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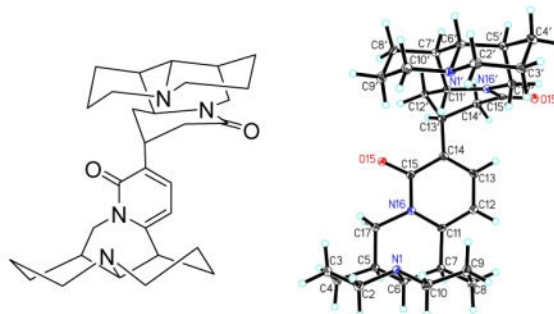
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Tetrahedron Lett. 2016 November 16; 57(46): 5047–5049. doi:10.1016/j.tetlet.2016.10.009.**Ochrocephalamine A, a new quinolizidine alkaloid from *Oxytropis ochrocephala* Bunge**Li-Na Liu^{1,3,||}, Jian-Qiang Ran^{1,||}, Li-Jun Li¹, Yu Zhao⁴, Masuo Goto⁴, Susan L. Morris-Natschke⁴, Kuo-Hsiung Lee^{4,5,*}, Bao-Yu Zhao^{2,*}, and Cheng-Jian Tan^{1,*}¹School of Ethnic Medicine, Guizhou Minzu University, Guiyang 550025, China²College of Veterinary Medicine, Northwest A&F University, Yangling, Shaanxi, 712100, China³Department of Biotechnology, Guizhou Medical University, Guiyang 550025, China⁴Natural Products Research Laboratories, UNC Eshelman School of Pharmacy, University of North Carolina, Chapel Hill, NC 27599, United States⁵Chinese Medicine Research and Development Center, China Medical University and Hospital, Taichung 401, Taiwan**Abstract**

One dimeric matrine-type alkaloid, ochrocephalamine A (**1**), was isolated from the poisonous plant *Oxytropis ochrocephala* Bunge. Its structure was elucidated by spectroscopic data and single-crystal X-ray diffraction. The insecticidal and cytotoxic activities of **1** were evaluated.

Graphical Abstract

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Supplementary Material

NMR spectra, bioactivity assay, and the X-ray crystallographic data (CIF) of **1** are available free of charge via the Internet at <http://>

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Keywords

Poisonous plant; *Oxytropis ochrocephala* Bunge, *Oxytropis ochrocephala*; Alkaloid; Quinolizidine alkaloid

The plant *Oxytropis ochrocephala* Bunge is a common poisonous plant found in the western grasslands of China¹⁻². After ingesting the plant, livestock gradually develop a chronic neurological disease, characterized by a staggering gait and muscular incoordination, giving rise to the vivid name 'locoweed' for this poisonous plant³. Previous phytochemical investigations on *O. ochrocephala* provided quinolizidine and indolizidine alkaloids, as well as flavonoids and saponins⁴⁻¹⁰.

Quinolizidine alkaloids are important natural products exhibiting a broad array of pharmacological activities, such as antineoplastic, antibacterial, antiviral, and insecticidal properties¹¹. Recently, we reported the synthesis of sophoridine derivatives as potential antitumor agents¹². As part of our ongoing research program to identify bioactive constituents, particularly from this alkaloid class, we obtained a new quinolizidine alkaloid, ochrocephalamine A (**1**), from *O. ochrocephala*. Herein, we describe the isolation, structural elucidation and bioactivity of this compound.

The whole plant (20.0 kg) of *O. ochrocephala* was percolated three times with 95% EtOH to give a crude extract (3.0 kg). The extract was concentrated to dryness under reduced pressure, followed by partitioning between CH₂Cl₂ and 2% HCl. The aqueous phase was then adjusted to pH 11 with 3% NaOH and extracted with CH₂Cl₂ to give crude alkaloids (100 g). The crude alkaloids were subjected to a silica gel column chromatography eluted with CH₂Cl₂/MeOH (1:0 to 0:1) to obtain fractions A, B and C. Fraction B was chromatographed on silica gel [CH₂Cl₂/MeOH (10:1)] and then RP-18 (30% MeOH) columns to yield ochrocephalamine A (**1**, 20 mg).

Ochrocephalamine A (**1**)¹³ was obtained as a light-yellow solid (CH₂Cl₂), and its molecular formula was established as C₃₀H₄₂N₄O₂ by HR-EI-MS (*m/z* 490.3296 [M]⁺, calcd 490.3308), requiring 12 degrees of unsaturation. The ¹H NMR data of **1** (Table 1) showed two groups of signals characteristic for a matrine-type alkaloid at δ_{H} 4.29 (1H, dd, *J* = 12.8, 4.0 Hz) and 3.11 (1H, *brt*, *J* = 12.8 Hz) and at 4.09 (1H, dd, *J* = 12.8, 7.2 Hz) and 3.80 (1H, *brt*, *J* = 12.8 Hz)¹⁴. In addition, two coupled olefinic protons [δ_{H} 7.30 (1H, d, *J* = 7.6 Hz), 6.51 (1H, d, *J* = 7.6 Hz)] were distinguished by NMR analysis. The ¹³C-NMR data (Table 1) accounted for 30 carbon signals, classified as four sp² quaternary carbons including two lactam-CO signals at δ_{C} 170.9 and 164.6, two sp² and eight sp³ methines, and 16 methylenes. Six sp² carbons accounted for 4 out of the 12 degrees of unsaturation, and the remaining 8 degrees required that **1** is an octacyclic system.

Comprehensive analysis of the 1D NMR data suggested that **1** contains two matrine-type alkaloid moieties, sophoramine and matrine¹⁵. Considering the octacyclic ring system of **1**, this compound was likely a dimeric alkaloid of sophoramine and matrine connected through a C-C bond. Accordingly, the methine of sophoramine at C-14 (δ_{C} 116.4) and methylene of matrine at C-13 (δ_{C} 19.2)¹⁵ were replaced by the quaternary carbon (δ_{C} 130.1) and methine

(δ_C 30.0), respectively, in **1**. The HMBC correlations from H-13 to C-15, C-11 and C-13', H₂-14' to C-15', C-13' and C-14, H-13' to C-15' and C-14', and H₂-12' to C-7', C-11', C-13' and C-14 confirmed the C-14–C-13' connection. Thus, the gross planar structure of ochrocephalamine A was established as shown (Figure 2a).

Determinations of the stereo configurations of **1** were particularly challenging. The ROESY correlations of H-17' α to H-5', H-6' to H-7' indicated H-5', H-6' and H-7' were α configuration¹⁰. H-11 α was assigned in β -configuration on the basis of the mutual ROESY between H-17' β and H-11'¹⁰. The relative configuration of H-13' could not be determined by a ROESY experiment due to unobserved correlations between H-11' and H-13' as well as H-11' and H-14'. Additionally, the relative configuration of H-5, H-6 and H-7 also can't be determined by ROESY experiment due to unobserved correlations between one of them with another confirmed proton configuration. Thus, we attempted to prepare crystals of **1** for X-ray determination. Eventually, a single crystal of this compound was obtained from EtOH. The connection between C-14 and C-13' was confirmed by X-ray crystallographic diffraction with CuK α radiation and the relative configurations of H-5, H-6, H-7 and H-13' were α -configuration (Figure 2b)^{13,16}. The absolute configurations of **1** were also determined by the refined Hooft parameter value 0.01(6) for 1892 Bijovet pairs with a probability of 1.000 as shown in Figure 2b¹⁶.

From a biogenetic point of view, matrine could react with known sophoridine through a Michael addition reaction to give the intermediate B. Aromatization of this intermediate would then yield **1**.

We evaluated the contact toxicity of **1** against *Spodoptera litura* third-instar larvae by the micro-drip method. One μ L of **1** (3 mg/mL in EtOH) was dropped on the pronotum of larvae. After five days, 50% of larvae were killed. In cytotoxicity assays, compound **1** was not active against A549, KB, KB-VIN, and MDA-MB-231 cell lines (ED₅₀ > 20 μ M).

O. ochrocephala is widely distributed in the grasslands of eastern and central Asia, Australia, and western North and South America, where it poses an extreme risk to grazing animals, resulting in a chronic neurological disease^{1,3}. The indolizidine alkaloid swainsonine is claimed as the constituent poisonous to livestock³. In view of the plant itself, poisonous secondary metabolites can improve the plant's chances of survival and reproduction by deterring animals, such as sheep, cattle, and horses. In addition, quinolizidine alkaloids^{8–9, 17}, such as **1**, play an important role in defense against insects. Biogenetically, indolizidine and quinolizidine alkaloids originate from the same precursor L-lysine¹⁸. The use of one biogenetic pathway to produce two different types of alkaloids involved in plant defense against both livestock and insects, respectively, is a smart and efficient strategy to combat the survival challenge.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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13. Ochrocephalamine A (**1**): yellow crystals (EtOH); mp 159–161 °C; ESIMS m/z 491 [M + H]⁺; HREIMS at 490.3296 [M]⁺ (calcd. 490.3308, C₃₀H₄₂N₄O₂); [α]_D²⁴ +32.2 (*c* 0.48, MeOH); UV (MeOH) λ_{max} (log *e*) 202 nm (4.2), 311 nm (3.9), 373 nm (2.3); IR (KBr) ν_{max} 2932, 2860, 2805, 2764, 2745, 1639, 1597, and 1554 cm⁻¹; ¹H and ¹³C NMR data (Table 1). Crystal data for **1**: C₃₀H₄₂N₄O₂·6(H₂O), *M* = 598.77, monoclinic, *a* = 13.0528(3) Å, *b* = 8.3014(2) Å, *c* = 14.8105(4) Å, *α* = 90.00°, *β* = 101.6050(10)°, *γ* = 90.00°, *V* = 1572.01(7) Å³, *T* = 100(2) K, space group *P*2₁, *Z* = 2, μ(CuKα) = 0.745 mm⁻¹, 13217 reflections measured, 4874 independent reflections (*R*_{int} = 0.0386). The final *R*_{*I*} values were 0.0521 (*I* > 2σ(*I*)). The final *wR*(*F*²) values were 0.1565 (*I* > 2σ(*I*)). The final *R*_{*J*} values were 0.0525 (all data). The final *wR*(*F*²) values were 0.1572 (all data). The goodness of fit on *F*² was 1.082. Flack parameter = -0.3(2). The Hooft parameter is 0.01(6) for 1892 Bijvoet pairs. Deposited to Cambridge Crystallographic Data Center with No. CCDC 1449099.
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- Ochrocephalamine A (**1**) was isolated from *Oxytropis ochrocephala* Bunge.
- The poisonous plant *O. ochrocephala* poses an extreme risk to grazing animals.
- Compound **1** is a dimeric matrine-type alkaloid.
- Its structure was solved from spectral data and single-crystal X-ray diffraction.
- Compound **1** showed contact toxicity against *Spodoptera litura* third-instar larvae.

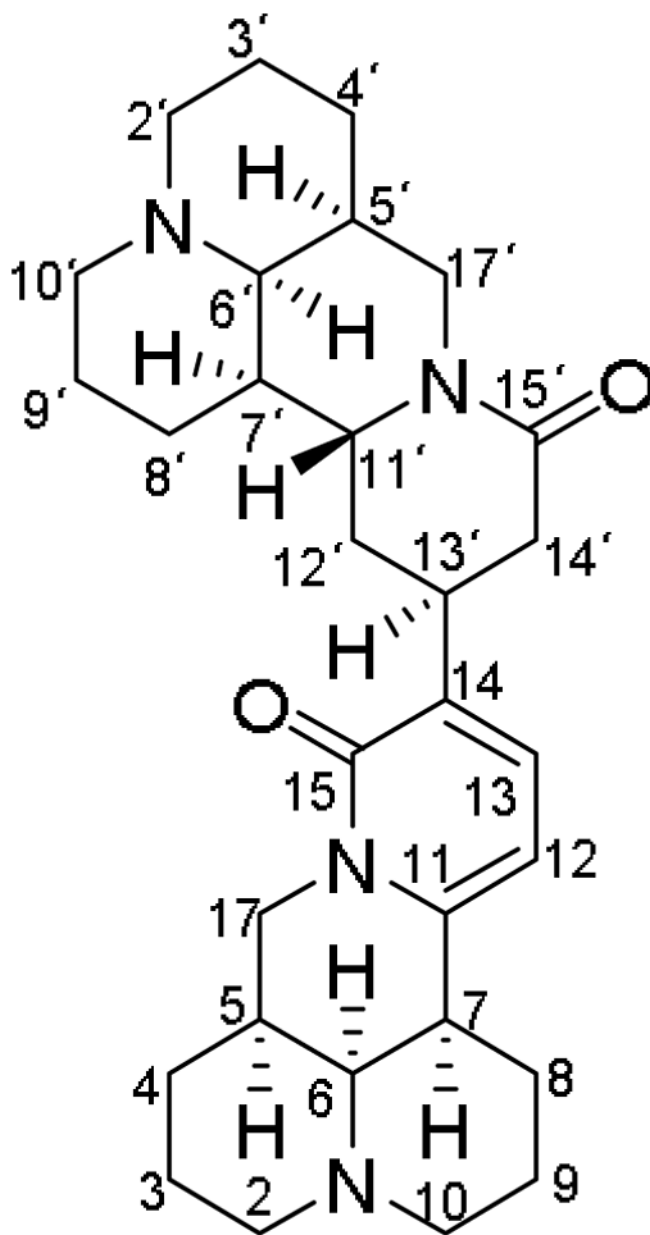


Figure 1.
The structure of ochrocephalamine A (1)

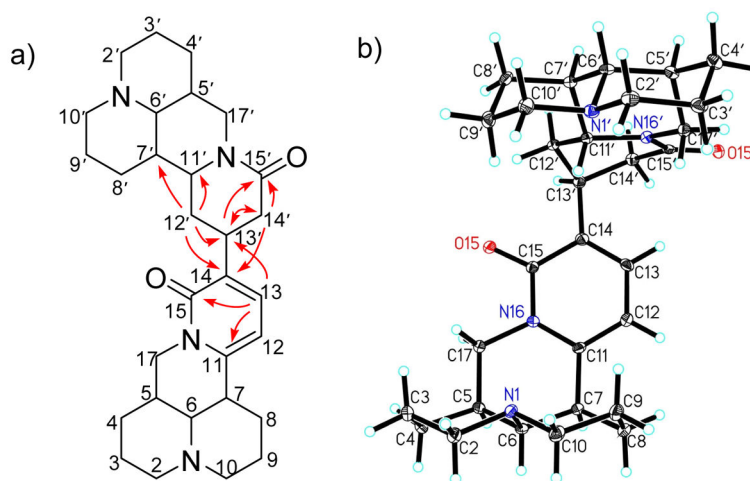
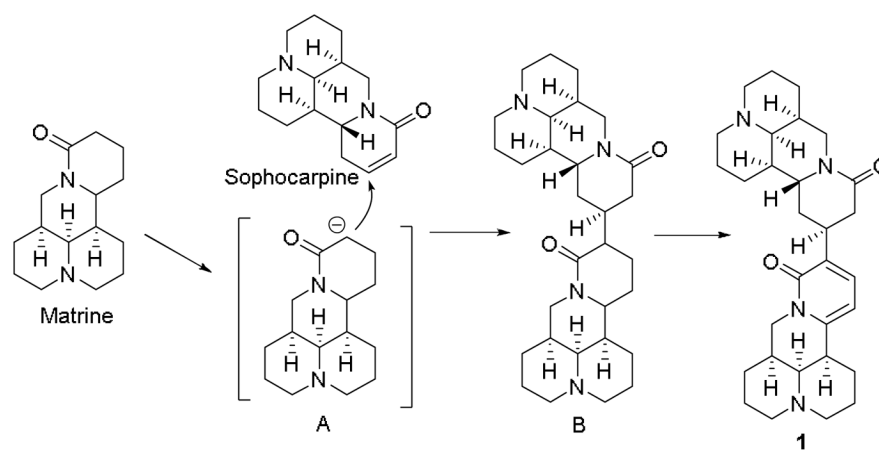


Figure 2.
a) key HMBC correlations of **1**, b) X-ray crystal structure of **1**.



Scheme 1.
Biogenetic pathway proposed of ochrocephalamine A (**1**)

Table 1
 ^1H (400 MHz) and ^{13}C NMR (100 MHz) Data of Ochrocephalamine A (1) in CD_3OD

| No | δ_{H} (mult, J) | δ_{C} | no. | δ_{H} (mult, J) | δ_{C} |
|----|--|---------------------|-----|--|---------------------|
| 2 | 2.82 (m) ^a 2.07 (m) ^a | 57.8 | 2' | 2.82 (m) ^a 2.07 (m) ^a | 58.3 |
| 3 | 1.56–1.65 (m) ^a | 22.2 | 3' | 1.56–1.65 (m) ^a | 21.6 |
| 4 | 1.79 (m) ^a | 27.8 | 4' | 1.67–1.71 (m) ^a | 28.7 |
| 5 | 2.25 (m) | 32.9 | 5' | 1.80 (m) | 37.5 |
| 6 | 2.33 (t, 4) | 61.5 | 6' | 2.29 (brs) | 65.6 |
| 7 | 3.03 (br-s) | 39.5 | 7' | 1.89 (m) | 42.6 |
| 8 | 2.15 (m) | 27.5 | 8' | 2.44–2.50(m) | 28.4 |
| 9 | 1.56–1.65 (m) ^a | 22.2 | 9' | 1.56–1.65 (m) ^a | 21.3 |
| 10 | 2.82 (m) ^a 2.07 (m) ^a | 57.6 | 10' | 2.82 (m) ^a 2.07 (m) ^a | 58.2 |
| 11 | | 147.7 | 11' | 3.94 (m) | 53.8 |
| 12 | 6.51 (d, 7.6) | 106.6 | 12' | 2.14 (m) | 29.8 |
| 13 | 7.30 (d, 7.6) | 135.7 | 13' | 3.38 (m) | 30.0 |
| 14 | | 130.1 | 14' | 2.62(dd, 16.0, 4.0) 2.46(dd, 16.0, 8.0) | 37.5 |
| 15 | | 164.6 | 15' | | 170.9 |
| 17 | 4.09 (dd, 12.8, 7.2) 3.80 (br-t, 12.8) | 45.5 | 17' | 4.29 (dd, 12.8, 4.0) 3.11 (br-t, 12.8) | 43.7 |

^a overlapped.