



Review

Current and Future Theranostic Applications of the Lipid-Calcium-Phosphate Nanoparticle Platform

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Received: 2015.12.14; Accepted: 2016.03.15; Published: 2016.04.27

Abstract

Over the last four years, the Lipid-Calcium-Phosphate (LCP) nanoparticle platform has shown success in a wide range of treatment strategies, recently including theranostics. The high specific drug loading of radiometals into LCP, coupled with its ability to efficiently encapsulate many types of cytotoxic agents, allows a broad range of theranostic applications, many of which are yet unexplored. In addition to providing an overview of current medical imaging modalities, this review highlights the current theranostic applications for LCP using SPECT and PET, and discusses potential future uses of the platform by comparing it with both systemically and locally delivered clinical radiotherapy options as well as introducing its applications as an MRI contrast agent. Strengths and weaknesses of LCP and of nanoparticles in general are discussed, as well as caveats regarding the use of fluorescence to determine the accumulation or biodistribution of a probe.

Key words: Cancer, Theranostic, SPECT, PET, SIRT, Nanoparticle, LCP.

Introduction

The fields of tumor imaging and cancer therapy have been steadily sliding closer together since the field recognized a need for targeted therapeutics. Targeted delivery must be coupled with verification of payload deposition in order to effectively monitor and analyze therapeutic outcomes. A wealth of theranostic agents have been developed to address the growing desire for this dual-purpose paradigm, and nanoparticle formulations have recently come to the forefront of theranostic research^{1,2}.

Calcium phosphate precipitates have been used as *in vitro* transfection agents since the 1970s³, when it was discovered that DNA could be co-precipitated in a solution containing calcium chloride and phosphate-buffered saline. Since that time, there has been a substantial effort to control particle size and aggregation in order to increase transfection efficiency and allow delivery *in vivo*. Some formulation strategies gained moderate success *in vitro*⁴⁻⁶, but it was not until 2010 that calcium phosphate

nanoparticles were successfully coated with a lipid bilayer⁷, and not until 2012 that these so-called Lipid-Calcium-Phosphate (LCP) nanoparticles showed efficient gene silencing *in vivo*⁸. Since that time, the two seminal papers have garnered nearly 300 citations, inspiring many other laboratories to use LCP themselves or innovate on its design⁹⁻¹⁴. The Huang lab has published nearly 20 original research articles using LCP, due to its versatility in encapsulating many types of therapeutic compounds, such as small molecule drugs¹⁵⁻¹⁷, siRNA¹⁸⁻²², DNA²³, peptides²⁴, and radionuclides^{25, 26}. The principles behind LCP formulation have also allowed the Huang lab to formulate lipid-coated nanoparticles with cores entirely made of cisplatin^{27, 28}. Over the last six years, LCP has transitioned from a novel formulation to a powerful tool used to probe for answers regarding PEGylation kinetics²⁹, the tumor microenvironment^{15, 18, 24, 25}, cancer vaccines^{18, 24}, and gene therapy²³. The Huang lab has now begun using LCP as a theranostic

agent, using imaging modalities such as SPECT and PET.

LCP is formulated using a reverse (water-in-oil) microemulsion system. In separate round-bottom flasks, water droplets containing either calcium or phosphate are stirred in an oil phase containing cyclohexane and the surfactant igepal. The surfactant maintains the oil-water interface and stabilizes nano-sized water droplets in the oil. When the two emulsions are mixed and the separate droplets collide, the soluble calcium and phosphate react to form nanoprecipitates, still suspended in the water droplets. Addition of the phospholipid DOPA soon after mixing allows the formation of a single lipid layer at the oil-water interface in which the hydrophilic head of DOPA inserts inside the water droplet and its hydrophobic tail remains in the oil phase. The phosphate head group of the DOPA co-precipitates into the calcium-phosphate nanoprecipitate, positioning the hydrophobic tail to remain in the oil. Over time, the co-precipitated DOPA replaces more and more igepal in the water-oil interface, and eventually the calcium phosphate precipitates are so well coated with DOPA that they are able to be dissolved in organic solvents such as chloroform or dichloromethane after washing with ethanol. The final LCP nanoparticle, shown in Figure 1, is generated by adding free lipids such as DOTAP,

cholesterol, and DSPE-PEG2000, also in organic solvent, to the cores. The solvent is then evaporated to leave a thin film of lipids and cores on the edge of the vial. Addition of warm water/sucrose solution and brief sonication is all that is needed to resuspend the final particles for testing or injection into mice.

Encapsulation of therapeutics into LCP occurs during the calcium-phosphate nanoprecipitation portion of the procedure. Phosphorylated prodrugs, nucleotide mimics, siRNA, and DNA are all ideal candidates for encapsulation into LCP because the phosphate groups on all of these molecules will co-precipitate in the calcium-phosphate LCP core. Radionuclides such as indium and lutetium can similarly be encapsulated via their low solubility with phosphate. Because calcium-phosphate is acid-sensitive, after the LCP is delivered to the cellular endosome, the late endosome's acidic environment will dissolve the calcium-phosphate core⁷. The increase in endosomal salt concentration increases the osmolarity in the endosome, destabilizing it and releasing the therapeutics into the cytosol. The cell avoids calcium-induced toxicity by quickly lowering cytosolic calcium via the mitochondrial calcium uniporter and then by more slowly removing the excess intracellular calcium via the plasma membrane calcium ATPase³⁰.

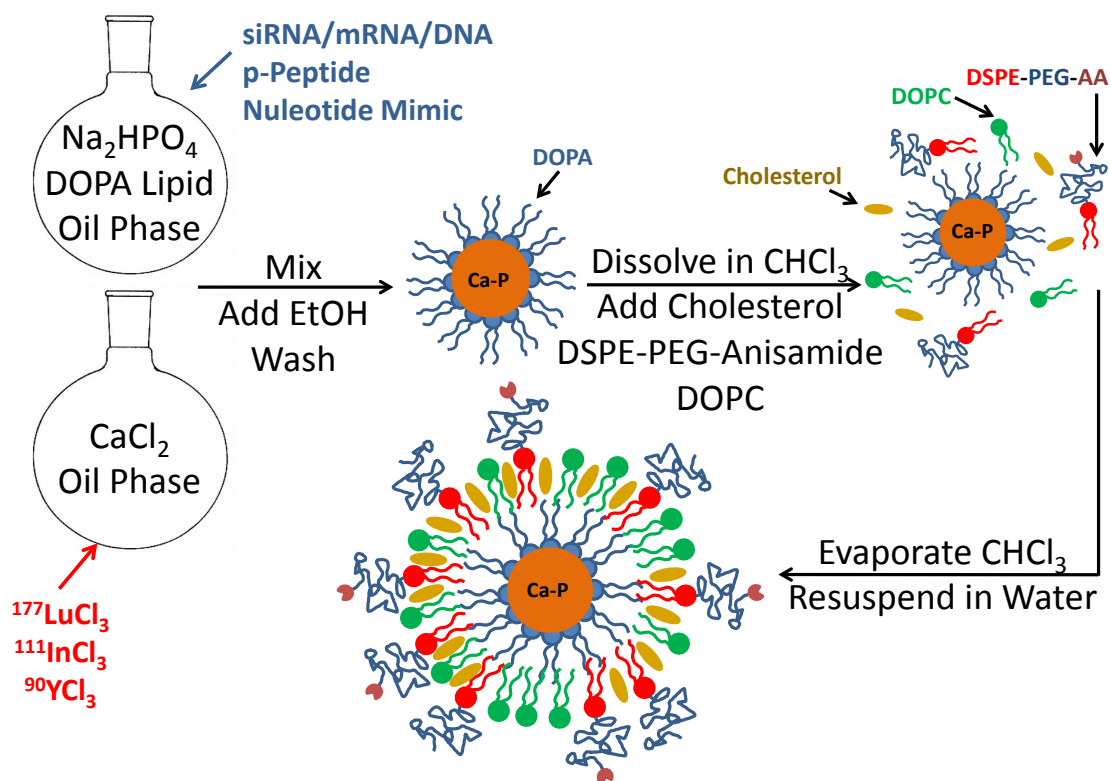


Figure 1: Schematic of LCP nanoparticle synthesis. LCP is capable of encapsulating siRNA, mRNA, DNA, phosphorylated peptides, nucleotide mimics, and radioactive isotopes in its calcium phosphate core. Source: Adapted from Satterlee et al²⁵. Copyright © 2015 Elsevier B.V. License number: 3815560646760

Herein we aim to review current imaging techniques and compare the theranostic capabilities of LCP to clinical practices that combine therapy and imaging. This review will focus its discussion on the nuclear imaging modalities SPECT and PET.

Single Photon Emission Computed Tomography (SPECT)

SPECT imaging detects γ photon emissions from a decaying radioactive source inside the body³¹. Radionuclides with a high abundance of γ decay at energies around 100-200 keV³², such as ¹¹¹In, ^{99m}Tc, and ¹²³I, are ideal for detection using SPECT, as their γ energies are high enough to avoid attenuation in tissue but low enough to be adequately captured by the SPECT detectors. Because the source emits photons in all directions, in order to maximize spatial resolution, a collimator must be used to block all photons that are not approaching the detector from the region or angle of interest. More selective collimators increase the resolution of the image but also reduce signal to the detectors, requiring longer detection times or higher doses of radionuclide to maintain adequate image reconstruction³³. SPECT is often run in combination with a computed tomography (CT) scan to provide anatomical contrast.

Positron Emission Tomography (PET)

PET imaging is similar to SPECT imaging, as the PET detectors also detect γ photons from a radioactive source inside the body³². In contrast to SPECT, the photons detected in PET are generated from an annihilation reaction between a positron emitted by the source and a nearby electron present in the body. The annihilation reaction converts the electron-positron pair into 511 keV photons that travel in opposite directions toward the PET detector. By only counting photon pairs that are detected at a 180° angle from each other, all other noise not attributed to positron emission can be disregarded, eliminating the need for collimation. By also calculating the small differences in detector arrival time of the photon pair—referred to as time-of-flight—, more accurate spatial localization of the radionuclide can be determined³⁴. The unique imaging properties of positron emitters allow PET imaging to be more sensitive than SPECT, but PET must also be coupled with an imaging modality like CT or MRI for anatomical contrast.

Comparing two common PET agents, copper-64 (⁶⁴Cu) and gallium-68 (⁶⁸Ga), will demonstrate the additional considerations that must be made when planning a PET imaging study. Half-life must first be considered, as many PET agents have very short half-lives and must be produced and purified on-site.

⁶⁴Cu has a relatively long half-life of 12.7 h, allowing it to be shipped overnight without decaying prohibitively, unlike ⁶⁸Ga which has a half-life of just 68 minutes. Fortunately, ⁶⁸Ga is generated by the slow decay of Germanium-68 (⁶⁸Ge) and can be quickly purified from its parent on-site³⁵, unlike ⁶⁴Cu which must be generated by neutron or proton bombardment in a nuclear reactor or cyclotron³⁶. Half-life also affects the total dose of nuclide that must be given to achieve the same decay rate during imaging. It is desirable to inject a low total dose with a high rate of decay such that a large portion of the decay will occur during the imaging experiment. In a one-hour PET imaging session using ⁶⁸Ga, half of the remaining ⁶⁸Ga atoms will decay during the imaging experiment, but if ⁶⁴Cu is used, less than 10% of the ⁶⁴Cu will decay during the experiment, leaving the rest to decay over the next few days. In order to produce the same quality image, both ⁶⁴Cu and ⁶⁸Ga need to be dosed at similar positron decay rates, and since ⁶⁸Ga decays more quickly, fewer total atoms also need to be given, further lowering the required dose.

The energy with which a positron is emitted, $\beta^+_{\max} = 653$ keV for ⁶⁴Cu and $\beta^+_{\max} = 1899$ keV for ⁶⁸Ga, also affects the resolution of the image. A positron must be at rest energy before annihilating with an electron, and any extra energy is released as the positron proceeds away from its parent nucleus and collides with other matter³⁷. Positrons ejected at higher energies must release more energy before annihilation and therefore must travel further away from the nucleus before the resulting two γ photons are generated. Because the PET detectors can only detect where in space the photons were generated, and not where the positron was emitted, a longer positron range will generate larger areas between the positron emitter and the photon generation, thus decreasing the spatial resolution of the image. The average positron ranges in water for ⁶⁴Cu and ⁶⁸Ga are 0.55 mm and 2.8 mm, respectively³⁸, which is not a negligible difference and should be considered when planning an experiment.

Clinical Nuclear Imaging

In the clinic, SPECT and PET contrast agents are often used diagnostically for blood vessel perfusion imaging or tumor diagnosis. Radiometals have recently seen more use in the clinic, and are often chelated to small molecules or targeted proteins. A huge amount of effort has gone into discovering bifunctional chelators that stably bind both the radiometal and the tumor-targeting moiety³⁹⁻⁴³. Because each radiometal ion can differ in atomic size, oxidation state, and electron configuration, a chelator that stably binds one metal may not be suitable for

another. Stability tests must include exposing the chelated metal to serum conditions, as an inadequate chelator may have higher affinity for an endogenous metal ion or allow reduction of its metal into a less stable state⁴⁴. The expected circulation time of the radionuclide must also be considered, as a nuclide chelated to a protein with a circulation half-life on the order of days would need a much more stable chelator than one on a particle with a half-life of an hour or less. If an unsuitable chelator is chosen to bind a radionuclide, the nuclide may prematurely shed in circulation and skew the biodistribution data, as highly charged heavy metals such as ¹¹¹In(III) or ¹⁷⁷Lu(III) prefer to accumulate in the bone, and actively metabolized metals such as copper may be filtered through the liver and into the intestines.

There are several typical radioisotopes used diagnostically as SPECT and PET contrast agents. Current clinical uses involve chelation or conjugation of the isotopes to small molecules for blood vessel perfusion imaging or tumor diagnosis. Classically, the γ emitter ^{99m}Tc has been injected intravenously into patients after chelation to one of several small molecules such as exametazime and sestamibi⁴⁵. The lipophilic conjugate formed can accumulate in tumors and allow detection of many types of cancer via SPECT. In several clinical trials, ^{99m}Tc has also been conjugated to the apoptosis marker Annexin V to gauge a patient's response to treatment⁴⁶. A similar agent exists for PET imaging. Incorporation of the positron emitter ¹⁸F as the 2' group on glucose molecules yields ¹⁸Fluoro-deoxyglucose (¹⁸FDG), which can accumulate in tumors due to a tumor's high glucose uptake and the inability for cells to metabolize FDG after uptake. ¹¹¹In is also currently used as a diagnostic agent to measure tumor burden or metastasis by chelating it to an antibody targeted to an overexpressed tumor receptor⁴⁷. The clinically used ProstaScint® is an ¹¹¹In-conjugated monoclonal antibody against prostate specific membrane antigen used to diagnose prostate cancer. In some cases, such as in the phase II clinical trial for a doxorubicin-linked polymeric micelle termed PK1, pharmacokinetics and biodistribution of the micelle were determined by conjugating the γ emitter ¹²³I onto a small amount of PK1 analog during one treatment cycle⁴⁸. The imaging analog contained an additional methacryloyltyrosinamide group whose tyrosine moiety was labeled with ¹²³I via the Iodogen® labeling procedure⁴⁹. These imaging strategies have laid the groundwork for those therapeutics that can simultaneously image and treat the tumor.

Radioimmunotherapy

Another recent effort has been placed on

formulations that can accumulate in the tumor to provide diagnosis while simultaneously treating the tumor mass, and so far the most successful radio-theranostics in the clinic seem to be those used in radioimmunotherapy (RIT). RIT is a treatment strategy in which a therapeutic radioisotope such as ⁹⁰Y or ¹⁷⁷Lu is chelated and covalently bound to an antibody that will selectively target the tumor. ⁹⁰Y has a higher β energy and a longer β range in tissue, while ¹⁷⁷Lu decays via both β and γ emission, allowing SPECT imaging with simultaneous therapy. The radioimmunotherapeutic Zevalin®, which uses ⁹⁰Y, was the first in its class to receive FDA approval and is currently seeing success in the clinic⁵⁰. Zevalin® is used against Non-Hodgkin's Lymphoma (NHL) and uses the murine anti-CD20 antibody Ibritumomab against B-cells as a targeting and therapeutic agent. Unfortunately, the antibodies used in this therapy are known to cause sometimes serious toxicity via complement activation and subsequent cytokine release⁵¹⁻⁵³. Antibodies used in RIT are also very expensive, especially those that have been humanized or otherwise modified to reduce toxicity.

An effective alternative to RIT may be to use a peptide instead of an antibody to target the chelated radionuclide to the tumor, which would be less toxic and less expensive. This approach has been termed Peptide Receptor Radionuclide Therapy (PRRT) and the most successful designs have used a somatostatin analog called octreotate to target the somatostatin receptor overexpressed in neuroendocrine tumors⁵⁴⁻⁵⁶. The novel molecule DOTA-TATE (Figure 2) was generated by covalently linking the chelator DOTA to octreotate, allowing chelation of radionuclides such as ¹⁷⁷Lu, ⁹⁰Y, ⁶⁴Cu, and ¹¹¹In for imaging and therapy. PRRT with octreotate was recently approved by the FDA for use in patients with neuroendocrine cancer. The dual therapy (β) and imaging (γ) capabilities of ¹⁷⁷Lu allow simultaneous treatment and dosimetry of patients receiving ¹⁷⁷Lu-DOTA-TATE⁵⁷. The absorbed dose to healthy organs, especially the kidneys and bone marrow, was calculated from γ -scintigraphy and SPECT/CT images in order to provide the maximum therapeutic dose without exceeding the maximum acceptable dose to healthy tissue.

SPECT Imaging with LCP

The LCP formulation procedure allows efficient encapsulation of radiometals without the use of a chelator, and as of 2015, two articles have been published using LCP for SPECT imaging. The first encapsulated the γ emitter ¹¹¹In³⁺ into the core of LCP as an imaging agent to measure lymph node accumulation of LCP, initiating research on treating lymph node metastasis²⁶. The second article

encapsulated the γ and β emitter $^{177}\text{Lu}^{3+}$ as both an imaging and therapeutic agent²⁵. It was discovered that the 3^+ charge on radiometals like ^{177}Lu enabled very selective encapsulation in the LCP core, as their solubility product with phosphate are many orders of magnitude lower than that of calcium and phosphate (k_{sp} lutetium-phosphate = 2×10^{-25} ; k_{sp} calcium-phosphate = 1×10^{-7}). Consequently, when ^{177}Lu was co-precipitated into the LCP core, a 70% encapsulation efficiency of ^{177}Lu was maintained as the feed ratio of lutetium to calcium was increased by four orders of magnitude, allowing a tunable loading of radionuclide in a single-sized batch of LCP. Because only a trace amount of ^{177}Lu is required to provide therapeutic effect, small-scale batches are still sufficient to produce ^{177}Lu -LCP particles at human-level therapeutic dosing. Even at the highest stable feed ratio of Lu:Ca, ^{177}Lu -LCP maintains its encapsulation efficiency and morphology in particles that are still overwhelmingly composed of calcium phosphate (1:1000 Lu:Ca input ratio). This eliminates the risk of heavy metal toxicity that would be presented in neutron-activatable theranostic nanoparticles, which can be loaded with as much as 20 wt% of the heavy metal ion⁵⁸. In this case, the nanoparticles are bombarded with neutrons in a nuclear reactor to produce radioactive ^{166}Ho from stable ^{165}Ho . Only a small fraction of the ^{165}Ho is converted, increasing the dose of nanoparticles and heavy metal that must be given, but this method does decrease radioactive handling during fabrication.

^{177}Lu has a max β energy of ~ 500 keV and γ energies of 113 and 208 keV. The γ energies are in the ideal range for efficient SPECT imaging and allow the biodistribution and tumor accumulation of ^{177}Lu -LCP to be measured using SPECT/CT (Figure 3A) and γ scintillation²⁵. The therapeutic β emission from ^{177}Lu -LCP was found to cause significant tumor inhibition via DNA damage-induced H2AX phosphorylation, followed by cellular apoptosis. A change in the tumor microenvironment's stromal structure was also detected, resulting in a decrease in fibroblast and collagen organization. Furthermore, the cytotoxic β emission from ^{177}Lu also induced photons in the visible spectrum from the surrounding media. This Cerenkov radiation is generated from the β electron moving faster than the speed of light in tissue and was detected using optical imaging in a live mouse (Figure 3B). Although Cerenkov imaging does not provide a 3D reconstruction of biodistribution as SPECT does, it does allow less expensive and faster image acquisition that would be especially useful in determining early biodistribution kinetics using multiple scans in quick succession. Cerenkov radiation also allows imaging of other β -emitting

nuclides such as ^{90}Y that would otherwise not be available for imaging studies.

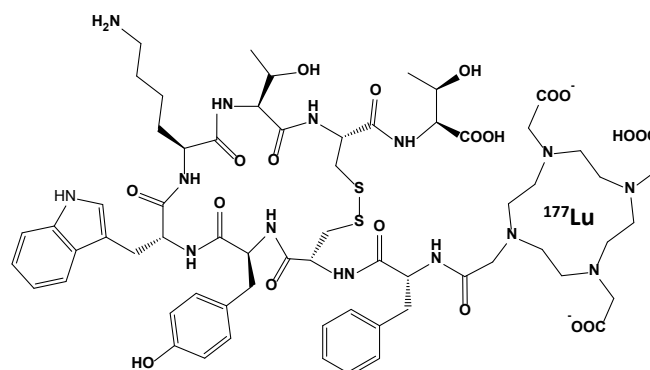


Figure 2: Chemical Structure of the PRRT agent ^{177}Lu -DOTA-TATE

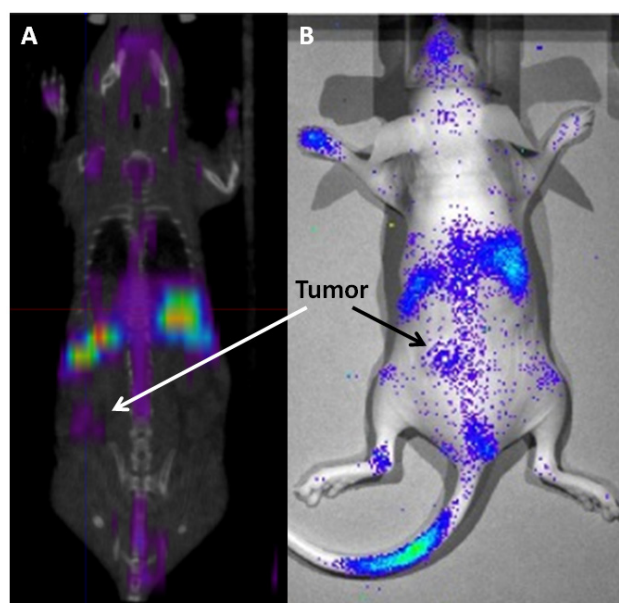


Figure 3: (A) SPECT/CT and (B) Cerenkov imaging of a live athymic nude mouse bearing a subcutaneous UMUC3/3T3 stroma-rich bladder cancer tumor at $t = 24$ h after i.v. injection of 2.5 mCi of ^{177}Lu -LCP. Tumor accumulation of ^{177}Lu is clearly visible above background, although not as high as liver and spleen accumulation. SPECT and Cerenkov agree on the overall biodistribution of ^{177}Lu . Source: Adapted from Satterlee et al²⁵. Copyright © 2015 Elsevier B.V. License number: 3815560646760

Local Delivery of Radiation Therapy

When delivered systemically, nanoparticle-based therapeutics are generally too large to clear from the body through the kidneys, and therefore tend to accumulate in the body's other clearing organs, which are primarily the liver and spleen. Splenocytes and the liver's Kupffer cells will internalize these nanoparticles and attempt to metabolize the contents⁵⁹, so when delivering therapeutic siRNA or DNA, this off-target delivery is not of much consequence. Additionally, since siRNA is generally chosen to knock down upregulated mRNA that is not abundant in healthy cells, and gene

therapy often seeks to reintroduce a lost or downregulated gene that is present in healthy cells, off-target effects resulting from delivery to hepatocytes or other healthy cells are dampened. In contrast, when radiation is delivered, the cytotoxic decay products can reach several millimeters away from the delivered source, indiscriminately acting on healthy and cancerous cells alike. Although cancer cells are more susceptible to DNA damage due to their increased proliferation rate and decreased DNA repair capacity, care must be taken to minimize the dose to off-target organs. Thus, current internal radiation therapies focus on local delivery of the radiation in order to increase the absorbed dose to the tumor and limit dose to healthy tissues.

Brachytherapy

Brachytherapy is a type of clinical radiotherapy in which the therapeutic radioactive source is permanently or temporarily placed in or near the tumor site. The majority of brachytherapy procedures are performed in prostate cancer, and placement of the source(s) is pre-planned using transrectal ultrasound to maximize dose to the tumor while minimizing dose to healthy tissue, allowing safe dose escalation to nearly double that for external beam radiation therapy⁶⁰. In high dose rate (HDR) brachytherapy, sealed sources of Iridium-192 are temporarily placed within the prostate via intraperineal catheterization, while in low dose rate (LDR) brachytherapy, sealed radioactive seeds measuring ~0.8 mm x 4.5 mm are permanently deposited at precise intervals in the prostate. The radioisotopes Cesium-131, Iodine-125, and Palladium-103 are preferred for LDR brachytherapy because their low-energy emissions provide rapid dose fall-off with increasing distance from the source, minimizing off-target dose⁶¹. To decrease incidence of seed movement or loss, each row of seeds can be sutured together to create a strand of several seeds⁶². LDR seeds are typically placed 1 cm apart⁶³, which allows a tunable dose to different areas of the prostate based on the activity encapsulated in each seed.

The new concept of nanobrachytherapy is currently limited to simple intratumoral injection of radioactive nanoparticles⁶⁴, but there may be some value in delivering nanoparticles that can evenly diffuse within the tumor. Major difficulties would be in reproducible dosimetry and clearance of the nanoparticles out of the tumor, as clearance would affect dosimetry as well as convey off-target toxicity to the clearing organ. In order to control diffusion and minimize clearance, nanoparticle penetration through tissue could be tuned by modifying their charge and PEGylation density. Highly loaded ¹⁷⁷Lu-LCP may be

a good probe to quantify nanoparticle diffusion and clearance in the tumor, both by live animal imaging and in frozen tissue sections. In its current formulation, LCP may be too acid-sensitive to ensure minimal clearance of the radioisotope over the initial decay of the delivered isotope, but a similar precipitate with a lower k_{sp} such as $\text{Cu}(\text{PO}_4)$ may more effectively resist dissolution while maintaining the preferential encapsulation of trivalent radiometals. Interestingly, excess cellular copper is known to generate reactive oxygen-dependent DNA damage, which could itself provide an effective combination therapy with the delivered radionuclide.

Selective Internal Radiation Therapy (SIRT)

SIRT is another clinically approved local delivery method for internal radiation therapy. This approach delivers radioactive microspheres loaded with ⁹⁰Y to the liver via a surgically implanted catheter that is fed up the hepatic artery. It is known that while the portal vein provides the majority of the blood supply to healthy hepatocytes in the liver, liver tumors are fed by the hepatic artery. Thus, introduction of microspheres through the hepatic artery selectively target the tumor regions. Commercially available microsphere formulations such as Sir-Spheres by Sirtex are approximately 30 μm in diameter, which prevent cellular uptake and deep tumor penetration, but also help retain the microspheres around the tumor site while minimizing clearance through the hepatic capillaries. SIRT is currently in several phase II and III clinical trials for use in combination with front-line chemotherapeutics against hepatocellular carcinoma (HCC) and advanced metastatic colorectal cancer^{65, 66}. The data from recent clinical trials show that ⁹⁰Y-SIRT + systemic chemotherapy provides significantly more hepatic progression-free survival than chemotherapy alone while maintaining tolerable liver and lung toxicity.

Although local hepatic delivery using ⁹⁰Y-microspheres for SIRT has shown success, there are areas in which it can be improved. Most notably, the lack of tumor penetration by the microspheres requires a high-energy β emitter with a long depth of penetration to reach the deeper tumor tissue. To maximize the absorbed dose to deep tumor cells, a large dose must be provided at the tumor periphery, which increases toxicity to healthy liver tissue. A future ¹⁷⁷Lu-LCP-SIRT therapy may be able to address these issues. With a diameter of just 40-60 nm, the much smaller ¹⁷⁷Lu-LCP could penetrate much deeper into the tumor mass, and the addition of an appropriate targeting ligand would allow selective uptake of the particles into the tumor cells themselves,

bringing the radioactive payload closer to the target DNA in the cell nucleus. The lower energy and shorter depth of penetration of ^{177}Lu 's β emissions may then be adequate to treat the entire tumor at a lower total dose and with less toxicity to nearby healthy tissue, although the efficacy of ^{177}Lu as the encapsulated radionuclide should be compared with ^{90}Y . If ^{177}Lu -LCP is effective, the γ decay from ^{177}Lu could then allow simultaneous confirmation of ^{177}Lu -LCP accumulation in the tumor mass via SPECT imaging. Additionally, ^{177}Lu -LCP could be easily co-loaded with a phosphorylated chemotherapeutic for localized chemoradiation therapy.

The much smaller size of LCP compared to Sir-Spheres also presents new challenges regarding the clearance of ^{177}Lu away from the tumor site, not only from those particles that are injected into the hepatic artery and do not enter the tumor mass, but also from any radiation that initially enters the tumor and exits at a later time. To ensure maximal tumor accumulation and minimal clearance of ^{177}Lu -LCP through the vena cava, simple changes to the surface of the particle should be made to encourage interaction with the tumor cell membrane and increase receptor-mediated endocytosis, such as modifying the outer leaflet with cationic lipids, lowering the surface PEG density, and adding a high affinity targeting ligand. Peptide targeting ligands may be the most successful class of ligand for this purpose, as they generally bind with higher affinity than small molecule ligands, and are less bulky, expensive, and immunostimulatory than antibodies. Peptide ligands against the $\alpha_6\beta_1$ receptor overexpressed in human colon cancer cells may be successful against liver metastasis^{67, 68}, and similarly, overexpression of glypican-3 (GPC-3) in HCC has allowed synthesis and use of targeted peptides^{69, 70}. Changes in the nanoparticle injection speed should also be considered, as a slower infusion of particles into the tumor will avoid saturation of the surrounding blood vessels and tumor cell receptors. The introduction of novel nanodelivery approaches does indeed present new challenges in local delivery of radioisotopes, but the advantages posed by these new methods should make overcoming these challenges a worthwhile venture.

Original Research: PET Imaging with LCP

The successful encapsulation of ^{177}Lu and ^{111}In in the LCP core^{25, 26} prompted experiments to encapsulate a radionuclide into an LCP formulation already containing chemotherapeutics in order to better study its kinetics and biodistribution. The LCP formulation that was chosen contained the cytotoxic nucleotide analog gemcitabine monophosphate, and

had previously been published as a monotherapy and in combination with siRNA^{16, 19, 20}. In previously unpublished experiments from this lab, trace ^{64}Cu was encapsulated in the GMP-LCP core via co-precipitation and achieved a moderate encapsulation efficiency of ~50% because of its slightly lower solubility with phosphate when compared to calcium (a difference in K_{sp} of 3-4 orders of magnitude). Using PET/CT, we were able to record a continuous dynamic scan documenting the first two hours after intravenous injection into mice (Supplementary Video S1), followed by additional time points at 16, 22, and 48 h after injection. As shown in Figures 4 and 5A, tumor accumulation of ^{64}Cu -GMP-LCP reached a maximum at $t = 22$ h and was clearly visible in the PET image. Figure 5A shows that while max tumor uptake was sustained throughout the study, liver uptake peaked just 20 min after injection and showed a continuous decrease over time. This time-dependent decrease in liver accumulation was not consistent with our experiments using ^{177}Lu and ^{111}In , in which liver accumulation of the probes remained high after 24 h.

The difference in $^{177}\text{Lu}/^{111}\text{In}$ and ^{64}Cu biodistribution is likely because of downstream action after the radionuclide cargoes are released from LCP via acid-sensitive dissolution in the cellular endosome. Because trivalent radiometals like ^{177}Lu are not metabolically active, they are not actively excreted in the bile and intestines by the liver. When free $^{177}\text{LuCl}_3$ is injected intravenously, intestinal accumulation of ^{177}Lu is not observed²⁵. In contrast, copper is a micronutrient that is actively metabolized in the liver, secreted into the bile, and excreted into the intestines⁷¹. Previously published time-dependent PET data of free $^{64}\text{CuCl}_2$ injected intravenously shows high initial liver uptake, followed by a decrease in liver accumulation via washout through the intestines⁷². Figure 5C similarly shows that as ^{64}Cu is excreted from the liver, the intestines begin to show ^{64}Cu accumulation. Because free ^{64}Cu can only be excreted after release from LCP, this may suggest that LCP releases its payload within just a few hours of deposition.

Quick copper clearance from off-target organs can be desirable if one wants to decrease background signal or use copper itself as a theranostic agent⁷², but when using ^{64}Cu as a probe for nanoparticle biodistribution, it is important that ^{64}Cu pharmacokinetics accurately represent those of the nanoparticle. Problematic copper metabolism after delivery can be remedied by chelating the copper to render it metabolically inactive. For many years, the most effective copper chelators were the tetraazamacrocyclics DOTA and TETA, which

admittedly had well-documented stability issues *in vivo*, allowing ^{64}Cu to transchelate onto ceruloplasmin, the main copper-binding enzyme in the liver^{73, 41}. The chelator TE2A has shown better ^{64}Cu stability than DOTA and TETA, allowing faster and more complete off-target clearance *in vivo*^{74, 75}, and recently, a class of bifunctional, bicyclic chelators called sarcophagines has shown excellent ability to stably chelate copper for tumor imaging^{76, 77}. Sarcophagines could potentially be covalently attached to the distal ends of PEG or outer leaflet lipids on nanoparticles in order to provide stable PET imaging. To achieve reproducible results, chelator density on the nanoparticle must be optimized, excess chelators must be removed before addition of ^{64}Cu , and unincorporated ^{64}Cu —if any—must be purified away before injection. Alternatively, it may be possible to incorporate chelated ^{64}Cu directly into the nanoparticle core. A chelator that remains highly charged after ^{64}Cu incorporation may co-precipitate with calcium phosphate in the LCP core, eliminating the need for surface chelation and accurately representing the biodistribution of the LCP core while also maintaining the metabolic inertness of the isotope.

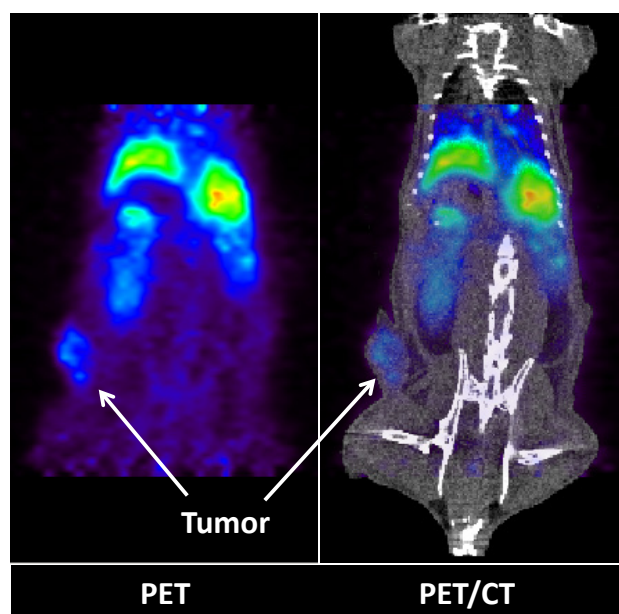
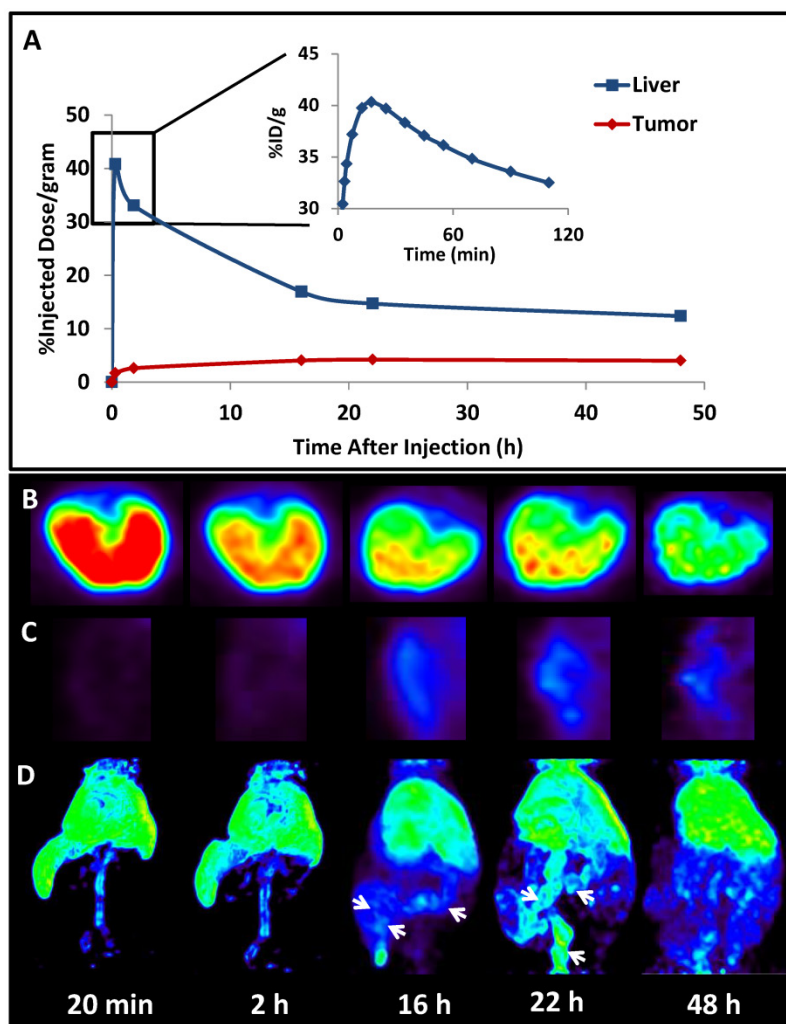


Figure 4: PET (left) and PET/CT (right) images of a live athymic nude mouse bearing a subcutaneous A549 lung cancer tumor. Images show 2-D coronal sections at $t = 22\text{h}$ after ^{64}Cu -GMP-LCP administration. Liver and tumor accumulation of ^{64}Cu are both clearly visible. Dose of $^{64}\text{Cu} = 500 \mu\text{Ci}$; dose of GMP = 5 mg/kg.

Figure 5: Biodistribution kinetics in a live athymic nude mouse bearing a subcutaneous A549 lung cancer tumor: (A) Quantification of ^{64}Cu from ^{64}Cu -GMP-LCP in liver and tumor, calculated from PET reconstruction ROIs; (B) PET images of 2-D transverse liver sections at different time points after injection; (C) PET images of 2-D coronal tumor sections at different time points after injection; (D) 3-D reconstruction of ^{64}Cu biodistribution from ^{64}Cu -GMP-LCP at different time points after injection. Arrows at 16 h and 22 h time points show accumulation of ^{64}Cu in the intestines. Dose of $^{64}\text{Cu} = 500 \mu\text{Ci}$; dose of GMP = 5 mg/kg.



Future Directions: Magnetic Resonance Imaging (MRI) with LCP

MRI uses the concept of nuclear magnetic resonance to reconstruct anatomical images. By using a large magnet to manipulate the spin and magnetic moment of hydrogen atoms in the body, and then measuring their relaxation times, the distribution and structural characteristics of the hydrogen atoms can be measured⁷⁸. MRI contrast agents such as iron oxide or gadolinium can themselves be magnetized when under the influence of an external magnetic field, which shortens both the T1 and T2 relaxation times of nearby hydrogen atoms⁷⁹. Thus, to localize the contrast agents, the affected hydrogen

atoms are measured, and not the contrast agents themselves. This requires that a baseline reading be completed before adding the contrast agents.

LCP has not yet been utilized in MRI for any purpose, but there are definitely some interesting theranostic applications for this imaging modality, since gadolinium is another trivalent cation that should be easily incorporated into the LCP core. One advantage that MRI has over SPECT and PET is that it does not use radioactivity to generate an image, which makes formulation and optimization of a theranostic nanoparticle formulation safer and less expensive. What functionally separates gadolinium and MRI from SPECT and PET imaging, however, is the fact that gadolinium must interact with the hydrogen atoms found mostly in water or tissue to generate its contrast, unlike radioisotopes that decay independent of their surroundings. The magnetic moment generated by gadolinium on water decreases with distance to the sixth power, which means that gadolinium encapsulated in LCP and surrounded by calcium phosphate and lipid membrane will provide a much lower signal than free or released gadolinium. Our previous experiments with ^{64}Cu have provided some insight into the release profile of LCP in the liver, but using MRI to quantify gadolinium release from LCP could show relative release rates in all organs, determine if any release occurs in circulation, and measure whether the low pH of the tumor does indeed cause more rapid LCP dissolution. This type of information would not be available if gadolinium were chelated to the outside of the particle, as it has been repeatedly shown that chelated gadolinium retains its ability to magnetize water^{80, 81}.

This approach can be taken one step further. By encapsulating or chelating ^{64}Cu to LCP as described above and also encapsulating gadolinium in the core, combined PET/MRI could be used to simultaneously measure the LCP biodistribution and release, showing which organ regions contain both released and unreleased LCP over time. This type of information could provide insight into what kind of dose the tumor and other organs are receiving, as faster release of a small molecule drug or siRNA from LCP should cause more acute toxicity than slow, sustained release.

Optical Imaging: Use Caution

Optical imaging generally uses fluorescent or luminescent contrast agents with emission spectrums in or near the visible light spectrum. Fluorescent compounds require excitation by light before they are able to emit light of their own, while other luminescent compounds can utilize chemical reactions or biological processes to emit light without an incident excitation wavelength. The wide range of

available fluorescent probes and the low cost of optical imaging make fluorescent theranostics a very desirable modality for both *in vitro* and *in vivo* work, although attenuation and quenching *in vivo* can be problematic.

A discussion on fluorescent theranostics has been conspicuously missing from this review, in part because of a recent article published by the Huang lab that has detailed several caveats when dealing with *in vivo* and *ex vivo* quantification of fluorescent signal⁸². Because of high tissue autofluorescence at lower wavelengths in the visible spectrum⁸³, red and near-infrared (NIR) probes are extensively used to quantify nanoparticle biodistribution. In order to determine if these fluorescent probes accurately measure nanoparticle accumulation, the referenced article compared the biodistribution of LCP using three different quantification methods: Texas red oligonucleotide fluorescence, ^3H -oligonucleotide β emission, and ^{111}In γ emission. For consistency, each contrast agent was loaded into the LCP core. While the biodistributions of ^3H and ^{111}In agreed, the high amounts of blood in the liver, spleen, and other organs quenched and attenuated the fluorescent signal in those organs, inaccurately causing measurements of higher relative fluorescence in the tumor. Even after perfusion to remove circulating blood from the liver, fluorescence quenching was not completely eliminated. This phenomenon also occurred when using a near-infrared dye. The quenching occurs at least in part because the secondary absorption peaks of hemoglobin and oxyhemoglobin present around the red and NIR emission spectra absorb the light emitted from the fluorescent probes, effectively lowering their signals in blood-rich organs such as the liver. Signal quenching could also occur because of greater tissue thickness of liver vs tumor, but the referenced article also compared tissue homogenates to show that equal amounts of different tissues will yield different fluorescence readings after adding equal concentrations of a fluorescent probe. These results suggest that while comparing fluorescent signals in the same type of organ can provide qualitative results, comparing relative fluorescence across different organ types, especially when comparing blood-rich organs like the liver to sparsely vascularized tumors, is an inaccurate quantitative method and should not be used unless adequate standards and controls are generated.

Conclusions

The versatile LCP platform has proven itself to be a successful theranostic agent and it is clear that there are many more future opportunities for LCP to

combine simultaneous therapy and imaging *in vivo*. Our lab has been primarily known as a formulation lab, but we have recently focused on innovating upon LCP in order to broaden therapeutic approaches and learn more about tumor biology. LCP's success has not been based on changes in formulation as much as on innovative applications in cancer vaccine therapy, gene therapy, and theranostics that provide solutions in areas where therapeutic options are lacking. In 2015, nearly 3,000 peer-reviewed articles were published in the field of cancer nanotechnology, and many had imaging or theranostic components to aid in tumor detection and treatment. It is surely an exciting time in this field, as hundreds of minds collaborate against cancer, but as competition increases, the focus must not be on simply publishing novel papers, but on broadening our knowledge of cancer and improving treatment options for others.

Supplementary Material

Supplementary video S1.

<http://www.thno.org/v06p0918s1.mpg>

Abbreviations

DOPA: 1,2-dioleoyl-*sn*-glycero-3-phosphate

DOTAP: 1,2-dioleoyl-3-trimethylammonium-propane

DSPE-PEG2000:

N-(Carbonyl-methoxypolyethyleneglycol 2000)-1,2-distearoyl-*sn*-glycero-3-phosphoethanolamine, sodium salt

K_{sp}: Solubility Product Constant

LCP: Lipid-Calcium-Phosphate

Lu: Lutetium

NIR: Near-Infrared

SPECT: Single Photon Emission Computed Tomography

PET: Positron Emission Tomography

MRI: Magnetic Resonance Imaging

SIRT: Selective Internal Radiation Therapy

Acknowledgment

ABS is supported by the National Science Foundation's Graduate Research Fellowship Program and by the Graduate School at the University of North Carolina at Chapel Hill. The work in this lab is supported by NIH grants DK100664, CA194067, and CA149387.

We thank the Small Animal Imaging facility at the UNC Biomedical Research Imaging Center, especially staff Jon Frank and Joseph Merrill, for providing the PET/CT imaging service.

Original Animal Studies

Animal Studies used to produce original

research in Figures 4 and 5 utilized 6–8 week old female athymic nude mice. These mice were purchased from the National Cancer Institute (Bethesda, MD) and bred at the Division of Laboratory Animal Medicine at the University of North Carolina-Chapel Hill. All work performed on these animals was approved by the Institutional Animal Care and Use Committee at the University of North Carolina-Chapel Hill.

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Competing Interests

The authors have declared that no competing interest exists.

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