

REVIEW



The global epidemiology of carbapenemase-producing Enterobacteriaceae

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Carbapenemase-producing Enterobacteriaceae (CPE) are an important and increasing threat to global health. Both clonal spread and plasmid-mediated transmission contribute to the ongoing rise in incidence of these bacteria. Among the 4 classes of β -lactamases defined by the Ambler classification system, the carbapenemases that confer carbapenem resistance in Enterobacteriaceae belong to 3 of them: Class A (K. pneumoniae carbapenemases, KPC), Class B (metallo- β -lactamases, MBL including New Delhi metallo- β -lactamases, NDM) and Class D (OXA-48-like carbapenemases). KPC-producing CPE are the most commonly occurring CPE in the United States. MBL-producing CPE have been most commonly associated with the Indian Subcontinent as well as with specific countries in Europe, including Romania, Denmark, Spain, and Hungary. The epicenter of OXA-48-like-producing is in Turkey and surrounding countries. Detailed knowledge of the epidemiology and molecular characteristics of CPE is essential to stem the spread of these pathogens.

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Introduction

Resistance against antibacterials in clinically relevant bacteria is one of the most imminent threats to public health and especially to our most vulnerable patient populations. The World Health Organization has recognized antimicrobial resistance (AMR) as “a global health security threat that requires action across government sectors and society as a whole.”¹ The Centers for Disease Control and Prevention (CDC) has estimated the excess direct healthcare costs associated with AMR to be as high as \$20 billion, and additional costs to society for lost productivity as high as \$35 billion a year in the United States alone.² Reliable global estimates are needed for the proportion of total infections that are caused by multidrug resistant (MDR) bacteria, and the proportion for each bacterial species isolated from clinical samples that displays an MDR phenotype. However, obtaining the data to calculate these proportions is hampered by incomplete reporting, practice variance in diagnostics and changing definitions for the MDR phenotype. Nonetheless, important progress in this respect has been made by organizations such as various tracking programs led by the CDC (www.cdc.gov), the Centers for Disease Dynamics, Economics and Policy (CDDEP, www.cddep.org), the Antibacterial Resistance Leadership Group (ARLG, www.arlg.org), and in Europe by the European

Centers for Disease Control and Prevention (eCDC, ecdc.europa.eu) and the Combatting Bacterial Resistance in Europe project (COMBACTE, www.combacte.com).

Enterobacteriaceae are a family of bacteria that encompass many bacteria that are commonly isolated from clinical cultures, including *Escherichia coli*, *Klebsiella* spp., and *Enterobacter* spp. From the perspective of antimicrobial resistance, Enterobacteriaceae are especially important as they are a common cause of community-associated as well as healthcare-associated infections.³

In recent years, we have witnessed the emergence of carbapenemase-producing Enterobacteriaceae (CPE). For now, infections caused by CPE in the US are generally healthcare-associated, although community-associated infections are beginning to emerge.^{3,4} The threat of CPE is substantial as carbapenems have traditionally been used in the treatment of infections caused by extended-spectrum β -lactamase-producing Enterobacteriaceae (ESBL-E), and are still considered a last-line of defense against Enterobacteriaceae to date. Few antibiotics retain activity against CPE due to the ability of carbapenemases to hydrolyze most other β -lactam antibiotics as well as the frequent coexistence in CPE isolates of additional mechanisms of resistance against other antibiotic classes such as fluoroquinolones and aminoglycosides. The remaining therapeutic options are less than

desirable secondary to concerns over the lack of efficacy and their toxicity profiles.⁵ In addition, rates of resistance to these agents of last resort such as tigecycline and polymyxins are increasing.^{6–9} Recently, ceftazidime-avibactam has become available and other novel agents with anti-CPE activity are in phase 3 clinical trials.¹⁰ Nonetheless, it is likely that history will repeat itself and resistance against these newer agents will also develop in time.

In this review, we describe the current epidemiology of CPE.

Definitions and terminology

In the past, the field of antibacterial resistance research has been plagued by the lack of standardized definitions for resistance phenotypes. In 2012, Magiorakos and colleagues proposed consensus definitions for MDR, extensively-drug-resistant (XDR) and pandrug-resistant (PDR) bacteria, generated by experts representing the CDC and eCDC.¹¹ Using these criteria, almost all currently encountered CPE would be considered MDR, and a substantial subset of CPE would be considered XDR.

Additional issues with terminology have been introduced by the concurrent use of the terms carbapenem-resistant Enterobacteriaceae (CRE), CPE and carbapenemase-producing carbapenem-resistant Enterobacteriaceae (CP-CRE). Also, the terms carbapenem-resistant organisms (CRO) and carbapenemase-producing organisms (CPO) are occasionally used. The CDC had initially defined CRE as those Enterobacteriaceae which were non-susceptible to ≥ 1 carbapenem and were resistant to 3rd generation cephalosporins. In their November 2015 update, this definition was revised. CRE are now defined as any Enterobacteriaceae which are *resistant* (excluding intermediate resistance) to any carbapenem antimicrobial or are documented to produce a carbapenemase. In addition, for those Enterobacteriaceae which may have intrinsic reduced susceptibility to imipenem such as *Proteus mirabilis*, resistance to a non-imipenem carbapenem is required.¹² The CDC acknowledges that this definition lacks specificity for CPE, especially in low-prevalence areas. The CDC Toolkit therefore recommends that laboratories confirm carbapenemase production by performing molecular testing for the presence of carbapenemases. In this review, we will specifically focus on the epidemiology of CPE.

Horizontal gene transfer and clonal expansion

An important question that remains largely unresolved is whether the main driver of spread of carbapenemases within Enterobacteriaceae is clonal expansion and transmission of successful CPE clonal lineages that stably maintain carbapenemase genes, or whether horizontal

transfer of carbapenemase genes through mobile genetic elements such as plasmids containing them is primarily responsible. The global spread of sequence type (ST) 258 KPC-producing *K. pneumoniae* is an argument in favor of the former.^{13,14} ST258 *K. pneumoniae* is considered a “high risk international clone,” similar to ST131 *E. coli*.¹⁵ In addition, specific clades within ST258 have been associated with carriage of specific *bla*_{KPC} genes: ST258A – corresponding to clade I – is found to be highly associated with *bla*_{KPC-2} whereas ST258B (clade II) tends to carry *bla*_{KPC-3}.¹⁶ This suggests that through recombination events and transference of mobile genetic elements, including transposons and plasmids, *bla*_{KPC-2} and *bla*_{KPC-3} have become associated with specific clones of *K. pneumoniae*, and that this association has remained intact as bacteria spread from one person to the next. These subtypes also differ in the genetic region responsible for capsular polysaccharide biosynthesis.

However, evidence for outbreaks primarily related to horizontal gene transfer has also been reported. In a 5-year single center CPE outbreak investigation, *bla*_{KPC} was found in 66 different strains of Enterobacteriaceae. These 66 strains consisted of 13 different species, including *Klebsiella*, *Enterobacter*, and *Citrobacter*.¹⁷ In addition to person-to-person spread of *bla*_{KPC} carrying bacteria, they found evidence for the transfer of plasmids between various bacteria, as well as for the transfer of *bla*_{KPC} containing transposons between plasmids.¹⁷ In addition, an outbreak investigation from Norway revealed that intergenous spread mediated through a *bla*_{KPC-2} containing plasmid was responsible for transmission of carbapenem resistance from *K. pneumoniae* to *Enterobacter asburiae*.¹⁸

It is clear that there is continuous interplay between bacterial clones and mobile genetic elements that carry resistance genes. In contrast to the close association between ST258 and *bla*_{KPC}, *bla*_{OXA-48} is more closely associated with IncLM type plasmids irrespective of ST type.¹⁵ In conclusion, both plasmid-mediated spread that involves horizontal transmission of resistance genes between bacteria, as well as clonal expansion and transmission likely contribute to the ongoing global CPE epidemic. Evidence is strong that clonal expansion is responsible for a substantial portion of transmitted cases. However, plasmid-mediated transmission is harder to detect and may have been underestimated in reports to date.

Where are CPE found?

Among the 4 classes of β -lactamases defined by the Ambler classification system, the carbapenemases that confer carbapenem resistance in Enterobacteriaceae

belong to 3 of them: Class A, Class B, and Class D.¹⁹ Class A enzymes include the *Klebsiella pneumoniae* carbapenemase (KPC) family, as well as much less commonly encountered nonmetallocarbapenemase type A (NMC-A) and SME enzymes, which may be found in *E. cloacae* and *S. marcescens*, respectively. KPC enzymes are the most commonly encountered enzymatic cause for carbapenem resistance in the US. The closely related genes *bla*_{KPC-2} and *bla*_{KPC-3} account for most of *bla*_{KPC}. Class B enzymes include the metallo- β -lactamases (MBL), such as the New-Delhi-metallo- β -lactamases (NDM), the IMP family of carbapenemases, and the Verona integron-encoded metallo- β -lactamases (VIM). In a zinc-dependent manner, these enzymes hydrolyze a broad variety of β -lactams, but are unable to hydrolyze monobactams such as aztreonam. Class D carbapenemases produced by Enterobacteriaceae include the oxacillinase (OXA)-48-like β -lactamases. OXA-48-like carbapenemases in isolation induce a relatively weak hydrolysis of penicillins and carbapenems but not cephalosporins. As a consequence they may be more difficult to detect, and have been called “the phantom menace.”²⁰ Unfortunately, high level carbapenem resistance may occur when these enzymes are found in combination with other β -lactamases such as ESBL, or with porin changes leading to permeability defects.

Where are KPC-producing CPE found?

Carbapenemases of the KPC family have the most extensive global distribution of all carbapenemases associated with Enterobacteriaceae. The first KPC-producing CPE in the United States was isolated from a patient in North Carolina.²¹ This strain was identified through the Project Intensive Care Antimicrobial Resistance Epidemiology (ICARE) of the CDC, another example of the importance of such surveillance programs.²² This initial report was followed by a large number of cases of KPC-producing CPE reported from New York City area hospitals.¹⁴ In a New York-based multicenter survey, the prevalence of *bla*_{KPC} within *K. pneumoniae* isolates peaked at 36% in 2006.²³ Of great interest, a notable decline has since been recorded to 25% in 2009 and 13% in 2013–2014.²³ Spread to the Great Lakes region of the United States has been described. The Consortium on resistance against carbapenems in *Klebsiella* and other Enterobacteriaceae (CRACKLE) is a prospective, multicenter, observational study of hospitals in Ohio, Pennsylvania and Michigan.¹⁶ Data from CRACKLE showed endemicity of KPC-producing CPE, primarily *K. pneumoniae* of ST258, in the region. The carbapenemase genes responsible for carbapenem resistance were *bla*_{KPC-2} and *bla*_{KPC-3} in more than 90% of all CPE in this study.¹⁶ Patients with CPE

infection and/or colonization have been recognized in both community hospitals as well as tertiary care hospitals in the CRACKLE study.

In the most recent assessment from the CDC, KPC-producing CPE have been reported from every state in the United States except for Maine and Idaho.²⁴ Nonetheless, infections caused by KPC-producing CPE remain a very small subset of all infections caused by Enterobacteriaceae in the US. In a CDC study spanning 2012–2013, the incidence of CRE in 7 US communities was estimated between 0.35 and 4.80 annual incident CRE cases/100,000 population, with an overall estimate of 2.94 annual incident CRE cases/100,000 population.²⁵ However, these data are limited by the small number of CRE isolates that were confirmed to be CPE. Less than a third of CRE isolates were available for testing. Of these 188 isolates, 47% tested positive for a carbapenemase and the only identified carbapenemases were part of the KPC family.²⁵ Detailed data on the incidence of KPC-producing CPE in the US remain scarce.

KPC-producing CPE are also widespread in South and Central America. Several studies have described their epidemiology in Colombia.^{26–28} For example, a recent 2 y (2012–2014) surveillance study in 5 hospitals – sized between 140 and 754 beds – in Medellin revealed that 166 patients had KPC-producing CPE during the study period. Of interest, a high frequency of non-ST258 (62%) was found in this study.²⁹ In a multi-national observational study spanning 7 Latin American nations (Argentina, Colombia, Ecuador, Guatemala, Mexico, Peru and Venezuela), 255 patients with bloodstream infections caused by Enterobacteriaceae were included. Of these patients, 21% had a CPE, which were mostly KPC producers (83%).³⁰ As in most series of CPE infections, *K. pneumoniae* was the most commonly isolated Enterobacteriaceae species. Similar to the US, it is difficult to reliably estimate the exact incidence of CPE infections in various Latin American countries.

In Europe, the highest incidence of KPC-producing CPE is found in Mediterranean countries, especially Italy and Greece.³¹ These 2 countries were the only 2 European countries reported to have an “endemic situation” for KPC in 2014–2015.³¹ While other carbapenemases are present in Italy and Greece, KPC remains the most common etiology of carbapenem resistance.³¹ In a single-center study from Italy that reported on CPKP during 2012–2014, 432 of a total of 436 carbapenemase-producing strains were found to produce KPC.³² In a large, retrospective, multi-center study conducted in 5 Italian hospitals, 661 patients with KPC-producing *K. pneumoniae* were included over a 4 y period. Of these, the majority carried *bla*_{KPC-3} (75%), and the remainder carried *bla*_{KPC-2}.³³ These were selected from a total of

3,449 patients with culture positivity for KPC-producing *K. pneumoniae*. In other words, on average around 170 patients had positive cultures for KPC-producing *K. pneumoniae* per participating hospital per year.³³ The impact of KPC in Greece has been similarly dramatic. In a 10-year single-center study, a large increase in the number of KPC-producing *K. pneumoniae* cases was reported.³⁴ Prior to 2008, no KPC producers were found. From 2008 onwards, the prevalence of KPC producers increased and by 2014 the majority of *K. pneumoniae* isolates carried a *bla*_{KPC} gene.³⁴ In addition, the overall rate of carbapenemase-producing *K. pneumoniae* bloodstream infection (BSI) increased from 0.5 cases per 10,000 patient-days in 2005 to 4.2 cases per 10,000 patient-days, while non-carbapenemase-producing *K. pneumoniae* BSI rates did not decrease.³⁴

In the Middle East, a significant epidemic of KPC-producing CPE has evolved. Of note, a national interventional strategy to contain the spread of CPE within Israel was started in 2007 after a large spike in the number of such cases was detected nationwide. This successful intervention resulted in a dramatic drop of hospital-acquired CPE cases from 55.5 to 4.8 cases/100,000 patient-days.³⁵ Unfortunately, ongoing unrest in the Middle East threatens these advances, as exemplified by the finding of CPE in wounded Syrian patients admitted to northern Israeli hospitals.³⁶

KPC enzymes have also been reported in Asia, especially China. In a study of carbapenem-resistant *E. coli* strains from Shanghai, the majority of CPE (13/16) produced KPC.³⁷ When patients at a Chinese tertiary care hospital were screened for CRE rectal carriage in a 2011–2012 study, 4 of 303 were found to have KPC-producing Enterobacteriaceae.³⁸ In a study from 2011, *bla*_{KPC-2} was present in 71% of 109 ertapenem-resistant *K. pneumoniae* isolates, often in combination with CTX-M type ESBL enzymes.³⁹ The Chinese report of *bla*_{KPC-2} in a hypervirulent K1 strain is especially concerning as it may be the harbinger of the spread of strains that combine hypervirulence with carbapenem resistance.⁴⁰ In contrast, *bla*_{KPC} has been infrequently found in Enterobacteriaceae from patients in India, where NDM-type MBL is the predominant carbapenemase.^{41,42}

Few data are available regarding the epidemiology of KPC in Africa. A single-center study from Tanzania indicated that KPC was an unusual etiology for carbapenem resistance; among 29 carbapenemase-producing *K. pneumoniae*, only 3 were KPC producers.⁴³ KPC producers have also been reported from South Africa.⁴⁴

Where are MBL-producing CPE found?

The substantial burden of carbapenemases in the MBL class appears to lie in Asia. Especially, the New Delhi

Metallo- β -lactamases (NDM) are of concern in this region. The first patient in whom *bla*_{NDM-1} was detected was a Swedish patient who traveled to India in 2007, and acquired a *K. pneumoniae* urinary tract infection. This isolate displayed carbapenem resistance that was mediated through production of a novel carbapenemase designated NDM-1.⁴⁵ A follow-up study showed that NDM-1-mediated carbapenem resistance was widespread in India, Pakistan, and Bangladesh.⁴⁶ Since then, *bla*_{NDM}-positive Enterobacteriaceae have become increasingly common in India. Of note, *bla*_{NDM-1} carrying CPE were not only isolated from patients in India, but importantly also in public tap water and seepage water.⁴⁷ In a recent study from Mumbai, out of 111 CPE, 106 were NDM producers. In addition, 21 NDM producers were found to produce additional carbapenemases as well (17 isolates were positive for OXA-48-type carbapenemase and 4 for VIM-type MBL).⁴¹ Of great concern, they have also been recognized as a source of community-associated infections.⁴⁸ NDM producers have also been described in Enterobacteriaceae in China, but appear to be less widespread as compared to India. In a multicenter study looking at 90 patients with *E. coli* BSI in Shanghai between 2013–2014, no NDM producers were found.⁴⁹ Similarly, in an evaluation of 71 CRE from Hong Kong from 2010–2012, only 9 were confirmed to be CPE. Of these 9 isolates, 3 produced IMP-4, 3 NDM-1 and 3 KPC-2.⁵⁰ A recent worrying observation is the colocalization of *bla*_{NDM-9} and the plasmid-mediated colistin resistance gene *mcr-1* within an *E. coli* strain recovered from chicken meat sold in Guangzhou, China.⁵¹

In Europe, NDM producers are most commonly found in Romania, Poland and Denmark, where “inter-regional spread” (epidemiological stage 4) is deemed to be present.³¹ A large national Polish study from 2012–2014 identified 374 patients with NDM-producing CPE.⁵² Most of these cases were epidemiologically linked and thought to be part of a large multi-regional, multi-center outbreak. Outbreak isolates included *K. pneumoniae* and *E. coli*, and several contained similar transposons.⁵² In Spain, Italy and Hungary, VIM is the predominant MBL; in these countries, “inter-regional spread” (epidemiological stage 4) of VIM producers has been documented.³¹ In addition, contact with healthcare in countries that are endemic for NDM-producers such as India, has been linked to cases presenting in the UK and other European countries.⁵³

In the US and Canada, MBL have remained an uncommon etiology for carbapenem resistance in Enterobacteriaceae.⁵⁴ In 2012, an outbreak of *bla*_{NDM-1} producing *K. pneumoniae* was reported from Denver. Routes of transmission were speculated to include colonized patients who went undetected. The route of introduction into the Denver healthcare system was not established.⁵⁵ As of April 2016, 157 NDM-producing

CPE from 25 states were reported to the CDC.²⁴ A recent outbreak from Illinois of NDM-producing CPE was found to be associated with the use of endoscopes.⁵⁶ VIM-producing and IMP-producing CPE are even less common; per the CDC, only 17 isolates from 7 states; and 10 isolates from 5 states have been reported as of April 2016, respectively.²⁴

MBL-producing Enterobacteriaceae isolates have also been reported from several Latin American countries. In a multinational survey spanning 2012–2014, VIM-producing CPE were recovered from Mexico and NDM-1-producing CPE from Venezuela.⁵⁷ As noted above, KPC enzymes are the most common etiology of carbapenem resistance in Latin America; in the observational multi-national study on CPE BSI in 7 Latin American nations, 9% and 8% of CPE harbored *bla*_{VIM} and *bla*_{NDM}, respectively.³⁰ Similarly, in a multi-center Colombian study of 193 carbapenem-resistant *K. pneumoniae* isolates, only 1 isolates was found to be positive for an MBL gene (*bla*_{VIM}).²⁹ MBL-producing CPE have also been reported from Brazil.⁵⁸ Of concern, *bla*_{NDM-1} was found in the water near Rio de Janeiro in the time period leading up to the 2016 Olympics.⁵⁹

MBL enzymes have been reported in CPE recovered from patients in various African countries for the past decade or so. However, the current magnitude of the MBL-mediated CPE epidemic in Africa is difficult to estimate due to scarcity of data. In a multinational survey spanning 2012–2014, NDM-producing CPE were recovered from Kenya, Nigeria and South Africa and VIM-producing CPE from Nigeria and South Africa.⁵⁷ In an early report from Tunisia, 11 patients admitted in 2005 with VIM-4-producing CPE were described.⁶⁰ A more recent study evaluating bacteria recovered from 2 polluted Tunisian rivers in 2010 documented the frequent occurrence of VIM and IMP enzymes in *K. pneumoniae* isolates.⁶¹ The first recognized occurrence of NDM-1 in South Africa was in 2010.⁶² This first case has been followed by several others from South Africa, raising concerns that NDM-producing CPE are becoming endemic.^{63,64} Risk factors for acquisition of NDM-producing CPE in South Africa were similar to other reports and included comorbid conditions, mechanical ventilation, and prior use of piperacillin-tazobactam.⁶⁴ Of note, both local and imported cases of NDM-1 producing CPE seem to occur in South Africa.⁶³ VIM-1-producing CPE have also been described from South Africa.⁶⁵ In addition, NDM enzymes have been reported from several other African countries including Egypt, Morocco, Algeria, Kenya, Cameroon and Tanzania.^{43,66-70}

Where are OXA-producing CPE found?

OXA-48-like carbapenemases remain extremely rare as a cause of carbapenem resistance in Enterobacteriaceae in

the US.⁷¹ Per CDC data, OXA-48-like-producing CPE were detected only in 43 patients from 19 states as of August 2015.⁷¹ In contrast, they are relatively commonly found in Europe, especially in Mediterranean countries.²⁰ The first reported OXA-48-producing Enterobacteriaceae was a *Klebsiella pneumoniae* strain that was isolated in Turkey in 2001.⁷² OXA-48-like producing *Klebsiella pneumoniae* clones have persisted in Turkey as a cause of nosocomial infections. Turkey was reported as having the highest epidemiologic level (stage 5 “endemic situation”) of these strains in 2014–2015.³¹ In a recent study from Turkey, 92% of CPE were OXA-48-like producers.⁷³ In Spain, France, Belgium and Romania, the epidemiologic stage was deemed “inter-regional spread,” or stage 4, in 2014–2015.³¹ In addition to Europe, OXA-48-like enzymes have been found worldwide in Enterobacteriaceae. Examples of areas with spread of OXA-48-like producing CPE include the Middle East (e.g., United Arab Emirates, Saudi Arabia, Lebanon, Israel), Africa (e.g., Libya, Egypt, Algeria, Morocco, South Africa), Asia (e.g., Russia, India, China, Taiwan, Thailand), and South America (e.g., Argentina, Brazil, Colombia).^{29,74-89} It is important to note that, since most clinical microbiology laboratories do not test for the presence of OXA-48-like enzymes and the associated phenotype (i.e., low-level carbapenem resistance) may be difficult to recognize, the incidence of OXA-48-like-producing CPE is likely underestimated.

Who is at risk for acquiring CPE?

Safdar and Maki outlined a framework for the commonality of risk factors for the acquisition of MDR organisms in their landmark article.⁹⁰ This framework applies to the acquisition of CPE as well. Similar to other MDR bacteria, important risk factors for colonization with CPE include prior antibiotic usage, healthcare and long term care exposure, chronic comorbid conditions, and the presence of invasive catheters and drains.^{16,25,91-95} A potential common pathway for these risk factors – in addition to the obvious increased risk of exposure to other patients and/or healthcare workers who are colonized with CPE – is the disturbance of the microbiome.⁹⁶

Accordingly, in the CRACKLE study patients were elderly; the median age of patients with carbapenem-resistant *K. pneumoniae* was 70 y (interquartile range [IQR] 58–81 years).¹⁶ A slight female predominance was observed; 60% of patients were women. Comorbid conditions were common; 56% had a documented history of diabetes mellitus, 57% had heart disease, 26% of patients carried the diagnosis of renal insufficiency. The median Charlson Comorbidity Index was 4 (IQR 2–6).¹⁶ Similarly, patients with CDC-defined CRE had a median age of 66 (with a range of <1 to 100), 59% were female. In

the CDC study, a lower Charlson Comorbidity Index was noted (a median of 2, with a range of 0–12), perhaps as a consequence of a lower percentage of patients included in the study who had carbapenemase-producing CRE. Still only 9% of patients did not have any underlying condition. Indwelling devices were also common in this CDC study, 75% of patients had a urinary catheter in place, 43% a central venous catheter, and 39% a feeding tube.²⁵ In addition to these traditional risk factors, travel to endemic areas is obviously an important risk factor.⁹⁷ Especially healthcare exposure in endemic areas including “medical tourism” plays an important role in the spread of CPE.⁹⁷

Conclusions

Carbapenemases have a global distribution, but substantial variability exists on the continental, national, regional, and even center-to-center levels. Awareness of the prevalence and incidence of the specific mechanisms of carbapenem resistance within Enterobacteriaceae is crucial in the prevention of their spread and selection of appropriate treatment options.

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No potential conflicts of interest were disclosed.

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References

- [1] Organization WH. Antimicrobial Resistance: Global Report on Surveillance. 2014; <http://www.who.int/drugresistance/documents/surveillancereport/en/>
- [2] Centers for Disease Control and Prevention. ANTIBIOTIC RESISTANCE THREATS in the United States, 2013. 2013; <http://www.cdc.gov/drugresistance/threat-report-2013/>
- [3] van Duin D, Paterson DL. Multidrug-resistant bacteria in the community: Trends and lessons learned. *Infect Dis Clin North Am* 2016; 30:377-90; PMID:27208764; <https://doi.org/10.1016/j.idc.2016.02.004>
- [4] Khatri A, Naeger Murphy N, Wiest P, Osborn M, Garber K, Hecker M, Hurlless K, Rudin SD, Jacobs MR, Kalayjian RC, et al. Community-Acquired Pyelonephritis in Pregnancy Caused by KPC-Producing *Klebsiella pneumoniae*. *Antimicrob Agents Chemother* 2015; 59:4375-8; PMID:26185273; <https://doi.org/10.1128/AAC.00553-15>
- [5] van Duin D, Kaye KS, Neuner EA, Bonomo RA. Carbapenem-resistant Enterobacteriaceae: a review of treatment and outcomes. *Diagn Microbiol Infect Dis* 2013; 75:115-20; PMID:23290507; <https://doi.org/10.1016/j.diagmicrobio.2012.11.009>

- [6] Van Duin D, Cober E, Richter S, Perez F, Cline M, Kaye K, Kalayjian RC, Salata RA, Evans S, Fowler VG, Jr., et al. Tigecycline therapy for carbapenem-resistant *Klebsiella pneumoniae* (CRKP) bacteriuria leads to tigecycline resistance. *Clin Microbiol Infect* 2014; 20:O1117-20; PMID:24931918; <https://doi.org/10.1111/1469-0691.12714>
- [7] van Duin D, Cober E, Richter S, Perez F, Kalayjian RC, Salata RA, Evans S, Fowler VG, Bonomo RA, Kaye KS. Residence in skilled nursing facilities is associated with tigecycline non-susceptibility in carbapenem-resistant *klebsiella pneumoniae*. *Infect Control Hosp Epidemiol* 2015; 36(8); 942-8.
- [8] van Duin D, Doi Y. Outbreak of colistin-resistant, carbapenemase-producing *klebsiella pneumoniae*: Are we at the end of the road? *J Clin Microbiol* 2015; 53:3116-7; PMID:26202122; <https://doi.org/10.1128/JCM.01399-15>
- [9] Liu YY, Wang Y, Walsh TR, Yi LX, Zhang R, Spencer J, Doi Y, Tian G, Dong B, Huang X, et al. Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. *Lancet Infect Dis* 2016; 16:161-8; PMID:26603172; [https://doi.org/10.1016/S1473-3099\(15\)00424-7](https://doi.org/10.1016/S1473-3099(15)00424-7)
- [10] van Duin D, Bonomo RA. Ceftazidime/avibactam and ceftolozane/tazobactam: “Second generation” beta-Lactam/beta-lactamase combinations. *Clin Infect Dis* 2016; 63:234-41; PMID:27098166
- [11] Magiorakos AP, Srinivasan A, Carey RB, Carmeli Y, Falagas ME, Giske CG, Harbarth S, Hindler JF, Kahlmeter G, Olsson-Liljequist B, et al. Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. *Clin Microbiol Infect* 2012; 18:268-81; PMID:21793988; <https://doi.org/10.1111/j.1469-0691.2011.03570.x>
- [12] Centers for Disease Control and Prevention. Facility Guidance for Control of Carbapenem-resistant Enterobacteriaceae (CRE) November 2015. Update - CRE Toolkit 2015; www.cdc.gov
- [13] Kitchel B, Rasheed JK, Patel JB, Srinivasan A, Navon-Venezia S, Carmeli Y, Brolund A, Giske CG. Molecular epidemiology of KPC-producing *Klebsiella pneumoniae* isolates in the United States: clonal expansion of multilocus sequence type 258. *Antimicrob Agents Chemother* 2009; 53:3365-70; PMID:19506063; <https://doi.org/10.1128/AAC.00126-09>
- [14] Munoz-Price LS, Poirel L, Bonomo RA, Schwaber MJ, Daikos GL, Cormican M, Cornaglia G, Garau J, Gniadkowski M, Hayden MK, et al. Clinical epidemiology of the global expansion of *Klebsiella pneumoniae* carbapenemases. *Lancet Infect Dis* 2013; 13:785-96; PMID:23969216; [https://doi.org/10.1016/S1473-3099\(13\)70190-7](https://doi.org/10.1016/S1473-3099(13)70190-7)
- [15] Mathers AJ, Peirano G, Pitout JD. The role of epidemic resistance plasmids and international high-risk clones in the spread of multidrug-resistant Enterobacteriaceae. *Clin Microbiol Rev* 2015; 28:565-91; PMID:25926236; <https://doi.org/10.1128/CMR.00116-14>
- [16] van Duin D, Perez F, Rudin SD, Cober E, Hanrahan J, Ziegler J, Webber R, Fox J, Mason P, Richter SS, et al. Surveillance of carbapenem-resistant *klebsiella pneumoniae*: Tracking molecular epidemiology and outcomes through a regional network. *Antimicrob Agents*

- Chemother 2014; 58:4035-41; PMID:24798270; <https://doi.org/10.1128/AAC.02636-14>
- [17] Sheppard AE, Stoesser N, Wilson DJ, Sebra R, Kasarskis A, Anson LW, Giess A, Pankhurst LJ, Vaughan A, Grim CJ, et al. Nested russian doll-like genetic mobility drives rapid dissemination of the carbapenem resistance gene blaKPC. *Antimicrob Agents Chemother* 2016; 60:3767-78; PMID:27067320; <https://doi.org/10.1128/AAC.00464-16>
- [18] Tofteland S, Naseer U, Lislevand JH, Sundsfjord A, Samuelsen O. A long-term low-frequency hospital outbreak of KPC-producing *Klebsiella pneumoniae* involving Intergenous plasmid diffusion and a persisting environmental reservoir. *PLoS One* 2013; 8:e59015; PMID:23536849; <https://doi.org/10.1371/journal.pone.0059015>
- [19] Nordmann P, Poirel L. Emerging carbapenemases in Gram-negative aerobes. *Clin Microbiol Infect* 2002; 8:321-31; PMID:12084099; <https://doi.org/10.1046/j.1469-0691.2002.00401.x>
- [20] Poirel L, Potron A, Nordmann P. OXA-48-like carbapenemases: the phantom menace. *J Antimicrob Chemother* 2012; 67:1597-606; PMID:22499996; <https://doi.org/10.1093/jac/dks121>
- [21] Yigit H, Queenan AM, Anderson GJ, Domenech-Sanchez A, Biddle JW, Steward CD, Alberti S, Bush K, Tenover FC. Novel carbapenem-hydrolyzing beta-lactamase, KPC-1, from a carbapenem-resistant strain of *Klebsiella pneumoniae*. *Antimicrob Agents Chemother* 2001; 45:1151-61; PMID:11257029; <https://doi.org/10.1128/AAC.45.4.1151-1161.2001>
- [22] Fridkin SK, Steward CD, Edwards JR, Pryor ER, McGowan JE, Jr., Archibald LK, Gaynes RP, Tenover FC. Surveillance of antimicrobial use and antimicrobial resistance in United States hospitals: project ICARE phase 2. Project Intensive Care Antimicrobial Resistance Epidemiology (ICARE) hospitals. *Clin Infect Dis* 1999; 29:245-52; PMID:10476720; <https://doi.org/10.1086/520193>
- [23] Abdallah M, Olafisoye O, Cortes C, Urban C, Landman D, Ghitan M, Collins B, Bratu S, Quale J. Rise and fall of KPC-producing *Klebsiella pneumoniae* in New York City. *J Antimicrob Chemother* 2016; [Epub ahead of print] PMID:27353464; <https://doi.org/10.1093/jac/dkw242>
- [24] Centers for Disease Control and Prevention. Tracking CRE. 2016; <http://www.cdc.gov/hai/organisms/cre/TrackingCRE.html> accessed 5/23/2016
- [25] Guh AY, Bulens SN, Mu Y, Jacob JT, Reno J, Scott J, Wilson LE, Vaeth E, Lynfield R, Shaw KM, et al. Epidemiology of carbapenem-resistant enterobacteriaceae in 7 US communities, 2012-2013. *JAMA* 2015; 314:1479-87; PMID:26436831; <https://doi.org/10.1001/jama.2015.12480>
- [26] Diaz A, Ortiz DC, Trujillo M, Garces C, Jaimes F, Restrepo AV. Clinical characteristics of carbapenem-resistant *klebsiella pneumoniae* infections in ill and colonized children in colombia. *Pediatr Infect Dis J* 2016; 35:237-41; PMID:26569194; <https://doi.org/10.1097/INF.0000000000000987>
- [27] Cuzon G, Naas T, Correa A, Quinn JP, Villegas MV, Nordmann P. Dissemination of the KPC-2 carbapenemase in non-*Klebsiella pneumoniae* enterobacterial isolates from Colombia. *Int J Antimicrob Agents* 2013; 42:59-62; PMID:23664581; <https://doi.org/10.1016/j.ijantimicag.2013.04.002>
- [28] Mojica MF, Correa A, Vargas DA, Maya JJ, Montealegre MC, Rojas LJ, Ruiz SJ, Quinn JP, Villegas MV, Colombian Nosocomial Bacterial Resistance Study G. Molecular correlates of the spread of KPC-producing Enterobacteriaceae in Colombia. *Int J Antimicrob Agents* 2012; 40:277-9; PMID:22789725; <https://doi.org/10.1016/j.ijantimicag.2012.05.006>
- [29] Ocampo AM, Chen L, Cienfuegos AV, Roncancio G, Chavda KD, Kreiswirth BN, Jimenez JN. A two-year surveillance in five colombian tertiary care hospitals reveals high frequency of non-CG258 clones of carbapenem-resistant *klebsiella pneumoniae* with distinct clinical characteristics. *Antimicrob Agents Chemother* 2016; 60:332-42; <https://doi.org/10.1128/AAC.01775-15>
- [30] Villegas MV, Pallares CJ, Escandon-Vargas K, Hernandez-Gomez C, Correa A, Alvarez C, Rosso F, Matta L, Luna C, Zurita J, et al. Characterization and clinical impact of bloodstream infection caused by carbapenemase-producing enterobacteriaceae in seven latin american countries. *PLoS One* 2016; 11:e0154092; PMID:27104910; <https://doi.org/10.1371/journal.pone.0154092>
- [31] Albiger B, Glasner C, Struelens MJ, Grundmann H, Monnet DL, European Survey of Carbapenemase-Producing Enterobacteriaceae working g. Carbapenemase-producing Enterobacteriaceae in Europe: assessment by national experts from 38 countries, May 2015. *Euro Surveill* 2015; 20(45):pii=30062; PMID:26675038; <https://doi.org/10.2807/15607917.ES.2015.20.45.30062>
- [32] Parisi SG, Bartolini A, Santacatterina E, Castellani E, Ghirardo R, Berto A, Franchin E, Menegotto N, De Canale E, Tommasini T, et al. Prevalence of *Klebsiella pneumoniae* strains producing carbapenemases and increase of resistance to colistin in an Italian teaching hospital from January 2012 To December 2014. *BMC Infect Dis* 2015; 15:244; PMID:26116560; <https://doi.org/10.1186/s12879-015-0996-7>
- [33] Tumbarello M, Trecarichi EM, De Rosa FG, Giannella M, Giacobbe DR, Bassetti M, Losito AR, Bartoletti M, Del Bono V, Corcione S, et al. Infections caused by KPC-producing *Klebsiella pneumoniae*: differences in therapy and mortality in a multicentre study. *J Antimicrob Chemother* 2015; 70:2133-43; PMID:25900159; <https://doi.org/10.1093/jac/dkv200>
- [34] Spyropoulou A, Papadimitriou-Olivgeris M, Bartzavali C, Vamvakopoulou S, Marangos M, Spiliopoulou I, Anastassiou ED, Christofidou M. A ten-year surveillance study of carbapenemase-producing *Klebsiella pneumoniae* in a tertiary care Greek university hospital: predominance of KPC- over VIM- or NDM-producing isolates. *J Med Microbiol* 2016; 65:240-6; PMID:26698320; <https://doi.org/10.1099/jmm.0.000217>
- [35] Schwaber MJ, Carmeli Y. An ongoing national intervention to contain the spread of carbapenem-resistant enterobacteriaceae. *Clin Infect Dis* 2014; 58:697-703; PMID:24304707; <https://doi.org/10.1093/cid/cit795>
- [36] Lerner A, Solter E, Rachi E, Adler A, Rechnitzer H, Miron D, Krupnick L, Sela S, Aga E, Ziv Y, et al. Detection and characterization of carbapenemase-producing Enterobacteriaceae in wounded Syrian patients admitted to hospitals in northern Israel. *Eur J Clin Microbiol Infect Dis* 2016; 35:149-54; PMID:26581423; <https://doi.org/10.1007/s10096-015-2520-9>

- [37] Zhang F, Zhu D, Xie L, Guo X, Ni Y, Sun J. Molecular epidemiology of carbapenemase-producing *Escherichia coli* and the prevalence of ST131 subclone H30 in Shanghai, China. *Eur J Clin Microbiol Infect Dis* 2015; 34:1263-9; PMID:25759112; <https://doi.org/10.1007/s10096-015-2356-3>
- [38] Zhao ZC, Xu XH, Liu MB, Wu J, Lin J, Li B. Fecal carriage of carbapenem-resistant Enterobacteriaceae in a Chinese university hospital. *Am J Infect Control* 2014; 42:e61-4; PMID:24773806; <https://doi.org/10.1016/j.ajic.2014.01.024>
- [39] Chen S, Hu F, Xu X, Liu Y, Wu W, Zhu D, Wang H. High prevalence of KPC-2-type carbapenemase coupled with CTX-M-type extended-spectrum beta-lactamases in carbapenem-resistant *Klebsiella pneumoniae* in a teaching hospital in China. *Antimicrob Agents Chemother* 2011; 55:2493-4; PMID:21321140; <https://doi.org/10.1128/AAC.00047-11>
- [40] Wei DD, Wan LG, Deng Q, Liu Y. Emergence of KPC-producing *Klebsiella pneumoniae* hypervirulent clone of capsular serotype K1 that belongs to sequence type 11 in Mainland China. *Diagn Microbiol Infect Dis* 2016; 85:192-4; PMID:27049969; <https://doi.org/10.1016/j.diagmicrobio.2015.03.012>
- [41] Kazi M, Drego L, Nikam C, Ajbani K, Soman R, Shetty A, Rodrigues C. Molecular characterization of carbapenem-resistant Enterobacteriaceae at a tertiary care laboratory in Mumbai. *Eur J Clin Microbiol Infect Dis* 2015; 34:467-72; PMID:25260787; <https://doi.org/10.1007/s10096-014-2249-x>
- [42] Shanmugam P, Meenakshisundaram J, Jayaraman P. blaKPC gene Detection in Clinical Isolates of Carbapenem Resistant Enterobacteriaceae in a Tertiary Care Hospital. *J Clin Diagn Res* 2013; 7:2736-8; PMID:24551626
- [43] Mushi MF, Mshana SE, Imirzalioglu C, Bwanga F. Carbapenemase genes among multidrug resistant gram negative clinical isolates from a tertiary hospital in Mwanza, Tanzania. *BioMed Res Int* 2014; 2014:303104; PMID:24707481; <https://doi.org/10.1155/2014/303104>
- [44] Brink AJ, Coetzee J, Clay CG, Sithole S, Richards GA, Poirer L, Nordmann P. Emergence of New Delhi metallo-beta-lactamase (NDM-1) and *Klebsiella pneumoniae* carbapenemase (KPC-2) in South Africa. *J Clin Microbiol* 2012; 50:525-7; PMID:22116157; <https://doi.org/10.1128/JCM.05956-11>
- [45] Yong D, Toleman MA, Giske CG, Cho HS, Sundman K, Lee K, Walsh TR. Characterization of a new metallo-beta-lactamase gene, bla(NDM-1), and a novel erythromycin esterase gene carried on a unique genetic structure in *Klebsiella pneumoniae* sequence type 14 from India. *Antimicrob Agents Chemother* 2009; 53:5046-54; PMID:19770275; <https://doi.org/10.1128/AAC.00774-09>
- [46] Kumarasamy KK, Toleman MA, Walsh TR, Bagaria J, Butt F, Balakrishnan R, Chaudhary U, Doumith M, Giske CG, Irfan S, et al. Emergence of a new antibiotic resistance mechanism in India, Pakistan, and the UK: a molecular, biological, and epidemiological study. *Lancet Infect Dis* 2010; 10:597-602; PMID:20705517; [https://doi.org/10.1016/S1473-3099\(10\)70143-2](https://doi.org/10.1016/S1473-3099(10)70143-2)
- [47] Walsh TR, Weeks J, Livermore DM, Toleman MA. Dissemination of NDM-1 positive bacteria in the New Delhi environment and its implications for human health: an environmental point prevalence study. *Lancet Infect Dis* 2011; 11:355-62; PMID:21478057; [https://doi.org/10.1016/S1473-3099\(11\)70059-7](https://doi.org/10.1016/S1473-3099(11)70059-7)
- [48] Borah VV, Saikia KK, Chandra P, Hazarika NK, Chakravarty R. New Delhi metallo-beta-lactamase and extended spectrum beta-lactamases co-producing isolates are high in community-acquired urinary infections in Assam as detected by a novel multiplex polymerase chain reaction assay. *Indian J Med Microbiol* 2016; 34:173-82; PMID:27080768; <https://doi.org/10.4103/0255-0857.176853>
- [49] Wang S, Zhao SY, Xiao SZ, Gu FF, Liu QZ, Tang J, Guo XK, Ni YX, Han LZ. Antimicrobial Resistance and Molecular Epidemiology of *Escherichia coli* Causing Bloodstream Infections in Three Hospitals in Shanghai, China. *PLoS One* 2016; 11:e0147740; PMID:26824702; <https://doi.org/10.1371/journal.pone.0147740>
- [50] Ho PL, Cheung YY, Wang Y, Lo WU, Lai EL, Chow KH, Cheng VC. Characterization of carbapenem-resistant *Escherichia coli* and *Klebsiella pneumoniae* from a healthcare region in Hong Kong. *Eur J Clin Microbiol Infect Dis* 2016; 35:379-85; PMID:26740321; <https://doi.org/10.1007/s10096-015-2550-3>
- [51] Yao X, Doi Y, Zeng L, Lv L, Liu JH. Carbapenem-resistant and colistin-resistant *Escherichia coli* co-producing NDM-9 and MCR-1. *Lancet Infect Dis* 2016; 16:288-9; PMID:26842777; [https://doi.org/10.1016/S1473-3099\(16\)00057-8](https://doi.org/10.1016/S1473-3099(16)00057-8)
- [52] Baraniak A, Izdebski R, Fielt J, Gawryszewska I, Bojarska K, Herda M, Literacka E, Zabicka D, Tomczak H, Pewinska N, et al. NDM-producing Enterobacteriaceae in Poland, 2012-14: inter-regional outbreak of *Klebsiella pneumoniae* ST11 and sporadic cases. *J Antimicrob Chemother* 2016; 71:85-91; PMID:26386745; <https://doi.org/10.1093/jac/dkv282>
- [53] Darley E, Weeks J, Jones L, Daniels V, Wootton M, MacGowan A, Walsh T. NDM-1 polymicrobial infections including *Vibrio cholerae*. *Lancet* 2012; 380:1358; PMID:23063285; [https://doi.org/10.1016/S0140-6736\(12\)60911-8](https://doi.org/10.1016/S0140-6736(12)60911-8)
- [54] Rasheed JK, Kitchel B, Zhu W, Anderson KF, Clark NC, Ferraro MJ, Savard P, Humphries RM, Kallen AJ, Limbago BM. New Delhi metallo-beta-lactamase-producing Enterobacteriaceae, United States. *Emerg Infect Dis* 2013; 19:870-8; PMID:23731823; <https://doi.org/10.3201/eid1906.121515>
- [55] Centers for Disease C, Prevention. Notes from the field: hospital outbreak of carbapenem-resistant *Klebsiella pneumoniae* producing New Delhi metallo-beta-lactamase-Denver, Colorado, 2012. *MMWR Morb Mortal Wkly Rep* 2013; 62:108; PMID:23407128
- [56] Centers for Disease C, Prevention. Notes from the Field: New Delhi metallo-beta-lactamase-producing *Escherichia coli* associated with endoscopic retrograde cholangiopancreatography - Illinois, 2013. *MMWR Morb Mortal Wkly Rep* 2014; 62:1051; PMID:24381080
- [57] Kazmierczak KM, Rabine S, Hackel M, McLaughlin RE, Biedenbach DJ, Bouchillon SK, Sahn DF, Bradford PA. Multiyear, Multinational Survey of the Incidence and Global Distribution of Metallo-beta-Lactamase-Producing Enterobacteriaceae and *Pseudomonas aeruginosa*. *Antimicrob*

- Agents Chemother 2015; 60:1067-78; PMID:26643349; <https://doi.org/10.1128/AAC.02379-15>
- [58] Rozales FP, Ribeiro VB, Magagnin CM, Pagano M, Lutz L, Falci DR, Machado A, Barth AL, Zavascki AP. Emergence of NDM-1-producing Enterobacteriaceae in Porto Alegre, Brazil. *Int J Infect Dis* 2014; 25:79-81; PMID:24857802; <https://doi.org/10.1016/j.ijid.2014.01.005>
- [59] de Araujo CF, Silva DM, Carneiro MT, Ribeiro S, Fontana-Maurell M, Alvarez P, Asensi MD, Zahner V, Carvalho-Assef AP. Detection of Carbapenemase Genes in Aquatic Environments in Rio de Janeiro, Brazil. *Antimicrob Agents Chemother* 2016; 60:4380-3; PMID:27139469; <https://doi.org/10.1128/AAC.02753-15>
- [60] Ktari S, Arlet G, Mnif B, Gautier V, Mahjoubi F, Ben Jmeaa M, Bouaziz M, Hammami A. Emergence of multi-drug-resistant *Klebsiella pneumoniae* isolates producing VIM-4 metallo-beta-lactamase, CTX-M-15 extended-spectrum beta-lactamase, and CMY-4 AmpC beta-lactamase in a Tunisian university hospital. *Antimicrob Agents Chemother* 2006; 50:4198-201; PMID:17015633; <https://doi.org/10.1128/AAC.00663-06>
- [61] Chouchani C, Marrakchi R, Henriques I, Correia A. Occurrence of IMP-8, IMP-10, and IMP-13 metallo-beta-lactamases located on class I integrons and other extended-spectrum beta-lactamases in bacterial isolates from Tunisian rivers. *Scand J Infect Dis* 2013; 45:95-103; PMID:22992193; <https://doi.org/10.3109/00365548.2012.717712>
- [62] Lowman W, Sriruttan C, Nana T, Bosman N, Duse A, Venturas J, Clay C, Coetzee J. NDM-1 has arrived: first report of a carbapenem resistance mechanism in South Africa. *S Afr Med J* 2011; 101:873-5; PMID:22273027
- [63] Rubin JE, Peirano G, Peer AK, Govind CN, Pitout JD. NDM-1-producing Enterobacteriaceae from South Africa: moving towards endemicity? *Diagn Microbiol Infect Dis* 2014; 79:378-80; PMID:24853768; <https://doi.org/10.1016/j.diagmicrobio.2014.04.003>
- [64] de Jager P, Chirwa T, Naidoo S, Perovic O, Thomas J. Nosocomial outbreak of New Delhi metallo-beta-lactamase-1-producing gram-negative bacteria in south africa: A case-control study. *PLoS One* 2015; 10:e0123337; PMID:25909482; <https://doi.org/10.1371/journal.pone.0123337>
- [65] Peirano G, Moolman J, Pitondo-Silva A, Pitout JD. The characteristics of VIM-1-producing *Klebsiella pneumoniae* from South Africa. *Scand J Infect Dis* 2012; 44:74-8; PMID:21954935; <https://doi.org/10.3109/00365548.2011.614276>
- [66] Poirel L, Benouda A, Hays C, Nordmann P. Emergence of NDM-1-producing *Klebsiella pneumoniae* in Morocco. *J Antimicrob Chemother* 2011; 66:2781-3; PMID:21930570; <https://doi.org/10.1093/jac/dkr384>
- [67] Sassi A, Loucif L, Gupta SK, Dekhil M, Chettibi H, Rolain JM. NDM-5 carbapenemase-encoding gene in multidrug-resistant clinical isolates of *Escherichia coli* from Algeria. *Antimicrob Agents Chemother* 2014; 58:5606-8; PMID:24982080; <https://doi.org/10.1128/AAC.02818-13>
- [68] Poirel L, Revathi G, Bernabeu S, Nordmann P. Detection of NDM-1-producing *Klebsiella pneumoniae* in Kenya. *Antimicrob Agents Chemother* 2011; 55:934-6; PMID:21115785; <https://doi.org/10.1128/AAC.01247-10>
- [69] Dortet L, Poirel L, Anguel N, Nordmann P. New Delhi metallo-beta-lactamase 4-producing *Escherichia coli* in Cameroon. *Emerg Infect Dis* 2012; 18:1540-2; PMID:22932298; <https://doi.org/10.3201/eid1809.120011>
- [70] Khalifa HO, Soliman AM, Ahmed AM, Shimamoto T, Shimamoto T. NDM-4 and NDM-5-Producing *Klebsiella pneumoniae* Coinfection in a 6-Month-old Infant. *Antimicrob Agents Chemother* 2016; 60:4416-7; PMID:27185797
- [71] Lyman M, Walters M, Lonsway D, Rasheed K, Limbago B, Kallen A. Notes from the field: carbapenem-resistant enterobacteriaceae producing OXA-48-like carbapenemases—United States, 2010–2015. *MMWR Morb Mortal Wkly Rep* 2015; 64:1315-6; PMID:26633574; <https://doi.org/10.15585/mmwr.mm6447a3>
- [72] Poirel L, Heritier C, Tolun V, Nordmann P. Emergence of oxacillinase-mediated resistance to imipenem in *Klebsiella pneumoniae*. *Antimicrob Agents Chemother* 2004; 48:15-22; PMID:14693513; <https://doi.org/10.1128/AAC.48.1.15-22.2004>
- [73] Baran I, Aksu N. Phenotypic and genotypic characteristics of carbapenem-resistant Enterobacteriaceae in a tertiary-level reference hospital in Turkey. *Ann Clin Microbiol Antimicrob* 2016; 15:20; PMID:27048322; <https://doi.org/10.1186/s12941-016-0136-2>
- [74] Lunha K, Chanawong A, Lulitanond A, Wilailuckana C, Charoensri N, Wonglakorn L, Saenjamla P, Chaimanee P, Angkititrakul S, Chetchotisakd P. High-level carbapenem-resistant OXA-48-producing *Klebsiella pneumoniae* with a novel OmpK36 variant and low-level, carbapenem-resistant, non-porin-deficient, OXA-181-producing *Escherichia coli* from Thailand. *Diagn Microbiol Infect Dis* 2016; 85:221-6; PMID:27041106
- [75] Adler A, Hussein O, Ben-David D, Masarwa S, Navon-Venezia S, Schwaber MJ, Carmeli Y. Post-Acute-Care Hospital Carbapenem-Resistant Enterobacteriaceae Working G. Persistence of *Klebsiella pneumoniae* ST258 as the predominant clone of carbapenemase-producing Enterobacteriaceae in post-acute-care hospitals in Israel, 2008–13. *J Antimicrob Chemother* 2015; 70:89-92; PMID:25204343; <https://doi.org/10.1093/jac/dku333>
- [76] Ahn C, Butt AA, Rivera JI, Yaqoob M, Hag S, Khalil A, Pitout M, Doi Y. OXA-48-producing Enterobacteriaceae causing bacteremia, United Arab Emirates. *Int J Infect Dis* 2015; 30:36-7; PMID:25462183; <https://doi.org/10.1016/j.ijid.2014.11.008>
- [77] Hammoudi D, Ayoub Moubareck C, Aires J, Adaime A, Barakat A, Fayad N, Hakime N, Houmani M, Itani T, Najjar Z, et al. Countrywide spread of OXA-48 carbapenemase in Lebanon: surveillance and genetic characterization of carbapenem-non-susceptible Enterobacteriaceae in 10 hospitals over a one-year period. *Int J Infect Dis* 2014; 29:139-44; PMID:25449248; <https://doi.org/10.1016/j.ijid.2014.07.017>
- [78] Poirel L, Abdelaziz MO, Bernabeu S, Nordmann P. Occurrence of OXA-48 and VIM-1 carbapenemase-producing Enterobacteriaceae in Egypt. *Int J Antimicrob Agents* 2013; 41:90-1; PMID:23040010; <https://doi.org/10.1016/j.ijantimicag.2012.08.015>
- [79] Lafeuille E, Decre D, Mahjoub-Messai F, Bidet P, Arlet G, Bingen E. OXA-48 carbapenemase-producing *Klebsiella pneumoniae* isolated from Libyan patients. *Microbial drug resistance* 2013; 19:491-7; PMID:23808959; <https://doi.org/10.1089/mdr.2012.0219>

- [80] Cuzon G, Bentchouala C, Vogel A, Hery M, Lezzar A, Smati F, Dortet L, Naas T. First outbreak of OXA-48-positive carbapenem-resistant *Klebsiella pneumoniae* isolates in Constantine, Algeria. *Int J Antimicrob Agents* 2015; 46:725-7; PMID:26453148; <https://doi.org/10.1016/j.ijantimicag.2015.08.005>
- [81] Barguigua A, Zerouali K, Katty K, El Otmani F, Timinouni M, Elmdaghri N. Occurrence of OXA-48 and NDM-1 carbapenemase-producing *Klebsiella pneumoniae* in a Moroccan university hospital in Casablanca, Morocco. *Infect Genet Evol* 2015; 31:142-8; PMID:25620375; <https://doi.org/10.1016/j.meegid.2015.01.010>
- [82] Lascols C, Hackel M, Marshall SH, Hujer AM, Bouchillon S, Badal R, Hoban D, Bonomo RA. Increasing prevalence and dissemination of NDM-1 metallo-beta-lactamase in India: data from the SMART study (2009). *J Antimicrob Chemother* 2011; 66:1992-7; PMID:21676902; <https://doi.org/10.1093/jac/dkr240>
- [83] Liu Y, Feng Y, Wu W, Xie Y, Wang X, Zhang X, Chen X, Zong Z. First Report of OXA-181-Producing *Escherichia coli* in China and Characterization of the Isolate Using Whole-Genome Sequencing. *Antimicrob Agents Chemother* 2015; 59:5022-5; PMID:26014927; <https://doi.org/10.1128/AAC.00442-15>
- [84] Ma L, Wang JT, Wu TL, Siu LK, Chuang YC, Lin JC, Lu MC, Lu PL. Emergence of OXA-48-Producing *Klebsiella pneumoniae* in Taiwan. *PLoS One* 2015; 10:e0139152; PMID:26414183; <https://doi.org/10.1371/journal.pone.0139152>
- [85] Fursova NK, Astashkin EI, Knyazeva AI, Kartsev NN, Leonova ES, Ershova ON, Alexandrova IA, Kurdyumova NV, Sazikina SY, Volozhantsev NV, et al. The spread of bla OXA-48 and bla OXA-244 carbapenemase genes among *Klebsiella pneumoniae*, *Proteus mirabilis* and *Enterobacter* spp. isolated in Moscow, Russia. *Ann Clin Microbiol Antimicrob* 2015; 14:46; PMID:26526183; <https://doi.org/10.1186/s12941-015-0108-y>
- [86] Al-Agamy MH, Shibl AM, Elkhizzi NA, Meunier D, Turton JF, Livermore DM. Persistence of *Klebsiella pneumoniae* clones with OXA-48 or NDM carbapenemases causing bacteraemias in a Riyadh hospital. *Diagn Microbiol Infect Dis* 2013; 76:214-6; PMID:23518186; <https://doi.org/10.1016/j.diagmicrobio.2013.02.006>
- [87] Poirel L, Castanheira M, Carrer A, Rodriguez CP, Jones RN, Smayevsky J, Nordmann P. OXA-163, an OXA-48-related class D beta-lactamase with extended activity toward expanded-spectrum cephalosporins. *Antimicrob Agents Chemother* 2011; 55:2546-51; PMID:21422200; <https://doi.org/10.1128/AAC.00022-11>
- [88] Pereira PS, Borghi M, de Araujo CF, Aires CA, Oliveira JC, Asensi MD, Carvalho-Assef AP. Clonal dissemination of OXA-370-producing *Klebsiella pneumoniae* in Rio de Janeiro, Brazil. *Antimicrob Agents Chemother* 2015; 59:4453-6; PMID:25987619; <https://doi.org/10.1128/AAC.04243-14>
- [89] Brink AJ, Coetzee J, Corcoran C, Clay CG, Hari-Makkan D, Jacobson RK, Richards GA, Feldman C, Nutt L, van Greune J, et al. Emergence of OXA-48 and OXA-181 carbapenemases among Enterobacteriaceae in South Africa and evidence of in vivo selection of colistin resistance as a consequence of selective decontamination of the gastrointestinal tract. *J Clin Microbiol* 2013; 51:369-72; PMID:23152549; <https://doi.org/10.1128/JCM.02234-12>
- [90] Safdar N, Maki DG. The commonality of risk factors for nosocomial colonization and infection with antimicrobial-resistant *Staphylococcus aureus*, enterococcus, gram-negative bacilli, *Clostridium difficile*, and *Candida*. *Ann Intern Med* 2002; 136:834-44; PMID:12044132; <https://doi.org/10.7326/0003-4819-136-11-200206040-00013>
- [91] Dhar S, Martin ET, Lephart PR, McRoberts JP, Chopra T, Burger TT, Tal-Jasper R, Hayakawa K, Ofer-Friedman H, Lazarovitch T, et al. Risk factors and outcomes for carbapenem-resistant *klebsiella pneumoniae* isolation, stratified by its multilocus sequence typing: ST258 versus non-ST258. *Open Forum Infect Dis* 2016; 3:ofv213; PMID:26885543; <https://doi.org/10.1093/ofid/ofv213>
- [92] Bart Y, Paul M, Eluk O, Geffen Y, Rabino G, Hussein K. Risk factors for recurrence of carbapenem-resistant enterobacteriaceae carriage: case-control study. *Infect Control Hosp Epidemiol* 2015; 36:936-41; PMID:25869912; <https://doi.org/10.1017/ice.2015.82>
- [93] Bhargava A, Hayakawa K, Silverman E, Haider S, Alluri KC, Datla S, Diviti S, Kuchipudi V, Muppavarapu KS, Lephart PR, et al. Risk factors for colonization due to carbapenem-resistant Enterobacteriaceae among patients exposed to long-term acute care and acute care facilities. *Infect Control Hosp Epidemiol* 2014; 35:398-405; PMID:24602945; <https://doi.org/10.1086/675614>
- [94] Falagas ME, Rafailidis PI, Kofteridis D, Vartzili S, Chelvatzoglu FC, Papaioannou V, Maraki S, Samonis G, Michalopoulos A. Risk factors of carbapenem-resistant *Klebsiella pneumoniae* infections: a matched case control study. *J Antimicrob Chemother* 2007; 60:1124-30; PMID:17884829; <https://doi.org/10.1093/jac/dkm356>
- [95] Teo J, Cai Y, Tang S, Lee W, Tan TY, Tan TT, Kwa AL. Risk factors, molecular epidemiology and outcomes of ertapenem-resistant, carbapenem-susceptible Enterobacteriaceae: a case-case-control study. *PLoS One* 7:e34254; PMID:22461908; <https://doi.org/10.1371/journal.pone.0034254>
- [96] Caballero S, Carter R, Ke X, Susac B, Leiner IM, Kim GJ, Miller L, Ling L, Manova K, Pamer EG. Distinct but spatially overlapping intestinal niches for vancomycin-resistant enterococcus faecium and carbapenem-resistant *klebsiella pneumoniae*. *PLoS Pathog* 2015; 11:e1005132; PMID:26334306; <https://doi.org/10.1371/journal.ppat.1005132>
- [97] van der Bij AK, Pitout JD. The role of international travel in the worldwide spread of multiresistant Enterobacteriaceae. *J Antimicrob Chemother* 2012; 67:2090-100; PMID:22678728; <https://doi.org/10.1093/jac/dks214>