

Nitrate reductase activity and oxalate content of sugar-beet leaves

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Summary

The nitrate and oxalate content of sugar-beet leaves increases with their ageing, when the plants are grown on nitrate medium.

The present research investigated the influence of high oxalate concentrations on the activity of nitrate reductase. The results showed that oxalate has no substantial effect on the activity of this enzyme.

Introduction

The metabolic conversion of nitrates into organic N gives rise to an equivalent quantity of carboxylates in plant tissues. Leaves of sugar-beet plants, grown on nitrate medium, accumulate considerable amounts of carboxylates, oxalate being the prevailing organic anion. Young leaves have a lower carboxylate and oxalate content than old leaves (van Egmond, 1971). In sugar-beet plants supplied with nitrate van Egmond and Houba (1970) observed a rapid decline in production of organic N, expressed as the quantity of nitrate reduced per gram (dry matter) leaf blade per day. It was suggested that the steadily accumulating oxalate might cause a repression of nitrate metabolism. For this reason a further investigation was made of the effect of varying amounts of oxalate on the activity of the enzyme nitrate reductase (reduced nicotinamide-adenine dinucleotide nitrate oxidoreductase E.C. 1.6.6.1.).

Material and methods

Diploid sugar-beet seeds were germinated in shallow dishes filled with sand moistened with demineralized water. After germination the equal sized seedlings were placed on a well-aerated nutrient solution. The composition of the nutrient solution was the same as used by van Egmond and Houba (1970), containing nitrogen as 6 meq NO_3/l . The nutrient solution was renewed at regular intervals in order to avoid possible nutrient shortages. The plants were grown in a growth chamber. The daily light period was 14 hours, while the temperature was 25° C during the light period, and 17° C during the dark period. The relative humidity of the air was 70 %. The experiments were carried out with plants 4 and 8 weeks old. To determine the enzyme activity, organic anions, total nitrogen and nitrate, samples were taken from comparable leaves of different plants.

A part of a fresh leaf sample was analysed for nitrate reductase activity (NRA), the remainder of the leaf material was dried at 70° C for 24 hours. The dried material was finely ground and analysed for organic anions, nitrate and total nitrogen as described by Houba et al., (1971).

Determination of NRA: 2.00 g of fresh leaf material were cut into small pieces with scissors, and homogenized together with 25 ml of extraction solution. This extraction solution, a sucrose-cysteine-phosphate buffer of pH 7.5, consisted of 1 litre 0.1 M phosphate (0.08 M Na₂HPO₄ + 0.02 M KH₂PO₄) and 0.5 M sucrose + 100 ml 0.1 M cysteine + 10 ml 0.003 M EDTA. For homogenization a cooled homogenizer was used for 30 seconds at 40,000 rev/min. Care was taken to carry out all the preparatory work from weighing until incubation in a cold room at 1° C. A 2-ml aliquot of the homogenate was incubated with 10 ml reagent (6 ml 0.1 M phosphate buffer pH 7.5 + 3 ml reduced NAD solution of 0.5 mg/ml + 1 ml 0.1 M KNO₃) in a centrifuge tube. The nitrate content in the leaf material used in these experiments was so high that omitting extra nitrate ions from the incubation solution had no effect on the amount of nitrite produced. Incubation continued for 30 minutes in a water bath at 30° C. The reaction was stopped with 1 ml 2 M BaCl₂ and the precipitate centrifuged for 10 minutes at 17,000 g. The supernatant was poured into a test tube and mixed with 5 ml 1 % (w/v in 2.5 N HCl) sulphanylamide and 5 ml 0.02% (w/v) N-1-naphthylethylene diamine HCl. After 30 minutes the optical density was measured with a Beckmann Model B spectrophotometer at 540 nm. Standard series (0 to 30 × 10⁻⁵ M NO₂) were prepared from KNO₂. Extraction and assay conditions were in agreement with the conditions described by Sanderson and Cocking (1964), Bowerman and Goodman (1971) and Wallace and Pate (1965).

Experimental

Experiment I concerns the nitrate reductase activity (NRA) in leaves of varying age. Leaves of the same age (leaf number) were taken from 4-week-old sugar-beet plants and mixed to form one sample. NRA was determined in samples of the 6 oldest leaves the leaves younger than Leaf 6 were still unexpanded. Dried samples were analysed for carboxylates, nitrate and total nitrogen.

In Experiment II sampling and analysis were made in the same way, but 8 weeks old plants were used. There were 20 leaves per plant, of which about 14 were fully expanded. This leaf material was used in several experiments. The effect of oxalate on NRA was investigated in two ways: in Experiment IIIa fresh samples of young and old leaves were mixed in varying proportions, the mixed samples were homogenized and NRA was determined; in Experiment IIIb oxalate was added in increasing amounts, corresponding to 0, 1000, 2000, 3000 and 4000 meq oxalate per kg dry matter of the leaf, to a suspension with a relatively high NRA, prepared from leaves of average age. NRA was determined after adding the oxalate.

In Experiment IV, young and old leaves of 8-week-old plants were homogenized and the two homogenates were mixed in varying proportions. NRA was determined in the mixtures.

Results and discussion

The results of Experiment I are given in Fig. 1. Note that when the leaf is older, it contains more nitrate and oxalate, and less organic N, while NRA is lower. A lower rate of nitrate reduction in small, unexpanded, young leaves compared with expanded young

NITRATE REDUCTASE ACTIVITY AND OXALATE CONTENT OF SUGAR-BEET LEAVES

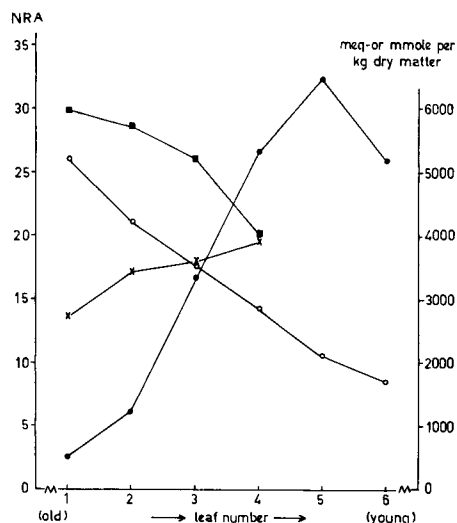


Fig. 1. Nitrate reductase activity, oxalate, organic N and nitrate content of leaves of different age. Experiment I: 4-week-old sugar-beet plants. Left hand ordinate: ● NRA ($\mu\text{mol NO}_2^-$ produced per gram leaf dry matter per hour). Right hand ordinate: ○ oxalate; x organic N; ■ nitrate ($10 \times$). meq or mmol per kg d.m.

leaves is also reported by Martin (1971) in bean plants and by Wallace and Pate (1965, 1967) in peas and cocklebur. Leaf 5 and Leaf 6 were not analysed for total nitrogen and nitrate, because no material was left.

It should be noticed that the fall in NRA with leaf age is relatively much greater than the decline of organic N. This disproportionality means that the decline in NRA with age is not simply due to dilution of the nitrogenous substance in the leaf.

Nitrate reductase is known to be an adaptive enzyme, and if the pool of nitrate within the tissues is depleted, a fall in NRA may be expected. However Fig. 1 shows that all leaves had high nitrate contents, and the older leaves with low NRA contained more nitrate than the younger leaves with high NRA. From this it seems more likely that accumulation of nitrate is promoted by a low NRA.

Table 1 records the content of the various carboxylates in these sugar-beet leaves. From the apex to the base of the plant the carboxylate content of the leaves increases considerably, and oxalate is the prevailing carboxylate, especially in the old bottom leaves. These data and Fig. 1 show that increase in oxalate content and decrease in NRA occur simultaneously.

Table 1. Carboxylate content of leaves of different age in meq/kg dry matter.

Leaf number	Fumarate	Succinate	Malonate	Oxalate	Malate	Citrate	Sum carboxylates
1 old	40	16	44	5236	76	424	5836
2	56	32	52	4220	92	496	4989
3	56	24	36	3552	52	384	4084
4	40	64	28	2878	76	384	3554
5	72	32	28	2128	124	296	2680
6 young	64	40	40	1744	124	300	2312

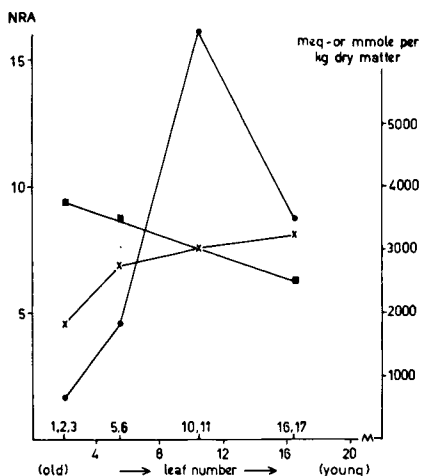


Fig. 2. Nitrate reductase activity, organic N and nitrate content of leaves of different age. Experiment II: 8-week-old sugar-beet plants. Left hand ordinate: ● NRA ($\mu\text{mol NO}_2^-$ produced per gram leaf dry matter per hour). Right hand ordinate: x organic N; ■ nitrate (5 ×); meq or mmol per kg d.m.

The results of Experiment II are summarized in Fig. 2. Although obtained with older plants, the results are similar to those of Experiment I. Again, a drop in NRA is observed when passing from the expanded leaves to the still unexpanded leaves.

Fig. 3 shows the results of NRA determinations on homogenates of mixtures of young and old leaves, as determined in Experiment IIIa. The broken line is calculated from NRA in the unmixed leaf samples of Experiment II and their proportion in the mixture, as the two samples differed in dry matter content the line curves. It is seen that the values of NRA determined in the mixtures are higher than the calculated ones. Since the leaves are large, and the enzyme may not be equally distributed over the leaf, sampling errors may be involved. In Experiment IV this error is avoided.

In Experiment IIIb a homogenate of high NRA was incubated after adding various amounts of potassium oxalate. Fig. 4 shows that NRA fell to some extent, but very little compared with the decline of NRA with leaf age (compare Fig. 2). The leaves used for

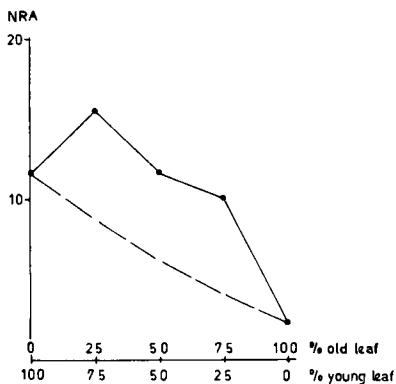


Fig. 3. Nitrate reductase activity ($\mu\text{mol NO}_2^-$ produced per gram leaf dry matter per hour) of mixtures of young and old sugar-beet leaves. Experiment III: 8-week-old plants.

NITRATE REDUCTASE ACTIVITY AND OXALATE CONTENT OF SUGAR-BEET LEAVES

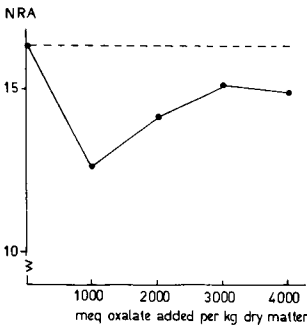


Fig. 4. Nitrate reductase activity ($\mu\text{mol NO}_2^-$ produced per gram leaf dry matter per hour) of a sugar-beet leaf after addition of increasing amounts of oxalate during the incubation, Experiment III; 8-week-old plants.

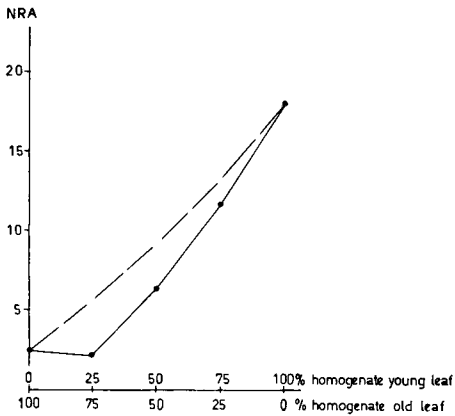


Fig. 5. Nitrate reductase activity ($\mu\text{mol NO}_2^-$ produced per gram leaf dry matter per hour) of mixtures of homogenates of old and young leaves, Experiment IV; 8-week-old plants.

the homogenate contained about 2000 meq oxalate/kg dry matter so that the maximum addition of potassium oxalate raised the actual oxalate concentration to a value comparable with 6000 meq oxalate/ kg dry matter in the leaf. Such a high level of oxalate occurs in the matured bottom leaves of very low NRA, but the addition of extra oxalate had little effect on NRA of the homogenate. From this it follows that the increase in oxalate concentration associated with natural ageing of the leaf does not interfere with the activity of nitrate reductase within the leaf. This is confirmed by the results of Experiment IV given in Fig. 5. Fig. 5 shows that the mixtures of homogenates prepared from young and old leaves respectively had nitrate reductase activities which differed only little from the values calculated from the NRA's of the original homogenates and their proportion in the mixture, as represented by the broken line.

Conclusion

Although a decrease in NRA in sugar-beet leaves occurs simultaneously with an increase in oxalate content of these leaves, the present experiments prove that oxalate as such does not cause a NRA decrease in vitro.

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