Revista peruana de biología 27(1): 061 - 066 (2020) doi: http://dx.doi.org/10.15381/rpb.v27i1.17596 ISSN-L 1561-0837; eISSN: 1727-9933 Universidad Nacional Mayor de San Marcos

Trabajos presentados al *I Congreso Internacional de Biotecnología e innovación (ICBi)*, 9 - 12 de julio de 2018, Universidad Nacional Agraria La Molina, Lima, Perú. Editoras: Ilanit Samolski Klein Maria Lucila Hernández-Macedo Gretty Katherina Villena Chávez

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Citación

Mamani J, Chávez J, Apumayta E, Gil-Kodaka P. 2020. Antioxidant activity and total phenolic content in Caulerpa filiformis (Chlorophyta) from Sechura Bay and Paracas Bay, Peru. I Congreso Internacional de Biotecnología e innovación (ICBi), Revista peruana de biología número especial 27(1): 061- 066 (Marzo 2020). doi: http://dx.doi.org/10.15381/rpb. v27i1.17596

SECCIÓN II: INDUSTRIAL AND ENVIRONMETAL BIOTECHNOLOGY ARTICLE

Actividad antioxidante y contenido fenólico total en Caulerpa filiformis (Chlorophyta) de Bahía de Sechura y Bahía de Paracas, Perú

Antioxidant activity and total phenolic content in Caulerpa filiformis (Chlorophyta) from Sechura Bay and Paracas Bay, Peru

Abstract

In Peru, Caulerpa filiformis is a marine algae listed as an invasive species. For years, its distribution has been considered to be in the north coast (Isla Lobos de Afuera and Piura) until a recent report of its distribution in the central coast (Ancash, Lima, and Ica). The present investigation aims to determine the main groups of secondary metabolites, total phenol content, and antioxidant activity of the methanolic extract of C. filiformis from Sechura Bay (Piura) and Paracas Bay (Ica). The main chemical groups were determined through phytochemical screening, the content of phenols by the Folin-Ciocalteu method, and antioxidant activity by the ABTS method (2,2-azinobis-[3-ethylbenzothiazoline-6-sulfonic acid]) and 2,2'-diphenyl-1-picrylhydrazyl (DPPH). The phytochemical screening of the methanolic extract of C. filiformis from Sechura Bay and Paracas Bay revealed the presence of carbohydrates, polyphenols, tannins, flavonoids, lipids, alkaloids, steroids, and triterpenes for both extracts. The total phenol content of the extract of C. filiformis from Sechura Bay $(39.31 \pm 0.39 \text{ mg of AGE/g extract})$ was significantly higher (p < 0.05) than that from Paracas Bay (18.78 \pm 0.31 mg of AGE/g extract). In the ABTS and DPPH assays, the antioxidant capacity of the Sechura C. filiformis extract (IC50 = 3.49 ± 0.01 and $2.18 \pm 0.02 \text{ mg/mL}$) was significantly higher (p < 0.05) than that of the Paracas C. filiform is extract (IC50 = 6.41 ± 0.02 and 2.42 ± 0.04 mg/mL). These findings suggest that the methanolic extract of C. filiformis is a source of secondary metabolites with an antioxidant potential.

Resumen

En Perú, Caulerpa filiformis es una macroalga catalogada como especie invasora. Durante años, su distribución fue considerada en la costa norte (Isla Lobos de Afuera y Piura) hasta un informe reciente de su distribución en la costa central (Ancash, Lima e Ica). El objetivo de esta investigación es determinar los principales grupos de metabolitos secundarios, contenido total de fenol y actividad antioxidante del extracto metanólico de C. filiformis de Bahía de Sechura (Piura) y Bahía de Paracas (Ica). Los principales grupos químicos se determinaron mediante análisis fitoquímico, el contenido de fenoles mediante el método Folin-Ciocalteu y la actividad antioxidante mediante el método ABTS (ácido 2,2-azinobis- [3-etilbenzotiazolina-6-sulfónico]) y 2, 2'-difenil-1-picrylhydrazyl (DPPH). El examen fitoquímico del extracto metanólico de C. filiformis de ambas bahías revelaron la presencia de carbohidratos, polifenoles, taninos, flavonoides, lípidos, alcaloides, esteroides y triterpenos. El contenido total de fenol del extracto de C. filiformis de Bahía de Sechura (39.31 ± 0.39 mg de extracto de AGE / g) fue significativamente mayor (p <0.05) que el de Bahía de Paracas (18.78 ± 0.31 mg de extracto de AGE / g). En los ensayos ABTS y DPPH, la capacidad antioxidante del extracto de Sechura (IC50 = 3.49 ± 0.01 y 2.18 ± 0.02 mg / mL) fue significativamente mayor (p < 0.05) que la del extracto de Paracas C. *filiformis* (IC50 = 6.41 ± 0.02 y 2.42 ± 0.04 mg / mL). Estos hallazgos sugieren que el extracto metanólico de C. filiformis es una fuente de metabolitos secundarios con potencial antioxidante.

Palabras clave:

Contenido fenólico total; actividad antioxidante; metabolitos secundarios; Caulerpa filiformis; Chlorophyta.

Keywords:

Total phenolic content; antioxidant activity; secondary metabolites; *Caulerpa filiformis*; Chlorophyta.

Journal home page: http://revistasinvestigacion.unmsm.edu.pe/index.php/rpb/index

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Introduction

Macroalgae have been used as important source of highly nutritious food for thousands of years. They have also been used as fodder, fertilizer, and in the field of medicine, mainly in Asian countries (Kolanjinathan et al. 2014). They also produce bioactive compounds, including polyphenols, terpenoids, carotenoids, and tocopherols that possess antibacterial, antiviral, antifungal, antioxidant, anti-inflammatory, antitumor, and allelopathic properties (Michalak & Chojnacka, 2015). In addition, they are important because they have polysaccharides, called phycocolloids that are used in the food industry. These polysaccharides are the agar, carrageenan, and alginates obtained from some species of red (Rhodophyta) and brown (Phaeophyceae) algae.

In the Peruvian coast only few species of macroalgae are used. That is, some red algae species, such as *Chondracanthus chamissoi* "yuyo" and *Pyropia* sp. "cochayuyo" are used for food; *Gracilariopsis lemaneiformis* "pelillo" are exported for the agar industry; and the Phaeophyceae (brown algae) species, such as *Macrocystis* spp. "sargazo" and *Lessonia* spp. "aracanto" are exported for the alginate industry. However, Chlorophyta species (green algae) do not have a commercial interest yet because studies of their chemical compounds are scanty.

Caulerpa filiformis (Chlorophyta) was initially described by Howe (1914) for the north coast (Lobos de Afuera Island and Piura); however, later it was introduced in the central coast (Ancash, Lima, and Ica). *Caulerpa filiformis* is recognized as an invasive species for its rapid range expansion, colonizing new habitats such occurred in Paracas Bay (MINAM 2014; Ramsar 2015). In addition, its high productivity in places such as Paracas Bay causes a large amount of this algae to be stranded on the banks, where it accumulates and decomposes, generating bad odors, pollution, and a landscaping impact (personal observation), especially in spring (Pariona 2018).

Caulerpa have aroused interest worldwide for their secondary metabolites and for some activities useful for pharmaceutical industries, such as those that include antidiabetic (Sharma & Rhyu, 2014), antinociceptive and anti-inflammatory (Matta et al. 2015), antiviral (Nicoletti et al. 1999), antitumor (Cavas et al. 2006), antioxidant (Nguyen et al. 2011), antimicrobial (Vairappan, 2004), and anti-inflammatory (Stirk et al. 2003) properties.

The importance of knowing the antioxidant activity of *C. filiformis* is due to its possible potential as a natural source of antioxidants. Therefore, *C. filiformis* could be considered for use in the treatment of diseases, such as cancer, diabetes, and hypertension, whose pathophysiology is associated with the overproduction of reactive oxygen species (oxidative stress) (Leiva 2000). Currently, *C. filiformis* is not subject to any type of use, and studies on its biochemistry or pharmacological properties are scarce (Egg et al. 2015; Hernández et al. 2015). Therefore, the present study aims to generate knowledge about the main chemical groups (secondary metabolites) and the antioxidant properties of *C. filiformis* from Sechura Bay and Paracas Bay collected in the spring season where the highest biomass has been recorded.

Material and methods

Algae collection.- First, *C. filiformis* was collected from the shallow submareal in two areas of the Peruvian coastline. Blanca Beach in Sechura Bay (5°49'50.8"S; 81°0'21.2"W) at 3 m depth and Atenas Beach in Paracas Bay (13°49'13.5" S, 76°18'1.8" W) at a 1.5 m depth in the spring season (September and October 2017, respectively). The distance among them are 1000 km approximately. Figure 1 shows the sites of collection. The samples were cleaned and washed *in situ* with seawater and transported to the laboratory, where they were immediately washed with potable water to remove excess sand and epiphytic organisms. They were then allowed to drain and dried at 40 °C.

Extract preparation.- The dry samples were subsequently pulverized, followed by the extraction of 5 g of this powder with 100 mL of 99.98% methanol at room temperature for 24 h on a magnetic stirrer. The mixture was centrifuged at 4500 rpm for 15 min at 10 °C, and the obtained supernatant was used in the experiment. The supernatants constituted the methanolic extracts of the *C. filiformis* composition (E_s for Sechura Bay and E_p for Paracas Bay) stored at -20 °C until use.



Figure 1. Sampling sites of Caulerpa filiformis from Sechura Bay and Paracas Bay.

Preliminary Phytochemical Screening.- The methanolic extracts of *C. filiformis* from Sechura Bay (ES) and Paracas Bay (EP) were initially fractionated following the phytochemical guidelines (Rondina and Coussio, 1969) simplified in Figure 2.



Figure 2. Fractionation of the methanolic extract of *C. filiformis* (S = Sechura and P = Paracas).

The typical qualitative reactions of the coloration and the precipitation of the chemical groups were applied for the chemical analysis of the fractions obtained (i.e., FA, FB, FC, and FD).

Determination of the Total Phenol Content.- The total phenol content in the extracts of *C. filiformis* was determined according to the Folin–Ciocalteu test performed by López et al. (2011). Accordingly, 100 μ L of the methanolic extract of *C. filiformis* (standard solution), 8.4 mL of distilled water, and 1 mL of an aqueous solution of Na₂CO₃ (20% w/v) were mixed. Subsequently, 500 μ L of the Folin–Ciocalteu reagent was added. This mixture was kept stirred for 30 min at room temperature in the dark. The absorbance of the reaction mixture was measured at 765 nm using a spectrophotometer (Pharo300-Spectroquant). The results were expressed in mg of a gallic acid equivalent/g extract of the sample (mg AGE/g extract). The presented data corresponded to the average of three measurements.

Antioxidant activity.-

1. **ABTS assay**: The antioxidant activity of the methanolic extract of *C. filiformis* using the radical ABTS (2,2-azinobis-[3-ethylbenzothiazoline-6-sulfonic acid]) was determined according to the method adapted by Arnao *et al.* (2001). The ABTS radical (7.84 mg/mL) was previously activated with potassium persulfate (1.32 mg/mL) in the dark at room temperature for 12 h. The radical ABTS+ solution was obtained as a result of this reaction. This solution was diluted with 80% methanol until an absorbance of 1.1 ± 0.02 measured in a spectrophotometer at 734 nm. Next, 150 µL of the algae extract (standard solution) was reacted with 2,850 µL of the diluted solution of ABTS in a test tube. The mixture was immediately stirred and

allowed to react in the dark at room temperature for 30 min. The absorbance of the reaction mixture was measured in a spectrophotometer at 734 nm. Trolox was used as a positive standard. The antioxidant activity was expressed in percentage of inhibition of the radical **ABTS+**.

2. DPPH assay: The antioxidant activity of the methanolic extract of *C. filiformis* using the DPPH radical (2,2'-diphenyl-1-picrylhydrazyl) was determined according to the method developed by Mensor *et al.* (2001). Accordingly, 0.8 mL of the methanolic solution of DPPH (0.118 mg/mL) was mixed with 2 mL of the seaweed extract (standard solution) in a test tube. The mixture was then immediately stirred and kept at rest for 30 min in the dark. The reaction mixture absorbance was measured in a spectrophotometer at 517 nm. Gallic acid was used as a positive standard. The antioxidant activity was expressed in percentage of inhibition of the DPPH radical.

The following equation was used in both cases:

Inhibition % =
$$\frac{A_C - (A_M - A_{BM})}{A_C} \times 100$$

- A_c = control absorbance
- $A_{_M}$ = absorbance of the reaction of the sample or the standard solution
- A_{BM} = absorbance of the blank sample or a blank of the standard solution

The EC_{50} value (mg/mL) is the effective concentration in which 50% of free radicals are neutralized. This value was calculated using a linear regression, whose value is the average of three measurements.

Statistical Analysis.- The experiments were performed in triplicate, and the results presented corresponded to the average \pm SD. A regression analysis was used to calculate the EC₅₀ value. Statistical comparisons were made using the analysis of variance using Paleontological Statistics program (PAST) version 2.17. The differences were considered significant when the p-value was less than 0.05 (p < 0.05).

Results and Discussion

Photochemical screening.- The general phytochemical study of the *C. filiformis* composition indicated nine groups of chemical compounds for both samples (i.e. Sechura Bay and Paracas Bay) (Table 1). Among these compounds, carbohydrates, substances of a phenolic nature (e.g. flavonoids and tannins), lipids, steroids, triterpenes, and alkaloids highlighting lipids in FA_s and steroids in the FB_s for the Sechura samples were present. According to the studies performed in other *Caulerpa* species, phenolic compounds (i.e., tannins and flavonoids), carbohydrates, terpenoids, and steroids were the common chemical groups present in *Caulerpa* (Karthick et al. 2014; Azhagu Raj et al. 2015).

Table 1. Preliminary phytochemical screening of the extracts of
the composite of Caulerpa filiformis from Sechura Bay and Paracas
Bay. (-): Negative, (+): Mild positive, (++): Moderate positive, and
(+++): Marked positive.

Fraction	Chemical groups	Sechura Bay	Paracas Bay
A	Carbohydrates	++	++
	Flavonoids	+	+
	Lipids	+++	++
	Tannins	++	++
	Phenolic oxidrils	++	++
В	Anthraquinones	-	-
	Steroids	+++	++
	Triterpenes	++	++
	Cardenolides	-	-
С	Alkaloids	++	++
	Cardenolides	-	-
	Leucoanthocyanidins	-	-
D	Quaternary ammonium salts	-	-
	Steroids	-	-
	Triterpenes	-	-
	Flavonoids	-	-

The secondary metabolites produced by *Caulerpa* possess antiviral (Nicoletti et al. 1999), anti-inflammatory (Stirk et al. 2003), antimicrobial (Vairappan, 2004), antitumor (Cavas et al. 2006), and antioxidant (Nguyen et al. 2011) properties. However, the concentration of these compounds can vary according to habitat, collection time, exposure to light, and availability of nutrients. Therefore, the results presented herein could vary if *C. filiformis* were collected at another time of the year.

Total phenol content.- Table 2 presents the yield and content of the total phenols of the methanolic extract of *C. filiformis*. The yield of the methanol extract of *C. filiformis* from Sechura Bay was lower than that obtained from Paracas Bay. However, the total phenolic content of the methanol extract of *C. filiformis* from Sechura Bay (39.31 \pm 0.39 mg AGE/g of extract) was significantly higher (p < 0.05) than that obtained with *C. filiformis* from Paracas Bay (18.78 \pm 0.31 mg AGE/g extract).

Table 2. Yield (% (w/w) of dry seaweed) and total phenol content (CFT) of the methanolic extract of *Caulerpa filiformis* from Sechura Bay and Paracas Bay. The CFT values are the mean \pm standard deviation (SD), n = 3.

Sampling site	Yield (%)	CFT (mg AGE/g extract)
Sechura Bay	8.32	39.31 ± 0.39
Paracas Bay	15.72	18.78 ± 0.31

The results indicated that *C. filiformis* from Sechura Bay produced more than twice the amount of phenolic compounds compared to that from Paracas Bay, with samples obtained at the same time of the year. Similar values of the total phenol content in the range of 23.12 to 38.93 mg AGE/g seaweed extract were found in other *Caulerpa* species, such as *C. racemosa*, *C. peltate*, and *C. taxifolia* in India (Vinayak et al. 2011).

The phenolic compounds with a greater antioxidant activity in plants are flavonoids. These compounds have also been found in some algae. The antioxidant activity is not exclusive to phenolic compounds; other groups of chemical compounds, such as lipids and polysaccharides, also have antioxidant properties (Michalak and Chojnacka, 2015). Recent studies indicated that most marine algae produce chemical compounds with antioxidant activities (Chew et al. 2008), including some *Caulerpa* species, such as *C. lentilifera* and *C. racemosa* (Nguyen et al. 2011; Chia et al. 2015), whose activity is mainly attributed to phenolic compounds.

Antioxidant activity.-

ABTS radical uptake activity.- The antioxidant activity of the methanolic extract of *C. filiformis* from Paracas Bay was 79.60 \pm 0.40%, whereas that from Sechura Bay was 78.49 \pm 0.32%. By contrast, the antioxidant capacity in terms of IC50 (mg/mL) of the methanol extract from Sechura Bay was significantly higher (p < 0.05) compared to that from Paracas Bay, obtaining values of 2.55 \pm 0.01 and 4.62 \pm 0.01, respectively (Table 3). However, the antioxidant capacity of *C. filiformis* from both sites was significantly lower (p < 0.05) compared to the Trolox standard. Therefore, these results indicated that extracts of *C. filiformis* from Sechura Bay have a higher antioxidant capacity compared to those from Paracas Bay, although the latter presented a higher extract yield.

Table 3. Antioxidant activity of the methanolic extract of *Caulerpa filiformis* from Sechura Bay and Paracas Bay determined by the ABTS assay. The values are the average \pm standard deviation (SD); n = 3; and AA = antioxidant activity.

Coursela.	ABTS assay		
Sample	AA ± DS (%)	EC ₅₀ (mg/mL)	
Trolox standard	_*	0.070 ± 0.001	
C. filiformis from Sechura Bay	78.490 ± 0.322	2.546 ± 0.007	
C. filiformis from Paracas Bay	79.599 ± 0.399	4.624 ± 0.014	

*=The test was not performed.

The antioxidant capacity of *Caulerpa* was studied in the form of extracts with an antioxidant potential, which varied in the same or different species (e.g., *C. racemosa* exhibited the capacity of trapping free radicals (ABTS) with an IC50 = 0.709 ± 0.02 mg/mL, greater than that obtained in the analyzed samples). However, similar studies in other algae (i.e. *P. australis* and *S. polycystum*) indicated that brown algae have a greater antioxidant potential compared to *C. racemosa* (Gany et al. 2014).

DPPH radical uptake activity.- The antioxidant activity of the methanolic extract of *C. filiformis* from Paracas Bay was $84.65 \pm 1.56\%$, whereas that from Sechura Bay was $72.53 \pm 1.50\%$. By contrast, the antioxidant capacity in terms of IC50 (mg/mL) of the methanol extract from Sechura Bay was higher compared to that from Paracas

Bay, with values of 2.19 ± 0.02 and 3.22 ± 0.05 , respectively (Table 4). However, the results of the antioxidant activity obtained from both sites of *C. filiformis* were significantly lower (p < 0.05) compared to the standard gallic acid.

Table 4. Antioxidant activity of the methanolic extract of *Caulerpa filiformis* from Sechura Bay and Paracas Bay determined by the DPPH assay. The values are the average \pm standard deviation (SD); n = 3; and AA = antioxidant activity.

Commis	DPPH assay		
Sample	AA ± DS (%)	EC ₅₀ (mg/mL)	
Gallic acid standard	-	0.009 ± 0.001	
C. filiformis de Sechura	72.014 ± 0.601	2.194 ± 0.021	
C. filiformis de Paracas	84.651 ± 2.631	3.218 ± 0.054	

In this assay and in the ABTS assay, the extract of *C. filiformis* from Sechura Bay evidently had a greater antioxidant capacity compared to that from Paracas Bay. Similar studies in *Caulerpa* also showed a greater ability of the extract to trap free radicals (DPPH).

Correlation between antioxidant activity and phenol content.- The antioxidant activity of the macroalgae extracts could be mainly related to the content of phenolic compounds without ruling out the synergistic action between their compounds (Chew et al. 2008; Abdallah et al. 2017). Therefore, the correlation between the antioxidant activity of *C. filiformis* and the phenol content was determined, finding a direct and significant correlation for both the ABTS test (ρ (rho) = 0.641, p = 0.025) and DPPH (ρ) (rho) = 0.625, p = 0.03).

Conclusions.- In the present study, *C. filiformis* from Sechura Bay and Paracas Bay was proven to be an important source of chemical groups, mainly phenolic compounds. A higher extract yield was obtained in *C. filiformis* from Paracas Bay. The total phenolic content of the methanol extract of *C. filiformis* from Sechura Bay was significantly higher compared to that from Paracas Bay. In the ABTS and DPPH assays, the methanolic extract of *C. filiformis* from Paracas Bay had a significantly higher percentage of inhibition of radicals (p < 0.05) compared to that from Sechura Bay. However, the antioxidant capacity (EC₅₀) of the Sechura extract was significantly higher (p < 0.05) than that of Paracas Bay, indicating that the extract of *C. filiformis* from Sechura Bay has a greater antioxidant capacity.

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Agradecimientos / Acknowledgments:

Our thanks to $\ensuremath{\mathsf{Ernesto}}$ Pariona for the support in sampling and for the maps in Figure 1.

Conflicto de intereses / Competing interests:

The authors declare that there is no conflict of interest regarding the publication of this article.

Rol de los autores / Authors Roles:

PGK, JC performed the experimental design. JM, EA performed the experiments. JM Drafted the manuscript. JM, JC, EA, PGK Revised and approved the manuscript

Fuentes de financiamiento / Funding:

This research was funded by FONDECYT 129-2015 and carried out at the Institute of Biochemistry and Molecular Biology at the Universidad Nacional Agraria La Molina, Lima, Peru.

Aspectos éticos / legales; Ethics / legals:

Resolución Jefatural N°11-2016-SERNAMP-RNP/J

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