%G80-34%MgCl ₂ -60°C	0.888 ^g	0.845 ^l	0.9911 ^b
5%G80-34%CaCl ₂ -25°C	0.76 ^b	1.1732 ^m	0.9967 ^b
5%G80-34%CaCl ₂ -60°C	0.816 ^d	0.3313 ^j	0.9977 ^d
2%G80-34%CaCl ₂ -25°C	0.88 ^g	0.0771 ^d	0.9955 ^b
2%G80-34%CaCl ₂ -60°C	0.898 ^h	0.0355 ^b	0.9938 ^b
5%G80-43%ZnCl ₂ -25°C	0.852 ^f	0.3025 ⁱ	0.999 ^d
5%G80-43%ZnCl ₂ -60°C	0.835 ^e	0.1329 ^f	1 ^d
2%G80-43%ZnCl ₂ -25°C	0.904 ^h	0.0549 ^c	0.9971 ^b
2%G80-43%ZnCl ₂ -60°C	0.941 ^j	0.0266 ^a	0.9963 ^b
5%G80-43%CaCl ₂ -25°C	0.875 ^g	1.5158 ⁿ	0.9925 ^b
5%G80-43%CaCl ₂ -60°C	0.882 ^g	0.2669 ^h	0.9995 ^d
2%G80-43%CaCl ₂ -25°C	0.923 ⁱ	0.1026 ^e	0.997 ^b
2%G80-43%CaCl ₂ -60°C	0.934 ^j	0.039 ^b	0.9802 ^a

Superscripts with different letters in the same column indicate significant differences ($p \leq 0.05$).

DISCUSSION

Structural disorganization Process of Starch in Chloride Salt Solutions. The structure of G80 granules includes the hilum locates in the center of granule, loosely-packed semicrystalline growth

1
2
3
4 rings around the hilum formed mainly by amylopectin, and a thick and compact outer layer formed
5
6 primarily by entangled amylose chains.^{5, 38-39}
7
8

9 The outer layer of starch granules exhibits a semipermeable-membrane-like behavior for both
10 cations and anions, which prevents ions at low concentrations from entering the granule.²² Since there
11 are no cavities and channels on G80 granules,⁴⁰⁻⁴¹ the high salt concentration is needed for the
12 penetration of ions through the outer layer. Once ions have passed through the outer layer and reached
13 the hilum, they will destruct the semicrystalline lamellar regularity firstly, as reflected by the
14 disappearance of birefringence and a significant reduction in X_c . The outer layer will require an even
15 higher ion concentration to be disintegrated.
16
17
18
19
20
21
22
23
24
25
26
27

28 **Aggregation of Starch Chains to Form Nanoparticles.** Large amounts of nanoparticles can be
29 seen in regenerated starch in all cases (see SEM results). These nanoparticles should not be the
30 inorganic salt crystals (XRD results). Previous studies have shown that coordinated complexes may
31 form between starch chains and some metal cations since the positively charged cations can be paired
32 with the isolated charges on the oxygen atoms of starch hydroxyl groups.⁴²⁻⁴⁵ Therefore, the
33 nanoparticles observed could be considered as a form of starch–metal cation complexes, which can
34 stabilize starch chains in the solution.
35
36
37
38
39
40
41
42
43
44
45
46

47 Moreover, different chloride salt solutions could result in different structures of starch–metal cation
48 complex. The starch– Mg^{2+} complex presents a V-type crystalline structure. Regarding this, the
49 coordination of Mg^{2+} with starch may allow the main chains of amylose or the long branches of starch
50 macromolecules to form single helices, which then align into crystals under the neutral condition.
51 However, such alignment may not be possible in the $ZnCl_2$ or $CaCl_2$ solution, where an acidic or
52
53
54
55
56
57
58
59
60

1
2
3
4 alkaline condition may restrict the formation of any crystalline structure. Therefore, starch–Zn²⁺ and
5
6 starch–Ca²⁺ complexes are amorphous.
7

8
9 Early work has involved the preparation of starch–metal cation complexes, such as the bismuth (III)
10
11 and bismuth (V) derivatives of starch,⁴⁶⁻⁴⁷ starch–aluminum complex,⁴⁸ starch–copper complex and
12
13 starch–iron complex.⁴⁹ These complexes are potential to be used in pharmaceutical applications such
14
15 as for skin diseases, gastric ulcers, or hair growth. In this work, we have revealed a facile method to
16
17 prepare starch–metal cation nanoparticles. The characteristics including embedding properties of these
18
19 nanoparticles for active ingredients will be studied in the future.
20
21
22
23
24

25
26 **Structural Disorganization Ability of Ions on Starch.** Based on the residues and the degree of
27
28 crystallinity, the structural disorganization ability of three chloride salt solutions on G80 starch
29
30 granules follows the order of CaCl₂ > ZnCl₂ > MgCl₂. Then, what is the key factor to determine this
31
32 ability?
33
34

35
36 For the structural disorganization mechanism of starch in ILs, previous studies have indicated that
37
38 the anions have a protonation effect and can weaken the hydrogen bonding between starch chains, thus
39
40 promoting the phase transition of granules.⁵⁰⁻⁵¹ Moreover, the cations can improve the protonation
41
42 effect of anions by their ionic radius.⁵²⁻⁵⁴ In other words, the larger the size of the cation, the stronger
43
44 effect does the anion have. Moreover, besides the hydrogen-bonding capacity of IL anion, the viscosity
45
46 of water/IL mixtures also plays a key role in the disruption of the starch structure.⁵⁵
47
48
49
50

51
52 The three chloride salt solutions studies here present similar viscosities. The ionic radius of Ca²⁺ is
53
54 100 pm, that of Mg²⁺ is 72 pm, and that of Zn²⁺ is 74 pm,⁵⁶ which show no obvious difference as well.
55
56 Therefore, the key factor should be attributed to the synergistic effect of ion. Specifically, based on the
57
58
59
60

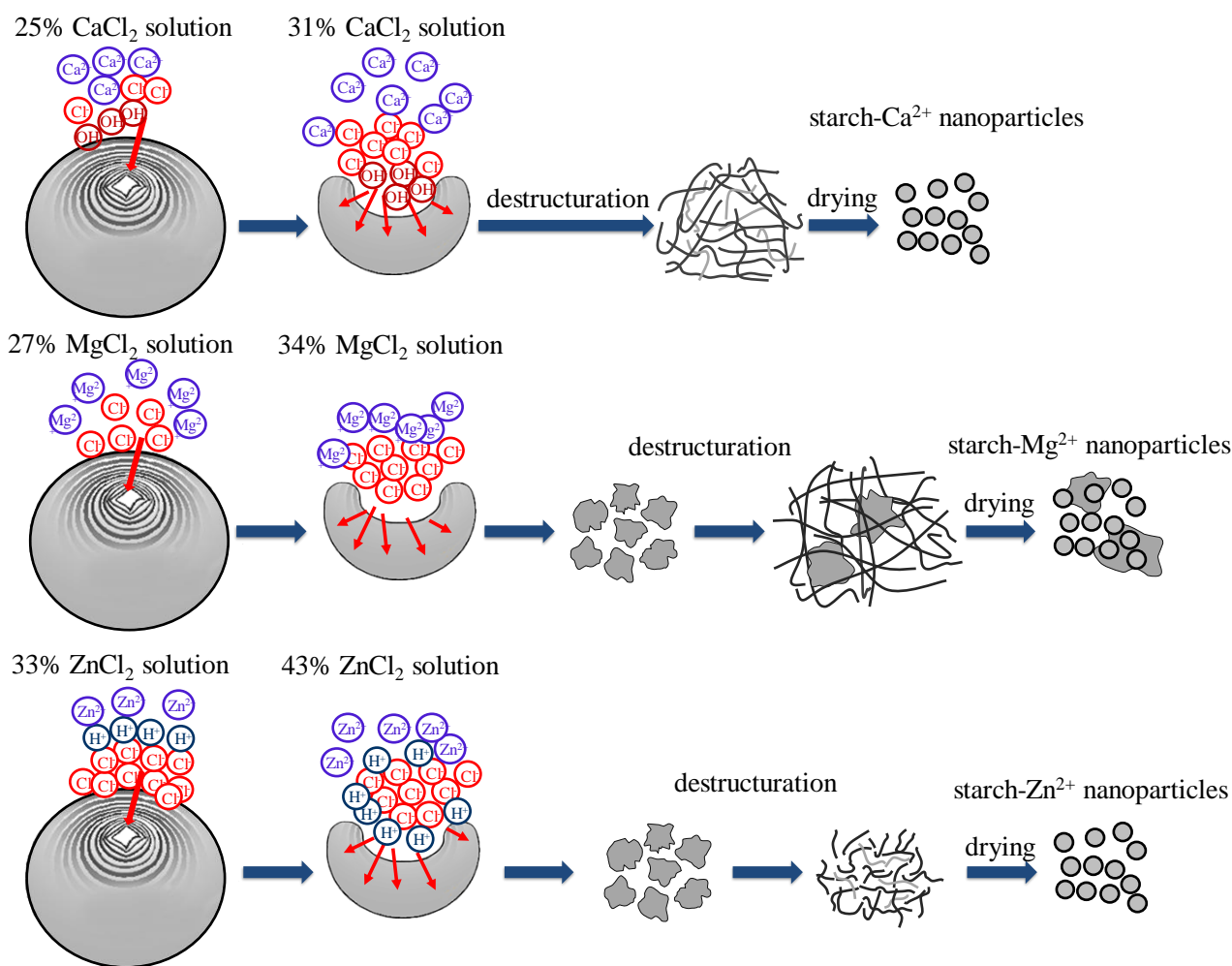
1
2
3
4 acid–base proton theory, Cl^- may accept a proton and break the hydrogen bonds between starch chains;
5
6 however, its protonation effect is very weak.⁵⁷ With the pH of these salt solutions noticed, it could be
7
8 possible that H^+ and OH^- ions assist the structural disorganization.
9
10

11
12 For the alkaline CaCl_2 solution, both Cl^- and OH^- anions are strong hydrogen-acceptors.⁵⁸⁻⁵⁹
13
14 Therefore, the structural disorganization ability of the CaCl_2 solution is the highest, and all residues
15
16 changed into nanoparticles within a short time. When the acidic ZnCl_2 solution is used, the acid
17
18 hydrolysis by H^+ facilitates the destruction of starch granules, accompanied by the degradation of
19
20 starch chains. For MgCl_2 solution, which is pH-neutral, the concentration of H^+ and OH^- ions can be
21
22 too low to facilitate the protonation by Cl^- , resulting in incomplete structural disorganization of starch
23
24 granules (Fig. 7).
25
26
27
28
29

30
31 The threshold salt concentration to destruct starch granules follows the order of $\text{CaCl}_2 < \text{MgCl}_2 <$
32
33 ZnCl_2 . The reason should be the influence of H^+ or OH^- ions on the protonation effect of Cl^- .
34
35 Specifically, OH^- anions can destroy the hydrogen bonds without varying the structural
36
37 disorganization effect of Cl^- ; thus, the threshold concentration of CaCl_2 solution is the lowest. However,
38
39 although H^+ can hydrolyze starch chains, it hinders the protonation effect of Cl^- anions, as H^+ can
40
41 accept the electron offered by Cl^- . Therefore, a greater number of Cl^- ions (a higher concentration of
42
43 ZnCl_2) are needed to destruct starch granules.
44
45
46
47
48

49
50 Besides HAS, we found all these chloride salt solutions can fully destroy normal and waxy starch
51
52 granules as well, and the dissolution of waxy and normal starches is easier than that of HAS, namely
53
54 with lower salt concentrations and a shorter time. This may be explained by the looser granule structure
55
56 of waxy and normal starches than that of HAS. The details regarding the dissolution of different
57
58
59
60

1
2
3
4 starches with different amylose contents by these chloride salt solutions will be discussed in our future
5
6
7 work. Moreover, the dissolution mechanism is worth to be further investigated using theoretical
8
9
10 chemistry approaches (e.g. molecular modelling simulations) and electrochemical studies.



47 Fig. 7. Schematic diagram representing the structural disorganization of starch by metal chloride salt
48 solutions.
49

55 CONCLUSIONS

56
57
58 Here, we have compared the effects of three chloride salt solutions on the structural disorganization of
59
60

1
2
3
4 G80 HAS granules. The crystalline structure of starch granules can be destroyed in 33 wt% ZnCl₂, 27
5
6 wt% MgCl₂, and 25 wt% CaCl₂ solutions, and the complete destruction of the granules needs 43 wt%
7
8 ZnCl₂, 34 wt% MgCl₂, and 31 wt% CaCl₂ solutions at 50 °C. In other words, the threshold salt
9
10 concentration follows the order of CaCl₂ < MgCl₂ < ZnCl₂. This should be mainly caused by the
11
12 protonation effect of Cl⁻ anions on the hydrogen bonds, which is promoted by OH⁻ and hindered by
13
14 H⁺. Meanwhile, the degradation of starch by OH⁻ and H⁺ is inevitable. The structural disorganization
15
16 ability of the salt solutions follows the order of CaCl₂ > ZnCl₂ > MgCl₂. Specifically, starch chains
17
18 dissociate in the CaCl₂ and ZnCl₂ solutions totally. Nevertheless, in MgCl₂ solutions, although most
19
20 starch chains were detached, small amounts of granule residues remain.
21
22
23
24
25
26
27

28 Surprisingly, the regenerated starch from the chloride salt solutions is in the form of nanoparticles,
29
30 which are likely to be formed by the complexation between the dissociated starch chains and the metal
31
32 cations. XRD results indicate the different crystalline structures of these complexes, with Mg²⁺ cations
33
34 promoting the formation of V-type crystallites whereas Ca²⁺ cations lead to completely amorphous
35
36 nanoparticles. Besides, we also found that the water absorption ratio of regenerated starch is 1.5 to 3.5
37
38 times that of the control (treated in water). Thus, we demonstrate a facile method using non-
39
40 derivatizing metal chloride salts to prepare absorbent biopolymer nanoparticles, which can be
41
42 potentially used for the encapsulation and controlled release of active ingredients in food and
43
44 pharmaceutical applications.
45
46
47
48
49
50
51

52 In all, this work has shown that HAS can be fully dissolved by cheap metal chloride salt solutions
53
54 without derivatization effect. This finding could lead to 'green' processes for easy modification of
55
56 HAS and for the preparation of plasticized starch-based ion-conductive materials.
57
58
59
60

1
2
3
4
5
6
7 AUTHOR INFORMATION
8
9

10 **Corresponding Authors**
11

12
13 * Email address: liu_peng@gzhu.edu.cn (P. Liu)
14

15
16 * Email address: f.xie@uq.edu.au; fwhsieh@gmail.com (F. Xie)
17
18

19 **Author Contributions**
20

21
22 P.L. and F.X. conceived and designed the study. Y.L. and C.M. conducted experiments and data
23
24 analysis. The manuscript was written through the contributions of all authors. All authors have given
25
26 approval to the final version of the manuscript. #These authors contributed equally.
27
28
29

30 **Notes**
31

32
33 The authors declare no competing financial interest.
34
35

36 **ACKNOWLEDGMENT**
37

38
39 This research was funded by the Natural Science Foundation of Guangdong Province (Project No.
40
41 2018A0303130048), the Pearl River S&T Nova Program of Guangzhou (Grant No. 201610010019),
42
43 and the National Natural Science Foundation of China (NSFC) (Project No. 21106023) F. Xie
44
45 acknowledges support from the European Union's Horizon 2020 research and innovation programme
46
47 under the Marie Skłodowska-Curie grant agreement No. 798225. F. Xie also acknowledges support
48
49 from the Guangxi Key Laboratory of Polysaccharide Materials and Modification, Guangxi University
50
51 for Nationalities, China (Grant No. GXPSMM18ZD-02).
52
53
54
55
56
57

58 **REFERENCES**
59
60

- 1
2
3
4 (1) Haq, F.; Yu, H.; Wang, L.; Teng, L.; Haroon, M.; Khan, R. U.; Mehmood, S.; Ullah, R. S.; Khan,
5
6 A.; Nazir, A. Advances in chemical modifications of starches and their applications. *Carbohydrate*
7
8 *Research* **2019**, *476*, 12-35, DOI: 10.1016/j.carres.2019.02.007.
9
10
11
12 (2) Li, H.; Gidley, M. J.; Dhital, S. High-Amylose Starches to Bridge the “Fiber Gap”: Development,
13
14 Structure, and Nutritional Functionality. *Comprehensive Reviews in Food Science and Food Safety*
15
16 **2019**, *18* (2), 362-379, DOI: 10.1111/1541-4337.12416.
17
18
19
20 (3) Vilaplana, F., Meng, D., Hasjim, J., Gilbert, R. G. Two-dimensional macromolecular distributions
21
22 reveal detailed architectural features in high-amylose starches. *Carbohydrate Polymers* **2014**, *113*,
23
24 539-551, DOI: 10.1016/j.carbpol.2014.07.050.
25
26
27
28 (4) Yang, J.; Xie, F.; Wen, W.; Chen, L.; Shang, X.; Liu, P. Understanding the structural features of
29
30 high-amylose maize starch through hydrothermal treatment. *International journal of biological*
31
32 *macromolecules* **2016**, *84*, 268-274, DOI: 10.1016/j.ijbiomac.2015.12.033.
33
34
35
36 (5) Jane, J.-L. Current understanding on starch granule structures. *Journal of Applied Glycoscience*
37
38 **2006**, *53*(3), 205-213, DOI: 10.5458/jag.53.205.
39
40
41
42 (6) Benyerbah, N.; Ispas-Szabo, P.; Sakeer, K.; Chapdelaine, D.; Mateescu, M. A. Ampholytic and
43
44 Polyelectrolytic Starch as Matrices for Controlled Drug Delivery. *Pharmaceutics* **2019**, *11* (6), 253,
45
46 DOI: 10.3390/pharmaceutics11060253.
47
48
49
50 (7) Tang, H.; Fan, S.; Li, Y.; Dong, S. Amylose: Acetylation, Optimization, and Characterization. *J*
51
52 *Food Sci* **2019**, *84* (4), 738-745, DOI: 10.1111/1750-3841.14487.
53
54
55
56 (8) Luo, Z.; Zou, J.; Chen, H.; Cheng, W.; Fu, X.; Xiao, Z. Synthesis and characterization of amylose-
57
58 zinc inclusion complexes. *Carbohydr Polym* **2016**, *137*, 314-320, DOI: 10.1016/j.carbpol.2015.10.100.
59
60

- 1
2
3
4 (9) Li, M.; Liu, P.; Zou, W.; Yu, L.; Xie, F.; Pu, H.; Liu, H.; Chen, L. Extrusion processing and
5
6 characterization of edible starch films with different amylose contents. *Journal of Food Engineering*
7
8 **2011**, *106* (1), 95-101, DOI: 10.1016/j.jfoodeng.2011.04.021.
9
10
11
12 (10) Ren, F.; Wang, J.; Yu, J.; Xiang, F.; Wang, S.; Wang, S.; Copeland, L. Dissolution of Maize Starch
13
14 in Aqueous Ionic Liquids: The Role of Alkyl Chain Length of Cation and Water:Ionic Liquid Ratio.
15
16 *ACS Sustainable Chemistry & Engineering* **2019**, *7* (7), 6898-6905, DOI:
17
18 10.1021/acssuschemeng.8b06432.
19
20
21
22 (11) Biswas, A.; Shogren, R.; Stevenson, D.; Willett, J.; Bhowmik, P. K. Ionic liquids as solvents for
23
24 biopolymers: Acylation of starch and zein protein. *Carbohydrate polymers* **2006**, *66* (4), 546-550, DOI:
25
26 10.1016/j.carbpol.2006.04.005.
27
28
29
30 (12) Wang, Y.; Xie, W. Synthesis of cationic starch with a high degree of substitution in an ionic liquid.
31
32 *Carbohydrate Polymers* **2010**, *80* (4), 1172-1177, DOI: 10.1016/j.carbpol.2010.01.042.
33
34
35
36 (13) Zarski, A.; Ptak, S.; Siemion, P.; Kapusniak, J. Esterification of potato starch by a biocatalysed
37
38 reaction in an ionic liquid. *Carbohydr Polym* **2016**, *137*, 657-663, DOI: 10.1016/j.carbpol.2015.11.029.
39
40
41
42 (14) Xie, W.; Zhang, Y.; Liu, Y. Homogenous carboxymethylation of starch using 1-butyl-3-
43
44 methylimidazolium chloride ionic liquid medium as a solvent. *Carbohydrate polymers* **2011**, *85* (4),
45
46 792-797, DOI: 10.1016/j.carbpol.2011.03.047.
47
48
49
50 (15) Gao, J.; Luo, Z.-G.; Luo, F.-X. Ionic liquids as solvents for dissolution of corn starch and
51
52 homogeneous synthesis of fatty-acid starch esters without catalysts. *Carbohydrate polymers* **2012**, *89*
53
54 (4), 1215-1221, DOI: 10.1016/j.carbpol.2012.03.096.
55
56
57
58 (16) Chen, X.; Souvanhthong, B.; Wang, H.; Zheng, H.; Wang, X.; Huo, M. Polyoxometalate-based
59
60

1
2
3
4 ionic liquid as thermoregulated and environmentally friendly catalyst for starch oxidation. *Applied*
5
6 *Catalysis B: Environmental* **2013**, *138*, 161-166, DOI: 10.1016/j.apcatb.2013.02.028.

7
8
9 (17) Chen, X.; Liu, P.; Shang, X.; Xie, F.; Jiang, H.; Wang, J. Investigation of rheological properties
10 and conformation of cassava starch in zinc chloride solution. *Starch -Stärke* **2017**, *69* (9-10), 1600384,
11
12
13
14
15 DOI: 10.1002/star.201600384.

16
17 (18) Jane, J. L. Mechanism of starch gelatinization in neutral salt solutions. *Starch -Stärke* **1993**, *45*
18
19
20
21
22 (5), 161-166, DOI: 10.1002/star.19930450502.

23 (19) Lee, J. H.; You, S.; Kweon, D.-K.; Chung, H.-J.; Lim, S.-T. Dissolution behaviors of waxy maize
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
amylopectin in aqueous-DMSO solutions containing NaCl and CaCl₂. *Food Hydrocolloids* **2014**, *35*,
115-121, DOI: 10.1016/j.foodhyd.2013.05.003.

(20) Lin, M.; Shang, X.; Liu, P.; Xie, F.; Chen, X.; Sun, Y.; Wan, J. Zinc chloride aqueous solution as
a solvent for starch. *Carbohydrate polymers* **2016**, *136*, 266-273, DOI: 10.1016/j.carbpol.2015.09.007.

(21) Liu, P.; Li, Y.; Shang, X.; Xie, F. Starch-zinc complex and its reinforcement effect on starch-based
materials. *Carbohydr Polym* **2019**, *206*, 528-538, DOI: 10.1016/j.carbpol.2018.11.034.

(22) Nicol, T. W. J.; Isobe, N.; Clark, J. H.; Matubayasi, N.; Shimizu, S. The mechanism of salt effects
on starch gelatinization from a statistical thermodynamic perspective. *Food Hydrocolloids* **2019**, *87*,
593-601, DOI: 10.1016/j.foodhyd.2018.08.042.

(23) Liu, H.; Yu, L.; Simon, G.; Zhang, X.; Dean, K.; Chen, L. Effect of annealing and pressure on
microstructure of cornstarches with different amylose/amylopectin ratios. *Carbohydr Res* **2009**, *344*
(3), 350-354, DOI: 10.1016/j.carres.2008.11.014.

(24) Chen, P.; Yu, L.; Kealy, T.; Chen, L.; Li, L. Phase transition of starch granules observed by

1
2
3
4 microscope under shearless and shear conditions. *Carbohydrate Polymers* **2007**, *68* (3), 495-501, DOI:

5
6
7 10.1016/j.carbpol.2006.11.002.

8
9 (25) Mateyawa, S.; Xie, D. F.; Truss, R. W.; Halley, P. J.; Nicholson, T. M.; Shamshina, J. L.; Rogers,

10
11 R. D.; Boehm, M. W.; McNally, T. Effect of the ionic liquid 1-ethyl-3-methylimidazolium acetate on

12
13 the phase transition of starch: dissolution or gelatinization? *Carbohydr Polym* **2013**, *94* (1), 520-530,

14
15
16
17 DOI: /10.1016/j.carbpol.2013.01.024.

18
19 (26) Cheetham, N. W. H.; Tao, L. Variation in crystalline type with amylose content in maize starch

20
21
22 granules: an X-ray powder diffraction study. *Carbohydrate Polymers* **1998**, *36* (4), 277-284, DOI:

23
24
25
26 10.1016/S0144-8617(98)00007-1.

27
28 (27) Lopez-Rubio, A.; Flanagan, B. M.; Gilbert, E. P.; Gidley, M. J. A novel approach for calculating

29
30 starch crystallinity and its correlation with double helix content: a combined XRD and NMR study.

31
32
33 *Biopolymers* **2008**, *89* (9), 761-768, DOI: 10.1002/bip.21005.

34
35 (28) Chen, P.; Xie, F.; Zhao, L.; Qiao, Q.; Liu, X. Effect of acid hydrolysis on the multi-scale structure

36
37
38 change of starch with different amylose content. *Food Hydrocolloids* **2017**, *69*, 359-368, DOI:

39
40
41
42 10.1016/j.foodhyd.2017.03.003.

43
44 (29) Kong, L.; Ziegler, G. R. Molecular encapsulation of ascorbyl palmitate in preformed V-type starch

45
46
47 and amylose. *Carbohydrate Polymers* **2014**, *111*, 256-263, DOI: 10.1016/j.carbpol.2014.04.033.

48
49 (30) Vicentini, N.; Dupuy, N.; Leitzelman, M.; Cereda, M.; Sobral, P. Prediction of cassava starch

50
51
52 edible film properties by chemometric analysis of infrared spectra. *Spectroscopy Letters* **2005**, *38* (6),

53
54
55 749-767, DOI: 10.1080/00387010500316080.

56
57 (31) Weiqing, L.; Tatiana, B. Dissolution of unmodified waxy starch in ionic liquid and solution

58
59
60

1
2
3
4 rheological properties. *Carbohydrate Polymers* **2013**, *93* (1), 199-206, DOI:
5
6 10.1016/j.carbpol.2012.01.090.
7

8
9 (32) Wolf, B. A. Polyelectrolytes Revisited: Reliable Determination of Intrinsic Viscosities.
10
11 *Macromolecular Rapid Communications* **2007**, *28* (2), 164-170, DOI: 10.1002/marc.200600650.
12
13

14 (33) Botelho da Silva, S.; Krolicka, M.; van den Broek, L. A. M.; Frissen, A. E.; Boeriu, C. G. Water-
15
16 soluble chitosan derivatives and pH-responsive hydrogels by selective C-6 oxidation mediated by
17
18 TEMPO-laccase redox system. *Carbohydrate Polymers* **2018**, *186*, 299-309, DOI:
19
20 10.1016/j.carbpol.2018.01.050.
21
22
23

24 (34) Trinh, K. S.; Dang, T. B. Structural, physicochemical, and functional properties of electrolyzed
25
26 cassava starch. *International journal of food science* **2019**, *2019*, 9290627, DOI:
27
28 10.1155/2019/9290627.
29
30
31

32 (35) Xie, F.; Yu, L.; Su, B.; Liu, P.; Wang, J.; Liu, H.; Chen, L. Rheological properties of starches with
33
34 different amylose/amylopectin ratios. *Journal of Cereal Science* **2009**, *49*, 371-377, DOI:
35
36 10.1016/j.jcs.2009.01.002.
37
38
39

40 (36) Xie, F.; Pollet, E.; Halley, P. J.; Averous, L. Starch-based nano-biocomposites. *Progress in*
41
42 *polymer science* **2013**, *38* (10-11), 1590-1628, DOI: 10.1016/j.progpolymsci.2013.05.002.
43
44
45

46 (37) Lancuški, A.; Vasilyev, G.; Putaux, J.-L.; Zussman, E. Rheological properties and
47
48 electrospinnability of high-amylose starch in formic acid. *Biomacromolecules* **2015**, *16* (8), 2529-2536,
49
50 DOI: 10.1021/acs.biomac.5b00817.
51
52

53 (38) Jiang, H.; Campbell, M.; Blanco, M.; Jane, J.-L. Characterization of maize amylose-extender (ae)
54
55 mutant starches: Part II. Structures and properties of starch residues remaining after enzymatic
56
57
58
59
60

1
2
3
4 hydrolysis at boiling-water temperature. *Carbohydrate Polymers* **2010**, *80* (1), 1-12, DOI:
5
6 10.1016/j.carbpol.2009.10.060.
7

8
9 (39) Buléon, A.; Colonna, P.; Planchot, V.; Ball, S. Starch granules: structure and biosynthesis.
10
11 *International journal of biological macromolecules* **1998**, *23* (2), 85-112, DOI: 10.1016/S0141-
12
13 8130(98)00040-3.
14
15

16
17 (40) Chen, P.; Yu, L.; Simon, G. P.; Liu, X.; Dean, K.; Chen, L. Internal structures and phase-transitions
18
19 of starch granules during gelatinization. *Carbohydrate Polymers* **2011**, *83* (4), 1975-1983, DOI:
20
21 10.1016/j.carbpol.2010.11.001.
22
23

24
25 (41) Chen, P.; Yu, L.; Chen, L.; Li, X. X. Morphology and microstructure of maize starches with
26
27 different amylose/amylopectin content. *Starch-Starke* **2006**, *58* (12), 611-615, DOI:
28
29 10.1002/star.200500529.
30
31

32
33 (42) Tyrlik, S. K.; Tomasik, P.; Anderegg, J. W.; Baczkowicz, M. Titanium (IV) starch complexes.
34
35 *Carbohydrate Polymers* **1997**, *34* (1), 1-7, DOI: 10.1016/S0144-8617(97)00102-1.
36
37

38
39 (43) Luo, Z.; Cheng, W.; Chen, H.; Fu, X.; Peng, X.; Luo, F.; Nie, L. Preparation and properties of
40
41 enzyme-modified cassava starch-zinc complexes. *J Agric Food Chem* **2013**, *61* (19), 4631-4638, DOI:
42
43 10.1021/jf4016015.
44
45

46
47 (44) Wei, W.; Wu, S. Depolymerization of cellulose into high-value chemicals by using synergy of
48
49 zinc chloride hydrate and sulfate ion promoted titania catalyst. *Bioresource Technology* **2017**, *241*,
50
51 760-766, DOI: 10.1016/j.biortech.2017.06.004.
52
53

54
55 (45) Szymońska, J.; Molenda, M.; Wiczorek, J. Study of quantitative interactions of potato and corn
56
57 starch granules with ions in diluted solutions of heavy metal salts. *Carbohydrate Polymers* **2015**, *134*,
58
59
60

1
2
3
4 102-109, DOI: 10.1016/j.carbpol.2015.07.041.
5

6
7 (46) Bączkiewicz, M.; Wójtowicz, D.; Anderegg, J. W.; Schilling, C. H.; Tomasik, P. Starch complexes
8
9 with bismuth (III) and (V). *Carbohydrate Polymers* **2003**, *52* (3), 263-268, DOI: 10.1016/S0144-
10
11 8617(02)00294-1.
12

13
14
15 (47) Yang, Y.; Ouyang, R.; Xu, L.; Guo, N.; Li, W.; Feng, K.; Ouyang, L.; Yang, Z.; Zhou, S.; Miao,
16
17 Y. Review: Bismuth complexes: synthesis and applications in biomedicine. *Journal of Coordination*
18
19 *Chemistry* **2015**, *68* (3), 379-397, DOI: 10.1080/00958972.2014.999672.
20
21

22
23 (48) Marusza, K.; Tomasik, P. Aluminium and arsenic(III) starchates. **1994**, *46* (1), 13-17, DOI:
24
25 10.1002/star.19940460105.
26

27
28 (49) Somsook, E.; Hinsin, D.; Buakhrong, P.; Teanchai, R.; Mophan, N.; Pohmakotr, M.; Shiowatana,
29
30 J. Interactions between iron(III) and sucrose, dextran, or starch in complexes. *Carbohydrate Polymers*
31
32 **2005**, *61* (3), 281-287, DOI: 10.1016/j.carbpol.2005.04.019.
33
34

35
36 (50) Jordan, T.; Schmidt, S.; Liebert, T.; Heinze, T. Molten imidazole – A starch solvent. *Green*
37
38 *Chemistry* **2014**, *16* (4), 1967-1973, DOI: 10.1039/C3GC41818A.
39
40

41
42 (51) Sciarini, L.; Rolland-Sabaté, A.; Guilois, S.; Decaen, P.; Leroy, E.; Le Bail, P. Understanding the
43
44 destructuration of starch in water–ionic liquid mixtures. *Green Chemistry* **2015**, *17* (1), 291-299, DOI:
45
46 10.1039/C4GC01248H.
47

48
49 (52) Ji, G.; Luo, Z.; Xiao, Z.; Peng, X. Synthesis of starch nanoparticles in a novel microemulsion with
50
51 two ILs substituting two phases. *Journal of materials science* **2016**, *51* (15), 7085-7092, DOI:
52
53 10.1007/s10853-016-9952-1.
54
55

56
57 (53) Zhang, B.; Chen, L.; Xie, F.; Li, X.; Truss, R. W.; Halley, P. J.; Shamshina, J. L.; Rogers, R. D.;
58
59
60

1
2
3
4 McNally, T. Understanding the structural disorganization of starch in water–ionic liquid solutions.

5
6
7 *Physical Chemistry Chemical Physics* **2015**, *17* (21), 13860-13871, DOI: 10.1039/C5CP01176K.

8
9 (54) Papanyan, Z.; Roth, C.; Wittler, K.; Reimann, S.; Ludwig, R. The dissolution of polyols in salt
10 solutions and ionic liquids at molecular level: Ions, counter ions, and hofmeister effects.
11
12
13
14
15 *ChemPhysChem* **2013**, *14* (16), 3667-3671, DOI: 10.1002/cphc.201300465.

16
17 (55) Ren, F.; Wang, J.; Luan, H.; Yu, J.; Copeland, L.; Wang, S.; Wang, S. Dissolution behavior of
18 maize starch in aqueous ionic liquids: effect of anionic structure and water/ionic liquid ratio. *ACS*
19
20
21
22
23 *Omega* **2019**, *4* (12), 14981-14986, DOI: 10.1021/acsomega.9b01768.

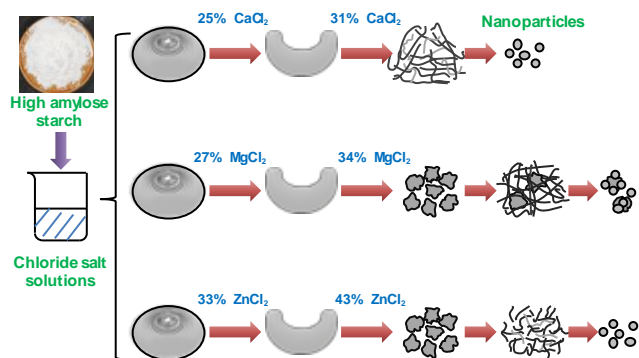
24
25 (56) Shannon, R. D., Revised effective ionic radii and systematic studies of interatomic distances in
26 halides and chalcogenides. *Acta Crystallographica Section A* **1976**, *32* (5), 751-767, DOI:
27
28
29
30
31 10.1107/S0567739476001551.

32
33 (57) Chen, J.; Xie, F.; Li, X.; Chen, L. Ionic liquids for the preparation of biopolymer materials for
34 drug/gene delivery: a review. *Green Chemistry* **2018**, *20* (18), 4169-4200 DOI: 10.1039/C8GC01120F.

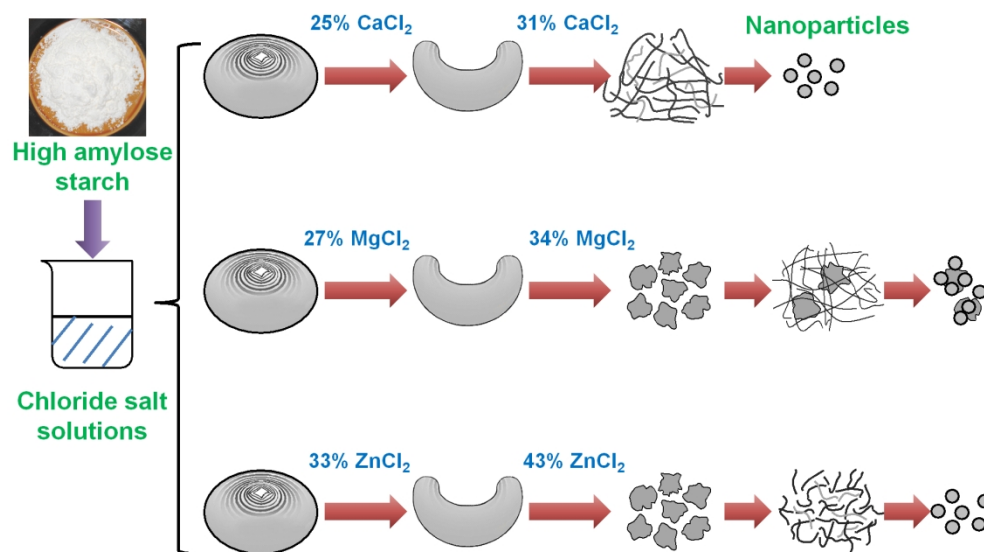
35
36 (58) Dupont, J.; Suarez, P. A. Physico-chemical processes in imidazolium ionic liquids. *Physical*
37
38
39
40
41
42 *Chemistry Chemical Physics* **2006**, *8* (21), 2441-2452, DOI: 10.1039/B602046A.

43
44 (59) Remsing, R. C.; Swatloski, R. P.; Rogers, R. D.; Moyna, G. Mechanism of cellulose dissolution
45 in the ionic liquid 1-n-butyl-3-methylimidazolium chloride: a ^{13}C and $^{35/37}\text{Cl}$ NMR relaxation study on
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60 model systems. *Chemical Communications* **2006**, *2006* (12), 1271-1273, DOI: 10.1039/B600586C.

- For Table of Content Use Only -



A facile method to prepare starch nanoparticles using cheap metal chloride salts is disclosed, leading to advanced applications of biopolymer.



ToC graphic

532x297mm (96 x 96 DPI)

25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60